

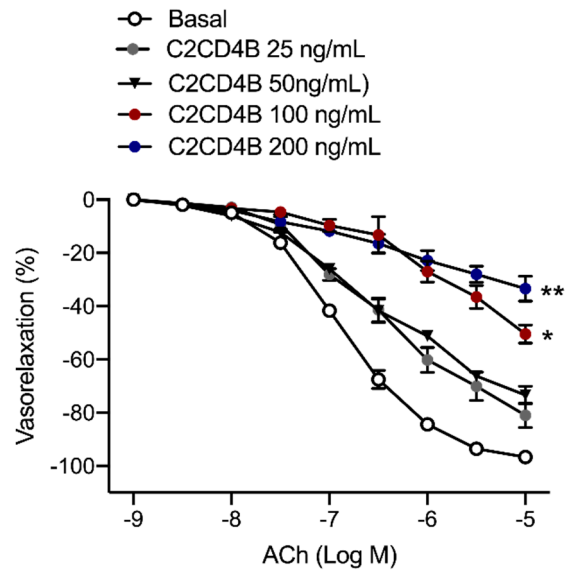
## **SUPPLEMENTARY DATA**

### **C2CD4B evokes oxidative stress and vascular dysfunction via a PI3K/Akt/PKC $\alpha$ -signaling pathway**

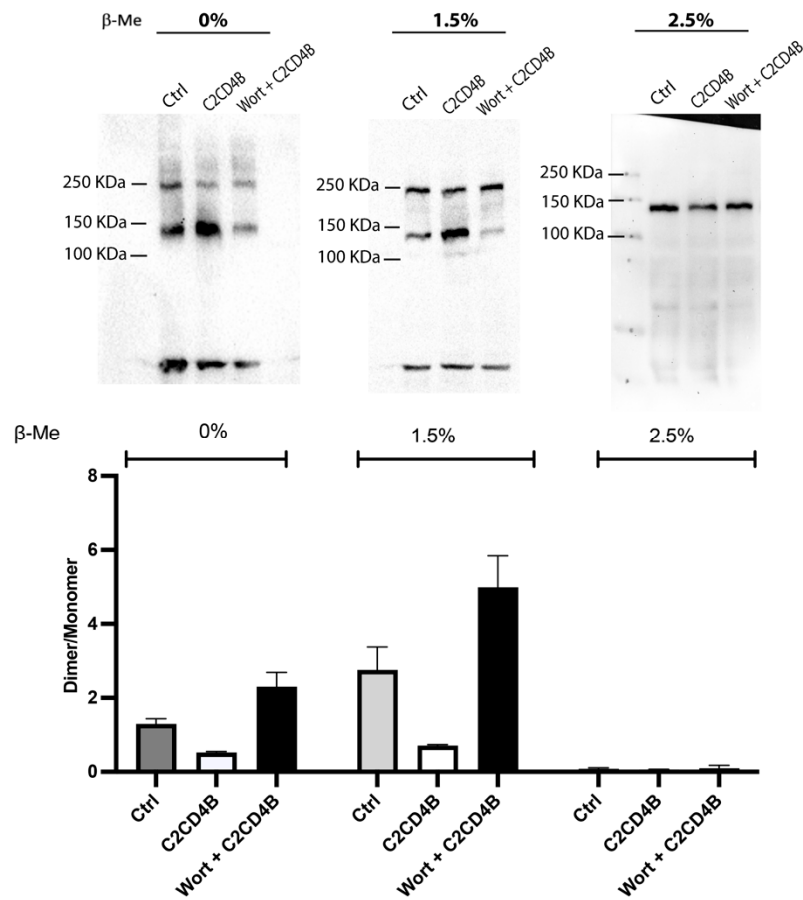
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### **Supplementary results**

The ability of C2CD4B to promote PI3K-dependent eNOS uncoupling was further corroborated by additional experiments in which we test the effect of increasing concentrations of  $\beta$ -ME on the ratio between monomer and dimer, as  $\beta$ -ME is reported to influence the dimeric and monomeric form of eNOS [57]. Our result demonstrated that, while in the presence of 0%  $\beta$ -Me oligomers and the monomer form were markedly present, at 1.5%  $\beta$ -Me oligomers were not clearly detectable, with a marked increase of dimer signals. Conversely, 2.5% of  $\beta$ -Me completely abolished the formation of both oligomers and dimeric form of eNOS, resulting in the visualization only of the monomeric form (Supplementary Figure 2).



**Supplementary Figure S1.** Experimental setup for the assessment of the effects of recombinant C2CD4B on mice mesenteric arteries. Acetylcholine (ACh)-evoked vasorelaxation in mice mesenteric arteries exposed to vehicle (basal) or different doses of recombinant C2CD4B protein for 1 hour; (n = 3).



**Supplementary Figure S2.** Effect of incubation of cell lysates from endothelial cells in the presence or the absence of the reducing agent  $\beta$ -mercaptoethanol (1.5%, 2.5%) on levels of endothelial nitric oxide synthase (eNOS). Representative western blot and densitometric analyses of 3 independent experiments evaluating eNOS dimers and eNOS monomers protein expression in HUVECs treated with vehicle (ctrl), recombinant C2CD4B alone, or pre-treated with wortmannin. Dimer/monomer eNOS were examined by low-temperature SDS-PAGE without (non-reducing) or with (reducing) 2-mercaptoethanol ( $\beta$ -Me, 1.5%, or 2.5%).