



Review

Discovering the Potential Mechanisms of Medicinal Mushrooms Antidepressant Activity: A Review

Jan Lazur *, Kamil Hnatyk , Katarzyna Kała , Katarzyna Sułkowska-Ziaja and Bożena Muszyńska *

Department of Pharmaceutical Botany, Faculty of Pharmacy, Jagiellonian University Medical College, Medyczna 9 Street, 30-688 Kraków, Poland

* Correspondence: jan.lazur@doctoral.uj.edu.pl (J.L.); bozena.muszynska@uj.edu.pl (B.M.)

Abstract: Major Depression Disease is a common mental illness that affects more than 322 million people worldwide and it is one of the leading causes of mental and physical disability. The etiology of depression is a complex interplay of psychological, social, and biological factors. Currently, psychopharmacotherapy is based mainly on the monoamine theory, which states that depression is caused by an insufficient level of monoamines such as serotonin, norepinephrine, and/or dopamine. Due to the relatively low efficacy of the typical antidepressant and the high prevalence of treatment-resistant depression (~30%), seeking new ways of prophylaxis, adjuvant therapy, or novel compounds with antidepressant activity, is a priority. According to studies that analyzed mushroom consumption patterns and depression prevalence, it was concluded that mushroom ingestion lowers the odds of depression. Medicinal mushrooms are considered functional foods because of their ability to synthesize and accumulate different types of metabolites, which enhance their health-promoting properties. The review aims to explain the antidepressant activity of edible/medicinal mushrooms by elucidating the mechanism from different perspectives: edible mushrooms as a source of serotonin precursors and psilocybin as a rapid-acting antidepressant. These compounds exhibit anti-neuroinflammatory and antioxidant activities that impact neurotrophin expression, the neurogenesis process, and influence on the gut–brain axis.

Keywords: L-tryptophan; psilocybin; BDNF; neurogenesis; neuroinflammation; antioxidant activity; gut–brain axis



Citation: Lazur, J.; Hnatyk, K.; Kała, K.; Sułkowska-Ziaja, K.; Muszyńska, B. Discovering the Potential Mechanisms of Medicinal Mushrooms Antidepressant Activity: A Review. *Antioxidants* **2023**, *12*, 623. <https://doi.org/10.3390/antiox12030623>

Academic Editors: Xiangyang Zhang, Zezhi Li and Dongmei Wang

Received: 31 January 2023
Revised: 24 February 2023
Accepted: 27 February 2023
Published: 2 March 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

As defined by the World Health Organization (WHO), depression is a common mental illness that affects more than 322 million people worldwide [1]. The typical symptoms of depression are persistent sadness, as well as the inability to feel feelings of happiness (anhedonia), sleep problems, loss of appetite, general fatigue, and cognitive problems. Depression is one of the leading causes of mental and physical disability worldwide [2].

The etiology of depression is a complex interplay of psychological, social, and biological factors. People who have experienced adverse life events, such as the death of a loved one or prolonged unemployment, are at an increased risk of developing depression. The presence of physical diseases such as cardiovascular and neurodegenerative diseases may increase the risk of depression [3,4].

Interestingly, the COVID-19 pandemic increased depressive symptoms by five percentage points, from 27.8 to 32.8% of adults in the United States in 2021 compared to the first months of 2020 [5]. The increased risk of depressive symptoms and the development of depression may be related to the so-called pandemic environment and the introduction of “lockdowns” that restrict social activities in many countries. There is also growing evidence of COVID-19 disease and an increased risk of depression in recovered adults [6]. The mechanism of the development of depressive symptoms in recovered adults is not well understood yet. One of the possible explanations for this phenomenon may be related

to the so-called “cytokine storm”—abnormally high levels of pro-inflammatory cytokines such as IL-1 β , IL-6, IL-12, and tumor necrosis factor-alpha (TNF- α) and interferon gamma (INF- γ). A cytokine storm can contribute to neurotoxicity, blood barrier disruption, or even acute necrotizing encephalopathy [7].

Clinical psychopharmacology is a branch of science that deals with the description of the use of pharmacological agents for the treatment of specific psychopathological symptoms. The beginning of this field of knowledge dates to the 1940s and 1950s [8]. The first antidepressant drug was iproniazid, introduced in the treatment of tuberculosis in 1952 [9]. In tuberculosis patients treated with this drug, a significant improvement in mood was observed, unprecedented in patients in this clinical state [10]. After a few years, the mechanism of action of iproniazid was described, as it turned out to be an irreversible inhibitor of the monoamine oxidase (MAO) enzyme, which in turn led to an increase in the concentration of biogenic amines in the brain [11]. Iproniazid became a precursor drug for the first antidepressants, MAO inhibitors, including tricyclopromine and phenelzine. Currently, iproniazid is not registered for the treatment of depression due to side effects, including liver damage [11].

Treatment of depression is based on the theory of monoamines introduced in the 1960s, which states that this disease is caused by a decreased level of monoamines (serotonin, norepinephrine, and dopamine) in the brain [12]. The mechanism of action of drugs used in the first line of depression treatment is inhibition of neuronal reuptake of monoamines from synaptic clefts, as in the case of selective serotonin reuptake inhibitors (SSRIs), for example, fluoxetine, citalopram, or sertraline, which reduce the activity of serotonin transporters [13]. Although these drugs are potent antidepressants, the cause of depression is not simply insufficient monoamine levels. SSRIs cause an immediate increase in serotonin transmission, while it takes several weeks for mood-elevating activity to develop in treated patients, which is associated with changes in the expression of serotonin-dependent receptors. Recent data on the development of depression have extended the theory of monoamines to include neurotrophic and neurogenic hypotheses [14,15]. Decreased levels of brain-derived neurotrophic factor (BDNF) are involved in the pathogenesis of depression [16]. BDNF is required for neurogenesis and neuroplasticity in the hippocampus [17]. In people with depression, BDNF expression is decreased in the limbic area of the brain due to neuronal atrophy. Serotonin and its receptors are involved in the regulation of BDNF levels and neurogenesis in the adult hippocampus. Chronic treatment with an SSRI has been shown to increase BDNF levels in humans and rodents [18,19]. Altered levels of other neurotrophins, such as neurotrophin-3 (NT-3), neurotrophin-4 (NT-4), and nerve growth factor (NGF), are also observed in patients with depressive disorders [20].

Medicinal/edible mushrooms and their mycelia from *in vitro* cultures are receiving increasing scientific attention for their potential to promote health. They are considered functional foods because of their ability to synthesize and accumulate different types of metabolites, which enhance their health-promoting properties and can be used as a supplement to the human diet. Studies show the multidirectional activity of medicinal mushrooms and their mycelium, including antioxidant, anticancer, anti-inflammatory, and immunostimulatory effects. Increasingly, there is also evidence of antidepressant activity [21–23].

Researchers at Penn State University published a research paper describing the link between eating mushrooms and depression [24]. The main conclusion of this population-based study, which analyzed mushroom consumption among US residents from 2005 to 2016, was that mushroom consumers are less likely to suffer from depression [24]. The results are consistent with previous small clinical studies [25–27]. However, the studies presented above did not investigate the potential mechanisms of the antidepressant effect of edible mushrooms.

This review aims to explain the antidepressant activity of edible/medicinal mushrooms by elucidating mechanisms from different perspectives, starting with answering the question of whether edible mushrooms can be a good source of indole compounds,

such as L-tryptophan (Trp)—a precursor of the brain serotonin synthesis pathway. Although psilocybin-containing mushrooms are considered inedible, psilocybin and its active constituent, psilocin, is a real candidate for being classified as a rapid-acting antidepressant (RAAD) which is especially important for patients suffering from treatment-resistant depression (TRD). That is why this review also describes the current knowledge about the mechanism of action of psilocybin action and summarizes the current progress of clinical trials considering the usage of psilocybin in TRD. The review summarizes the anti-inflammatory effect of the administration of edible/medicinal mushrooms in alleviating neuroinflammation and the influence of analysis on the activity of the kynurenic pathway for in vitro and in vivo models. Furthermore, the neurotrophic and neurogenic activity of selected edible/medicinal mushrooms of in vitro and in vivo models was summarized. The last part of this review focuses on summarizing the current knowledge of edible/medicinal mushroom species—extracts or isolated substances on the gut microbiota, which has been extensively studied over the last five years.

2. L-Tryptophan Derivatives—Essential Compounds for Serotonin Synthesis

L-Tryptophan (Trp) and its derivatives, such as 5-hydroxy-L-tryptophan (5-OH-L-Trp), and tryptamine, are related to biochemical reactions that lead to serotonin synthesis in the brain's neurotransmitters, lower levels of which are observed in clinically depressed patients [28]. These compounds have been shown to scavenge free radicals and protect cells against oxidative stress, potentially reducing the risk of certain diseases such as cancer, neurodegenerative diseases, and depression [29].

Trp is an essential amino acid and is considered an exogenic amino acid for the human body. Although its importance is the synthesis of various proteins, Trp is a precursor of serotonin (5-hydroxytryptamine) in the brain and gut. The biosynthetic pathway of serotonin is presented in Figure 1.

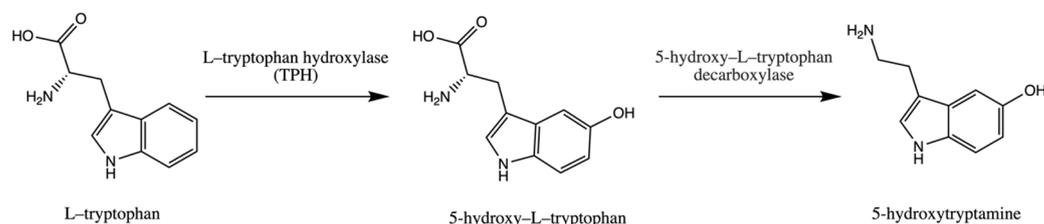


Figure 1. Pathway of serotonin (5-hydroxytryptamine) synthesis.

The serotonin metabolic pathway starts with the hydroxylation of Trp to 5-OH-L-Trp, which is decarboxylated to 5-hydroxytryptamine (serotonin). The limiting stage of serotonin synthesis is Trp hydroxylation by the enzyme Trp hydroxylase (TPH) and is not saturated at physiological brain tryptophan concentrations; therefore, serotonin synthesis in the brain is assumed to be directly connected with tryptophan transport into the brain [30–32].

Trp can be transported to the brain through a nutrient amino acid transporter protein that is involved in the transport of large neutral amino acids (LNAAs) such as valine, leucine, isoleucine, tyrosine, phenylalanine, and methionine from the bloodstream to the brain through the blood-brain barrier (BBB) [33]. The content of Trp that crosses the BBB by the nutrient amino acid transporter depends on the ratio of Trp and other LNAAs in plasma [33]. After meal ingestion, the levels of Trp and other LNAAs in plasma increase. As a result of a relatively low increase in Trp in comparison to other essential amino acids in plasma concentration, the plasma Trp/LNAA ratio decreases, and consequently, a reduced Trp influx to the brain is observed [33].

There are several factors that can influence Trp influx to the brain by influencing LNAA concentration in plasma, such as the ingestion of carbohydrates, the intake of protein amounts, or exercise. Ingestion of dietary carbohydrates led to elevated insulin levels. Insulin promotes the uptake of LNAAs in skeletal muscle, which leads to an increase in the Trp/LNAA ratio and consequently to Trp influx into brain tissue [34]. L-Tryptophan

is not transported to muscle tissue because it bonds with albumin, while other LNAAs are not.

Trp obtained from food can be transformed into serotonin in a limited amount. In mammals, approximately 95% of Trp is metabolized through the kynurenic metabolic pathway, whose products exhibit biological activity [35].

Fruiting bodies of edible mushrooms are a good source of non-hallucinogenic indole compounds such as Trp, 5-OH-L-Trp, and tryptamine (Table 1) [36–38].

Table 1. Content of L-tryptophan, 5-hydroxy-L-tryptophan and tryptamine in fruiting bodies of selected medicinal mushrooms.

Species	Source	L-Tryptophan (mg/100 g dw)	Tryptamine (mg/100 g dw)	5-Hydroxy-L-tryptophan (mg/100 g dw)	Ref.
<i>Pleurotus citrinopileatus</i>	Fruiting bodies from commercial cultivation	1.29 ± 0.08	1.29 ± 0.08	128.89 ± 10.67	[39]
<i>Pleurotus djamor</i>		24.84 ± 0.97	3.54 ± 0.18	193.95 ± 17.69	
<i>Pleurotus ostreatus</i>		5.79 ± 0.06	1.04 ± 0.15	67.45 ± 7.94	
<i>Auricularia cornea</i>		0.16 ± 0.050	2.77 ± 0.050	7.35 ± 0.200	
<i>Armillaria mellea</i>	Fruiting bodies from natural habitats	4.47 ± 0.01	2.74 ± 0.01	- ^z	[41]
<i>Imleria badia</i>		0.68 ± 0.047	0.18 ± 0.001	- ^z	
<i>Boletus edulis</i>		0.39 ± 0.020	1.17 ± 0.003	0.18 ± 0.001	
<i>Lactarius deliciosus</i>		- ^z	- ^z	0.25 ± 0.017	
<i>Hydnum repandum</i>		0.37 ± 0.09	1.46 ± 1.05	n/a	
<i>Cantharellus cibarius</i>		0.01 ± 0.002	0.01 ± 0.002	0.02 ± 0.003	
<i>Agaricus bisporus</i>		0.39 ± 0.020	0.06 ± 0.003	- ^z	
<i>Sarcodon imbricatus</i>	13.01 ± 0.01	22.12 ± 0.03	n/a	[40]	
<i>Macrolepiota procera</i>	3.47 ± 0.050	0.92 ± 0.040	22.94 ± 0.500		
<i>Suillus bovinus</i>	25.90 ± 0.200	3.15 ± 0.050	15.83 ± 0.500		
<i>Tricholoma equestre</i>	2.851 ± 0.242	2.011 ± 0.141	0.586 ± 0.041		
<i>Calocera viscosa</i>		1.26 ± 0.04	n/a	11.88 ± 0.19	[44]

n/a—data not available, ^z concentration lower than 0.001 mg/100 g dw.

The highest content of Trp and its hydroxylated derivative was observed in *Pleurotus djamor* (respectively, 24.84 and 193.95 mg/100 g dw) and *Suillus bovinus* (respectively, 25.9 and 15.83 mg/100 g dw) [39].

According to scientific data, nonfungal sources that contain high levels of Trp include soy seeds (680 mg/100 g dw), pumpkin seeds (580 mg/100 g dw), and spirulina (930 mg/100 g dw) [45]. Other researchers described that transgenic soybean plants were found to accumulate Trp at levels as high as 380 to 480 mg/100 g dw of seed flour, up to a 12-fold increase compared to Trp levels in non-transgenic seeds [46]. Wheat—durum 169 mg/100 g dw, rye 125 mg/100 g dw, barley 165 mg/100 g dw, chickpea 220 mg/100 g dw, lentil—red 139 mg/100 g dw, and kidney beans 240 mg/100 g dw are also considered good natural sources of Trp [45]. In contrast, the seeds of the *Griffonia simplicifolia* plant are considered one of the best natural sources of 5-OH-L-Trp [47,48]. Its content can be as high as 156 mg/g dw (16% of the seed weight) [48]. However, Maffei points out that mushrooms can also be a good source of this substance [47]. When analyzing the plant sources of Trp, it turns out that tomatoes are a good source (14.71 mg/100 g dw), while smaller amounts were determined in strawberries (5.7 mg/100 g dw), lettuce (2.5 mg/100 g dw), spinach (0.65 mg/100 g dw) or chicory (0.08 mg/100 g dw) [49]. Thus, it appears that both mycelium and fruiting bodies can provide an alternative source of Trp and Trp derivatives. The content of biologically active substances in mushroom samples is mainly measured after extracting them with various solvents such as methanol and ethanol from raw, lyophilized fruiting bodies. To determine the usage of selected edible mushrooms

as sources of indole compounds, the influence of various types of thermal preparation of edible mushrooms on the content of biological active substances was analyzed [40,41]. Thermal processing (dry material suspended in water and thermostated at 100 °C for 60 min in a Soxhlet apparatus) was shown to result in approximately 2 times lower indole compound content after thermal processing compared to the unprocessed. However, Trp content increases relatively in processed samples compared to that of unprocessed one. The increase in Trp content can be explained by the fact that 5-OH-L-Trp or serotonin degradation at higher temperatures [40]. The results were confirmed in another study [41]. In conclusion, the method of preparing meals with mushrooms can affect the content of indole compounds because of their sensitivity to elevated temperature. However, thermally processed mushrooms remain a good source of Trp and 5-OH-L-Trp [40,41].

Today, dietary supplements containing lyophilized fruiting bodies, extracts, or even mycelium from edible mushroom species are available in community pharmacies or in stores with so-called healthy food. Mycelium can be obtained through in vitro cultures initiated from specially prepared parts of the fruiting body, the hymenial area. One of the most important advantages of in vitro cultures is the fact that the content of biological active substances does not differ between batches because the condition of the in vitro culture is monitored and maintained at specific parameters depending on the mushroom species. Studies have shown that the content of indole compounds in biomass from in vitro cultures can be much higher than that in fruiting bodies [50]. The content of selected indole compounds in mycelia from in vitro cultures is presented in Table 2.

Table 2. Content of selected non-hallucinogenic indole compounds in mycelia of selected edible mushroom.

Species	L-Tryptophan (mg/100 g dw)	Tryptamine (mg/100 g dw)	5-Hydroxy-L-tryptophan (mg/100 g dw)	Ref.
<i>Pleurotus citrinopileatus</i>	7.82 ± 0.31	3.71 ± 0.34	368.67 ± 23.53	[39]
<i>Pleurotus djamor</i>	24.34 ± 1.44	n/d	703.56 ± 37.79	
<i>Pleurotus ostreatus</i>	1.89 ± 0.04	1.03 ± 0.15	120.11 ± 20.12	[51]
<i>Agaricus bisporus</i>	14.00 ± 0.300	0.48 ± 0.050	12.50 ± 0.671	
<i>Sarcodon imbricatus</i>	1.31 ± 0.6	4.11 ± 0.26	n/a	[52]
<i>Tricholoma equestre</i>	1.036 ± 0.093	0.598 ± 0.048	0.344 ± 0.031	[44]
<i>Imleria badia</i>	0.827 ± 0.049	0.409 ± 0.045	- ^z	
<i>Calocera viscosa</i>	1.79 ± 0.05	n/a	11.42 ± 0.20	[53]
<i>Cantharellus cibarius</i>	0.64 ± 0.013	- ^z	12.52 ± 0.671	[54]

n/a—data not available, n/d—not detected, ^z concentration lower than 0.001 mg/100 g dw.

In most cases, the content of Trp is higher in mycelia than in the fruiting bodies of selected edible mushroom species, especially *Pleurotus citrinopileatus* and *Pleurotus djamor*. The most notable change can be observed in the content of 5-OH-L-Trp, which is almost four times higher in the mycelium compared to the fruiting bodies [39]. Another advantage of making mycelium from edible mushroom species a dietary supplement is that powdered mycelium or mycelial extract does not have to be thermally processed, so thermolabile substances will not degrade.

Modification of the composition of the in vitro medium, such as the addition of indole precursors—anthranilic acid and serine—can have a positive influence on the content of indole compounds in mycelia [55]. In one experimental study, in vitro culture medium was supplemented with various concentrations of serine or anthranilic acid (0.1–0.75 g/L). For in vitro cultures of *A. bisporus* and *I. badia*, the most optimal precursor concentration was 0.5 g/L of serine for *A. bisporus* or 0.5 g/L of anthranilic acid for both species analyzed according to the content of indole compounds. The addition of 0.5 g/L of serine to *A. bisporus* in vitro cultures resulted in the highest total concentration of indole com-

pounds (186.37 mg/100 g dw). The addition of 0.5 g/L anthranilic acid to in vitro cultures of *I. badia* and *A. bisporus* resulted in the highest total concentration of indole compounds, 352.06 dw and >200 mg/100 g dw [55].

The liberation of biological substances, which is the number of substances released from the matrix of food or dietary supplement formula (tablets, hard capsules), can be measured in vitro using models of the human gastrointestinal tract. Therefore, only substances free of their matrix can be absorbed in the gastrointestinal tract. The analysis of the liberation of indole compounds was performed for *Agaricus bisporus* mycelia [56]. In the study, the content of indole compounds in artificial gastric and intestinal juice was measured after 5 time points—15, 30, 60, 90, and 120 min of incubation. The highest 5-OH-L-Trp content was established between 91.99 and 324.64 mg/100 g dw after 30 min of digestion in artificial gastric juice and after 150 min of incubation in artificial intestinal juice [56]. In a similar study on the release of indole compounds from fruiting bodies and *Tricholoma equestre* mycelia, 5-OH-L-Trp was released in the highest amount from freeze-dried mycelia after 120 min of incubation in artificial gastric juice (352.47 mg/100 g dw) and after 15 min of incubation in artificial gastric juice in the case of fruiting bodies (281.56 mg/100 g dw) [57]. For the fruiting bodies of *Suillus bovinus*, the highest content of released 5-OH-L-Trp was observed after 120 min of incubation in artificial gastric juice (237 mg/100 g dw) (for liberation study of *Imleria badia*, *Boletus edulis*, *Cantharellus cibarius*, *Lactarius deliciosus*, *Leccinum scabrum*, *Armillaria mellea*, *Suillus luteus*, *Pleurotus ostreatus*, *Auricularia polytricha*, see [58]). Based on the studies mentioned above, it can be concluded that indole compounds are released in the highest amount in artificial gastric juice compared to artificial intestinal juice. Trp is not readily liberated, regardless of whether it is from fruiting bodies or mycelia from in vitro cultures. However, 5-OH-L-Trp was one of the indole compounds that was released at the highest amount, regardless of the species analyzed [56–58].

Another important factor that should be considered when fruiting bodies or mycelium are thought to be a source of indole compounds is the bioavailability of these compounds. Bioavailability is a term used to describe the percentage or amount of a xenobiotic that reaches the systemic circulation [59]. In the case of the bioavailability analysis of secondary metabolites such as indole compounds, it will be the amount of indole compound that reaches the systemic circulation. The evaluation of the bioavailability of natural compounds in humans is rare due to requirements and restrictions imposed by ethics commissions, therefore, alternative methods involving, for example, the colon epithelial cells (CaCo-2) cell line are used to estimate the bioavailability of active substances [60]. In the study of indole absorption from *Imleria badia* mycelia, the CaCo-2 cell line was used to measure active transport, while semi-permeable membranes were used in the passive transport model after release of biological active substances in the human gastrointestinal tract model. The bioavailability of 5-hydroxy-L-tryptophan ranged from 5.21 to 11.92% using active transport modes (depending on mycelial in vitro culture conditions—an addition of zinc (VI) sulfate or zinc hydrogen aspartate). Through the passive transport model, 5-hydroxy-L-tryptophan accounted for 2% of the compound released into artificial digestive juices [61].

3. Tryptamine Derivatives—Psilocybin as a Potential Rapid Acting Antidepressant

Among patients with Major Depressive Disorder (MDD), almost 30% suffer from a treatment-resistant one [62]. To date, there is no one definition of TRD, but the most common criteria found in the literature are: failure to respond to at least two antidepressants with different mechanisms of action treatment, confirmation of adequate dosage, and duration of treatment longer than 4 weeks for each antidepressant without effect [63].

Patients suffering from TRD or for those for whom antidepressant action should be obtained in a shorter time compared to conventional — for example, patients at high risk of suicide may benefit from a novel group of antidepressants, rapid-acting antidepressants (RAADs) [64].

The definition of RAADs has not yet been specified, but contrary to conventional antidepressants, which require a few weeks to produce significant antidepressant action, they need one or a few doses to produce a significant impact on depressive symptoms or even remission, especially in a group of patients who did not respond to first-line treatment [65]. There are several mechanisms that can be responsible for the rapid antidepressant action of some drug candidates, for example, NMDA receptor antagonism, muscarinic receptors, and classic psychedelic drugs such as psilocybin or LSD, which influence serotonergic activity. The first drug registered for the TRD with a rapid-acting mechanism is esketamine, the ketamine enantiomer, approved by the US Food and Drug Administration and the European Medicines Agency in the form of a nasal spray called Spravato® [66]. Esketamine is a non-competitive NMDA receptor antagonist. It selectively induces an antagonist effect in the event of excessive activation of NMDA that leads to an increased concentration of extrasynaptic glutamate and activation of neuroplasticity pathways [67]. The high effectiveness of esketamine in the treatment of TRD leads researchers to seek other candidates for RAADs such as psilocybin.

Psilocybin (3-[2-(dimethylamino)ethyl]-1H-indol-4-yl dihydrogen phosphate) is a natural substance which is a secondary metabolite found in the following genera: *Psilocybe*, *Copelandia*, *Pluteus*, *Gymnopilus*, *Pholiotina*, *Galerina*, *Inocybe* [68–73]. It is not biologically active and must be dephosphorylated to become psilocin, a psychoactive compound. Psilocin, as a classic psychedelic, has an agonist or partially agonist effect on 5-HT_{2A} receptors, which is a possible explanation of the hallucinogenic effect of this compound as these receptors are highly expressed in the visual cortex [74]. The following theory was confirmed by the administration of a selective 5-HT_{2A} antagonist, ketanserin, to humans, which attenuated the hallucinatory effect of psilocybin [75].

Administration of psilocybin induces down-regulation of 5-HT_{2A} receptors, that over-expression is observed in patients with major depression disorder [76,77]. Another theory postulates that downregulation of 5-HT_{2A} receptors may be influenced by increased synthesis of BDNF in the medial prefrontal cortex (mPFC) after administration of psilocybin. Elevated levels of BDNF can be explained by modulation of AMPA and NMDA receptors through the effect of psilocybin administration as an agonist to 5-HT_{2A} receptors [78]. Activation of this receptor has a positive influence on cerebral neuroplasticity by increasing BDNF synthesis and increasing c-FOS factor expression in the anterior cingulate cortex and mPFC—areas of the brain implicated in depression [79]. The secondary mechanism of antidepressant action is its anti-inflammatory effect of psilocybin by decreasing levels of TNF- α and IL-1 β , which was demonstrated in the human U937 macrophage cell line by administering psilocybin-containing mushrooms' water extract [80]. An increase in the level of pro-inflammatory cytokines, such as TNF- α , is one of the causes of the activation of the kynurenic pathway in microglia and the production of neurotoxic compounds, hydroxykynurenine and quinolinic acid [81].

The use of psychedelics in the treatment of MDD has gained scientific attention due to the relatively low therapeutic effectiveness of current psychopharmacological approaches and the increasing knowledge of the pathophysiology of depression. Recent meta-analysis of psychedelic therapy for depressive symptoms carried out by the Ko team revealed that the definitive clinical efficacy of the use of psychedelics such as psilocybin, LSD, or ayahuasca for depressive symptoms has not been demonstrated, partly due to the lack of a sufficient number of randomized clinical trials [82]. The largest clinical trial on the use of psilocybin in treatment-resistant depression was carried out by the Goodwin team with participants from ten countries in Europe and North America. The trial consisted of 233 participants, divided into three groups that were given a single dose of 25, 10, and 1 mg (control) of psilocybin. The change in value of the Montgomery-Åsberg Depression Rating Scale, the tool that is used to stratify the severity of depressive episodes in adults, from baseline to week 3 was the primary endpoint of the trial. Psilocybin at a single dose of 25 mg reduced depression scores significantly more than a 1-mg dose over a period of 3 weeks, but was associated with adverse effects such as headache, nausea, and dizziness [83].

Although there is no definitive verdict on the use of psychedelics in the treatment of Major Depressive Disorder, based on randomized clinical trials conducted with psilocybin, short- and long-term reductions in depressive symptoms have been observed [83].

Gotvaldová and her team performed a quantitative analysis of tryptamine derivatives in fruiting bodies of genera such as *Psilocybe*, *Pluteus*, and *Inocybe* [84]. The highest content of psilocybin in the analyzed species of *Psilocybe* genera was in *Pmexicana* (3.29–3.93 mg/g dw), *P. caerulipes* (2.23–5.67 mg/g dw), *P. cyanescens* (2.34–13.8 mg/d gw), and *P. serbica var. moravica* (5.65–14.16 mg/g dw). The highest concentration of psilocybin in *Pluteus* genera was found in *P. americanus* (1.17–2.43 mg/g dw). In *Inocybe* genera, the amount of psilocybin did not exceed 0.282 mg/g dw (*I. corydalina*) [84]. Psilocybin is not present in edible mushrooms; concentration of this compound in *Agaricus bisporus* was lower than the limit of detection [84].

Few biotechnological attempts have been made to increase the content of tryptamine derivatives in fruiting bodies or mycelia in mushroom species containing psilocybin [85,86]. The most interesting biotechnological method for obtaining psilocybin is the production of this compound, which was proposed by the Milne team using metabolically engineered yeast (*Saccharomyces cerevisiae*), whose productivity was determined at 120.3 mg/L of psilocybin. Due to further modification of the transformed metabolic pathway with *P. cubensis* cytochrome P450 reductase, it was possible to obtain a production of psilocybin of 627 mg/L, which allows a relatively cheap production of psilocybin on an industrial scale [85]. The highest production of psilocybin was observed in genetically modified *Escherichia coli* in which 1.16 g/L was observed through the biotransformation of 4-hydroxindole, serine, and methionine. However, the method would be difficult to implement in an industrial setting because of the high price of substrates [86].

Microdosing is the practice of repeatedly using low doses of psychedelics such as psilocybin. It is believed that the consumption of low doses of psilocybin (around 0.5 g per dose) can improve cognitive performance, stimulate creativity, and increase stamina. Microdoses do not induce hallucinations, contrary to the regular dose used for recreational use [87]. Possession or consumption of psychedelics such as LSD or psilocybin is illegal in most countries, but there is increasing evidence of the use of microdoses of psychedelics. What is the main motivation for the consumption of “magic mushrooms”? In the survey that collected the responses from 1116 respondents through an online questionnaire, it was found that the main motivations for microdosing psychedelics were performance improvement (37%), mood improvement (29%), and curiosity (15%) [88]. The consumption of microdoses of psychedelics is considered safe by those who decide to try it for the first time, but it showed that almost 20% of consumers experienced some acute psychological or physical negative effects [88]. There are more studies describing microdosing phenomena among humans, but due to their type (online surveys, observation, and open-label studies), the results are prone to confirmatory bias, as many lack a control group or are based on self-selected samples [88–90]. Some respondents reported a positive effect of microdosing on their cognitive abilities and creativity. However, it should be considered that in most cases, this may be due to the approach associated with the expectation of the positive effects of respondents and the researchers themselves conducting the observational study. A double-blind, placebo-controlled study of psilocybin microdosing carried out with 35 participants revealed no evidence to support enhanced cognitive or creative function. Low doses of psilocybin (0.5 g of dried fruiting bodies of *P. cubensis* two times a week, the total dose equals to 1.0 g) even resulted in small cognitive impairments [91].

So far, the results of clinical trials on the use of psilocybin in controlled clinical conditions give hope to patients suffering from TRD. In contrast to the antidepressants currently used, their effect appears several hours after the first dose of the preparation and lasts several days, which means that the total number of doses is lower than in the case of drugs administered every day, which can positively affect adherence to medical recommendations. More clinical trials are needed before psilocybin can be approved for the treatment of patients with TRD, especially those evaluating long-term antidepressant effects.

4. Anti-Inflammatory Activity of Medicinal Mushrooms in Beating Depression

The link between immune system, inflammation and depression was observed for the first time when IFN- α therapies, which activate inflammatory antiviral response, were introduced as a treatment for hepatitis C. Patients treated with interferon developed depression-like behaviors after 4 weeks of treatment initiation [92]. Patients with MDD have been observed to have higher levels of pro-inflammatory mediators such as IL-6, IL-12 and C-reactive protein compared to nondepressed individuals [93,94]. Patients with TRD are more likely to have elevated pro-inflammatory markers [95].

Peripheral inflammation can affect the central nervous system in many ways. It starts by having a negative impact on the permeability of the blood-brain barrier, which makes cytokines and immune cells more likely to cross to the brain [96]. The possible crossing of the proinflammatory cytokines to the brain may alter the kynurenic pathways, which are correlated with tryptophan availability.

The activity of enzymes involved in the kynurenic pathway can be modulated by glucocorticosteroids and/or pro-inflammatory cytokines. The hypothesis of depression induction caused by tryptophan depletion in the brain was first stated by Fuch et al. in 2002 [97]. However, research carried out by Dunn and Welch demonstrated that administration of LPS and/or proinflammatory cytokine IL-1 to mice increases brain tryptophan and serotonin concentration [98]. The O'Connor team conducted a similar observation that showed that LPS administered to rodents resulted in increased kynurenine content in the brains of mice and brain tryptophan and serotonin [99]. In human studies, patients treated with IFN- α showed that tryptophan concentrations in cerebrospinal fluid were stable despite a decrease in Trp blood level [100].

Based on the observations mentioned above, it is unlikely that depressive symptoms can be caused or worsened by tryptophan depletion in the brain by shifting tryptophan to the kynurenic pathway. An alternate hypothesis is that alterations in concentrations of products of the kynurenic pathway may play a role in the development of depression [81,101].

Kynurenine can be metabolized in several ways, depending on the cell type in which kynurenine is produced, transported, and metabolized. In microglia, kynurenine is broken down to 3-hydroxykynurenine and quinolinic acid, which are neurotoxic. Neurotoxicity of these compounds is caused by the generation of reactive oxygen species that may damage neural cells and act as agonists in the NMDA receptor [102]. Reactive oxygen species can promote the production of proinflammatory cytokines via NF- κ B pathway [103]. In astrocytes, kynurenine is degraded to kynurenic acid known for its neuroprotective activity by acting as an antagonist of the NMDA and α -7 nicotinic acetylcholine receptor [104]. The intact neuron can metabolize kynurenine to picolinic acid, which is also neuroprotective.

In patients with depression, variations in the levels of the products of the kynurenic pathway have been observed. In a meta-analysis carried out by Ogyu et al. it was observed that in depressed patients decreased level of kynurenine and kynurenic acid was observed whereas depression free patients were observed with higher level of quinolinic acid [105].

Neuroinflammation may be beneficial because activation of microglia is necessary to eliminate the threat in the form of infection, injury, or toxic metabolites [102]. Although chronic neuroinflammation can lead to overproduction of pro-inflammatory cytokines and production of neurotoxins (products of the kynurenic pathway described above) that can lead to neuronal death and, consequently, loss of neuronal volume in areas responsible for mood regulation such as the PFC or the hippocampus. Additionally, neuroinflammation may be an important part of the pattern of neurodegenerative diseases such as Alzheimer's and Parkinson's disease [106]. In vitro and in vivo studies showing the anti-inflammatory activity are presented in Table 3.

Table 3. Anti-inflammatory activity of selected medicinal/edible mushroom species.

Species	Active Substance	Experience Model	Result	Ref.
<i>Lentinula edodes</i>	β -glucan	In vivo: C57BL/6J mice Inflammation: High-fat diet-induced (HFD)	<p>↓Neuroinflammation in hippocampus and prefrontal cortex</p> <ul style="list-style-type: none"> • ↓TNF-α, ↓IL-6, ↓IL-1B in hippocampus) • ↓Microglia proliferation and ↓TNF-α, ↓IL-6, ↓IL-1B in PFC 	[107]
		Lentinan In vitro: mice microglia BV-2 cell line Inflammation: LPS induced	<p>↓Microglia inflammation</p> <ul style="list-style-type: none"> • ↓TNF-a, ↓IL-1B • ↑IL-10, ↑BDNF <p>Promoted conversion of microglial phenotype from M1 (proinflammatory) to M2 (inflammatory resolving) status</p>	[108]
<i>Inonotus obliquus</i>	Lanostane triterpenes: Inonotusols H-N	In vitro: mice microglia BV-2 cell line Inflammation: LPS induced	<p>↓Microglia inflammation</p> <ul style="list-style-type: none"> • ↓NO generation • ↓iNOS expression inhibition (the strongest interaction with inonotusols I) 	[109]
<i>Cyathus africanus</i>	Cyanthane diterpenoids	In vitro: mice microglia BV-2 cell line, PC-12 cell line Inflammation: LPS induced	<p>↓Microglia inflammation</p> <ul style="list-style-type: none"> • ↓NO generation in BV-2 cell line • ↑Neurotrophic activity through NGF • ↑Neurite outgrowth in PC-12 cell line 	[110]
<i>Auricularia polytricha</i>	Ethanol and hexane extracts	In vitro: mice microglia BV-2 cell line Inflammation: BPA induced	<p>↓Neuroinflammation by regulation of NG-kB signaling pathway</p> <p>↑Antioxidant activity</p> <ul style="list-style-type: none"> • ↑SOD-1 enzyme activity 	[111]
<i>Sanguinoderma rugosum</i>	Mycelial extracts	In vitro: mice hippocampal neuronal HT-22 cell line Inflammation: glutamate induced	<p>↑Cell viability in pretreated with mycelial extract cell line after glutamate inflammation induction</p> <p>The main constituents responsible for the action were: linoleic acid, ergosterol and ethyl linoleate (GC-MS analysis)</p>	[112]

Table 3. Cont.

Species	Active Substance	Experience Model	Result	Ref.
<i>Hericium erinaceus</i>	Erinacine A (EA)/ <i>H. erinaceus</i> extract (HEM)	In vivo: male Sprague-Dawley rats Inflammation: LPS injection intranigrally In vitro: mice microglia BV-2 cell line, CTX TNA2 cell line Inflammation: LPS, IFN- γ induced	In vivo: Both, EA and HEM showed neuroprotective effect on motor dysfunction in rats with LPS-induced neuronal damage. \downarrow Neuroinflammation <ul style="list-style-type: none"> \downarrowTNF-α, \downarrowIL-1β, \downarrowiNOS gene expression BDNF levels did not differ from control and experimental groups In vitro: \downarrow Microglia inflammation <ul style="list-style-type: none"> \downarrowNO generation in BV-2 cell line after EA pretreatment \downarrowiNOS expression in BV-2 cell line after EA pretreatment \downarrow Astrocytes inflammation <ul style="list-style-type: none"> \downarrowTNF-α expression in CTX TNA2 cell line after EA pretreatment The expression levels of IL-1 β and iNOS at the same level, regardless whether pretreated with EA or not	[113]
<i>Grifola frondosa</i>	o-Orsellinaldehyde	In vitro: primary microglia and astrocytes from mice brain Inflammation: LPS induced	\downarrow Neuroinflammation <ul style="list-style-type: none"> \downarrowNO generation \downarrowiNOS, \downarrowHO-1 expression \downarrowNF-κB pathway activation in microglia Promoted conversion of microglial phenotype from M1 (proinflammatory) to M2 (inflammatory resolving) status	[114]
-	Ergothioneine	In vitro: hCMEC/D3 human brain endothelial cell line Inflammation: 7-ketocholesterol induced	\downarrow Neuroinflammation \downarrow IL-1 β , \downarrow IL-6, \downarrow IL-8, \downarrow TNF- α , \downarrow COX-2 mRNA expression after ergothioneine addition	[115]
<i>Cordyceps militaris</i>	<i>C. militaris</i> powdered fruiting bodies	In vivo: AD mice Inflammation: intracerebroventricular injection of A β ₁₋₄₂	\downarrow Neuroinflammation <ul style="list-style-type: none"> \downarrowiNOS, \downarrowCOX-2 gene expression \uparrow Neuronal plasticity <ul style="list-style-type: none"> \uparrowBDNF expression 	[116]
<i>Lignosus rhinocerotis</i>	Lipid rich fraction of the sclerotium	In vitro: mice microglia BV-2 cell line Inflammation: LPS induced	\downarrow Neuroinflammation <ul style="list-style-type: none"> \downarrowNO production \downarrowiNOS, \downarrowCOX-2 gene expression \uparrowHO-1, \uparrowNQO-1, \uparrowNrf2 gene expression 	[117]

Table 3. Cont.

Species	Active Substance	Experience Model	Result	Ref.
	Hot aqueous extract (HAE), an ethanol extract (EE), fractions from the HAE and EE, crude polysaccharides	In vitro: mice microglia BV-2 cell line Inflammation: LPS induced	↓Neuroinflammation 500 µg/mL of HAE → ↓NO production by 88.95% * 250 µg/mL of an n-butanol fraction → ↓NO production by 86.5% * 250 µg/mL of an ethyl acetate fraction of HAE → ↓NO production by 85.93% * * Compared to control group	[118]
<i>Coriolus versicolor</i>	<i>C. versicolor</i> biomass (mycelium and primordia)	Clinical trial: 40 patients with Meniere's disease (MD)	↓Oxidation stress in group treated with <i>Coriolus</i> supplement ↑Hsp70, ↑HO-1 proteins level in lymphocytes and plasma ↑γ-GC liase activity ↑GSH/GSSG ratio in patients treated with <i>Coriolus</i> ↑GSH level in patient treated group	[119]
	<i>C. versicolor</i> biomass (mycelium and primordia)	In vivo: male Sprague–Dawley rats Inflammation: no induction	↑Neuroprotection ↑LXA4 level in rat brain, especially in cortex and hippocampus ↑Hsp72, ↑OH-1, ↑thioredoxin proteins level	[120]
<i>Ganoderma lucidum</i>	Deacetyl ganoderic acid F (DeGA F)	In vivo: zebrafish and C57BL/6J mice Inflammation: LPS induced In vitro: mice microglia BV-2 cell line Inflammation: LPS induced	In vivo: ↓Inflammation in zebrafish • ↓NO production ↓Neuroinflammation in mice • ↓TNF-α, ↓IL-6, ↓iNOS, ↓p-Akt, ↓p-IKKα/protein levels DeGA F attenuated LPS-induced cell morphology changes In vitro: ↓Neuroinflammation through NF-κB signaling pathway ↓p65 nuclear protein level and migration reduction ↓p-Akt, ↓p-IKKα/β, ↓p-IκBα expression	[121]
<i>Armillaria mellea</i>	Fr.2 fraction (5-hydroxymethylfurfural, vanillic acid, syringate)	In vitro: mice microglia BV-2 cell line Inflammation: LPS induced	↓Neuroinflammation • ↓NO, ↓IL-6, ↓IL-1β, ↓TNF-α in a dose depended manner • ↓phosphorylation level of NF-κB p65, IκB-α and JNKs pathway	[122]

↓—decrease, ↑—increase.

Research on potential anti-neuroinflammatory activity is based on activity analysis of TLR4 and NF-κB pathways in cells in order to observe whether the addition of investigated mushroom species extract/isolated substance exhibits this activity.

Toll-like receptors (TLRs) are a group of transmembrane receptors responsible for the recognition of pathogen-associated molecular patterns (PAMPs) or danger-associated

molecular patterns (DAMPs). The activation of TLR4 receptor by bacterial lipopolysaccharide (LPS), viral proteins and polysaccharides results in the production of inflammatory substances, which are essential in order to produce effective immune response [123]. Activation of TLR4 leads to activation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B). NF- κ B is one of the most important and versatile family of transcriptional factors which are associated with inflammation and immunity [124]. After stimulation, NF- κ B factors complex are being freed and in free form (p50 and p65) translocate to nucleus which activate transcription and liberation of pro-inflammatory mediator, such as inducible nitric oxide synthase (iNOS) and NO, prostaglandin E2 (PGE2) and cyclooxygenase-2 (COX-2) and proinflammatory cytokines such as IFN- γ , IL-1 β , IL-6, and TNF- α [124].

5. Medicinal Mushrooms and Their Impact on Neurotrophins—Neurotrophin and Neurogenesis-Based Depression

The neurogenesis hypothesis of depression postulates that depressive behaviors can be attributed to insufficient or altered production and maturation of new neurons—adult neurogenesis [125,126]. The reduced volume of the hippocampus and mPFC is one of the most described neural abnormalities in depressed patients. However, it is still not concluded whether depression is a cause of neuronal atrophy or whether neuronal atrophy is a cause of depression. Conclusions from observation of hippocampus and mPFC volumes in first-onset MDD are not conclusive [89]. A meta-analysis revealed that a reduction in hippocampal volume is observed in patients with a history of MDD of at least two years [127]. Currently, there are three different approaches according to the cause versus consequence debate on the reduction of hippocampal and mPFC volume in depressed patients [128]. The causative one postulates that reduced mPFC and hippocampal-volume cause initial depression, which is supported by human studies that showed that patients with reduced hippocampal and mPFC volume have a higher chance of onset of MDD [129,130]. In the opposite approach, the reduced volume of the hippocampus and mPFC is caused by multiple episodes of depression—the hypothesis is supported by longitudinal studies that showed that the decreased volume of structures is more obvious when MDD does not relapse [131,132]. The third approach is that reduced volumes of mPFC and hippocampus do not have to be related to MDD itself, as there are some studies showing reduced volumes of these structures in healthy subjects exposed to chronic stress in life [133].

The proteins involved in the regulation of cell proliferation, maintaining synaptic plasticity, and neural functions are neurotrophins, such as NGF, NT-3 and NT-4 [134]. Brain-derived neurotrophic factors are the most studied member of the NGF family. In general, neurotrophins are synthesized as proneurotrophins that are processed intra- or extracellularly to be secreted in a mature and biologically active form. Proneurotrophins react with the p75 neurotrophin receptor (p75^{NTR}) and thus mediate neuronal death, leading to decreased synaptic plasticity, while mature neurotrophins bind to a particular tyrosine kinase receptor (Trk), leading to promotion of survival and differentiation by increasing the branching of axons and dendrites [135–139]. Clinical evidence supporting the neurotrophic hypothesis of depression is based on postmortem studies that demonstrated that BDNF levels are decreased in the cerebral cortex of depressed and suicide subjects [140,141]. Studies confirmed that in depressed patients, a reduced volume of the PFC and hippocampus was observed, which can be explained by a decrease in the signaling of BDNF-TrkB [142–144]. It was suggested, in a very simplified way, that an increased level of mature BDNF through TrkB signaling may produce an ‘antidepressant’ response, while pro-BDNF through p75^{NTR} signaling may exhibit the opposite effect [145].

There is research on the impact of the extract from the fruiting bodies or mycelia or isolated substances from medicinal/edible mushroom species on the survival of neuron cells in in vitro and in vivo models and analyses of the influence on neurotrophin expression—BDNF, particularly.

An antidepressant activity of polysaccharide–peptide (PGL) isolated from *Ganoderma lucidum* spores was measured in mice. PGL reversed depression behaviors in mice after acute and chronic administration. The administration of isolated PGL resulted in upregulation of BDNF/TrkB expression in the prefrontal cortex [146].

Hericium erinaceus (Lion's mane) is one of the most investigated medicinal mushroom species in terms of antidepressant activity. The popularity of this species is particularly caused by the presence of erinacines, a group of biologically active substances that is a stimulator of NGF [147].

Erinacines are chemical components that can be classified as cyathin diterpenoids that are found in the mycelium of *H. erinaceus* but not in fruiting bodies. To date, 15 erinacines have been isolated and described—the neuroprotective effect shows erinacine A–I [148]. In a study in rats with induced ischemic brain injury, administration of mycelium or isolated erinacine A resulted in inhibition of neuronal cell death. Erinacine A acts as a reactive oxygen species scavenger and inhibitor of the iNOS/p38 MAPK and CHOP proteins, which protect neurons from death caused by ischemic injury. In addition, the increase in pro-inflammatory cytokine levels observed in the control group after ischemic injury was reversed after administration of erinacine A in a dose-dependent manner [149]. In another study, a standardized aqueous extract of *H. erinaceus* was administered to male C57BL/6 mice subjected to chronic restraint stress for 4 weeks. Upregulation of mRNA and increased expression of proteins related to neurogenesis such as BDNF, doublecortin, nestin, synaptophysin, tropomyosin receptor kinase B (TrkB) were observed in mice fed *H. erinaceus* extract. Furthermore, bromodeoxyuridine positive cells were observed in the hippocampus, indicating enhanced neurogenesis [150]. Neurotrophic group of substances isolated from the fruiting bodies of *H. erinaceus*, isoindolinones—hericerin, isohericerinol A, N-de-phenylethyl isohericerin and corallocin A. Increased NGF production in C6 glioma cells was observed. Increased expression of NGF, BDNF, and synaptophysin (SYP) was observed in the C6-N2a cell line [151].

Biological activity and safety data obtained in the course of many in vitro and in vivo studies of *H. erinaceus* led to the conducting of a few clinical trials in humans. The first randomized, double-blind, placebo-controlled clinical trial was conducted in a small group of 30 women, who in the experimental group ingested powdered fruiting bodies of *H. erinaceus* in the form of cookies. Based on the result comparison of the Center for Epidemiologic Studies Depression Scale (CES-D) and the Indefinite Complaints Index (ICI), it can be concluded that *H. erinaceus* ingestion for at least 4 weeks (0.5 g of powdered fruiting bodies) may result in reduced depression-like symptoms and anxiety [152]. In a more recent clinical trial, whose objective was to investigate the effect of *H. erinaceus* on hearing degeneration in elderly patients ($n = 80$), it was proved that administration of *H. erinaceus* mycelium could reduce hearing loss, especially for high frequencies and speech recognition. *H. erinaceus* administration promoted the concentrations of BDNF and NGF in patients over 65 years old better than in younger patients [153].

Another species of the *Hericium* genera, *Hericium coralloides*, is a source of the benzofuranone and isoindolinone structure groups—corrallocins A–C. The greatest impact on the stimulation of neurite outgrowth from PC12 cells was observed in the case of the addition of corrallocin C to the culture medium—the same compound was shown to be the NGF—inducer in the 1321N1 astrocyte cell line. The expression of BDNF mRNA was evaluated in 1321N1 astrocytes after stimulation with corrallocins A–C. The highest level of BDNF mRNA was observed after corrallocin C stimulation [154].

2 α -hydroxy-inotodiol (2 α -HI)—a lanostane tripterpenoid isolated from *Inonotus obliquus* exhibited the most remarkable neuroprotective activity among 10 isolated structures from this species. Neuroprotective activity was measured in vitro in H₂O₂-induced SH-SY5Y cells. 2 α -HI showed neuroprotective effects through activation of Nrg2 and BDNF/TrkB/ERK/CREB pathway. The result of the in vitro study was confirmed in vivo in zebrafish [155].

The 28 kDa polysaccharide peptide isolated from *Ganoderma lucidum* spores showed antidepressant activity in mouse model of depression induced by unpredictable chronic

mild stress. The antidepressant mechanism of action was the upregulation of BDNF in the PFC [146].

Polyoxygenated cyanthane diterpenoids, in addition to their anti-neuroinflammatory activity, showed neurogenesis inducing activity by NGF-induced neurite growth activity in PC-12 cells after the addition of isolated diterpenoids in all concentrations tested [110].

β -glucans depending on their chemical structure, may have different effects on cognition. The difference in biological activity of three β -glucans from mushroom (β -(1,3)/(1,6)-glucan isolated from *L. edodes*), curdlan (β -(1,3)-glucan) and oat bran (β -(1,3)/(1,4)-glucan) was evaluated in animal model (C57BL/6J mice) [156]. All three glucans had a positive effect on temporal order recognition memory. The expression of BDNF and postsynaptic protein 95 increased in the PFC. In addition, only in supplementation with mushroom β -glucan, post-synaptic thickness of synaptic ultrastructure was observed [156]. Lentinan, an β -(1,3)/(1,6)-glucan polysaccharide found in *L. edodes* inhibited neuroinflammation and enhanced remyelination in the cuprizone (CPZ) mouse model [108]. Lentinan reversed neuronal injury as well as motor dysfunctions through dectin-1 receptor. It shows that lentinan could be a novel therapeutic agent that can reduce demyelination, which is present in multiple sclerosis [108].

6. Medicinal Mushroom and the Brain–Gut Axis

The human gut microbiota has become the subject of intensive research in recent years. The brain–gut axis refers to the bilateral link between the intestinal microbiome and the central nervous system. The gut microbiota plays an important role in the processing of carbohydrates, the production of some vitamins (B group vitamins and vitamin K), or the synthesis of short-chain fatty acids (SCFAs).

The gut microbiota also plays a crucial role in modulating the pharmacokinetic parameters of some drugs, including those that act on the CNS. The presence of *Helicobacter pylori* can induce a reduction in L-dopa absorption, which can have a negative impact on the time of onset, duration, and quality of life parameters in patients with Alzheimer's disease [157]. The presence of *Clostridium leptum* in the gastrointestinal tract of humans may affect nitrazepam metabolism due to the presence of the nitroreductase enzyme. As a result, 7-aminonitrazepam is produced, which is known for its toxic activity [158]. Similar observations were made in the biotransformation of clonazepam to 7-aminoclonazepam, leading to a higher risk of toxicity [159]. In another study, the presence of *Clostridium sporogenes* and the reductase activity found in this strain altered the metabolism of zonisamide—an anticonvulsant leading to increased drug activity [160].

In addition to the possible interaction between the gut microbiota and drugs that can result in suboptimal drug activity, altered gut microbiota composition may be one of the pathomechanisms that explain some neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease, multiple sclerosis, MDD, or diseases with the predominant symptom of anxiety [161]. The connection between the gut microbiota and depression may be attributed to an inflammation-based theory: alteration of the gut microbiota in a direction favorable growth of pro-inflammatory taxa such as *Enterobacteriaceae*, *Eggerthella*, *Desulfovibrio* may contribute to the initiation of inflammation [162–164]. Increased pro-inflammatory mediators can interfere with the blood-brain barrier and cause inflammation, degradation in the CNS, and penetration of immune cells into the CNS [165]. Interestingly, there are some bacterial taxa that produce anti-inflammatory agents, such as SCFAs. Example taxa are *Faecalibacterium*, *Coprococcus*, *Clostridium XIVa* species [166,167], which can produce butyrate, and *Megamonas*, which can produce acetate and propionate [168]. In the animal model, transplanting feces from depressed patients into mice can produce depression-like behaviors [169]. The cause of depression like behavior development may be attributed to inflammation-based theory of depression as hippocampal levels of IFN- γ and TNF- α were significantly increased in mice which received transplant [170].

In one study, the composition of the fecal microbiome in MDD patients differed significantly increased *Bacteroidetes*, *Proteobacteria*, *Actinobacteria* and fewer *Firmicutes* were

observed compared to healthy individuals [171]. The systematic review on the gut microbiota in anxiety and depression pointed out that in patients with depressive disorders, higher pro-inflammatory species and a lower abundance of taxa producing SCFAs are observed [172].

Changes in diet can produce a rapid effect on the human gut microbiota [173]. For example, the ingestion of omega-3 polyunsaturated fatty acid dietary supplements can positively impact the growth of butyrate-producing bacteria in healthy middle-aged volunteers [174]. The depletion of dietary fiber in the diet of mice resulted in intestinal barrier dysfunction by promoting the growth of bacteria that degrade colonic mucus [175]. Based on research on the link between food and changes in the gut microbiota, the common sentence “You are what you eat” takes on a new meaning.

Dietary prebiotics, according to the terminology established during the 6th meeting of the International Scientific Association of Probiotics and Prebiotics in 2008, can be defined as “a selectively fermented ingredient that results in specific changes in the composition and/or activity of the gastrointestinal microbiota, thus conferring benefit(s) on host health” [176]. Edible mushrooms are a source of polysaccharides, naturally occurring substances that exhibit multidirectional biological activity and can be classified as prebiotics. Their activity is affected by its chemical composition such as monosaccharide composition, molecular weight, and the type of glycosidic bond [177].

Edible mushrooms are considered as a potential source of prebiotics due to the presence of polysaccharides such as α -, β -glucans, chitin that meet the conditions of being prebiotics: they are resistant to conditions of the upper intestinal tract (acidity), stable to food processing and fermented by the gut microbiota, which leads to the production of substrates needed for efficient growth of the gut microbiota. Current knowledge about the influence of edible mushrooms on the microbiota and gut health is summarized in Table 4.

Table 4. Selected medicinal mushroom species effect on gut microbiota.

Medicinal Mushroom Species	Active Ingredient	Experimental Model	Gut Microbiota Alteration in Treated Group	Biological Activity	Ref.
<i>Ganoderma lucidum</i>	Ethanollic extract	In vivo: C57BL6/J mice Dextran sulphate sodium administration (DSS)-induced colitis	↑ <i>Turicibacter</i> ↑ <i>Bifidobacterium</i> ↑ <i>Parabacteroides</i> abundance ↓ <i>Escherichia/Shigella</i> ↓ <i>Bacteroides</i> ↓ <i>Staphylococcus</i> abundance	Restoration of the intestinal barrier • ↑MUC2, ↑ZO-1, ↑claudin-3, and ↑occludin in DDS-induced colitis group with supplementation of ethanollic extract. Reduction of local bowel inflammation • ↓iNOS, ↓COX-2 activity	[178]
	Ethanollic extract of <i>G. lucidum</i> mycelium (GLAA)	In vivo: ICR mice, SPF Gut microbiota depleted by antibiotic	↑ <i>Bifidobacteriaceae</i> <i>Bifidobacterium</i> ↑ <i>Lactobacillaceae</i> <i>Lactobacillus (L. reuteri)</i> ↑ <i>Porphyromonadaceae</i> <i>Odoribacter</i> ↑ <i>Erysipelotrichales</i> <i>Turicibacter</i> ↓ <i>Ruminococcaceae</i> <i>Oscillibacter</i> ↓ <i>Anaerotignum</i> ↓ <i>Roseburia</i>	Addition of GLAA reduced required time for sleep induction by 50% in pentobarbital-induced hypnosis model ↑5-HT in the hypothalamus in treated with GLAA group (50 and 100 mg/kg)	[179]

Table 4. Cont.

Medicinal Mushroom Species	Active Ingredient	Experimental Model	Gut Microbiota Alteration in Treated Group	Biological Activity	Ref.
	Hydroalcoholic extracts	In vivo: C57BL/6 mice High-cholesterol diet (HCD)	Reversed gut dysbiosis caused by HCD ↑ <i>Lactobacillus</i> ↓ <i>Bacteroides acidifaciens</i> ↓ <i>Mucispirillum schaedleri</i> ↓ <i>Parabacteroides distasonis</i>	<i>G. lucidum</i> extracts revealed to be a novel transcriptome modulator—prevention of metabolic disorders associated with hypercholesterolemia	[180]
	Polysaccharides from hot water extraction	In vivo: Sprague-Dawley rats (SPF) HFD and streptozotocin-induced type 2 diabetes mellitus	↑ <i>Blautia</i> ↑ <i>Dehalobacterium</i> ↑ <i>Parabacteroides</i> ↑ <i>Bacteroides</i> ↓ <i>Aerococcus</i> ↓ <i>Ruminococcus</i> ↓ <i>Corynebacterium</i> ↓ <i>Proteus</i>	Administration of mushroom polysaccharides showed antidiabetic effect	[181]
<i>Antrodia cinnamomea</i>	Exopolysaccharides	In vivo: ICR mice Antibiotics induced gut dysbiosis (lincomycin)	↑ <i>Lactobacillus</i> ↑ <i>Roseburia</i> ↑ <i>Ligilactobacillus</i> ↑ <i>Lachnospiraceae_NK4A136</i> group abundance ↓ <i>Enterococcus Shigella</i> abundance	Reduced peripheral inflammation ↓IL-6 and ↓TNF- α serum levels compared to control group	[182]
	Isolated polysaccharides	In vivo: Lingnan yellow-heathered female chickens LPS induced liver inflammation	Restrained the decline of beneficial cecal microbiota (typically <i>Lactobacillus</i> , <i>Faecalibacterium</i> , and <i>Christensenellaceae R-7</i> group)	Hepatoprotective effect—reversed LPS-induced liver inflammation ↓TLR4/NF- κ B signaling pathway expression in the liver tissues	[183]
<i>Morchella esculenta</i>	Isolated polysaccharides	In vivo: BALB/c mice (SPF) HFD	↑ <i>Lactobacillus</i> ↓ <i>Enterococcus</i>	Improved intestinal barrier • ↑TJs proteins ↓Inflammation in bowel • ↓iNOS, ↓COX-2, ↓TNF- α , and ↓IL-6 overexpression (immunoblotting) • ↓TLR4 inflammatory related signaling pathways	[184]
<i>Lyophyllum decastes</i>	Isolated polysaccharides	In vivo: C57BL/7J mice HFD	↓ <i>Firmicutes/Bacteroidetes</i> ratio ↑ <i>Lactobacillus johnsonii</i> <i>Bacteroides intestinalis</i>	Antiobesity effect of extracted polysaccharides due to microbiota alteration and secondary bile-acids production activation	[185]

Table 4. Cont.

Medicinal Mushroom Species	Active Ingredient	Experimental Model	Gut Microbiota Alteration in Treated Group	Biological Activity	Ref.
<i>Wolfiporia cocos</i>	Including water-soluble polysaccharides (PCX), water-insoluble polysaccharides (PCY), and triterpenoid saponins (PCZ)	In vivo: healthy Kunming mice	The strongest altering gut microbiota activity was found in PCY fraction: ↑ <i>Lactobacillus</i> In mice fed with PCX: ↑ <i>Deferribacterota</i>	PCX: ↑IL-10 levels in the liver, spleen tissues and serum, PCY: ↓IFN-γ level in the liver, PCZ: ↓TNF-α level in liver and spleen PCX and PCZ: ↑IFN-γ	[186]
	Triterpenoid fraction	In vivo: Spague–Daley rats Unpredictable mild stress model (CUMS)	Restoration of altered gut microbiota by CUMS: ↓ <i>Firmicutes/Bacteroidetes</i> ratio	↑BDNF, ↑NGF in the hippocampus	[187]
<i>Auricularia polytricha</i>	Freeze dried ethanolic extracts of fruiting bodies	In vivo: ICR mice (SPF)	↑ <i>Ruminococcaceae</i> ↑ <i>Lachnospiraceae</i> ↑ <i>Prevotellaceae</i>	Anti-inflammatory activity: ↓F-κB and MAPK/ERK1/2 signaling pathway	[111]
<i>Flammulina velutipes</i>	Freeze dried ethanolic extracts of fruiting bodies	DSS-induced intestinal dysbiosis	↑ <i>Bifidobacteriaceae</i> ↑ <i>Erysipelotrichaceae</i>	↑Keap1/Nrf2 signaling pathways	
<i>Pleurotus citrinopileatus</i>	Isolated Polysaccharide–peptide I (PSI), Polysaccharide–peptide II (PSII)	In vitro batch culture: fecal samples from healthy human volunteers on YCFA medium	PSI: ↓ <i>Escherichia-Schigella</i> PSII: ↑ <i>Bifidobacterium</i> ↑ <i>Lactococcus</i> ↑ <i>Lactobacillus</i> ↑ <i>Desulfovibrionaceae</i> ↑ <i>Lachnospiraceae</i> ↑ <i>Odoribacter</i> ↑ <i>Coriobacteriaceae</i> ↑ <i>Blautia</i> ↓ <i>Escherichia-Schigella</i>	Administration of PSI resulted in higher production of SCFAs	[188]
<i>Pleurotus eryngii</i>	Water soluble polysaccharide fraction	In vivo: C57BL/6J mice HFD	↑ <i>Anaerostipes</i> ↑ <i>Clostridium</i> ↑ <i>Lactococcus</i> ↓ <i>Roseburia</i> ↓ <i>Lactobacillus</i>		[189]
	Powdered fruiting bodies	In vivo: CD-1 mice DSS-induced colitis	↑ <i>Odoribacteraceae</i> ↑ <i>Adlercreutzia</i> ↑ <i>Akkermanisa</i> ↑ <i>Lactobacillus</i> ↑ <i>Anaerostipes</i> ↑ <i>Allobaculum</i> ↓ <i>Actinobacteria</i> ↓ <i>Mollicutes</i> ↓ <i>Desulfovibrionaceae</i> ↓ <i>Enterococcaceae</i> ↓ <i>Turicibacter</i> ↓ <i>Dorea</i> ↓ <i>Bacteroides</i> ↓ <i>Prevotella</i>	↑Production of SFCAs in dose-depended manner ↓Local—colon inflammation • ↓IL-1β, ↓IL-17 No effect on IL-2, IL-17A, IFN-γ,	[190]

Table 4. Cont.

Medicinal Mushroom Species	Active Ingredient	Experimental Model	Gut Microbiota Alteration in Treated Group	Biological Activity	Ref.
<i>Pleurotus ostreatus</i>	Powdered fruiting bodies	In vivo: C57BL/6J mice HFD-induced obesity	↑ <i>Oscillospira</i> ↑ <i>Lactobacillus</i> group ↑ <i>Bifidobacterium</i> ↑ <i>Anaerostipes</i> ↑ <i>Anaerovorax</i> ↑ <i>Anaerofustis</i> ↑ <i>Ruminococcus</i> ↑ <i>Coprococcus</i> ↓ <i>Bacteroides</i> ↓ <i>Roseburia</i> ↓ <i>Acinetobacter</i> ↓ <i>Agrobacterium</i> ↓ <i>Microbacterium</i> ↓ <i>Novosphingobium</i> ↓ <i>Streptococcus</i> ↓ <i>Prevotella</i> ↓ <i>Sphingomonas</i> ↓ <i>Macrocooccus</i> ↓ <i>Lactococcus</i> Compared to HFD only		[191]
	Powdered fruiting bodies	In vivo: piglets (crossbred: Duroc × Large white × Landrace)	↑ <i>Prevotella</i> ↑ <i>Anaerovibrio</i> ↑ <i>Veillonellaceae</i>	↑Production of SFCAs in group with supplementation of <i>P. ostreatus</i> ,	[192]
<i>Pleurotus sajor-caju</i>	Powdered mycelium	In vivo: Zucker rats	↑ <i>Faecalibaculum</i> ↑ <i>Bifidobacterium</i> ↑ <i>Roseburia</i> ↑ <i>Blautia</i> ↓ <i>Escherichia-Shigella</i>	↑Production of SCFAs in the colon	[193]
<i>Agaricus bisporus</i>	Roasted fruiting bodies	Open label crossover study in healthy adults ($n = 32$)	↑ <i>Bacteroides</i> ↑ <i>Parabacteroides</i> ↑ <i>Coprococcus</i> ↑ <i>Sutterella</i> ↑ <i>Anaerostipes</i> Comparing to diet with meat		[194]
<i>Agaricus bisporus</i> <i>Agaricus bisporus brunnescens</i>	Powdered fruiting bodies	In vivo: C57BL/J mice	Stable Firmicutes/Bacteroidetes ratio ↑ <i>Verrucomicrobia</i> ↑ <i>Akkermansiaceae</i> ↑ <i>Tannerellaceae</i> ↓ <i>Prevotellaceae</i>	↓Inflammation related genes levels such as IL-6, Nox-2 (Portobello only), Hmox-1	[195]

Table 4. Cont.

Medicinal Mushroom Species	Active Ingredient	Experimental Model	Gut Microbiota Alteration in Treated Group	Biological Activity	Ref.
<i>Armillaria mellea</i>	<i>A. mellea</i> fermentation liquor	In vivo: Sprague–Dawley rats Insomnia model by the p-chlorophenylalanine (PCPA) induction	<ul style="list-style-type: none"> ↑<i>Lachnospiraceae</i> NK4A136 group ↑<i>Lachnospiraceae</i> the [<i>Eubacterium</i>] <i>xylanophilum</i> group ↑<i>Ruminococcus</i> ↑<i>Candidatus</i> ↑<i>Saccharimonas</i> ↑the [<i>Eubacterium</i>] <i>coprostanoligenes</i> group ↓<i>Lactobacillus</i> ↓<i>Maribaculum</i> ↓<i>Prevotellaceae</i> UCG-001 	<p>Administration of AFL resulted in ↑5-HT_{1A} and 5-HT_{2A} proteins in hippocampus comparing to model group</p> <p>↓Peripheral inflammation reduced after AFL administration in dose-dependent manner</p> <ul style="list-style-type: none"> • ↓IL-6, ↓TNF-α, ↓IL-1β 	[196]
	Isolated polysaccharide—lentinan	In vivo: C57BL/6J mice, HFD	<ul style="list-style-type: none"> ↑<i>Actinobacteria</i> ↑<i>Firmicutes</i> ↑<i>Bifidobacterium</i> ↓<i>Proteobacteria</i> ↓<i>Epsilonbacteraeota</i> 	<p>Addition of lentinan to HD diet resulted in improved intestinal barrier</p> <ul style="list-style-type: none"> • ↑TJs proteins (occludin, ZO-1) at mRNA and protein level <p>Improved glucose tolerance</p>	[197]
<i>Lentinula edodes</i>	Isolated β-glucan	In vivo: C57BL/6J mice HFD	<ul style="list-style-type: none"> ↑<i>Clostridiales</i> ↑<i>Lachnospiraceae</i> ↑<i>Ruminococcaceae</i> 	<p>Addition of lentinan to HFD resulted in improved intestinal barrier</p> <ul style="list-style-type: none"> • ↑TJs proteins (occludin) at protein level <p>↓Serum LPS → ↓peripheral inflammation</p>	[107]
	Polysaccharides	In vivo: C57BL/6J male mice DSS-induced colitis	<p>In DDS-induced group:</p> <ul style="list-style-type: none"> ↑<i>Bacteroides</i> ↑<i>Helicobacter</i> ↑<i>Parasutterella</i> ↓<i>Firmicutes</i> <p>Addition of FMP to diet reversed negative gut microbiota alteration</p>	<p>Improved intestinal barrier after mushroom addition to diet:</p> <ul style="list-style-type: none"> • ↑TJs proteins (occludin, ZO-1) level <p>Local inflammation in colon</p> <ul style="list-style-type: none"> • ↓TNF-α, ↓IL-1β, ↓IL-6 • ↑Production of SCFAs 	[198]
<i>Hericium erinaceus</i>	Powderized <i>H. erinaceus</i> fruiting bodies	In vivo: 11-years old dogs	<ul style="list-style-type: none"> ↑<i>Bacteroidetes</i> ↑<i>Bacteroidales</i> ↓<i>Firmicutes</i> ↓<i>Streptococcus</i> ↓<i>Tyzzereella</i> ↓<i>Campylobacteraceae</i> ↓<i>Campylocaber</i> 		[199]

Table 4. Cont.

Medicinal Mushroom Species	Active Ingredient	Experimental Model	Gut Microbiota Alteration in Treated Group	Biological Activity	Ref.
	Powdered fruiting bodies (as dietary supplement)	A pilot study with 13 healthy adults	↑ <i>Roseburia faecis</i> ↑ <i>Faecalibacterium prausnitzii</i> ↑ <i>Eubacterium rectale</i> ↑ <i>Fusicatenibacter saccharivorans</i> ↑ <i>Kineothrix alysoides</i> ↑ <i>Gemmiger formicilis</i> ↑ <i>Dorea longicatena</i> ↓ <i>Streptococcus thermophilus</i> ↓ <i>Roseburia intestinalis</i> ↓ <i>Bacteroides caccae</i> ↓ <i>Anaerostipes hadrus</i>	Short supplementation (7 days) resulted in increased alpha and beta diversity—it may produce short term effect retention.	[200]
<i>Dictyophora indusiata</i>	Different fractions of polysaccharides: soluble (DIPX) and insoluble in water (DIPY)	In vivo: Kunming mice	In DIPY group: ↓ <i>Lachnospiraceae</i> ↑ <i>Lactobacillus</i> Gut microbiota composition in DIPX group did not differ to the control group	↑Production of SCFAs in DIPY group Production of SCFAs in DIPX groups did not differ comparing to the control group	[201]
	Isolated polysaccharide	In vivo: BALB/c mice HFD	Addition of polysaccharide resulted in reversion of HFD-induced gut alteration: ↓ <i>Firmicutes/Bacteroidetes</i> ratio	Improved intestinal barrier after polysaccharide supplementation to diet: ↑claudin-1, ↑occludin, and ↑zonula occludens (ZO-1) ↓Serum LPS → ↓peripheral inflammation	[202]
<i>Helvella leucopus</i>	Isolated polysaccharide (p-HLP)	In vivo: C57BL/6 DSS-induced colitis	↑ <i>Bacteroidaceae</i> ↑ <i>Prevotellaceae</i> ↑ <i>Akkermansiaceae</i>	↓Local inflammation • ↓IL-6, ↓IL-1b, ↓TNF-α, ↓iNOS, ↓COX-2 at mRNA level • ↑IL-10 at mRNA level	[203]
<i>Tremella fuciformis</i>	Isolated polysaccharides	In vivo: C57BL/6 DSS-induced colitis	↑ <i>Lactobacillaceae</i> ↑ <i>Lactobacillus</i> ↑ <i>Marinifilaceae</i> ↓ <i>Helicobacter</i> ↓ <i>Ruminococcaceae</i>	↓ Local inflammation • ↓IFN-γ, ↓IL-1β, ↓IL-6, ↓TGF-β, ↓TNF-α • ↑IL-10 • ↑PPAR-γ activity	[204]
<i>Grifola frondosa</i>	Isolated polysaccharides	In vivo: Wistar rats HFD	↑ <i>Helicobacter</i> ↑ <i>Intestinimonas</i> ↑ <i>Barnesiella</i> ↑ <i>Parasutterella</i> ↑ <i>Ruminococcus</i> ↑ <i>Flavonifracter</i> ↓ <i>Clostridium-XVIII</i> ↓ <i>Turicibacter</i>		[205]

Table 4. Cont.

Medicinal Mushroom Species	Active Ingredient	Experimental Model	Gut Microbiota Alteration in Treated Group	Biological Activity	Ref.
	Isolated heteropolysaccharide (GFP-N)	In vivo: ICR mice T2DM induced	↑ <i>Porphyromonas gingivalis</i> ↑ <i>Akkermansia muciniphila</i> ↑ <i>Lactobacillus acidophilus</i> ↑ <i>Tannerella forsythia</i> ↑ <i>Bacteroides acidifaciens</i> ↑ <i>Roseburia intestinalis</i>	Improved oral glucose test, alleviated insulin resistance, decreased the fast blood glucose level	[206]

↓—decrease, ↑—increase.

The influence of mushroom ingestion on the gut microbiota can be measured by different approaches. There is increasing research on the impact of mushroom administration on the use of animal models, mostly rodents. The tested substance can be powdered fruiting bodies or mycelium of selected mushroom species or an isolated polysaccharide—mostly the one that cannot be hydrolyzed in an acidic environment.

The analyzes comprises evaluation of microbiota alteration after a certain amount of time of mushroom administration—analyzes of compositional complexity of a bacterial community within a site that increases with the number of present species and with the evenness of their relative abundances (Alpha diversity) and highlight taxonomical differences between pairs of samples (Beta diversity) [207]. Methods for the complex analysis of the gut microbiota are based on 16Sr RNA high-throughput sequencing and metabolomics [121]. Another factor, the *Firmicutes/Bacteroidetes* ratio (F/B ratio) during may be important biomarkers that prove no negative effect on the gut microbiota [208]. For example, the addition of powdered *A. bisporus* fruiting bodies to mice resulted in a stable F/B ratio during the course of the experiment, indicating that supplementation with *A. bisporus* in the diet did not result in any negative alteration of the intestinal microbiota [195].

Another important factor contributing to the potential mood-elevating activity of medicinal/edible mushrooms is restoring the intestinal barrier; by this means, the levels of so-called tight junction proteins are measured such as ZO-1, claudin-3, and occludin using the Western blot method [209]. Ethanolic extract of *G. lucidum* administration to: C57BL6/J mice with DDS-induced colitis for 14 days that disrupted the intestinal barrier resulted in recovery of intestinal barrier function by increasing the level of junction proteins, the levels of which, after treatment, were at the same level as in the control group [178]. Disruption of the intestinal barrier may result in some diseases such as irritable bowel syndrome (IBS), obesity, non-alcoholic fatty liver disease (NAFLD), type 2 diabetes mellitus [210–213]. A disrupted intestinal barrier—the core of the hypothesis of “leaky gut”— may be an important factor in the pathophysiology of depression, as the leakage of bacteria into the bloodstream and pro-inflammatory substances such as endotoxins that increase inflammation can increase the odds of depression [214,215].

Interestingly, psychological stress can affect the permeability of the intestinal barrier by the so-called hypothalamic-pituitary-adrenal axis (HPA axis) [216]. Stress can activate the HPA axis, which leads to cortisol production and release from the adrenal cortex in response to adrenocorticotrophic hormone (ACTH). Cortisol can increase gut permeability and thus, cause the penetration of bacterial LPS into the bloodstream, which leads to peripheral inflammation.

7. Conclusions

The International Society for Nutritional Psychiatry Research pointed out that human nutrition should be taken into account, especially in patients suffering from MDD or anxiety disorders [217]. There are many studies that show the effect of human nutrition on human health. For example, in the study that involved patients with type 2 diabetes divided into two groups: Mediterranean diet with nuts and the control group, the chances of depression

in the experimental group were 41% lower compared to the control group [218]. The high content of omega-3 polyunsaturated fatty acids in the Mediterranean diet is believed to be responsible for lowering the risk of depression due to their antioxidant activity, as well as the rich content of polyphenols and reduced intake of red meat [219,220].

A plethora of evidence suggests that the introduction of medicinal/edible mushrooms to the daily diet in the form of mushroom-containing dishes and dietary supplements may lower the risk of depression development. The review summarized the potential mechanisms of antidepressant action from different perspectives, which are illustrated in Figure 2.

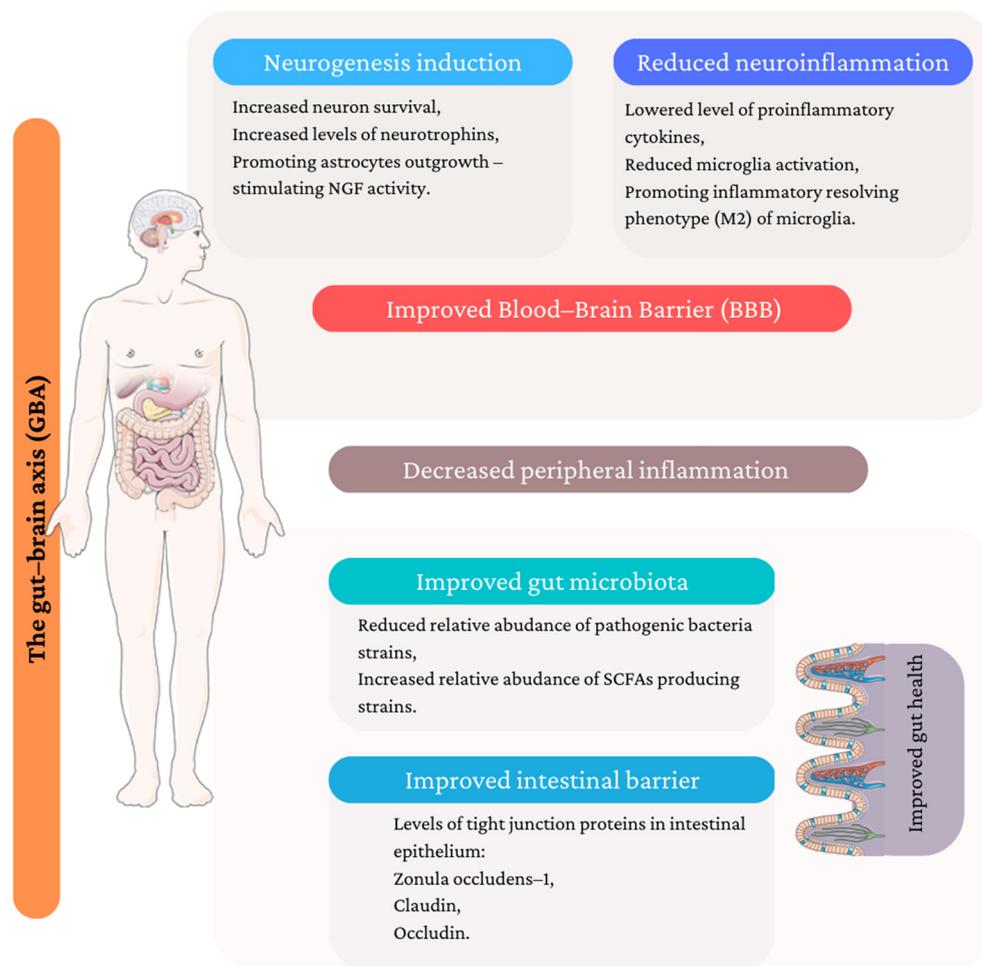


Figure 2. Potential mechanism of antidepressant activity of medicinal/edible mushroom. (Figure composed using Servier Medical Art: <http://smart.servier.com/> (accessed on 14 January 2023)).

In conclusion, edible mushrooms should be considered a valuable daily dietary source with potential multidirectional antidepressant activity.

Author Contributions: Conceptualization, J.L.; investigation, J.L. and K.H.; writing—original draft preparation, J.L., K.H. and B.M.; writing—review and editing, J.L., K.K., K.S.-Z. and B.M.; visualization, J.L.; supervision, B.M. All authors have read and agreed to the published version of the manuscript.

Funding: The article was funded as a part of the research project supported by the Polish Ministry of Science and Higher Education (Grants PL: N42/DBS/000271). Publication of this article was also co-funded by the Priority Research Area qLife under the program “Excellence Initiative—Research University” at the Jagiellonian University in Krakow (06/IDUB/2019/94).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data is contained within the article.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

World Health Organization (WHO), tumor necrosis factor-alpha (TNF- α), interferon gamma (INF- γ), monoamine oxidase (MAO), selective serotonin reuptake inhibitors (SSRIs), brain-derived neurotrophic factor (BDNF), Major Depressive Disorder (MDD), neurotrophin-3 (NT-3), neurotrophin-4 (NT-4), nerve growth factor (NGF), L-tryptophan (Trp), rapid acting antidepressant (RAAD), treatment-resistant depression (TRD), 5-hydroxy-L-tryptophan (5-OH-L-Trp), Trp hydroxylase (TPH), large neutral amino acids (LNAAs), blood-brain barrier (BBB), colon epithelial cells (CaCo-2), rapid-acting antidepressants (RAADs), medial prefrontal cortex (mPFC), high-fat diet (HFD), erinacine A (EA), *H. erinaceus* extract (HEM), hot aqueous extract (HAE), ethanol extract (EE), Meniere's disease (MD), deacetyl ganoderic acid F (DeGA F), toll-like receptors (TLRs), pathogen-associated molecular patterns (PAMPs), danger-associated molecular patterns (DAMPs), lipopolysaccharide (LPS), nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), inducible nitric oxide synthase (iNOS), nitric oxide (NO), cyclooxygenase-2 (COX-2), p75 neurotrophin receptor (p75^{NTR}), tyrosine kinase receptor (Trk), polysaccharide-peptide (PGL), tropomyosin receptor kinase B (TrkB), synaptophysin (SYP), Center for Epidemiologic Studies Depression Scale (CES-D), Indefinite Complaints Index (ICI), 2 α -hydroxy-inotodiol (2 α -HI), cuprizone (CPZ), short-chain fatty acids (SCFAs), dextran sulphate sodium administration (DSS), ethanolic extract of *G. lucidum* mycelium (GLAA), high-cholesterol diet (HCD), Specific free-pathogen (SFC), water-soluble polysaccharides (PCX), water-insoluble polysaccharides (PCY), triterpenoid saponins (PCZ), unpredictable mild stress model (CUMS), polysaccharide-peptide I (PSI), polysaccharide-peptide II (PSII), p-chlorophenylalanine (PCPA), alkaline phosphatase (ALP), different fractions of polysaccharides: soluble in water (DIPX), different fractions of polysaccharides: insoluble in water (DIPY), zonula occludens (ZO-1), irritable bowel syndrome (IBS), non-alcoholic fatty liver disease (NAFLD), hypothalamic-pituitary-adrenal axis (HPA axis), adrenocorticotrophic hormone (ACTH).

References

- Moreno-Agostino, D.; Wu, Y.-T.; Daskalopoulou, C.; Hasan, M.T.; Huisman, M.; Prina, M. Global trends in the prevalence and incidence of depression: a systematic review and meta-analysis. *J. Affect. Disord.* **2021**, *281*, 235–243. [[CrossRef](#)] [[PubMed](#)]
- Uher, R.; Payne, J.L.; Pavlova, B.; Perlis, R.H. Major Depressive Disorder in DSM-5: Implications for clinical practice and research of changes from DSM-IV. *Depress. Anxiety* **2014**, *31*, 459–471. [[CrossRef](#)] [[PubMed](#)]
- Dhar, A.K.; Barton, D.A. Depression and the link with cardiovascular disease. *Front. Psychiatry* **2016**, *7*, 33. [[CrossRef](#)] [[PubMed](#)]
- Galts, C.P.C.; Bettio, L.E.B.; Jewett, D.C.; Yang, C.C.; Brocardo, P.S.; Rodrigues, A.L.S.; Thacker, J.S.; Gil-Mohapel, J. Depression in neurodegenerative diseases: Common mechanisms and current treatment options. *Neurosci. Biobehav. Rev.* **2019**, *102*, 56–84. [[CrossRef](#)]
- Ettman, C.K.; Cohen, G.H.; Abdalla, S.M.; Sampson, L.; Trinquart, L.; Castrucci, B.C.; Bork, R.H.; Clark, M.A.; Wilson, I.; Vivier, P.M.; et al. Persistent depressive symptoms during COVID-19: A national, population-representative, longitudinal study of U.S. adults. *Lancet Reg. Health Am.* **2022**, *5*, 100091. [[CrossRef](#)]
- Calcagno, N.; Colombo, E.; Maranzano, A.; Pasquini, J.; Keller Sarmiento, I.J.; Trogu, F.; Silani, V. Rising evidence for neurological involvement in COVID-19 Pandemic. *Neurol. Sci.* **2020**, *41*, 1339. [[CrossRef](#)]
- Aghagoli, G.; Gallo Marin, B.; Katchur, N.J.; Chaves-Sell, F.; Asaad, W.F.; Murphy, S.A. Neurological involvement in COVID-19 and potential mechanisms: A review. *Neurocrit. Care* **2021**, *34*, 1062–1071. [[CrossRef](#)]
- Ban, T.A. Pharmacotherapy of mental illness—A historical analysis. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2001**, *25*, 709–727. [[CrossRef](#)]
- Bloch, R.G.; Dooneief, A.S.; Buchberg, A.S.; Sellman, S. The clinical effect of isoniazid and iproniazid in the treatment of pulmonary tuberculosis. *Ann. Intern. Med.* **1954**, *40*, 881–900. [[CrossRef](#)]
- Ayd, F.J. A preliminary report on mersalid. *Am. J. Psychiatry* **1957**, *114*, 459. [[CrossRef](#)]
- Goldberg, L.I. Monoamine Oxidase Inhibitors. *JAMA* **1964**, *190*, 456–462. [[CrossRef](#)] [[PubMed](#)]
- Perez-Caballero, L.; Torres-Sanchez, S.; Romero-López-Alberca, C.; González-Saiz, F.; Mico, J.A.; Berrocoso, E. Monoaminergic system and depression. *Cell Tissue Res.* **2019**, *377*, 107–113. [[CrossRef](#)] [[PubMed](#)]

13. Vaswani, M.; Linda, F.K.; Ramesh, S. Role of selective serotonin reuptake inhibitors in psychiatric disorders: A comprehensive review. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2003**, *27*, 85–102. [[CrossRef](#)] [[PubMed](#)]
14. Schmidt, H.D.; Duman, R.S. The role of neurotrophic factors in adult hippocampal neurogenesis, antidepressant treatments and animal models of depressive-like behavior. *Behav. Pharmacol.* **2007**, *18*, 391–418. [[CrossRef](#)] [[PubMed](#)]
15. Castrén, E.; Vöikar, V.; Rantamäki, T. Role of neurotrophic factors in depression. *Curr. Opin. Pharmacol.* **2007**, *7*, 18–21. [[CrossRef](#)]
16. Brunoni, A.R.; Lopes, M.; Fregni, F. A systematic review and meta-analysis of clinical studies on Major Depression and BDNF levels: Implications for the role of neuroplasticity in depression. *Int. J. Neuropsychopharmacol.* **2008**, *11*, 1169–1180. [[CrossRef](#)]
17. Sairanen, M.; Lucas, G.; Ernfors, P.; Castrén, M.; Castrén, E. Brain-Derived Neurotrophic Factor and antidepressant drugs have different but coordinated effects on neuronal turnover, proliferation, and survival in the adult dentate gyrus. *J. Neurosci.* **2005**, *25*, 1089–1094. [[CrossRef](#)]
18. Nestler, E.J.; Barrot, M.; DiLeone, R.J.; Eisch, A.J.; Gold, S.J.; Monteggia, L.M. Neurobiology of depression. *Neuron* **2002**, *34*, 13–25. [[CrossRef](#)]
19. Nibuya, M.; Morinobu, S.; Duman, R.S. Regulation of BDNF and TrkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. *J. Neurosci.* **1995**, *15*, 7539–7547. [[CrossRef](#)]
20. Neto, F.L.; Borges, G.; Torres-Sanchez, S.; Mico, J.A.; Berrocoso, E. Neurotrophins role in depression neurobiology: A review of basic and clinical evidence. *Curr. Neuropharmacol.* **2011**, *9*, 530–552. [[CrossRef](#)]
21. Muszyńska, B.; Grzywacz-Kisielewska, A.; Kała, K.; Gdula-Argasińska, J. Anti-inflammatory properties of edible mushrooms: A review. *Food Chem.* **2018**, *243*, 373–381. [[CrossRef](#)] [[PubMed](#)]
22. Chen, S.Y.; Ho, K.J.; Hsieh, Y.J.; Wang, L.T.; Mau, J.L. Contents of lovastatin, γ -aminobutyric acid and ergothioneine in mushroom fruiting bodies and mycelia. *LWT* **2012**, *47*, 274–278. [[CrossRef](#)]
23. Muszyńska, B.; Kała, K.; Włodarczyk, A.; Krakowska, A.; Ostachowicz, B.; Gdula-Argasińska, J.; Suchocki, P. *Lentinula edodes* as a source of bioelements released into artificial digestive juices and potential anti-inflammatory material. *Biol. Trace Elem. Res.* **2020**, *194*, 603–613. [[CrossRef](#)]
24. Ba, D.M.; Gao, X.; Al-Shaar, L.; Muscat, J.E.; Chinchilli, V.M.; Beelman, R.B.; Richie, J.P. Mushroom intake and depression: A population-based study using data from the US National Health and Nutrition Examination Survey (NHANES), 2005–2016. *J. Affect. Disord.* **2021**, *294*, 686–692. [[CrossRef](#)]
25. Vigna, L.; Morelli, F.; Agnelli, G.M.; Napolitano, F.; Ratto, D.; Occhinegro, A.; di Iorio, C.; Savino, E.; Girometta, C.; Brandalise, F.; et al. *Hericium erinaceus* improves mood and sleep disorders in patients affected by overweight or obesity: Could circulating Pro-BDNF and BDNF be potential biomarkers? *Evid. Based Complement. Altern. Med.* **2019**, *2019*, 7861297. [[CrossRef](#)] [[PubMed](#)]
26. Okamura, H.; Anno, N.; Tsuda, A.; Inokuchi, T.; Uchimura, N.; Inanaga, K. The effects of *Herichium erinaceus* on sleep quality and subjective well-being among female undergraduate students: A pilot study. *Pers. Med. Universe* **2015**, *4*, 76–78. [[CrossRef](#)]
27. Park, S.K.; Oh, C.M.; Ryoo, J.H.; Jung, J.Y. The protective effect of mushroom consumption on depressive symptoms in Korean population. *Sci. Rep.* **2022**, *12*, 21914. [[CrossRef](#)]
28. Bell, C.J.; Hood, S.D.; Nutt, D.J. Acute tryptophan depletion. Part II: Clinical Effects and Implications. *Aust. N. Z. J. Psychiatry* **2005**, *39*, 565–574. [[CrossRef](#)]
29. Oladele, J.O.; Oladiji, A.T.; Oladele, O.T.; Oyeleke, O.M. Reactive oxygen species in neurodegenerative diseases: Implications in pathogenesis and treatment strategies. In *Reactive Oxygen Species*; Ahmad, R., Ed.; IntechOpen: London, UK, 2021. [[CrossRef](#)]
30. Pardridge, W.M. Blood-Brain Barrier carrier-mediated transport and brain metabolism of amino acids. *Neurochem. Res.* **1998**, *23*, 635–644. [[CrossRef](#)]
31. Silber, B.Y.; Schmitt, J.A.J. Effects of tryptophan loading on human cognition, mood, and sleep. *Neurosci. Biobehav. Rev.* **2010**, *34*, 387–407. [[CrossRef](#)]
32. Ruddick, J.P.; Evans, A.K.; Nutt, D.J.; Lightman, S.L.; Rook, G.A.W.; Lowry, C.A. Tryptophan metabolism in the Central Nervous System: Medical implications. *Expert Rev. Mol. Med.* **2006**, *8*, 1–27. [[CrossRef](#)] [[PubMed](#)]
33. Fernstrom, J.D. Large neutral amino acids: Dietary effects on brain neurochemistry and function. *Amino Acids* **2013**, *45*, 419–430. [[CrossRef](#)] [[PubMed](#)]
34. Tovar, A.R.; Tews, J.K.; Torres, N.; Harper, A.E. Neutral amino acid transport into rat skeletal muscle: Competition, adaptive regulation, and effects of insulin. *Metabolism* **1991**, *40*, 410–419. [[CrossRef](#)] [[PubMed](#)]
35. Cervenka, I.; Agudelo, L.Z.; Ruas, J.L. Kynurenines: Tryptophan's metabolites in exercise, inflammation, and mental health. *Science* **2017**, *357*, eaaf9794. [[CrossRef](#)] [[PubMed](#)]
36. Muszyńska, B.; Sułkowska-Ziaja, K.; Łojewski, M.; Opoka, W.; Zając, M.; Rojowski, J. Edible mushroom in prophylaxis and treatment of human diseases. *Med. Int. Rev.* **2013**, *101*, 170–183.
37. Muszyńska, B.; Sułkowska-Ziaja, K.; Ekiert, H. Indole compounds in some culinary-medicinal higher basidiomycetes from Poland. *Int. J. Med. Mushrooms* **2011**, *13*, 449–454. [[CrossRef](#)]
38. Muszyńska, B.; Sułkowska-Ziaja, K.; Maślanka, A.; Rojowski, J.; Opoka, W.; Łojewski, M. Analysis of indole compounds quantity in biomass of edible Basidiomycota species from in vitro cultures and from cultures supplemented with L-tryptophan. *Acta Chromatogr.* **2016**, *28*, 223–238. [[CrossRef](#)]

39. Krakowska, A.; Zięba, P.; Włodarczyk, A.; Kała, K.; Sułkowska-Ziaja, K.; Bernaś, E.; Sękara, A.; Ostachowicz, B.; Muszyńska, B. Selected edible medicinal mushrooms from *Pleurotus* genus as an answer for human civilization diseases. *Food Chem.* **2020**, *327*, 127084. [[CrossRef](#)]
40. Muszyńska, B.; Sułkowska-Ziaja, K.; Wójcik, A. Levels of physiologically active indole derivatives in the fruiting bodies of some edible mushrooms (Basidiomycota) before and after thermal processing. *Mycoscience* **2013**, *54*, 321–326. [[CrossRef](#)]
41. Muszyńska, B.; Sułkowska-Ziaja, K. Analysis of indole compounds in edible Basidiomycota species after thermal processing. *Food Chem.* **2012**, *132*, 455–459. [[CrossRef](#)]
42. Sułkowska-Ziaja, K.; Muszyńska, B.; Szewczyk, A. Antioxidant components of selected indigenous edible mushrooms of the obsolete order Aphyllophorales. *Rev. Iberoam. Micol.* **2015**, *32*, 99–102. [[CrossRef](#)] [[PubMed](#)]
43. Muszyńska, B.; Sułkowska-Ziaja, K.; Ekiert, H. Indole compounds in fruiting bodies of some edible Basidiomycota species. *Food Chem.* **2011**, *125*, 1306–1308. [[CrossRef](#)]
44. Muszyńska, B.; Sułkowska-Ziaja, K.; Ekiert, H. Indole compounds in fruiting bodies of some selected macromycetes species and in their mycelia cultured in vitro. *Pharmazie* **2009**, *64*, 479–480. [[CrossRef](#)]
45. Çevikkalp, S.A.; Löker, G.B.; Yaman, M.; Amoutzopoulos, B. A Simplified HPLC method for determination of tryptophan in some cereals and legumes. *Food Chem.* **2016**, *193*, 26–29. [[CrossRef](#)] [[PubMed](#)]
46. Friedman, M. Analysis, nutrition, and health benefits of tryptophan. *Int. J. Tryptophan Res.* **2018**, *11*, 1178646918802282. [[CrossRef](#)] [[PubMed](#)]
47. Maffei, M.E. 5-Hydroxytryptophan (5-HTP): Natural occurrence, analysis, biosynthesis, biotechnology, physiology and toxicology. *Int. J. Mol. Sci.* **2020**, *22*, 181. [[CrossRef](#)]
48. Vigliante, I.; Mannino, G.; Maffei, M.E. Chemical characterization and DNA fingerprinting of *Griffonia simplicifolia* Baill. *Molecules* **2019**, *24*, 1032. [[CrossRef](#)]
49. Ly, D.; Kang, K.; Choi, J.Y.; Ishihara, A.; Back, K.; Lee, S.G. HPLC Analysis of serotonin, tryptamine, tyramine, and the hydroxycinnamic acid amides of serotonin and tyramine in food vegetables. *J. Med. Food* **2008**, *11*, 385–389. [[CrossRef](#)]
50. Muszyńska, B.; Kała, K.; Firlej, A.; Sułkowska-Ziaja, K. *Cantharellus cibarius*—culinary-medicinal mushroom content and biological activity. *Acta Pol. Pharm.* **2016**, *73*, 589–598.
51. Muszyńska, B.; Sułkowska-Ziaja, K.; Hałaszczyk, P.; Krężałek, R.; Łojewski, M. Analysis of indole derivatives in methanolic extracts from mycelium of *Agaricus bisporus* cultured in vitro on liquid Oddoux medium. *Folia Biol. Oecologica* **2014**, *10*, 66–72. [[CrossRef](#)]
52. Sułkowska-Ziaja, K.; Muszyńska, B.; Ekiert, H. Analysis of indole compounds from the fruiting bodies and the culture mycelia of *Sarcodon imbricatus*. *Mycoscience* **2014**, *55*, 164–167. [[CrossRef](#)]
53. Muszyńska, B.; Sułkowska-Ziaja, K. Analysis of indole compounds in fruiting bodies and in mycelia from in vitro cultures of *Calocera viscosa* (Basidiomycota). *Acta Mycol.* **2012**, *47*, 57–64. [[CrossRef](#)]
54. Muszyńska, B.; Sułkowska-Ziaja, K.; Ekiert, H. Analysis of indole compounds in methanolic extracts from the fruiting bodies of *Cantharellus cibarius* (the Chanterelle) and from the mycelium of this species cultured in vitro. *J. Food Sci. Technol.* **2013**, *50*, 1233–1237. [[CrossRef](#)]
55. Opoka, W.; Kała, K.; Krężałek, R.; Sułkowska-Ziaja, K.; Maślanka, A.; Muszyńska, B. TLC–densitometry analysis of indole compounds in mycelial culture of *Imleria badia* and *Agaricus bisporus* enriched with precursors—serine or anthranilic acid. *Acta Chromatogr.* **2018**, *30*, 236–242. [[CrossRef](#)]
56. Muszyńska, B.; Kała, K.; Sułkowska-Ziaja, K.; Krakowska, A.; Opoka, W. *Agaricus bisporus* and its in vitro culture as a source of indole compounds released into artificial digestive juices. *Food Chem.* **2016**, *199*, 509–515. [[CrossRef](#)]
57. Kała, K.; Sułkowska-Ziaja, K.; Rojowski, J.; Opoka, W.; Muszyńska, B. *Tricholoma equestre* species as a source of indole compounds and zinc released into artificial digestive juices. *Med. Int. Rev.* **2016**, *106*, 35–39.
58. Kała, K.; Krakowska, A.; Sułkowska-Ziaja, K.; Szewczyk, A.; Reczyński, W.; Opoka, W.; Muszyńska, B. Kinetics of extracted bioactive components from mushrooms in artificial digestive juices. *Int. J. Food Prop.* **2016**, *20*, 1796–1817. [[CrossRef](#)]
59. Saghir, S.A. Absorption. *Encycl. Toxicol. Third Ed.* **2014**, 1–6. [[CrossRef](#)]
60. Fang, Y.; Cao, W.; Xia, M.; Pan, S.; Xu, X. Study of structure and permeability relationship of flavonoids in Caco-2 Cells. *Nutrients* **2017**, *9*, 1301. [[CrossRef](#)]
61. Kała, K.; Krakowska, A.; Gdula-Argasińska, J.; Opoka, W.; Muszyńska, B. Assessing the bioavailability of zinc and indole compounds from mycelial cultures of the Bay Mushroom *Imleria badia* (Agaricomycetes) using in vitro models. *Int. J. Med. Mushrooms* **2019**, *21*, 343–352. [[CrossRef](#)]
62. Voineskos, D.; Daskalakis, Z.J.; Blumberger, D.M. Management of Treatment-Resistant Depression: Challenges and strategies. *Neuropsychiatr. Dis. Treat.* **2020**, *16*, 221–234. [[CrossRef](#)] [[PubMed](#)]
63. Gaynes, B.N.; Lux, L.; Gartlehner, G.; Asher, G.; Forman-Hoffman, V.; Green, J.; Boland, E.; Weber, R.P.; Randolph, C.; Bann, C.; et al. Defining Treatment-Resistant Depression. *Depress. Anxiety* **2020**, *37*, 134–145. [[CrossRef](#)] [[PubMed](#)]
64. Riggs, L.M.; Gould, T.D. Ketamine and the future of Rapid-Acting Antidepressants. *Annu. Rev. Clin. Psychol.* **2021**, *17*, 207–231. [[CrossRef](#)] [[PubMed](#)]
65. Witkin, J.M.; Martin, A.E.; Golani, L.K.; Xu, N.Z.; Smith, J.L. Rapid-Acting Antidepressants. *Adv. Pharmacol.* **2019**, *86*, 47–96. [[CrossRef](#)]

66. McIntyre, R.S.; Rosenblat, J.D.; Nemeroff, C.B.; Sanacora, G.; Murrrough, J.W.; Berk, M.; Brietzke, E.; Dodd, S.; Gorwood, P.; Ho, R.; et al. Synthesizing the evidence for ketamine and esketamine in Treatment-Resistant Depression: An international expert opinion on the available evidence and implementation. *Am. J. Psychiatry* **2021**, *178*, 383–399. [[CrossRef](#)]
67. Machado-Vieira, R.; Henter, I.D.; Zarate, C.A. New targets for Rapid Antidepressant Action. *Prog. Neurobiol.* **2017**, *152*, 21–37. [[CrossRef](#)]
68. Kosentka, P.; Sprague, S.L.; Ryberg, M.; Gartz, J.; May, A.L.; Campagna, S.R.; Matheny, P.B. Evolution of the toxins muscarine and psilocybin in a family of mushroom-forming fungi. *PLoS ONE* **2013**, *8*, e64646. [[CrossRef](#)]
69. Kreisel, H.; Lindquest, U. *Gymnopilus purpuratus*, ein psilocybinhaltiger Pilz adventiv in Berzirk Rostock. *Z. Mykol.* **1988**, *54*, 73–76.
70. Halama, M.; Poliwoda, A.; Jasicka-Misiak, I.; Wieczorek, P.P.; Rutkowski, R. *Pholiotina cyanopus*, a rare fungus producing psychoactive tryptamines. *Open Life Sci.* **2015**, *10*, 40–51. [[CrossRef](#)]
71. Saupe, S.G. Occurrence of psilocybin/psilocin in *Pluteus salicinus* (Pluteaceae). *Mycologia* **2018**, *73*, 781–784. [[CrossRef](#)]
72. Stijve, T. Psilocin, psilocybin, serotonin and urea in *Panaeolus cyanescens* from various origin. *Pers. Mol. Phylogeny Evol. Fungi* **1992**, *15*, 117–121.
73. Froese, T.; Guzmán, G.; Guzmán-Dávalos, L. On the origin of the genus Psilocybe and its potential ritual use in ancient Africa and Europe. *Econ. Bot.* **2016**, *70*, 103–114. [[CrossRef](#)]
74. Mahapatra, A.; Gupta, R. Role of psilocybin in the treatment of depression. *Ther. Adv. Psychopharmacol.* **2017**, *7*, 54. [[CrossRef](#)] [[PubMed](#)]
75. Quednow, B.B.; Komater, M.; Geyer, M.A.; Vollenweider, F.X. Psilocybin-induced deficits in automatic and controlled inhibition are attenuated by ketanserin in healthy human volunteers. *Neuropsychopharmacology* **2012**, *37*, 630–640. [[CrossRef](#)]
76. Eison, A.S.; Mullins, U.L. Regulation of central 5-HT_{2A} receptors: A review of in vivo studies. *Behav. Brain Res.* **1996**, *73*, 177–181. [[CrossRef](#)]
77. Van Oekelen, D.; Luyten, W.H.M.L.; Leysen, J.E. 5-HT_{2A} and 5-HT_{2C} receptors and their atypical regulation properties. *Life Sci.* **2003**, *72*, 2429–2449. [[CrossRef](#)]
78. Gill, H.; Gill, B.; Chen-Li, D.; El-Halabi, S.; Rodrigues, N.B.; Cha, D.S.; Lipsitz, O.; Lee, Y.; Rosenblat, J.D.; Majeed, A.; et al. The emerging role of psilocybin and MDMA in the treatment of mental illness. *Expert Rev. Neurother.* **2020**, *20*, 1263–1273. [[CrossRef](#)]
79. Dos Santos, R.G.; Hallak, J.E.C. Therapeutic use of serotonergic hallucinogens: A review of the evidence and of the biological and psychological mechanisms. *Neurosci. Biobehav. Rev.* **2020**, *108*, 423–434. [[CrossRef](#)]
80. Nkadameng, S.M.; Steinmann, C.M.L.; Eloff, J.N. Anti-inflammatory effects of four psilocybin-containing magic mushroom water extracts in vitro on 15-lipoxygenase activity and on lipopolysaccharide-induced cyclooxygenase-2 and inflammatory cytokines in human U937 macrophage cells. *J. Inflamm. Res.* **2021**, *14*, 3729–3738. [[CrossRef](#)]
81. Schwarcz, R.; Bruno, J.P.; Muchowski, P.J.; Wu, H.Q. Kynurenines in the mammalian brain: When physiology meets pathology. *Nat. Rev. Neurosci.* **2012**, *13*, 465–477. [[CrossRef](#)]
82. Ko, K.; Kopra, E.I.; Cleare, A.J.; Rucker, J.J. Psychedelic therapy for depressive symptoms: A systematic review and meta-analysis. *J. Affect. Disord.* **2023**, *322*, 194–204. [[CrossRef](#)] [[PubMed](#)]
83. Goodwin, G.M.; Aaronson, S.T.; Alvarez, O.; Arden, P.C.; Baker, A.; Bennett, J.C.; Bird, C.; Blom, R.E.; Brennan, C.; Bruschi, D.; et al. Single-dose psilocybin for a treatment-resistant episode of major depression. *N. Engl. J. Med.* **2022**, *387*, 1637–1648. [[CrossRef](#)] [[PubMed](#)]
84. Gotvaldová, K.; Borovička, J.; Hájková, K.; Cihlářová, P.; Rockefeller, A.; Kuchař, M. Extensive collection of psychotropic mushrooms with determination of their tryptamine alkaloids †. *Int. J. Mol. Sci.* **2022**, *23*, 14068. [[CrossRef](#)]
85. Milne, N.; Thomsen, P.; Mølgaard Knudsen, N.; Rubaszka, P.; Kristensen, M.; Borodina, I. Metabolic engineering of *Saccharomyces cerevisiae* for the de novo production of psilocybin and related tryptamine derivatives. *Metab. Eng.* **2020**, *60*, 25–36. [[CrossRef](#)]
86. Adams, A.M.; Kaplan, N.A.; Wei, Z.; Brinton, J.D.; Monnier, C.S.; Enacopol, A.L.; Ramelot, T.A.; Jones, J.A. In vivo production of psilocybin in *E. coli*. *Metab. Eng.* **2019**, *56*, 111–119. [[CrossRef](#)]
87. Vollenweider, F.X.; Komater, M. The neurobiology of psychedelic drugs: Implications for the treatment of mood disorders. *Nat. Rev. Neurosci.* **2010**, *11*, 642–651. [[CrossRef](#)]
88. Hutten, N.R.P.W.; Mason, N.L.; Dolder, P.C.; Kuypers, K.P.C. Motives and side-effects of microdosing with psychedelics among users. *Int. J. Neuropsychopharmacol.* **2019**, *22*, 426. [[CrossRef](#)]
89. Polito, V.; Stevenson, R.J. A systematic study of microdosing psychedelics. *PLoS ONE* **2019**, *14*, e0211023. [[CrossRef](#)]
90. Prochazkova, L.; Lippelt, D.P.; Colzato, L.S.; Kuchar, M.; Sjoerds, Z.; Hommel, B. Exploring the effect of microdosing psychedelics on creativity in an open-label natural setting. *Psychopharmacology* **2018**, *235*, 3401–3413. [[CrossRef](#)]
91. Cavanna, F.; Muller, S.; de la Fuente, L.A.; Zamberlan, F.; Palmucci, M.; Janeckova, L.; Kuchar, M.; Pallavicini, C.; Tagliazucchi, E. Microdosing with psilocybin mushrooms: A double-blind placebo-controlled study. *Transl. Psychiatry* **2022**, *12*, 307. [[CrossRef](#)]
92. Nesić, Z.; Delić, D.; Prostran, M.; Stojanović, R.; Vucković, S.; Todorović, Z. Psychiatric adverse effects induced by recombinant interferon alfa in patients with chronic hepatitis C. *Med. Pregl.* **2004**, *57*, 219–226. [[CrossRef](#)] [[PubMed](#)]
93. Ng, A.; Tam, W.W.; Zhang, M.W.; Ho, C.S.; Husain, S.F.; McIntyre, R.S.; Ho, R.C. IL-1 β , IL-6, TNF- α and CRP in elderly patients with depression or Alzheimer’s disease: Systematic review and meta-analysis. *Sci. Rep.* **2018**, *8*, 12050. [[CrossRef](#)] [[PubMed](#)]

94. Osimo, E.F.; Pillinger, T.; Rodriguez, I.M.; Khandaker, G.M.; Pariante, C.M.; Howes, O.D. Inflammatory markers in depression: A meta-analysis of mean differences and variability in 5,166 patients and 5,083 controls. *Brain Behav. Immun.* **2020**, *87*, 901–909. [[CrossRef](#)] [[PubMed](#)]
95. Carvalho, L.A.; Torre, J.P.; Papadopoulos, A.S.; Poon, L.; Juruena, M.F.; Markopoulou, K.; Cleare, A.J.; Pariante, C.M. Lack of clinical therapeutic benefit of antidepressants is associated overall activation of the inflammatory system. *J. Affect. Disord.* **2013**, *148*, 136–140. [[CrossRef](#)]
96. Varatharaj, A.; Galea, I. The Blood-Brain Barrier in systemic inflammation. *Brain Behav. Immun.* **2017**, *60*, 1–12. [[CrossRef](#)]
97. Widner, B.; Laich, A.; Sperner-Unterweger, B.; Ledochowski, M.; Fuchs, D. Neopterin production, tryptophan degradation, and mental depression—What is the link? *Brain Behav. Immun.* **2002**, *16*, 590–595. [[CrossRef](#)]
98. Dunn, A.J.; Welch, J. Stress-and endotoxin-induced increases in brain tryptophan and serotonin metabolism depend on sympathetic nervous system activity. *J. Neurochem.* **1991**, *57*, 1615–1622. [[CrossRef](#)]
99. O'Connor, J.C.; Lawson, M.A.; André, C.; Moreau, M.; Lestage, J.; Castanon, N.; Kelley, K.W.; Dantzer, R. Lipopolysaccharide-induced depressive-like behavior is mediated by indoleamine 2,3-dioxygenase activation in mice. *Mol. Psychiatry* **2009**, *14*, 511–522. [[CrossRef](#)]
100. Raison, C.L.; Dantzer, R.; Kelley, K.W.; Lawson, M.A.; Woolwine, B.J.; Vogt, G.; Spivey, J.R.; Saito, K.; Miller, A.H. CSF concentrations of brain tryptophan and kynurenines during immune stimulation with IFN- α : Relationship to CNS immune responses and depression. *Mol. Psychiatry* **2010**, *15*, 393–403. [[CrossRef](#)]
101. Maes, M.; Leonard, B.E.; Myint, A.M.; Kubera, M.; Verkerk, R. The new '5-HT' hypothesis of depression: Cell-mediated immune activation induces indoleamine 2,3-dioxygenase, which leads to lower plasma tryptophan and an increased synthesis of detrimental tryptophan catabolites (TRYCATs), both of which contribute to the onset of depression. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2011**, *35*, 702–721. [[CrossRef](#)]
102. Guidetti, P.; Eastman, C.L.; Schwarcz, R. Metabolism of [5-3H]Kynurenine in the rat brain in vivo: Evidence for the existence of a functional kynurenine pathway. *J. Neurochem.* **1995**, *65*, 2621–2632. [[CrossRef](#)] [[PubMed](#)]
103. Sanlioglu, S.; Williams, C.M.; Samavati, L.; Butler, N.S.; Wang, G.; McCray, P.B.; Ritchie, T.C.; Hunninghake, G.W.; Zandi, E.; Engelhardt, J.F. Lipopolysaccharide induces Rac1-dependent reactive oxygen species formation and coordinates Tumor Necrosis Factor- α secretion through IKK Regulation of NF-KB. *J. Biol. Chem.* **2001**, *276*, 30188–30198. [[CrossRef](#)] [[PubMed](#)]
104. Guillemain, G.J.; Kerr, S.J.; Smythe, G.A.; Smith, D.G.; Kapoor, V.; Armati, P.J.; Croitoru, J.; Brew, B.J. Kynurenine pathway metabolism in human astrocytes: A paradox for neuronal protection. *J. Neurochem.* **2001**, *78*, 842–853. [[CrossRef](#)]
105. Ogyu, K.; Kubo, K.; Noda, Y.; Iwata, Y.; Tsugawa, S.; Omura, Y.; Wada, M.; Tarumi, R.; Plitman, E.; Moriguchi, S.; et al. Kynurenine pathway in depression: A systematic review and meta-analysis. *Neurosci. Biobehav. Rev.* **2018**, *90*, 16–25. [[CrossRef](#)]
106. Chen, W.W.; Zhang, X.; Huang, W.J. Role of neuroinflammation in neurodegenerative diseases (review). *Mol. Med. Rep.* **2016**, *13*, 3391–3396. [[CrossRef](#)]
107. Pan, W.; Jiang, P.; Zhao, J.; Shi, H.; Zhang, P.; Yang, X.; Biazik, J.; Hu, M.; Hua, H.; Ge, X.; et al. β -Glucan from *Lentinula edodes* prevents cognitive impairments in high-fat diet-induced obese mice: Involvement of colon-brain axis. *J. Transl. Med.* **2021**, *19*, 54. [[CrossRef](#)]
108. Zhang, D.; Jia, Y.; Sun, X.; Li, H.; Yin, M.; Li, H.; Dai, L.; Han, L.; Wang, L.; Qian, M.; et al. The dectin-1 receptor signaling pathway mediates the remyelination effect of lentinan through suppression of neuroinflammation and conversion of microglia. *J. Immunol. Res.* **2022**, *2022*, 3002304. [[CrossRef](#)]
109. Maza, P.A.M.A.; Lee, J.-H.; Kim, Y.-S.; Sun, G.-M.; Sung, Y.-J.; Ponomarenko, L.P.; Stonik, V.A.; Ryu, M.; Kwak, J.-Y. Inotodiol from *Inonotus obliquus* Chaga mushroom induces atypical maturation in dendritic cells. *Front. Immunol.* **2021**, *12*, 650841. [[CrossRef](#)] [[PubMed](#)]
110. Wei, J.; Guo, W.H.; Cao, C.Y.; Kou, R.W.; Xu, Y.Z.; Górecki, M.; di Bari, L.; Pescitelli, G.; Gao, J.M. Polyoxygenated cyathane diterpenoids from the mushroom *Cyathus africanus*, and their neurotrophic and anti-neuroinflammatory activities. *Sci. Rep.* **2018**, *8*, 2175. [[CrossRef](#)]
111. Nguepi Tsopmejo, I.S.; Ding, M.; Wei, J.; Zhao, C.; Jiang, Y.; Li, Y.; Song, H. *Auricularia polytricha* and *Flammulina velutipes* ameliorate inflammation and modulate the gut microbiota via regulation of NF-KB and Keap1/Nrf2 signaling pathways on DSS-induced inflammatory bowel disease. *Food Biosci.* **2022**, *47*, 101426. [[CrossRef](#)]
112. Sam, S.E.; Sim, K.S.; Rahman, S.N.S.A.; Tan, Y.S. Neuroprotective properties of wild medicinal mushroom, *Sanguinoderma rugosum* (Agaricomycetes), extracts against glutamate-induced hippocampal cells. *Int. J. Med. Mushrooms* **2022**, *24*, 35–50. [[CrossRef](#)] [[PubMed](#)]
113. Lee, S.L.; Hsu, J.Y.; Chen, T.C.; Huang, C.C.; Wu, T.Y.; Chin, T.Y. Erinacine a prevents lipopolysaccharide-mediated glial cell activation to protect dopaminergic neurons against inflammatory factor-induced cell death in vitro and in vivo. *Int. J. Mol. Sci.* **2022**, *23*, 810. [[CrossRef](#)] [[PubMed](#)]
114. Tomas-Hernandez, S.; Blanco, J.; Garcia-Vallvé, S.; Pujadas, G.; Ojeda-Montes, M.J.; Gimeno, A.; Arola, L.; Minghetti, L.; Beltrán-Debón, R.; Mulero, M. Anti-inflammatory and immunomodulatory effects of the *Grifola frondosa* natural compound o-Orsellinaldehyde on LPS-challenged murine primary glial cells. Roles of NF-K β and MAPK. *Pharmaceutics* **2021**, *13*, 806. [[CrossRef](#)] [[PubMed](#)]
115. Koh, S.S.; Ooi, S.C.Y.; Lui, N.M.Y.; Qiong, C.; Ho, L.T.Y.; Cheah, I.K.M.; Halliwell, B.; Herr, D.R.; Ong, W.Y. Effect of ergothioneine on 7-ketocholesterol-induced endothelial injury. *Neuromolecular Med.* **2021**, *23*, 184. [[CrossRef](#)] [[PubMed](#)]

116. He, M.T.; Park, C.H.; Cho, E.J. Caterpillar medicinal mushroom, *Cordyceps militaris* (Ascomycota), attenuates A β 1–42-induced amyloidogenesis and inflammatory response by suppressing amyloid precursor protein progression and P38 MAPK/JNK Activation. *Int. J. Med. Mushrooms* **2021**, *23*, 71–83. [[CrossRef](#)]
117. Nallathamby, N.; Lakshmanan, H.; Vidyadaran, S.; Malek, S.N.A.; Phan, C.W.; Sabaratnam, V. Lipid-rich fraction of the sclerotium of tiger milk mushroom *Lignosus rhinocerotis* (Agaricomycetes) attenuates LPS-induced inflammation in BV2 cells via Nrf2 Pathway. *Braz. J. Pharm. Sci.* **2020**, *56*. [[CrossRef](#)]
118. Seow, S.L.S.; Naidu, M.; Sabaratnam, V.; Vidyadaran, S.; Wong, K.H. Tiger's milk medicinal mushroom, *Lignosus rhinocerotis* (Agaricomycetes) sclerotium inhibits nitric oxide production in LPS-stimulated BV2 Microglia. *Int. J. Med. Mushrooms* **2017**, *19*, 405–418. [[CrossRef](#)]
119. Scuto, M.; di Mauro, P.; Ontario, M.L.; Amato, C.; Modafferi, S.; Ciavardelli, D.; Salinaro, A.T.; Maiolino, L.; Calabrese, V. Nutritional mushroom treatment in Meniere's disease with *Coriolus versicolor*: A rationale for therapeutic intervention in neuroinflammation and antineurodegeneration. *Int. J. Mol. Sci.* **2020**, *21*, 284. [[CrossRef](#)]
120. Trovato-Salinaro, A.; Siracusa, R.; di Paola, R.; Scuto, M.; Fronte, V.; Koverech, G.; Luca, M.; Serra, A.; Toscano, M.A.; Petralia, A.; et al. Redox modulation of cellular stress response and lipoxin A4 expression by *Coriolus versicolor* in rat brain: Relevance to Alzheimer's disease pathogenesis. *Neurotoxicology* **2016**, *53*, 350–358. [[CrossRef](#)]
121. Sheng, F.; Zhang, L.; Wang, S.; Yang, L.; Li, P. Deacetyl ganoderic acid F inhibits LPS-induced neural inflammation via NF-KB pathway both in vitro and in vivo. *Nutrients* **2020**, *12*, 85. [[CrossRef](#)]
122. Geng, Y.; Zhu, S.; Cheng, P.; Lu, Z.M.; Xu, H.Y.; Shi, J.S.; Xu, Z.H. Bioassay-guided fractionation of ethyl acetate extract from *Armillaria mellea* attenuates inflammatory response in lipopolysaccharide (LPS) stimulated BV-2 microglia. *Phytomedicine* **2017**, *26*, 55–61. [[CrossRef](#)] [[PubMed](#)]
123. Moresco, E.M.Y.; LaVine, D.; Beutler, B. Toll-like Receptors. *Curr. Biol.* **2011**, *21*, R488–R493. [[CrossRef](#)] [[PubMed](#)]
124. Kaltschmidt, B.; Kaltschmidt, C. NF-B in the nervous system. *Cold Spring Harb. Perspect. Biol.* **2009**, *1*, a001271. [[CrossRef](#)] [[PubMed](#)]
125. Sahay, A.; Hen, R. Adult hippocampal neurogenesis in depression. *Nat. Neurosci.* **2007**, *10*, 1110–1115. [[CrossRef](#)] [[PubMed](#)]
126. Sapolsky, R.M. Glucocorticoids and hippocampal atrophy in neuropsychiatric disorders. *Arch. Gen. Psychiatry* **2000**, *57*, 925–935. [[CrossRef](#)] [[PubMed](#)]
127. McKinnon, M.C.; Yucel, K.; Nazarov, A.; MacQueen, G.M. A meta-analysis examining clinical predictors of hippocampal volume in patients with Major Depressive Disorder. *J. Psych. Neurosci.* **2009**, *34*, 41–54.
128. Belleau, E.L.; Treadway, M.T.; Pizzagalli, D.A. The impact of stress and Major Depressive Disorder on hippocampal and medial prefrontal cortex morphology. *Biol. Psychiatry* **2019**, *85*, 443–453. [[CrossRef](#)]
129. Amico, F.; Meisenzahl, E.; Koutsouleris, N.; Reiser, M.; Möller, H.J.; Frodl, T. Structural MRI correlates for vulnerability and resilience to Major Depressive Disorder. *J. Psych. Neurosci.* **2011**, *36*, 15–22. [[CrossRef](#)]
130. Chen, M.C.; Hamilton, J.P.; Gotlib, I.H. Decreased hippocampal volume in healthy girls at risk of depression. *Arch. Gen. Psychiatry* **2010**, *67*, 270–276. [[CrossRef](#)]
131. Taylor, W.D.; McQuoid, D.R.; Payne, M.E.; Zannas, A.S.; MacFall, J.R.; Steffens, D.C. Hippocampus atrophy and the longitudinal course of late-life depression. *Am. J. Geriatr. Psychiatry* **2014**, *22*, 1504–1512. [[CrossRef](#)]
132. Frodl, T.S.; Koutsouleris, N.; Bottlender, R.; Born, C.; Jäger, M.; Scupin, I.; Reiser, M.; Möller, H.J.; Meisenzahl, E.M. Depression-related variation in brain morphology over 3 years: Effects of stress? *Arch. Gen. Psychiatry* **2008**, *65*, 1156–1165. [[CrossRef](#)] [[PubMed](#)]
133. Papagni, S.A.; Benetti, S.; Arulanantham, S.; McCrory, E.; McGuire, P.; Mechelli, A. Effects of stressful life events on human brain structure: A longitudinal voxel-based morphometry study. *Stress* **2011**, *14*, 227–232. [[CrossRef](#)]
134. Huang, E.J.; Reichardt, L.F. Neurotrophins: Roles in neuronal development and function. *Annu. Rev. Neurosci.* **2001**, *24*, 677. [[CrossRef](#)]
135. Matsumoto, T.; Rauskolb, S.; Polack, M.; Klose, J.; Kolbeck, R.; Korte, M.; Barde, Y.A. Biosynthesis and processing of endogenous BDNF: CNS neurons store and secrete BDNF, not pro-BDNF. *Nat. Neurosci.* **2008**, *11*, 131–133. [[CrossRef](#)] [[PubMed](#)]
136. Yang, J.; Siao, C.J.; Nagappan, G.; Marinic, T.; Jing, D.; McGrath, K.; Chen, Z.Y.; Mark, W.; Tessarollo, L.; Lee, F.S.; et al. Neuronal release of ProBDNF. *Nat. Neurosci.* **2009**, *12*, 113–115. [[CrossRef](#)] [[PubMed](#)]
137. Chao, M. Neurotrophins and their receptors: A convergence point for many signalling pathways. *Nat. Rev. Neurosci.* **2003**, *4*, 299–309. [[CrossRef](#)] [[PubMed](#)]
138. Frade, J.M.; Rodriguez-Tebar, A.; Barde, Y.A. Induction of cell death by endogenous Nerve Growth Factor through its P75 receptor. *Nature* **1996**, *383*, 166–168. [[CrossRef](#)]
139. Lee, R.; Kermani, P.; Teng, K.K.; Hempstead, B.L. Regulation of cell survival by secreted proneurotrophins. *Science* **2001**, *294*, 1945–1948. [[CrossRef](#)]
140. Drevets, W.C.; Price, J.L.; Furey, M.L. Brain structural and functional abnormalities in mood disorders: Implications for neurocircuitry models of depression. *Brain Struct. Funct.* **2008**, *213*, 93–118. [[CrossRef](#)]
141. MacQueen, G.; Frodl, T. The hippocampus in major depression: Evidence for the convergence of the bench and bedside in psychiatric research? *Mol. Psych.* **2010**, *16*, 252–264. [[CrossRef](#)]
142. Duman, R.S.; Aghajanian, G.K.; Sanacora, G.; Krystal, J.H. Synaptic plasticity and depression: New insights from stress and Rapid-Acting Antidepressants. *Nat. Med.* **2016**, *22*, 238–249. [[CrossRef](#)] [[PubMed](#)]

143. Duman, R.S.; Monteggia, L.M. A Neurotrophic model for stress-related mood disorders. *Biol. Psychiatry* **2006**, *59*, 1116–1127. [[CrossRef](#)] [[PubMed](#)]
144. Dwivedi, Y. Brain-Derived Neurotrophic Factor: Role in depression and suicide. *Neuropsychiatr. Dis. Treat.* **2009**, *5*, 433–449. [[CrossRef](#)] [[PubMed](#)]
145. Martinowich, K.; Manji, H.; Lu, B. New insights into BDNF function in depression and anxiety. *Nat. Neurosci.* **2007**, *10*, 1089–1093. [[CrossRef](#)]
146. Zhao, S.; Rong, C.; Gao, Y.; Wu, L.; Luo, X.; Song, S.; Liu, Y.; Wong, J.H.; Wang, H.; Yi, L.; et al. Antidepressant-like effect of *Ganoderma lucidum* spore polysaccharide-peptide mediated by upregulation of prefrontal cortex Brain-Derived Neurotrophic Factor. *Appl. Microbiol. Biotechnol.* **2021**, *105*, 8675–8688. [[CrossRef](#)]
147. Ma, B.-J.; Shen, J.-W.; Yu, H.-Y.; Ruan, Y.; Wu, T.-T.; Zhao, X. Hericenones and erinacines: Stimulators of Nerve Growth Factor (NGF) biosynthesis in *Hericum erinaceus*. *Mycology* **2010**, *1*, 92–98. [[CrossRef](#)]
148. Li, I.C.; Lee, L.Y.; Tzeng, T.T.; Chen, W.P.; Chen, Y.P.; Shiao, Y.J.; Chen, C.C. Neurohealth properties of *Hericum erinaceus* mycelia enriched with erinacines. *Behav. Neurol.* **2018**, *2018*, 5802634. [[CrossRef](#)]
149. Lee, K.F.; Chen, J.H.; Teng, C.C.; Shen, C.H.; Hsieh, M.C.; Lu, C.C.; Lee, K.C.; Lee, L.Y.; Chen, W.P.; Chen, C.C.; et al. Protective effects of *Hericum erinaceus* mycelium and its isolated erinacine A against ischemia-injury-induced neuronal cell death via the inhibition of INOS/P38 MAPK and nitrotyrosine. *Int. J. Mol. Sci.* **2014**, *15*, 15073–15089. [[CrossRef](#)]
150. Chong, P.S.; Poon, C.H.; Roy, J.; Tsui, K.C.; Lew, S.Y.; Phang, M.W.L.; Tan, R.J.Y.; Cheng, P.G.; Fung, M.L.; Wong, K.H.; et al. Neurogenesis-dependent antidepressant-like activity of *Hericum erinaceus* in an animal model of depression. *Chin. Med.* **2021**, *16*, 132. [[CrossRef](#)]
151. Ryu, S.H.; Hong, S.M.; Khan, Z.; Lee, S.K.; Vishwanath, M.; Turk, A.; Yeon, S.W.; Jo, Y.H.; Lee, D.H.; Lee, J.K.; et al. Neurotrophic isoindolinones from the fruiting bodies of *Hericum erinaceus*. *Bioorg. Med. Chem. Lett.* **2021**, *31*, 127714. [[CrossRef](#)]
152. Nagano, M.; Shimizu, K.; Kondo, R.; Hayashi, C.; Sato, D.; Kitagawa, K.; Ohnuki, K. Reduction of depression and anxiety by 4 weeks *Hericum erinaceus* intake. *Biomed. Res.* **2010**, *31*, 231–237. [[CrossRef](#)] [[PubMed](#)]
153. Chan, Y.C.; Lin, T.C.; Chen, C.C.; Lee, L.Y.; Chen, W.P.; Liu, Y.Z.; Hwang, J.H. Effects of erinacine A-enriched *Hericum erinaceus* on elderly hearing-impaired patients: A double-blind, randomized, placebo-controlled clinical trial. *J. Funct. Foods* **2022**, *97*, 105220. [[CrossRef](#)]
154. Wittstein, K.; Rascher, M.; Rupcic, Z.; Löwen, E.; Winter, B.; Köster, R.W.; Stadler, M. Coralloicins A-C, Nerve Growth and Brain-Derived Neurotrophic Factor inducing metabolites from the mushroom *Hericum coralloides*. *J. Nat. Prod.* **2016**, *79*, 2264–2269. [[CrossRef](#)] [[PubMed](#)]
155. Kou, R.-W.; Xia, B.; Han, R.; Li, Z.-Q.; Yang, J.-R.; Yin, X.; Gao, Y.-Q.; Gao, J.-M. Neuroprotective effects of a new triterpenoid from edible mushroom on oxidative stress and apoptosis through the BDNF/TrkB/ERK/CREB and Nrf2 signaling pathway in vitro and in vivo. *Food Funct.* **2022**, *13*, 12121–12134. [[CrossRef](#)] [[PubMed](#)]
156. Hu, M.; Zhang, P.; Wang, R.; Zhou, M.; Pang, N.; Cui, X.; Ge, X.; Liu, X.; Huang, X.F.; Yu, Y. Three different types of β -glucans enhance cognition: The role of the gut-brain axis. *Front. Nutr.* **2022**, *9*, 312. [[CrossRef](#)]
157. Hashim, H.; Azmin, S.; Razlan, H.; Yahya, N.W.; Tan, H.J.; Manaf, M.R.A.; Ibrahim, N.M. Eradication of *Helicobacter pylori* infection improves levodopa action, clinical symptoms and quality of life in patients with Parkinson's Disease. *PLoS ONE* **2014**, *9*, e112330. [[CrossRef](#)]
158. Rafii, F.; Sutherland, J.B.; Hansen, E.B.; Cerniglia, C.E. Reduction of nitrazepam by *Clostridium leptum*, a nitroreductase-producing bacterium isolated from the human intestinal tract. *Clin. Infect. Dis.* **1997**, *25* (Suppl. 2), S121–S122. [[CrossRef](#)]
159. Elmer, G.W.; Rummel, R.P. Role of the intestinal microflora in clonazepam metabolism in the rat. *Xenobiotica* **1984**, *14*, 829–840. [[CrossRef](#)]
160. Kitamura, S.; Sugihara, K.; Kuwasako, M.; Tatsumi, K. The role of mammalian intestinal bacteria in the reductive metabolism of zonisamide. *J. Pharm. Pharmacol.* **1997**, *49*, 253–256. [[CrossRef](#)]
161. Roy Sarkar, S.; Banerjee, S. Gut microbiota in neurodegenerative disorders. *J. Neuroimmunol.* **2019**, *328*, 98–104. [[CrossRef](#)]
162. Belizário, J.E.; Faintuch, J.; Garay-Malpartida, M. New frontiers for treatment of metabolic diseases. *Mediat. Inflamm.* **2018**, *2018*, 2037838. [[CrossRef](#)] [[PubMed](#)]
163. Devkota, S.; Wang, Y.; Musch, M.W.; Leone, V.; Fehlner-Peach, H.; Nadimpalli, A.; Antonopoulos, D.A.; Jabri, B.; Chang, E.B. Dietary-fat-induced taurocholic acid promotes pathobiont expansion and colitis in Il10^{-/-} Mice. *Nature* **2012**, *487*, 104–108. [[CrossRef](#)] [[PubMed](#)]
164. Pedersen, C.; Ijaz, U.Z.; Gallagher, E.; Horton, F.; Ellis, R.J.; Jaiyeola, E.; Duparc, T.; Russell-Jones, D.; Hinton, P.; Cani, P.D.; et al. Fecal Enterobacteriales enrichment is associated with increased in vivo intestinal permeability in humans. *Physiol. Rep.* **2018**, *6*, e13649. [[CrossRef](#)] [[PubMed](#)]
165. Bengtsson, S. Gut microbiota, immune development and function. *Pharmacol. Res.* **2013**, *69*, 87–113. [[CrossRef](#)] [[PubMed](#)]
166. Van den Abbeele, P.; Belzer, C.; Goossens, M.; Kleerebezem, M.; de Vos, W.M.; Thas, O.; de Weirdt, R.; Kerckhof, F.M.; van de Wiele, T. Butyrate-producing *Clostridium* Cluster XIVa species specifically colonize mucins in an in vitro gut model. *ISME J.* **2013**, *7*, 949. [[CrossRef](#)] [[PubMed](#)]
167. Rivièrè, A.; Selak, M.; Lantin, D.; Leroy, F.; de Vuyst, L. Bifidobacteria and butyrate-producing colon bacteria: Importance and strategies for their stimulation in the human gut. *Front. Microbiol.* **2016**, *7*, 979. [[CrossRef](#)]

168. Sakon, H.; Nagai, F.; Morotomi, M.; Tanaka, R. *Sutterella parvirubra* Sp. Nov. and *Megamonas funiformis* Sp. Nov., Isolated from human faeces. *Int. J. Syst. Evol. Microbiol.* **2008**, *58*, 970–975. [[CrossRef](#)]
169. Stilling, R.M.; Dinan, T.G.; Cryan, J.F. Microbial genes, brain & behaviour—epigenetic regulation of the gut-brain axis. *Genes Brain Behav.* **2014**, *13*, 69–86. [[CrossRef](#)]
170. Li, N.; Wang, Q.; Wang, Y.; Sun, A.; Lin, Y.; Jin, Y.; Li, X. Fecal microbiota transplantation from chronic unpredictable mild stress mice donors affects anxiety-like and depression-like behavior in recipient mice via the gut microbiota-inflammation-brain axis. *Stress* **2019**, *22*, 592–602. [[CrossRef](#)]
171. Jiang, H.; Ling, Z.; Zhang, Y.; Mao, H.; Ma, Z.; Yin, Y.; Wang, W.; Tang, W.; Tan, Z.; Shi, J.; et al. Altered fecal microbiota composition in patients with Major Depressive Disorder. *Brain Behav. Immun.* **2015**, *48*, 186–194. [[CrossRef](#)]
172. Simpson, C.A.; Diaz-Arteche, C.; Eliby, D.; Schwartz, O.S.; Simmons, J.G.; Cowan, C.S.M. The gut microbiota in anxiety and depression—A systematic review. *Clin. Psychol. Rev.* **2021**, *83*, 101943. [[CrossRef](#)]
173. Li, C. Understanding interactions among diet, host and gut microbiota for personalized nutrition. *Life Sci.* **2023**, *312*, 121265. [[CrossRef](#)] [[PubMed](#)]
174. Watson, H.; Mitra, S.; Croden, F.C.; Taylor, M.; Wood, H.M.; Perry, S.L.; Spencer, J.A.; Quirke, P.; Toogood, G.J.; Lawton, C.L.; et al. A randomised trial of the effect of omega-3 polyunsaturated fatty acid supplements on the human intestinal microbiota. *Gut* **2018**, *67*, 1974–1983. [[CrossRef](#)] [[PubMed](#)]
175. Desai, M.S.; Seekatz, A.M.; Koropatkin, N.M.; Kamada, N.; Hickey, C.A.; Wolter, M.; Pudlo, N.A.; Kitamoto, S.; Terrapon, N.; Muller, A.; et al. A dietary fiber-deprived gut microbiota degrades the colonic mucus barrier and enhances pathogen susceptibility. *Cell* **2016**, *167*, 1339–1353.e21. [[CrossRef](#)] [[PubMed](#)]
176. Scott, K.P.; Tuohy, K.M.; Mach -Istituto, F.E.; San, A.; All’adige, M.; Gibson, G.R.; Rastall, R.A.; Hotchkiss, A.; Dubert-Ferrandon, A.; Gareau, M.; et al. Bioactive food ingredients for safe and health-promoting functional foods view project neurodevelopmental impact of preterm birth in a porcine model view project dietary prebiotics: Current status and new definition. *Food Sci. Technol. Bull.* **2010**, *7*, 1–19. [[CrossRef](#)]
177. Zhang, S.; Lei, L.; Zhou, Y.; Ye, F.; Zhao, G. Roles of mushroom polysaccharides in chronic disease management. *J. Integr. Agric.* **2022**, *21*, 1839–1866. [[CrossRef](#)]
178. Li, M.; Yu, L.; Zhai, Q.; Liu, B.; Zhao, J.; Chen, W.; Tian, F. *Ganoderma lucidum* ethanol extraction promotes dextran sulphate sodium induced colitis recovery and modulation in microbiota. *Foods* **2022**, *11*, 4023. [[CrossRef](#)] [[PubMed](#)]
179. Yao, C.; Wang, Z.; Jiang, H.; Yan, R.; Huang, Q.; Wang, Y.; Xie, H.; Zou, Y.; Yu, Y.; Lv, L. *Ganoderma lucidum* promotes sleep through a gut microbiota-dependent and serotonin-involved pathway in mice. *Sci. Rep.* **2021**, *11*, 13660. [[CrossRef](#)]
180. Romero-Córdoba, S.L.; Salido-Guadarrama, I.; Meneses, M.E.; Cosentino, G.; Iorio, M.V.; Tagliabue, E.; Torres, N.; Sánchez-Tapia, M.; Bonilla, M.; Castillo, I.; et al. Mexican *Ganoderma lucidum* extracts decrease lipogenesis modulating transcriptional metabolic networks and gut microbiota in C57BL/6 mice fed with a high-cholesterol diet. *Nutrients* **2020**, *13*, 38. [[CrossRef](#)]
181. Chen, M.; Xiao, D.; Liu, W.; Song, Y.; Zou, B.; Li, L.; Li, P.; Cai, Y.; Liu, D.; Liao, Q.; et al. Intake of *Ganoderma lucidum* polysaccharides reverses the disturbed gut microbiota and metabolism in type 2 diabetic rats. *Int. J. Biol. Macromol.* **2020**, *155*, 890–902. [[CrossRef](#)]
182. Lu, C.-L.; Lee, B.H.; Ren, Y.-L.; Ji, D.; Rao, S.-Q.; Li, H.-X.; Yang, Z.-Q. Effects of exopolysaccharides from *Antrodia cinnamomea* on inflammation and intestinal microbiota disturbance induced by antibiotics in mice. *Food Biosci.* **2022**, *50*, 102116. [[CrossRef](#)]
183. Ye, J.; Zhang, C.; Fan, Q.; Lin, X.; Wang, Y.; Azzam, M.; Alhotan, R.; Alqhtani, A.; Jiang, S. *Antrodia cinnamomea* polysaccharide improves liver antioxidant, anti-inflammatory capacity, and cecal flora structure of slow-growing broiler breeds challenged with lipopolysaccharide. *Front. Vet. Sci.* **2022**, *9*, 1559. [[CrossRef](#)] [[PubMed](#)]
184. Rehman, A.U.; Khan, A.I.; Xin, Y.; Liang, W. *Morchella esculenta* polysaccharide attenuate obesity, inflammation and modulate gut microbiota. *AMB Express* **2022**, *12*, 114. [[CrossRef](#)]
185. Wang, T.; Han, J.; Dai, H.; Sun, J.; Ren, J.; Wang, W.; Qiao, S.; Liu, C.; Sun, L.; Liu, S.; et al. Polysaccharides from *Lyophyllum decastes* reduce obesity by altering gut microbiota and increasing energy expenditure. *Carbohydr. Polym.* **2022**, *295*, 119862. [[CrossRef](#)] [[PubMed](#)]
186. Lai, Y.; Yu, H.; Deng, H.; Fang, Q.; Lei, H.; Liu, L.; Wu, N.; Guo, X.; Song, C. Three main metabolites from *Wolfiporia cocos* (F. A. Wolf) Ryvarden & Gilb regulate the gut microbiota in mice: A comparative study using microbiome-metabolomics. *Front. Pharmacol.* **2022**, *13*, 2852. [[CrossRef](#)]
187. Gao, X.; Feng, Y.; Xue, H.; Meng, M.; Qin, X. Antidepressant-like effect of triterpenoids extracts from *Poria Cocos* on the CUMS rats by 16S RRNA gene sequencing and LC-MS metabolomics. *J. Liq. Chromatogr. Relat. Technol.* **2020**, *43*, 494–507. [[CrossRef](#)]
188. Huang, Y.; Gao, Y.; Pi, X.; Zhao, S.; Liu, W. In vitro hepatoprotective and human gut microbiota modulation of polysaccharide-peptides in *Pleurotus citrinopileatus*. *Front. Cell Infect. Microbiol.* **2022**, *12*, 892049. [[CrossRef](#)]
189. Nakahara, D.; Nan, C.; Mori, K.; Hanayama, M.; Kikuchi, H.; Hirai, S.; Egashira, Y. Effect of mushroom polysaccharides from *Pleurotus eryngii* on obesity and gut microbiota in mice fed a high-fat diet. *Eur. J. Nutr.* **2020**, *59*, 3231–3244. [[CrossRef](#)]
190. Hu, Q.; Yuan, B.; Wu, X.; Du, H.; Gu, M.; Han, Y.; Yang, W.; Song, M.; Xiao, H. Dietary intake of *Pleurotus eryngii* ameliorated dextran-sodium-sulfate-induced colitis in mice. *Mol. Nutr. Food Res.* **2019**, *63*, 1801265. [[CrossRef](#)]
191. Hu, Y.; Xu, J.; Sheng, Y.; Liu, J.; Li, H.; Guo, M.; Xu, W.; Luo, Y.; Huang, K.; He, X. *Pleurotus ostreatus* ameliorates obesity by modulating the gut microbiota in obese mice induced by high-fat diet. *Nutrients* **2022**, *14*, 1868. [[CrossRef](#)]

192. Adams, S.; Che, D.; Hailong, J.; Zhao, B.; Rui, H.; Danquah, K.; Qin, G. Effects of pulverized oyster mushroom (*Pleurotus ostreatus*) on diarrhea incidence, growth performance, immunity, and microbial composition in piglets. *J. Sci. Food Agric.* **2019**, *99*, 3616–3627. [[CrossRef](#)] [[PubMed](#)]
193. Maheshwari, G.; Gessner, D.K.; Neuhaus, K.; Most, E.; Zorn, H.; Eder, K.; Ringseis, R. Influence of a biotechnologically produced oyster mushroom (*Pleurotus Sajor-Caju*) on the gut microbiota and microbial metabolites in obese Zucker rats. *J. Agric. Food Chem.* **2021**, *69*, 1524–1535. [[CrossRef](#)] [[PubMed](#)]
194. Hess, J.; Wang, Q.; Gould, T.; Slavin, J. Impact of *Agaricus bisporus* mushroom consumption on gut health markers in healthy adults. *Nutrients* **2018**, *10*, 1402. [[CrossRef](#)] [[PubMed](#)]
195. García-Sanmartín, J.; Bobadilla, M.; Mirpuri, E.; Grifoll, V.; Pérez-Clavijo, M.; Martínez, A. Agaricus mushroom-enriched diets modulate the microbiota-gut-brain axis and reduce brain oxidative stress in mice. *Antioxidants* **2022**, *11*, 695. [[CrossRef](#)]
196. Yao, L.; Lv, J.; Duan, C.; An, X.; Zhang, C.; Li, D.; Li, C.; Liu, S. *Armillaria mellea* fermentation liquor ameliorates p-chlorophenylalanine-induced insomnia associated with the modulation of serotonergic system and gut microbiota in rats. *J. Food Biochem.* **2022**, *46*, e14075. [[CrossRef](#)]
197. Yang, X.; Zheng, M.; Zhou, M.; Zhou, L.; Ge, X.; Pang, N.; Li, H.; Li, X.; Li, M.; Zhang, J.; et al. Lentinan supplementation protects the gut–liver axis and prevents steatohepatitis: The role of gut microbiota involved. *Front. Nutr.* **2022**, *8*, 1286. [[CrossRef](#)]
198. Zou, M.; Wang, Y.; Liu, Y.; Xiong, S.; Zhang, L.; Wang, J. Huangshan floral mushroom polysaccharide ameliorates dextran sulfate sodium-induced colitis in mice by modulating Th17/Treg balance in a gut microbiota-dependent manner. *Mol. Nutr. Food Res.* **2022**, *67*, 2200408. [[CrossRef](#)]
199. Cho, H.-W.; Choi, S.; Seo, K.; Kim, K.H.; Jeon, J.-H.; Kim, C.H.; Lim, S.; Jeong, S.; Chun, J.L. Gut microbiota profiling in aged dogs after feeding pet food contained *Hericium erinaceus*. *J. Anim. Sci. Technol.* **2022**, *64*, 937–949. [[CrossRef](#)]
200. Xie, X.Q.; Geng, Y.; Guan, Q.; Ren, Y.; Guo, L.; Lv, Q.; Lu, Z.M.; Shi, J.S.; Xu, Z.H. Influence of short-term consumption of *Hericium erinaceus* on serum biochemical markers and the changes of the gut microbiota: A pilot study. *Nutrients* **2021**, *13*, 1008. [[CrossRef](#)]
201. Lai, Y.; Fang, Q.; Guo, X.; Lei, H.; Zhou, Q.; Wu, N.; Song, C. Effect of polysaccharides from *Dictyophora indusiata* on regulating gut microbiota and short-chain fatty acids in mice. *J. Food Meas. Charact.* **2022**, *17*, 1–11. [[CrossRef](#)]
202. Kanwal, S.; Aliya, S.; Xin, Y. Anti-obesity effect of *Dictyophora indusiata* mushroom polysaccharide (DIP) in high fat diet-induced obesity via regulating inflammatory cascades and intestinal microbiome. *Front. Endocrinol.* **2020**, *11*, 558874. [[CrossRef](#)] [[PubMed](#)]
203. Abdureyim, Z.; Wang, L.; Tao, Q.; Xu, J.; Yimiti, D.; Gao, Q. Bachu mushroom polysaccharide alleviates colonic injury by modulating the gut microbiota. *Comput. Math. Methods Med.* **2022**, *2022*, 1353724. [[CrossRef](#)] [[PubMed](#)]
204. Xu, Y.; Xie, L.; Zhang, Z.; Zhang, W.; Tang, J.; He, X.; Zhou, J.; Peng, W. *Tremella fuciformis* polysaccharides inhibited colonic inflammation in dextran sulfate sodium-treated mice via Foxp3+ T cells, gut microbiota, and bacterial metabolites. *Front. Immunol.* **2021**, *12*, 948. [[CrossRef](#)] [[PubMed](#)]
205. Li, L.; Guo, W.L.; Zhang, W.; Xu, J.X.; Qian, M.; Bai, W.D.; Zhang, Y.Y.; Rao, P.F.; Ni, L.; Lv, X.C. *Grifola frondosa* polysaccharides ameliorate lipid metabolic disorders and gut microbiota dysbiosis in high-fat diet fed rats. *Food Funct.* **2019**, *10*, 2560–2572. [[CrossRef](#)]
206. Chen, Y.; Liu, D.; Wang, D.; Lai, S.; Zhong, R.; Liu, Y.; Yang, C.; Liu, B.; Sarker, M.R.; Zhao, C. Hypoglycemic activity and gut microbiota regulation of a novel polysaccharide from *Grifola frondosa* in type 2 diabetic mice. *Food Chem. Toxicol.* **2019**, *126*, 295–302. [[CrossRef](#)]
207. Finotello, F.; Mastrorilli, E.; di Camillo, B. Measuring the diversity of the human microbiota with targeted next-generation sequencing. *Brief. Bioinform.* **2018**, *19*, 679–692. [[CrossRef](#)]
208. Di Pierro, F. Gut microbiota parameters potentially useful in clinical perspective. *Microorganisms* **2021**, *9*, 2402. [[CrossRef](#)]
209. Bischoff, S.C.; Barbara, G.; Buurman, W.; Ockhuizen, T.; Schulzke, J.D.; Serino, M.; Tilg, H.; Watson, A.; Wells, J.M. Intestinal permeability—A new target for disease prevention and therapy. *BMC Gastroenterol.* **2014**, *14*, 189. [[CrossRef](#)]
210. Kerckhoffs, A.P.M.; Akkermans, L.M.A.; de Smet, M.B.M.; Besselink, M.G.H.; Hietbrink, F.; Bartelink, I.H.; Busschers, W.B.; Samsom, M.; Renooij, W. Intestinal permeability in irritable bowel syndrome patients: Effects of NSAIDs. *Dig. Dis. Sci.* **2010**, *55*, 716–723. [[CrossRef](#)]
211. Ley, R.E.; Turnbaugh, P.J.; Klein, S.; Gordon, J.I. Human gut microbes associated with obesity. *Nature* **2006**, *444*, 1022–1023. [[CrossRef](#)]
212. Turnbaugh, P.J.; Ley, R.E.; Mahowald, M.A.; Magrini, V.; Mardis, E.R.; Gordon, J.I. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* **2006**, *444*, 1027–1031. [[CrossRef](#)]
213. Qin, J.; Li, Y.; Cai, Z.; Li, S.; Zhu, J.; Zhang, F.; Liang, S.; Zhang, W.; Guan, Y.; Shen, D.; et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature* **2012**, *490*, 55–60. [[CrossRef](#)] [[PubMed](#)]
214. Madison, A.; Kiecolt-Glaser, J.K. Stress, depression, diet, and the gut microbiota: Human–bacteria interactions at the core of psychoneuroimmunology and nutrition. *Curr. Opin. Behav. Sci.* **2019**, *28*, 105. [[CrossRef](#)] [[PubMed](#)]
215. Ohlsson, L.; Gustafsson, A.; Lavant, E.; Suneson, K.; Brundin, L.; Westrin, Å.; Ljunggren, L.; Lindqvist, D. Leaky gut biomarkers in depression and suicidal behavior. *Acta Psychiatr. Scand.* **2019**, *139*, 185. [[CrossRef](#)] [[PubMed](#)]
216. Peirce, J.M.; Alviña, K. The role of inflammation and the gut microbiome in depression and anxiety. *J. Neurosci. Res.* **2019**, *97*, 1223–1241. [[CrossRef](#)]

217. Kris-Etherton, P.M.; Petersen, K.S.; Hibbeln, J.R.; Hurley, D.; Kolick, V.; Peoples, S.; Rodriguez, N.; Woodward-Lopez, G. Nutrition and behavioral health disorders: Depression and anxiety. *Nutr. Rev.* **2021**, *79*, 247–260. [[CrossRef](#)]
218. Sánchez-Villegas, A.; Martínez-González, M.A.; Estruch, R.; Salas-Salvadó, J.; Corella, D.; Covas, M.I.; Arós, F.; Romaguera, D.; Gómez-Gracia, E.; Lapetra, J.; et al. Mediterranean dietary pattern and depression: The PREDIMED Randomized Trial. *BMC Med.* **2013**, *11*, 208. [[CrossRef](#)]
219. Appleton, K.M.; Sallis, H.M.; Perry, R.; Ness, A.R.; Churchill, R. Omega-3 fatty acids for depression in adults. *Cochrane Database Syst. Rev.* **2015**, *2015*, CD004692. [[CrossRef](#)]
220. Sofi, F.; Macchi, C.; Abbate, R.; Gensini, G.F.; Casini, A. Mediterranean diet and health. *BioFactors* **2013**, *39*, 335–342. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.