



## Article

# The Effect of Holder Pasteurization and Different Variants on Breast Milk Antioxidants

Réka Anna Vass<sup>1,2,3,\*</sup>, Éva Mikó<sup>2,4</sup>, Csenge Gál<sup>2</sup>, Tamás Kószegi<sup>2,5</sup>, Csaba I. Vass<sup>3</sup>, Szilvia Bokor<sup>2,6</sup>, Dénes Molnár<sup>2,6</sup> , Simone Funke<sup>1,2</sup>, Kálmán Kovács<sup>1,2</sup>, József Bódis<sup>1,2,7</sup> and Tibor Ertl<sup>1,2</sup>

<sup>1</sup> Department of Obstetrics and Gynecology, Medical School University of Pécs, 7624 Pécs, Hungary

<sup>2</sup> National Laboratory on Human Reproduction, University of Pécs, 7624 Pécs, Hungary

<sup>3</sup> Obstetrics and Gynecology, Magyar Imre Hospital, 8400 Ajka, Hungary

<sup>4</sup> Department of Microbiology, Medical School University of Pécs, 7624 Pécs, Hungary

<sup>5</sup> Department of Laboratory Medicine, Medical School University of Pécs, 7624 Pécs, Hungary

<sup>6</sup> Department of Pediatrics, Medical School University of Pécs, 7624 Pécs, Hungary

<sup>7</sup> HUN-REN-PTE Human Reproduction Research Group, 7624 Pécs, Hungary

\* Correspondence: vass.reka@pte.hu; Tel.: +36-30-253-2000

**Abstract:** Background: After birth, breast milk (BM) is a known essential source of antioxidants for infants. We analyzed the non-enzymatic total antioxidant capacity (TAC), oxygen radical absorbance capacity (ORAC), and glutathione, calcium, transferrin, and total protein levels of human breast milk before and after Holder pasteurization (HoP). Methods: The collected donor BM samples were pasteurized with HoP. Results: HoP decreased TAC (−12.6%), ORAC (−12.1%), transferrin (−98.3%), and total protein (−21.4%) levels; HoP did not influence the glutathione concentration, and it increased the total calcium (+25.5%) concentration. Mothers who gave birth via Cesarean section had significantly lower TAC in their BM. TAC and glutathione levels were elevated in the BM of mothers over the age of 30. BM produced in the summer had higher glutathione and calcium levels compared to BM produced in the winter. The glutathione concentration in term milk samples was significantly higher in the first two months of lactation compared to the period between the third and sixth months. The transferrin level of BM for female infants was significantly higher than the BM for boys, and mothers with a BMI above 30 had increased transferrin in their samples. Conclusions: Antioxidant levels in human milk are influenced by numerous factors. Environmental and maternal factors, the postpartum age at breast milk collection, and Holder pasteurization of the milk influence the antioxidant intake of the infant.

**Keywords:** glutathione; TAC; ORAC; transferrin; calcium; total protein; breast milk; donor milk



**Citation:** Vass, R.A.; Mikó, É.; Gál, C.; Kószegi, T.; Vass, C.I.; Bokor, S.; Molnár, D.; Funke, S.; Kovács, K.; Bódis, J.; et al. The Effect of Holder Pasteurization and Different Variants on Breast Milk Antioxidants. *Antioxidants* **2023**, *12*, 1857. <https://doi.org/10.3390/antiox12101857>

Academic Editor: Alessandra Napolitano

Received: 14 August 2023  
Revised: 28 September 2023  
Accepted: 10 October 2023  
Published: 13 October 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Human breast milk (BM) provides all the necessary macro- and micronutrients and many of the non-nutritive bioactive molecules needed for an infant's development, survival, and well-being [1]. Antioxidants are compounds that help to protect the body against damage caused by free radicals, which are unstable molecules that harm cells and contribute to the development of disease. Reactive oxygen species (ROS) contribute to biological homeostasis and play a significant role in cell signaling in both psychological and pathophysiological processes, but they also cause molecular or even cell damage, necrosis, apoptosis, and DNA oxidation [2,3]. Increased oxidative stress is a known causative factor for mortality, bronchopulmonary dysplasia (BPD), and retinopathy of prematurity (ROP), especially in very-low-birth-weight infants [4–6]. Total antioxidant capacity (TAC) refers to the overall antioxidant capacity of a substance, including the combined effects of various antioxidants. In the context of breast milk, non-enzymatic TAC represents the collective antioxidant capacity of the antioxidants present in breast milk [7,8]. These antioxidants

include vitamins (such as vitamin C, vitamin E, and beta-carotene), minerals (such as selenium and zinc), and other bioactive compounds (such as bilirubin) [9]. The non-enzymatic antioxidants in breast milk protect the infant's cells from oxidative damage, support the developing immune system, and support physiological development [6]. The oxygen radical absorbance capacity (ORAC) assay measures a fluorescent signal that is quenched in the presence of ROS [10]. BM TAC can be estimated with ORAC, a standardized and validated method to measure the antioxidant capacity in biological samples *in vitro*. It assesses the ability of antioxidants to neutralize free radicals and reduce oxidative stress [8–10]. BM contains a dynamic and diverse array of anti- and prooxidants, and their interactions affect the ORAC level [8].

Glutathione is a powerful antioxidant that is naturally present in the human body. It plays a crucial role in protecting cells from oxidative damage caused by free radicals and toxins. Glutathione plays a crucial role in supporting the immune system by regulating immune responses, thereby enhancing the function of immune cells. In breastfed infants, glutathione in breast milk may contribute to their ability to handle and eliminate certain toxins more effectively [11,12].

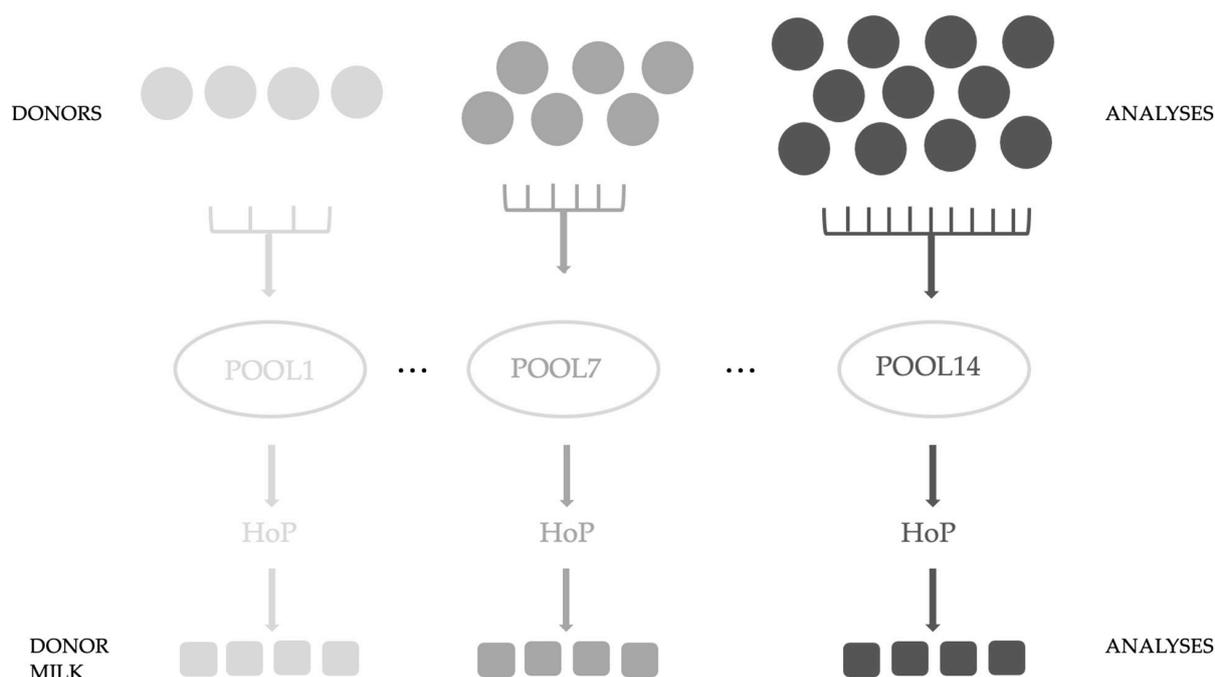
Transferrin is a protein that plays a vital role in iron transport and absorption [13]. While transferrin is primarily found in the blood, it is also present in small amounts in breast milk [14]. Transferrin in breast milk helps facilitate the absorption and utilization of iron by the infant. It is necessary for the production of red blood cells and for the proper functioning of various enzymes and metabolic processes. Once absorbed, iron is utilized for various physiological processes, including the production of hemoglobin and the support of overall growth and development. Iron is an integral component of many proteins and enzymes most relevant in our context of peroxidase and catalase [15]. Iron concentrations in human milk are low (0.2–0.4 mg/L), yet the iron in milk is highly bioavailable [16].

The calcium concentration in breast milk does not change with the stage of lactation. A previous review and meta-analysis reported no significant differences in milk calcium concentration between lactation stages, adolescent and adult mothers, preterm and term infants, exclusive and mixed breastfeeding, with or without calcium supplementation, between nutritional statuses, country income categories, continents, and measurement methods in calcium concentration [17].

Breast milk is important for premature and mature infants as it provides the nutrients and immune-boosting factors needed for their development and helps to reduce the risk of complications [18]. Antioxidants in BM directly influence the development of the gastrointestinal tract and, through absorption, impact organ maturation and infant development [2,6,7]. Since their intrauterine development is disrupted due to preterm birth, preterm infants are predisposed to oxidative stress, and they need certain maternal protective factors. After birth, BM is the exclusive source of these maternal protective compounds. When own mother's milk is not available, donor milk is considered the best feeding alternative.

## 2. Materials and Methods

In this study, registered and approved donor mothers of the Breast Milk Collection Center (BMCC) (Unified Health Institution at Pécs, Hungary) were recruited, who, following the center's protocol, donated freshly pumped milk ( $n = 122$ ). Our study was conducted with the approval of the Regional and Local Research Ethics Committee of the University of Pécs, Pécs, Hungary (PTE KK 7072-2018). Waivers for participant consent were obtained. For analysis, 3 mL was poured and stored separately at  $-80\text{ }^{\circ}\text{C}$  until laboratory measurements. The protocol of the BMCC was followed during our study. Our aim was to examine the effect of HoP on donor milk samples. Pool sizes were variable from 4 to 11 samples; we examined 14 pools. Figure 1. shows the experimental design (Figure 1.)



**Figure 1.** Experimental design of our study.

The samples were analyzed first individually and after they were pooled and Holder pasteurized (30 min at 62.5 °C) in the laboratory of the Unified Health Institution. We took five samples for later analyses; three samples were used in the present experiment. All samples from the pooled and Holder pasteurized donor milk were stored at  $-80$  °C until laboratory measurements were taken. First, we sonicated the BM samples and centrifuged them at  $15,000 \times g$  for 15 min. The skimmed milk was transferred for analysis according to the previously described preparation methods [19,20]. For the measurement of glutathione, every sample was analyzed, while the other factors were detected in the first 6 pools.

For TAC determination, two different assays were used: enhanced chemiluminescence (ECL) and ORAC. In the ECL assay, a fully validated luminol-peroxidase-4-iodophenol-hydrogen peroxide-based technique was applied [10]. In all analyses, first 20  $\mu\text{L}$  of blank/standard/sample and then 270  $\mu\text{L}$  of horseradish peroxidase-ECL reagent were pipetted into 96-well white optical plates (Optiplates, Per-Form Hungaria Ltd., Budapest, Hungary). Then, 20  $\mu\text{L}$  of hydrogen peroxide solution was injected into the wells by a Biotek Synergy HT plate reader (Agilent, Santa Clara, CA, USA) and the developing luminescence signals were monitored kinetically for 10 min. Measurements were carried out in duplicate. A standard curve for Trolox calibrators was established by using the area under the curve (AUC) of the luminescence signals, and the TAC of the samples was calculated from the equation of the standard curve. The TAC values of the samples were given as Trolox equivalent in  $\mu\text{mol/L}$  or  $\text{mmol/L}$ .

For the ORAC technique, 25  $\mu\text{L}$  of blank/standard/sample was mixed with 150  $\mu\text{L}$   $\text{Na}_2$ - fluorescein in a black optical plate (Optiplates), and 25  $\mu\text{L}$  of AAPH oxidant (2,2'-azo-bis(2-amidinopropane) dihydrochloride, Merck, Darmstadt, Germany) was injected into the wells by the Biotek Synergy HT plate reader. Kinetic measurement of fluorescence quenching was performed at 490/520 nm wavelengths for 80 min [10]. For the calculation, the AUC values obtained for the blanks/standards/samples were used, and the TAC was calculated as described for the ECL assay. The BM samples were diluted 20-fold with tri-distilled water, and measurements were carried out in duplicate. The TAC, ORAC, calcium, transferrin, and total protein concentration were detected in 59 breast milk samples at the fully accredited Department of Laboratory Medicine, University of Pécs.

The total glutathione level was detected with a colorimetric detection kit (ThermoFisher Scientific, Frederick, MD, USA) based on the manufacturer's instructions. Next,

50  $\mu\text{L}$  standards or samples were added to the wells. After additional steps, the absorbance was read at 405 nm, and the concentrations were expressed in mM. The total glutathione level was measured in 122 samples.

Calcium, transferrin, and total protein levels were measured using a fully automatized Cobas c analyzer system (Roche Diagnostics, Mannheim, Germany). The lower limit of detection for calcium was 0.20 mmol/L, for transferrin, it was 1.5 mg/L, and for total protein, it was 40 mg/L.

To test the data normality, Shapiro–Wilk tests were performed with GraphPad (La Jolla, CA, USA). Paired *t*-tests or *t*-tests were used for further analysis. The repeated measures one-way ANOVA test with the post hoc Dunnett’s test was applied to compare the effect of Holder pasteurization. Differences were considered statistically significant if the *p*-values were <0.05. The study was powered to detect moderate effect sizes (Cohen’s *d* = 0.6). The results are presented as the mean  $\pm$  SEM. Maternal age and body mass index (BMI), infant gender, seasonal differences, and duration of lactation at the time of sampling were also analyzed.

### 3. Results

For the measurement of TAC, ORAC, transferrin, calcium, and total protein, we analyzed 59 BM samples. The mean maternal age was  $32.2 \pm 0.6$  years, the mean BMI was  $25.2 \pm 0.5$ , and the mean infant gestational age was  $38.3 \pm 0.4$  weeks. In total, 25 BM samples were donated by mothers who had undergone a Cesarean section (CS), and 34 samples were collected after spontaneous delivery. Female infants were delivered by 27 mothers, and 32 mothers gave birth to male infants.

The total glutathione level was measured in 122 BM samples. The average maternal age of the donors for these samples was  $32.8 \pm 0.4$  years, the average maternal BMI was  $26.4 \pm 0.6$ , and the average gestational age of the newborns was  $38.9 \pm 0.2$  weeks. In this study, there were 52 samples donated after CS, while after spontaneous delivery, 70 samples were donated. Out of 122 samples, 57 were produced for female infants and 65 were produced for males.

After HoP, their glutathione levels did not change; however, their TAC, ORAC, total protein, and transferrin levels were significantly lower after HoP. Their calcium concentration was higher after HoP (Table 1).

**Table 1.** The effect of Holder pasteurization (HoP) on different factors.

	Raw	HoP	<i>p</i> -Value
Glutathione (mM) (n = 112)	0.11 $\pm$ 0.02	0.12 $\pm$ 0.10	0.575
TAC-ECL ( $\mu\text{M}$ ) (n = 59)	127.31 $\pm$ 6.24	111.27 $\pm$ 5.26	0.028
TAC-ORAC ( $\mu\text{M}$ ) (n = 59)	3602.34 $\pm$ 104.13	3163.13 $\pm$ 787.94	0.001
Calcium (mM) (n = 59)	5.08 $\pm$ 0.15	6.38 $\pm$ 0.10	<0.0001
Total protein (g/L) (n = 59)	4.91 $\pm$ 0.16	3.86 $\pm$ 0.11	<0.0001
Transferrin (mg/L) (n = 59)	80.82 $\pm$ 10.85	1.34 $\pm$ 0.09	<0.0001

HoP: Holder pasteurization; TAC: total antioxidant capacity; ORAC: oxygen radical absorbance capacity.

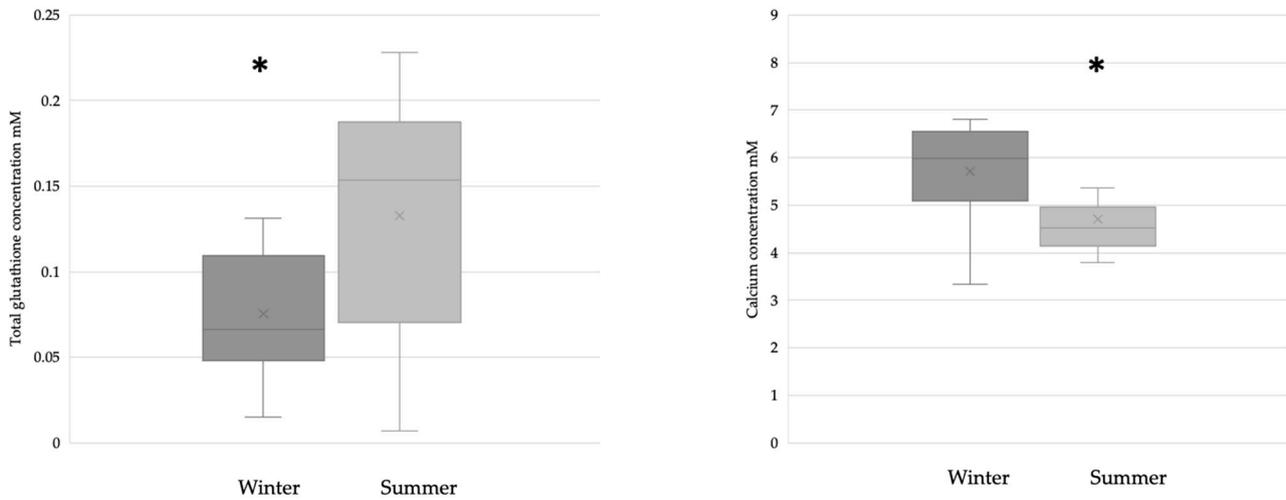
The transferrin concentration was higher in the BM produced for female infants than in the BM produced for male infants. The glutathione, TAC, ORAC, calcium, and protein levels were similar between the two groups. Mothers delivering spontaneously had significantly higher TAC concentrations than mothers giving birth by CS; otherwise, the mode of delivery did not influence the other antioxidants in breast milk. Mothers whose BMI was above 30 after delivery and during breastfeeding had higher BM transferrin content than mothers with a BMI under 30. Maternal BMI had no impact on glutathione, TAC, ORAC, calcium, or protein levels. Maternal age did not influence ORAC, calcium, protein, and transferrin content, but the total glutathione and TAC concentrations were significantly higher in the BM of mothers above the age of 30 (Table 2).

**Table 2.** Antioxidants and total protein in human milk.

		Glutathione mM	TAC μM	ORAC μM	Calcium mM	Total Protein g/L	Transferrin mg/L
Infant gender	Girl	0.08 ± 0.01 (n = 57)	134.06 ± 8.08 (n = 27)	3752.71 ± 122.34 (n = 27)	5.21 ± 0.21 (n = 27)	5.02 ± 0.22 (n = 27)	<b>108.39 ± 16.62</b> (n = 27)
	Boy	0.15 ± 0.05 (n = 65)	114.82 ± 9.83 (n = 32)	3406.66 ± 192.41 (n = 32)	4.78 ± 0.22 (n = 32)	4.74 ± 0.27 (n = 32)	<b>44.22 ± 8.54 *</b> (n = 32)
Delivery	Vaginal delivery	0.13 ± 0.04 (n = 70)	<b>142.07 ± 7.96</b> (n = 25)	3733.45 ± 149.79 (n = 25)	5.05 ± 0.25 (n = 25)	5.07 ± 0.25 (n = 25)	69.1 ± 16.5 (n = 25)
	C-section	0.08 ± 0.01 (n = 52)	<b>109.94 ± 8.75 *</b> (n = 34)	3507.41 ± 153.97 (n = 34)	5.02 ± 0.17 (n = 34)	4.77 ± 0.24 (n = 34)	94.16 ± 15.68 (n = 34)
Maternal BMI	<30	0.12 ± 0.03 (n = 90)	130.21 ± 7.07 (n = 41)	3623.63 ± 132.36 (n = 41)	4.99 ± 0.21 (n = 41)	4.84 ± 0.23 (n = 41)	<b>52.8 ± 7.94</b> (n = 41)
	>30	0.07 ± 0.01 (n = 32)	105.97 ± 13.83 (n = 18)	3605.48 ± 192.65 (n = 18)	5.12 ± 0.24 (n = 18)	5.06 ± 0.24 (n = 18)	<b>139.87 ± 24.30 *</b> (n = 18)
Maternal age	<30	<b>0.06 ± 0.01</b> (n = 51)	<b>99.61 ± 9.36</b> (n = 34)	3690.39 ± 193.75 (n = 34)	5.15 ± 0.28 (n = 34)	4.65 ± 0.25 (n = 34)	81.63 ± 20.04 (n = 34)
	>30	<b>0.11 ± 0.01 *</b> (n = 71)	<b>134.07 ± 7.56 *</b> (n = 25)	3592.23 ± 134.22 (n = 25)	4.99 ± 0.19 (n = 25)	5.01 ± 0.22 (n = 25)	82.76 ± 13.93 (n = 25)

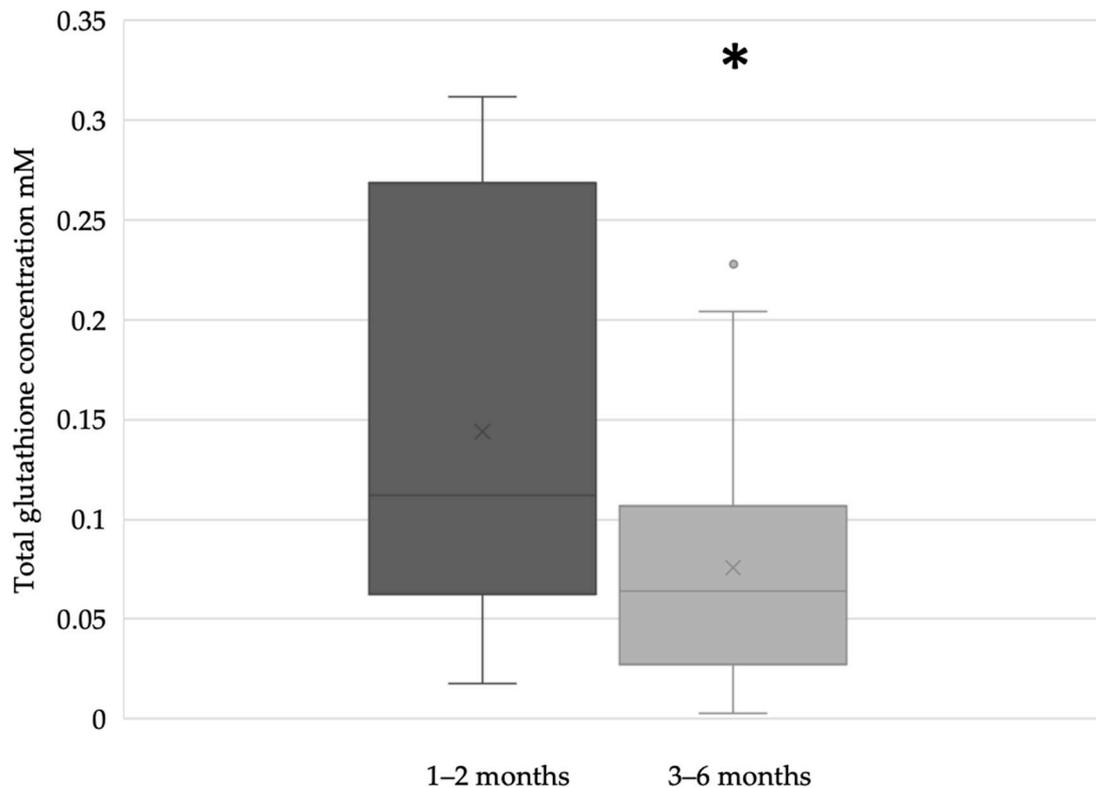
\*  $p < 0.05$ . TAC: total antioxidant capacity; ORAC: oxygen radical absorbance capacity; BMI: body mass index; C-section: Cesarean section.

The breast milk samples collected during the winter ( $n = 28$ ) had significantly lower glutathione levels than the samples collected during the summer ( $n = 31$ ). The calcium concentration was significantly lower in samples collected during the summer (winter  $n = 17$ ; summer  $n = 21$ ) (Figure 2).



**Figure 2.** Glutathione (winter  $n = 28$ ; summer  $n = 31$ ) and calcium (winter  $n = 17$ ; summer  $n = 21$ ) levels in BM samples collected during the winter and summer. In the case of glutathione,  $* p = 0.0167$ ; in the case of calcium,  $* p = 0.0128$ .

In the term breast milk samples, the glutathione concentration was significantly higher during the first two months ( $n = 51$ ) of lactation compared to the period between the third and sixth months ( $n = 71$ ) of breastfeeding (Figure 3).



**Figure 3.** Glutathione concentration of term breast milk during the first 6 months of lactation ( $* p = 0.0213$ ).

#### 4. Discussion

Breastfeeding is a critical aspect of postnatal adaptation; it plays a crucial role in providing optimal nutrition and promoting bonding and emotional attachment between the mother and child. Breast milk is highly nutritious and provides numerous benefits for the infant's growth and development. It contains a balance of essential nutrients, antibodies, enzymes, and other bioactive compounds that support the immune system, digestive health, and overall well-being. Breastfeeding is associated with a lower incidence of a variety of oxidative stress-related illnesses in premature infants [21]. Also, it is known to be a rich source of glutathione, which contributes to the antioxidant protection provided to infants during breastfeeding. Numerous studies have proven the differences between own mother's milk, donor milk, and infant formula [22,23]. All of them reported poorer bioactive factor, hormone, and immunoglobulin content in formula compared to donor milk or own mother's milk. Poorer growth and developmental outcomes have been reported in infants receiving pasteurized donor milk compared to infants receiving unpasteurized human milk [24,25]. The value of antioxidant richness in BM is conceivably important to protect nursing infants against oxidative stress [26,27].

Preterm infants are exposed to a wide range of stressors, e.g., blood tests, infections, phototherapy, oxygen supplementation, parenteral nutrition, and therapeutic interventions during their care. Although ROS contribute to homeostasis, they also participate in cell signaling both in physiological and pathophysiological processes, and ROS can cause molecular and cell damage [2,3]. An investigation found that the phototherapy of jaundiced neonates resulted in increased oxidative stress [28]. As an explanation for the decrease in antioxidant capacity of BM with HoP, thermally induced denaturation is the most likely mechanism. HoP is known to change the composition of BM and decrease the concentration of different hormones and compounds [1,19,20,22]. Other preservation techniques, like refrigeration, also change the composition of BM [20]. A previous study showed that the antioxidant activity of BM decreased significantly from the 21st day of cold storage (at 4 °C and −20 °C) [29]. A recent meta-analysis of the effect of HoP on the antioxidant properties of human milk showed inconclusive results regarding the effect of pasteurization on the TAC of BM [7]. Some studies proved a reduction in TAC after HoP compared with untreated BM, while others detected no influence of Holder pasteurization [8].

Glutathione is a known antioxidant in human milk. It deactivates oxygen-derived free radicals and eliminates toxins, carcinogens, and malonic dialdehyde [30]. Silvestre et al. investigated the effects of HoP, and in contrast to our results, they found that HoP reduced glutathione concentrations in human milk by 46%. They investigated the effect of high-temperature, short-time pasteurization and described no concentration changes in glutathione levels after the procedure [31]. The antioxidant levels of colostrum, mature milk, and transitional milk are different [32]. Our results showed that the glutathione content of BM decreases with time. The ability of the neonatal intestine to mitigate radical accumulation plays a role in its capacity to overcome oxidative stress. Lipid peroxidation is known to preferentially target polyunsaturated fatty acids, and oxidative injury of necrotizing enterocolitis leads to deregulation of the glutathione defense system [11].

BM's non-enzymatic TAC is believed to play a significant role in protecting the infant from oxidative stress and supporting its overall health and development. TAC is a general measure to indicate the level of free radicals scavenged by a test solution that is commonly used to assess the antioxidant status of human milk samples. TAC provides a general assessment of the antioxidant capacity of a given bioactive component, while the quantification of nutrient antioxidants, specific antioxidant enzymes, conjugated dienes, or lipoprotein oxidation provides more specific information [33–35]. With age, BM melatonin concentration was shown to be decreased [36]. No previous data were found about TAC or total glutathione levels in BM related to maternal age. Our present results suggest a correlation between maternal age and TAC and glutathione concentration, which might be a compensatory mechanism of age-related processes. However, it is important to understand

that the precise impact and significance of non-enzymatic TAC in BM on infant health are still areas of ongoing research.

In a previous study, the BM of mothers with gestational diabetes had similar ORAC levels compared to non-diabetic mothers. BM ORAC was positively correlated with BM ascorbic acid in mothers with gestational diabetes [8]. Colostrum showed significantly higher ORAC values compared with mature milk [32].

The calcium concentrations detected in our study were similar to previous results [37,38]. The HoP calcium concentration was found to be elevated; presumably, the heat treatment unbound the calcium in the BM. This phenomenon was observed with Il-7 [39] and TSH [20] in previous studies. No differences in calcium concentration were detected based on gestational age [40]. A review found that conditions like familial hypophosphatemia and hyperparathyroidism affect BM calcium concentrations, but other environmental parameters did not influence calcium concentration [41]. Minerals, like calcium in milk, are particularly important for infant skeletal development and may reflect maternal characteristics [42]. In a previous study on children, compared to winter, children in the spring and summer had significantly lower plasma calcium concentrations [43]. The absorption of calcium is controlled by vitamin D from the small intestine [44]. Theoretically, the serum calcium concentration in the summer is higher than in the winter; we did not find dietary differences among the food consumption of the involved women. Seasonal diversity in T cell activity may also be associated with seasonal changes in blood calcium levels [45]. These results suggest that other factors may influence the calcium content of BM. The total protein level of BM was not affected by HoP in our present study, in agreement with previous reports [23,46,47].

A limited amount of information is available on the presence of transferrin in breast milk [48,49]. The present work shows that HoP vigorously decreases the concentration of transferrin in BM. BM produced for female infants contains higher levels of transferrin than milk produced for boys. An earlier study reported that infant girls had higher hemoglobin and serum ferritin concentrations than boys [50]. The serum ferritin level was found to be elevated in individuals with increased BMI values [51], while in our study, the transferrin level was higher in the BM of mothers with a BMI above 30.

It is important to note that the content of antioxidants and other components in human milk may vary depending on various factors such as the mother's diet, overall health, stage of lactation, and mode of delivery. Additionally, the pasteurization process, such as HoP, is known to affect the levels of certain components in human milk. However, despite any potential changes, human milk remains an excellent source of nutrition and immune protection for infants [52,53]. Earlier findings shed light on how the hormonal components of milk have sex-specific effects on offspring growth during early postnatal life with varying temporal windows of sensitivity [54,55]. The total dietary antioxidant capacity of patients' diets significantly depended on the season and was highest in the summer [56]. It is known that maternal diet influences the composition of BM [51,52,57,58] and may result in epigenetic changes [59]. Our results demonstrate that the antioxidant content of BM is influenced by the seasons and may reflect maternal diet as well.

The antioxidant properties of human milk limit the consequences of excessive oxidative damage. After birth, especially in premature infants, the gastrointestinal tract is under development, which leads to incomplete or slow protein digestion [60], promoting the absorption and bioavailability of BM components. Previous research has demonstrated that the addition of antioxidants to infant formula increases infant resistance to oxidative stress [27,61]. Continuous ROS exposure can induce metabolic changes such as hyperglycemia in extremely-low-birth-weight infants [62].

Hormone levels (e.g., leptin) [63] and supplementation (e.g., thyroxin) [64] show a connection with developmental outcomes in early childhood, suggesting that continuous monitoring of antioxidant levels or supplementation during intensive care should be investigated in clinical trials. Anti- and prooxidants control a sensitive balance in newborns via multiple factors, such as immunoglobulins, short-chain fatty acids, and cytokines (which

can be found in BM), and, through absorption and local effects, influence their reaction to intensive care treatment, survival, and development [65–68]. The impact of oxidative stress and the balance or imbalance of pro- and antioxidants control intrauterine development and postnatal life [69–71]. Obstetrical complications, like the premature rupture of membranes [72] or preeclampsia [73], also intensify OS-induced processes. During postnatal adaptation oxidative stress may have an impact on adult diseases affecting the cardiovascular or endocrine system [74,75]. OS, similar to inflammation, promotes aging-related pathologies, endothelial dysfunction, and adverse pregnancy outcomes [76]. Maternal nutrition influences the nutritional programming of the offspring; in later life, cardiovascular diseases, metabolic syndrome, diabetes, insulin resistance, glucose intolerance, fertility issues [77,78], and hypertension may develop in adulthood [79]. Dysfunction of hypothalamic appetite control results in obesity through increased lipogenesis [80]. Preterm infants with a higher total antioxidant status are more likely to be protected from free radicals, blocking ROS accumulation and OS [81]. BM is a known source of antioxidant capacity, providing and supporting breastfed preterm neonates [82]. Antioxidant treatment has become a potential and predictably essential therapeutic strategy in the treatment of preterm newborns with bronchopulmonary dysplasia [83,84] or necrotizing enterocolitis [85,86]. The prevention of chronic morbidities of extremely premature newborns by different therapeutic options and adjuvant perinatal strategies are already highlighted in clinical practice [87–89]; therefore, supplementing antioxidants during intensive care should be investigated.

Our study has limitations. We only examined the chosen antioxidants in BM samples before and after HoP but revealed higher transferrin levels in the BM produced for female infants and in the BM of mothers with BMIs of 30 or above. Our results suggest that some antioxidants are present in higher concentrations in BM in the summer than in the winter. Elevated maternal age was associated with higher glutathione and TAC levels in BM, and after vaginal delivery, glutathione was present in higher concentrations in BM than after Cesarean section. Our results have augmented our knowledge about the effects of HoP: glutathione concentration was not impacted by HoP, but TAC, ORAC, and transferrin levels were reduced.

## 5. Conclusions

Breast milk is considered the most optimal feeding option for an infant, with it having greater benefits in the case of preterm birth [90–93]. Breastfeeding influences the rate of OS bias postnatal adaptation, through impacting insulin sensitivity, choline and prostaglandin metabolism, and lipid profile during early infancy [94–96]. Although HoP modifies the antioxidant content of BM, donor milk is still considered the most suitable alternative to a mother's own BM. HoP reduced the TAC, ORAC, and transferrin concentration in BM. The clinical significance of the changes in BM antioxidants with HoP is unknown; further research is necessary to improve our knowledge. Transferrin levels are higher in BM produced for female infants, the TAC concentration was found to be elevated in BM after vaginal delivery, and an elevated maternal BMI resulted in higher transferrin levels. Our present results demonstrate that the antioxidant intake of the infant is influenced by the gender of the infant and maternal and environmental factors. Knowing that these antioxidant compounds actively influence physiological processes, antioxidant supplementation and guidance of human milk banks in testing different pasteurization processes in order to maximize the preservation of antioxidant properties are highly recommended.

**Author Contributions:** Conceptualization, R.A.V., É.M. and T.E.; methodology, R.A.V., É.M. and T.K.; investigation, R.A.V., É.M., C.G., T.K. and C.I.V.; writing—original draft preparation, R.A.V.; writing—review and editing, R.A.V., É.M., C.G., T.K., C.I.V., J.B., K.K., S.F., S.B., D.M. and T.E.; project administration, R.A.V., S.F., C.I.V., S.B. and D.M.; funding acquisition, R.A.V., É.M., J.B., K.K. and T.E. All authors have read and agreed to the published version of the manuscript.

**Funding:** Project no. RRF-2.3.1-21-2022-00012, entitled the National Laboratory on Human Reproduction, has been implemented with support provided by the Recovery and Resilience Facility of the European Union within the framework of the Programme Széchenyi Plan Plus. This work was supported by the Medical School of the University of Pécs.

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Regional and Local Research Ethics Committee of the University of Pécs, Pécs, Hungary (PTE KK 7072-2018).

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study.

**Data Availability Statement:** Data applied in this study are available from the corresponding author upon request.

**Acknowledgments:** We are grateful to the recruited mothers and infants. We thank Katalin Szalaváry and the PMJV Health Institution for their help. We acknowledge Edward F. Bell for his advice and help in editing the manuscript. Project no. RRF-2.3.1-21-2022-00012, entitled the National Laboratory on Human Reproduction, has been implemented with support provided by the Recovery and Resilience Facility of the European Union within the framework of the Programme Széchenyi Plan Plus. The project has received funding from the HUN-REN Hungarian Research Network.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Vass, R.A.; Kiss, G.; Bell, E.F.; Roghair, R.D.; Miseta, A.; Bódis, J.; Funke, S.; Ertl, T. Breast milk for term and preterm infants—own mother’s milk or donor milk? *Nutrients* **2021**, *13*, 424. [[CrossRef](#)]
2. Moore, T.A.; Ahmad, I.M.; Zimmerman, M.C. Oxidative stress and preterm birth: An integrative review. *Biol. Res. Nurs.* **2018**, *20*, 497–512. [[CrossRef](#)] [[PubMed](#)]
3. Mauchart, P.; Vass, R.A.; Nagy, B.; Sulyok, E.; Bódis, J.; Kovács, K. Oxidative stress in assisted reproductive techniques, with a focus on an underestimated risk factor. *Curr. Issues Mol. Biol.* **2023**, *45*, 1272–1286. [[CrossRef](#)] [[PubMed](#)]
4. Hwang, J.H.; Lee, E.H.; Kim, E.A. Retinopathy of Prematurity among very-low-birth-weight infants in Korea: Incidence, treatment, and risk factors. *J. Korean Med. Sci.* **2015**, *30*, 88–94. [[CrossRef](#)] [[PubMed](#)]
5. Perrone, S.; Tataranno, M.L.; Buonocore, G. Oxidative stress and bronchopulmonary dysplasia. *J. Clin. Neonatol.* **2012**, *1*, 109–114. [[PubMed](#)]
6. Silvers, K.M.; Gibson, A.T.; Russell, J.M.; Powers, H.J. Antioxidant activity, packed cell transfusions, and outcome in premature infants. *Arch. Dis. Child. Fetal Neonatal Ed.* **1998**, *78*, 214–219. [[CrossRef](#)] [[PubMed](#)]
7. Juncker, H.G.; Ruhé, E.J.M.; Burchell, G.L.; van den Akker, C.H.P.; Korosi, A.; van Goudoever, J.B.; van Keulen, B.J. The effect of pasteurization on the antioxidant properties of human milk: A literature review. *Antioxidants* **2021**, *10*, 1737. [[CrossRef](#)]
8. Churchill, M.; Zawawi, H.; Elisia, I.; Seider, M.; Noseworthy, R.; Thompson, A.; Glenn, A.J.; Ramdath, D.D.; O’Connor, D.; Darling, P.; et al. The antioxidant capacity of breast milk and plasma of women with or without gestational diabetes mellitus. *Antioxidants* **2023**, *12*, 842. [[CrossRef](#)]
9. Cloetens, L.; Panee, J.; Åkesson, B. The antioxidant capacity of milk—the application of different methods in vitro and in vivo. *Cell Mol. Biol.* **2013**, *59*, 43–57.
10. Kőszegi, T.; Sali, N.; Raknić, M.; Horváth-Szalai, Z.; Csepregi, R.; Končić, M.Z.; Papp, N.; Poór, M. A novel luminol-based enhanced chemiluminescence antioxidant capacity microplate assay for use in different biological matrices. *J. Pharmacol. Toxicol. Methods* **2017**, *88*, 153–159. [[CrossRef](#)]
11. Golubkova, A.; Leiva, T.; Snyder, K.; Schlegel, C.; Bonvicino, S.M.; Agbaga, M.P.; Brush, R.S.; Hansen, J.M.; Vitiello, P.F.; Hunter, C.J. Response of the glutathione (GSH) antioxidant defense system to oxidative injury in necrotizing enterocolitis. *Antioxidants* **2023**, *12*, 1385. [[CrossRef](#)] [[PubMed](#)]
12. Di Giacomo, C.; Malfa, G.A.; Tomasello, B.; Bianchi, S.; Acquaviva, R. Natural compounds and glutathione: Beyond mere antioxidants. *Antioxidants* **2023**, *12*, 1445. [[CrossRef](#)] [[PubMed](#)]
13. Gomme, P.T.; McCann, K.B.; Bertolini, J. Transferrin: Structure, function and potential therapeutic actions. *Drug Discov. Today* **2005**, *10*, 267–273. [[CrossRef](#)] [[PubMed](#)]
14. Blanc, B.; Isliker, H. Isolation and characterization of the red siderophilic protein from maternal milk: Lactotransferrin. *Bull. Soc. Chim. Biol.* **1961**, *43*, 929–943.
15. Hao, L.; Shan, Q.; Wei, J.; Ma, F.; Sun, P. Lactoferrin: Major physiological functions and applications. *Curr. Protein Pept. Sci.* **2019**, *20*, 139–144. [[CrossRef](#)]
16. EFSA NDA. Scientific Opinion on the essential composition of infant and follow-on formulae. *EFSA J.* **2014**, *12*, 3760. [[CrossRef](#)]
17. Rios-Leyvraz, M.; Yao, Q. Calcium, zinc, and vitamin D in breast milk: A systematic review and meta-analysis. *Int. Breastfeed. J.* **2023**, *18*, 27. [[CrossRef](#)]

18. Vass, R.A.; Kemeny, A.; Dergez, T.; Ertl, T.; Reglodi, D.; Jungling, A.; Tamas, A. Distribution of bioactive factors in human milk samples. *Int. Breastfeed. J.* **2019**, *14*, 9. [[CrossRef](#)]
19. Vass, R.A.; Bell, E.F.; Colaizy, T.T.; Schmelzel, M.L.; Johnson, K.J.; Walker, J.R.; Ertl, T.; Roghair, R.D. Hormone levels in preterm and donor human milk before and after Holder pasteurization. *Pediatr. Res.* **2020**, *88*, 612–617. [[CrossRef](#)]
20. Vass, R.A.; Roghair, R.D.; Bell, E.F.; Colaizy, T.T.; Schmelzel, M.L.; Johnson, K.J.; Walker, J.R.; Ertl, T. Pituitary glycoprotein hormones in human milk before and after pasteurization or refrigeration. *Nutrients* **2020**, *12*, 687. [[CrossRef](#)]
21. Shoji, H.; Shimizu, T. Effect of human breast milk on biological metabolism in infants. *Pediatr. Int.* **2019**, *61*, 6–15. [[CrossRef](#)] [[PubMed](#)]
22. Vass, R.A.; Kiss, G.; Bell, E.F.; Miseta, A.; Bódis, J.; Funke, S.; Bokor, S.; Molnár, D.; Kósa, B.; Kiss, A.A.; et al. Thyroxine and thyroid-stimulating hormone in own mother's milk, donor milk, and infant formula. *Life* **2022**, *12*, 584. [[CrossRef](#)] [[PubMed](#)]
23. Vass, R.A.; Bell, E.F.; Roghair, R.D.; Kiss, G.; Funke, S.; Bokor, S.; Molnár, D.; Miseta, A.; Bódis, J.; Kovács, K.; et al. Insulin, testosterone, and albumin in term and preterm breast milk, donor milk, and infant formula. *Nutrients* **2023**, *15*, 1476. [[CrossRef](#)]
24. Montjaux-Regis, N.; Cristini, C.; Arnaud, C.; Glorieux, I.; Vanpee, M.; Casper, C. Improved growth of preterm infants receiving mother's own raw milk compared with pasteurized donor milk. *Acta Paediatr.* **2011**, *100*, 1548–1554. [[CrossRef](#)] [[PubMed](#)]
25. Hard, A.L.; Nilsson, A.K.; Lund, A.M.; Hansen-Pupp, I.; Smith, L.E.H.; Hellstrom, A. Review shows that donor milk does not promote the growth and development of preterm infants as well as maternal milk. *Acta Paediatr.* **2019**, *108*, 998–1007. [[CrossRef](#)]
26. Tsopmo, A.; Friel, J.K. Human milk has anti-oxidant properties to protect premature infants. *Curr. Pediatr. Rev.* **2007**, *3*, 45–51. [[CrossRef](#)]
27. Friel, J.K.; Martin, S.M.; Langdon, M.; Herzberg, G.R.; Buettner, G.R. Milk from mothers of both premature and full-term infants provides better antioxidant protection than does infant formula. *Pediatr. Res.* **2002**, *51*, 612–618. [[CrossRef](#)]
28. Aycicek, A.; Erel, O. Total oxidant/antioxidant status in jaundiced newborns before and after phototherapy. *J. Pediatr.* **2007**, *83*, 319–322. [[CrossRef](#)]
29. Ribeiro, V.P.D.; Tinoco, R.B.; Chamon, A.L.B.; Pessoa, I.S.; Santos, T.C.D.; Silva, R.S.; Fronza, M. The influence of time and temperature on human milk storage antioxidant properties, oxidative stress, and total protein. *J. Hum. Lact.* **2023**, *39*, 308–314. [[CrossRef](#)]
30. Ankrah, N.A.; Appiah-Opong, R.; Dzokoto, C. Human breastmilk storage and the glutathione content. *J. Trop. Pediatr.* **2000**, *46*, 111–113. [[CrossRef](#)]
31. Silvestre, D.; Miranda, M.; Muriach, M.; Almansa, I.; Jareño, E.; Romero, F.J. Antioxidant capacity of human milk: Effect of thermal conditions for the pasteurization. *Acta Paediatr.* **2008**, *97*, 1070–1074. [[CrossRef](#)]
32. Zarban, A.; Taheri, F.; Chahkandi, T.; Sharifzadeh, G.; Khorashadizadeh, M. Antioxidant and radical scavenging activity of human colostrum, transitional and mature milk. *J. Clin. Biochem. Nutr.* **2009**, *45*, 150–154. [[CrossRef](#)]
33. Kusano, C.; Ferrari, B. Total antioxidant capacity: A biomarker in biomedical and nutritional studies. *J. Cell. Mol. Biol.* **2008**, *7*, 1–15.
34. Elisia, I.; Kitts, D.D. Quantification of hexanal as an index of lipid oxidation in human milk and association with antioxidant components. *J. Clin. Biochem. Nutr.* **2011**, *49*, 147–152. [[CrossRef](#)]
35. Kitts, D.D.; Hu, C. Biological and chemical assessment of antioxidant activity of sugar-lysine model Maillard reaction products. *Ann. N.Y. Acad. Sci.* **2005**, *1043*, 501–512. [[CrossRef](#)] [[PubMed](#)]
36. Gila-Díaz, A.; Herranz Carrillo, G.; Cañas, S.; Saenz de Pipaón, M.; Martínez-Orgado, J.A.; Rodríguez-Rodríguez, P.; López de Pablo, Á.L.; Martín-Cabrejas, M.A.; Ramiro-Cortijo, D.; Arribas, S.M. Influence of maternal age and gestational age on breast milk antioxidants during the first month of lactation. *Nutrients* **2020**, *12*, 2569. [[CrossRef](#)] [[PubMed](#)]
37. Wu, X.; Jackson, R.T.; Khan, S.A.; Ahuja, J.; Pehrsson, P.R. Human milk nutrient composition in the United States: Current knowledge, challenges, and research needs. *Curr. Dev. Nutr.* **2018**, *2*, nzy025. [[CrossRef](#)]
38. Yang, T.; Zhang, L.; Bao, W.; Rong, S. Nutritional composition of breast milk in Chinese women: A systematic review. *Asia Pac. J. Clin. Nutr.* **2018**, *27*, 491–502.
39. Peila, C.; Moro, G.E.; Bertino, E.; Cavallarin, L.; Giribaldi, M.; Giuliani, F.; Cresi, F.; Coscia, A. The effect of Holder Pasteurization on nutrients and biologically-active components in donor human milk: A review. *Nutrients* **2016**, *8*, 477. [[CrossRef](#)]
40. Gidrewicz, D.A.; Fenton, T.R. A systematic review and meta-analysis of the nutrient content of preterm and term breast milk. *BMC Pediatr.* **2014**, *14*, 216. [[CrossRef](#)]
41. Dorea, J.G. Calcium and phosphorus in human milk. *Nutr. Res.* **1999**, *19*, 709–739. [[CrossRef](#)]
42. Hinde, K.; Foster, A.B.; Landis, L.M.; Rendina, D.; Oftedal, O.T.; Power, M.L. Daughter dearest: Sex-biased calcium in mother's milk among rhesus macaques. *Am. J. Phys. Anthropol.* **2013**, *151*, 144–150. [[CrossRef](#)]
43. Zhang, M.; Zhai, R.; Liu, J.; Guang, H.; Li, B.; Zhang, S. Seasonal variation of blood calcium levels in children aged 1–10. *J. Clin. Lab. Anal.* **2016**, *30*, 741–744. [[CrossRef](#)]
44. Peacock, M. Calcium metabolism in health and disease. *Clin. J. Am. Soc. Nephrol.* **2010**, *1*, S23–S30. [[CrossRef](#)] [[PubMed](#)]
45. Khoo, A.L.; Koenen, H.J.; Chai, L.Y.; Sweep, F.C.; Netea, M.G.; van der Ven, A.J.; Joosten, I. Seasonal variation in vitamin D(3) levels is paralleled by changes in the peripheral blood human T cell compartment. *PLoS ONE* **2012**, *7*, 29250. [[CrossRef](#)] [[PubMed](#)]
46. Binder, C.; Baumgartner-Parzer, S.; Gard, L.-I.; Berger, A.; Thajer, A. Human Milk Processing and Its Effect on Protein and Leptin Concentrations. *Nutrients* **2023**, *15*, 347. [[CrossRef](#)]

47. Ley, S.H.; Hanley, A.J.; Stone, D.; O'Connor, D.L. Effects of pasteurization on adiponectin and insulin concentrations in donor human milk. *Pediatr. Res.* **2011**, *70*, 278–281. [[CrossRef](#)] [[PubMed](#)]
48. Saarinen, U.M.; Siimes, M.A.; Dallman, P.R. Iron absorption in infants: High bioavailability of breast milk iron as indicated by the extrinsic tag method of iron absorption and by the concentration of serum ferritin. *J. Pediatr.* **1977**, *91*, 36–39. [[CrossRef](#)]
49. Bullen, J.J.; Rogers, H.J.; Leigh, L. Iron-binding proteins in milk and resistance to *Escherichia coli* infection in infants. *Br. Med. J.* **1972**, *1*, 69–75. [[CrossRef](#)]
50. Wieringa, F.T.; Berger, J.; Dijkhuizen, M.A.; Hidayat, A.; Ninh, N.X.; Utomo, B.; Wasantwisut, E.; Winichagoon, P. Sex differences in prevalence of anaemia and iron deficiency in infancy in a large multi-country trial in South-East Asia. *Br. J. Nutr.* **2007**, *98*, 1070–1076. [[CrossRef](#)]
51. Alam, F.; Memon, A.S.; Fatima, S.S. Increased body mass index may lead to hyperferritinemia irrespective of body iron stores. *Pak. J. Med. Sci.* **2015**, *31*, 1521–1526. [[PubMed](#)]
52. Keikha, M.; Shayan-Moghadam, R.; Bahreynian, M.; Kelishadi, R. Nutritional supplements and mother's milk composition: A systematic review of interventional studies. *Int. Breastfeed. J.* **2021**, *16*, 1. [[CrossRef](#)] [[PubMed](#)]
53. Ramiro-Cortijo, D.; Herranz Carrillo, G.; Singh, P.; Rebollo-Hernanz, M.; Rodríguez-Rodríguez, P.; Ruvira, S.; Martín-Trueba, M.; Martín, C.R.; Arribas, S.M. Maternal and neonatal factors modulating breast milk cytokines in the first month of lactation. *Antioxidants* **2023**, *12*, 996. [[CrossRef](#)]
54. Bermejo-Haro, M.Y.; Camacho-Pacheco, R.T.; Brito-Pérez, Y.; Mancilla-Herrera, I. The hormonal physiology of immune components in breast milk and their impact on the infant immune response. *Mol. Cell Endocrinol.* **2023**, *572*, 111956. [[CrossRef](#)] [[PubMed](#)]
55. Petrullo, L.; Hinde, K.; Lu, A. Steroid hormone concentrations in milk predict sex-specific offspring growth in a nonhuman primate. *Am. J. Hum. Biol.* **2019**, *31*, e23315. [[CrossRef](#)]
56. Czlapka-Matyasik, M.; Gramza-Michalowska, A. The total dietary antioxidant capacity, its seasonal variability, and dietary sources in cardiovascular patients. *Antioxidants* **2023**, *12*, 292. [[CrossRef](#)]
57. Simon Sarkadi, L.; Zhang, M.; Muránszky, G.; Vass, R.A.; Matsyura, O.; Benes, E.; Vari, S.G. Fatty acid composition of milk from mothers with normal weight, obesity, or gestational diabetes. *Life* **2022**, *12*, 1093. [[CrossRef](#)]
58. Ramiro-Cortijo, D.; Singh, P.; Liu, Y.; Medina-Morales, E.; Yakah, W.; Freedman, S.D.; Martín, C.R. Breast milk lipids and fatty acids in regulating neonatal intestinal development and protecting against intestinal injury. *Nutrients* **2020**, *12*, 534. [[CrossRef](#)]
59. Bokor, S.; Vass, R.A.; Funke, S.; Ertl, T.; Molnár, D. Epigenetic effect of maternal methyl-group donor intake on offspring's health and disease. *Life* **2022**, *12*, 609. [[CrossRef](#)]
60. Dallas, D.C.; Underwood, M.A.; Zivkovic, A.M.; German, J.B. Digestion of protein in premature and term Infants. *J. Nutr. Disord. Ther.* **2012**, *2*, 112. [[CrossRef](#)]
61. Pozzo, L.; Cirrincione, S.; Russo, R.; Karamać, M.; Amarowicz, R.; Coscia, A.; Antoniazzi, S.; Cavallarin, L.; Giribaldi, M. Comparison of oxidative status of human milk, human milk fortifiers and preterm infant formulas. *Foods* **2019**, *8*, 458. [[CrossRef](#)]
62. Turai, R.; Schandl, M.F.; Dergez, T.; Vass, R.A.; Kvárik, T.; Horányi, E.; Balika, D.; Mammel, B.; Gyarmati, J.; Fónai, F.; et al. Early and late complications of hyperglycemic extremely low birth-weight infants. *Orv. Hetil.* **2019**, *160*, 1270–1278. [[CrossRef](#)]
63. Roghair, R.D.; Colaizy, T.T.; Steinbreker, B.; Vass, R.A.; Hsu, E.; Dagle, D.; Chatmethakul, T. Neonatal leptin levels predict the early childhood developmental assessment scores of preterm infants. *Nutrients* **2023**, *15*, 1967. [[CrossRef](#)] [[PubMed](#)]
64. Ng, S.M.; Turner, M.A.; Weindling, A.M. Neurodevelopmental outcomes at 42 months after thyroxine supplementation in infants below 28 weeks' gestation: A randomized controlled trial. *Thyroid* **2020**, *30*, 948–954. [[CrossRef](#)]
65. Sulyok, E.; Farkas, B.; Bodis, J. Pathomechanisms of prenatally programmed adult diseases. *Antioxidants* **2023**, *12*, 1354. [[CrossRef](#)]
66. Barker, D.J.P. In utero programming of chronic disease. *Clin. Sci.* **1998**, *95*, 115–128. [[CrossRef](#)]
67. Barker, D.J.; Osmond, C.; Forsén, T.J.; Kajantie, E.; Eriksson, J.G. Trajectories of growth among children who have coronary events as adults. *N. Engl. J. Med.* **2005**, *353*, 1802–1809. [[CrossRef](#)]
68. Kuzawa, C.W. Fetal origins of developmental plasticity: Are fetal cues reliable predictors of future nutritional environment? *Am. J. Hum. Biol.* **2005**, *17*, 5–21. [[CrossRef](#)] [[PubMed](#)]
69. Hussain, T.; Murtaza, G.; Metwally, E.; Kalhor, D.H.; Kalhor, M.S.; Rahu, B.A.; Sahito, R.G.A.; Yin, Y.; Yang, H.; Chughtai, M.I.; et al. The role of oxidative stress and antioxidant balance in pregnancy. *Mediat. Inflamm.* **2021**, *2021*, 9962860. [[CrossRef](#)]
70. Mistry, H.D.; Williams, P.J. The importance of antioxidant micronutrients in pregnancy. *Oxid. Med. Cell. Longev.* **2011**, *2011*, 841749. [[CrossRef](#)] [[PubMed](#)]
71. Yang, X.; Hu, R.; Shi, M.; Wang, L.; Yan, J.; Gong, J.; Zhang, Q.; He, J.; Wu, S. Placental malfunction, fetal survival and development caused by sow metabolic disorder: The impact of maternal oxidative stress. *Antioxidants* **2023**, *12*, 360. [[CrossRef](#)] [[PubMed](#)]
72. Musilova, I.; Tothova, L.; Menon, R.; Vlkova, B.; Celec, P.; Hornychova, H.; Kutova, R.; Andrys, C.; Stepan, M.; Kacerovsky, M. Umbilical cord blood markers of oxidative stress in pregnancies complicated by preterm prelabor rupture of membranes. *J. Matern. Fetal Neonatal Med.* **2016**, *29*, 1900–1910. [[CrossRef](#)] [[PubMed](#)]
73. Han, C.; Huang, P.; Lyu, M.; Dong, J. Oxidative stress and preeclampsia-associated prothrombotic state. *Antioxidants* **2020**, *9*, 1139. [[CrossRef](#)]
74. Marseglia, L.; D'Angelo, G.; Manti, S.; Arrigo, T.; Barberi, I.; Reiter, R.J.; Gitto, E. Oxidative stress-mediated aging during the fetal and perinatal periods. *Oxid. Med. Cell Longev.* **2014**, *2014*, 358375. [[CrossRef](#)]

75. DeFreitas, M.J.; Katsoufis, C.P.; Benny, M.; Young, K.; Kulandavelu, S.; Ahn, H.; Sfakianaki, A.; Abitbol, C.L. Educational Review: The Impact of Perinatal Oxidative Stress on the Developing Kidney. *Front. Pediatr.* **2022**, *10*, 853722. [[CrossRef](#)]
76. Sultana, Z.; Maiti, K.; Aitken, J.; Morris, J.; Dedman, L.; Smith, R. Oxidative stress placental ageing-related pathogenesis and adverse pregnancy outcomes. *Am. J. Reprod. Immunol.* **2017**, *77*, 12653. [[CrossRef](#)]
77. Gluckman, P.D. Editorial. nutrition, glucocorticoid, birth size and adult disease. *J. Clin. Endocrinol. Metab.* **2001**, *142*, 1689–1691. [[CrossRef](#)] [[PubMed](#)]
78. Kanaka-Gantenbein, C. Fetal origins of adult diabetes. *Ann. N.Y. Acad. Sci.* **2010**, *1205*, 99–105. [[CrossRef](#)]
79. Symonds, M.E.; Stephenson, T.; Gardner, D.S.; Budge, H. Long-term effects of nutritional programming of the embryo and fetus. mechanisms and critical windows. *Reprod. Fertil. Dev.* **2007**, *19*, 53–63. [[CrossRef](#)]
80. Ross, M.G.; Desai, M. Developmental programming of offspring obesity, adipogenesis and appetite. *Clin. Obstet. Gynecol.* **2013**, *56*, 529–536. [[CrossRef](#)]
81. Chrustek, A.; Dombrowska-Pali, A.; Olszewska-Slonina, D. Analysis of the composition and antioxidant status of breast milk in women giving birth prematurely and on time. *PLoS ONE* **2021**, *16*, 0255252. [[CrossRef](#)]
82. Yang, X.; Jiang, S.; Deng, X.; Luo, Z.; Chen, A.; Yu, R. Effects of antioxidants in human milk on bronchopulmonary dysplasia prevention and treatment: A review. *Front Nutr* **2022**, *9*, 924036. [[CrossRef](#)] [[PubMed](#)]
83. Rudloff, I.; Cho, S.X.; Bui, C.B.; McLean, C.; Veldman, A.; Berger, P.J.; Nold, M.F.; Nold-Petry, C.A. Refining anti-inflammatory therapy strategies for bronchopulmonary dysplasia. *J. Cell Mol. Med.* **2017**, *21*, 1128–1138. [[CrossRef](#)] [[PubMed](#)]
84. Gao, R.; Li, Z.; Ai, D.; Ma, J.; Chen, C.; Liu, X. Interleukin-24 as a pulmonary target cytokine in bronchopulmonary dysplasia. *Cell Biochem. Biophys.* **2021**, *79*, 311–320. [[CrossRef](#)] [[PubMed](#)]
85. Altobelli, E.; Angeletti, P.M.; Verrotti, A.; Petrocelli, R. The impact of human milk on necrotizing enterocolitis: A systematic review and meta-analysis. *Nutrients* **2020**, *12*, 1322. [[CrossRef](#)]
86. Sami, A.S.; Frazer, L.C.; Miller, C.M.; Singh, D.K.; Clodfelter, L.G.; Orgel, K.A.; Good, M. The role of human milk nutrients in preventing necrotizing enterocolitis. *Front. Pediatr.* **2023**, *11*, 1188050. [[CrossRef](#)]
87. Tang, W.; Gao, T.; Cao, Y.; Zhou, W.; Song, D.; Wang, L. Narrative review of perinatal management of extremely preterm infants: What's the evidence? *Pediatr. Med.* **2022**, *5*, 37. [[CrossRef](#)]
88. Siffel, C.; Hirst, A.K.; Sarda, S.P.; Kuzniecicz, M.W.; Li, D.K. The clinical burden of extremely preterm birth in a large medical records database in the United States: Mortality and survival associated with selected complications. *Early Hum. Dev.* **2022**, *171*, 105613. [[CrossRef](#)]
89. Balázs, G.; Balajthy, A.; Seri, I.; Hegyi, T.; Ertl, T.; Szabó, T.; Röszer, T.; Papp, Á.; Balla, J.; Gáll, T.; et al. Prevention of chronic morbidities in extremely premature newborns with LISA-nCPAP respiratory therapy and adjuvant perinatal strategies. *Antioxidants* **2023**, *12*, 1149. [[CrossRef](#)]
90. Vohr, B.R.; Poindexter, B.B.; Dusick, A.M.; McKinley, L.T.; Wright, L.L.; Langer, J.C.; Poole, W.K.; NICHD Neonatal Research Network. Beneficial effects of breast milk in the neonatal intensive care unit on the developmental outcome of extremely low birth weight infants at 18 months of age. *Pediatrics* **2006**, *118*, 115–123. [[CrossRef](#)]
91. Vohr, B.R.; Poindexter, B.B.; Dusick, A.M.; McKinley, L.T.; Higgins, R.D.; Langer, J.C.; Poole, W.K.; National Institute of Child Health and Human Development National Research Network. Persistent beneficial effects of breast milk ingested in the neonatal intensive care unit on outcomes of extremely low birth weight infants at 30 months of age. *Pediatrics* **2007**, *120*, e953–e959. [[CrossRef](#)]
92. Arslanoglu, S.; Boquien, C.Y.; King, C.; Lamireau, D.; Tonetto, P.; Barnett, D.; Bertino, E.; Gaya, A.; Gebauer, C.; Grovslie, A.; et al. Fortification of human milk for preterm infants: Update and recommendations of the European Milk Bank Association (EMBA) working group on human milk fortification. *Front. Pediatr.* **2019**, *7*, 76. [[CrossRef](#)]
93. Sudeep, K.C.; Kumar, J.; Ray, S.; Dutta, S.; Aggarwal, R.; Kumar, P. Oral Application of colostrum and mother's own milk in preterm infants—a randomized, controlled trial. *Indian J. Pediatr.* **2022**, *89*, 579–586. [[CrossRef](#)] [[PubMed](#)]
94. Lembo, C.; Buonocore, G.; Perrone, S. Oxidative stress in preterm newborns. *Antioxidants* **2021**, *10*, 1672. [[CrossRef](#)] [[PubMed](#)]
95. Agarwal, A.; Aponte-Mellado, A.; Premkumar, B.J.; Shaman, A.; Gupta, S. The effects of oxidative stress on female reproduction: A review. *Reprod. Biol. Endocrinol.* **2012**, *10*, 49. [[CrossRef](#)] [[PubMed](#)]
96. Saker, M.; Soulimane Mokhtari, N.; Merzouk, S.A.; Merzouk, H.; Belarbi, B.; Narce, M. Oxidant and antioxidant status in mothers and their newborns according to birthweight. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2008**, *141*, 95–99. [[CrossRef](#)]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.