

Review



## Intersection between Redox Homeostasis and Autophagy: Valuable Insights into Neurodegeneration

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Abstract: Autophagy, a main degradation pathway for maintaining cellular homeostasis, and redox homeostasis have recently been considered to play protective roles in neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis. Increased levels of reactive oxygen species (ROS) in neurons can induce mitochondrial damage and protein aggregation, thereby resulting in neurodegeneration. Oxidative stress is one of the major activation signals for the induction of autophagy. Upon activation, autophagy can remove ROS, damaged mitochondria, and aggregated proteins from the cells. Thus, autophagy can be an effective strategy to maintain redox homeostasis in the brain. However, the interaction between redox homeostasis and autophagy is not clearly elucidated. In this review, we discuss recent studies on the relationship between redox homeostasis and autophagy associated with neurodegenerative diseases and propose that autophagy induction through pharmacological intervention or genetic activation might be a promising strategy to treat these disorders.

Keywords: autophagy; redox homeostasis; ROS; neurodegenerative diseases

## 1. Introduction

Neurodegenerative diseases are a heterogeneous group of disorders characterized by progressive loss of the structure or function of neurons. The major risk factor for the development of neurodegenerative diseases is aging; therefore, the prevalence of these disorders increases as the average lifespan increases. However, effective treatments for most neurodegenerative diseases are still lacking. Over the past decades, various studies demonstrated that the progression of neurodegenerative diseases is accompanied by decreased antioxidant levels, increased oxidative stress, and decreased autophagy. Oxidative stress can induce autophagy, which relieves the cytotoxicity caused by accumulated oxidative stress. Here, we describe the dysregulation of redox homeostasis that is observed in major neurodegenerative diseases, including Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis. We then discuss the molecular pathways related to oxidative stress and autophagy in neurodegenerative diseases. Finally, we suggest that manipulation of autophagy might be an effective strategy to alleviate oxidative stress in patients suffering from neurodegenerative diseases.

## 2. Main Players Involved in Maintaining Redox Homeostasis

Cells continually generate small, highly reactive molecules called reactive oxygen species (ROS) due to incomplete reduction of oxygen during aerobic metabolism or immune response. ROS include superoxide anion  $(O_2^{\bullet-})$ , hydrogen peroxide  $(H_2O_2)$ , hydroxyl radical ( $^{\bullet}OH$ ), and 4-hydroxynonenal (HNE). ROS can be generated from endogenous sources. The mitochondrial electron transport chain is recognized as one of the major modes of cellular ROS generation [1]. NADPH oxidases, which share the capacity to transport electrons across the plasma membrane, are also able to generate superoxide and other



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**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). downstream ROS [2]. Cytochrome P450 generates ROS and lipid peroxidation products which might interact with DNA to create oxidative DNA adducts [3]. ROS generally cause detrimental effects, such as DNA damage, lipid peroxidation, and protein oxidation, in cells, ultimately leading to cellular stress or pathological conditions. Therefore, under normal physiological conditions, the formation and elimination of ROS are tightly regulated by endogenous antioxidants and ROS scavengers, to maintain homeostasis and avoid oxidative stress and toxicity. The antioxidants that are produced endogenously, include superoxide dismutase (SOD), catalase (CAT), glutathione (GSH), and peroxiredoxins (Prxs). When electrophiles, such as ROS, are increased in turn, antioxidant responses, such as NF- $\kappa$ B or Keap1-Nrf2-mediated signaling cascade, are upregulated to restore the original steady-state [4]. These redox reactions are important for cellular homeostasis to regulate cell signaling, proliferation, and survival [5].

Superoxide anion  $(O_2^{\bullet-})$  is produced by one-electron reduction of dioxygen, which occurs in the mitochondrial electron chain reaction or NADPH oxidase-mediated immune response in phagocytic cells [6]. Owing to its high reactivity, the generation of  $O_2^{\bullet-}$ , even at low levels, is essential for the oxygen-dependent killing of invading pathogens in phagocytic cells [7]. However, there is a proper defense mechanism to remove  $O_2^{\bullet-}$ , as it is harmful to cells when present in excess. SOD is an antioxidant enzyme that catalyzes the dismutation of  $O_2^{\bullet-}$  into molecular oxygen and hydrogen peroxide, which is the first defense mechanism against ROS. Almost all organisms that use  $O_2^{\bullet-}$  for mitochondrial respiration express SOD1 in the mitochondrial intermembrane space and cytoplasm, SOD2 in the mitochondrial matrix, and SOD3 in the extracellular region [8].

Hydrogen peroxide  $(H_2O_2)$  has an unstable peroxide bond, which can oxidize proteins, lipids, and DNA. Mitochondrial respiratory chain is a continuous source of  $H_2O_2$  resulting from the dismutation of  $O_2^{\bullet-}$  by SOD [9]. Upon  $H_2O_2$  generation, CAT rapidly detoxifies it to molecular oxygen and  $H_2O$  [10]. CAT, which is enriched in peroxisomes, contains four iron-containing heme groups that allow it to react with  $H_2O_2$ . Glutathione system also eliminates  $H_2O_2$  from organisms [11].  $H_2O_2$  is reduced by glutathione peroxidase, which transfers the energy of the peroxide to GSH, a small sulfur-containing tripeptide. Since glutathione exists in reduced (GSH) and oxidized (GSSG) states, the ratio of GSH to GSSG can be a sensor of oxidative stress [12]. As  $H_2O_2$  has a lower activity than the hydroxyl radical, it remains in the nucleus for a longer time, which further damages DNA. Although  $H_2O_2$  has cytotoxic effects, it also acts as a signaling molecule in regulating various biological processes, such as immune cell activation and vascular remodeling [13]. Therefore, its production is highly regulated as part of redox homeostasis.

Hydroxyl radical (•OH), the three-electron reduction state of  $O_2^{\bullet-}$ , is an extremely reactive and short-lived ROS. It is produced by reduction of  $H_2O_2$  or due to the immune responses in macrophages and microglia [14]. The hydroxyl radical damages almost all types of molecules, such as nucleic acids, lipids, and amino acids. In addition, the hydroxyl radical attacks the plasma membrane, causing membrane damage. Due to its high reactivity, the hydroxyl radical cannot be used as a substrate for any enzyme and therefore cannot be removed by enzymatic reactions [15]. The hydroxyl radical is neutralized by toxic reactions with adjacent oxidizable molecules.

Lipid peroxidation is a process that occurs when free radicals attack nucleophilic polyunsaturated fatty acids in lipids. Since lipids are particularly abundant in the cell membrane, lipid peroxidation leads to membrane lipid degradation and cell damage. It produces lipid hydroperoxides (LOOH), such as malondialdehyde, 4-hydroxynonenal (HNE), 4-oxo-2-nonenal, and acrolein, thereby causing oxidative stress. For example, modification of mRNA expression or the activity of antioxidant enzymes increases, as the amount of HNE increases due to lipid peroxidation [16]. Thus, HNE is often regarded as a marker of oxidative stress.

The accumulation of ROS, resulting from the failure to maintain redox homeostasis, permanently induces increased oxidative stress, thereby leading to the functional loss of proteins, cellular apoptosis, mutagenesis, carcinogenesis, and fibrosis [17–19]. Increased ROS levels are associated with oxidative DNA damage and genomic instability [20]. NADPH oxidase-derived ROS were identified as the main cause of fibrotic diseases [21]. These harmful effects are known to cause various diseases, including cancer, metabolic disorders, chronic inflammation, cardiovascular diseases, and neurodegenerative diseases (Figure 1) [22–24].



Cancer, metabolic disorders, and neurodegenerative diseases

**Figure 1.** Schematic of the biological processes regulating cellular redox homeostasis. ROS are generated from various sources due to incomplete reduction of oxygen, causing oxidative stress. The elimination of ROS is tightly regulated by antioxidants, such as SOD, to maintain homeostasis and avoid oxidative stress. However, dysregulation of oxidative stress induces various diseases such as cancer, metabolic disorders, and neurodegenerative diseases.  $O_2^{\bullet-}$ : superoxide anion,  $H_2O_2$ : hydrogen peroxide,  $^{\bullet}OH$ : hydroxyl radical, HNE: 4-hydroxynonenal, SOD: superoxide dismutase, CAT: catalase, GSH: glutathione, and Prx: peroxiredoxin.

#### 3. Dysregulation of Redox Homeostasis in Neurodegenerative Diseases

ROS, at lower levels, generally act as a small molecule messenger for cell signaling in the brain, whereas, at higher levels, they induce protein misfolding and neuronal cell death [18,25,26]. The brain requires large amounts of oxygen for mitochondrial respiration, which inevitably generates ROS. Conversely, the brain is very susceptible to excessive formation of ROS, as it is rich in polyunsaturated fatty acids, which are the preferred substrates of ROS, and it shows a low expression of antioxidants such as GSH [27,28]. Therefore, it was recognized that oxidative stress contributes to the pathogenesis of neurodegenerative diseases [24,29].

Although it is unclear whether it is the cause or result, a decrease in antioxidant levels and an increase in ROS levels are commonly observed in the nervous systems of patients with neurodegenerative diseases as well as animal models of such diseases, including Parkinson's disease, Alzheimer's disease, and amyotrophic lateral sclerosis [30–34]. Furthermore, aging, the major risk factor for most neurodegenerative diseases, induces loss of mitochondrial function and an imbalance in GSH homeostasis [35,36]. As a result, aginginduced pro-oxidizing states in the mitochondria and cells cause synaptic dysfunction, such as impairment of synaptic activity and neurotransmission [37]. In particular, increased ROS levels results in the thermodynamically unstable redox states of proteins, leading to their misfolding [38]. Various studies reported that oxidative stress triggers the formation of malfunctional proteins in the central nervous system, leading to abnormal release of neurotransmitters, and the formation of protein aggregates, which is a hallmark of several neurodegenerative diseases [26,39].

#### 3.1. Alzheimer's Disease

Alzheimer's disease (AD), the most common neurodegenerative disease, is a progressive brain disorder that accounts for approximately 70% of dementia cases. AD is mainly characterized by the formation of extracellular amyloid-beta (A $\beta$ ) plaques, neurofibrillary tau tangles, and neuronal loss. Disruption of redox homeostasis was also observed in patients with AD [23,27]. Brain tissues or cells derived from patients with AD showed increased levels of ROS as well as oxidized proteins, and defects in antioxidant defense mechanisms [40–43]. Furthermore, the APP/PS1 double transgenic mice, which is a commonly used animal model of AD, showed a significant increase in ROS levels, which resulted in reduced neurogenesis in the brain [44]. In addition, increased ROS levels by H<sub>2</sub>O<sub>2</sub> or NO caused AD pathology, such as the occurrence of brain lesions and memory deficits [45,46].

The formation of A $\beta$  plaques or tau neurofibrillary tangles in the brain appears to be directly related to the impairment of redox homeostasis. A $\beta$ , a small peptide derived from the cleavage of amyloid precursor protein (APP) by BACE1 and  $\gamma$ -secretase, forms oligomeric fibrils in the brains of patients with AD. As mentioned above, ROS facilitate the formation of neurofibrillary tangles by promoting the misfolding and cross-linking of proteins [23,47,48]. Some studies reported the potential role of Aβ monomers as antioxidants, depending on the metal ion concentration in vitro [49,50]. In contrast, other studies showed that oligometric A $\beta$ , the dominant form in patients with AD, promotes ROS production in cooperation with metal ions in vitro and in vivo [51-54]. In addition, inhibition of the  $\gamma$ -secretase activity was reported to reduce the levels of ROS and oxidize proteins, and to enhance resistance to oxidative stress [55]. A $\beta$  activates NADPH oxidase and interferes with Ca<sup>2+</sup> homeostasis and mitochondrial membrane potential, leading to impaired ROS homeostasis [53,56–58]. Tau neurofibrillary tangles, which contain oxidized molecules, also produce ROS by interacting with Cu<sup>2+</sup>, which in turn increases the formation of tau aggregates [59]. Furthermore, tau aggregation activates NADPH oxidase, which accelerates ROS-mediated neuronal death [60].

#### 3.2. Parkinson's Disease

Parkinson's disease (PD) is a chronic neurodegenerative disorder that mainly affects the motor system; its primary symptoms are rigidity, tremor, bradykinesia, and postural instability. PD is characterized by the loss of dopaminergic neurons in the substantia nigra and accumulation of abnormal  $\alpha$ -synuclein aggregates called Lewy bodies. The neuropathological characterization of PD appears to be directly related to the production of ROS, owing to mitochondrial dysfunction, neuroinflammation, and genetic mutations. Brain tissues derived from patients with PD showed defects in mitochondrial complex I and an increase in the levels of oxidized proteins [61]. Conversely, a systemic partial defect in the mitochondrial complex I is sufficient to reproduce the neuropathological features observed in patients with PD. Mutations in DJ-1 cause autosomal recessive forms of PD. Oxidized DJ-1 was observed in the brains of patients with idiopathic PD [62]. DJ-1 knock-out (KO) mice exhibited a significant increase in  $\alpha$ -synuclein levels in the substantia nigra, increased levels of oxidized dopamine, and diminished the activity of glucocerebrosidase [63]. A reduction in GSH levels, which causes redox imbalance, was observed in the substantia nigra of patients with PD [64].

Oxidative stress is thought to be directly linked to PD pathogenesis. First, oxidative stress modifies and disrupts the activity of glucocerebrosidase, encoded by GBA1, whose mutations are commonly known genetic risk factors involved in the development of PD [65]. Second,  $\alpha$ -synuclein accumulation appears to be directly affected by oxidative stress.  $\alpha$ -Synuclein binds to ubiquitin and forms Lewy bodies. Oxidative stress leads

to post-translational modifications of  $\alpha$ -synuclein, such as HNE- $\alpha$ -synuclein and n- $\alpha$ synuclein, which are more toxic than unmodified  $\alpha$ -synuclein and are prone to form oligomers [66]. In particular, dopaminergic neurons in PD are susceptible to oxidative stress, as the dopamine metabolism generates ROS [67]. Levels of oxidized cholesterol metabolites derived from ROS were found to be increased in the cortex of patients with PD, resulting in accelerated generation of  $\alpha$ -synuclein aggregates [68]. Brain tissues derived from patients with PD also showed oxidized nucleic acids, lipids, and proteins generated due to oxidative damage, indicating that oxidative damage of nucleic acids is probably a major risk factor for PD [69]. It is plausible that oxidative stress can trigger mutations resulting in cellular dysfunction, thereby increasing the chance of spontaneous mutations induced by this process [70]. The cytokine tumor necrosis factor (TNF), a critical regulator of the immune system, can contribute to neurodegeneration by promoting the generation and release of ROS or by exacerbating ROS generation by activating NADPH oxidase [71].

#### 3.3. Amyotrophic Lateral Sclerosis

Amyotrophic lateral sclerosis (ALS) is a progressive neurological disease characterized by the death of motor neurons and leads to the impairment of voluntary muscle movement. Sporadic or familial ALS cases are caused by diverse gene mutations, including those in TAR DNA-binding protein 43 (TDP-43) and SOD1, the antioxidant enzyme [72]. SOD1 catalyzes the conversion of  $O_2^{\bullet-}$  to  $H_2O_2$  with  $Cu^{2+}$ . Therefore, the autosomal dominant mutations in SOD1, which account for up to 20% of familial ALS, increase the oxidative stress prior to the onset of ALS symptoms [73–75]. In addition, increased SOD1 activity boosted the production of ROS in the mitochondrial intermembrane space, resulting in morphological and functional impairment of the mitochondria [76,77]. Cells bearing the SOD1-G93A mutation are more vulnerable to ROS exposure, leading to the death of motor neurons [76,78]. Conversely, inhibition of SOD1 activity or treatment with antioxidants ameliorates the toxicity of motor neurons [76,79]. This increase in oxidative stress in cells caused SOD1 and TDP-43 protein aggregates, which are hallmarks of ALS [80,81]. As a result, the brain and motor neurons of patients with ALS as well as those of animal models of ALS exhibited increased levels of oxidized proteins, lipid peroxidation, and ROS, and decreased levels of antioxidants [82–87]. Furthermore, antioxidant treatment showed protective effects against neuronal survival and motor neuron impairment and decreased the ALS score [88,89].

# 4. Upregulation of Autophagy: An Effective Strategy to Maintain Redox Homeostasis in the Brain

#### 4.1. General Description of Autophagy

Autophagy is a major intracellular degradation process that is important for the survival and homeostasis of cells [90,91]. Stress-induced autophagy mediates the clearance of protein aggregates, damaged intracellular organelles, and ROS. Autophagy is induced by a variety of stress stimuli such as starvation, ER stress, hypoxia, mitochondrial dysfunction, and oxidative stress. Notably, ROS were reported to be crucial early inducers of autophagy as they are indispensable for the recruitment of LC3 into autophagosomes [92]. Autophagy is initiated by the formation and expansion of the phagophore to form the autophagosome, a double-membrane vesicle that sequesters cargoes to be degraded [93]. Autophagosomes then fuse with lysosomes to form autolysosomes, and the materials contained within them are degraded by the lysosomal enzymes. The degradation of cargoes by lysosomes generates nutrients that enable cells to survive [94]. After degradation, new lysosomes are formed from existing autolysosomes through autophagic lysosome reformation (ALR), a recently described lysosome biogenesis mechanism [95].

Increasing evidence suggests that autophagic impairments are closely linked to the pathogenesis of neurodegenerative disorders [96]. Mutations in autophagic receptors, such as p62 and optineurin (OPTN), are associated with a variety of neurodegenerative diseases, including PD, AD, and ALS [97,98]. The autophagic receptors are required

for the interaction and autophagosomal recruitment of specific cargoes for degradation. Accumulation of protein aggregates, mostly due to impaired autophagic degradation, is a common feature of neurodegenerative diseases. Therefore, it is reasonable that the malfunction of the autophagic receptors is related to the etiology of neurodegenerative diseases. Conversely, diverse genes associated with neurodegenerative diseases were shown to function in autophagy [96,99]. For example, PTEN-induced kinase 1 (PINK1) and parkin, which are often mutated in patients with early onset autosomal recessive PD, are well-known to be critical for mitophagy, a selective autophagy that degrades damaged mitochondria [100]. Presenilin-1, which is associated with familial AD, regulates the maturation of lysosomal v-ATPase and affects the autophagic degradation of cargoes [101].

#### 4.2. Regulatory Mechanism of ROS-Mediated Autophagy

Several studies demonstrated that oxidation of cellular molecules by ROS, such as peroxidation of unsaturated lipids and DNA or the production of mitochondrial ROS, is sufficient for activating autophagy, which can mitigate the cytotoxic stress caused by ROS (Figure 2) [102–104]. Conversely, inhibition of autophagy by lysosomal inhibitors causes an increase in ROS-induced cytotoxicity [105,106]. ROS can directly induce autophagy by activating protein kinases, including AMP-activated protein kinase (AMPK) and c-Jun N-terminal kinase (JNK), which are essential for the activation of autophagy [107–109]. For example, ROS directly oxidize Cys299/Cys304 of AMPK and activate its kinase activity without reducing the cellular ATP levels [110]. Mitochondrial ROS production also activates AMPK and triggers an antioxidant response [111,112]. In addition, ROS activate JNK by stimulating the ER stress response [102]. ROS can also influence autophagy via the oxidation of core autophagy proteins, such as ATG4 and TFEB [113,114].



**Figure 2.** ROS induce autophagy and mitophagy. Damaged mitochondria and cellular stresses generate ROS. ROS activate AMPK and inhibit mTORC1, leading to sequential activation of the autophagy initiation complex, the ULK1 and VPS34 complex. The activated autophagy initiation complex induces autophagosome formation. Autophagosomes engulf cargoes, such as damaged organelles and proteins, and fuse with lysosomes. Autolysosomes degrade the autophagic cargoes. Damaged mitochondria are targeted for degradation by a form of selective autophagy, called mitophagy. PINK1 is recruited to damaged mitochondria and phosphorylates parkin, an E3 ubiquitin ligase, and mitophagy receptors such as BNIP3L/NIX. Parkin-mediated poly-ubiquitination of OMM proteins leads to the recognition of damaged mitochondria by autophagic receptors. These autophagic receptors interact with the ATG8 family proteins and transport the damaged mitochondria to autophagosomes for their degradation. Detailed pathway is described in Section 4. AMPK: AMP-activated protein kinase, BNIP3L: Bcl2 interacting protein 3 like, mTORC1: mammalian target of rapamycin complex 1, OMM: outer mitochondria membrane proteins, and PINK1: PTEN-induced kinase 1.

#### 4.2.1. ROS-Mediated Regulation of Mitophagy

Mitochondrial dysfunction is strongly associated with various diseases, including neurodegenerative disorders. Therefore, it is important to minimize mitochondrial dysfunction to counteract the development of numerous human diseases [115]. Mutations in genes encoding mitochondrial quality control proteins were found in patients with PD, indicating their failure to protect against stress-induced mitochondrial dysfunction [116]. Dysfunctional mitochondria can be degraded by mitophagy. Engulfment of mitochondria by autophagosomes can occur under various conditions, including starvation and oxidative stress, and when the mitochondrial function is impaired [117,118]. ROS produced by mitochondria act as signaling molecules for stress-induced autophagy, and ROS produced by damaged mitochondria induce mitophagy for their elimination [118].

In response to ROS, parkin, an E3 ubiquitin ligase, is selectively recruited to damaged mitochondria, promoting mitochondrial degradation [119]. PINK1, a mitochondrial serine/threonine-protein kinase, is required for the recruitment of parkin to the damaged mitochondrial membrane and activates parkin ligase activity [120]. Parkin mediates the formation of poly-ubiquitin chains in clustered mitochondria. Subsequently, p62, the autophagic receptor that recognizes poly-ubiquitin chains, is recruited to the ubiquitinated mitochondria for their clearance [121]. Moreover, VDAC1, a voltage-gated channel protein in the mitochondria, was identified as a mitophagy target through parkin-mediatedubiquitylation. By interacting with LC3, mitophagy receptors, such as p62, mediate the targeting of damaged mitochondria into autophagosomes, for degradation [122]. Furthermore, BNIP3L/NIX, a BH3-only member of the BCL2 family, serves in mitochondrial clearance as a receptor that targets organelles for mitophagy [123].

#### 4.2.2. Transcriptional and Post-Transcriptional Regulation of Autophagy by ROS

TFEB is a master regulator of lysosomal biogenesis and autophagy [124]. Inactivated TFEB is retained in the cytoplasm, whereas activated TFEB translocates to the nucleus and promotes the transcription of genes essential for the autophagy–lysosome system. ROS directly oxidize the Cys212 residue of TFEB [113]. This oxidation inhibits the interaction of TFEB with RRAG GTPases, which promotes the translocation of TFEB to the nucleus, ultimately leading to the upregulation of lysosomal and autophagic pathways. ROS also induce transient receptor potential mucolipin 1 (TRPML1) channel-mediated lysosomal  $Ca^{2+}$  release [125]. Increased  $Ca^{2+}$  levels in the cytoplasm stimulate TFEB activity, which in turn promotes mitochondrial protein transcription, thereby reducing mitochondrial fragmentation, and recovering mitochondrial function [126,127]. Furthermore, overexpression of TFEB stabilizes Nrf2 by inhibiting DDB1 and CUL4-associated factor 11 (DCAF11), an E3 ligase of Nrf2, which enhances the transcription of Nrf2-response antioxidant genes, such as heme oxygenase-1 (HO-1), SOD2, and glutathione-s-transferase  $\mu1$  (GSTM1) [128].

Beclin-1, a component of the autophagy initiation complex (the VPS34 complex) that is essential for autophagy initiation, is regulated by various post-translational modifications [93,129]. ROS increase the protein levels and activity of Beclin-1 through the pro-apoptotic protein Bcl-2 [130,131]. Under normal conditions, Bcl-2 inhibits Beclin-1 activity via a direct binding to the BH domain within Beclin-1 [132,133]. Conversely, nutrient deprivation stimulates the JNK1-mediated phosphorylation of Bcl-2, leading to interference with its binding to Beclin-1 [134]. ROS decrease Bcl-2 protein levels through JNK-mediated Bcl-2 phosphorylation [135,136]. As a result, ROS increase Beclin-1 protein levels and promote autophagy initiation while simultaneously alleviating oxidative stress caused by ROS [130,131].

p62 is a representative autophagic receptor that recognizes ubiquitin-positive protein aggregates or mitochondria [137]. However, recent studies reported that p62 has a novel regulatory role in the Nrf2-Keap1 pathway, under oxidative stress conditions [138,139]. Nrf2 is a transcription factor that regulates several genes that encode antioxidants. Under normal conditions, the E3 ubiquitin ligase cullin 3 (CUL3) and ring-box 1 (RBX1) ubiquitinate Keap1, which interacts with Nrf2, leading to proteasomal degradation of Nrf2 [140].

Oxidative stress causes structural changes in Keap1 by generating an internal disulfide bond, resulting in the dissociation of Nrf2. The released Nrf2 binds to an antioxidant response element (ARE) sequence in target genes and suppresses the ROS-mediated toxicity [141,142]. p62 also interacts with Keap1 through the Kelch domain of Keap1, which is the Nrf2-binding site, and inhibits Keap1-mediated Nrf2 repression by producing Keap1 protein aggregates that are degraded by the proteasomal or autophagy–lysosomal system [138,143–145]. As the promoter of p62 contains an ARE sequence, activated Nrf2 can promote p62 transcription in response to oxidative stress [145]. The increased p62 levels can further activate Nrf2 [145,146]. Furthermore, since many promoters of autophagy core proteins, such as ATG5, ATG7, ATG9, ATG16L1, GABARAPL1, LAMP2A, and ULK1, contain ARE sequences, the brain tissues of Nrf2-deficient mice exhibit reduced protein levels of various autophagy regulators in response to ROS [147,148]. As a result, p62 and Nrf2 form a positive feedback loop to attenuate oxidative stress through activation of autophagy (Figure 3).



**Figure 3.** Autophagy-mediated regulation of ROS homeostasis. ROS promote autophagic activity by dissociating TFEB from RRAG GTPase and Beclin-1 from Bcl-2. ROS also activate Nrf2 through a conformational change in Keap1, leading to the dissociation of the Keap1-Nrf2 complex. Therefore, cells can ameliorate increased oxidative stress by the TFEB-and Nrf2-dependent upregulation of autophagic activity and antioxidant proteins. The detailed defense mechanisms are described in Section 4. TFEB: transcription factor EB, DCAF11: DDB1 and Cul4 associated factor 11, RBX1: Ring-box 1, CUL3: Cullin 3, ARE: antioxidant response element, JNK: c-Jun N-terminal kinase, HO1: Heme oxygenase 1, SOD2: Superoxide dismutase 2, and GSTM1: Glutathione-S-transferase μ1.

## 5. Autophagy as a Promising Therapeutic Strategy for Neurodegenerative Diseases

As described above, an imbalance in redox homeostasis contributes to the pathology of various neurodegenerative diseases, including AD, PD, and ALS, which exhibit increased ROS levels, as well as decreased levels of antioxidant proteins [37,42,76,87,149]. Various studies showed that adequate relief from oxidative stress can ameliorate the symptoms of neurodegenerative diseases, such as motor neuron death and cognitive dysfunction [45,53,76,149–151]. Therefore, induction of autophagy to eliminate ROS might be an appropriate therapeutic strategy for neurodegenerative diseases.

Treatment with small molecules can induce autophagy in an mTOR-dependent or -independent manner. Rapamycin, a representative autophagy inducer, inhibits the kinase activity of mTORC1. Rapamycin treatment reportedly reduced ROS production and improved abnormalities in neurodegenerative disease models [151–153]. For example, rapamycin treatment showed a protective effect by reducing intracellular ROS, as well

as increasing antioxidant protein levels in vitro and in vivo [152,153]. Furthermore, rapamycin treatment recovered impaired synaptic function, neurotransmission, and cognitive ability, by reversing impaired redox homeostasis in animal models of AD [151].

Excessive use of mTOR-dependent autophagy inducers can cause various side effects, as mTOR signaling is important for global translation. Trehalose induces autophagy by activating AMPK or TFEB in an mTOR-independent manner [154]. It was suggested that trehalose might exhibit a protective effect against oxidative stress by scavenging ROS via the upregulation of SOD activity in plants [155]. In mammalian cells, trehalose causes the dissociation of Nrf2 from Keap1 through upregulation of the p62-mediated Nrf2 positive feedback loop [156]. Trehalose also promotes autophagy, especially mitophagy, resulting in the amelioration of oxidative stress [157].

Since ROS directly affect the function of autophagy genes associated with various neurodegenerative diseases, upregulation of autophagy genes is considered to be a preventive and therapeutic strategy for controlling these diseases. Therefore, several studies examined whether genetic activation of autophagy genes could relieve oxidative stress and the symptoms of neurodegenerative diseases in animal models.

Overexpression of TFEB enhances the degradation of substrates, lipid droplets, and damaged mitochondria, and alleviates the phenotypes associated with neurodegenerative diseases, such as PD and AD, by promoting autophagy and lysosomal biogenesis [124]. Moreover, adeno-associated virus (AAV)-mediated overexpression of TFEB was reported to show neuroprotective effects in rat nigral dopaminergic neurons [158]. Notably, the injection of viral vectors induced local protection of neuronal cell bodies and improved dopamine neurotransmission.

In response to ROS stimulation, CHK2 binds to Beclin-1 and phosphorylates it at Ser90/Ser93, thereby promoting autophagy via Beclin-1 release from Bcl-2 sequestration [159]. Beclin-1-deficient mice showed disrupted neuronal autophagy, abnormal APP processing, and increased neurodegeneration [160]. Lentivirus-mediated Beclin-1 transfer into the brain of  $\alpha$ -synuclein transgenic mice relieved the synaptic and dendritic pathology with reduced  $\alpha$ -synuclein accumulation [161]. Therefore, upregulation of Beclin-1 levels might have therapeutic potential in AD and PD.

Parkin functions as a critical regulator of mitophagy. Mutations in the parkin gene are linked to an autosomal recessive form, known as autosomal recessive juvenile parkinsonism [162]. Parkin overexpression by lentivirus ameliorated the dopaminergic neuronal cell death induced by  $\alpha$ -synuclein overexpression in the rat substantia nigra [163]. In addition, parkin overexpression, using recombinant AAV (rAAV) in striatum of macaque monkeys, reduced the accumulation of co-expressed  $\alpha$ -synuclein [164]. Therefore, parkin gene therapy is thought to be effective against  $\alpha$ -synucleinopathy, suggesting its potential suitability for patients with PD [165].

DJ-1 is known to orchestrate a defense mechanism against oxidative stress, as its deletion leads to increased mitochondrial oxidative stress [166]. Dopamine adducts can modify  $\alpha$ -synuclein and promote its aggregation; a recent study showed that loss of DJ-1 function is associated with Lewy body pathology [167]. As described earlier, DJ-1 KO mice showed increased levels of  $\alpha$ -synuclein aggregation in the substantia nigra, accompanied by increased levels of oxidized dopamine [63]. Conversely, overexpression of DJ-1 in vitro effectively reduced the  $\alpha$ -synuclein levels [168,169].

## 6. Conclusions

In this review, we discussed the intersection between redox homeostasis and autophagy, focusing on the pathogenesis of neurodegenerative diseases. Increasing evidence further clarifies that the disruption of redox homeostasis due to various reasons is a major cause of neurodegenerative diseases. When redox homeostasis is disrupted, cells upregulate antioxidant responses to restore it. Various studies revealed the potential role of autophagy in exerting a protective effect against oxidative stress, along with other antioxidant responses. In this context, it is necessary to elucidate the regulatory roles of autophagy in redox homeostasis in greater details. It is expected that optimal upregulation of autophagy could be a desirable therapeutic strategy to ameliorate neurodegenerative diseases.

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## References

- 1. Boveris, A.; Oshino, N.; Chance, B. The cellular production of hydrogen peroxide. *Biochem. J.* **1972**, *128*, 617–630. [CrossRef] [PubMed]
- Bedard, K.; Krause, K.H. The NOX family of ROS-generating NADPH oxidases: Physiology and pathophysiology. *Physiol. Rev.* 2007, 87, 245–313. [CrossRef] [PubMed]
- Linhart, K.; Bartsch, H.; Seitz, H.K. The role of reactive oxygen species (ROS) and cytochrome P-450 2E1 in the generation of carcinogenic etheno-DNA adducts. *Redox Biol.* 2014, 3, 56–62. [CrossRef] [PubMed]
- 4. Brigelius-Flohe, R.; Flohe, L. Basic principles and emerging concepts in the redox control of transcription factors. *Antioxid. Redox Signal.* **2011**, *15*, 2335–2381. [CrossRef] [PubMed]
- Ray, P.D.; Huang, B.W.; Tsuji, Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell. Signal.* 2012, 24, 981–990. [CrossRef]
- 6. Turrens, J.F. Mitochondrial formation of reactive oxygen species. J. Physiol. 2003, 552, 335–344. [CrossRef]
- Cathcart, M.K. Regulation of superoxide anion production by NADPH oxidase in monocytes/macrophages: Contributions to atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 2004, 24, 23–28. [CrossRef]
- Fukai, T.; Ushio-Fukai, M. Superoxide dismutases: Role in redox signaling, vascular function, and diseases. *Antioxid. Redox Signal.* 2011, 15, 1583–1606. [CrossRef]
- Wong, H.S.; Dighe, P.A.; Mezera, V.; Monternier, P.A.; Brand, M.D. Production of superoxide and hydrogen peroxide from specific mitochondrial sites under different bioenergetic conditions. J. Biol. Chem. 2017, 292, 16804–16809. [CrossRef]
- 10. Alfonso-Pireto, M.; Biarnes, X.; Vidossich, P.; Rovira, C. The molecular mechanism of the catalase reaction. *J. Am. Chem. Soc.* 2009, 131, 11751–11761. [CrossRef]
- 11. Reiter, R.J.; Melchiorri, D.; Sewerynek, E.; Poeggeler, B.; Barlow-Walden, L.; Cjuang, J.; Ortiz, G.G.; Acuna-Castroviejo, D. A review of the evidence supporting melatonin's role as an antioxidant. *J. Pienal. Res.* **1995**, *18*, 1–11. [CrossRef]
- 12. Halprin, K.M.; Ohkawara, A. The measurement of glutathione in human epidermis using glutathione reductase. *J. Investig. Dermatol.* **1967**, *48*, 149–152. [CrossRef]
- 13. Veal, E.A.; Day, A.M.; Morgan, B.A. Hydrogen peroxide sensing and signaling. Mol. Cell 2007, 26, 1–14. [CrossRef]
- 14. Hoidal, J.R.; Beall, G.D.; Repine, J.E. Production of hydroxyl radical by human alveolar macrophages. *Infect. Immun.* **1979**, *26*, 1088–1092. [CrossRef]
- 15. Hrycay, E.G.; Bandiera, S.M. Involvement of Cytochrome P450 in Reactive Oxygen Species Formation and Cancer. *Adv. Pharmacol.* **2015**, *74*, 35–84. [CrossRef]
- 16. Zhang, H.; Court, N.; Forman, H.J. Submicromolar concentrations of 4-hydroxynonenal induce glutamate cysteine ligase expression in HBE1 cells. *Redox Rep.* 2007, *12*, 101–106. [CrossRef]
- 17. Ursini, F.; Maiorino, M.; Forman, H.J. Redox homeostasis: The Golden Mean of healthy living. *Redox Biol.* **2016**, *8*, 205–215. [CrossRef] [PubMed]
- 18. Gandhi, S.; Abramov, A.Y. Mechanism of oxidative stress in neurodegeneration. *Oxid. Med. Cell. Longev.* **2012**, 2012, 428010. [CrossRef] [PubMed]
- Alfadda, A.A.; Sallam, R.M. Reactive oxygen species in health and disease. J. Biomed. Biotechnol. 2012, 2012, 936486. [CrossRef] [PubMed]
- 20. Holmstrom, K.M.; Finkel, T. Cellular mechanisms and physiological consequences of redox-dependent signalling. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 411–421. [CrossRef] [PubMed]
- Babalola, O.; Mamalis, A.; Lev-Tov, H.; Jagdeo, J. NADPH oxidase enzymes in skin fibrosis: Molecular targets and therapeutic agents. Arch. Dermatol. Res. 2014, 306, 313–330. [CrossRef] [PubMed]
- 22. McBean, G.J.; Aslan, M.; Griffiths, H.R.; Torrao, R.C. Thiol redox homeostasis in neurodegenerative disease. *Redox Biol.* 2015, *5*, 186–194. [CrossRef] [PubMed]
- 23. Cheignon, C.; Tomas, M.; Bonnefont-Rousselot, D.; Faller, P.; Hureau, C.; Collin, F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol.* 2018, 14, 450–464. [CrossRef] [PubMed]
- 24. Halliwell, B. Free radicals and antioxidants: Updating a personal view. Nutr. Rev. 2012, 70, 257–265. [CrossRef] [PubMed]

- Hekimi, S.; Lapointe, J.; Wen, Y. Taking a "good" look at free radicals in the aging process. *Trends Cell Biol.* 2011, 21, 569–576. [CrossRef] [PubMed]
- 26. Bao, L.; Avshalumov, M.V.; Patel, J.C.; Lee, C.R.; Miller, E.W.; Chang, C.J.; Rice, M.E. Mitochondria are the source of hydrogen peroxide for dynamic brain-cell signaling. *J. Neurosci.* 2009, 29, 9002–9010. [CrossRef]
- 27. Wang, X.; Wang, W.; Li, L.; Perry, G.; Lee, H.G.; Zhu, X. Oxidative stress and mitochondrial dysfunction in Alzheime's disease. *Biochim. Biophys. Acta* 2014, 1842, 1240–1247. [CrossRef] [PubMed]
- 28. Raps, S.P.; Lai, J.C.; Hertz, L.; Cooper, A.J. Glutathione is present in high concentrations in cultured astrocytes but not in cultured neurons. *Brain Res.* **1989**, *493*, 398–401. [CrossRef]
- 29. Formella, I.; Svahn, A.J.; Radford, R.A.W.; Don, E.K.; Cole, N.J.; Hogan, A.; Lee, A.; Chung, R.S.; Morsch, M. Real-time visualization of oxidative stress-mediated neurodegeneration of individual spinal motor neurons in vivo. *Redox Biol.* **2018**, *19*, 226–234. [CrossRef]
- 30. Wiedemann, F.R.; Manfredi, G.; Mawrin, C.; Beal, M.F.; Schone, E.A. Mitochondrial DNA and respiratory chain function in spinal cords of ALS patients. *J. Neurochem.* 2002, *80*, 616–625. [CrossRef]
- 31. Mattiazzi, M.; D'Aurelio, M.; Gajewski, C.D.; Martushova, K.; Kiaei, M.; Beal, M.F.; Manfredi, G. Mutated human SOD1 causes dysfunction of oxidative phosphorylation in mitochondria of transgenic mice. J. Biol. Chem. 2002, 277, 29626–29633. [CrossRef]
- 32. Mandal, P.K.; Saharan, S.; Tripathi, M.; Murari, G. Brain glutathione levels—A novel biomarker for mild cognitive impairment and Alzheimer's disease. *Biol. Psychiatry* 2015, *78*, 702–710. [CrossRef]
- 33. Harish, G.; Mahadevan, A.; Srinivas Bharath, M.M.; Shankar, S.K. Alteration in glutathione content and associated enzyme activities in the synaptic terminals but not in the non-synaptic mitochondria from the frontal cortex of Parkinson's disease brains. *Neurochem. Res.* **2013**, *38*, 186–200. [CrossRef] [PubMed]
- 34. Pearce, R.K.; Owen, A.; Daniel, S.; Jenner, P.; Marsden, S.D. Alterations in the distribution of glutathione in the substantia nigra in Parkinson's disease. *J. Neural Transm.* **1997**, *104*, 661–677. [CrossRef] [PubMed]
- Rebrin, I.; Kamzalov, S.; Sohal, R.S. Effects of age and caloric restriction on glutathione redox state in mice. *Free Radic. Biol. Med.* 2003, *35*, 626–635. [CrossRef]
- Aliev, G.; Liu, J.; Shenk, J.C.; Fischbach, K.; Pacheco, G.J.; Chen, S.G.; Obrenovich, M.E.; Ward, W.F.; Richardson, A.G.; Smith, M.A.; et al. Neuronal mitochondrial amelioration by feeding acetyl-L-carnitine and lipoic acid to aged rats. *J. Cell. Mol. Med.* 2009, 13, 320–333. [CrossRef] [PubMed]
- 37. Tonnies, E.; Trushina, E. Oxidative Stress, Synaptic Dysfunction, and Alzheimer's Disease. J. Alzheimers Dis. 2017, 57, 1105–1121. [CrossRef]
- 38. Squier, T.C. Oxidative stress and protein aggregation during biological aging. Exp. Gerontol. 2001, 36, 1539–1550. [CrossRef]
- Levy, E.; El Banna, N.; Baille, D.; Heneman-Masurel, A.; Truchet, S.; Rezaei, H.; Huang, M.E.; Beringue, V.; Martin, D.; Vernis, L. Causative Links between Protein Aggregation and Oxidative Stress: A Review. *Int. J. Mol. Sci.* 2019, 20, 3896. [CrossRef] [PubMed]
- Cosin-Tomas, M.; Senserrich, J.; Arumi-Planas, M.; Alquezar, C.; Pallas, M.; Martin-Requero, A.; Sunol, C.; Kaliman, P.; Sanfeliu, C. Role of Resveratrol and Selenium on Oxidative Stress and Expression of Antioxidant and Anti-Aging Genes in Immortalized Lymphocytes from Alzheimer's Disease Patients. *Nutrients* 2019, *11*, 1764. [CrossRef]
- 41. Smith, M.A.; Richey Harris, P.L.; Sayre, L.M.; Beckman, J.S.; Perry, G. Widespread peroxynitrite-mediated damage in Alzheimer's disease. *J. Neurosci.* 1997, 17, 2653–2657. [CrossRef]
- Leutner, S.; Czech, C.; Schindowski, K.; Touchet, N.; Eckert, A.; Muller, W.E. Reduced antioxidant enzyme activity in brains of mice transgenic for human presenilin-1 with single or multiple mutations. *Neurosci. Lett.* 2000, 292, 87–90. [CrossRef]
- 43. Doria, M.; Nury, T.; Delmas, D.; Moreau, T.; Lizard, G.; Vejux, A. Protective function of autophagy during VLCFA-induced cytotoxicity in a neurodegenerative cell model. *Free Radic. Biol. Med.* **2019**, *137*, 46–58. [CrossRef] [PubMed]
- 44. Hamilton, A.; Holscher, C. The effect of ageing on neurogenesis and oxidative stress in the APP(swe)/PS1(deltaE9) mouse model of Alzheimer's disease. *Brain Res.* 2012, 1449, 83–93. [CrossRef] [PubMed]
- Yan, J.; Pang, Y.; Zhuang, J.; Lin, H.; Zhang, Q.; Han, L.; Ke, P.; Zhuang, J.; Huang, X. Selenepezil, a Selenium-Containing Compound, Exerts Neuroprotective Effect via Modulation of the Keap1-Nrf2-ARE Pathway and Attenuates Abeta-Induced Cognitive Impairment in Vivo. ACS Chem. Neurosci. 2019, 10, 2903–2914. [CrossRef] [PubMed]
- 46. Aliev, G.; Palacios, H.H.; Lipsitt, A.E.; Fischbach, K.; Lamb, B.T.; Obrenovich, M.E.; Morales, L.; Gasimov, E.; Bragin, V. Nitric oxide as an initiator of brain lesions during the development of Alzheimer disease. *Neurotox. Res.* 2009, *16*, 293–305. [CrossRef]
- 47. Mattson, M.P. Free radicals and disruption of neuronal ion homeostasis in AD: A role for amyloid beta-peptide? *Neurobiol. Aging* **1995**, *16*, 679–682. [CrossRef]
- 48. Sayre, L.M.; Zelasko, D.A.; Harris, P.L.; Perry, G.; Salomon, R.G.; Smith, M.A. 4-Hydroxynonenal-derived advanced lipid peroxidation end products are increased in Alzheimer's disease. *J. Neurochem.* **1997**, *68*, 2092–2097. [CrossRef] [PubMed]
- 49. Mitra, S.; Prasad, P.; Chakraborty, S. A Unified View of Assessing the Pro-oxidant versus Antioxidant Nature of Amyloid Beta Conformers. *Chembiochem* **2018**, *19*, 2360–2371. [CrossRef]
- 50. Baruch-Suchodolsky, R.; Fischer, B. Abeta40, either soluble or aggregated, is a remarkably potent antioxidant in cell-free oxidative systems. *Biochemistry* **2009**, *48*, 4354–4370. [CrossRef]

- Hensley, K.; Carney, J.M.; Mattson, M.P.; Aksenova, M.; Harris, M.; Wu, J.F.; Floyd, R.A.; Butterfield, D.A. A model for betaamyloid aggregation and neurotoxicity based on free radical generation by the peptide: Relevance to Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 1994, 91, 3270–3274. [CrossRef] [PubMed]
- 52. Cheignon, C.; Faller, P.; Testemale, D.; Hureau, C.; Collin, F. Metal-catalyzed oxidation of Abeta and the resulting reorganization of Cu binding sites promote ROS production. *Metallomics* **2016**, *8*, 1081–1089. [CrossRef]
- 53. Sotolongo, K.; Ghiso, J.; Rostagno, A. Nrf2 activation through the PI3K/GSK-3 axis protects neuronal cells from Abeta-mediated oxidative and metabolic damage. *Alzheimers Res. Ther.* **2020**, *12*, 13. [CrossRef]
- 54. Ejaz, H.W.; Wang, W.; Lang, M. Copper Toxicity Links to Pathogenesis of Alzheimer's Disease and Therapeutics Approaches. *Int. J. Mol. Sci.* **2020**, *21*, 7660. [CrossRef] [PubMed]
- 55. Sheng, B.; Gong, K.; Niu, Y.; Liu, L.; Yan, Y.; Lu, G.; Zhang, L.; Hu, M.; Zhao, N.; Zhang, X.; et al. Inhibition of gamma-secretase activity reduces Abeta production, reduces oxidative stress, increases mitochondrial activity and leads to reduced vulnerability to apoptosis: Implications for the treatment of Alzheimer's disease. *Free Radic. Biol. Med.* 2009, 46, 1362–1375. [CrossRef] [PubMed]
- Shelat, P.B.; Chalimoniuk, M.; Wang, J.H.; Strosznajder, J.B.; Lee, J.C.; Sun, A.Y.; Simonyi, A.; Sun, G.Y. Amyloid beta peptide and NMDA induce ROS from NADPH oxidase and AA release from cytosolic phospholipase A2 in cortical neurons. *J. Neurochem.* 2008, 106, 45–55. [CrossRef] [PubMed]
- 57. Reddy, P.H. Amyloid precursor protein-mediated free radicals and oxidative damage: Implications for the development and progression of Alzheimer's disease. *J. Neurochem.* **2006**, *96*, 1–13. [CrossRef] [PubMed]
- 58. Ferreiro, E.; Oliveira, C.R.; Pereira, C.M.F. The release of calcium from the endoplasmic reticulum induced by amyloid-beta and prion peptides activates the mitochondrial apoptotic pathway. *Neurobiol. Dis.* **2008**, *30*, 331–342. [CrossRef] [PubMed]
- Ahmadi, S.; Zhu, S.; Sharma, R.; Wu, B.; Soong, R.; Dutta Majumdar, R.; Wilson, D.J.; Simpson, A.J.; Kraatz, H.B. Aggregation of Microtubule Binding Repeats of Tau Protein is Promoted by Cu(2). ACS Omega 2019, 4, 5356–5366. [CrossRef]
- Esteras, N.; Kundel, F.; Amodeo, G.F.; Pavlov, E.V.; Klenerman, D.; Abramov, A.Y. Insoluble tau aggregates induce neuronal death through modification of membrane ion conductance, activation of voltage-gated calcium channels and NADPH oxidase. *FEBS J.* 2020. [CrossRef]
- 61. Betarbet, R.; Sherer, T.B.; Mackenzie, G.; Garcia-Osuna, M.; Panov, A.V.; Greenamyre, J.T. Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nat. Neurosci.* 2000, *3*, 1301–1306. [CrossRef]
- 62. Saito, Y.; Hamakubo, T.; Yoshida, Y.; Ogawa, Y.; Hara, Y.; Fujimura, H.; Imai, Y.; Iwanari, H.; Mochizuki, Y.; Shichiri, M.; et al. Preparation and application of monoclonal antibodies against oxidized DJ-1. Significant elevation of oxidized DJ-1 in erythrocytes of early-stage Parkinson disease patients. *Neurosci. Lett.* **2009**, *465*, 1–5. [CrossRef]
- Burbulla, L.F.; Song, P.; Mazzulli, J.R.; Zampese, E.; Wong, Y.C.; Jeon, S.; Santos, D.P.; Blanz, J.; Obermaier, C.D.; Strojny, C.; et al. Dopamine oxidation mediates mitochondrial and lysosomal dysfunction in Parkinson's disease. *Science* 2017, 357, 1255–1261. [CrossRef] [PubMed]
- 64. Perry, T.L.; Godin, D.V.; Hansen, S. Parkinson's disease: A disorder due to nigral glutathione deficiency? *Neurosci. Lett.* **1982**, *33*, 305–310. [CrossRef]
- 65. Do, J.; McKinney, C.; Sharma, P.; Sidransky, E. Glucocerebrosidase and its relevance to Parkinson disease. *Mol. Neurodegener.* 2019, 14, 36. [CrossRef] [PubMed]
- Xiang, W.; Schlachetzki, J.C.; Helling, S.; Bussmann, J.C.; Berlinghof, M.; Schaffer, T.E.; Marcus, K.; Winkler, J.; Klucken, J.; Becker, C.M. Oxidative stress-induced posttranslational modifications of alpha-synuclein: Specific modification of alpha-synuclein by 4-hydroxy-2-nonenal increases dopaminergic toxicity. *Mol. Cell. Neurosci.* 2013, 54, 71–83. [CrossRef]
- 67. Lotharius, J.; Brundin, P. Pathogenesis of Parkinson's disease: Dopamine, vesicles and alpha-synuclein. *Nat. Rev. Neurosci.* 2002, *3*, 932–942. [CrossRef]
- Bosco, D.A.; Fowler, D.M.; Zhang, Q.; Nieva, J.; Powers, E.T.; Wentworth, P., Jr.; Lerner, R.A.; Kelly, J.W. Elevated levels of oxidized cholesterol metabolites in Lewy body disease brains accelerate alpha-synuclein fibrilization. *Nat. Chem. Biol.* 2006, 2, 249–253. [CrossRef]
- 69. Nakabeppu, Y.; Tsuchimoto, D.; Yamaguchi, H.; Sakumi, K. Oxidative damage in nucleic acids and Parkinson's disease. *J. Neurosci. Res.* 2007, *85*, 919–934. [CrossRef]
- 70. Puspita, L.; Chung, S.Y.; Shim, J.W. Oxidative stress and cellular pathologies in Parkinson's disease. *Mol. Brain* **2017**, *10*, 53. [CrossRef]
- 71. Fischer, R.; Maier, O. Interrelation of oxidative stress and inflammation in neurodegenerative disease: Role of TNF. *Oxid Med. Cell. Longev.* **2015**, 2015, 610813. [CrossRef]
- 72. SChen, S.; Sayana, P.; Zhang, X.; Le, W. Genetics of amyotrophic lateral sclerosis: An update. *Mol. Neurodegener.* 2013, 8. [CrossRef]
- Xiao, Y.; Karam, C.; Yi, J.; Zhang, L.; Li, X.; Yoon, D.; Wang, H.; Dhakal, K.; Ramlow, P.; Yu, T.; et al. ROS-related mitochondrial dysfunction in skeletal muscle of an ALS mouse model during the disease progression. *Pharmacol. Res.* 2018, 138, 25–36. [CrossRef] [PubMed]
- 74. Ahmed, M.S.; Hung, W.Y.; Zu, J.S.; Hockberger, P.; Siddique, T. Increased reactive oxygen species in familial amyotrophic lateral sclerosis with mutations in SOD1. *J. Neurol. Sci.* 2000, 176, 88–94. [CrossRef]

- Li, Q.; Spencer, N.Y.; Pantazis, N.J.; Engelhardt, J.F. Alsin and SOD1(G93A) proteins regulate endosomal reactive oxygen species production by glial cells and proinflammatory pathways responsible for neurotoxicity. *J. Biol. Chem.* 2011, 286, 40151–40162. [CrossRef] [PubMed]
- 76. Rojas, F.; Gonzalez, D.; Cortes, N.; Ampuero, E.; Hernandez, D.E.; Fritz, E.; Abarzua, S.; Martinez, A.; Elorza, A.A.; Alvarez, A.; et al. Reactive oxygen species trigger motoneuron death in non-cell-autonomous models of ALS through activation of c-Abl signaling. *Front. Cell. Neurosci.* 2015, 9, 203. [CrossRef]
- 77. Ahtoniemi, T.; Jaronen, M.; Keksa-Goldsteine, V.; Goldsteins, G.; Koistinaho, J. Mutant SOD1 from spinal cord of G93A rats is destabilized and binds to inner mitochondrial membrane. *Neurobiol. Dis.* **2008**, *32*, 479–485. [CrossRef] [PubMed]
- Richardson, K.; Allen, S.P.; Mortiboys, H.; Grierson, A.J.; Wharton, S.B.; Ince, P.G.; Shaw, P.J.; Heath, P.R. The effect of SOD1 mutation on cellular bioenergetic profile and viability in response to oxidative stress and influence of mutation-type. *PLoS ONE* 2013, *8*, e68256. [CrossRef] [PubMed]
- 79. Guareschi, S.; Cova, E.; Cereda, C.; Ceroni, M.; Donetti, E.; Bosco, D.A.; Trotti, D.; Pasinelli, P. An over-oxidized form of superoxide dismutase found in sporadic amyotrophic lateral sclerosis with bulbar onset shares a toxic mechanism with mutant SOD1. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 5074–5079. [CrossRef]
- Cohen, T.J.; Hwang, A.W.; Unger, T.; Trojanowski, J.Q.; Lee, V.M. Redox signalling directly regulates TDP-43 via cysteine oxidation and disulphide cross-linking. *EMBO J.* 2012, *31*, 1241–1252. [CrossRef]
- 81. Bakavayev, S.; Chetrit, N.; Zvagelsky, T.; Mansour, R.; Vyazmensky, M.; Barak, Z.; Israelson, A.; Engel, S. Cu/Zn-superoxide dismutase and wild-type like fALS SOD1 mutants produce cytotoxic quantities of H2O2 via cysteine-dependent redox short-circuit. *Sci. Rep.* **2019**, *9*, 10826. [CrossRef]
- Ikawa, M.; Okazawa, H.; Tsujikawa, T.; Matsunaga, A.; Yamamura, O.; Mori, T.; Hamano, T.; Kiyono, Y.; Nakamoto, Y.; Toneda, M. Increased oxidative stress is related to disease severity in the ALS motor cortex: A PET study. *Neurology* 2015, *84*, 2033–2039. [CrossRef] [PubMed]
- Shibata, N.; Nagai, R.; Uchida, K.; Horiuchi, S.; Yamada, S.; Hirano, A.; Kawaguchi, M.; Yamamoto, T.; Sasaki, S.; Kobayashi, M. Morphological evidence for lipid peroxidation and protein glycoxidation in spinal cords from sporadic amyotrophic lateral sclerosis patients. *Brain Res.* 2001, *917*, 97–104. [CrossRef]
- 84. Ferrante, R.J.; Shinobu, L.A.; Schulz, J.B.; Matthews, R.T.; Thomas, C.E.; Kowall, N.W.; Gurney, M.E.; Beal, M.F. Increased 3-nitrotyrosine and oxidative damage in mice with a human copper/zinc superoxide dismutase mutation. *Ann. Neurol.* **1997**, 42, 326–334. [CrossRef] [PubMed]
- 85. Andrus, P.K.; Fleck, T.J.; Gurney, M.E.; Hall, E.D. Protein oxidative damage in a transgenic mouse model of familial amyotrophic lateral sclerosis. *J. Neurochem.* **1998**, *71*, 2041–2048. [CrossRef] [PubMed]
- Cova, E.; Bongioanni, P.; Cereda, C.; Metelli, M.R.; Salvaneschi, L.; Bernuzzi, S.; Guareschi, S.; Rossi, B.; Ceroni, M. Time course of oxidant markers and antioxidant defenses in subgroups of amyotrophic lateral sclerosis patients. *Neurochem. Int.* 2010, 56, 687–693. [CrossRef]
- 87. Ehrhart, J.; Smith, A.J.; Kuzmin-Nichols, N.; Zesiewicz, T.A.; Jahan, I.; Shytle, R.D.; Kim, S.H.; Sanberg, C.D.; Vu, T.H.; Gooch, C.L.; et al. Humoral factors in ALS patients during disease progression. *J. Neuroinflamm.* **2015**, *12*, 127. [CrossRef] [PubMed]
- Zhang, C.; Yang, Y.; Liang, W.; Wang, T.; Wang, S.; Wang, X.; Wang, Y.; Jiang, H.; Feng, H. Neuroprotection by urate on the mutant hSOD1-related cellular and Drosophila models of amyotrophic lateral sclerosis: Implication for GSH synthesis via activating Akt/GSK3beta/Nrf2/GCLC pathways. *Brain Res. Bull.* 2019, 146, 287–301. [CrossRef] [PubMed]
- 89. Yoshino, H.; Kimura, A. Investigation of the therapeutic effects of edaravone, a free radical scavenger, on amyotrophic lateral sclerosis (Phase II study). *Amyotroph. Lateral Scler.* **2006**, *7*, 241–245. [CrossRef]
- 90. Boya, P.; Reggiori, F.; Codogno, P. Emerging regulation and functions of autophagy. Nat. Cell Biol. 2013, 15, 713–720. [CrossRef]
- 91. Mizushima, N.; Komatsu, M. Autophagy: Renovation of cells and tissues. Cell 2011, 147, 728–741. [CrossRef]
- 92. Filomeni, G.; De Zio, D.; Cecconi, F. Oxidative stress and autophagy: The clash between damage and metabolic needs. *Cell Death Differ.* **2015**, *22*, 377–388. [CrossRef]
- 93. Antonioli, M.; Di Rienzo, M.; Piacentini, M.; Fimia, G.M. Emerging Mechanisms in Initiating and Terminating Autophagy. *Trends Biochem. Sci.* 2017, 42, 28–41. [CrossRef]
- 94. Nakatogawa, H.; Suzuki, K.; Kamada, Y.; Ohsumi, Y. Dynamics and diversity in autophagy mechanisms: Lessons from yeast. *Nat. Rev. Mol. Cell Biol.* **2009**, *10*, 458–467. [CrossRef]
- 95. Yu, L.; McPhee, C.K.; Zheng, L.; Mardones, G.A.; Rong, Y.; Peng, J.; Mi, N.; Zhao, Y.; Liu, Z.; Wan, F.; et al. Termination of autophagy and reformation of lysosomes regulated by mTOR. *Nature* **2010**, *465*, 942–946. [CrossRef]
- Menzies, F.M.; Fleming, A.; Caricasole, A.; Bento, C.F.; Andrews, S.P.; Ashkenazi, A.; Fullgrabe, J.; Jackson, A.; Jimenez Sanchez, M.; Karabiyik, C.; et al. Autophagy and Neurodegeneration: Pathogenic Mechanisms and Therapeutic Opportunities. *Neuron* 2017, 93, 1015–1034. [CrossRef]
- 97. Deng, Z.; Purtell, K.; Lachance, V.; Wold, M.S.; Chen, S.; Yue, Z. Autophagy Receptors and Neurodegenerative Diseases. *Trends Cell Biol.* **2017**, *27*, 491–504. [CrossRef] [PubMed]
- 98. Park, H.; Kang, J.H.; Lee, S. Autophagy in Neurodegenerative Diseases: A Hunter for Aggregates. *Int. J. Mol. Sci.* 2020, 21, 3369. [CrossRef] [PubMed]
- 99. Scrivo, A.; Bourdenx, M.; Pampliega, O.; Cuervo, A.M. Selective autophagy as a potential therapeutic target for neurodegenerative disorders. *Lancet Neurol.* 2018, *17*, 802–815. [CrossRef]

- 100. Pickrell, A.M.; Youle, R.J. The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron* **2015**, *85*, 257–273. [CrossRef] [PubMed]
- Lee, J.H.; Yu, W.H.; Kumar, A.; Lee, S.; Mohan, P.S.; Peterhoff, C.M.; Wolfe, D.M.; Martinez-Vicente, M.; Massey, A.C.; Sovak, G.; et al. Lysosomal proteolysis and autophagy require presenilin 1 and are disrupted by Alzheimer-related PS1 mutations. *Cell* 2010, 141, 1146–1158. [CrossRef]
- Haberzettl, P.; Hill, B.G. Oxidized lipids activate autophagy in a JNK-dependent manner by stimulating the endoplasmic reticulum stress response. *Redox Biol.* 2013, 1, 56–64. [CrossRef] [PubMed]
- Galati, S.; Boni, C.; Gerra, M.C.; Lazzaretti, M.; Buschini, A. Autophagy: A Player in response to Oxidative Stress and DNA Damage. Oxid. Med. Cell. Longev. 2019, 2019, 5692958. [CrossRef] [PubMed]
- 104. Hill, B.G.; Haberzettl, P.; Ahmed, Y.; Srivastava, S.; Bhatnagar, A. Unsaturated lipid peroxidation-derived aldehydes activate autophagy in vascular smooth-muscle cells. *Biochem. J.* **2008**, *410*, 525–534. [CrossRef] [PubMed]
- 105. Zhao, L.; Li, H.; Wang, Y.; Zheng, A.; Cao, L.; Liu, J. Autophagy Deficiency Leads to Impaired Antioxidant Defense via p62-FOXO1/3 Axis. Oxid. Med. Cell. Longev. 2019, 2019, 2526314. [CrossRef]
- Xiong, Y.; Contento, A.L.; Nguyen, P.Q.; Bassham, D.C. Degradation of oxidized proteins by autophagy during oxidative stress in Arabidopsis. *Plant Physiol.* 2007, 143, 291–299. [CrossRef]
- 107. Kim, J.; Kundu, M.; Viollet, B.; Guan, K.L. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat. Cell Biol.* **2011**, *13*, 132–141. [CrossRef]
- 108. Zhou, Y.Y.; Li, Y.; Jiang, W.Q.; Zhou, L.F. MAPK/JNK signalling: A potential autophagy regulation pathway. *Biosci. Rep.* 2015, 35. [CrossRef]
- 109. Xu, P.; Das, M.; Reilly, J.; Davis, R.J. JNK regulates FoxO-dependent autophagy in neurons. *Genes Dev.* 2011, 25, 310–322. [CrossRef]
- 110. Zmijewski, J.W.; Banerjee, S.; Bae, H.; Friggeri, A.; Lazarowski, E.R.; Abraham, E. Exposure to hydrogen peroxide induces oxidation and activation of AMP-activated protein kinase. *J. Biol. Chem.* **2010**, *285*, 33154–33164. [CrossRef]
- 111. Hinchy, E.C.; Gruszczyk, A.V.; Willows, R.; Navaratnam, N.; Hall, A.R.; Bates, G.; Bright, T.P.; Krieg, T.; Carling, D.; Murphy, M.P. Mitochondria-derived ROS activate AMP-activated protein kinase (AMPK) indirectly. J. Biol. Chem. 2018, 293, 17208–17217. [CrossRef]
- Rabinovitch, R.C.; Samborska, B.; Faubert, B.; Ma, E.H.; Gravel, S.P.; Andrzejewski, S.; Raissi, T.C.; Pause, A.; St-Pierre, J.; Jones, R.G. AMPK Maintains Cellular Metabolic Homeostasis through Regulation of Mitochondrial Reactive Oxygen Species. *Cell Rep.* 2017, 21, 1–9. [CrossRef]
- 113. Wang, H.; Wang, N.; Xu, D.; Ma, Q.; Chen, Y.; Xu, S.; Xia, Q.; Zhang, Y.; Prehn, J.H.M.; Wang, G.; et al. Oxidation of multiple MiT/TFE transcription factors links oxidative stress to transcriptional control of autophagy and lysosome biogenesis. *Autophagy* 2020, 16, 1683–1696. [CrossRef]
- 114. Scherz-Shouval, R.; Shvets, E.; Fass, E.; Shorer, H.; Gil, L.; Elazar, Z. Reactive oxygen species are essential for autophagy and specifically regulate the activity of Atg4. *EMBO J.* **2007**, *26*, 1749–1760. [CrossRef] [PubMed]
- Wallace, D.C. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: A dawn for evolutionary medicine. *Annu. Rev. Genet.* 2005, 39, 359–407. [CrossRef] [PubMed]
- 116. Valente, E.M.; Abou-Sleiman, P.M.; Caputo, V.; Muqit, M.M.; Harvey, K.; Gispert, S.; Ali, Z.; Del Turco, D.; Bentivoglio, A.R.; Healy, D.G.; et al. Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science* **2004**, *304*, 1158–1160. [CrossRef]
- 117. Bampton, E.T.; Goemans, C.G.; Niranjan, D.; Mizushima, N.; Tolkovsky, A.M. The dynamics of autophagy visualized in live cells: From autophagosome formation to fusion with endo/lysosomes. *Autophagy* **2005**, *1*, 23–36. [CrossRef]
- 118. Kim, I.; Rodriguez-Enriquez, S.; Lemasters, J.J. Selective degradation of mitochondria by mitophagy. *Arch. Biochem. Biophys.* 2007, 462, 245–253. [CrossRef] [PubMed]
- Narendra, D.; Tanaka, A.; Suen, D.F.; Youle, R.J. Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. J. Cell Biol. 2008, 183, 795–803. [CrossRef]
- 120. Vives-Bauza, C.; Zhou, C.; Huang, Y.; Cui, M.; de Vries, R.L.; Kim, J.; May, J.; Tocilescu, M.A.; Liu, W.; Ko, H.S.; et al. PINK1dependent recruitment of Parkin to mitochondria in mitophagy. *Proc. Natl. Acad. Sci. USA* 2010, 107, 378–383. [CrossRef]
- Geisler, S.; Holmstrom, K.M.; Skujat, D.; Fiesel, F.C.; Rothfuss, O.C.; Kahle, P.J.; Springer, W. PINK1/Parkin-mediated mitophagy is dependent on VDAC1 and p62/SQSTM1. *Nat. Cell Biol.* 2010, *12*, 119–131. [CrossRef] [PubMed]
- 122. Abu-Hamad, S.; Arbel, N.; Calo, D.; Arzoine, L.; Israelson, A.; Keinan, N.; Ben-Romano, R.; Friedman, O.; Shoshan-Barmatz, V. The VDAC1 N-terminus is essential both for apoptosis and the protective effect of anti-apoptotic proteins. *J. Cell Sci.* 2009, 122, 1906–1916. [CrossRef]
- 123. Novak, I.; Kirkin, V.; McEwan, D.G.; Zhang, J.; Wild, P.; Rozenknop, A.; Rogov, V.; Lohr, F.; Popovic, D.; Occhipinti, A.; et al. Nix is a selective autophagy receptor for mitochondrial clearance. *EMBO Rep.* **2010**, *11*, 45–51. [CrossRef]
- 124. Napolitano, G.; Ballabio, A. TFEB at a glance. J. Cell Sci. 2016, 129, 2475–2481. [CrossRef]
- 125. Zhang, X.; Cheng, X.; Yu, L.; Yang, J.; Calvo, R.; Patnaik, S.; Hu, X.; Gao, Q.; Yang, M.; Lawas, M.; et al. MCOLN1 is a ROS sensor in lysosomes that regulates autophagy. *Nat. Commun.* 2016, 7, 12109. [CrossRef]
- 126. Ivankovic, D.; Chau, K.Y.; Schapira, A.H.; Gegg, M.E. Mitochondrial and lysosomal biogenesis are activated following PINK1/parkin-mediated mitophagy. J. Neurochem. 2016, 136, 388–402. [CrossRef]

- 127. Gambardella, G.; Staiano, L.; Moretti, M.N.; Cegli, R.; Fagnocchi, L.; Tullio, G.; Polletti, S.; Braccia, C.; Armirotti, A.; Zippo, A.; et al. GADD34 is a modulator of autophagy during starvation. *Sci. Adv.* **2020**, *6*, eabb0205. [CrossRef]
- 128. Park, J.Y.; Kim, S.; Sohn, H.Y.; Koh, Y.H.; Jo, C. TFEB activates Nrf2 by repressing its E3 ubiquitin ligase DCAF11 and promoting phosphorylation of p62. *Sci. Rep.* 2019, *9*, 14354. [CrossRef]
- 129. Hill, S.M.; Wrobel, L.; Rubinsztein, D.C. Post-translational modifications of Beclin 1 provide multiple strategies for autophagy regulation. *Cell Death Differ.* 2019, 26, 617–629. [CrossRef]
- 130. Wang, X.; Sun, D.; Hu, Y.; Xu, X.; Jiang, W.; Shang, H.; Cui, D. The roles of oxidative stress and Beclin-1 in the autophagosome clearance impairment triggered by cardiac arrest. *Free Radic. Biol. Med.* **2019**, *136*, 87–95. [CrossRef]
- 131. Zhang, H.; Kong, X.; Kang, J.; Su, J.; Li, Y.; Zhong, J.; Sun, L. Oxidative stress induces parallel autophagy and mitochondria dysfunction in human glioma U251 cells. *Toxicol. Sci.* 2009, *110*, 376–388. [CrossRef]
- 132. Erlich, S.; Mizrachy, L.; Segev, O.; Lindenboim, L.; Zmira, O.; Adi-Harel, S.; Hirsch, J.A.; Stein, R.; Pinkas-Kramarski, R. Differential interactions between Beclin 1 and Bcl-2 family members. *Autophagy* **2007**, *3*, 561–568. [CrossRef]
- Pattingre, S.; Tassa, A.; Qu, X.; Garuti, R.; Liang, X.H.; Mizushima, N.; Packer, M.; Schneider, M.D.; Levine, B. Bcl-2 antiapoptotic proteins inhibit Beclin 1-dependent autophagy. *Cell* 2005, 122, 927–939. [CrossRef]
- 134. Wei, Y.; Sinha, S.; Levine, B. Dual role of JNK1-mediated phosphorylation of Bcl-2 in autophagy and apoptosis regulation. *Autophagy* **2008**, *4*, 949–951. [CrossRef]
- 135. Hildeman, D.A.; Mitchell, T.; Aronow, B.; Wojciechowski, S.; Kappler, J.; Marrack, P. Control of Bcl-2 expression by reactive oxygen species. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 15035–15040. [CrossRef] [PubMed]
- Liu, G.Y.; Jiang, X.X.; Zhu, X.; He, W.Y.; Kuang, Y.L.; Ren, K.; Lin, Y.; Gou, X. ROS activates JNK-mediated autophagy to counteract apoptosis in mouse mesenchymal stem cells in vitro. *Acta Pharmacol. Sin.* 2015, *36*, 1473–1479. [CrossRef]
- 137. Lippai, M.; Low, P. The role of the selective adaptor p62 and ubiquitin-like proteins in autophagy. *BioMed Res. Int.* 2014, 2014, 832704. [CrossRef]
- 138. Komatsu, M.; Kurokawa, H.; Waguri, S.; Taguchi, K.; Kobayashi, A.; Ichimura, Y.; Sou, Y.S.; Ueno, I.; Sakamoto, A.; Tong, K.I.; et al. The selective autophagy substrate p62 activates the stress responsive transcription factor Nrf2 through inactivation of Keap1. Nat. Cell Biol. 2010, 12, 213–223. [CrossRef] [PubMed]
- Copple, I.M.; Lister, A.; Obeng, A.D.; Kitteringham, N.R.; Jenkins, R.E.; Layfield, R.; Foster, B.J.; Goldring, C.E.; Park, B.K. Physical and functional interaction of sequestosome 1 with Keap1 regulates the Keap1-Nrf2 cell defense pathway. *J. Biol. Chem.* 2010, 285, 16782–16788. [CrossRef] [PubMed]
- 140. Furukawa, M.; Xiong, Y. BTB protein Keap1 targets antioxidant transcription factor Nrf2 for ubiquitination by the Cullin 3-Roc1 ligase. *Mol. Cell. Biol.* 2005, *25*, 162–171. [CrossRef]
- 141. DeNicola, G.M.; Karreth, F.A.; Humpton, T.J.; Gopinathan, A.; Wei, C.; Frese, K.; Mangal, D.; Yu, K.H.; Yeo, C.J.; Calhoun, E.S.; et al. Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis. *Nature* 2011, 475, 106–109. [CrossRef]
- 142. Ma, Q. Role of nrf2 in oxidative stress and toxicity. Annu. Rev. Pharmacol. Toxicol. 2013, 53, 401–426. [CrossRef]
- 143. Lau, A.; Wang, X.J.; Zhao, F.; Villeneuve, N.F.; Wu, T.; Jiang, T.; Sun, Z.; White, E.; Zhang, D.D. A noncanonical mechanism of Nrf2 activation by autophagy deficiency: Direct interaction between Keap1 and p62. *Mol. Cell. Biol.* **2010**, *30*, 3275–3285. [CrossRef]
- 144. Ichimura, Y.; Waguri, S.; Sou, Y.S.; Kageyama, S.; Hasegawa, J.; Ishimura, R.; Saito, T.; Yang, Y.; Kouno, T.; Fukutomi, T.; et al. Phosphorylation of p62 activates the Keap1-Nrf2 pathway during selective autophagy. *Mol. Cell* **2013**, *51*, 618–631. [CrossRef]
- 145. Jain, A.; Lamark, T.; Sjottem, E.; Larsen, K.B.; Awuh, J.A.; Overvatn, A.; McMahon, M.; Hayes, J.D.; Johansen, T. p62/SQSTM1 is a target gene for transcription factor NRF2 and creates a positive feedback loop by inducing antioxidant response element-driven gene transcription. J. Biol. Chem. 2010, 285, 22576–22591. [CrossRef]
- 146. Lacher, S.E.; Levings, D.C.; Freeman, S.; Slattery, M. Identification of a functional antioxidant response element at the HIF1A locus. *Redox Biol.* **2018**, *19*, 401–411. [CrossRef]
- 147. Pajares, M.; Rojo, A.I.; Arias, E.; Diaz-Carretero, A.; Cuervo, A.M.; Cuadrado, A. Transcription factor NFE2L2/NRF2 modulates chaperone-mediated autophagy through the regulation of LAMP2A. *Autophagy* **2018**, *14*, 1310–1322. [CrossRef] [PubMed]
- 148. Pajares, M.; Jimenez-Moreno, N.; Garcia-Yague, A.J.; Escoll, M.; de Ceballos, M.L.; Van Leuven, F.; Rabano, A.; Yamamoto, M.; Rojo, A.I.; Cuadrado, A. Transcription factor NFE2L2/NRF2 is a regulator of macroautophagy genes. *Autophagy* 2016, 12, 1902–1916. [CrossRef]
- 149. Hettiarachchi, N.T.; Boyle, J.P.; Dallas, M.L.; Al-Owais, M.M.; Scragg, J.L.; Peers, C. Heme oxygenase-1 derived carbon monoxide suppresses Abeta1-42 toxicity in astrocytes. *Cell Death Dis.* **2017**, *8*, e2884. [CrossRef]
- Du, S.; Zhang, Y.; Yang, J.; Liu, X.; Wang, Y.; Xu, B.; Jia, J. Curcumin Alleviates beta Amyloid-Induced Neurotoxicity in HT22 Cells via Upregulating SOD2. J. Mol. Neurosci. 2019, 67, 540–549. [CrossRef]
- 151. Singh, A.K.; Kashyap, M.P.; Tripathi, V.K.; Singh, S.; Garg, G.; Rizvi, S.I. Neuroprotection Through Rapamycin-Induced Activation of Autophagy and PI3K/Akt1/mTOR/CREB Signaling Against Amyloid-beta-Induced Oxidative Stress, Synaptic/Neurotransmission Dysfunction, and Neurodegeneration in Adult Rats. *Mol. Neurobiol.* 2017, 54, 5815–5828. [CrossRef] [PubMed]

- 152. Martinez-Cisuelo, V.; Gomez, J.; Garcia-Junceda, I.; Naudi, A.; Cabre, R.; Mota-Martorell, N.; Lopez-Torres, M.; Gonzalez-Sanchez, M.; Pamplona, R.; Barja, G. Rapamycin reverses age-related increases in mitochondrial ROS production at complex I, oxidative stress, accumulation of mtDNA fragments inside nuclear DNA, and lipofuscin level, and increases autophagy, in the liver of middle-aged mice. *Exp. Gerontol.* 2016, *83*, 130–138. [CrossRef]
- 153. Shin, Y.J.; Cho, D.Y.; Chung, T.Y.; Han, S.B.; Hyon, J.Y.; Wee, W.R. Rapamycin reduces reactive oxygen species in cultured human corneal endothelial cells. *Curr. Eye Res.* 2011, *36*, 1116–1122. [CrossRef]
- 154. Rusmini, P.; Cortese, K.; Crippa, V.; Cristofani, R.; Cicardi, M.E.; Ferrari, V.; Vezzoli, G.; Tedesco, B.; Meroni, M.; Messi, E.; et al. Trehalose induces autophagy via lysosomal-mediated TFEB activation in models of motoneuron degeneration. *Autophagy* 2019, 15, 631–651. [CrossRef] [PubMed]
- 155. Luo, Y.; Li, W.-M.; Wang, W. Trehalose: Protector of antioxidant enzymes or reactive oxygen species scavenger under heat stress? *Environ. Exp. Bot.* **2008**, *63*, 378–384. [CrossRef]
- 156. Mizunoe, Y.; Kobayashi, M.; Sudo, Y.; Watanabe, S.; Yasukawa, H.; Natori, D.; Hoshino, A.; Negishi, A.; Okita, N.; Komatsu, M.; et al. Trehalose protects against oxidative stress by regulating the Keap1-Nrf2 and autophagy pathways. *Redox Biol.* 2018, 15, 115–124. [CrossRef]
- 157. Tang, Q.; Zheng, G.; Feng, Z.; Chen, Y.; Lou, Y.; Wang, C.; Zhang, X.; Zhang, Y.; Xu, H.; Shang, P.; et al. Trehalose ameliorates oxidative stress-mediated mitochondrial dysfunction and ER stress via selective autophagy stimulation and autophagic flux restoration in osteoarthritis development. *Cell Death Dis.* **2017**, *8*, e3081. [CrossRef]
- 158. Arotcarena, M.L.; Bourdenx, M.; Dutheil, N.; Thiolat, M.L.; Doudnikoff, E.; Dovero, S.; Ballabio, A.; Fernagut, P.O.; Meissner, W.G.; Bezard, E.; et al. Transcription factor EB overexpression prevents neurodegeneration in experimental synucleinopathies. *JCI Insight* 2019, 4. [CrossRef]
- 159. Guo, Q.Q.; Wang, S.S.; Zhang, S.S.; Xu, H.D.; Li, X.M.; Guan, Y.; Yi, F.; Zhou, T.T.; Jiang, B.; Bai, N.; et al. ATM-CHK2-Beclin 1 axis promotes autophagy to maintain ROS homeostasis under oxidative stress. *EMBO J.* **2020**, *39*, e103111. [CrossRef]
- 160. Pickford, F.; Masliah, E.; Britschgi, M.; Lucin, K.; Narasimhan, R.; Jaeger, P.A.; Small, S.; Spencer, B.; Rockenstein, E.; Levine, B.; et al. The autophagy-related protein beclin 1 shows reduced expression in early Alzheimer disease and regulates amyloid beta accumulation in mice. J. Clin. Investig. 2008, 118, 2190–2199. [CrossRef]
- 161. Spencer, B.; Potkar, R.; Trejo, M.; Rockenstein, E.; Patrick, C.; Gindi, R.; Adame, A.; Wyss-Coray, T.; Masliah, E. Beclin 1 gene transfer activates autophagy and ameliorates the neurodegenerative pathology in alpha-synuclein models of Parkinson's and Lewy body diseases. *J. Neurosci.* 2009, 29, 13578–13588. [CrossRef] [PubMed]
- 162. Kitada, T.; Asakawa, S.; Hattori, N.; Matsumine, H.; Yamamura, Y.; Minoshima, S.; Yokochi, M.; Mizuno, Y.; Shimizu, N. Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature* **1998**, *392*, 605–608. [CrossRef]
- 163. Bianco, C.L.; Schneider, B.L.; Bauer, M.; Sajadi, A.; Brice, A.; Iwatsubo, T.; Aebischer, P. Lentiviral vector delivery of parkin prevents dopaminergic degeneration in an alpha-synuclein rat model of Parkinson's disease. *Proc. Natl. Acad. Sci. USA* 2004, 101, 17510–17515. [CrossRef] [PubMed]
- 164. Yasuda, T.; Miyachi, S.; Kitagawa, R.; Wada, K.; Nihira, T.; Ren, Y.R.; Hirai, Y.; Ageyama, N.; Terao, K.; Shimada, T.; et al. Neuronal specificity of α-synuclein toxicity and effect of Parkin co-expression in primates. *Neuroscience* 2007, 144, 743–753. [CrossRef] [PubMed]
- 165. Yamada, M.; Mizuno, Y.; Mochizuki, H. Parkin gene therapy for alpha-synucleinopathy: A rat model of Parkinson's disease. *Hum. Gene Ther.* **2005**, *16*, 262–270. [CrossRef] [PubMed]
- 166. Guzman, J.N.; Sanchez-Padilla, J.; Wokosin, D.; Kondapalli, J.; Ilijic, E.; Schumacker, P.T.; Surmeier, D.J. Oxidant stress evoked by pacemaking in dopaminergic neurons is attenuated by DJ-1. *Nature* **2010**, *468*, 696–700. [CrossRef]
- 167. Taipa, R.; Pereira, C.; Reis, I.; Alonso, I.; Bastos-Lima, A.; Melo-Pires, M.; Magalhaes, M. DJ-1 linked parkinsonism (PARK7) is associated with Lewy body pathology. *Brain* 2016, 139, 1680–1687. [CrossRef]
- 168. Zondler, L.; Miller-Fleming, L.; Repici, M.; Goncalves, S.; Tenreiro, S.; Rosado-Ramos, R.; Betzer, C.; Straatman, K.R.; Jensen, P.H.; Giorgini, F.; et al. DJ-1 interactions with alpha-synuclein attenuate aggregation and cellular toxicity in models of Parkinson's disease. *Cell Death Dis.* 2014, 5, e1350. [CrossRef]
- 169. Xu, C.Y.; Kang, W.Y.; Chen, Y.M.; Jiang, T.F.; Zhang, J.; Zhang, L.N.; Ding, J.Q.; Liu, J.; Chen, S.D. DJ-1 Inhibits alpha-Synuclein Aggregation by Regulating Chaperone-Mediated Autophagy. *Front. Aging Neurosci.* **2017**, *9*, 308. [CrossRef]