

Supplemental Figure 1. Role of *DWARF* in cold tolerance in tomato.

(A-C) The relative electrolyte leakage (REL, A), the maximum quantum efficiency of photosystem II (Fv/Fm, B) and the relative expression level of COR47-like (C) in dwf mutant, wild type (WT) and DWARF -overexpressing transgenic plants (DWF: OE) with or without cold treatment. For cold treatment, plants were exposed to 8°C for 24 hours and subsequently exposed to 4°C for another six days. COR47-like was detected from leaf samples collected at 8 h after a cold at 8°C. Data are the means of three biological replicates ( $\pm$ SD) (A, C) or eight replicates (B). Different letters indicate significant differences according to Tukey's test at 0.05% level.



B

Α



## Supplemental Figure 2. Silencing efficiency and the cold tolerance of *CBF1* and *CBF1/2/3* gene silenced plants in response to EBR treatment.

(A) The relative expression levels of *CBF1* and three *CBFs* in *CBF1* silenced (pTRV-*CBF1*) and *CBF1/2/3*-cosilenced (pTRV-*CBF1/2/3*) plants respectively. Relative gene expression was calculated using the control (pTRV) plants as 1. (B) The phenotypes of pTRV-*CBF1* and pTRV-*CBF1/2/3* plants with or without cold treatment. For cold treatment, plants were exposed to 8°C for 24 hours and subsequently exposed to 4°C for another six days. 24 hours before cold treatment, the plants were pre-treated with 200 nM 24-epibrassinolide (EBR) or distilled water as the control.



Supplemental Figure 3. The relative expression of *BZR1* from tomato leaves exposed to  $25^{\circ}$  or  $8^{\circ}$  for different periods of time.

Data are the means of three biological replicates  $(\pm SD)$  shown by vertical error bars.





(A-C) The phenotypes (A), relative electrolyte leakage (REL, B) and maximum photochemical efficiency of PSII (Fv/Fm, C) in bzr1 mutant, wild type (WT) and 35S:BZR1-3HA-overexpression plants (BZR1:OE) with and without cold treatment. For cold treatment, plants were exposed to 8°C for 24 hours and subsequently exposed to 4°C for another six days. 24 hours before cold treatment, the plants were foliar applied with 200 nM 24-epibrassinolide (EBR) or distilled water as the control. Data are the means of three biological replicates ( $\pm$ SD) (B) or eight replicates (C). Different letters indicate significant differences according to Tukey's test at 0.05% level.



# Supplemental Figure 5. Yeast-one hybrid analysis of BZR1 binding to the promoters of *CBF1* and *CBF3* in tomato.

The 475 - and 388 bp fragments from *CBF1* and *CBF3* promoter sequences indicated in (Figure 2D) were cloned into pAbAi vector respectively. Interaction was determined on SD medium lacking leucine in the presence of 100 ng/ml Aureobasidin A ( $-Leu+AbA^{100}$ ). AD-empty and pAbAi-*CBFs* were used as negative controls.



**Supplemental Figure 6. Silencing efficiency of** *RBOH1* in *BZR1***:OE** plants. Relative gene expression was calculated using the pTRV plants as 1.



#### Supplemental Figure 7. Role of *RBOH1* in BR-induced cold tolerance.

(A-C) The phenotypes (A) relative electrolyte leakage (REL, B) and maximum photochemical efficiency of PSII (Fv/Fm, C) in the WT and *RBOH1*-RNAi plants with or without cold treatment. For cold treatment, plants were exposed to 8°C for 24 hours and subsequently exposed to 4°C for another six days. 24 hours before cold treatment, the plants were foliar applied with 200 nM 24-epibrassinolide (EBR) or distilled water as the control. Data are the means of three biological replicates (±SD) (B) or eight replicates (C). Different letters indicate significant differences according to Tukey's test at 0.05% level.

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### Supplemental Figure 8. The GSH and GSSG contents in WT and *RBOH1*-RNAi plants under cold condition.

(A and B) The GSH (A) and GSSG contents (B) in WT and *RBOH1*-RNAi plants exposed to 8°C for 12 hours. 24 hours before cold treatment, the plants were foliar applied with distilled water, 5 mM 6-aminonicotinamide (6-AN), 200 nM 24-epibrassinolide (EBR) or 5 mM 6-AN followed with 200 nM EBR, respectively. Data are the means of three biological replicates ( $\pm$ SD) shown by vertical error bars. Different letters indicate significant differences according to Tukey's test at 0.05% level.



#### Supplemental Figure 9. *RBOH1* plays a role in BR-induced cold tolerance by redox regulation.

(A-C) The phenotypes (A), relative electrolyte leakage (REL, B) and maximum photochemical efficiency of PSII (Fv/Fm, C) in wild type (WT) and *RBOH1*-RNAi plants with and without cold treatment. For cold treatment, plants were exposed to 8°C for 24 hours and subsequently exposed to 4°C for another six days. 24 hours before cold treatment, the plants were foliar applied with distilled water, 5 mM 6-aminonicotinamide (6-AN), 200 nM 24-epibrassinolide (EBR) or 5 mM 6-AN followed with 200 nM EBR, respectively. Data are the means of three biological replicates (±SD) (B) or eight replicates (C). Different letters indicate significant differences according to Tukey's test at 0.05% level.



### Supplemental Figure 10. H<sub>2</sub>O<sub>2</sub> and GSH treatment rescue the cold tolerance of *RBOH1*-silenced plants.

(A-C) The phenotypes (A), relative electrolyte leakage (REL, B) and maximum photochemical efficiency of PSII (Fv/Fm, C) in wild type (WT) and *RBOH1*-RNAi plants with and without cold treatment. For cold treatment, plants were exposed to 8°C for one day and subsequently exposed to 4°C for another six days. 12 hours Before cold treatment, Plants were foliar applied with distilled water, 5 mM H<sub>2</sub>O<sub>2</sub> and 5mM GSH respectively. Data are the means of three biological replicates (±SD) (B) or eight replicates (C). Different letters indicate significant differences according to Tukey's test at 0.05% level.

Supplemental Table 1. Primers used for VIGS constructs

Vector	Forward primer	Reverse Primer
pTRV2-CBF1	5'-CCGgaattcTATGCTACCTCCACCT-3'	5'-GCtctagaAACCCAACAAGTTTCT-3'
pTRV2-CBF1/2/3	5'-CGCggatccAGGGGAATCAGGAAGAGGAAT-3'	5'-GCtctagaGAAGATTTCGACGGCCTGAG-3'
pTRV2-RBOH1	5'-CGCgagctcCGTTCAGCTCTCATTACCATGG-3'	5'-CCGctcgagCCGAAGATAGATGTGTGTGTACCG-3'

Supplemental Table 2. Primers used for qRT-PCR analysis

gene	Accession number	Forward primer	<b>Reverse Primer</b>
CBF1	Solyc03g026280	5'-GTGACTTCGTGGATGAGGAG-3'	5'-AGGCATCAGTTTCCACACAA-3'
CBF2	Solyc03g124110	5'-TTCGATCGGAAGAAGTTTCA-3'	5'-CAAGTAATCCTGGCATGGAA-3'
CBF3	Solyc03g026270	5'-TGCCGGGTTTACTTACGAAT-3'	5'-TCAGCTTCCACATGATCTCC-3'
RBOH1	Solyc08g081690	5'-TCCAGCACAAGATTACCG-3'	5'-CCTCCATTGCGACGAT-3'
BZR1	Solyc04g079980	5'-TAGCCCGATTCCATCTTACC-3'	5'-TAATGGTGGTAGCGACAAGG-3'
COR47-like	Solyc04g082200	5'-TCTAGTAGCTCCAGTGATG-3'	5'-TCTCCTCTGTTTCCTCGT-3'
ACTIN	Solyc11g005330	5'-TGTCCCTATTTACGAGGGTTATGC-3'	5'-CAGTTAAATCACGACCAGCAAGAT-3'
UBI3	Solyc01g056940	5'-GCCGACTACAACATCCAGAAGG-3'	5'-TGCAACACAGCGAGCTTAACC-3'

Supplemental Table 3. Multiple reaction monitoring conditions used for LC-MS/MS analysis

Compound	Precursor (m/z)	Ion product (m/z)	Cone (V)	Collision Energy (eV)
brassinolide	708.43	160.96	70	50
[26- <sup>2</sup> H <sub>3</sub> ]-brassinolide	711.43	160.96	70	50
castasterone	692.5	160.96	75	45
[26- <sup>2</sup> H <sub>3</sub> ]-castasterone	695.5	160.96	75	45
28-norcastasterone	678.46	160.96	50	45
[28- <sup>2</sup> H <sub>3</sub> ]-norcastasterone	681.46	160.96	50	45

Supplemental Table 4. Primers used for	pAbAi-baits and AD-prey	constructs
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Vector	Forward primer	Reverse Primer
pAbAi-CBF1	5'-GGggtaccTACACATGTTTCTCAATTTTACA-3'	5'-CCctcgagTTGAAAAGATAGTGGAAGGT-3'
pAbAi-CBF3	5'-GGggtaccGAGATTTTACGTGTCGTTCTGTTGA-3'	5'-CCctcgagGGCCTGATCAATTGGTTAGGATG-3'
pGADT7-BZR1	5'-CCcatatgATGTGGGAAGGTGGAGGGTTG-3'	5'-CGCcccgggTCACATCCGAGCAGTCCCAC-3'

Supplemental Table 5. Triners used for Chill-qr Cix analysis			
DNA fragments	Forward primer	Reverse Primer	
CBF1	5'-GTTTCTCAATTTTACACGTG-3'	5'-GATATGCTTGGAATTGG-3'	
CBF2	5'-TAGAAAGTTTGCCACAT-3'	5'- CGGTATTACACGGAGTT-3'	
CBF3	5'-GTTAGACGCACGGAAGAT-3'	5'-GAACACGGAGTTAGAGGG-3'	
P1	5'-ATCCTGACTCCAACACGACT-3'	5'-GGTCACAAACTTAGCTTGAACG-3'	
P2	5'-CTTTTGTTTTGCTATTGGTA-3'	5'-CGTAAAGAAAACCATAAATC-3'	
P3	5'-CATCTGTTTCATTCTATACGAGTC-3'	5'-AATGGTGAGGAAGTGAGGGT-3'	

Supplemental Table 5. Primers used for ChIP-qPCR analysis

Supplemental Table 6. BR contents in tomato leaves under control and cold conditions
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Treatment	Brassinolide (ng/g FW)	castasterone (ng/g FW)	28-norcastasterone (ng/g FW)
25°C	$0.087 \pm 0.008$	$0.257 \pm 0.011$	$0.816 \pm 0.033$
8°C	$0.191 \pm 0.018$	$0.325 \pm 0.017$	$0.900 \pm 0.041$