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Diffuse Correlation Spectroscopy at Short Source-Detector Separations: Simulations, Experiments and Theoretical Modeling

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Abstract: Diffuse correlation spectroscopy (DCS) has widely been used as a non-invasive optical technique to measure tissue perfusion in vivo. DCS measurements are quantified to yield information about moving scatterers using photon diffusion theory and are therefore obtained at long source-detector separations (SDS). However, short SDS DCS could be used for measuring perfusion in small animal models or endoscopically in clinical studies. Here, we investigate the errors in analytically retrieved flow coefficients from simulated and experimental data acquired at short SDS. Monte Carlo (MC) simulations of photon correlation transport was programmed to simulate DCS measurements and used to (a) examine the accuracy and validity of theoretical analyses, and (b) model experimental measurements made on phantoms at short SDS. Experiments consisted of measurements from a series of optical phantoms containing an embedded flow channel. Both the fluid flow rate and depth of the flow channel from the liquid surface were varied. Inputs to MC simulations required to model experiments were obtained from corrected theoretical analyses. Results show that the widely used theoretical DCS model is robust for quantifying relative changes in flow. We also show that retrieved flow coefficients at short SDS can be scaled to retrieve absolute values via MC simulations.

Keywords: turbid media; dynamic light scattering; radiative transport; tissue phantoms

1. Introduction

Diffuse correlation spectroscopy (DCS) is a well-established optical technique capable of sensing blood flow in biological tissue using multiply scattered light and has widely been applied to quantify tissue perfusion in vivo [1–13]. The light source used in DCS is a long coherence length laser, which is delivered to a medium of interest through a large-core optical fiber while the back-scattered (reflected) intensity is collected using a small-core (single mode) fiber, placed at a fixed distance (the source-detector separation SDS) from the center of the source fiber. The detected signal is transmitted to a photon counter that in turn is electronically coupled to a hardware correlator board [14,15]. Experimental DCS measurements yield the normalized intensity autocorrelation function, $g_2(\tau)$ measured for a given fiber geometry, which in turn is quantified by fitting via an analytical expression to extract flow-dependent coefficients that characterize the dynamically scattered photon transport [15–17].

Theoretically, solutions to the unnormalized field correlation $G_1(\tau)$ are found by considering it to be governed by the modified photon diffusion equation

$$\left[D\nabla^2 - \mu_a - \frac{1}{3}\mu_s'k_0^2\langle\Delta r^2(\tau)\rangle \right] G_1(r, \tau) = S(r) \quad (1)$$

As discussed previously [15–17], D in Equation (1) is the photon diffusion coefficient of the medium (determined from its absorption and reduced scattering coefficients μ_a, μ'_s as $D = [3(\mu_a + \mu'_s)]^{-1}$). $\langle \Delta r^2(\tau) \rangle$ is a correlation-time (τ) dependent term and represents the ensemble-averaged, mean-squared displacement of moving scatterers in time τ . k_0 is the wave-vector associated with the laser source and $S(r)$ describes the spatial geometry of the source to be modeled [15]. Two models are commonly used for $\langle \Delta r^2(\tau) \rangle$: the Brownian flow model (where $\langle \Delta r^2(\tau) \rangle = 6D_B\tau$) with Brownian flow coefficient D_B ; the random flow model (where $\langle \Delta r^2(\tau) \rangle = V^2\tau^2$) where V is the average volumetric flow-speed of the scattering particles [5,15,16,18–20].

In a semi-infinite turbid medium, for a collimated pencil-beam input, closed form analytical solutions to $G_1(\tau)$ in cylindrical geometry is given by

$$G_1(\tau) = \frac{1}{4\pi D} \left[\frac{\exp(-K(\tau) \cdot r_1)}{r_1} - \frac{\exp(-K(\tau) \cdot r_2)}{r_2} \right] \quad (2)$$

Here, $K(\tau)$ is dependent on the medium's absorption, scattering and correlation-transport properties as $K(\tau) = 3\mu_a\mu'_s + \mu_s'^2 k_0^2 \Delta r^2(\tau)$. r_1 and r_2 are given as $r_1^2 = \rho^2 + z_0^2$ and $r_2^2 = \rho^2 + (z_0 + z_b)^2$, where ρ is the SDS (i.e., the distance from the detection point on the surface of the semi-infinite medium and the collimated input source). z_0 and z_b are dependent on the medium's optical properties (via the boundary conditions specified) as $z_0 = 1/\mu'_s$ and $z_b = 2AD$ and A is obtained from the relative refractive index between the medium and detector, as discussed previously [17,19,21]. To match theoretical predictions to experimental measurements the unnormalized field correlation $G_1(\tau)$ is transformed into the normalized intensity correlation via the Siegert relation, which gives that $g_2(\tau) = 1 + \beta[g_1(\tau)]^2$, where $g_1(\tau) = G_1(\tau)/G_1(0)$, and β is a scalar constant that depends on detection geometry [15,16].

Equation (2) is derived from the formalism of photon diffusion theory and as such would be expected to carry constraints on its validity (dictating geometry of measurements and the choice of laser wavelengths) [22–26]. To satisfy these constraints, experimental measurements of DCS typically use long SDS (>1 cm) and near-infrared laser sources. However, there could be clinical opportunities for using endoscopically compatible DCS probes to assess tissue perfusion in vivo, which will need to use short SDS for signal acquisition. Further, recent reports have employed DCS measurements at short SDS (<1 cm) for quantifying tissue perfusion in small animal models [27–31]. Surprisingly, these studies all reported that the classical diffusion-theory based theoretical expression in Equation (2) still provided good parametrization of relative changes in tissue perfusion even at short SDS used. Here, we further investigate the utility of Equation (2) to parametrize absolute and relative changes in flow in turbid media measured experimentally data in controlled flow-phantoms at short SDS. Additionally, we also investigate the use of Equation (2) to parametrize numerically simulated data at short SDS.

Monte Carlo (MC) based modeling of photon migration provide direct, numerical solutions to the radiative transport equation and have been extensively used in the field of tissue spectroscopy [32]. They are well developed and have been used to quantify experimental measurements of diffuse reflectance obtained in phantoms, animal models and human studies [33–37]. MC simulations can be programmed to incorporate heterogeneous tissue architectures and can be used to accurately model diffuse reflectance and correlation even at short SDS [32,38]. Thus, MC based photon transport methods have been used to compute normalized field correlation in semi-infinite media, simulate intensity correlation decays in complex layered scattering media and to validate improved theoretical models [31,38–42].

Here, we first examine the impact of using Equation (2) to quantitatively retrieve $\langle \Delta r^2(\tau) \rangle$ from $g_1(\tau)$ that were simulated using MC models of semi-infinite media spanning a range of absorption, scattering and Brownian flow diffusion coefficient D_B at varying SDS. Next, we acquired DCS measurements at a fixed short SDS of 0.15 cm, experimentally, in a set of flow-phantoms that included an embedded flow-channel (containing the same liquid as the phantom). The phantoms were designed to investigate how two physical parameters—the depth of the flow-channel from the optical probe and the flow type and rate of fluid in the channel—influenced DCS measurements obtained on the surface.

In Section 2, we describe the experimental methods used including the instrumentation, the design of liquid phantoms and the construction of the flow-channel. We then outline the MC modeling approach to simulate photon correlation transport and the tissue models simulated for analysis, discuss the use of Equation (2) to parametrize simulated and experimental data and provide a description of the goodness-of-fit metrics used. Section 3 presents the results of the analysis of the MC simulations of semi-infinite models, followed by results from theoretical analysis of the experimental phantom, and then show the comparison between MC models developed to simulate experimental measurements on flow phantoms.

2. Materials and Methods

2.1. Experiments

2.1.1. Instrumentation

Experimental DCS measurements were obtained using a home-built system that employed a stabilized diode laser centered at 785 nm and coherence length of nearly 3 m (I0785SH0100B-TH-L; Innovative Photonic Solutions, NJ, USA) as the source. The laser was coupled into one end of an SMA terminated fiber (diameter 200 μm , NA 0.22) of a fiber optical probe. The sampling end of the probe consisted of a detector fiber (diameter 8.2 μm) and was custom-fabricated (Gulf Photonics, Oldsmar, FL, USA) with SDS of 0.15 cm. The distal end of the detector fiber was optically coupled to a single photon counting module (ID100; ID Quantique, Geneva, Switzerland), which in turn was electronically connected to a digital correlation board (LS Instruments, Fribourg, Switzerland). The correlator was controlled by a computer and recorded DCS scans as normalized intensity autocorrelation functions, $g_2(\tau)$.

2.1.2. Liquid Phantoms

Liquid phantom media with required optical properties were prepared by mixing pre-determined volumes of an absorbing solution (with negligible scattering) and scattering solution (with negligible absorption), and diluting the resulting solution with deionized (DI) water, using protocols described previously [33,43]. A suspension of 1.0 μm diameter polystyrene microspheres (Polybead microspheres, 07310, Polysciences Inc., Warrington, PA, USA) was used as the scatterer and its scattering coefficient was calculated from Mie theory [44]. The absorber solution was prepared by dissolving bovine hemoglobin (H3760; MilliporeSigma, MO, USA) in DI water, whose absorption coefficient was experimentally measured using a transmission spectrophotometer (Cary 100 UV-Vis, Agilent Technologies, Wilmington, DE, USA). The volume-fractions of absorber and scatter mixed together determined the absorption and scattering coefficient of the final solution [33]. The liquid media used for phantoms were prepared with absorption coefficient of $\mu_a = 0.075 \text{ cm}^{-1}$ and reduced scattering coefficient $\mu'_s = 12 \text{ cm}^{-1}$. These values for absorption and scattering at 785 nm (the laser wavelength) are within ranges previously reported for different tissue types including bone, abdomen and white matter of the brain [45–47].

2.1.3. Flow-Chamber Description

Figure 1a shows a photograph of the phantom used and its schematic illustration in Figure 1b. A plastic chamber with dimensions of 5.5 cm \times 5.5 cm \times 1.0 cm was filled with the liquid phantom described above (having $\mu_a = 0.075 \text{ cm}^{-1}$; $\mu'_s = 12 \text{ cm}^{-1}$ at 785 nm) and the chamber was placed atop a homogenous solid phantom block with similar optical properties as the liquid phantom at 785 nm. The chamber housed a clear plastic tube (inner diameter = 1.6mm, outer diameter = 3.2 mm; 8000–0004; Thermo Fisher Scientific Inc., Miami, OK, USA) that was attached to two opposite sides of the chamber using moldable glue (Sugru; FormFormForm Ltd., London, UK) and ran along the bottom surface of the plastic chamber, parallel to one edge. One end of the plastic tube was connected to a syringe pump (75900–00; Cole Parmer, Vernon Hills, IL, USA) that was controlled via software to produce a

given volume flow rate F , while the other end of this tube drained into a sink. Assuming laminar flow, the pump flow rate was converted to an average fluid flow speed in the channel as $F = V_{ch}A$, where A is the cross-sectional area of the channel and V_{ch} the flow speed. Since the same liquid phantom added to the chamber was also loaded into the syringe pump, both the actively pumped and surrounding liquid media had identical optical transport coefficients. F was set via software to one of four values: F_0 (pump turned off; 0 mL/h), F_1 (1.44 mL/h), F_2 (3.60 mL/h) or F_3 (5.76 mL/h). Given a circular tube of radius 0.08 cm, the average channel flow speeds were calculated to be 0.02 cm/s, 0.05 cm/s and 0.08 cm/s at F_1 , F_2 , and F_3 , respectively. These flow rates gave speeds in ranges reported for moving red blood cells in capillaries [48].

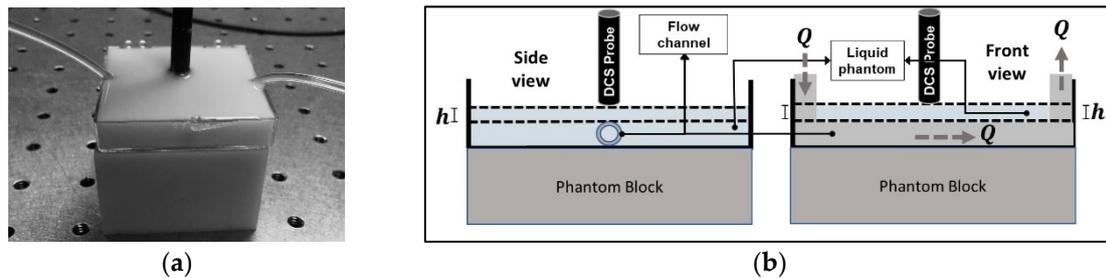


Figure 1. Photograph of the phantom flow chamber (a) and schematic of the flow chamber's construction and measurement system (b). The gray arrows in (b) show the direction of the fluid flow. The flow rate F was controlled using a motorized syringe pump (see text). h is the distance from of the liquid surface to the top of the flow channel, which was controlled by addition of specific volumes of the liquid phantom to the chamber. The chamber (edges shown as thick black lines in (b)) was placed atop a homogeneous tissue phantom block with optical properties matched to the liquid phantom at the laser wavelength. The DCS probe was positioned directly above the flow channel and at center of the flow chamber.

2.1.4. Phantom Preparation and Measurement Protocol

Two primary parameters were varied in the flow phantoms as shown in Figure 1b—the volume flow rate F through the flow channel and the depth D of the flow channel from the fluid surface. The flow rate F was cycled through for four rates (F_0 – F_3), for four cycles. The depth of the flow-channel from the liquid surface was held fixed (from D_0 to D_3) for each flow speed cycle.

The plastic flow chamber had equal length and breadth ($L = 5.5$ cm), thus addition of a given volume v of liquid to the chamber would raise the height of the fluid surface by $\Delta h = v/L^2$. For the first phantom to be measured, the chamber was filled with prepared liquid media such that liquid surface was visually assessed to have just submerged the flow channel. The depth of the flow channel from this first phantom was assumed to be D_0 . DCS measurements were acquired in this phantom by cycling through the four different flow rates (F_0 – F_3), giving four measurements (labeled D_0F_0 – D_0F_3).

Next, 1.0 mL of phantom was added to the container, which was estimated to increase the depth of the channel from the surface by approximately $\Delta h = 0.03$ cm, as done previously [49]. The depth of the flow channel from fluid surface was $D_1 = D_0 + \Delta h$. DCS measurements were acquired for a full cycle of pump volume flow rates (F_0 – F_3) and gave phantoms D_1F_0 – D_1F_3 . The process was repeated twice—each time, 1 mL of liquid was added to the chamber (increasing depth of channel by $2\Delta h$ and $3\Delta h$). Thus, DCS measurements were obtained for 16 phantoms, corresponding to each depth and flow-rate used and were sequentially measured in sets of 4, starting from D_0F_0 – D_0F_3 and ending in D_3F_0 – D_3F_3 . It is worth noting that this experimental protocol implied that in all but the first set of four measurements, the pump had been active for at least one cycle of operation before additional media was added to the chamber (i.e., D_1F_0 was measured chronologically after D_0F_3).

2.1.5. Data Acquisition

For each phantom, prepared phantom liquid media (with viscosity nearly that of water) was added to the container, stirred gently and allowed to come to rest (3–5 min). DCS measurements were obtained by moving the optical fiber probe vertically toward the liquid surface. The probe was situated at the center of the phantom container and oriented such that the line connecting the center of the source and detector fiber was perpendicular to the flow channel and was positioned using custom probe holder (mounted on a linear optical stage that allowed the probe head to be moved away or toward the liquid surface). Once the liquid phantom was prepared, the probe was moved till it just broke the surface tension of the liquid medium. Five scans were recorded for each phantom with changing flow condition. Each DCS scan was acquired for 1s and had count rates ranging between 80–100 kHz. The probe was then removed, new media added, and the process repeated for depths D_1 – D_3 .

2.2. Monte Carlo Simulations of Correlation Transport

Light propagation in tissue-like media can be numerically simulated by construction of stochastic trajectories of individual photon packets, where the variables governing the evolution of these trajectories are determined by an input tissue model’s optical properties and sampled using MC techniques [32,50]. In these simulations, several (10^6 – 10^8) individual photon packets are launched and aggregate quantities of interest (such as the photon flux, fluence, absorbance and fluorescence) are tracked spatio-temporally, for a given tissue model. For the studies here, we modified a time-resolved MC photon transport model (previously described in detail [49,51]) to include simulate of photon field correlations in reflection geometry. We note that MC modelling of photon transport in scattering media are well discussed elsewhere [50,51], and we therefore confine description of aspects pertinent only to tracking of field correlations in the MC model.

Correlation transport was integrated into the MC model by adding a new variable (programmed into the simulation) to track the field correlation carried by each photon packet, as it was transported from the source to the detector, following the methodology described before [15,39,40]. To track the decay of the normalized photon-field correlation $g_1(\tau)$, it was assumed that moving particles were uniformly distributed in each tissue layer modeled, moved uncorrelated to each other and that all photon scattering events were from dynamically moving particles. Under those conditions, $g_1(\tau)$ was expressed as

$$g_1(\tau) = \exp\left[-\frac{1}{6}q^2\langle\Delta r^2(\tau)\rangle\right] \tag{3}$$

In Equation (3), $\langle\Delta r^2(\tau)\rangle$ represents the mean-squared displacement term of particles contributing to dynamic scattering in time τ , while q is the magnitude of momentum transfer due to a scattering event. Each scattering event undergone by a photon packet tracked in the MC simulation provides specific q associated with it. Thus, in the geometry of the MC model, if \vec{k}_1 and \vec{k}_2 represented directions associated the incoming and scattered photon packets, then $q^2 = 2k^2(1 - \cos \theta)$, where θ is the single-scattering angle (i.e., the angle between \vec{k}_1 and \vec{k}_2) sampled by the MC model, for the photon packet being tracked [15,40]. Since $|\vec{k}_1| = |\vec{k}_2| = k = 2\pi/\lambda$, and λ (the wavelength of the laser) is known, Equation (3) was used to update this variable, through the flight of the photon packet simulated using

$$g_1^{N+1}(\tau) \rightarrow g_1^N(\tau) \cdot \exp\left[-\frac{1}{6}q^2 \cdot \Delta r_i^2(\tau)\right] \tag{4}$$

In Equation (4) the superscripts N and $N + 1$ refer to the field correlation variable (for a given photon packet within the MC simulation) before the N^{th} and $(N + 1)^{th}$ scattering event, respectively with q^2 being computed at the N^{th} scattering event. $\Delta r_i^2(\tau)$ was the mean-squared displacement of the scattering particles in layer i (where the scattering event occurred) and was set either to $6D_B\tau$ for Brownian flow, or to $V^2\tau^2$ for random flow [15]. Thus, inputs to the MC simulations (for each layer) required flow coefficients D_B (or V^2) together with the layer optical properties. $\Delta r_i^2(\tau)$ in Equation (4)

was computed (depending on inputs either as Brownian or as Random flow) dynamically, through the path of each photon packet tracked.

Each photon packet launched had initial value of $g_1^1(\tau) = 1$. If a photon packet exiting the medium was detected by the simulation, the photon packet's field correlation variable was multiplied with the exiting photon weight (to account for all path-length dependent absorption losses) and accumulated into a global field correlation variable, $g_1(\tau, \rho)$, where ρ indexed the spatial detector location for the exiting photon. At the end of the simulation, $g_1(\tau, \rho)$ was normalized such that $g_1(0, \rho) = 1$ (i.e., $g_1(\tau, \rho)$ was divided by the number of photons detected, for each detection-bin). The MC model used here yielded surface reflectance $g_1(\tau, \rho)$ for photons exiting into annular, concentric rings on the surface of the medium (ρ identifies the distance of the collection annulus from relative to a source at the origin).

2.3. Tissue Models Simulated

2.3.1. Semi-Infinite Models

The first set of simulations consisted of 210 runs modeling different semi-infinite media with each run producing $g_1(\tau)$ for 20 SDS (uniformly spaced between 0.1–2 cm). The optical properties of the media simulated obtained by permuting through five scattering coefficients (μ'_s : 3 – 15 cm⁻¹), seven absorption coefficients (μ_a : 0.05 – 0.5 cm⁻¹) and six Brownian flow coefficients (D_B : 5×10^{-9} – 3×10^{-8} cm²/s) following the ranges as reported recently [31]. These MC simulations were compared against theoretical calculations to assess both the predictions of theory (forward model), as well as parametrization of the simulations (inverse model), as described in Section 2.4.

2.3.2. Three-Layer Models

The second set of MC simulations modeled experimental measurements. Each measurement from a phantom (for given depth and flow speed) was modeled as a 3-layered slab (following the schematic in Figure 1b). Layer 1 was liquid covering the flow chamber, whose layer thickness represented depth of channel from surface. Layer 2 was the flow channel having fixed thickness of 0.15 cm (to match the inner diameter of the tubing used) and Layer 3 representing the solid phantom block (thickness of 5 cm).

We note that this modeling architecture does not account for the plastic tube-wall (present experimentally). Modeling the tube wall as an additional layer with (with low values of absorption, scattering and Brownian correlation-decay coefficient relative to the liquid layer) did not change simulated outputs relative to an equivalent 3-layer model and thus we chose to represent flow phantoms as a 3-layer model. Since Layer 1 and Layer 2 contained the same liquid medium, and since the optical coefficients for Layer 3 (solid block) was similar to that of the liquid phantom, we modeled all layers as having identical transport coefficients and refractive indices, matching those of the liquid phantom used.

Table 1 shows the required inputs (with values for experimentally known parameters) needed to simulate photon correlation from the three-layer phantom models. In Layers 1 and 3 the correlation flow coefficients were modeled as diffusive with a given Brownian diffusion coefficient; for Layer 2 it was modeled as diffusive only when the pump was inactive (phantom F_0). The flow in Layer 2 was modeled as random flow, when the fluid was actively pumped. The Brownian flow-coefficient inputs for layers 1 and 3 were kept fixed for all flow speeds (for fixed depth). Input values of all Brownian flow coefficients needed were obtained using theoretical analyses (described in Section 3.3.1).

Increasing depth of the flow channel were modeled via MC simulations by increasing the thickness of Layer 1 (from $z_1 = D_0$) in increments of 0.03 cm, to match experimentally expected values. The thickness of layer 1 at depth D_0 needed to be empirically determined (described in Section 3.3.2). Changes in channel flow rates were modeled by varying flow coefficients used for Layer 2 such that the relative change in flow F_2/F_1 and F_3/F_1 in the MC models matched experiments. The flow speed in

Layer 2 at flow rate F_1 (1.44 mL/h) was needed and was also empirically determined (described in Section 3.3.2). MC predictions of $g_2(\tau)$ simulated for experimental phantoms were obtained for the experimental SDS of 0.15 cm.

Table 1. Optical coefficients used for simulations of three-layered tissue models.

Layer #	μ_a (1/cm)	μ_s (1/cm)	g	Refractive Index, n	z_1 (cm)	$\langle \Delta r^2(\tau) \rangle$
Layer 1					h	$6D_B\tau$
Layer 2	0.075	120	0.9	1.35	0.15	$6D_B\tau$ or $V^2\tau^2$
Layer 3					5	$6D_B\tau$

2.4. Theoretical Analyses

Both the simulated and measured data were presented as normalized intensity field correlation for theoretical analyses. Since measurements provide normalized intensity correlations, the theoretical unnormalized field correlation from Equation (2) was transformed into the intensity correlation via the Siegert relation ($g_2(\tau) = 1 + \beta|g_1(\tau)|^2$) as mentioned before [15,16] and thus experimental data were fitted using Equation (2) via a non-linear least-squares optimizing algorithm (lsqcurvefit in MATLAB) to retrieve coefficients describing $\langle \Delta r^2(\tau) \rangle$ and β .

Two functional forms used previously to describe $\langle \Delta r^2(\tau) \rangle$ were used—the Brownian flow model or the random-flow model [16,17]. When scatterers are assumed as moving diffusively in the medium $\langle \Delta r^2(\tau) \rangle = 6D_B\tau$. Scatterers moving under actively pumped flow are modeled as random flow with $\langle \Delta r^2(\tau) \rangle = V^2\tau^2$. These two flow models can also be combined together as a shear-flow model where the mean squared displacement contains both terms, $\langle \Delta r^2(\tau) \rangle = D_B\tau + V^2\tau^2$ [52]. Experimental data were analyzed using three flow models: for Brownian flow model D_B was obtained, for the random flow the speed of the particles V was obtained, and for the shear-flow model both coefficients D_B and V were obtained. All other variables needed to evaluate Equation (2)—i.e., the optical properties of the medium, SDS, refractive index of medium and detector—were set matched to experimentally used values.

Equation (2) was used in two ways to analyze MC simulated data. The first (as forward model) was to employ it to calculate $G_1(\tau)$ from inputs of a given simulation, to match it directly against to MC simulations (at any needed SDS). The second way was as the inverse model as used for the analysis of experiments. However, to keep this procedure identical to processing of experimental data, simulated $g_1(\tau)$ data were first translated into $g_2(\tau)$ (using a fixed value of $\beta = 0.48$) before least-squares fitting. As in the experimental analysis, the fit coefficients describing $\Delta r^2(\tau)$ and β were obtained. All other variables needed to evaluate Equation (2)—i.e., the optical properties of the medium, SDS, refractive index of medium and detector—were matched to simulated inputs.

2.5. Goodness-of-Fits: Fit Residuals

The results of fitting the measured, simulated and calculated correlation functions across the different experimental phantoms and MC models were quantified by using a non-negative scalar value χ^2 , as a measure of the goodness-of-fit. This was calculated as $\chi^2 = \sum_{m=1}^M [g_2^{meas}(\tau_m) - g_2^{fit}(\tau_m)]^2$, where g_2^{meas} was the intensity correlation (measured or simulated) and g_2^{fit} was the fitted intensity correlation (using Equation (2) or simulated). A perfect fit would have $\chi^2 = 0$, and thus the lower the χ^2 the better the goodness-of-fit. The index m corresponds to discrete correlation-time bins (as output by the hardware correlator using a multi-tau data accumulation scheme). For analysis here, there were a total of $M = 178$ logarithmically spaced bins (i.e., the correlation times were evenly spaced on a log scale), matching experimental data with $\tau_1 = 1 \mu\text{s}$ and $\tau_M = 1 \text{s}$.

3. Results

3.1. Simulations in Semi-Infinite Phantoms

3.1.1. Forward Theoretical Calculations vs. Simulations

Figure 2 presents direct comparisons of MC simulated $g_2(\tau)$ (symbols) and forward calculations using Equation (2) (lines). Each plot shows data from five different tissue models (squares: $\mu'_s = 3/\text{cm}$; triangles: $\mu'_s = 6/\text{cm}$; circles: $\mu'_s = 9/\text{cm}$; asterisks: $\mu'_s = 12/\text{cm}$; diamonds: $\mu'_s = 15/\text{cm}$). Figure 2a,b show data for media with absorption $\mu_a = 0.05/\text{cm}$, at SDS of 0.15 and 1.5 cm respectively. All simulated and theoretical models shown in Figure 2 had fixed Brownian flow coefficient of $D_B = 2 \times 10^{-8} \text{ cm}^2/\text{s}$. Data in Figure 2c,d) are for media with higher absorption $\mu_a = 0.42/\text{cm}$, at same SDS as Figure 2a,b respectively. The χ^2 fit-residuals are shown within legends for each simulated model in Figure 2a and d and are in accordance with known limitations of diffusion theory—i.e., data simulated for long SDS were fit better Figure 2c,d, relative to data from shorter SDS Figure 2a,c and simulated media with lower albedo (where the albedo $a = \mu'_s / (\mu'_s + \mu_a)$) were fit worse relative, to media with higher albedo.

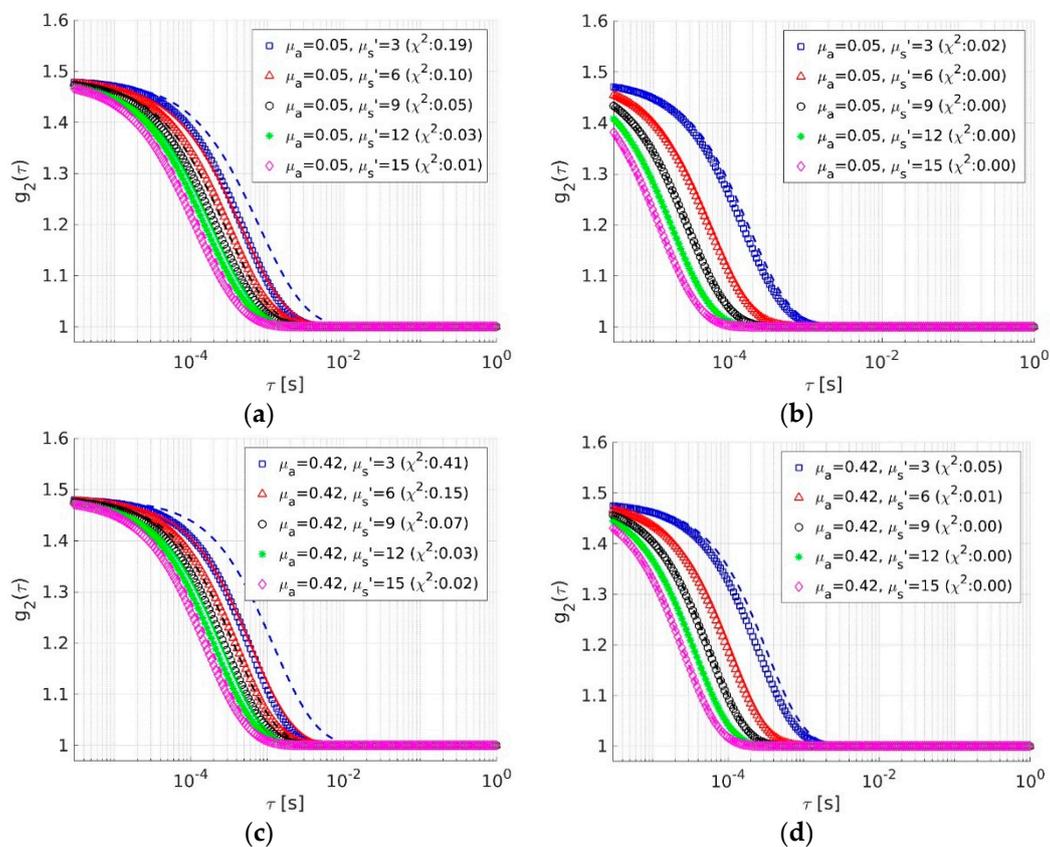


Figure 2. MC simulations (symbols) and forward theoretical calculations using Equation (2) (lines) for media with low and high absorption (top and bottom rows) at SDS of 0.15 and 1.5 cm (left and right columns). Each figure shows data for 5 media with varying scattering distributed between $\mu'_s = 3/\text{cm}$ (squares) to $\mu'_s = 15/\text{cm}$ (diamonds).

3.1.2. Fitting Simulations Using Theory

Direct calculations of $g_2(\tau)$ from diffusion theory deviated at short SDS relative to MC predictions, but Equation (2) could be instead used to freely fit simulated data for D_B the Brownian diffusion flow coefficient as described in Section 2.4. Symbols in Figure 3a–d show the same simulated data as in Figure 2a–d, respectively, while lines show fitted diffusion theory calculations. It is clear that the theoretical data fit simulations near perfectly, even at short SDS of 0.15 cm Figure 3a,c and for low

albedo media. Although the numerical fits in Figure 3 matched numerical simulations much better than direct forward calculations Figure 2, these numerical fits are obtained at the cost of errors in retrieved flow coefficients.

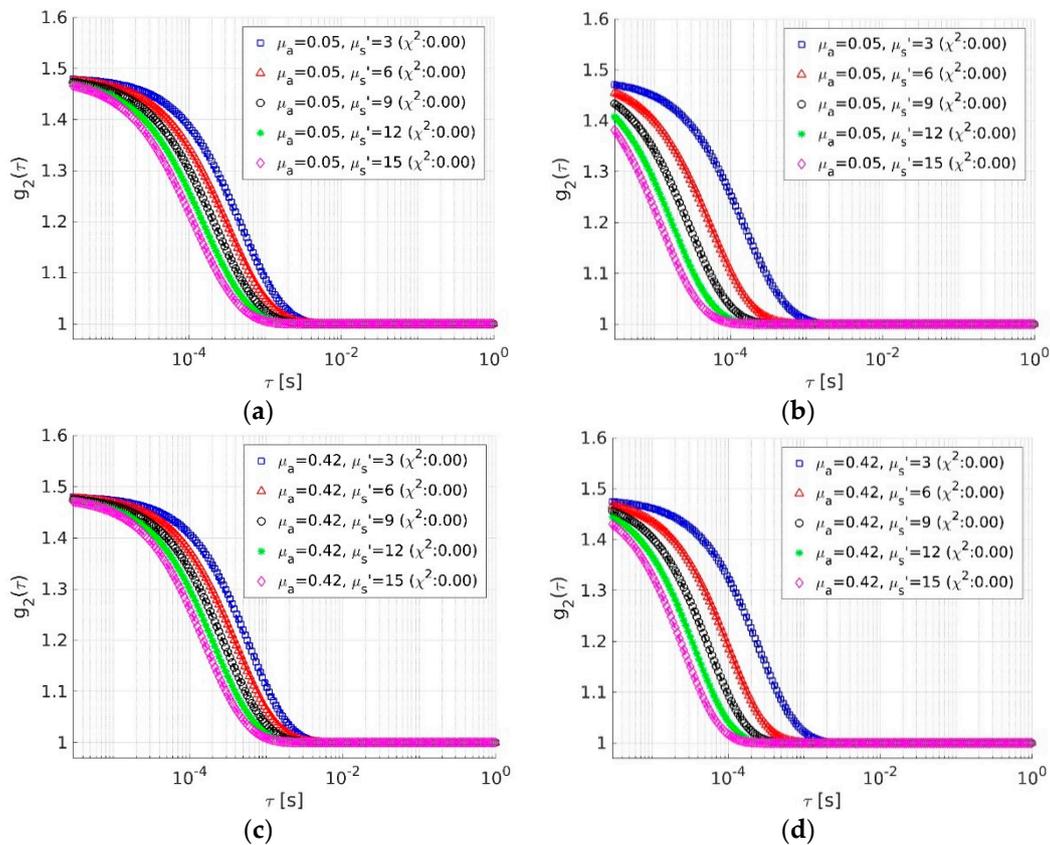


Figure 3. MC simulations (symbols) with fitted curves using Equation (2) (lines) via optimizing for the Brownian flow coefficient D_B (inverse model) to minimize χ^2 residuals. Simulated data shown in (a,d) are identical (in the same order) as Figure 2a,d. Data in (a,c) are for SDS of 0.15 cm (at low and high absorption coefficients respectively), while data in (b,d) are for SDS of 1.5 cm.

3.1.3. Errors from Theoretical Fits

In Figure 4, retrieved values of the absolute Brownian flow coefficients (from each fit) are shown as contour maps from 35 simulations (spanning 7 absorption and 5 scattering coefficients) with fixed Brownian flow coefficient $D_B = 2 \times 10^{-8} \text{ cm}^2/\text{s}$, for three different SDS (Figure 4a: SDS = 0.15 cm; Figure 4b: SDS = 0.45 cm; Figure 4c: SDS = 0.75 cm). With decreasing SDS, the retrieved values of D_B increased from its expected (known) value of $D_B = 2 \times 10^{-8} \text{ cm}^2/\text{s}$. Figure 4d–f show the same data as Figure 4a–c, but as percent-errors relative to the true input value (of $D_B = 2 \times 10^{-8} \text{ cm}^2/\text{s}$). As seen before, errors associated with retrieved flow coefficients were highest for small SDS and low albedos. These data also indicate that extracted D_B depended on the optical properties of the media, and the SDS used.

Figure 5 shows the averaged percent-error for extracted D_B with varying SDS, for the six D_B coefficients simulated (squares: $5 \times 10^{-9} \text{ cm}^2/\text{s}$; up-triangles: $1 \times 10^{-8} \text{ cm}^2/\text{s}$; circles: $1.5 \times 10^{-8} \text{ cm}^2/\text{s}$; asterisk: $2 \times 10^{-8} \text{ cm}^2/\text{s}$; diamonds: $2.5 \times 10^{-8} \text{ cm}^2/\text{s}$; down-triangles: $3 \times 10^{-8} \text{ cm}^2/\text{s}$). For each line (i.e., at fixed D_B), the percent errors at each SDS shown in Figure 5 was the mean across all 35 simulated models (spanning seven μ_a and five μ_s' values with given D_B), while error-bars are standard-deviations. Abscissa of each line (for each D_B) are shown staggered in Figure 5 to show six distinct markers, since the computed errors (and distributions) were nearly identical across all D_B values. In other words, the percent-error in retrieved D_B was independent of the flow coefficient used and only depended on

the optical properties of the medium and the SDS used. These data are in agreement with findings published in a recent report that investigated short separation DCS with MC simulations [31].

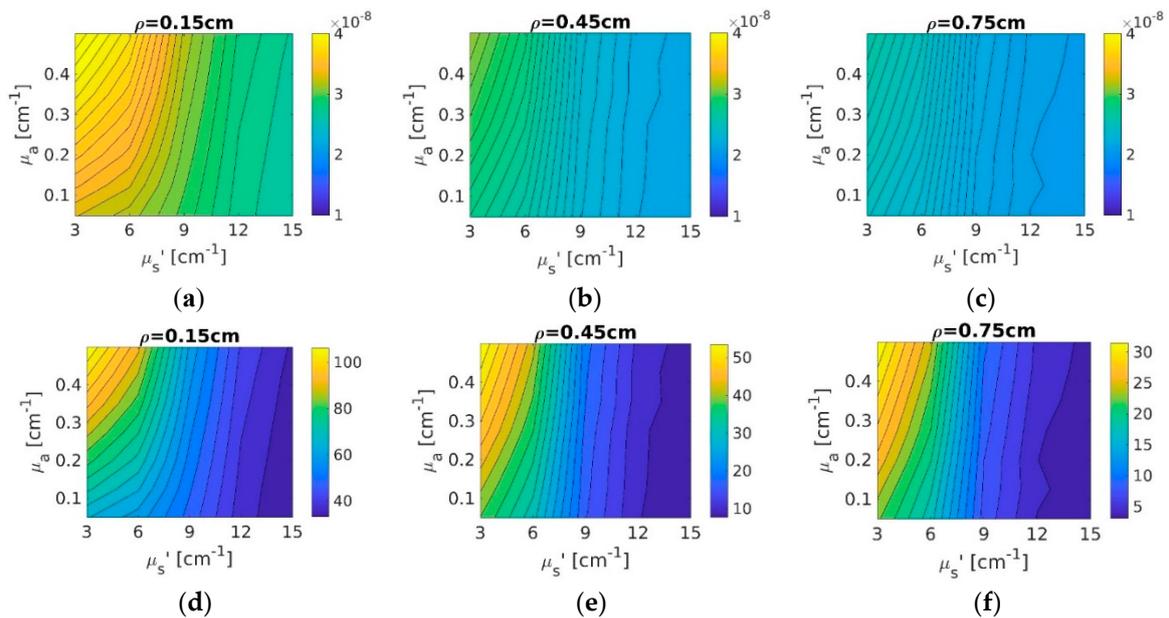


Figure 4. Retrieved flow coefficients obtained via using Equation (2) to fit simulations (top-row) and corresponding percent-errors (bottom-row) for 35 different MC simulations spanning seven μ_a and five μ_s' values and fixed $D_B = 2 \times 10^{-8}$ cm²/s. Data are shown as contour maps for 3 different SDS (left column: SDS = 0.15 cm; middle column: SDS = 0.45 cm; right column: SDS = 0.75 cm). Note that the color-bar scale is fixed for the top row but changes across SDS for the bottom row.

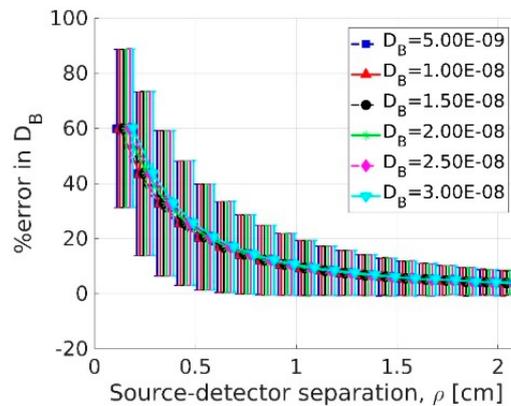


Figure 5. Percent-errors in retrieved Brownian flow coefficient D_B with varying SDS from analysis of simulations. Lines show data for 6 simulated D_B coefficients (identified in the legend). Symbols represent mean percent errors calculated across all available simulations for given SDS and D_B , while error bars shows standard deviations. Data for each point (in each line) was computed across 35 simulations run (see text) with varying absorption and scattering. The lines were identical to each other and are shown staggered only for visual clarity.

3.1.4. Relative Changes in Flow Coefficients: Simulations vs. Theory

Even though there were errors in absolute values of retrieved D_B coefficients when simulations at high albedo and/or short SDS were fit with Equation (2), relative changes in derived flow coefficients could be compared in media with given (fixed) optical scattering and absorption but changing D_B . Figure 6 shows the mean relative change in extracted D_B coefficient, at three different SDS—where the relative change in D_B at each SDS, was calculated as the ratio of the retrieved D_B coefficient for a given simulation (i.e., for given medium optical coefficients), by the D_B retrieved from a matched simulation

but with $D_B = 5 \times 10^{-9} \text{ cm}^2/\text{s}$. The mean value of the relative changes in D_B thus calculated across each of the 35 simulations (spanning all 7 μ_a and 5 μ'_s coefficients) are shown by three bars, each for different SDS in Figure 6. The red asterisks indicate error bars computed as the standard deviation across each of these 35 simulations and were nearly zero, indicating that predicted relative changes in D_B values from theoretical analyses, were near exactly as modeled by the simulations, across all SDS.

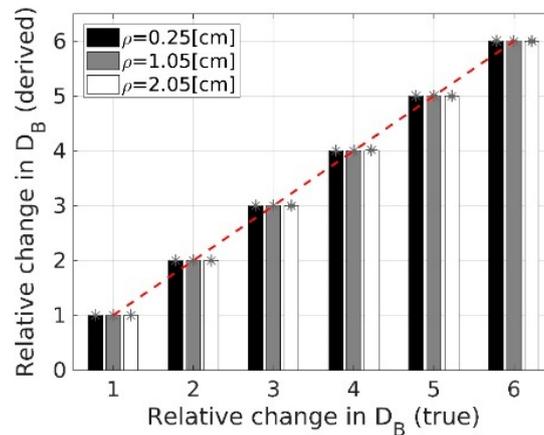


Figure 6. Relative change in extracted flow coefficients (relative to values obtained from media simulated with $D_B = 5 \times 10^{-9} \text{ cm}^2/\text{s}$) at three different SDS. The true relative change in D_B (x -axis) was obtained from input values to simulations, while the derived values (y -axis) were obtained from fitted coefficients (see text).

3.1.5. Scaling Factors: Linearly Correcting Retrieved D_B Coefficients

Given relative changes in D_B could be computed across optically identical media but with different flow coefficients, we derived correction factors (for each medium with given optical coefficients and required SDS). These correction factors were calculated as the ratio of the simulated (input) Brownian flow coefficients simulated, to the flow coefficient retrieved via fitting the simulation using Equation (2). Thus, retrieved D_B coefficients, for any medium, could be appropriately scaled and corrected to estimate an absolute flow coefficient. Figure 7 shows the correction factor maps obtained (across the 35 simulated media) at two different input D_B coefficients (Figure 7a,c: $D_B = 1 \times 10^{-8} \text{ cm}^2/\text{s}$; Figure 7b,d: $D_B = 3 \times 10^{-8} \text{ cm}^2/\text{s}$) for two different SDS (Figure 7a,b: 0.25 cm; Figure 7c,d: 1.05 cm). The similarity of these derived correction factors in Figure 7a–d indicate that they were nearly identical for either flow coefficient. Since a correction factor of unity would indicate that retrieved flow coefficient was the same as that modeled, the correction factors in Figure 7a,b (short SDS) are correspondingly lower, than those for Figure 7c,d (longer SDS).

Figure 8a shows the average absolute values of retrieved D_B coefficients, across the 6 different sets of simulations (with each set having fixed D_B ranging from $5 \times 10^{-9} \text{ cm}^2/\text{s}$ through $3 \times 10^{-8} \text{ cm}^2/\text{s}$ in intervals of $5 \times 10^{-9} \text{ cm}^2/\text{s}$), for four SDS (squares: 0.15 cm; triangles: 0.55 cm; circles: 1.05 cm; asterisks: 1.95 cm). The error bars show the standard deviation in retrieved D_B values across the 35 simulated media with varying optical properties (the dashed line is $y = x$). Figure 8b shows the average absolute value of the corrected D_B coefficients, for the same data shown in Figure 8a where each retrieved flow coefficient was corrected by multiplication with correction factors derived. Correction factors for each tissue model and SDS were obtained from the set simulations with input $D_B = 3 \times 10^{-8} \text{ cm}^2/\text{s}$. As was seen previously in Figure 6, corrected flow coefficients yielded values that were near exactly identical to the true (input) value for D_B , across all D_B coefficients.

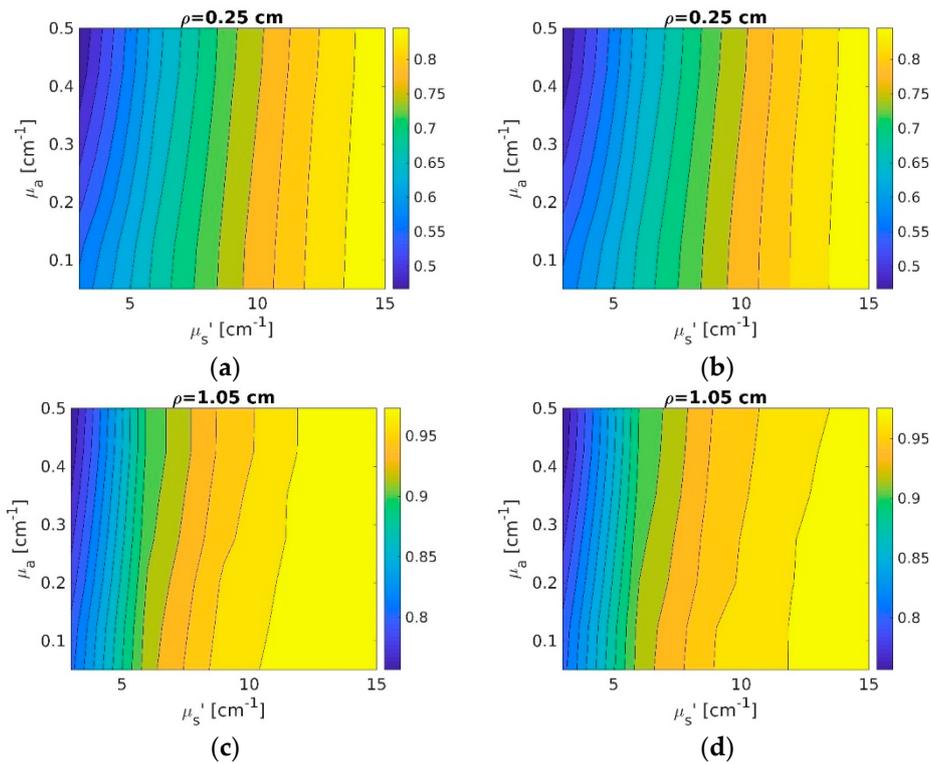


Figure 7. Scaling (correction) factors determined using simulations at two SDS (top: 0.25 cm; bottom: 1.05 cm) shown for two different media with two different flow coefficients (left: $D_B = 1 \times 10^{-8} \text{ cm}^2/\text{s}$; right: $D_B = 3 \times 10^{-8} \text{ cm}^2/\text{s}$). Each panel shows data for 35 MC simulations spanning seven μ_a and five μ_s' values (see text). Note that the color-bar scale changes for data in top and bottom rows.

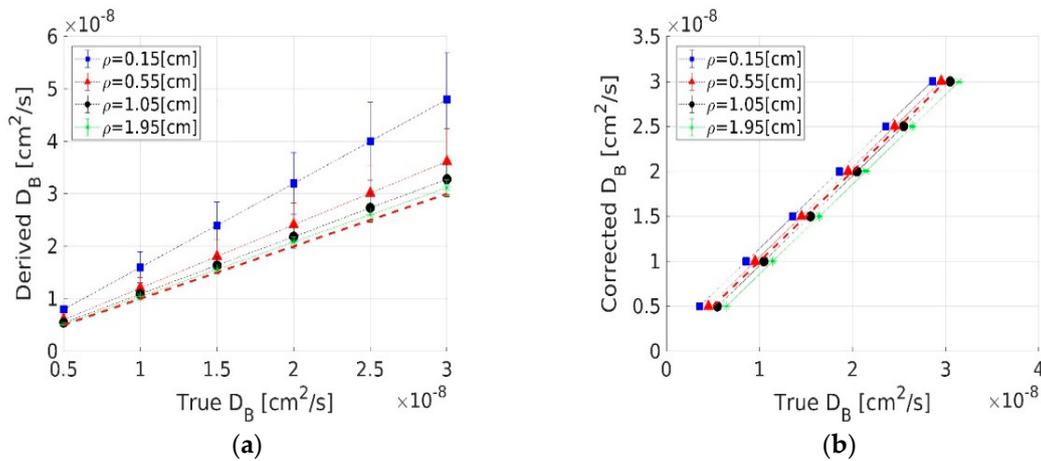


Figure 8. Estimated Brownian flow coefficients vs. simulated values. (a) shows the coefficients obtained directly by fitting simulated data, where each line shows data for different SDS. Symbols were average values obtained from 35 simulated media having fixed D_B (but changing optical properties); errorbars show standard deviations. (b) shows the same data in (a) after appropriate scaling by correction factors (see text). The errorbars in (b) are smaller than the size of symbols shown and different lines (for different SDS) are shown staggered only for clarity. The dashed line in Figure 8a,b shows $y = x$.

3.2. Diffusion-Theory Based Analysis of Experiments

3.2.1. Flow Models for Fitting Experimental Data

As described above, experimental phantoms included a flow channel with the fluid in the channel being either actively pumped (flow phantoms F_1 – F_3) or with pump turned off (flow phantom F_0) and

were designed to examine the impact of flow dynamics on DCS measurements. However, theoretical fits to these data could be obtained by using the Brownian flow model ($\langle \Delta r^2(\tau) \rangle = 6D_B\tau$), the random flow model ($\langle \Delta r^2(\tau) \rangle = V^2\tau^2$) or the shear-flow model ($\langle \Delta r^2(\tau) \rangle = D_B\tau + V^2\tau^2$).

Figure 9a,b show theoretical fits (lines) to an experimental acquisition of $g_2(\tau)$ (symbols) when the pump was switched off (phantom D_0F_0 Figure 9a) and when pump flow rate was $F_3 = 5.76$ mL/hr (phantom D_0F_3 , Figure 9b). Theoretical fits for the three different flow models used are shown by lines (dash-dotted: Brownian flow; solid: random flow; dashed: shear flow). As seen visually (and by χ^2 residuals in figure legends), the fits to data are poor for the random flow model in Figure 9a and for the Brownian flow model in Figure 9b, while they are excellent for the random flow model in Figure 9b and for the Brownian flow model in Figure 9a. Given that experimentally measured $g_2(\tau)$ had markedly different decay profiles (by design) in these two phantoms, the results are as anticipated. The shear-flow fit was however able to fit both experimental flow profiles with ($\chi^2 < 0.01$) and all experimental data presented below were analyzed using the shear-flow model. This fitting process yielded flow coefficients D_B and V , corresponding to the diffusive and the bulk flow components, respectively, for each phantom.

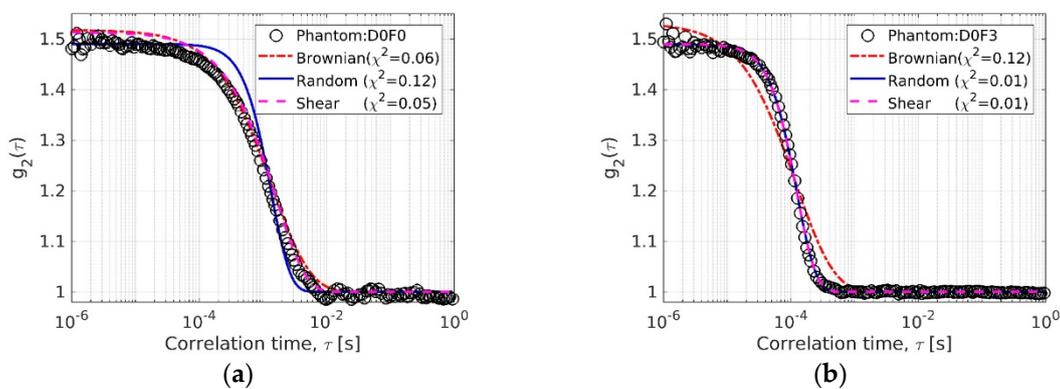


Figure 9. Experimental data (symbols) and fits using Equation (2) for three different flow models (dash-dotted: Brownian; solid: Random; dashed: shear) for two different flow phantoms. (a) shows representative data in phantom D_0F_0 (pump flow turned off) while (b) shows data from phantom D_0F_3 (highest pump flow).

3.2.2. Absolute vs. Relative Flow-Coefficients in Phantoms

Figure 10a shows extracted D_B coefficients in phantoms vs. the experimentally controlled pump flow rate. Each symbol-line shows data at fixed channel depth (squares: depth D_0 ; triangles: depth D_1 ; circles: depth D_2 ; asterisks: depth D_3). The derived Brownian flow coefficients D_B decreased with increasing flow rates, when the flow channel depth was shallowest, while it increased with increasing flow, as the height of the liquid surface above the flow channel was raised. Figure 10b shows the extracted speed of the moving particles V from phantoms vs. changing fluid flow speed in the channel. Each symbol-line shows data for fixed channel depth (as in Figure 10a). The pump-flow rate, F shown in Figure 10a was converted to a fluid flow speed in the channel (as described in Section 2.1.3) using $F = v_{ch}A$, where A was area of a tube with diameter equal to the inner diameter of the flow channel (0.16 cm). The derived speed of moving particles on the other hand increased with increasing fluid flow speed for the shallow channel depths (squares and triangles in Figure 10b). However, the derived flow speed did not increase linearly with pump flow rate in phantoms with deeper channels (circles and asterisks in Figure 10b).

Since the fluid flow in the channel was well-controlled experimentally, we examined relative changes in flow coefficients in the phantoms with actively pumped flow (i.e., across F_1 – F_3). Relative flow rates in experiments were calculated as ratio of the pump flow rates of F_2/F_1 and F_3/F_1 corresponding to relative flow rates of 2.5 and 4, respectively. Relative changes in phantoms (at each channel depth) were calculated by dividing derived flow coefficients by the flow coefficients obtained for phantoms

at pump flow F_1 . These data are shown in Figure 10c (for the Brownian flow coefficient ratios) and in Figure 10d (for the shear-flow speed coefficient ratios). Figure 10c,d only show three flow speeds (at each depth) as it does not include data from phantoms when the pump was inactive.

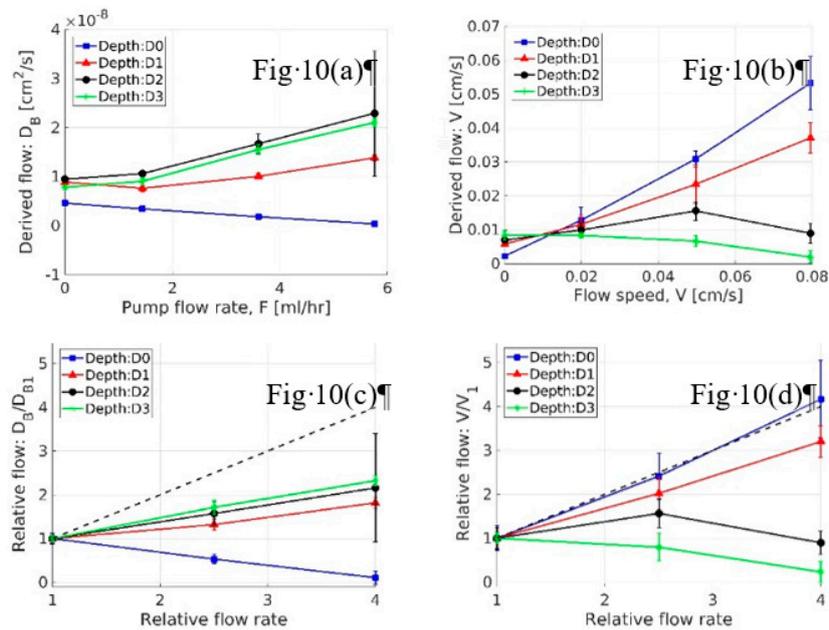


Figure 10. Derived flow coefficients obtained from fitting phantom data using Equation (2). (a,b) show the retrieved Brownian diffusion D_B and flow speed V , for each phantom. Each line shows fixed channel depth (shown in legend), symbols show mean values and error bars are standard deviations (across the five DCS scans obtained for each phantom). (c,d) show relative changes flow vs. relative change in input flow rates, normalized relative to flow phantoms F_1 . Dashed line in (c,d) show $y = x$.

3.3. Simulating Experimental Data Using MC

DCS measurements made on the experimental phantoms were modeled using MC simulations and required input coefficients to describe the phantom model (as described in Section 2.3). These modeling results are presented as two separate cases below, corresponding to phantoms with active flow (pump turned on) and those with inactive flow (pump turned off). In both these cases, the optical transport coefficients and refractive indices for all layers in the tissue model were set to optical coefficients and refractive index of the liquid phantom used, the thickness of layer 2 was matched to the flow channel diameter, and thickness of layer 3 was set to 5 cm (as noted in Table 1). Four input coefficients needed to be determined for simulating each MC phantom tissue model: the height of the liquid layer h above the flow channel, and the three flow coefficients for each of the three phantom tissue model layers.

3.3.1. Modeling Phantoms with Inactive Flow

The correlation transport coefficients in each layer of the simulated phantoms when the pump was inactive (i.e., for flow F_0 phantoms), were treated as identical across layers and were each set to the same Brownian flow coefficient D_B . Since the fluid in the channel was identical to the liquid media above it, the three-layer model in this case was equivalent to a semi-infinite medium and used $h = 0$. The input value of D_B to the MC phantom tissue model was derived by correcting the Brownian flow coefficient obtained from theoretical fitting—i.e., the retrieved D_B from experimental fits was first multiplied by 0.83 (the correction factor) and then used as input to the MC model. This correction factor was obtained from previously run semi-infinite MC simulations (described in Section 3.1.5 and Figure 7), at the optical transport coefficients of the liquid phantom ($\mu_a = 0.075/\text{cm}$ and $\mu'_s = 12/\text{cm}$), at the experimentally employed SDS (0.15 cm).

Since variations were observed in D_B coefficients across phantoms $D_0F_0 - D_3F_0$, (as seen in Figure 10a), the input Brownian coefficients for MC phantom models simulating different channel depths were obtained by correcting D_B from experimentally fitted data from each phantom at F_0 , at each channel depth. Thus, the D_B coefficient obtained from the four phantoms $D_0F_0 - D_3F_0$ were each multiplied by 0.83 to create input values to MC simulations (since phantoms had same optical properties and were measured at fixed SDS, they had the same correction factor). Figure 11a–d show representative $g_2(\tau)$ data measured in phantoms (symbols), the corresponding theoretically Brownian flow model fitted using Equation (2) (dashed line), and predicted using MC simulations (solid line), for phantoms $D_0F_0 - D_3F_0$, respectively. It is worth noting that the theoretical lines were obtained by fitting the measured data, while the MC data were directly computed (for given inputs). Table 2 lists the values of the input coefficients used in these simulations.

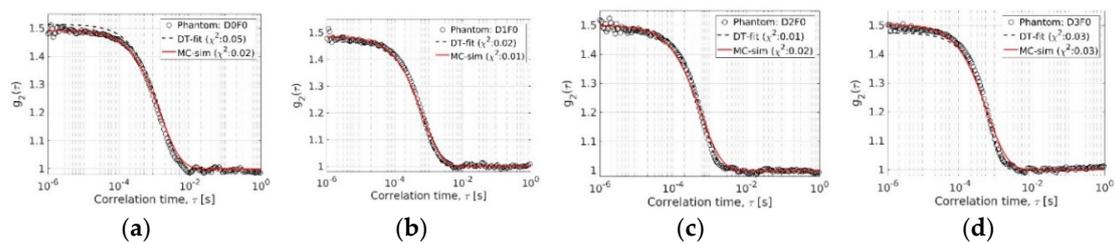


Figure 11. DCS measurements (symbols) for phantoms $D_0F_0 - D_3F_0$, (a–d, respectively) with theoretical fits using a Brownian-flow model (dashed line) and MC predictions (solid line). Each MC model used input D_B values obtained from scaling the theoretically fitted values (see text).

Table 2. Input coefficients for MC flow phantom models. Δh was constrained to be 0.03 cm experimentally while the relative change in fluid flow speed from flow rate of F_1 to F_2 was 2.5 (change from F_1 to F_3 was 4).

Phantom	Layer1 Thickness z_1 [cm]	D_B [cm ² /s] (Layers 1&3)	D_B [cm ² /s] (Layer 2)	V [cm ² /s] (Layer 2)
D0F0	$h_0 = 0.02$	3.6×10^{-9}	3.6×10^{-9}	-
D0F1			-	5.7×10^{-3}
D0F2			-	1.42×10^{-2}
D0F3			-	2.28×10^{-2}
D1F0	$h_1 = h_0 + \Delta h = 0.05$	7.9×10^{-9}	7.9×10^{-9}	-
D1F1			-	5.7×10^{-3}
D1F2			-	1.42×10^{-2}
D1F3			-	2.28×10^{-2}
D2F0	$h_2 = h_1 + \Delta h = 0.08$	8.9×10^{-9}	8.9×10^{-9}	-
D2F1			-	5.7×10^{-3}
D2F2			-	1.42×10^{-2}
D2F3			-	2.28×10^{-2}
D3F0	$h_3 = h_2 + \Delta h = 0.11$	9.3×10^{-9}	9.3×10^{-9}	-
D3F1			-	5.7×10^{-3}
D3F2			-	1.42×10^{-2}
D3F3			-	2.28×10^{-2}

3.3.2. Modeling Phantoms with Actively Pumped Flow

As described above, measurements from phantoms were obtained by changing both the depth of the flow channel and the fluid flow rate in the channel. There were thus 12 phantoms with actively pumped fluid flow—they are identified individually by a combining a flow rate ($F_1 - F_3$), with the channel depth ($D_0 - D_3$). In order to simulate measurements using MC models in these phantoms, inputs for the thickness of Layer 1 (height of the liquid layer h , above the flow channel) and flow

coefficients for the three layers were needed. For active pumping, correlation transport in Layer 2 (the flow channel) was modeled as random flow, while in Layers 1 and 3 (the liquid media above and below the flow channel, respectively) it was modeled as Brownian, with both layers having the same D_B . These D_B input values were obtained, for each depth, from corresponding MC models shown in Figure 11 (i.e., from phantoms with no active flow).

Input values for two remaining parameters—the depth of the flow channel from surface and the flow speed of the medium in the flow channel—had to be determined. Across 12 phantoms with active flow, relative changes (in both these parameters) were experimentally well controlled. Increases in channel depth (from one depth to the next) was experimentally estimated to approximately be $\Delta h = 0.03$ cm. Therefore, if the phantoms at depth D_0 had Layer 1 thickness of h_0 , then Layer 1 thickness for phantoms with flow channel at depth D_1 would be $h_1 = h_0 + \Delta h$ (and further as $h_2 = h_1 + \Delta h$ and $h_3 = h_2 + \Delta h$). Similarly, changes in flow speeds could be calculated from experimentally used flow rates—i.e., if the input flow speed V_1 was known for phantoms with flow F_1 , the flow speeds for flow F_2 and F_3 would scale as $V_2 = V_1 F_2 / F_1$ and $V_3 = V_1 F_3 / F_1$.

Input values for h_0 and V_1 were sought by running a set of simulations that varied h_0 (between 0–0.05 cm) and V_1 (between 0.04 and 0.001 cm/s) to find values that matched experimental measurements for phantom D_0F_1 with lowest χ^2 residuals. From these set of initial simulations, the tissue model with $h_0 = 0.02$ cm and $V_1 = 5.7 \times 10^{-3}$ cm/s, produced the lowest χ^2 residuals relative to experimental data acquired in phantom D0F1. Once h_0 and V_1 were known, required inputs for all 12 tissue models ($D_0F_1 - D_3F_1$, $D_0F_2 - D_3F_2$ and $D_0F_3 - D_3F_3$) were generated by scaling them across phantoms. The (scaled) input values of the coefficients (sought as inputs) for all the 3-layered tissue models constructed are shown in in Table 2.

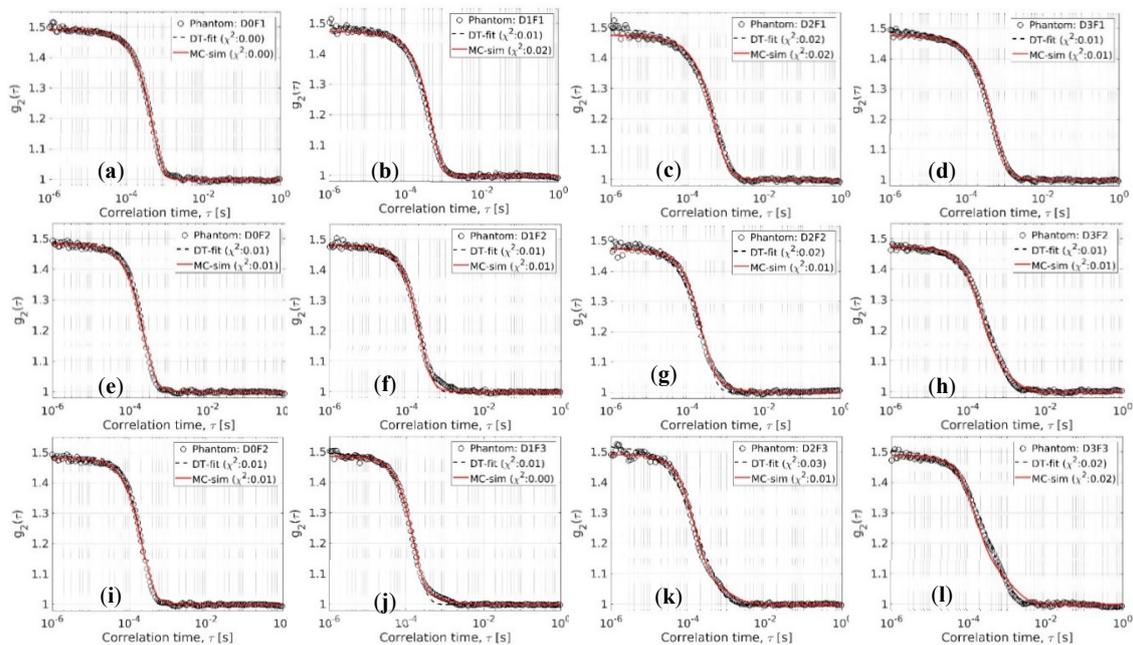


Figure 12. Comparisons of experimentally measured data (circles), diffusion theory-based fits (dashed line) and MC predictions (solid line) for the 12 phantoms with actively pumped. ((a–d) show these data when flow rate was $F_1 = 1.44$ mL/h for depths D_0 – D_3 . ((e–h) show the same sequence of depths at flow rate $F_2 = 3.6$ mL/h and (i–l) for flow rate $F_3 = 5.76$ mL/h).

Figure 12 shows comparisons of experimental measurements (symbols), diffusion theory fits with shear-flow model (dashed black lines) and the MC simulations (solid lines) for one representative measurement, for each of the 12 phantoms with active flow. Data in Figure 12 are organized such that moving across a row, changes channel depth (D_0 : Column 1; D_1 : Column 2; D_2 : Column 3; D_3 : Column 4) while moving down a column changes channel flow rate (F_1 : Row 1; F_2 : Row 2; F_3 : Row 3).

As seen in Figure 11, the data generated by the MC simulations matched experimental measurements as well as the fitted theoretical curves (χ^2 residuals are shown in each figure). Thus, the MC tissue model used could accurately represent the experimental phantom system used.

4. Discussion

We examine the quantitative impact of analyzing experimental DCS measurements made at short SDS using Equation (2), for different phantoms (with varying optical transport and flow properties) and use MC simulations to directly compare theoretical calculations across range of SDS. Simulations showed expected behavior, where long SDS DCS data well matched and short SDS data being poorly matched for direct, forward comparisons to calculations from Equation (2) Figure 2. However, DCS data obtained at short SDS were well fit using theory at the cost of errors in extracted flow coefficients Figures 3 and 4. Relative changes in perfusion could still be quantified exactly using theory, even for SDS < 0.1 cm Figures 5 and 6. as long as the optical properties of the media considered were not changing. MC simulations were shown to be accurate for calibrating and correcting theoretically retrieved coefficients, given the SDS and optical properties of the medium Figures 7 and 8.

Equation (2) could also directly be used to examine changes in flow in phantoms but required that flow dynamics include both Brownian and Random flow components Figure 9. As shown in Figure 10a,b, extracted both the diffusive flow and random flow coefficients extracted from theoretical analyses of experimental data (at SDS of 0.15 cm) increased with experimentally increasing flow rates. However, the Brownian flow component D_B changed proportionally with flow rate only when the flow channel was submerged deeper below the surface (estimated to be more than 500 μm here). While, for shallower channel depths the flow speed V linearly changed with experimentally controlled flow speed. Relative changes in extracted flow coefficients in phantom experiments when flow was actively pumped showed that the ratios of theoretically extracted flow speeds were almost exactly as expected experimentally, but only when the channel depth was shallowest Figure 10d.

These results taken together with relative flow changes extracted in MC simulations for semi-infinite media Figure 8, indicate that theoretical analysis of DCS measurements using Equation (2) is robust and can be used to quantitatively assess changes of relative flow within turbid media, even at short SDS as long as the optical properties and SDS do not change. These conclusions are in accordance with previous findings regarding the accuracy absolute flow rates vs. relative flow at longer SDS [13,16,53]. Our results support findings from recent studies, where Equation (2) was successfully used to examine relative changes in DCS flow coefficients from both simulations and in vivo studies in animal models, at short SDS [27,28,30,31]. However, our findings also indicate that quantification of relative perfusion changes reported by these studies would be estimated more accurately, if the relative changes in perfusion at the short SDS channels were computed with MC-derived correction factors with corresponding changes in media optical properties they measured.

We were also able to predict experimental measurements with MC simulations across all phantoms used Figures 11 and 12. These data showed excellent fits in all phantoms ($\chi^2 < 0.03$). Given that inputs to tissue models and experiments were tightly coupled and constrained, these data indicate good fidelity of the experimental phantom system used here. Although MC simulations for active flow phantoms matched changes in relative flow rates used experimentally, the absolute value of flow rates (listed in Table 2) were nearly 3.7 times lower than those expected—i.e., the average flow speed in the channel experimentally (given a tube of radius 0.08 cm, and flow rate $F_1 = 1.56 \text{ mL/h}$) would be $V_1 = 2.1 \times 10^{-2} \text{ cm/s}$ but the MC inputs for flow speed of in layer 2 was $V_1 = 5.7 \times 10^{-3} \text{ cm/s}$ (data in Figure 12b).

There could be a few different reasons for this issue: for e.g., our MC model assumed a flat velocity profile for correlation transport in the channel but a parabolic flow-profile may better physically represent our experimental system; the MC simulations also did not consider flow directions (relative to photon trajectories) in calculations of correlation transport in Equation (3); we also did not investigate simulations in semi-infinite media using random flow models to examine how they were impacted at

short SDS. Lastly, the phantom models though were carefully constructed and measured still includes uncertainty about the depth the channel from DCS probe may impact these results. These topics will be examined in future investigations.

Although Equation (2) is widely used to quantify DCS measurements mainly due to its simplicity and ease of use, there have been recent reports that have developed improved theoretical formulations to model DCS measurements directly using correlation transport modeled using the radiative transport theory [54,55]. Such approaches would be better suited for analyzing DCS measurements made at short SDS separations and could be investigated in more detail in the future using the MC simulations and experimental platform presented here.

In summary, we establish that the theoretical expression in Equation (2) used to model DCS measurements of dynamically back scattered reflectance from turbid media can be robust for fitting data obtained experimentally (and numerically) across a wide range of SDS. The parameters obtained from theoretically modeled fits of measurements can be used to quantitatively parametrize relative changes of fluid flow at short SDS only when the optical properties of the media are not changing, across changing flow conditions. Yet, derived flow coefficients using Equation (2) can be linearly corrected to provide absolute flow coefficients, if the optical properties and SDS are known, via MC generated calibration factors. Our findings show that such MC derived correction factors are applicable to DCS measurements that were made at with SDS as small as 0.15 cm.

Author Contributions: Both K.V. and S.Z. contributed to study conceptualization, methodology, investigation, data curation, writing the original draft, visualization. K.V. was responsible for software, writing—review and editing, formal analysis, supervision and validation of data.

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Conflicts of Interest: The authors declare no conflict of interest.

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