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Optimization of Lignin Extraction Variables by Response Surface Methodology from Pine Saw Dust, and Quantification of Major Structural Units in Isolated Lignin Fraction

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Abstract: A synergistic combination of dioxane, acetic acid, and HCl was investigated for lignin extraction from pine wood biomass. After initial screening of reagent combination, response surface methodology (RSM) was used to optimize the lignin yield with respect to the variables of time 24–72 h, solids loading 5–15%, and catalyst dose 5–15 mL. A quadratic model predicted 8.33% of the lignin yield, and it was further confirmed experimentally and through the analysis of variance (ANOVA). Lignin at optimum combination exhibited features in terms of derivatization followed by reductive cleavage (DFRC) with a value of (305 μ mol/gm), average molecular weights of 4358 and polydispersity of 1.65, and 2D heteronuclear single quantum coherence nuclear magnetic resonance spectrum (2D-HSQC NMR) analysis showing relative β -O-4 linkages (37.80%). From here it can be suggested that this fractionation can be one option for high quality lignin extraction from lignocellulosic biomass.

Keywords: pinewood; lignin extraction process; DFRC; 2D HSQC NMR

1. Introduction

Presently, the global community, with an aim to shape the future of bioeconomy, has put together rigorous efforts on the biorefining of biomass feedstocks [1]. These substrates, specifically the lignocelluloses due to their relative abundance and readily collectable nature, have become quite prominent candidates in this race [2]. However, their inclusive utilization still demands new technological processes to expand their boundaries for the production of sustainable products within the context of a biorefinery [3]. Among various lignocelluloses, pine wood biomass, a softwood, is an important feedstock with second highest contents (up to 30% on average) as lignin within their native biomass matrix [4].

This lignin, being an important macromolecule, is primarily composed of varying units of guaiacyl (G units in pine wood biomass), syringyl (S units), and p-hydroxyphenyl (H units) depending on the type of biomass species [5,6]. Lignin, with the presence of phenolic groups in its molecule, has become quite a versatile feedstock for the production of various sustainable materials [7]. These can be used for the production of rubbers, poly propylene, epoxy resins, and even carbon fibers [8–10]. Even more, this lignin can also be used as a partial substitute for the petroleum based resins minimizing thus the harmful impacts of toxic substances.

However, the lignin structure strongly varies with the adapted isolation process [11]. For example, lignin isolated via widely employed processes (such as Kraft, sulfite, organosolv, soda, etc.) differ widely not only among their molecular weights but also in their functional groups [11]. Although pulp and paper industry widely use the pulping processes (Kraft and sulfite) for lignin isolation, such kind of lignin offers less reactivity with



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). high impurities and even so with severely modified structures making them less likely to be used for wider industrial applications [12,13].

Literally, there have been some good strategies to isolate the proto lignin including the famous milled wood lignin (MWL) and enzymatic lignin processes [14,15]. Similarly, the organosolv processes, taking advantage of organic acids, alcohols, or even from their synergistic effects, have also been found effective to extract the prototype lignin ensuring the solvent recovery as an additional benefit [16]. Recently gamma valerolactone (GVL) has been investigated as a potential reagent for the dissolution and extraction of lignin from lingocelluloses [17,18]. This organic solvent is a self-sustained product within a lignocellulosic biorefinery and is capable enough to extract lignin at near neutral solvent conditions [19].

Typically, GVL/acid/water has been mostly applied for an enhanced delignification rate with an improved cellulose digestion [20]. Being miscible with water, it induces better hydrolytic effects for biomass to dissolve out lignin [21]. It has also been applied under synergetic effects with metal salts, microwave, and with acids [22,23].

Additionally, there is a fair deal of studies on the lignin isolation from biomass species using both the dioxane and acetic acid as it has been reported that the acidic conditions effectively break the most frequent arylalkyl ether bonds and lignin carbohydrate complex leading to the formation of low molecular weight lignin. However, increasing thermal and acidic severity can cause lignin condensation [24]. Acetsolv is another one such process in which xylan portion of the biomass is dissolved by breaking glycosidic bonds with the help of acetic acid and also this acid catalyzes the cleavage of β -O-4 linkages to yield soluble low weight aromatics. However, more severe conditions of hydrogen ions not only increase the hydrolysis but also degrade the secondary sugars [25]. Although acetic acid has been envisaged as one promising lignin isolation reagent equally effective for both softwood and hardwood biomass species, nonetheless, softwoods due to their relatively high portion of lignin contents demand for more harsh conditions [26]. Generally it has been applied as an acetic acid–water system catalyzed by HCl around 100 °C and 2 h of time [27]. However, under such conditions it has been previously found that the lignin, dissolved during such processes, can condense back during the cooling process [28].

Therefore, in this study, we tried several organosolv reagent combinations (GVL, dioxane, and acetic acid catalyzed by HCl) under mild thermal conditions and sorted one on the basis of its capacity to retain high quality of lignin extraction and further undertook its optimization using design expert software and employing Box Bhenken Design. After the optimization the lignin fraction was obtained and qualitatively analyzed in terms of its functional groups, molecular weights, and structural features and compared with the milled wood lignin.

2. Materials and Methods

2.1. Preparation of Biomass

Pine saw dust was collected from Gangwon province South Korea and was used as a model substrate in the experimentation. It was air dried, ground to 60 mesh size, and finally methanol extracted. The chemical composition of this saw dust was determined according to two step acid hydrolysis of National Renewable Energy laboratory's (NREL) standard analytical procedure and with wise method [29,30]. For the elemental analysis, an elemental analyzer (CHNS 932, LECO Corp., USA) was used and the contents of C, H, and N were measured, and O contents were calculated by subtracting C, H, and N contents from 100%. The heating value was determined by using a calorimeter Parr 6400 Calorimeter (Parr, USA). This saw dust had almost 5.4% moisture contents, 49.7% carbon, 5.3% hydrogen, 0.2% nitrogen, and 44.8% oxygen. Regarding its compositional analysis in terms of major units, it was 60.8% for holocellulose and 28% for lignin. There were also extractives as 8.7%, 0.4% ash, and 4% moisture. The higher heating value was determined to be 18.9 MJ/kg.

2.2. Fractionation of the Biomass Via Acid Catalyzed Process

Pine saw dust was subjected to lignin isolation experiments at various specified reagent combination along with HCl. For all the fractionation experiments, specially designed lab scale stainless steel cylindrical reactors, with a total volume of 500 mL, were used. A measured quantity of pine saw dust and dissolving reagents were loaded and the mixture was homogenized completely with glass stirrers. All the experiments were carried out at mild temperature of 70 °C and/or at room temperature [31,32].

2.3. Lignin Isolation

After being treated for the desired time, the biomass slurries were first subjected to filtration using a Buchner funnel attached with a vacuum assembly. The residue was shifted to 2 L of beakers and was diluted by adding deionized water to precipitate out the lignin. After this, it was subjected to centrifugation and finally freeze-dried and saved in the refrigerator for any further experiments.

2.4. Design of Experiments and Statistical Analysis

For the selection of the lignin isolation variables and their ranges, we first performed an initial screening with different reagent combinations (dioxane: acetic acid, GVL acetic: acid and GVL: dioxane all catalyzed by different HCl doses) under various temperatures and time intervals with 10% solids loading. These combinations then were performed and their lignin yield, G units' values, and molecular weights were analyzed accordingly. Keeping in view the lignin quality with highest G units, a combination of dioxane: acetic acid and HCl was chosen. For the selection of the ranges of lignin isolation variables, we selected the center point values as 10% solids loading, 48 h of time, and 10 mL of catalyst dose based on our preliminary trials and their ranges were defined around this center point. These three factors (time, solids loading, catalyst dose), specific to the lignin isolation, play a fundamental role [33]. Solids loading is important in a sense that high solids loading is recommended for the desired economic operation of the biobased refineries [34]. The catalyst dose and time is also crucial as they play an important role of the catalyst mass transport into the biomass cell walls [35]. Talking specifically for the case of the lignin isolation, room temperature is important as it somehow mimics the classical proto lignin isolation process i.e., milled wood lignin [36].

After selecting these variables and their ranges, to further sort out the optimum experimental conditions, response surface methodology (RSM) was used with lignin yield as response (Y). Three variables solids loading (X₁), catalyst dose (X₂), and time (X₃) were optimized and were tested in their ranges of 5–15% wt. for solids loading, 5–15 mL for HCl concentrations, and 24–72 h for time. Among from the experimental designs a Box-Behnken Design (BBD) with three center points was chosen to design the experiments. Data analysis was done with the help of Design Expert software (Version 8.0, State-Ease, Inc., USA). Lignin yield was calculated on the basis of initial weigh of lignin in pine saw dust.

2.5. Structural Characterization of Isolated Lignin Samples (GPC, DFRC, 2DHSQCNMR)

The recovered lignin fractions were analyzed by various qualitative analyses such as gel permeation chromatography (GPC), derivatization followed by reductive cleavage (DFRC), and 2D heteronuclear single quantum coherence nuclear magnetic resonance (HSQC NMR) spectrum by following the methods according to our previous studies [37]. Briefly, for GPC analysis, to determine the weight average (Mw) and number average (Mn) molecular weights of the lignin samples, the isolated lignin fraction was firstly put to acetylation by dissolving it in a 1:1 (v:v%) of pyridine and acetic anhydride solution. It was periodically shaken after every 30 min while maintaining a temperature of 105 °C for almost 2 h in the oven. Afterwards, it was washed and freeze dried and 20 mg of this acetylated lignin sample was dissolved in 2 mL of tetrahydrofuran (THF) followed by analyzing it on a GPC, Agilent 1200, USA (equipped with an ultraviolet detector (UV) on a PL-gel 5 mm Mixed-c. column. This was first calibrated with the standards of polystyrene

and THF was run as the mobile phase at a flow rate of 1 mL/min. After running all the samples, the GPC chromatograms were also drawn. To this end, after doing the standards calibration, the graphs were drawn between the peak intensity and the time intervals. As a typical rule of size exclusion, the higher molecular weight curve is detected earlier followed by the successive less molecular weight curves. Similarly, in our case, MWL peak was shown earlier on GPC chromatogram having 10,660 molecular weight followed by DA lignin with 4718 molecular weight. Lastly the ODA lignin peak was detected as having the lowest molecular weight (4358). For the DFRC analysis, 20 mg of lignin was dissolved in 5 mL of (AcOH + AcBr (4:1)) solution and kept at 50 °C for 1 h with periodic shaking. Afterwards it was dried and the residue was mixed with 2 mL of dioxane/acetic acid/water (5:4:1, v/v/v) solution followed by zinc dust (50 mg) addition. After 1 h, an internal standard tetracosane together with 20 mL of each of CH₂Cl₂ and saturated NH₄Cl were added and the pH of this whole mixture was set to around 2 with HCl and the phases were separated precisely. The organic phase was dried again on rotary evaporator and then finally subjected to acetylation and were quantified by GC-MS (Agilent 7890B USA). For 2D-HSQC-NMR, the lignin samples were prepared using 20 mg of lignin and dissolving it in 0.75 mL DMSO-D6 and then the analysis data was acquired with a Bruker AVANCE 600 spectrometer (Bruker, Germany).

3. Results and Discussion

3.1. Initial Experiments to Define Lignin Isolation Variables and Further Optimization by RSM

Initially to find out the experimental hotspot we performed some laboratory trials and tried different reagent combinations on the basis of previous literature survey. The list of the adapted reagent combinations of dioxane-acetic acid, GVL-acetic acid and GVL-dioxane catalyzed by HCl along with their operating conditions are mentioned in Table 1. As it can be seen in the table, the values in terms of lignin yield were quite noticeable for all the combinations with the highest value of 42% for GVL: acetic acid combination and the lowest one 2.5% with dioxane acetic acid conditions. These values are quite comparable with the previously published literature. For example, in a previous study, a lignin yield of 43% was obtained when organosolv reagent (H2O: EtOH/35:65 v/v) combination was employed at 162 °C for 78 min and with 1.6% H_2SO_4 [38]. In a similar other study, a lignin yield of 23.8% was obtained from corn stover when deep eutectic solvent (choline chloride and formic acid (1:2)) were employed at a higher temperature of 130 °C for 2 h of time [39]. In our reagent combination of dioxane and acetic acid with 10 mL of HCl dose and at 70 °C of temperature for 6 h of process time a lignin yield of 11% was obtained and in a previous study a slightly higher yield of 14% was obtained when flax fiber was subjected to a two-step lignin extraction process with the help of formic acid and hydrogen peroxide [40]. For our case of dioxane and acetic acid although the yield is much lower, this lower yield can be anticipated due to the rather less harsh thermal conditions and such low yields are possible as in one previous study a lignin yield of 5% was obtained when imidazolium ionic liquids were used to treat woody biomass around 100 °C temperature and long retention times from 8 to 24 h [41].

Table 1. Different reagent combinations for lignin yield, G units, M_w and polydispersity values.

Reagent (50% <i>v</i> / <i>v</i>)	Temperature (°C)	Time (h)	Lignin Yield (%)	G Units *	Mw	Mn	PDI *
Di:AA ¹ *	70	6	11	200 ± 20	4732	2290	2.06
Di:AA ² *	Room temp.	48	2.5	332 ± 30	4718	2473	1.90
GVL:AA ¹ *	70	6	42	100 ± 35	3604	1851	1.94
GVL:Di ² *	70	6	38	110 ± 40	4562	2064	2.21

Di: AA 2* = dioxane acetic acid with 5 mL HCl; GVL: AA 1* = GVL acetic acid with 10 mL HCl; GVL: Di 2* = GVL dioxane with 10 mL HCl; PDI * = Mw/Mn; G Units * = μ mol/g.

Since our objective was to extract high quality lignin, all these extracted samples were further subjected to the DFRC analysis and their results were quite different within

themselves. As can be seen in the table, the highest DFRC value was for dioxane: acetic acid combination when they were treated at room temperature for about 48 h with a value of 332 units. However, this value dropped significantly to 200 units when this combination was subjected to mild thermal severity 70 °C for about 6 h of time. Although in this case the yield was enhanced greatly to 11%, the DFRC value was dropped as well indicating that this combination did not preserve well the prototype structure of the isolated lignin. Moreover, the synergism of the gamma valerolactone (GVL) with both the acetic acid and dioxane also did not preserve well the higher DFRC values and these were 100 and 110, respectively, mainly due to the high thermal severity (70 °C) and high HCl dose (10 mL). For these two combinations, we did not observe any lignin yield with either 5 mL HCl or at room temperature. As far as the molecular weight and the polydispersity are concerned, these values were also in a narrow margin but the improved polydispersity (1.90) was achieved with the highest DFRC value. Therefore, keeping in view the high efficiency of the synergism of dioxane: acetic acid under HCl catalyzed conditions we chose this combination and named this initial lignin as dioxane: acetic acid lignin (DA lignin) to further optimize in terms of high yield and with high DFRC values. To this end, we chose Design Expert software to sort out the optimum combination via a statistical way using response surface methodology. We selected three variables: time from 24 to 72 h, solids loading from 5% to 15%, and HCl dose from 5 to 15 mL. All these variables with their values along with their different experimental combinations were performed accordingly and the response was tabulated and reported in Table 2.

Scheme 1	Time (X ₁)	Solids Loading (X ₂)	Catalyst Dose (X ₃)	Lignin Yield (%)
1	24	10	15	4.0
2	24	10	5	2.0
3	24	15	10	3.1
4	24	5	10	3.5
5	48	10	10	8.1
6	48	15	15	9.0
7	48	15	5	6.0
8	48	5	15	8.0
9	48	5	5	5.0
10	48	10	10	8.3
11	48	10	10	8.6
12	72	15	10	10.0
13	72	10	5	5.5
14	72	10	15	11.0
15	72	5	10	8.0

Table 2. Actual values of the experimental combinations.

For these performed experiments, the equation representing the adjusted quadratic model (model fitness with its significance suggested by software) in terms of coded factors is described in the following equation.

$Y = +8.33 + 2.742X_1 + 0.45X_2 + 1.69X_3 + 0.60 X_1X_2 + 0.88 X_1 X_3 + 0.000 X_2 X_3 - 1.78 X_1{}^2 - 0.40X_2{}^2 - 0.93 X_3{}^2 - 0.93 X_$

where the lignin yield is denoted by Y and the isolation variables are represented as X_1 time, X_2 as solids loading, and X_3 as catalyst dose. In order to study the model adequacy, the ANOVA table is presented in Table 3. For this model the adjusted R-squared value and R-squared values were 0.9857 and 0.9994, respectively.

As it can be seen in the Table 2, the lignin yield varied quite considerably with the variation of the extraction conditions. It became fairly prominent with the catalyst dose and the time of the extraction process adapted. This evidence was also found from the ANOVA table. It is evidenced there that the model is quite significant having a *p* value less than 0.0001. Moreover, the adjusted R squared value as well as the R squared value 0.9857 and 0.9994, respectively, are noticeably high showing thus the model adequacy. In our model, the independent variables of both the time and catalyst dose have a quite significant

impact on the lignin yield. For the time the *p* value is <0.0001 showing a prominent effect on the yield. This phenomenon is mainly regarded as time prolongation that can help to loosen the native biomass web to dissolve out the lignin portion of the biomass under such conditions [42]. In the case of the catalyst dose, the *p* value also exhibited a value lower than 0.0001. This prominent effect is highly likely due to the strong protonation effect of the catalyst HCl [40]. This acid catalyst, regarding lignin isolation, offers a better selective extraction retaining a good quality [43].

Scheme 2 Sum of Squares df Mean Square F Value p Value Prob > F 103.04 9 11.45 108.18 < 0.0001 Significant Model X₁—Time 59.95 59.95 566.47 1 < 0.0001 Significant -Solid loading 1.62 1 1.62 15.31 0.0113 Significant X₃—Catalyst 22.78 1 22.78 215.26 < 0.0001 X_1X_2 1.444 1.4413.61 0.0142 X_1X_3 3.06 1 3.06 28.94 0.0030 X_2X_3 0.000 0.000 0.000 1 0000 1 X^{2}_{1} 11.69 1 11.69 110.44 0.0001 X^{2}_{2} 0.60 0.60 5.700.0626 1 X^2_3 0.0027 3.19 1 3.19 30.12 Residual 0.535 0.11 Lack of fit 0.40 3 0.13 2.12 Not significant Pure error 0.13 2 0.063

Table 3. ANOVA for response surface optimization of the variables.

Adj R-squared 0.9857, R-squared 0.9949.

However, in the case of solids loading it was not found to be that significant with a comparatively higher value of 0.0113. For this variable, under our conditions it is anticipated that the catalyst was not penetrated enough to open up the recalcitrant structure to dissolve out the lignin portion. This interactive behavior is due to the fact that the higher biomass loading needed some additional thermomechanical and/or thermochemical treatments to expose the inner biomass surfaces for the catalyst transport as the outer biomass layer may swell, due to the acidic environment, on the surface offering a hindrance for the catalyst to penetrate deep inside [44].

For the interactive effects, it was found that only the effects of time and catalyst dose have p values (0.0030) less than 0.005. This interactive effect is quite obvious as both are significant even individually to cause the enhanced lignin yield. This similar trend was also obvious in our preliminary trials. Additionally, there are some previous studies where the similar trend was exhibited [45]. Therefore, it can be suggested that the synergism of both the time and HCl dose can lead to a better lignin yield but at room temperature. Regarding the interactive effects of time and solids loading, its p value was 0.0142 showing not a significant effect and a similar trend was also observed in the case of the solids loading and the catalyst, where the p value was also not significant and showed a value of 1.0000.

Under all of the aforementioned conditions, the optimal lignin isolation combination was found out to be at 48 h of time, 10% of solids loading, with 10 mL of catalyst dose with a predicted lignin yield of 8.33%. The predicted value was confirmed experimentally which yielded the lignin with 8.10%. Our results are fairly comparative with previously published studies. For example, in one similar study, almost 15% lignin yield was obtained by adapting typical Soxhelet apparatus. In that study, they used dioxane with 0.4% v/v HCl and 3% v/v water for the extraction of low molecular weight lignin [46]. In another study, a lignin yield of 5% lower than our yield was obtained when imidazolium ionic liquids were used to treat woody biomass around 100 °C temperature and long retention times from 8 to 24 h [41]. In another study a lignin yield of 7.2% was obtained when sugar cane bagasse was dissolved in different concentrations of dioxane in 1% NaOH to extract the lignin [47]. In one such study a lignin yield of Baruns's native lignin was almost 8% obtained from the woody biomass taking into account the proto lignin structure of the extracted lignin [48].

3.2. Effect of Process Variables on the Lignin Yield

In order to observe the experimental effects of all the three lignin isolation variables, 3D response surface graphs were also drawn (Figure 1). These plots exhibit the interaction of two variables while keeping the third at the center value. In Figure 1A, the effect of solids loading and time while keeping catalyst dose at a fixed value is shown. During the lignin isolation process, an elevated time and solids loading both led to a corresponding rise in the yield of lignin. However, after a certain time, a further increase of these variables resulted in less lignin yield. Moreover, in short time of the process no significant difference in the lignin yield was observed when then solids loading was varied f between their range of 5-15% w/w. Similarly, for the other two variables, the trend kept its similar behavior trend (Figure 1B,C).



Figure 1. 3D response surface plots for interactive behaviour of process variables: (**A**) solids loading and time, (**B**) catalyst dose and time, and (**C**) catalyst dose and solids loading.

As it is obvious from the figures, as solids loading and catalyst dose concentration rose, the yield concomitantly rose and then it decreased, indicating that neither harsh nor overly mild conditions had positive impacts on the lignin yield. However, the variables time and the catalyst strongly influenced the response (lignin yield) compared to the interaction of the process variables with solids loading, in our case only two process variables significantly contributed to the significance of quadratic model (value lower than p = 0.05); from a point of view of the interactive effect of process variables, time and catalyst dose have had a significant role on the lignin yield and this similar trend was also observed in our preliminary trials. From here, it can be anticipated that under our prescribed range of factors time and catalyst dose played a significant role to have a substantial impact on the lignin yield.

3.3. Molecular Weight Analysis of ODA and DA Lignin Samples

In order to further determine the molecular weight of the lignin extracted at the optimum conditions, named as optimum dioxane acetic acid (ODA) lignin and the previously sorted DA lignin, the GPC analysis was also performed. For the reference, milled wood lignin was also analyzed and the results are reported in Table 4. For the results of their molecular weights, the values are in comparison within each other but almost half of the milled wood lignin in terms of their weight average (M_w) value. Regarding their polydispersity values it was improved from 1.90 for DA lignin to 1.65 for ODA lignin. This is also somewhat expected under our adapted conditions as the low catalyst dose released less amount of lignin to be released but a high dose also helped to increase the yield but also led to the formation of more lower molecular weight fractions and this was also evident from our model values. As it can be seen in the table, the molecular weight of ODA lignin was quite high with a value of 4358 and a corresponding polydispersity of 1.65 but as expected the values for DA lignin were 4718 and 1.90, respectively. As compared with the milled wood lignin, it has high value of polydispersity of 2.06 and the molecular weight was 10,660 units almost double the extracted lignin. These values are in a fair comparison with the previously published literature. For example, in one such previous study, a mixture of ethanol/water (50/50 w/w) catalyzed by 1.2% w/w H₂SO₄ was used at 180 °C and 1 h of time to isolate the lignin from eucalyptus showed a molecular weight value of 5079 but with a rather higher polydispersity of 3.24 [49]. In another study, the acetic acid (87% aqueous w/w) catalyzed by HCl (0.6 gm of 36% w/w HCl) 114 °C for 80 min was used to extract lignin from bamboo shoot shell and this lignin also exhibited a quite low molecular weight of 2789 with 1.54 of improved polydispersity quite closed to our ODA lignin value (1.65) [50]. Similarly, acetosolv lignin isolated from siam weed stem at 107 °C for 3 h of time employing 93% acetic acid–water v/v with 0.1% (w/w of liquid phase) HCl as catalyst showed a high polydispersity of 2.83 but with rather low molecular weight of 2010 [27].

wive L samples).				
Scheme 3	$\mathbf{M}_{\mathbf{w}}$	M _n	(M _w /M _n) P	
MWL	10,660	5157	2.06	
ODA lignin	4358	2641	1.65	

2641 2473

1.90

4718

ODA lignin

DA lignin

Table 4. Molecular weights of Optimum Dioxane, Dioxane & Milled Wood Lignin (ODA, DA, and MWI samples)

For the GPC chromatograms, all of the three lignin samples exhibited the homogeneous curves showing their relative unique nature (Figure 2). In one of our previous studies, when the GVL was employed as a reagent for milled wood pine saw dust, the polydispersity was considerably high in the range of 2.54–2.96 but under this process condition the polydispersity was noticeably improved [51]. From here it can be anticipated that the thermal conditions may lead to a harsher condition for the lignin isolation even under mild reagent conditions.



Figure 2. GPC chromatogram of ODA, DA, and MWL samples.

3.4. DFRC Values of ODA and DA Lignin Samples

To investigate further the lignin structure in terms of their typical qualitative units of both the lignin samples ODA and DA and to subsequently compare it with milled wood lignin a DFRC method was also used to degrade and finally isolate the β -aryl ether structures with the help of the reagents mentioned in the materials section. Finally, these fractions were studied by GC-MS and the results are shown in Table 5. As it is obvious in the table, both the lignin fractions show only G types of units due to a typical softwood biomass nature. The DFRC values for the milled wood lignin is around 450 µmol/gm and for the case of ODA lignin this value is somehow comparable with a value of 305 slightly lower than the DA lignin value of 332. It can be anticipated here that the ODA lignin was somewhat in lower value of DFRC than the DA lignin due to increased amount of HCl addition as it has also been previously reported [24]. Our values stand prominent with the previously investigated articles. One such study revealed that a lignin sample isolated from giant reed biomass had a G units' value of 285.7 µmol/gm. This lignin was also isolated at rather mild conditions of 49 $^{\circ}$ C for 24 h of time using H₂O₂ and NaOH as reagents [45]. Similarly, maize lignin also showed a G units' value of 342 µmol/gm while this lignin was isolated via the classical milled wood lignin method. In the similar study, the value was fairly increased to 504 μ mol/gm when the bamboo lignin was isolated and characterized in the same way [52]. In one another study a value of these G units of 310 was obtained when the Eucalypt globulus woody biomass was subjected to lignin isolation. In this study lignin was extracted via a classical pulping method with Na₂O while keeping 30% sulfidity at a temperature of 80 °C for about 120 min of time interval [53]. As it can be anticipated, the harsh lignin isolation conditions may lead to the cleaving of the aryl-ether linkages which are responsible for the higher DFRC values in the lignin samples [54].

Table 5. DFRC values of the ODA, DA, and MWL samples.

Scheme	G Units	S Units	Total
MWL	450 ± 34	-	450 ± 34
ODA lignin	305 ± 25	-	305 ± 25
DA lignin	332 ± 30	-	332 ± 30

Nonetheless, it can be suggested that our adapted process was quite meaningful in isolating the lignin with reasonable DFCR values proving thus that this modified process without thermal conditions has led to the less degradation of the lignin.

3.5. 2D HSQC NMR Spectra

Finally, we also quantified the β -O-4 contents present in the extracted lignin samples by performing their 2D-HSQC-NMR analysis. For this, 20 mg of nonacetylated lignin was quantitatively used and was dissolved in 0.75 mL DMSO-D6 and then the analysis data was acquired with a Bruker AVANCE 600 spectrometer (Bruker, Germany). The relative quantification and the corresponding assignments of the A_{α} , B_{α} , and C_{α} are tabulated in Table 6 (β -O-4, β - β , and β -5, respectively) and the corresponding NMR figure is shown in Figure 3. The relative quantification of the present contents in the lignin samples were calculated accordingly. As can be seen in the table values. our extracted lignin, in comparison with MWL (46.00%), was slightly lower in β -O-4 contents ranging from 37.80% for ODA lignin to 40.00% for DA lignin. For the values of the other linkages in the lignin such as β - β and β -5 a similar trend was observed but the value of β -5 (7.00 and 6.00 for ODA and DA, respectively) was closer to milled wood lignin value of 5.10. All these values were found to be in good comparison with the other process values. For example, in one of our previous studies this was around 28% for β -O-4 contents when we adapted GVL as a sole reagent for the lignin isolation at a high temperature value ranging from 120 to 160 °C without any acid dosing. This can be anticipated as the thermal conditions surely modify the lignin structure as well as the acid concentration. Although our values found in this study are slightly lower than the classical milled wood lignin value, they are still high enough to be compared with the previously published literature. For example, values of 33.0 and 19.3 of β –O–4 linkages were reported when the baggase was treated with Ethanolamonium malonate (2He Mal) and Ethanolamonium malonate (2He Lac), respectively. However, their lignin extraction conditions were still harsh with 150 and 160 °C, respectively [55]. In a similar other study, a value of 32.4 was obtained when the lignin was isolated from wheat straw with 62% (wt%) formic acid at 130 °C for 20 min of time [56]. However, in another study, a value of as high as 44.4 β –O–4 was reported when Eucalyptus camaldulensis Dehnh was treated with 80% GVL/20% H₂O mixture catalyzed by 20 m M H₂SO₄ for 30 min of time [57]. Almost similar values of β –O–4 linkages were quantified as 45.54 units when abaca biomass was subjected to lignin isolation under alkaline conditions of 20 wt% NaOH with 10 wt.% H₂O₂ at a temperature of 105 °C for a process time of 3 h [58].

Table 6. Relative amounts of linkages in lignin samples.

	HO HO O S Z O		
	β-0-4	β-β	β-5
Pine MWL	46.0	3.0	5.1
ODA lignin	37.8	6.0	7.0
DA lignin	40.0	5.0	6.0



Figure 3. 2D HSQC NMR of ODA, DA, and MWL.

4. Conclusions

A new organosolv reagent combination was screened initially and statistically optimized subsequently to isolate high quality lignin fractions from pine saw dust at room temperature. Among from the three adapted process variables, only time and catalyst dose (HCl) played a significant role on the response of lignin yield. At the optimal combination of 48 h of time, 10% solids loading, and 10 mL HCl dose, a lignin yield as high as 8.10% was confirmed experimentally. A quadratic model fitness was suggested by the software to have a model significant value <0.0001 and with R-squared value of 0.9949. This optimal lignin fraction (optimum dioxane-acetic acid ODA lignin) exhibited fairly higher G units' values (305 μ mol/gm) together with the considerable β –O–4 contents (37.8%) worthy enough to be in comparison with the previously published studies. Moreover, this lignin fraction also showed a lower molecular weight with an improved polydispersity proving it to be a more uniform lignin fraction. This uniform molecular weight together with good proto lignin features make it a suitable candidate for diverse yet wider industrial applications including biocomposites, resins, and sustainable polymers. From this it can be conclusively suggested that this approach can be considered as one sustainable and green option for the fractionation of lignocelluloses specifically for the sole lignin extraction processes.

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