



PROTECTIVE EFFECT OF DIETARY TAURINE FROM ROS PRODUCTION IN EUROPEAN SEABASS UNDER CONDITIONS OF FORCED SWIMMING.

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Supplementary material S1

Generation of standard curves for cat, sod, and gpx genes

To generate the standard curves defined amounts of mRNAs at tenfold dilutions were subjected to qPCR using iTaqTM Universal Probes One-Step Kit (Bio-Rad, Italy) and Bio-Rad® CFX96TM Real-Time PCR System. The thermal cycling protocol was 10 min at 50 °C, 3 min at 95 °C, followed by 40 cycles consisting of 15 s at 95 °C, and 1 min at 60 °C. The cycle threshold (Ct) values obtained by amplification were used to create standard curves for target genes. This curve served as a basis for calculating the unknown mRNA copies of each gene in each liver and muscle RNA sample.

Table S1 Sequences of primers used to synthesize standard RNA with relative accession number.

	Symbol	Acc. nr.	Primer Sequence (5'-3')
Superoxide	sod	FJ860004	F:gtaatacgactcactatagggGTTGGAGACCTGGGAGATGT
dismutase			R:GAAAAGGAGCAATGAGGAG
Catalase	cat	FJ860003	F: gtaatac gact cactataggg ATGGTGTGGGACTTCTGGAG
			R: CGTTTCTACTGCAAGTTCCACT
Glutathione	gpx	FM01366	F: gtaatac gact cactataggg AGTTAATCCGGAATTCGTGAGA
peroxidase			R: CAACAACCAGGGACTACACTCA

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Table S2 Primer sequences and TaqMan® probe of each target gene.

Gene	Symbol	Nucleotide sequence (5'-3')
		F: TGGAGACCTGGGAGATGTAACTG
Superoxide Dismutase	sod	R: CAAGATAGACATCACGGACAAGA
		Taqman Probe: CAGGAGGAGATAACATTG
		F: ATGGTGTGGGACTTCTGGAG
Catalase	cat	R: CATCAGGTGTCTTTCTTGTTCAGC
		Taqman probe: TGAGGCCTGAGTGTCTG
		F: AGTTAATCCGGAATTCGTGAG
Glutathione Peroxidase	gpx	R:GTTTTACGACCTGACAGCTAAGCT
		Taqman probe: AATGGCTGGAAACGTG