

Thyroid Hormones in Hemolymph of Red King Crabs from the Barents Sea

Alexander G. Dvoretzky ¹, Elena V. Tipisova ², Viktoria A. Alikina ², Aleksandra E. Elfimova ²
and Vladimir G. Dvoretzky ^{1,*}

¹ Murmansk Marine Biological Institute of the Russian Academy of Sciences (MMBI RAS), 183010 Murmansk, Russia; ag-dvoretzky@yandex.ru

² N. Laverov Federal Center for Integrated Arctic Research of the Ural Branch of the Russian Academy of Sciences (FECIAR UrB RAS), 163000 Arkhangelsk, Russia; tipisova@rambler.ru (E.V.T.); victoria-popcova@yandex.ru (V.A.A.); a.elfimova86@mail.ru (A.E.E.)

* Correspondence: v-dvoretzky@yandex.ru

Simple Summary: In vertebrates, thyroid hormones, triiodothyronine, and thyroxine play a pivotal role in regulating development, metamorphosis, and metabolism. Recently, the presence of endogenous thyroid hormone signaling has also been reported in invertebrates. We assayed thyroid hormone profiles in the red king crab, an iconic commercially important species, which was introduced into the Barents Sea in the 1960s and now is supporting a large-scale profitable fishery. Both thyroid hormones were detected in the crab hemolymph. Their levels fluctuated significantly depending on the maturity status of the crabs and sampling season. These findings confirm that thyroid hormones are involved in the specific physiological processes of red king crabs and may be used in aquaculture.

Abstract: Only a few studies have reported the presence and levels of thyroid hormones in invertebrates. At the same time, the administration of these hormones has been shown to be a profitable method for inducing growth in crustaceans. Knowledge of thyroid hormone concentrations is required to involve this approach in crustacean aquaculture. For this reason, we examined hemolymph samples of the invasive red king crab *Paralithodes camtschaticus* from the Barents Sea population for the levels of triiodothyronine (T3) and thyroxine (T4). Radioimmunoassays indicated that T3 ranged from 0.26 to 1.55 $\mu\text{g dL}^{-1}$, whereas T4 varied from 52 to 203 $\mu\text{g dL}^{-1}$. Adult crabs demonstrated higher levels of T3 than juveniles, while juvenile crabs had higher levels of T4 in comparison to larger conspecifics. The thyroxine levels were consistent throughout the study period. In contrast, significant seasonal variations were found for T3. In immature crabs, the maximum concentration was found in October, whereas in adult crabs, the highest level of T3 was registered in August. Most likely, these results are explained by changes in physiological conditions of the crabs associated with their molting calendar and migration activity. Our data may have important implications for aquaculture of *Paralithodes camtschaticus* in the Barents Sea.

Keywords: *Paralithodes camtschaticus*; red king crab; thyroid hormones; triiodothyronine; thyroxine; aquaculture; Barents Sea



Citation: Dvoretzky, A.G.; Tipisova, E.V.; Alikina, V.A.; Elfimova, A.E.; Dvoretzky, V.G. Thyroid Hormones in Hemolymph of Red King Crabs from the Barents Sea. *Animals* **2022**, *12*, 379. <https://doi.org/10.3390/ani12030379>

Academic Editors: Eva Armero and Manuel Machado

Received: 27 December 2021

Accepted: 2 February 2022

Published: 4 February 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

There are two main thyroid hormones: 3',5',3,5-L-tetraiodothyronine (thyroxine), which is nowadays commonly called T4, and 3',5,3-L-triiodothyronine, T3 [1]. These hormones are composed of two tyrosyl residues linked through an ether linkage and substituted with 4 or 3 iodine residues, respectively. In vertebrates, triiodothyronine is the biologically active hormone, whereas thyroxine, the major thyroid hormone that is secreted from the thyroid gland consisting of follicles, is a precursor or prohormone. Deiodination of T4 leads to production of T3 and reverse T3 (rT3) [2]. Thyroids play a crucial role in regulating early development, metamorphosis, and metabolism in mammals, amphibians, and

fish, and most vertebrates are unable to grow and reach their normal adult form without them [3–6]. Non-vertebrate chordates, such as tunicates, cephalochordates, and the larvae of lampreys have the endostyle, which acts as a thyroid gland homolog [7,8]. Non-chordate invertebrates without thyroid follicles have been shown to accumulate iodo-compounds exogenously from seawater and through diets containing micro- and macroalgae [9]. Heyland et al. [10] suggested the presence of endogenous thyroid hormone signaling in echinoderms and mollusks, which may reflect the existence of iodination processes. There are only a few reports concerning thyroid hormone profiles in invertebrates, including terrestrial gastropods, marine bivalves, and mud crabs [11–13]. Because thyroid hormones are able to affect reproduction processes in invertebrates, these substances are considered promising agents in shellfish aquaculture [14–16].

The red king crab, *Paralithodes camtschaticus* (Tilesius, 1815), is an example of a well-organized and successful introduction of a non-indigenous species to a new place of habitat to establish a new population for commercial exploration. On the one hand, although some authors reported alterations in species composition, structure, and biomass of benthic communities at some coastal sites of the Barents after the crab introduction [17,18], these changes are not as critical as was predicted and the major fish stocks remained unaffected [19]. On the other hand, *Paralithodes camtschaticus* has become abundant both in the coastal waters and in the open Barents Sea and now its population supports a profitable fishery [19–22]. In Russia, this fishery was initiated in 2004 and stock indices of *Paralithodes camtschaticus* fluctuated over subsequent years due to range extension of the crabs, overfishing, and climate change [19,20,23,24]. Annual landings have increased from 9187 metric tons in 2018 to 9836 t in 2019 and 10,820 t in 2020 [21,25–27]. The total allowable catch for the 2021 fishery season was set at 11,815 t [25]. Moreover, the red crab stock now supports recreational fishing with an annual quota of 100 t [28].

Although many life-history traits of the Barents Sea red king crabs are well-studied [19–24,29–36], there is a lack of knowledge on their physiology, especially, hormone profiles in the hemolymph [37–39]. Recently, the presence of thyroid hormones has been reported in red king crabs [40], but those data are restricted and more detailed information is required for further research, monitoring, and aquaculture purposes.

For this reason, the aim of our study was to measure concentrations of thyroid hormones in hemolymph of red king crabs from the Barents Sea in relation to their sex, maturity status, and sampling season.

2. Materials and Methods

Red king crabs were collected by divers at 5–38 m and by using bottom traps at 50–60 m depths in the coastal waters of the Kola Peninsula (Southern Barents Sea, 69.26° N, 33.3° E–69.11° N, 36.1° E, Figure 1) in June and August 2008 and May, July, and October 2009.

After being collected, the crabs were visually inspected for sex and shell condition. Each crab was weighed and its size was measured using calipers across the carapace length (CL, the greatest straight-line distance from the posterior margin of the right eye orbit to the medial-posterior margin of the carapace) [41].

All the crabs used for biochemical analyses had new or old shells (2–20 months post ecdysis) [41].

Male red king crabs with CL < 90 mm were considered “juveniles” whereas males with the CL > 90 mm were classified as “adults” according to Dvoretzky and Dvoretzky [36]. This separation corresponds to the results of histological data: the individuals of *P. camtschaticus* larger than 90 mm CL had ripe spermatozoa in their testes in contrast to smaller specimens [39]. The same female groups were separated according to both size-at-maturity data (presence or absence of external eggs on their pleopods) [42] and histological data (mature females had oocyte diameter > 280 µm) [29].



Figure 1. Map of the study area.

Hemolymph samples (volume 0.6 mL) were taken from the articulating membrane between the dactyl and propodus of the third walking leg using a sterile syringe with a needle, both washed with an anticoagulant containing sodium citrate and EDTA to prevent coagulation of the samples [39]. Each hemolymph sample was placed in a sterile 2-mL plastic tube containing 1.4 mL of 96% ethanol. The samples were frozen and then transported to the laboratory at the Institute of Physiology of Natural Adaptations (Arkhangelsk, Russia) for biochemical analyses. The analyses were performed within 2–4 months after sampling.

Radioimmunoassays were carried out according to the protocols suggested by the manufacturer (A Beckman Coulter Company, Villepinte, France) using Immunotech Total T4 RIA KIT and Immunotech Total T3 RIA KIT. In the case of total thyroxine (TT4), a 20- μ L hemolymph sample and 500 μ L of 125 I-labelled T4 were added serially to an antibody-coated tube. In the case of total triiodothyronine (TT3), a 25- μ L hemolymph sample and 200 μ L of 125 I-labelled T3 were used in the assays. A tube containing 500 μ L of 125 I-labelled T4 (or 200 μ L of 125 I-labelled T3) was included in every assay to determine the total 125 I-activity (the sample “T”). Tubes with “total thyroxine” or “total triiodothyronine” were incubated with constant shaking (>280 rpm) at ambient temperature (18–25 °C) for 1 h. At the end of incubation, the radioactivity of samples was determined using a scintillation counter (ARIAN, producer OOO VITAKO, Moscow, Russia). The standard graphs were plotted and the concentrations of total thyroxine or total triiodothyronine were obtained from the standard plot. The results were multiplied by a dilution factor of 3.33. Because thyroxine was observed at concentrations above the established working range for the kit, the hemolymph samples were additionally diluted 2 or 3 times with 95% ethanol and the results were then multiplied by a dilution factor of 2 or 3. The analytical sensitivity of the total thyroxine kit was 1010.1 ng dL⁻¹ (13 nmol L⁻¹), the working RIA range was 1010.1–31,080 ng dL⁻¹ (13–400 nmol L⁻¹), and the coefficient of variation between samples was 6.2%. For the total triiodothyronine kit, these levels were 19.5 ng dL⁻¹ (0.3 nmol L⁻¹), 19.5–780 ng dL⁻¹ (0.3–12 nmol L⁻¹), and 6.3%, respectively.

Size and weight data were compared between male and female red king crabs and between juveniles and adults using a one-way analysis of variance (ANOVA) for normal data or non-parametric Kruskal–Wallis tests in cases of non-normal data distribution. The

same approaches were applied for the levels of thyroid hormones in relation to the sex and maturity status of red king crabs as well as sampling season. When the test was significant, mean values for different groups were compared using the Tukey–Kramer test for mean values or Bonferroni test for medians, and differences were considered significant at $p < 0.05$. Statistical analyses were carried out using Statistica 10. Mean values are presented with standard errors (\pm SE).

3. Results

A total of 23 juvenile crabs (16 females and 7 males) and 40 adult individuals (11 females and 29 males) were captured in the Southern Barents Sea (Figure 2). Their size and weight variations are summarized in Table 1.

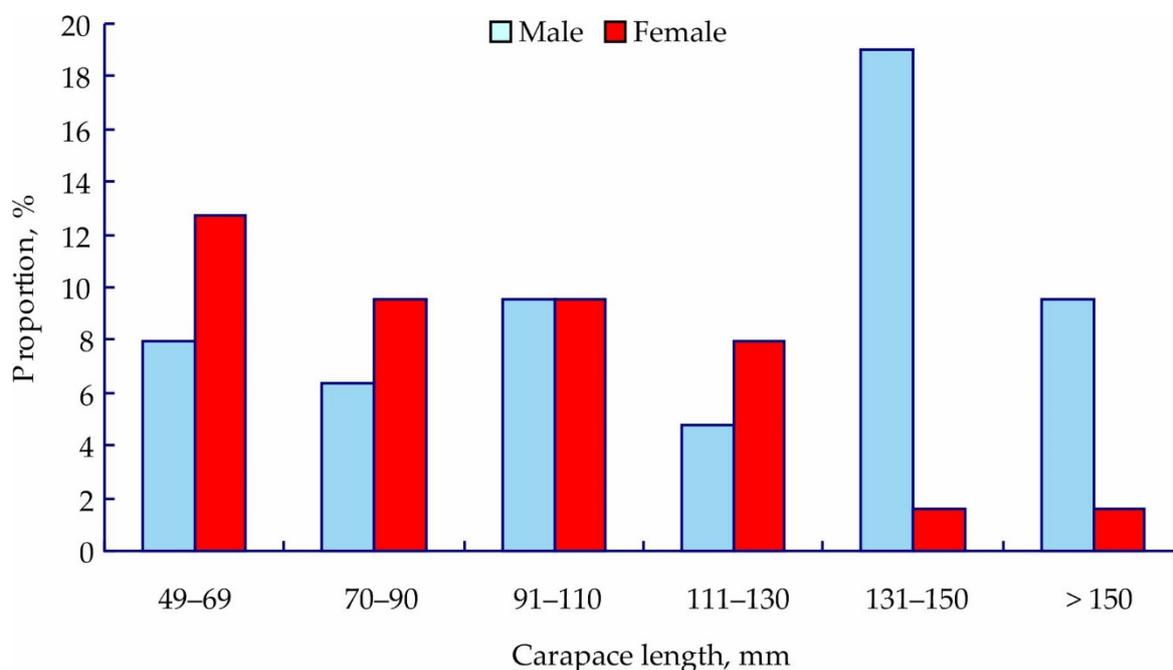


Figure 2. Size–frequency distribution of red king crabs from the coastal Barents Sea.

Table 1. Morphometric parameters of red king crabs used for biochemical analyses.

Sex	Maturity	N	Carapace Length, mm				Weight, g			
			X	SE	Min	Max	X	SE	Min	Max
M	Juv	7	66.8	4.3	54.6	83.1	239.7	47.7	131	432
M	Ad	29	130.7	4.3	85.6	162.7	2015.4	184.4	474	3805
M	Tot	36	118.3	5.6	54.6	162.7	1670.1	190.0	131	3805
F	Juv	16	71.5	3.8	49.0	91.7	293.3	40.5	97	532
F	Ad	11	120.1	5.1	99.8	158.0	1449.6	270.0	665	3820
F	Tot	27	91.3	5.6	49.0	158.0	764.4	156.2	97	3820
M + F	Juv	23	70.1	2.9	49.0	91.7	277.0	31.6	97	532
M + F	Ad	40	127.8	3.5	85.6	162.7	1859.8	156.4	474	3820

Note: M—male, F—female, Juv—juveniles, Ad—adults, tot—total, N—sample size, X—mean, SE—standard error, Min—minimum, Max—maximum.

Mean carapace length and weight in juvenile males and females were similar (ANOVA, $df = 1$, $F = 0.534$, $p = 0.473$ for CL and Kruskal–Wallis test, $df = 1$, $H = 0.599$, $p = 0.447$ for weight). The same result was obtained for adult specimens (ANOVA, $df = 1$, $F = 2.027$, $p = 0.155$ for CL and Kruskal–Wallis test, $df = 1$, $H = 2.248$, $p = 0.133$ for weight).

Biochemical assays have shown the presence of both triiodothyronine and thyroxine in the hemolymph of the red king crabs analyzed. Concentrations of T3 varied from 0.26

to $1.55 \mu\text{g dL}^{-1}$ (mean, $0.64 \pm 0.04 \mu\text{g dL}^{-1}$), while concentrations of T4 ranged from 52 to $203 \mu\text{g dL}^{-1}$ (mean, $129 \pm 5 \mu\text{g dL}^{-1}$). These levels demonstrated fluctuations depending on size, sex, and hormones (Figure 3). There were no significant differences between the T3 levels in juvenile males, juvenile females, and adult females, whereas juvenile concentrations of T3 were significantly lower in comparison to adult males (Table 2).

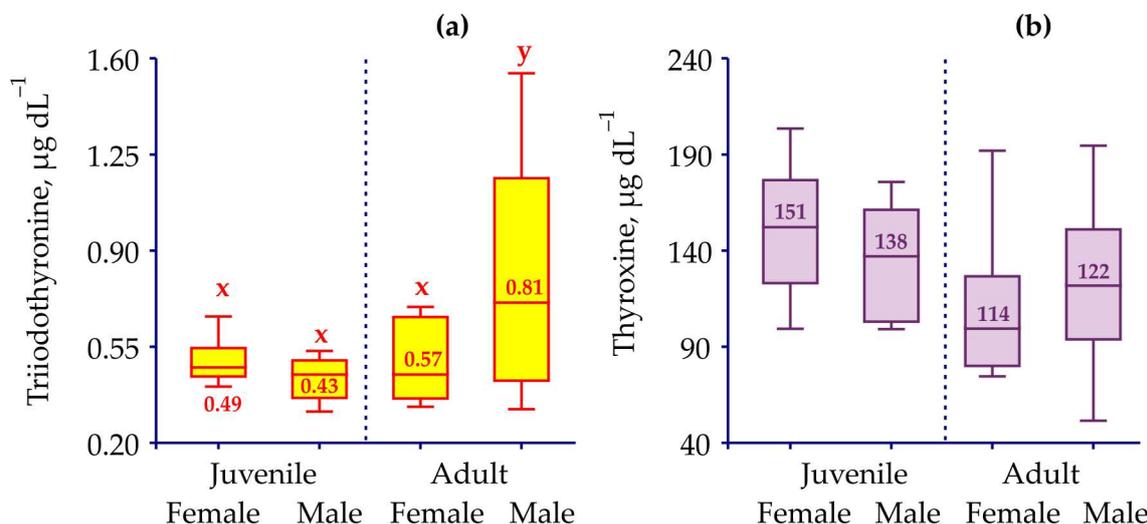


Figure 3. Variations in thyroid hormones in hemolymph of red king crabs (box plots) in relation to their sex and maturity status. Numbers indicate mean levels. Bars with the same letter are not significantly different ($p > 0.05$). (a) Triiodothyronine, (b) thyroxine.

Table 2. Results of ANOVA or Kruskal–Wallis tests (KWT) comparing the differences in thyroid hormone levels of red king crabs from the Barents Sea.

Hormone	Comparisons	Test	df	F (H)	p
T3	Juv-M vs. Juv-F	KWT	1	1.969	0.161
	Juv-M vs. Ad-M	KWT	1	5.284	0.022
	Juv-F vs. Ad-F	ANOVA	1	0.957	0.337
	Ad-M vs. Ad-F	ANOVA	1	3.393	0.073
	Tot-M vs. Tot-F	ANOVA	1	6.970	0.011
	Juv-Tot vs. Ad-tot	ANOVA	1	10.950	0.002
	July vs. Oct (Juv)	ANOVA	1	4.540	0.045
	May vs. June vs. Aug vs. Oct (Ad)	ANOVA	3	9.480	<0.001
T4	Juv-M vs. Juv-F	KWT	1	0.755	0.385
	Juv-M vs. Ad-M	KWT	1	1.122	0.290
	Juv-F vs. Ad-F	ANOVA	1	7.099	0.013
	Ad-M vs. Ad-F	ANOVA	1	0.345	0.560
	Tot-M vs. Tot-F	KWT	1	0.866	0.352
	Juv-Tot vs. Ad-tot	KWT	1	7.750	0.005
	July vs. Oct (Juv)	ANOVA	1	0.001	0.995
	May vs. June vs. Aug vs. Oct (Ad)	ANOVA	3	2.120	0.116

Note: T3—triiodothyronine, T4—thyroxine, M—male, F—female, Juv—juvenile, Ad—adult, tot—total, d—degree of freedom, F—F-ratio for ANOVA, H—chi-square for the Kruskal–Wallis test, p—probability level. Bold font indicates significant differences.

The adult male and female concentrations did not differ significantly while the combined T3 level in females ($0.52 \pm 0.04 \mu\text{g dL}^{-1}$) was significantly lower than in males ($0.74 \pm 0.06 \mu\text{g dL}^{-1}$) (Table 2). In general, juveniles exhibited lower levels of T3 than adult crabs (0.47 ± 0.02 vs. $0.74 \pm 0.06 \mu\text{g dL}^{-1}$). The comparisons of T4 levels in male and female red king crabs as well as in adult and juvenile specimens have shown

similar concentrations between almost all the combinations of the groups compared, except for juvenile and adult females (Figure 3, Table 2). Contrary to T3, the hemolymph of juvenile crabs contained higher concentrations of T4 than that of adult specimens (147 ± 6 vs. $112 \pm 6 \mu\text{g dL}^{-1}$) (Table 2).

Seasonal fluctuations in triiodothyronine and thyroxine concentrations are presented in Figure 4.

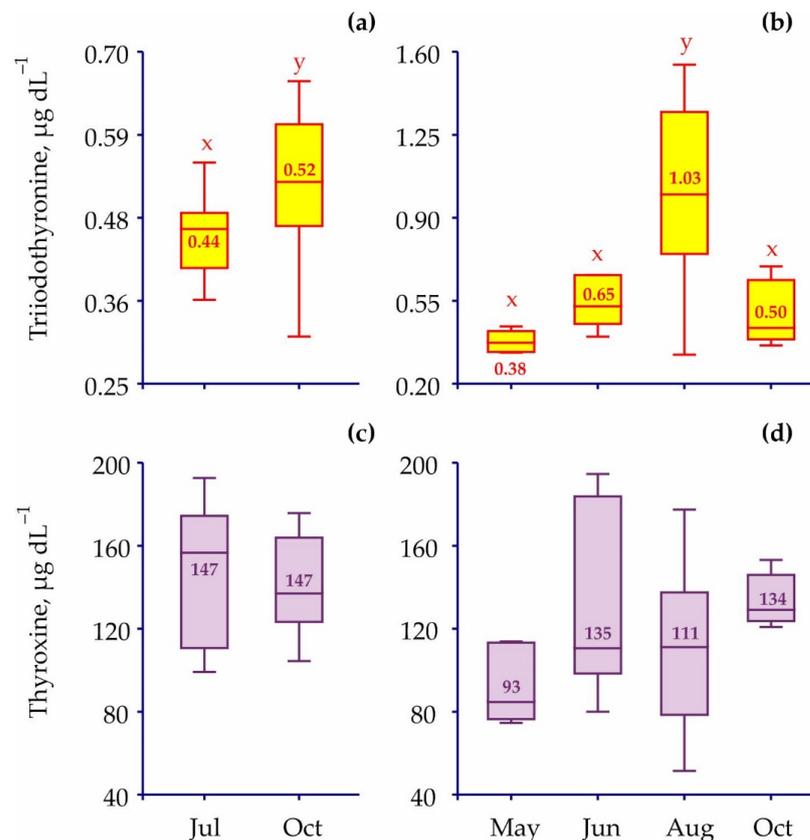


Figure 4. Seasonal variations in thyroid hormones in hemolymph of red king crabs from the coastal Barents Sea (box plots). Numbers indicate mean levels. Bars with the same letter are not significantly different ($p > 0.05$). (a) Triiodothyronine, juveniles; (b) triiodothyronine, adults; (c) thyroxine, juveniles; (d) thyroxine, adults.

In the case of juvenile red king crabs, the October level of T3 was significantly higher when compared to the July level (Table 2). Adult red king crabs demonstrated similar concentrations of triiodothyronine in May, June, and October, while a significant increase in the T3 level was observed in August. In contrast, the concentrations of thyroxine were consistent throughout the study period (Table 2).

4. Discussion

The most abundant cohort in this study was adult males with 131–150 mm CL. This size class includes commercial-sized individuals, i.e., the most important population group in terms of its ecological role and significance for fishing. According to size-at-age data [36], the age of the males used for biochemical assays in this study varied from 3 to 16 years. The predominance of males has been reported by other authors at western sites of the Kola Peninsula [30], while females usually dominate the catch at eastern sites [33,35]. Adult females were less abundant in the larger size classes because their growth rate is lower than that of males, reflecting the fact that females invest more energy into reproduction processes than into somatic growth than males do [24,35].

Our study confirmed that both juvenile and adult individuals of *Paralithodes camtschaticus* have detectable concentrations of T4 and T3 in their hemolymph, but the present levels were higher than those reported by Zenzerov and Tipisova [40], probably due to the lower sample size used in the previous study and restricted study period. Iromo et al. [11] studied thyroxin concentrations in hemolymph of female mud crabs of *Scylla serrata* from Tarakan Island, East Borneo, Indonesia, and found that immature crabs exhibited the lowest concentration of T4 (8.50 ± 2.17 ng mL⁻¹), maturing crabs have the intermediate level of T4 (11.23 ± 2.06 ng mL⁻¹), and the highest concentration was found in ripe females (15.33 ± 7.20 ng mL⁻¹). The same pattern was found for the ovaries of *Scylla serrata* (18.47 ± 2.06 vs. 27.07 ± 1.92 vs. 42.27 ± 3.17 ng mL⁻¹), indicating that thyroxin plays an important role in vitellogenesis. Thyroid hormones have also been found in other invertebrate taxa. Huang et al. [12] reported that larvae of the Pacific oyster *Crassostrea gigas* contain triiodothyronine and thyroxine in concentrations of 0.1–0.35 µg g⁻¹ and 0.5–4 µg g⁻¹, respectively. They concluded that these oysters have the ability to synthesize thyroid hormones endogenously. Later radioimmunoassay analyses have shown the presence of T4 (concentration 3.49 ± 0.55 µg dL⁻¹) in hemolymph of the giant African snail *Achatina fulica* [13]. The authors also found that the thyroxine level decreased significantly to 2.27 ± 0.16 µg dL⁻¹ as a result of 4-week starvation. All these findings support the opinion that levels of thyroid hormones do significantly correlate to physiological conditions of animals, rising or falling in critical phases of their life cycles [1]. We found that adult red king crabs had lower concentrations of T4 than juvenile ones and an opposite pattern was observed for T3. Adult crabs are known to expend more energy due to larger size, reproduction, and greater migration activity [29]. As a result, the concentration of T3 in their hemolymph is higher than in juvenile crabs. Taking into account that this hormone is synthesized from T4, it is not surprising that the level of T4 was lower than in smaller crabs. A similar pattern (an increased level of T3 and a stable level of T4) has been reported for oysters of *Crassostrea gigas* [12].

Most likely, seasonal patterns observed for triiodothyronine concentrations in the hemolymph of the red crabs can also be attributed to fluctuations in their physiological conditions. Indeed, in juveniles, the highest concentration of T3 occurred in October. According to published data, juvenile red king crabs at age 3–5 years molt 2 or 3 times per year in March–May and October–December [29,43]. The molting process is a high-energy-dependent event and higher triiodothyronine levels are expected in the period when mass molting occurs. Although ecdysteroids are considered the major molting hormones in crustaceans [44], recent studies have indicated that molt induction in black tiger shrimps, *Penaeus monodon*, led to expression of genes associated with vitellogenin, including thyroid hormone receptor associated protein [45].

Adult red king crabs exhibited the highest concentration of T3 in August, i.e., the month when both large females and males start to migrate to the open sea. The first migration peak occurs in March–April when the majority of males leave coastal sites after mating and spawning. In October–December, the coastal area is occupied by overwintering individuals of *Paralithodes camtschaticus* [29]. Taking into account that triiodothyronine has important effects on mitochondrial energetics [46] and can modulate several metabolic events similar to the effects of vertebrate thyroids [4], the higher levels of T3 may be associated with more intense motor activity of adult red king crabs. Juvenile red king crabs seem to spend less energy due to their smaller size and weight and less active lifestyle [20,29].

Thyroid hormones have been shown to induce a plethora of physiologic actions in crustaceans. Thus, exogenous application of thyroxine led to a significant increase in growth and molt cycle events in postlarvae and early juveniles of the shrimp *Penaeus monodon*, with the maximum growth increments at a dose of 3 µg L⁻¹ [47]. Roustaian and Gaik [48] also reported that exogenous application of T4 enhanced growth in larvae of the giant freshwater prawn *Macrobrachium rosenbergii* with the highest growth rates at doses of 1 and 2 µg L⁻¹, but precocious molt promoting events were not observed. Iromo et al. [14] showed

that maturation of female mud crabs *Scylla serrata* from vitellogenic stage 2 to 3 occurred 4 and 10 days earlier after injections of thyroxine at doses of 0.05 and 0.5 $\mu\text{g g}^{-1}$, respectively, but the second dose resulted in 100% mortality of experimental animals. Further studies showed that injections of T4 promoted the development of embryos to hatching eggs depending on dosage, with the best results at 0.15 and 0.1 $\mu\text{g g}^{-1}$ [15]. Adiputra et al. [49] have shown that injection of T4 at a dose of 0.1 $\mu\text{g g}^{-1}$ can induce a higher growth rate and better gonad development in male spiny lobsters of *Panulirus homarus*. Jayaprakas and Sambhu [50] found that thyroxine incorporated into a prepared diet promoted growth and feed utilization of white prawns of *Penaeus indicus*, being the most effective at 0.04 ppm. Recently, Pinandoyo et al. [51] have reported that T4 added to a diet for giant freshwater prawns stimulated their growth performance and decreased mortality rates, with the best results at a concentration of 10 mg kg^{-1} feed. Thus, thyroid hormones appear to promote growth and production in crustaceans.

Recently, Russian President Vladimir Putin has approved a strategy for the development of the Arctic zone of the Russian Federation for the period through 2035. Opportunities include further development of the aquaculture sector [27]. Coastal sites of the Kola Peninsula have favorable environmental conditions for invertebrate aquaculture [27,52], but profitable aquaculture cannot be organized without excellent knowledge regarding the ecology and physiology of target organisms [39,53,54]. The red king crab is a slow-growing species reaching the commercial size of 150 mm carapace width (or 137 mm carapace length) after 10 years of growth. Potential methods to decrease inter-molt intervals and improve growth performance of crustaceans include manipulation of their hormonal status by administration of steroid hormones [37,39,55] or double-stranded RNA (dsRNA) [56], dietary supplementation [50,51], and other methods [43]. Thyroid hormones may also be considered promoting agents for cultivation of *Paralithodes camtschaticus*. From this point of view, our results provide a baseline for further research in this field.

5. Conclusions

Although red king crabs lack a classical thyroid gland, they have the ability to utilize thyroid hormones which may be ingested exogenously, formed endogenously, or produced via catalysis (as has been suggested for echinoderms, mollusks, and mud crabs). Our data indicate that the hemolymph of adult crabs contains more triiodothyronine than juveniles, while immature crabs demonstrate higher thyroxine concentrations than mature individuals. Male and female red king crabs, both adults and juveniles, have similar concentrations of thyroid hormones. Seasonal changes in the triiodothyronine concentration reflect shifts in the physiological status of red king crabs associated with mass molting in juveniles and higher migration activity in adults. Further research should be focused on the exogenous administration of thyroid hormones to test their effects on growth rates, molting events, survival, and meat quality of red king crabs. Such studies would provide novel techniques for red king crab aquaculture in the Barents Sea.

Author Contributions: Conceptualization, A.G.D. and E.V.T.; data curation, A.G.D., A.E.E., V.A.A., A.E.E. and V.G.D.; validation, formal analysis, A.G.D., E.V.T., V.G.D., A.E.E. and V.A.A.; methodology, A.G.D. and E.V.T.; software, visualization, V.G.D.; project administration, A.G.D.; writing—original draft, A.G.D. and V.G.D.; writing—review and editing, A.G.D., V.G.D. and E.V.T. All authors have read and agreed to the published version of the manuscript.

Funding: This article is based upon work that was supported by the Ministry of Science and Higher Education of the Russian Federation under contracts to MMBI RAS and FECIAR UrB RAS.

Institutional Review Board Statement: This study was carried out in accordance with Russian legislation guidelines (Federal Act 52-F3, 24/04/1995) and approved by the Institutional Review of Murmansk Marine Biological Institute RAS (6732/253-ok, 01/12/2016).

Informed Consent Statement: Not applicable.

Data Availability Statement: The data are available on request from the corresponding author.

Acknowledgments: We are grateful to our MMBI colleagues for help in sampling and two anonymous reviewers for their valuable comments.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Hulbert, A.J. Thyroid hormones and their effects: A new perspective. *Biol. Rev.* **2000**, *75*, 519–631. [\[CrossRef\]](#)
2. Choksi, N.Y.; Jahnke, G.D.; Hilaire, C.S.; Shelby, M. Role of thyroid hormones in human and laboratory animal reproductive health. *Birth Defects Res. B* **2003**, *68*, 479–491. [\[CrossRef\]](#)
3. Mullur, R.; Liu, Y.-Y.; Brent, G.A. Thyroid hormone regulation of metabolism. *Physiol. Rev.* **2014**, *94*, 355–382. [\[CrossRef\]](#)
4. Sainath, S.B.; André, A.; Castro, L.F.C.; Santos, M.M. The evolutionary road to invertebrate thyroid hormone signaling: Perspectives for endocrine disruption processes. *Compar. Biochem. Physiol. C* **2019**, *223*, 124–138. [\[CrossRef\]](#)
5. Liotta, L.; Bionda, A.; Quartuccio, M.; De Nardo, F.; Visalli, R.; Fazio, E. Thyroid and lipidic profiles in nicastrese goats (*Capra hircus*) during pregnancy and postpartum period. *Animals* **2021**, *11*, 2386. [\[CrossRef\]](#)
6. Lall, S.P.; Kaushik, S.J. Nutrition and metabolism of minerals in fish. *Animals* **2021**, *11*, 2711. [\[CrossRef\]](#)
7. Crockford, S.J. Evolutionary roots of iodine and thyroid hormones in cell-cell signaling. *Integr. Comp. Biol.* **2009**, *49*, 155–166. [\[CrossRef\]](#)
8. Laudet, V. The origins and evolution of vertebrate metamorphosis. *Curr. Biol.* **2011**, *21*, R726–R737. [\[CrossRef\]](#)
9. Eales, J.G. Iodine metabolism and thyroid-related functions in organisms lacking thyroid follicles: Are thyroid hormones also vitamins? *Proc. Soc. Exp. Biol. Med.* **1997**, *214*, 302–317. [\[CrossRef\]](#)
10. Heyland, A.; Price, D.A.; Bodnarova-Buganova, M.; Moroz, L.L. Thyroid hormone metabolism and peroxidase function in two non-chordate animals. *J. Exp. Zool. B Mol. Dev. Evol.* **2006**, *306*, 551–566. [\[CrossRef\]](#)
11. Iromo, H.; Zairin, M., Jr.; Agus, M.S.; Manalu, W. Thyroxine distribution in the hemolymph, hepatopancreas, ovary, sponge, and larvae of female mud crabs (*Scylla serrata*) during ovarian maturation. *J. Crustac. Biol.* **2014**, *34*, 760–763. [\[CrossRef\]](#)
12. Huang, W.; Xu, F.; Qu, T.; Zhang, R.; Li, L.; Que, H.; Zhang, G. Identification of thyroid hormones and functional characterization of thyroid hormone receptor in the Pacific oyster *Crassostrea gigas* provide insight into evolution of the thyroid hormone system. *PLoS ONE* **2015**, *10*, e0144991. [\[CrossRef\]](#)
13. Lustrino, D.; Silva, A.C.M.; Araujo, I.G.; Tunholi, V.M.; Tunholi-Alves, V.M.; Castro, R.N.; Carvalho, D.P.; Pinheiro, J.; Marassi, M.P. Evidence of the presence of thyroid hormones in *Achatina fulica* snails. *An. Acad. Bras. Cienc.* **2017**, *89*, 2181–2188. [\[CrossRef\]](#)
14. Iromo, H.; Zairin, M., Jr.; Agus, M.S.; Manalu, W. Effectivity of thyroxine hormone supplementation in the ovarian maturation of females mud crab (*Scylla serrata*). *Pak. J. Biotechnol.* **2014**, *11*, 79–86.
15. Iromo, H.; Zairin, M., Jr.; Agus, M.S.; Manalu, W. Supplementation doses thyroxine hormone of broodstock mud crab (*Scylla serrata*) during ovarian maturation. *J. Aquac. Res. Dev.* **2015**, *6*, 12.
16. Iromo, H.; Jabarsyah, A.; Awaluddin. Reproduction of females mud crab (*Scylla serrata*) with thyroxine hormone supplementation in traditional ponds from north Borneo Indonesia. *Int. J. Fish. Aquat. Stud.* **2018**, *6*, 378–381.
17. Britayev, T.A.; Rzhavsky, A.V.; Pavlova, L.V.; Dvoretzkiy, A.G. Studies on impact of the alien red king crab (*Paralithodes camtschaticus*) on the shallow water benthic communities of the Barents Sea. *J. Appl. Ichthyol.* **2010**, *26*, 66–73. [\[CrossRef\]](#)
18. Oug, E.; Cochrane, S.; Sundet, J.H.; Norling, K.; Nilsson, H.C. Effects of the invasive red king crab (*Paralithodes camtschaticus*) on soft-bottom fauna in Varangerfjorden, Northern Norway. *Mar. Biodivers.* **2011**, *41*, 467–479. [\[CrossRef\]](#)
19. Dvoretzkiy, A.G.; Dvoretzkiy, V.G. Commercial fish and shellfish in the Barents Sea: Have introduced crab species affected the population trajectories of commercial fish? *Rev. Fish Biol. Fish.* **2015**, *25*, 297–322. [\[CrossRef\]](#)
20. Dvoretzkiy, A.G.; Dvoretzkiy, V.G. Red king crab (*Paralithodes camtschaticus*) fisheries in Russian waters: Historical review and present status. *Rev. Fish Biol. Fish.* **2018**, *28*, 331–353. [\[CrossRef\]](#)
21. Dvoretzkiy, A.G.; Dvoretzkiy, V.G. New echinoderm-crab epibiotic associations from the coastal Barents Sea. *Animals* **2021**, *11*, 917. [\[CrossRef\]](#)
22. Dvoretzkiy, A.G.; Dvoretzkiy, V.G. Epibiotic communities of common crab species in the coastal Barents Sea: Biodiversity and infestation patterns. *Diversity* **2022**, *14*, 6. [\[CrossRef\]](#)
23. Dvoretzkiy, A.G.; Dvoretzkiy, V.G. Inter-annual dynamics of the Barents Sea red king crab (*Paralithodes camtschaticus*) stock indices in relation to environmental factors. *Polar Sci.* **2016**, *10*, 541–552. [\[CrossRef\]](#)
24. Dvoretzkiy, A.G.; Dvoretzkiy, V.G. Effects of environmental factors on the abundance, biomass, and individual weight of juvenile red king crabs in the Barents Sea. *Front. Mar. Sci.* **2020**, *7*, 726. [\[CrossRef\]](#)
25. Bakanev, S.V.; Stesko, A.V. Red king crab. In *Materials Justifying Changes to the Previously Approved Total Allowable Catch in Fishing Areas in Inland Seas of the Russian Federation, on the Continental Shelf of the Russian Federation, in the Exclusive Economical Zone of the Russian Federation, in the Azov and Caspian Seas in 2021*; Sologub, D.O., Ed.; FGBUN VNIRO (Polar branch): Murmansk, Russia, 2021; pp. 2–16. (In Russian)
26. Dvoretzkiy, A.G.; Bichkaeva, F.A.; Baranova, N.F.; Dvoretzkiy, V.G. Fatty acid composition of the Barents Sea red king crab (*Paralithodes camtschaticus*) leg meat. *J. Food Compos. Anal.* **2021**, *98*, 103826. [\[CrossRef\]](#)
27. Dvoretzkiy, A.G.; Dvoretzkiy, V.G. Cucumaria in Russian waters of the Barents Sea: Biological aspects and aquaculture potential. *Front. Mar. Sci.* **2021**, *8*, 613453. [\[CrossRef\]](#)

28. Dvoretzky, A.G.; Dvoretzky, V.G. Renewal of the recreational red king crab fishery in Russian waters of the Barents Sea: Potential benefits and costs. *Mar. Policy* **2022**, *136*, 104916. [[CrossRef](#)]
29. Kuzmin, S.A.; Gudimova, E.N. *Introduction of the Kamchatka (Red King) Crab in the Barents Sea. Peculiarities of Biology, Perspectives of Fishery*; Publishing Kola Science Centre Russian Academy of Sciences: Apatity, Russia, 2002. (In Russian)
30. Sokolov, V.I.; Milyutin, D.M. Distribution, size-sex composition, and reserves of the red king crab (*Paralithodes camtschaticus*) in the upper sublittoral of the Kola Peninsula (the Barents Sea). *Zool. Zhurn.* **2006**, *85*, 158–171. (In Russian)
31. Pavlova, L.V. Red king crab trophic relations and its influence on bottom biocenoses. In *Biology and Physiology of the Red King Crab from the Coastal Zone of the Barents Sea*; Matishov, G.G., Ed.; Publishing Kola Scientific Centre Russian Academy of Sciences: Apatity, Russia, 2008; pp. 77–104. (In Russian)
32. Dvoretzky, A.G.; Dvoretzky, V.G. Distribution of amphipods *Ischyrocerus* on the red king crab, *Paralithodes camtschaticus*: Possible interactions with the host in the Barents Sea. *Estuar. Coast. Shelf Sci.* **2009**, *82*, 390–396. [[CrossRef](#)]
33. Dvoretzky, A.G.; Dvoretzky, V.G. Fouling community of the red king crab, *Paralithodes camtschaticus* (Tilesius 1815), in a subarctic fjord of the Barents Sea. *Polar Biol.* **2009**, *32*, 1047–1054. [[CrossRef](#)]
34. Dvoretzky, A.G.; Dvoretzky, V.G. Some aspects of the biology of the amphipods *Ischyrocerus anguipes* associated with the red king crab, *Paralithodes camtschaticus*, in the Barents Sea. *Polar Biol.* **2009**, *32*, 463–469. [[CrossRef](#)]
35. Dvoretzky, A.G.; Dvoretzky, V.G. Population dynamics of the invasive lithodid crab, *Paralithodes camtschaticus*, in a typical bay of the Barents Sea. *ICES J. Mar. Sci.* **2013**, *70*, 1255–1262. [[CrossRef](#)]
36. Dvoretzky, A.G.; Dvoretzky, V.G. Size-at-age of juvenile red king crab (*Paralithodes camtschaticus*) in the coastal Barents Sea. *Cah. Biol. Mar.* **2014**, *55*, 43–48.
37. Dvoretzky, A.G.; Dvoretzky, V.G. Hemolymph molting hormone concentrations in red king crabs from the Barents Sea. *Polar Biol.* **2010**, *33*, 1293–1298. [[CrossRef](#)]
38. Mota, V.C.; Siikavuopio, S.I.; James, P. Physiological responses to live air transport of red king crab (*Paralithodes camtschaticus*). *Fish. Res.* **2021**, *237*, 105882. [[CrossRef](#)]
39. Dvoretzky, A.G.; Tipisova, E.V.; Elfimova, A.E.; Alikina, V.A.; Dvoretzky, V.G. Sex hormones in hemolymph of red king crabs from the Barents Sea. *Animals* **2021**, *11*, 2149. [[CrossRef](#)]
40. Zenzerov, V.S.; Tipisova, E.V. Initial data on the level of thyroid hormones (Thyroxin, Triiodthyronine) and thyrotropic hormone of the hypophysis in the hemolymph of king crabs from the Barents Sea. *Dokl. Biol. Sci.* **2009**, *428*, 462–463. [[CrossRef](#)]
41. Donaldson, W.E.; Byersdorfer, S.E. *Biological Field Techniques for Lithodid Crabs. Fairbanks, Alaska Sea Grant College Program*; University of Alaska: Fairbanks, AK, USA, 2005.
42. Dvoretzky, A.G.; Dvoretzky, V.G. Size at maturity of female red king crab, *Paralithodes camtschaticus*, from the costal zone of Kola Peninsula (southern Barents Sea). *Cah. Biol. Mar.* **2015**, *56*, 49–54.
43. Dvoretzky, A.G.; Dvoretzky, V.G. Does spine removal affect molting process in the king red crab (*Paralithodes camtschaticus*) in the Barents Sea? *Aquaculture* **2012**, *326–329*, 173–177. [[CrossRef](#)]
44. Webster, S.G. Endocrinology of molting. In *The Natural History of Crustacea: Physiology*; Chang, E.S., Thiel, M., Eds.; Oxford University Press: Oxford, UK, 2015; pp. 1–35.
45. Uawisetwathana, U.; Leelatanawit, R.; Klanchui, A.; Prommoon, J.; Klinbunga, S.; Karoonuthaisiri, N. Insights into eyestalk ablation mechanism to induce ovarian maturation in the black tiger shrimp. *PLoS ONE* **2011**, *6*, e24427. [[CrossRef](#)]
46. Cheng, S.-Y.; Leonard, J.L.; Davis, P.J. Molecular aspects of thyroid hormone actions. *Endocrine Rev.* **2010**, *31*, 139–170. [[CrossRef](#)] [[PubMed](#)]
47. Pillai, S.M.; Verghese, P.U.; Ravichandran, P.; Roy, A.K. Effect of thyroxine on growth and moulting in *Penaeus monodon* Fabricius. *Ind. J. Anim. Sci.* **1987**, *57*, 241–245.
48. Roustaian, P.; Gaik, L.A. Effect of thyroxine immersion on larval survival, growth and postlarvae production of freshwater prawn, *Macrobrachium rosenbergii* (de Man). *Aquac. Res.* **2006**, *37*, 1378–1380. [[CrossRef](#)]
49. Adiputra, Y.T.; Zairin, M., Jr.; Suprayudi, M.A.; Manalu, W.; Brite, M. The effects of thyroxine hormone on gonadal maturation and growth of male spiny lobster (*Panulirus homarus*). *Malays. J. Sci.* **2020**, *39*, 30–40. [[CrossRef](#)]
50. Jayaprakas, V.; Sambhu, C. Growth characteristics of white prawn *Penaeus indicus* (Decapoda-Crustacea) under dietary administration of protein hormones. *Ind. J. Mar. Sci.* **1998**, *27*, 389–395.
51. Pinandoyo, D.H.; Windarto, S.; Herawati, V.E. The effects of addition thyroxin hormone on growth and the survival rate of giant prawn *Macrobrachium rosenbergii*. *Int. J. Fish. Aquat. Stud.* **2020**, *8*, 84–87.
52. Dvoretzky, A.G.; Dvoretzky, V.G. Aquaculture of green sea urchin in the Barents Sea: A brief review of Russian studies. *Rev. Aquac.* **2020**, *12*, 1280–1290. [[CrossRef](#)]
53. Syafaat, M.N.; Azra, M.N.; Waiho, K.; Fazhan, H.; Abol-Munafi, A.B.; Ishak, S.D.; Syahnon, M.; Ghazali, A.; Ma, H.; Ikhwanuddin, M. A Review of the nursery culture of mud crabs, genus *Scylla*: Current progress and future directions. *Animals* **2021**, *11*, 2034. [[CrossRef](#)]
54. Hachero-Cruzado, I.; Manchado, M. Dietary phospholipids enhance growth performance and modulate cold tolerance in meagre (*Argyrosomus regius*) juveniles. *Animals* **2021**, *11*, 2750. [[CrossRef](#)]

-
55. Subramoniam, T.; Kirubakaran, R. Endocrine regulation of vitellogenesis in lobsters. *J. Mar. Biol. Ass. India* **2010**, *52*, 229–236.
 56. Lezer, Y.; Aflalo, E.D.; Manor, R.; Sharabi, O.; Abilevich, L.K.; Sagi, A. On the safety of RNAi usage in aquaculture: The case of all-male prawn stocks generated through manipulation of the insulin-like androgenic gland hormone. *Aquaculture* **2015**, *435*, 157–166. [[CrossRef](#)]