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# Post-Mortem Dental Profile as a Powerful Tool in Animal Forensic Investigations—A Review

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Simple Summary: Teeth are the hardest anatomical structure of the animal body. As a result, even when preservation conditions are extremely poor and the rest of the skeleton decomposes, the dentition is often still preserved. The strong nature of the teeth means that they are often an invaluable biological source of information about a deceased animal. This is particularly important in forensic investigations resolving legal disputes involving animals and in circumstances where the animal body is recovered a long time after death. The post-mortem dental profile can contribute information such as species identification, sex, age-at-death, body size, geographical origin (provenance), and post-mortem interval. Although the dental profile may not lead to a positive identification, it can narrow the pool toward a presumptive identity. In this review, we briefly examine different dental techniques using characteristics of teeth as a means of identification of freshly deceased and skeletonised animals, highlighting the importance of dentition in the identification process in forensic contexts.

Abstract: Veterinary forensics is becoming more important in our society as a result of the growing demand for investigations related to crimes against animals or investigations of criminal deaths caused by animals. A veterinarian may participate as an expert witness or may be required to give forensic assistance, by providing knowledge of the specialty to establish a complete picture of the involvement of an animal and allowing the Courts to reach a verdict. By applying diverse dental profiling techniques, not only can species, sex, age-at-death, and body size of an animal be estimated, but also data about their geographical origin (provenance) and the post-mortem interval. This review concentrates on the dental techniques that use the characteristics of teeth as a means of identification of freshly deceased and skeletonised animals. Furthermore, this highlights the information that can be extracted about the animal from the post-mortem dental profile.

**Keywords:** veterinary forensics; carcasses; teeth; species identification; sex estimation; age-at-death estimation; body size estimation; geographical origin; post-mortem interval estimation; bite mark analysis



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#### 1. Animals in Forensic Sciences

In a broader sense, veterinary forensics can be defined as the application of veterinary science in resolving legal disputes involving animals (i.e., livestock, wild, exotic, and household animals) and animal derivatives [1]. Animals may be involved in two diverse ways: they may either be the victim of an assault or illegal act (i.e., maliciously or accidentally 'human-induced' injuries and/or insults to animals), or the perpetrator when the animal causes the incident (i.e., injuries caused to humans) [2]. This discipline is becoming more important in our society, increasing its frequency worldwide as a result of the growing

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demand for investigations related to crimes against animals or investigations of criminal deaths caused by animals [3]. The main fields in which a veterinarian may participate as an expert witness or may be required to provide forensic assistance are the following [4–6]:

- Assessment of animal welfare (includes survival-related factors: nutrition, environment, health; situation-related factor: behaviour; emotional-related factor: mental state). Includes giving an opinion as to whether an animal may be experiencing or has experienced pain, discomfort, or distress in the past. In some instances, this evaluation may incorporate experts from other scientific disciplines, such as nutritionists and animal behaviourists.
- 2. Determination of the time, the cause, and the circumstances of death of an animal, as well as other related investigations, such as the identification and interpretation of changes in the different body tissues, recognition of parasites and detection of signs of poisoning.
- 3. Verification of the history and provenance of live/dead animals and animal derivatives. Most of the cases are related to domestic species; however, other cases fall under the category of 'wildlife crime'. In this latter case, this field is related to whether national or international conservation legislation has been breached, such as the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES). This is a large field that includes offences and trapping of wild animals for sale, malicious poisoning, poaching, and the importation of endangered species or derivatives thereof.
- 4. Performing clinical or post-mortem examinations when animal abuse appears to be related to acts of violence towards humans.
- 5. Food safety. Ranges from animal welfare during slaughter (including religious slaughter) to meat inspection standards and the detection of contaminants, import, export, and correct identification of food.
- 6. *Human welfare*. Covers public health issues such as zoonotic infections, health and safety (both in the workplace and in care institutions), and personal injury cases (e.g., kicks, bites, venomous bites, scratches, etc.).
- 7. *Miscellaneous*. Miscellaneous cases relating to legal responsibility, negligence, nuisance, fraud, environmental pollution, damage to Crown property (e.g., the swans of Queen Elizabeth II in the UK), as well as other miscellaneous legislation and situations.

#### 1.1. The Animal as the Victim

There are many ways to inflict injuries and/or insults on animals maliciously or accidentally, including forms of physical, sexual, and psychological abuse [2,7]. Physical injuries may be the result of trauma events, excess heat or cold, immersion in water or other insults; these injuries are usually unintentional but may include non-accidental injuries. Injuries resulting from a sexual insult may be the result of attempted animal sexual abuse, or surgical or malicious damage of the urogenital region (including castration). These injuries can be the consequence of true sexual abuse or also due to normal veterinary/husbandry practices. The third insult, not legally accepted in some instances, is psychological in nature, which may be the result of taunting, teasing, or threatening an animal, or the deprivation of companionship or inappropriate social grouping. Recent behavioural studies conducted by forensic animal behaviourists document these forms of psychological effects [8], as well as the cultural conditions determining the manner of killing.

There have been extensive studies of some forms of 'human-induced' damage to animals; for example, non-accidental injury caused to dogs and cats [9,10], the effects of traps and snares [11,12], poisoning [13,14], and shooting on wildlife [12,15]. Death, injury, health concerns, pain, or distress may result from most of the examples cited above but the implications may differ depending on the species and the circumstances surrounding the attack [2,16]. Kellert and Felthous [17] proposed nine distinct motivations for animal cruelty when questioning groups of aggressive criminals, non-aggressive criminals, and non-criminals about their past experiences with animals:

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1. *To control an animal.* Excessive and cruel physical punishment may be employed to exert control or shape an animal's undesirable behaviours.

- 2. *To retaliate against an animal.* Extreme punishment or retaliation may be inflicted for suspected misbehaviour on the part of an animal.
- 3. To satisfy a prejudice against a species or breed. On numerous occasions, people designate certain groups of animals as good or bad. These beliefs may be related to cultural values, such as prejudice in our society against spiders or snakes.
- 4. *To express aggression through an animal.* Inculcating violent tendencies in the animal in order to express violent, aggressive behaviours toward other people or animals.
- 5. *To strengthen one's own aggressiveness*. Killing and abusing animals may be a way to enhance one's aggressive aptitudes or to impress others with the capacity for violence.
- 6. *To impress people for amusement.* Cruelty toward animals sometimes occurs as a means of creating amusement and 'entertaining' friends.
- 7. *To retaliate against another person.* Sometimes cruelty toward animals occurs as revenge against other people.
- 8. To divert hostility from a person to an animal. This deviated aggression usually involves authority figures whom the subject hates or fears but who they are afraid to aggress against. For example, in childhood, it is often easier to be violent to an animal than against a parent or adult
- 9. *Non-specific sadism*. This violence is related to the desire to inflict harm, suffering, or death on an animal in the absence of any particular provocation or especially hostile feelings toward an animal.

# 1.2. The Animal as the Perpetrator

The human–animal interaction dates back to prehistoric times, with animal domestication being practiced for thousands of years [18,19]. From ancient times, humans exploited animals for multiple reasons, generally having productive outcomes for humans, such as farming, obtaining food products (e.g., chickens as suppliers of eggs, pigs and poultry as meat sources), transportation of people or cargoes (e.g., mules for transport of supplies, horses as vehicles), recreation and entertainment (e.g., circus shows, horse racing), sports and hunting (e.g., sporting dogs) [20]. However, interactions between humans and animals can also be unproductive and unintentional, sometimes leading to injuries.

Animals may cause harm to humans or disrupt human activities in many ways, including: physical damage to property (e.g., cattle knocking down fences, birds destroying crops); causing noise (e.g., barking dogs, roosters crowing); producing unpleasant odours (e.g., a piggery or poultry house in the vicinity of a residential area); causing overwhelming fear (e.g., fear of spiders (arachnophobia) or dogs (cynophobia)); causing allergic reactions (e.g., hypersensitivity to animal derivatives, such as fur or feathers); and infecting humans with pathogens that cause a variety of zoonotic diseases (e.g., rabies, avian influenza) [2]. However, the continuous human–animal relationships in urban areas and the increasing human expansion into the animal's natural habitat, have led to an increased possibility of an animal attack on a human being causing serious injuries and even death [21].

The nature and incidence of animal attacks on humans varies between different regions of the world depending on the fauna present and the extent of the interactions between humans and animals. Many animals have been reported to attack and bite living humans, with most attacks being caused by the order of carnivorous mammals such as canids (e.g., dog, wolf; [22,23]), felids (e.g., cat, lion, tiger; [24,25]), ursids (e.g., brown bear, polar bear; [26,27]), and non-human Primates (e.g., chimpanzee, gorilla, macaque; [28,29]), as well as by ungulated mammals such as suids (e.g., domestic pig, wild boar; [30,31]) and hippopotamids (e.g., hippopotamus; [32]), by rodent mammals (e.g., rat, squirrel; [33,34]), by reptiles (e.g., crocodile, iguana; Komodo dragon; [35–37]), and even by sharks (e.g., white shark, tiger shark; [38–40]), among other animals.

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Similarly, animal scavenging is also relatively frequent in forensic investigations, where animal activity in outdoor settings is one of the main taphonomic agents that significantly affects the preservation of a human corpse when recovered from the area of deposition [41] (see Section 3.4. Bite Marks for more details on this topic). Thus, under varying conditions, any animal can attack a human corpse [42]. Generally, this is a scavenging process whereby the animal produces transportable units from the remains that can be moved to another place for later consumption [43,44], causing post-mortem modifications to a human corpse, altering characteristics of peri-mortem trauma, influencing decomposition rates, disarticulating and scattering body parts, mimicking or destroying actual forensic evidence, and affecting identification of the deceased [41].

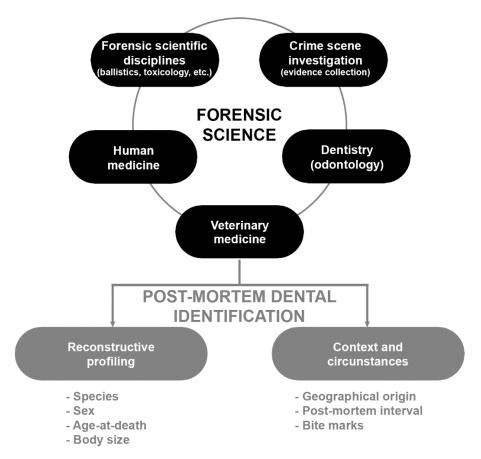
# 2. Role of the Veterinary Pathologist in Veterinary Forensic Investigations

The work of the forensic medical pathologist and the forensic veterinary pathologist is similar; however, there is an enormous difference: while the work of the former focuses on a single species (the human being), the work of the latter encompasses multiple species, with cases involving household animals (including exotic species), farm animals, and wild animals. In this way, multispecies forensic pathology makes it a complex and difficult discipline to manage [45]. The forensic veterinary pathologist is not only specifically concerned with the post-mortem examination of a deceased animal and documents the findings of the examination but is also involved in the collection of evidence and court proceedings.

In veterinary forensics, the identification of carcasses is of less importance compared to its counterpart in human forensic medicine, although the reliable identification of live animals can be crucial (e.g., in the resolution of criminal investigations where the animal is the causative agent of the injuries or death of a human being). However, when it is necessary to identify dead animals or their remains, the following methods can be used [2]: (i) external markings, colour patterns, etc.; (ii) external morphological features (e.g., shape of antlers, abnormal coloration, or wear of hooves); (iii) presence of external collars, chains, ear tags, and other human-introduced devices (e.g., transponders); (iv) surgical evidence (e.g., docked tail, prosthesis); and (v) osteological characteristics. In the latter case, the ultimate goal of analysing a set of skeletal remains is to estimate the biological profile (i.e., to establish a set of characteristics that an animal specimen possessed during their life), which can be used to determine identity after death. In veterinary science, the biological profile would include the taxonomic classification (i.e., class, order, family, genus, and species identification), sex, age-at-death, body size, health/disease status, and individualising characteristics [46].

The present review concentrates on the dental methods that use the characteristics of teeth as the means of identification of fresh deceased and skeletonised animals. The review is an attempt to highlight the importance of dentition in the identification process and its utility in estimating the biological profile and to show other information that can be extracted about the animal from the post-mortem dental profile. Figure 1 illustrates the integration of veterinary medicine within the forensic sciences, summarizing the main applications of dental profile in veterinary forensics.

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**Figure 1.** Diagram of the integration of veterinary medicine within the forensic sciences summarizing the main applications of dental profile in veterinary forensics.

# 3. Teeth as a Biological Source for Forensic Identification in Animal Remains

The distinct anatomy of the dentition and its resistance to decomposition makes it an invaluable source for biological studies and enables us to understand ancient and modern animal communities. Examination of the dentition is widely used by zooarchaeologists to identify animal skeletal remains [47,48], but it is also important in post-mortem forensic work. It is reported that the nineteenth-century French naturalist and zoologist Georges Cuvier, who established the sciences of comparative anatomy and palaeontology, said: 'Show me your teeth and I will tell you who you are' (translated from French; [49]). Animals' teeth are so varied and distinctive that they can be used to identify animal remains by veterinary forensics based on a single tooth.

# 3.1. Biological Profile

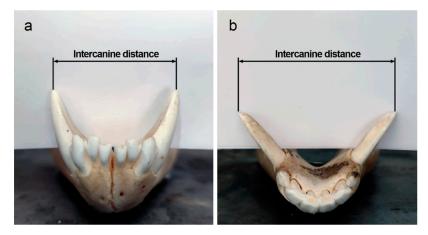
# 3.1.1. Species Identification

The comparative dental anatomy analysis is a classical technique for species identification, and it also correlates to the inter-species relationship among members of the same family (e.g., family of Felidae: includes cheetah, leopard, tiger, domestic cat, lynx, among others) [50,51]. The number and types of teeth present in the oral cavity is useful in genus identification. The number of teeth of each type, present in one maxillary or mandibular hemiarch, is referred to as the *dental formula*. For example, the dental formula for the permanent dentition of the family of Canidae (i.e., all dogs, wolves, foxes, and coyotes) is I3/3:C1/1:P4/4:M2/3, meaning that they have three incisors in both the maxilla and mandible (I3/3), one canine (C1/1), and four premolars (P4/4) in each upper and lower hemiarch, and two molars in the maxilla and three molars in the mandible (M2/3). This dental formula is different from a cow or goat, which is I0/3:C0/1:P3/3:M3/3, indicating that they have no incisors or canines in the maxillary arch. Looking at closely related

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species, the domestic cat has a dental formula of I3/3:C1/1:P3/2:P1/1, while the lynx has a dental formula of I3/3:C1/1:P2/2:M1/1 [52]. Eventually, the number of teeth may vary from the expected dental formula. In these situations, it is important to record which teeth are missing and why, as any deviation in the number of teeth from the dental formula must be considered, such as for example, genetic causes, ante-mortem tooth loss due to disease or trauma, post-mortem tooth loss, or unerupted or undeveloped tooth (agenesia).

Species identification or the distinction of closely related species can also be done using the metric and morphological characteristics of the teeth, applying statistically robust techniques and using advanced tools (e.g., geometric morphometrics) [51,53-59]. Furthermore, the variation of simple metric characteristics such as tooth size or jaw length can be key in resolving debates about whether a sample comprises a single species or includes more than one morphologically similar species [60]. For example, a high coefficient of variation in a dental sample can be an indication that corresponds to more than one species [61]. However, according to Hillson [59], the absolute size variation of individual teeth is less marked than the relative size variation between different elements of the dentition, so it is important to analyse several classes of teeth at the same time. Of particular importance is the *intercanine distance*, defined as the length between the two tips of the maxillary or mandibular canines (Figure 2). While the shape of the maxillary/mandibular dental arches can help differentiate between mammalian families (it is not possible to distinguish between members of the same family by the shape of their jaws alone), intercanine distance can help differentiate between species of varied sizes in the same family [62] (see Section 3.4.2. Human Deaths from Animal Bites for more information on this topic).



**Figure 2.** Intercanine distance as measured at the mandibular canine cusp tips in (**a**) *Canis lupus* and (**b**) *Sus scrofa.* Images courtesy of C. Tanga.

Non-metric dental traits (e.g., presence and size of cusps, form of fissures on occlusal surfaces of premolars and molars, form of ridges, presence of pits) also play a significant role in species identification. Some non-metric traits are normally scored as presence/absence or graded into categories defined by a set of rules [52]. In any case, the variation of these non-metric traits is used to distinguish between species [56]. For example, the pattern of folds exposed on the occlusal surface of persistently equine mandibular cheek teeth varies, this being one of the ways to distinguish different species of horse and donkey [63], although their reliability is currently being questioned [56].

In short, dental form (size + shape) is highly genetically controlled and well reflects phylogenetic relationships, making teeth useful to identify the taxonomy of animals [64]. Thus, species identification is based primarily on macroscopic inspection of dental form (e.g., [65]). In recent years, more complex tools (e.g., geometric morphometrics) and statistical procedures (e.g., machine learning algorithms, artificial intelligence) have allowed to analyse teeth and tooth marks with a higher precision [66–68]. However, when teeth are in a poor state of preservation, these traditional or advanced methods could be severely limited due to the difficulty or impossibility of observing

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species-specific dental anatomical characteristics. In this situation, histomorphometry of dental tissues (i.e., evaluating the organisation, composition, and structural components of enamel, dentine, and cementum) [52], immunological procedures [69], stable isotopes [70], and genetic tools (such as DNA sequencing, Single Nucleotide Polymorphism, Polymerase Chain Reaction–Restriction Fragment Length Polymorphism, and microsatellite analysis) [71,72] have become particularly useful and relatively applicative. However, all these methods require complex, time-consuming, and highly professional procedures. For these reasons, fast, accurate, and easy-to-use methods and techniques have been developed in recent years to identify the species origin from teeth samples. Non-destructive analytical chemistry (i.e., spectroscopy techniques) are constantly evolving and they are widely used in forensic science and practice. Thus, X-ray fluorescence [73–75] and Fourier transform infrared spectroscopy [76,77] can provide useful information on the elemental components of teeth to identify the species of animal remains. The main advantages of these spectroscopy techniques are that they provide fast and accurate results and do not require complex analytical procedures.

The investigation of criminal deaths caused by animals has increased considerably in recent decades, so that determining whether the death of a human being was caused by a domestic or wild animal (and its taxonomic classification) is a step of vital importance in forensic settings to determine legal responsibilities [22,62,78,79].

#### 3.1.2. Sex Estimation

Sexual dimorphism is the term that refers to differences between males and females of the same species [80]. This condition is common among mammals, but the levels of dimorphism vary between them, being generally higher in large mammals than in small mammals [52]. Sex is easily indicated by the presence/absence of the *baculum/baubellum* [81], but most frequently sexual dimorphism is identified by body measurements, particularly visible in body mass and size [82]. Sexual dimorphism is strongly present in those species with polygynous social ecology (i.e., a mating system in which one male lives and mates with multiple females but each female only mates with a single male), reflecting increased male—male competition for access to breeding females [83], rather than related to diet, habitat, or activity patterns [84].

Size-related sexual dimorphism is a common phenomenon in carnivores, particularly in the size of the skull, mandible, and teeth, with males on average being significantly larger than females [84–88], except in some animal species such as the spotted hyaena (*Crocuta crocuta*), where a reverse sexual dimorphism is observed [89]. In this order of mammals, sexual dimorphism in the size of the skull, canines, carnassial teeth, and molars is widespread, being more pronounced in the families of Felidae (e.g., [90,91]), Canidae (e.g., [82,92]), and Ursidae (e.g., [93,94]). These taxa contrast with other mammalian groups such as Primates, where diurnal species are generally more sexually dimorphic than nocturnal species, and terrestrial species tend to be more sexually dimorphic than arboreal species [91,95,96].

In general, dental sexual dimorphism of Primates centres on the canines [95,97–100] and, combined with the rest of the teeth in a discriminant analysis, can be used to assign a sex correctly in skeletal remains. It has been suggested that the sexual dimorphism in the dentition, centred on the canine teeth, is related to the so-called *field effect*, in which teeth closer to the canines tend to be more sexually dimorphic than those further away [101–103]. Among Primates, sexual dimorphism in the size of canine teeth ranges from minimal to extreme levels (e.g., lack of sexual dimorphism in *Aotus*, *Callicebus*, *Saguinus*; moderate in *Pan* and *Gorilla* [ca. 25–40%]; extreme in *Papio*, *Mandrillus*, and *Theropithecus* taxa (ca. 69–75%)) [98,104]; data of sexual dimorphism calculated from [105]). Humans fall at the low end of the range of Primate canine dimorphism, with male canine teeth that are up to 10% larger than those of females [98]. Dental sexual dimorphism is also marked in tusks, including marine mammals such as narwhals, walruses, and dugongs, and herbivorous terrestrial mammals such as elephants and hippopotami [52].

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Thus, in the resolution of a forensic investigation case, sex estimation is one of the first steps in reconstructing the biological profile of an animal that must be performed. The main reason for this is the fact that other vital information, such as age-at-death and body size, cannot be adequately obtained without prior sex estimation.

# 3.1.3. Age-at-Death Estimation

In the forensic context, age-at-death estimation (i.e., the amount of time that has passed from the birth to the given date of death) is one of parameters of the biological profile crucial to establish the identity of the animal [106]. The ageing of animals can be particularly important in disputes over purchases [107], control of hunting [108], violation of laws about animal conservation [109], age of abandonment or adoption [110,111], trade and imports [112], and deliberate acts of violence against animals [109], among others.

Age-at-death estimation can be applied to living animals or skeletonised remains [113]. The examination of bones, horns, and dentition has been proposed in ageing of carcasses, as well as the length or height of animal and the colour of the pelage [106]. However, the study of animal dentition is one of the most practical and accurate methods for estimating their age-at-death [114]. Dental age-at-death estimation has been widely accomplished in human forensic investigations and wildlife animal research [111]. The anatomical processes of development occur in the same way in humans and animals [115]; therefore, dental age-at-death estimation methods are focused on changes in growth and development of teeth, as well as the changes after their formation [116]. Several methods have been proposed for the estimation of dental age-at-death in animal forensic investigations, such as those based on (i) dental development and eruption, (ii) occlusal tooth wear, (iii) dental cementum annuli, and (iv) secondary dentine deposition.

Since dental growth and mineralisation follow a consistent sequence and clear-cut changes occur over a brief period, age-at-death can be estimated with reasonable reliability from the state of development [52]. In veterinary practice, age-at-death can be estimated by visual examination evaluating dental eruption, since the sequence and timing of the eruption of teeth provides a reference scale for age-at-death estimation; it can be studied since the tooth begins the process when the crown emerges from the crypt until it reaches the occlusal plane [117]. When estimating age-at-death through dental development, it is necessary to consider the different patterns and types of growth variation within species, taking into account: (i) the basic types of tooth development (i.e., monophyodont, diphyodont, or polyphyodont); (ii) the types of shape of dentition (i.e., homodont or heterodont); (iii) the anchorage of teeth (i.e., thecodont, acrodont, or pleurodont); (iv) the basic types of tooth crown (i.e., brachyodont or hypsodont); and (v) the basic types of jaw occlusal overlay (i.e., isognathus or anisognathus) [118]. For example, in many mammal species the tooth continues to grow after the main eruptive phase, which causes occlusion of the teeth and finishes with the closure of the root apex; for other mammals, the crown or root continues to grow throughout life and, in still other mammals, the root can be completed soon after the initial eruption [52]. The application of this method of age-at-death based on the evaluation of dental eruption is applied in humans only in children or young individuals; this situation must be considered in the case of some animals, for example the elephant, whose molars have an eruption sequence of 30 to 40 years [119]. Descriptive stages for dental development and eruption in relation to chronological age have been proposed in many animals of the order of carnivorous mammals such as canids (e.g., dog; [112,120,121]), felids (e.g., cat; [120]), and non-human Primates (e.g., lemur, galago, chimpanzee, gorilla, macaque; [122-125]), as well as ungulated mammals such as caprines (e.g., sheep, goat; [120,126,127]), bovids (e.g., cattle; [128]), and suids (e.g., pig; [120,129]), among others.

After the dentition is fully erupted, several researchers have proposed age-at-death estimation methods based on dental wear [117]. Once a tooth emerges from the gingivae, dental wear initiates as a consequence of the grinding of teeth against one another, and the contact with food, cheeks, and tongue [52]. Dental attrition of the permanent teeth has been extensively studied and is considered a classic method for age-at-death estimation in

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adult animals [119], visually assessing the loss of enamel and the amount of the dentine exposed [52]. The most used protocols for recording dental attrition are based on changes across the occlusal surface of premolars and molars, such as those developed by Payne [130] (a system based on sheep and goat remains) and Grant [131] (for cattle, sheep, goat, and pigs). Since the 1970s, an alternative approach has been proposed based on the measurement of crown height to evaluate its decrease through age (e.g., [132] in zebra, [133] in horse, [134] in red deer). Despite being a widely used method, it presents certain limitations [126], such as: (i) the assumption that there is a degree of correspondence between the tooth wear classes and the chronological age of the animal at death, (ii) the assumption that age ranges are of equal length, and (iii) the discrepancy of applying methods developed in ancient animal populations to contemporary animal populations or vice versa (i.e., to apply an age-at-death estimation method developed in an animal population of a specific chronology to another population of a different chronology).

Another age-at-death estimation method is based on the analysis of incremental structures in dental cementum [117]. The deposition of cementum is continuous throughout the life of the animal, providing a longitudinal record of factors affecting its growth, resulting in incremental bands correlated with seasonal growth in most species [135]. Each annual period of deposition is composed of a 'summer' layer (i.e., a wide and translucent layer that corresponds to the growth line) and a 'winter' layer (i.e., a narrow and hypermineralised layer that corresponds to the rest line) [136]. When longitudinal tooth sections are observed under a light-transmitting microscope using polarised light, translucent and opaque bands alternate as a result of the growth pattern; so, these bands can be related to the age of the animal and used to conduct the estimation of the age-at-death [135]. However, the regularity of deposition can be influenced by external factors (such as climatic variation or quality of food), but also by internal factors (such as feeding and reproductive habits) [136]. Thus, counting of dental cementum annuli is a very reliable and accurate method to provide estimates of age-at-death for seasonal animals, such as moose [136,137], red deer [136,138], black bear [139], polar bear [140,141], harbour seal [142], ringed seal [143], American badger [144], and feral pig [145], among others.

As in human dentition, the study of secondary dentine deposition inside the pulp chamber is also applied for age-at-death estimation in animals [52]. Secondary dentine is the dental tissue formed after root completion and its deposition is continuous inside the pulp cavity in the form of layers while the pulp remains vital. As a result, the pulp cavity reduces in volume with age [118]. The relationship between the pulp/tooth area ratio using dental radiographic images is the basis of this age-at-death estimation method and has been applied in several animal species such as cat [111], dog [146], coyote [147], and lion [148].

#### 3.1.4. Body Size Estimation

Body size is described in terms of body length or mass, since these two variables provide the greatest predictive value for understanding the animal's ecology [149]. For example, among living mammals, body size is usually described in terms of mass; however, when the animal is too large to be easily weighed or it is difficult to directly measure this variable in the field, withers height and body length are mainly used (e.g., whales) [150], although it has also been used for other domestic and small mammals [151–153].

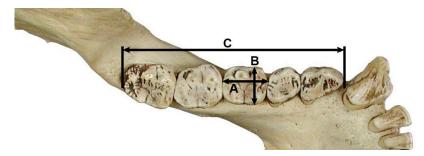
Limb-based estimations of body mass are the most common methods using either lengths [154,155] and/or midshaft cross-sectional dimensions [154,156] of long bones. They have the advantage that they are based on the relationship between body mass and the load borne by the limbs when they support the body on the ground [149]. However, because teeth are most frequently preserved in the skeletal record, their size is often used to estimate the body mass by biologists and palaeontologists [157–159].

For example, current approaches to body size estimation in mammals stem from Gould's paper [160] in which he proposed that the area of mammalian post-canine occlusal tooth area scales against body mass with positive allometry as a result of metabolism

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and/or or changes in dietary strategies (i.e., tooth size in mammals scale in a predictable manner to body mass to meet their metabolic needs) [157,161]. While several studies use the post-canine tooth row length to infer allometric relationships with body mass [157,162–164], other researchers have proven a strong relationship between body mass and the area of individual teeth, particularly the first molar (e.g., [103,165–172]). The first molar is the most useful tooth for this purpose because it tends to have relatively low levels of intraspecific variation and sexual dimorphism compared with the other tooth classes, probably as a consequence of its early eruption in the oral cavity during ontogeny [165,168,173,174]. Thus, inferring body mass from individual teeth adds to the metabolic scale assumption by assuming a relationship between the size of that tooth and that of the rest of the tooth row [149].

Numerous studies have performed regression equations based on post-canine tooth row length and/or mandibular first molar crown area (i.e., crown area = mesiodistal × buccolingual diameter) and have been developed for a variety of species of the class Mammalia, including ungulates (e.g., [158,159,172]), marsupials (e.g., [168]), carnivores (e.g., [163,171,174]), rodents (e.g., [164,175,176]), Primates (e.g., [157,165,177,178]), and even sharks (e.g., [179]) (Figure 3). Although the first molar is considered the tooth that has the least variation in its adjustment to body mass and, therefore, would be the ideal tooth to estimate body mass from a single tooth, regression equations are available for the other tooth classes of the dentition (e.g., [165,168,174,178]).



**Figure 3.** Example of three dental dimensions commonly used for estimating body mass using regression methods, illustrated on the left hemimandible of *Gorilla gorilla*. A = mesiodistal diameter of first molar; B = buccolingual diameter of first molar; C = mandibular post-canine tooth row length.

For example, in forensic settings, the examination of dental features and characteristics of a bite mark may help identify the animal that caused the biting injury (see Section 3.4. Bite Marks for more information on this topic). Useful clues for classification of the aggressor type include the estimation of the size of the mouth, calculated measuring the intercanine distance. This parameter can help to estimate the animal's breed and body size and can also help to distinguish between different sized species in the same family [62].

## 3.2. Geographical Origin (Provenance) Identification

An understanding of patterns of movement is fundamental for the knowledge of the ecological, life history, and behaviour of the majority of animals, which requires that specimens be tracked across time and space. Tracking animal movements can be done directly using some type of extrinsic marker (e.g., radio and satellite transmitters) or indirectly using intrinsic natural biological (e.g., fatty acid profiles, DNA techniques) or biogeochemical markers (e.g., concentration of trace elements, stable isotopes) [180,181]. Stable isotope techniques are the most widely used to link an animal to a specific geographical region and are based on the principle 'you are what you eat' (i.e., isotopic signatures of foods consumed are recorded in consumer tissues) [182].

Stable isotope ratios vary among biomes that animals inhabit and are incorporated into organism tissues from its diet. In this way, animals moving between isotopically different biomes can retain information of previous feeding locations for periods of time that depend on the turnover rates for the different organism tissues [181]. Keratin-based

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tissues (e.g., hair, feather, nail, claw, bill) are metabolically inert after synthesis, maintaining an isotopic record reflecting the location where the tissue was synthesised, so they are usually used to study seasonal movements. Conversely, metabolically active tissues' dietary and source information will correspond to a temporal integration, ranging from a few days or even hours (e.g., urine, faeces, blood plasma) [183–186] to several weeks (e.g., muscle, whole blood) [187–189] or even years (e.g., bone collagen) [190–192] (Table 1). Thus, investigations examining long-term movements use metabolically inert tissues, whereas studies on recent movements use metabolically active tissues with rapid turnover rates.

**Table 1.** Time periods represented by isotopic ratios in different body fluids and soft and hard tissues of animals.

Body Fluids and Tissues	Time Period Represented					
	Hours	Days	Weeks	Months	Years	
Urine	✓	<b>√</b>				
Faeces	$\checkmark$	$\checkmark$				
Blood plasma	$\checkmark$	$\checkmark$				
Blood cells		$\checkmark$	$\checkmark$			
Hair		$\checkmark$	$\checkmark$	$\checkmark$	<b>√</b> *	
Nail		$\checkmark$	$\checkmark$	$\checkmark$	<b>√</b> *	
Claw		$\checkmark$	$\checkmark$	$\checkmark$	<b>√</b> *	
Bill		$\checkmark$	$\checkmark$	$\checkmark$		
Feather		$\checkmark$	$\checkmark$	$\checkmark$		
Antler		$\checkmark$	$\checkmark$	$\checkmark$		
Muscle		$\checkmark$	$\checkmark$	$\checkmark$		
Bone (collagen)					$\checkmark$	
Tooth (dentine, enamel)					$\checkmark$	

<sup>\*</sup> Depending on their lengths, hair, nails, and claws can record incremental records of several years.

In the case of teeth, stable isotope analysis can be performed on either the organic or inorganic fraction. The organic fraction preserves proteins such as collagen, so the collagen contained in dentine can be used to assess short-term changes that occurred during puppyhood, as these tissues form in early life and undergo little remodelling [193]. The inorganic fraction is primarily formed by hydroxyapatite. Dental enamel, like bone, is mainly composed of hydroxyapatite; nevertheless, unlike bone (a relatively porous material composed of tiny hydroxyapatite crystals interspersed with approximately 30% of organic matter), dental enamel is essentially non-porous, composed of relatively large crystals including only minor amounts (ca. 2% or less) of organic matter [194]. Therefore, the dense crystalline structure of enamel makes it the preferred tissue for isotopic analysis, as it is less susceptible to diagenetic alterations compared to bone tissue [194,195]. Furthermore, dental enamel, unlike bone, is not remodelled during life, and therefore the isotopic signature of dental enamel is directly related to the environment and diet during the period of tooth formation [195].

In recent decades there has been an increase in the use of stable isotope analysis in wildlife and criminal forensic contexts as a means of determining the origin and movement of animals [196–199], with applications as varied as establishing the origins of ivory from elephant tusks (e.g., [200–203]), or the illegal animal trade or animal derivatives. An example of the latter case is the increase in seizures of jaguar (*Panthera onca*) body parts (e.g., skin, fat, meat, claws, tails, skulls, bones, teeth), which have been occurring since 2013 in different Latin American countries. This increase in seizures is linked to the high demand in Chinese markets for body parts of these felines to satisfy the demands of traditional Asian medicine [204,205]. In the particular case of teeth, these are used locally for decorative (e.g., jewellery and key chains), medicinal (e.g., therapeutic treatment for facial paralysis caused by a spell of misfortune; dental fillings), or for cultural purposes (e.g., necklaces used in traditional festivals; amulets to protect against bad luck

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or evil spirits) [206]. The application of stable isotope analysis in this context is considered a useful tool in countering wildlife trafficking efforts [207].

# 3.3. Post-Mortem Interval Estimation

The post-mortem interval (also known as the *time since death*) is the time between the death of an animal and the discovery of the body [208]. Knowing the time since death is essential in the investigation of human deaths, as well as in animal forensic investigations [209,210]. Various reasons have been proposed for estimating the post-mortem interval in animals: (i) inclusion or exclusion of individuals from a group of suspects and corroborating witness testimony [209]; (ii) differentiation of single or continuous episodes with animals [208,211]; (iii) identification of incidents of hunting out of season, poaching, negligent transport of animals or abuse of companion animals [208,211]; and (iv) application of legal deadlines (e.g., disposal of animal carcasses) [208,211]. In human forensic medicine, the study of the post-mortem interval is one of the most popular topics; however, in veterinary forensics, the number of studies is extremely limited [208–211]. Researchers must face a deficiency in the development of methodologies for a large number of species and, therefore, the obligation to apply methods developed in humans, lacking the appropriate validation to be applied in crimes against animals [209,210,212].

The most used methods of relevance to forensic veterinary pathology for estimating the post-mortem interval in animals' dead bodies are mainly based on temperature changes, muscular stiffening (also called *rigor mortis*), ocular changes, cadaveric lividity (*livor mortis*), decomposition processes, and entomology [210,212-214]. In the case of studies conducted on animal dentition, there is a limited amount of research based on morphological, histological, or molecular analysis [215]. For example, Akbulut et al. [216] analysed the relationship between the changes in the mineral density of enamel, the surface abrasion of hard dental tissues, and the estimation of time of death through micro-CT. The results of this study realised with rats' dentition showed that morphological changes in the microstructure of dental tissue can be considered a parameter for estimating the post-mortem interval in a forensic context. In the same way, a study by Granrud and Dabb [217], based on the exfoliation of the anterior dentition of pig teeth, showed a potential relationship with the post-mortem interval. Thus, an accumulated degree-days can be used to establish a minimum length of time since death; however, the authors indicated that future research is necessary. Mehendiratta et al. [218], on their part, observed a series of morphological and histological changes by analysing the dental pulp, showing that this approach could be applied in the early phases since after 144 h, dental pulp is not preserved (i.e., it is completely decomposed). Finally, Young et al. [219], analysing the dentition of buried pigs, studied the progressive changes in pulp coloration due to decomposition and the post-mortem pulp RNA degradation. The morphological and molecular evaluation showed a favourable estimation of the post-mortem interval through the differences in RNA decay and progressive colour changes. The authors highlighted the advantages of this method for its speed of application and low cost.

The small number and the results of the studies conducted on animal dentition for estimation of the post-mortem interval show the need to increase the analysis on this topic. All the studies point out the potential of their methods but emphasise the need for further research to give greater solidity to the results [216–219]. In the same way, they indicate the importance of developing studies on taphonomic factors to improve the precision of the post-mortem interval estimation. The choice of the study animal (mostly pig) represents the objective of application in human forensic cases. The majority choice of the pig is probably due to its similarity as an animal proxy to the human and not because of the need to study it as the animal victim or perpetrator. Similarly, the introduction and discussion of the studies reviewed are contextualised within human forensic activity and not in the animal context.

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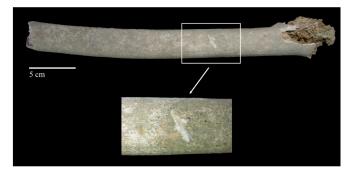
#### 3.4. Bite Marks

In forensic sciences, recognising and correctly identifying the actions of animals on human remains, but also on other faunal remains, is crucial, as this allows the collection of data about events that may have affected the body over a time, which may have ranged from the ante-mortem to post-mortem period [220]. In certain contexts or situations, animals can cause severe injuries that, on one hand, may lead to the death of the individual attacked and, on the other hand, can alter the corpse in the post-mortem period, either in relation to soft or hard tissues [221]. To reconstruct the forensic scene as reliably as possible and define how certain animal species acted on a human body, it is essential to correctly identify the nature of the injuries, the anatomical region affected, the circumstances in which they occurred and the agent that caused them, in order to avoid possible misjudgements with very disastrous implications in the forensic framework. At a macroscopic level, bite marks are among the signs most frequently found on the body of a victim, whether it is exposed in an open, outdoor, or enclosed environment. Bite marks can be defined as both superficial and deep marks left by teeth that affect, in diverse ways, both soft and hard tissues whose morphology varies depending on the size and shape of the maxillary/mandibular dental arches and the force exerted by the bite [222]. According to Binford's standards [223], four basic types of tooth marking can be recognised by the motion of animal teeth over the surface of bones: (i) punctures, (ii) pits, (iii) scores, and (iv) furrows.

Punctures are those regular-shaped marks left clearly and evidently at the point where the bone has collapsed under the pressure exercised by the teeth (e.g., canines). Since perforations are a direct consequence of the force that the animal's bite has exerted on the bone, the specific types of shapes and sizes will depend directly on the species involved (e.g., large and deep tooth marks could be due to the action of large carnivores such as canids or felids). However, puncture marks are not always circumscribed and well-defined and, in some cases, it is possible to find puncture marks associated with gnawing marks on the bone after the consumption of the soft tissue [223].

Gnawing generally proceeds from soft to hard bone; the animal first attacks the soft spongy parts of the bone and only later encounters progressively harder bone (i.e., compact bone), where pitting can occur. That is, the same actions of gnawing may result in pitting since the compact bone is strong enough to not collapse under the action of gnawing. According to Binford [223], when pitting is present rather than puncture marks, this indicates a prolonged gnawing action on the bone that cannot be attributed to either consumption or extraction of the meat from the skeleton, as is the case of killing.

Scoring is the result of either turning the bone against the teeth or dragging the teeth across relatively compact bone. The result is a scarring of the surface with close, linear morphological characteristics. In the case of tooth scoring, it is important to examine their characteristics carefully so as not to make the mistake of interpreting them as marks caused by different classes of bladed weapons [223] (Figure 4).



**Figure 4.** Human skeletal remains from the late medieval cemetery of Corfinio (12th–13th century CE, Italy). Anterior view of the diaphysis of the left femur from an adult individual showing a large linear mark that could be confused with a mark caused by a canine tooth dragging across compact bone; however, this large mark was caused by a sharp tool during a deficient excavation process (i.e., lesion of post-mortem origin). Image courtesy of C. Tanga from her bachelor's thesis [224].

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### 3.4.1. Animal Scavenging

Animal scavenging is one of the main post-mortem taphonomic processes responsible for the modification of a corpse, thus having a significant implication in forensic casework, especially in cases where the remains are deposited in an outdoor environment [225,226]. Identifying the nature of injuries made by domestic and/or wild animals on human bodies in which soft tissue is still present at the time of scavenging or on already skeletonised remains is still quite complex. In some cases, these can be misinterpreted as traumatic injuries caused by sharp weapons and gunshots, with signs of interpersonal violence that occurred ante-mortem or peri-mortem and with additional taphonomic variables [227]. Since the bite marks left by scavenging animals can lead to difficulties in the interpretation of the forensic investigation and consequently, also compromise the identification of a perpetrator and the interpretation of the cause of death, it is crucial to be able to recognise them, considering how the nature of the bite marks vary depending on the species involved, how behavioural patterns of predation vary at local and regional levels, as well as the dispersion or alteration of human remains induced by these scavengers [228–230].

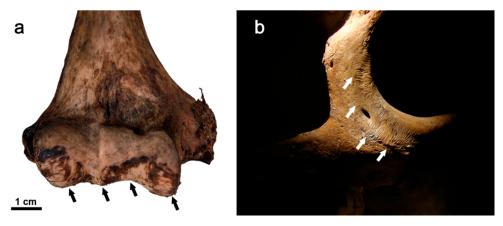
Generally, depending on the animal species involved in scavenging, both superficial injuries affecting only soft tissue and situations of disarticulation, dismemberment, and dispersal of body parts can be recorded, further complicating the recovery process and identification of human remains [231]. Therefore, knowledge of the regional fauna by veterinarians, biologists and forensic odontologists not only provides a better understanding of which animals feed on the carrion, but also of the reasons why they consume corpses. The involvement of these categories of experts in forensic investigations is proving to be increasingly necessary in the field of forensic activities since in complex situations it prevent errors when investigating bite marks caused by a possible animal attack on humans.

Although sometimes it is complex to identify the scavenger species that acted directly on the body, specialists can make use of some standard procedures beginning with macroscopic and direct observation of visible marks. This is followed by an assessment of the environment and geographical context of the finds, the pattern of scavenging on a body or skeletal element, the type and size of teeth marks, and finally, the additional faunal evidence associated with the forensic scene such as excrement, hides, or other organic elements. In more complex cases, molecular genetic analyses may also be performed to identify or exclude the perpetrator of certain injuries [232]. Therefore, in addition to the types of teeth marks, it is equally important to be aware of the different species of scavengers present around the forensic investigation as each of these leaves a different bite mark on soft tissues and bones. The main groups of scavengers that leave visible traces on human bones include (i) carnivores (e.g., canids, felids, and ursids), (ii) ungulates, (iii) rodents, and to a lesser extent (iv) birds. The mechanism of scavenging and the marks left on the soft tissues and bones between these species are quite different [233]. Large canids and felids (e.g., dogs, foxes, coyotes, wolves, lions, lynxes, cougars) are among the major carnivores on the planet capable of attacking or killing a human being, as well as being responsible for scavenging. In cases of displacement, it is important for forensic investigators to understand the behavioural patterns and patterns of animals that move locally; in fact, dismemberment and dispersion may occur differently depending on the species, but also on climate, season, and body size of the animal [234]. These variables must always be considered in a forensic investigation, as knowing how the local fauna acts in certain situations and how it approaches the remains are crucial, especially when recovering remains that are scattered over an area that may vary from a few to tens of metres [235].

Although the modalities and patterns of animal scavenging vary depending on the species involved, the behavioural pattern of predation and the availability of access to the body (e.g., open/enclosed environment, clothing, concealment by vegetation), the main cause of scavenging is the consumption of soft tissue and bone for nutritional purposes [236], which in some cases may be scarce in the ecosystems in which they live [237]. From a behavioural point of view, large carnivores first consume the soft tissues of a body

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from the thoracic cavity and only later turn their attention to the limb bones, leaving clear signs that can be traced back to both the action determined by claws and teeth [238]. Generally, the bite marks of canids and felids (puncture marks; see Section 3.4. Bite Marks) are recognisable in comparison to those of other animal species because the canines of these two families gnaw soft tissue or chew long bones to extract meat, fat, and bone marrow, leaving typical lesions in the form of pits and ovoid holes [239]. Like carnivores, rodents are also often found interacting with human remains in forensic contexts, leading in some cases to the dispersal of small bones, such as those of the hands and feet [240,241]. Rodent activity can be recognised by the presence of specific imprints known as gnawing marks ([223]; see Section 3.4. Bite Marks). Rodent bites are typically found post-mortem; these produce osseous changes through the gnawing action exerted by the upper and lower incisors constantly moving over skeletal remains (Figure 5). Unlike the bite marks of canids, those of rodents are distinguishable in that they do not usually affect soft tissue or leave claw marks but are mostly concentrated around the diaphysis of bones [226]; bone tissue, in addition to being a food source of calcium, is also an efficient wear tool for rodents to file away the continuing growth of incisors. Although not as significant, some species of herbivores, including ungulates (e.g., cervids), also constitute a taphonomic agent regarding the modification of human remains; these, in fact, in an open environment may proceed to consume bones due to a lack of phosphorus in nature. Teething, in this case, leaves a visible imprint in the form of erosion or exposure of the spongy tissue of the bone, sometimes associated with a series of parallel grooves smaller in size than those left by rodents. In long bones, the chewing of proximal or distal epiphyses by ungulates results in the formation of impressions whose morphology is termed *fork-shaped* [242–244].



**Figure 5.** Rodent modifications on human bones in the form of parallel striations from a forensic casework (indicated by arrows). Rodent gnawing signs of post-mortem origin (a) on the distal epiphysis of a right humerus and (b) on infraorbital and lateral margins of the right zygomatic bone. Images courtesy of J. Viciano.

# 3.4.2. Human Deaths from Animal Bites

In forensic contexts, there are frequent cases of death resulting from a violent predatory attack by certain animal species. Compared to the number of attacks by domestic animals on humans, those involving wild animals are much rarer and usually involve deceased humans [245] (see Section 3.4.1. Animal Scavenging). However, when tragic events of this type occur, there are several factors to take into consideration to understand the circumstances of the attack, the identity of the animal involved, the behavioural pattern in terms of both predation and feeding and finally, the consequences for the victim and their ante-mortem reactions [246]. In this way, in the case of a victim with physical evidence on the body of direct involvement of an animal predator, careful documentation and analysis of the anatomical region fatally involved, the position of the victim's body at the time of discovery, the age of the victim and the level of violence of the attack need to be noted; the size and morphology of the dentition involved, especially when there are

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clearly visible bite marks in soft tissue and bone, is helpful in correctly identifying the animal species responsible for the attack [247]. The analysis of morphological and metric characteristics of bite marks by veterinarians, biologists, and forensic odontologists can be particularly useful when trying to determine or discard the species, size, sex, and age of the biter involved in the attack [248]. Among domestic animals, dogs, especially in group situations, are the most frequently responsible for aggression against humans with often fatal consequences [249,250]. Injuries caused by a bite can vary from mild with non-fatal consequences, to profound with the consequent death of the victim, also including deep post-mortem lacerations and tears of soft tissue of the victim's body, caused by the action of canine teeth [251–253].

Thus, the analysis of bite marks in forensic contexts may provide sufficient scientific evidence that would help identify or exclude the animal perpetrator. This is especially useful when different animal species are involved in the area where the corpse was found, whose behavioural and injury patterns on the victim's body may be similar and whose misinterpretation may lead to errors and incorrect interpretations in the determination of the cause of death and the ante-mortem activities of the animal, often with serious consequences in the resolution of legal cases. An interesting case in this regard is the case report published by Fonseca and Palacios [22] concerning a case of animal predation on a male victim found in Argentina. The two experts, one an expert in forensic odontology and the other in animal biology, were involved by the competent judicial authorities to determine the nature of the injuries found on the victim's remains and to establish the animal species involved in the attack that turned out to be fatal for the victim. The multidisciplinary investigation aimed to obtain additional information on the manner of the attack and the animal species responsible for the fatal injuries, since the initial hypothesis concerned both the possibility that the attack was carried out by a cougar (*Puma concolor*) or by a pack of large dogs roaming the area where the victim's body was found. The main problem with the correct identification of the perpetrator of the death was the size and characteristics of the injuries found on the corpse, which are similar for both species. Following a careful assessment of the type of wounds, the anatomical position of the lesions, the tooth marks, the behavioural pattern of both animal species present in the environment, together with information of several eyewitnesses regarding previous aggressive behaviour of a pack of mixed-breed dogs of various sizes (e.g., Dogo Argentino, English Mastiff) found in the same area, the hypothesis regarding the dog pack being responsible for the attack and death of the victim was confirmed.

Another interesting situation involves canids. Wolves are considered a large and broadly distributed population in Europe. They preferably prey on wild animals and domestic ungulates (e.g., deer, cattle), causing great conflicts with humans and their economic interests in the case of livestock. Dogs are also widely distributed in Europe, coexisting with wolves, which means that wolf predations and bite marks sometimes can be confused with dog attacks [79,251]. There are plenty of cases in which the physical and circumstantial evidence at a crime scene refers to a fatal attack perpetrated against humans by wolves [23]. Wolves, however, are only under specific conditions responsible for violent attacks on humans, but like other large carnivores they participate in post-mortem scavenging. In this regard, a study by Toledo-González et al. [59] on the Iberian wolf and its associated bite marks to distinguish them from other canid species is interesting. The researchers described dental morphometric characteristics to aid the identification of the Iberian wolf's tooth/bite marks or to rule out other potential aggressors with great confidence.

Determining which predator species is responsible for killing a human is important, especially when there is the possibility of overlapping bite marks, as is the case with many carnivore species [62] (Table 2). In bite mark comparisons of sympatric animals (Note: *sympatry* is the term used to describe populations, varieties or species that occur in the same place at the same time [254]), measurements of the maxillary and mandibular intercanine distance are frequently used as an aid in identifying the different animal species responsible for a predatory or scavenging attack [255,256] (see Section 3.1.1. Species

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Identification for more information on this topic). The exclusion of one species over another in forensic contexts involving human remains, as well as in situations involving the interaction of wildlife, livestock, and humans, is crucial for determining appropriate management actions and for avoiding inappropriate human actions, such as the unjustified killing of certain predator species, mistakenly believed to be responsible for deadly and endangered attacks [79]. In relation to attacks by certain species on domestic livestock, Verzuh et al. [257] recently examined the mandibular and maxillary intercanine distance of Mexican wolves and sympatric carnivores (i.e., grey foxes, bobcats, coyotes, feral dogs, and cougars) in order to identify the measurements of intercanine distance that had the most potential to be multiple species and therefore, the least reliable measurements for bite mark analysis. The results of this study showed how indeed many of the measurements examined overlapped with each other, thus generating confusion in identification. However, it may also be useful for forensic investigators to know the measurement intervals considered problematic so that, together with the rest of the information obtained from a case, they can proceed with caution in the judicial investigation or at least not draw risky conclusions about the species involved in the predation.

Table 2. Ranges of intercanine distances in some species of the order Carnivora (data from [62,256–259].

Family	Species		Intercanine Distance (mm)		
		Common Name —	Maxilla	Mandible	
Felidae	Felis catus	Domestic cat	7.0-22.0	4.0-18.0	
	Felis catus	Feral cat	8.2-	21.0 *	
	Lynx canadensis	Canada lynx	22.7-27.6	21.9-27.8	
	Lynx lynx	Eurasian lynx	13.0-30.0	5.0-25.0	
	Lynx rufus	Bobcat	11.0-31.0	5.0-24.0	
	Puma concolor	Cougar	21.0-48.0	10.0-44.0	
Canidae	Canis latrans	Coyote	15.0-39.1	7.0-39.0	
	Canis lupus	Grey wolf	23.0-51.0	11.0-45.0	
	Canis lupus baileyi	Mexican wolf	31.4-49.8	27.8-43.7	
	Canis lupus familiaris	Domestic dog	13.0-48.0	6.0-49.0	
	Canis lupus familiaris	Feral dog	14.1-53.4	10.8-46.2	
	Urocyon cinereoargenteus	Gray fox	9.0-22.0	4.0-20.0	
	Vulpes lagopus	Arctic fox	19.4-24.2	16.4-20.8	
	Vulpes macrotis	Kit fox	13.8–17.5	11.3-15.5	
	Vulpes velox	Swift fox	15.0-17.0	13.6-15.2	
	Vulpes vulpes	Red fox	11.0-27.0	4.0-25.0	
Mephitidae	Mephitis mephitis	Striped skunk	12.0-16.4	10.3-14.8	
	Spilogale gracilis	Western spotted skunk	8.5–11.6	7.5–10.5	
Mustelidae	Genetta genetta	Spotted genet	9.0-	13.0 *	
	Gulo gulo	Wolverine	18.0-43.0	7.0-32.0	
	Lontra canadensis	Northern river otter	16.7–22.6	14.6-21.4	
	Martes americanus	American marten	8.6-14.6	8.2-12.5	
	Martes foina	Stone marten	11.0-	-15.0 *	
	Mustela erminea	Stoat	3.5–7.1	2.6-5.1	
	Mustela frenata	Long-tailed weasel	5.8-9.0	4.6-8.0	
	Mustela furo	Ferret	7.8–	13.6 *	
	Mustela lutreola	European mink	8.0-	12.0 *	
	Mustela nivalis	Least weasel	3.2-5.3	2.7-4.4	
	Mustela putorius	Polecat	8.0-	14.0 *	
	Neogale vison	American mink	8.3–12.4	6.2–10.1	
	Taxidea taxus	American badger	24.8-33.9	23.8-32.3	
Phocidae	Phoca vitulina	Harbour seal	27.5–36.3	23.0-36.1	
Procyonidae	Bassariscus astutus	Ringtail	9.5–12.2	8.6–11.8	
	Nasua narica	White-nosed coati	19.1–29.3	16.4–22.1	
	Procyon lotor	Raccoon	19.4–28.2	16.1–25.6	
Ursidae	Ursus americanus	Black bear	20.0–64.0	11.0-52.0	
	Ursus arctos horribilis	Grizzly bear	34.0–96.0	15.0–91.0	

<sup>\*</sup> Maxillary and mandibular intercanine distances are combined.

# 4. Application Potentialities in Other Contexts

So far, we have seen the importance of dentition in the identification process in forensic contexts. However, in addition to having an interest in the legal system, the analysis of

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animal dentition also has practical applications in other contexts where it is important to reliably identify the finding of animal remains, especially when they have suffered taphonomic alterations (e.g., fragmentation, cremation, cortical erosion) and teeth are the only available elements for analysis.

# 4.1. Archaeological and Palaeoanthropological Contexts

The analysis of bone modifications is the object of study in important disciplines such as bioarchaeology, palaeoanthropology, and zooarchaeology since, on the one hand, it allows understanding of the factors that caused such modifications, and on the other hand, allows reconstruction of the processes that led to the dislocation, fragmentation, and/or poor state of preservation of certain skeletal elements in the funerary context [260]. After the burial of an individual several physical, chemical, and biotic agents (e.g., fossorial species such as badgers, moles, voles, shrews, earthworms, beetles, ants; terrestrial species such as wild cats, lynxes, red foxes, raccoon dogs, brown bears, wild boars, squirrels; Note: fossorial species are those adapted to burrow into the ground and live primarily, but not solely, underground [261]) affect the body of the deceased. These agents are responsible for certain degradation processes that manifest themselves in the form of heterogeneous changes visible on the osteological remains (see Section 3.4. Bite Marks). It is necessary to take the utmost care when observing and analysing these macroscopic marks because in bioarchaeological and palaeoanthropological contexts it is common to find human remains that show clear traces of modifications related to animal action, but which, however, can easily be confused with the effect of pathological lesions or due to an incorrect excavation, transport, or storage of the remains by unqualified workers (Figure 6). The erroneous interpretation of the observed bone lesions can result in the loss of useful information for the interpretation of the post-depositional archaeological context and for further anthropological analyses. Among the bone lesions found most frequently in bioarchaeological contexts, those associated with the gnawing action of rodents or carnivores stand out [262], while in the palaeoanthropological context they are associated with the practice of scavenging by large carnivores [263]. In both cases, these bony lesions inform the researcher about the history of taphonomic damage after the death of an individual and before excavation/recovery.

The extent of osteological damage in archaeological contexts varies depending on the animal species involved, the environmental conditions, and the nature of the archaeological context (e.g., necropolis, isolated burials, caves) [264]. The attention of researchers to the carnivore tooth marks found on the skeletal remains of palaeoanthropological assemblages belonging to hominins is justified by the desire to understand the relationships and ways of living and sharing the same environments between large carnivores and humans; especially in prehistoric times, where some known examples of hominin remains show traces of bone modifications that could be associated with forms of feeding by certain animal species [265,266]. The possibility of identifying specific carnivores from the marks of their teeth left on hominin bones could provide sufficient evidence to demonstrate their presence in archaeological, palaeoanthropological and palaeoenvironmental times even when their skeletal remains are not preserved. Moreover, being able to distinguish which carnivores were active in the sites where faunal and human skeletal assemblages are recorded could provide useful information to reconstruct palaeoanthropological scenarios and to understand, if there were, patterns of competition between hominines and carnivores to access carcasses for nutritional purposes [267,268]. Furthermore, the interest in teeth marks left by carnivores on skeletal remains of hominines is justified by the fact that it is considered much more likely to find bite marks left by large carnivores on the bones of hominines than the other way around [266].

Interactions between hominins and large carnivores have occurred with great frequency and in different forms throughout the course of evolution, so much so that in many cases they have generated a series of reciprocal pressures [269–271]. From these interactions, the scenarios that generally seem to emerge include violent human–carnivore conflicts (associated

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with bone modifications), competition for the use of living habitats such as caves, exploitation, and scavenging of common prey or carcasses by large carnivores and hominins.

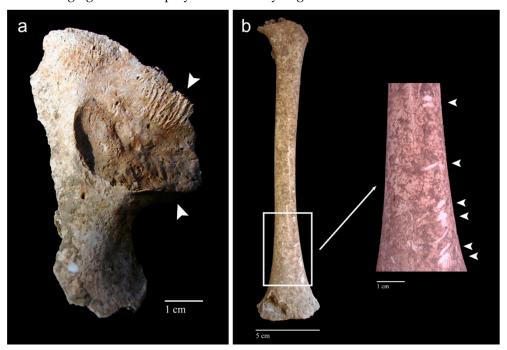


Figure 6. Human skeletal remains from the late medieval cemetery of Corfinio (12th–13th century CE, Italy). (a) Fragment of right coxal from a juvenile individual showing signs of rodent gnawing activity near the auricular surface (arrowheads). (b) Anterior view of the left femur belonging to a juvenile individual showing scratches that can be confused with signs of tooth marks; however, they are caused by a deficient excavation process using inadequate tools (i.e., lesions of post-mortem origin) (arrowheads). Images courtesy of C. Tanga from her bachelor's thesis [224].

Examples of human remains associated with animal scavenging activity are particularly recorded during the Pleistocene; in fact, several archaeological assemblages relating to both the accumulation of animal bones and hominins bearing signs of chewing by carnivores have been found at many sites from this period. The fact that prehistoric hominin remains show clear signs of animal biting is important evidence in palaeoanthropology and archaeology of the consumption of human remains by carnivores; however, it is not always possible to trace with absolute certainty the animal species directly involved in the scavenging and chewing of such remains [272].

Among the problematic issues related to the identification of both the type of bite mark and the animal species responsible for bone modification in archaeological contexts there is the interference of multiple physical and biological taphonomic agents, which may have succeeded one another and modified the initial context in a time interval ranging from the short to the long term. Another issue is the possible recurrence of events related to the chewing of the same remains by animals distinct from those responsible for the primary consumption event. In this regard, if traces of modification associated with different animal species are found on the same remains, a further problem concerns the possibility of establishing the sequence of the consumption action as well as understanding whether it is a predation phenomenon or secondary consumption of the carcass [273].

There are few cases in which the animal species involved has been identified with certainty or in which a relative hypothesis has been formulated. Interesting in this regard is the study by Daujeard et al. [274] on the remains of a Pleistocene human femur from North Africa with traces of teeth marks interpreted as resulting from the action of chewing for food by a large mammal, more probably a hyena. As in forensic cases, in palaeoanthropological contexts, bite marks have been recorded on fossil hominin remains applying the method of intercanine distance to identify the animal species involved in the formation of the

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tooth marks. An illustrative case is given by the analyses carried out on the remains of a skull belonging to a juvenile hominid from Swartkrans, South Africa. The analysis of the intercanine distance, together with the morphological study of the signs left by the dentition, led to the hypothesis that we were dealing with an attack by a leopard on a hominin [267,275].

### 4.2. Ecological Contexts: Study of the Biodiversity of Wild Animal Species

The identification techniques that are frequently employed in the legal system related to criminalistics can be applied in other contexts, where the identification of recovered osteological remains is also required, such as in ecological contexts. The study of wildlife remains is also important when unidentified carcasses are reported for monitoring and census purposes in parks and nature reserves in terms of health checks, hypotheses about the cause of death (e.g., investment or poaching), or the impact on biodiversity (e.g., habitat destruction, introduction of invasive non-native species, over-exploitation of resources, climate change). Censuses of various animal species, especially those considered at risk, have been conducted for many years in protected areas; in fact, the application of extrinsic marks (e.g., radio and satellite transmitters) on the wildlife in these areas is fundamental since it also allows the collection of additional information on the use of certain ecosystems. Reports of dead wildlife with extrinsic markings or animals that lack them are also of great interest, since their identification favours the collection of information that is useful not only for a better understanding of natural ecological phenomena, for planning actions aimed at protecting certain species, but also for assessing the risk of the spread of certain diseases transmissible to wildlife [276].

### 5. Concluding Remarks

There is no doubt that veterinary forensics is becoming increasingly important in our modern society, increasing the demand for investigations related to crimes against animals or investigations of criminal deaths of human beings involving animals. The high degree of mineralisation of dental tissues results in their hardness, durability, and resistance to post-mortem insults, so frequently the teeth are well preserved relative to bone tissue when an animal body is recovered a long time after death. Although the identification of carcasses in veterinary forensics is of less importance compared to its counterpart in human forensic medicine, the post-mortem dental profile of an animal can contribute essential information in resolving legal disputes involving animals and animal derivatives, such as species identification, sex, age-at-death, body size, geographical origin, and post-mortem interval. Moreover, the examination of dental features and characteristics of a bite mark may help identify the animal responsible for aggression against humans with often fatal consequences. This review points out the potential of dentition in the identification process in forensic contexts and emphasises the need for further research to give greater solidity to the results, helping the Courts in answering questions of interest to the legal system to reach a reliable verdict.

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#### References

1. Bailey, D. (Ed.) Introduction—What is the veterinary forensics? In *Practical Veterinary Forensics*; CAB International: Oxfordshire, UK, 2016; pp. 1–11.

- 2. Cooper, J.E.; Cooper, M.E. Special features of veterinary and comparative forensic medicine. In *Introduction to Veterinary and Comparative Forensic Medicine*; Cooper, J.E., Cooper, M.E., Eds.; Blackwell Publishing Ltd.: Oxford, UK, 2007; pp. 168–224.
- 3. De Siqueira, A.; Cuevas, S.E.C.; Salvagni, F.A.; Maiorka, P.C. Forensic veterinary pathology: Sharp injuries in animals. *Vet. Pathol.* **2016**, 53, 979–987. [CrossRef] [PubMed]
- 4. Forbes, N. An exacting science: The veterinary surgeon as expert witness. In Practice 2004, 26, 503–506. [CrossRef]
- 5. Cooper, J.E.; Cooper, M.E. Forensic veterinary medicine: A rapidly evolving discipline. *Forensic Sci. Med. Pathol.* **2008**, *4*, 75–82. [CrossRef]
- 6. Ledger, R.A.; Mellor, D.J. Forensic use of the five domains model for assessing suffering in cases of animal cruelty. *Animals* **2018**, 8, 101. [CrossRef] [PubMed]
- 7. Arkow, P.; Munro, H. The veterinary profession's roles in recognizing and preventing family violence: The experiences of the human medicine field and the development of diagnostic indicators of non-accidental injury. In *The International Handbook of Animal Abuse and Cruelty: Theory, Research, and Application*; Ascione, F., Ed.; Purdue University Press: West Lafayette, IN, USA, 2008; pp. 31–58.
- 8. McMillan, F.D.; Duffy, D.L.; Serpell, J.A. Mental health of dogs formerly used as 'breeding stock' in commercial breeding establishments. *Appl. Anim. Behav. Sci.* **2011**, *135*, 86–94. [CrossRef]
- 9. Munro, H.M.; Thrusfield, M.V. 'Battered pets': Non-accidental physical injuries found in dogs and cats. *J. Small Anim. Pract.* **2001**, 42, 279–290. [CrossRef]
- 10. Rebollada-Merino, A.; Bárcena, C.; Mayoral-Alegre, F.J.; García-Real, I.; Domínguez, L.; Rodríguez-Bertos, A. Forensic cases of suspected dog and cat abuse in the Community of Madrid (Spain), 2014–2019. *Forensic Sci. Int.* **2020**, *316*, 110522. [CrossRef] [PubMed]
- 11. Proulx, G.; Rodtka, D. Killing traps and snares in North America: The need for stricter checking time periods. *Animals* **2019**, *9*, 570. [CrossRef]
- 12. White, P.A.; Valkenburgh, B.V. Low-cost forensics reveal high rates of non-lethal snaring and shotgun injuries in Zambia's large carnivores. *Front. Conserv. Sci.* **2022**, *3*, 803381. [CrossRef]
- 13. Grilo, A.; Moreira, A.; Carrapiço, B.; Belas, A.; São Braz, B. Epidemiological study of pesticide poisoning in domestic animals and wildlife in Portugal: 2014–2020. *Front. Vet. Sci.* **2021**, *7*, 616293. [CrossRef]
- 14. Rial-Berriel, C.; Acosta-Dacal, A.; Zumbado, M.; Henríquez-Hernández, L.A.; Rodríguez-Hernández, Á.; Macías-Montes, A.; Boada, L.D.; Travieso-Aja, M.M.; Martin-Cruz, B.; Suárez-Pérez, A.; et al. Epidemiology of animal poisonings in the Canary Islands (Spain) during the period 2014–2021. *Toxics* 2021, 9, 267. [CrossRef] [PubMed]
- 15. Pankowski, F.; Bogiel, G.; Paśko, S.; Rzepiński, F.; Misiewicz, J.; Staszak, A.; Bonecka, J.; Dzierzęcka, M.; Bartyzel, B.J. Fatal gunshot injuries in the common buzzard *Buteo buteo* L. 1758—Imaging and ballistic findings. *Forensic Sci. Med. Pathol.* 2018, 14, 526–530. [CrossRef] [PubMed]
- Alleyne, E.; Parfitt, C. Adult-perpetrated animal abuse: A systematic literature review. *Trauma Violence Abuse* 2019, 20, 344–357.
  [CrossRef]
- 17. Kellert, S.R.; Felthous, A.R. Childhood cruelty toward animals among criminals and noncriminals. *Hum. Relat.* **1985**, *38*, 1113–1129. [CrossRef]
- 18. Ahmad, H.I.; Ahmad, M.J.; Jabbir, F.; Ahmar, S.; Ahmad, N.; Elokil, A.A.; Chen, J. The domestication makeup: Evolution, survival, and challenges. *Front. Ecol. Evol.* **2020**, *8*, 103. [CrossRef]
- 19. Cucchi, T.; Arbuckle, B. Animal domestication: From distant past to current development and issues. *Anim. Front.* **2021**, *11*, 6–9. [CrossRef]
- 20. Tanga, C.; Remigio, M.; Viciano, J. Transmission of zoonotic diseases in the daily life of ancient *Pompeii* and *Herculaneum* (79 CE, Italy): A review of animal-human-environment interactions through biological, historical and archaeological sources. *Animals* 2022, 12, 213. [CrossRef]
- 21. Errickson, D.; Thompson, T.J.U. Animal attacks and injuries: Anthropological findings. In *Encyclopedia of Forensic and Legal Medicine*, 2nd ed.; Payne-James, J., Byard, R.W., Eds.; Elsevier: Amsterdam, The Netherlands, 2016; pp. 143–147. [CrossRef]
- 22. Fonseca, G.M.; Palacios, R. An unusual case of predation: Dog pack or cougar attack? J. Forensic Sci. 2013, 58, 224–227. [CrossRef]
- 23. Butler, L.; Dale, B.; Beckmen, K.; Farley, S. *Findings Related to the March 2010 Fatal Wolf Attack Near Chignik Lake, Alaska*; ADF&G/DWC/WSP-2011-2; Alaska Department of Fish and Game, Division of Wildlife Conservation, Wildlife Special Publication: Juneau, AK, USA, 2011.
- 24. Ďatko, M.; Vojtíšek, T.; Hejna, P. A fatal lion attack. Forensic Sci. Med. Pathol. 2015, 11, 300–302. [CrossRef]
- 25. Kanchan, T.; Shekhawat, R.S.; Shetty, B.S.K.; Jayaram, L.; Meshram, V.P. Fatal captive tiger attack—A case report with review of literature. *J. Forensic Leg. Med.* **2021**, *78*, 102100. [CrossRef]
- 26. Floyd, T. Bear-inflicted human injury and fatality. Wilderness Environ. Med. 1999, 10, 75–87. [CrossRef]
- 27. Yilmaz, A.; Hosukler, E.; Erkol, Z.; Samurcu, H.; Gundogdu, V.; Celme, H.A.; Turan, T.; Dincer, G. Old female case who died as a result of bear attack. *Med. Sci.* **2021**, *10*, 623–626. [CrossRef]

Animals 2022, 12, 2038 22 of 30

28. Janda, D.H.; Ringler, D.H.; Hilliard, J.K.; Hankin, R.C.; Hankin, F.M. Nonhuman primate bites. *J. Orthop. Res.* **1990**, *8*, 146–150. [CrossRef]

- 29. Behera, C.; Chauhan, M.; Sikary, A.; Millo, T.; Byard, R.W. Injuries and deaths caused by non-human primate attacks: Paediatric vulnerability. *Med. Sci. Law* **2021**, *61*, *64*–*68*. [CrossRef] [PubMed]
- 30. Manipady, S.; Menezes, R.G.; Bastia, B.K. Death by attack from a wild boar. J. Clin. Forensic Med. 2006, 13, 89–91. [CrossRef]
- 31. Gitto, L.; Bonaccorso, L.; Bryant, S.M.; Serinelli, S. Death caused by a domestic pig attack. *Forensic Sci. Med. Pathol.* **2021**, 17, 469–474. [CrossRef]
- 32. Kovalenko, F.; Vassiliu, P.; Degiannis, K.; Doll, D. Complex femoral injuries after hippopotamus bite: A case report. *Albanian J. Trauma Emerg. Surg.* **2021**, *5*, 879–882. [CrossRef]
- 33. Yanai, O.; Goldin, L.; Hiss, J. Fatal rat bites. Harefuah 1999, 136, 611–613.
- 34. Skarentzos, S.K.; Papadopulos, N.; Deftereos, S.P.; Thomaidis, S.; Kambouri, K. Serious rodent bites to an 8-month-old infant due to child neglect. *Cureus* **2021**, *13*, e18493. [CrossRef]
- 35. Chattopadhyay, S.; Shee, B.; Sukul, B. Fatal crocodile attack. J. Forensic Leg. Med. 2013, 20, 1139–1141. [CrossRef]
- 36. Boyd, B.S.; Colon, F.; Doty, J.F.; Sanders, K.C. Beware of the dragon: A case report of a Komodo dragon attack. *Foot Ankle Orthop.* **2021**, *6*, 24730114211015623. [CrossRef] [PubMed]
- 37. Idani, M.; N'guessan, E.K.Z.; Millogo, M.; Badini, S.P.A.; Zango, A.; Ili, B.V.; Crezoit, G.E.; Konsem, T. Cranio-cervico-facial injuries due to crocodile bite: A rare observation in Yalgado Ouedraogo University Hospital (Burkina Faso). *J. Oral Med. Oral Surg.* 2022, 28, 12. [CrossRef]
- 38. Byard, R.W.; James, R.A.; Heath, K.J. Recovery of human remains after shark attack. *Am. J. Forensic Med. Pathol.* **2006**, 27, 256–259. [CrossRef] [PubMed]
- 39. Clua, E.; Reid, D. Features and motivation of a fatal attack by a juvenile white shark, *Carcharodon carcharias*, on a young male surfer in New Caledonia (South Pacific). *J. Forensic Leg. Med.* **2013**, 20, 551–554. [CrossRef] [PubMed]
- 40. Clua, E.; Bescond, P.M.; Reid, D. Fatal attack by a juvenile tiger shark, *Galeocerdo cuvier*, on a kitesurfer in New Caledonia (South Pacific). *J. Forensic Leg. Med.* **2014**, 25, 67–70. [CrossRef] [PubMed]
- 41. Indra, L.; Errickson, D.; Young, A.; Lösch, S. Uncovering forensic taphonomic agents: Animal scavenging in the European context. *Biology* **2022**, *11*, 601. [CrossRef]
- 42. Cattaneo, C.; Gibelli, D. Animal effects on bones. In *Forensic Anthropology*; Houck, M.M., Ed.; Academic Press: London, UK, 2017; pp. 31–35.
- 43. Young, A. The effects of terrestrial mammalian scavenging and avian scavenging on the body. In *Taphonomy of Human Remains: Forensic Analysis of the Dead and the Depositional Environment*; Schotsmans, E.M.J., Márquez-Grant, N., Forbes, S.L., Eds.; John Wiley & Sons Ltd.: Chichester, UK, 2017; pp. 212–234.
- 44. Pokines, J.T. Fauna dispersal, reconcentration, and gnawing damage to bone in terrestrial environments. In *Manual of Forensic Taphonomy*, 2nd ed.; Pokines, J.T., L'Abbé, E.N., Symes, S.A., Eds.; CRC Press: Boca Raton, FL, USA, 2022; pp. 295–359. [CrossRef]
- 45. Munro, R.; Munro, H.M.C. Some challenges in forensic veterinary pathology: A review. *J. Comp. Pathol.* **2013**, 149, 57–73. [CrossRef]
- 46. Kles, M.; Sutton, L. Forensic veterinary osteology. In *Veterinary Forensic Medicine and Forensic Sciences*; Byrd, J.H., Norris, P., Bradley-Siemens, N., Eds.; CRC Press: Boca Raton, FL, USA, 2021; pp. 199–224.
- 47. O'Connor, T. The Archaeology of Animal Bones; Texas A&M University Press: College Station, TX, USA, 2008.
- 48. Gifford-Gonzalez, D. Bone's intrinsic traits: Inferring species, sex, and age. In *An Introduction to Zooarchaeology*; Gifford-Gonzalez, D., Ed.; Springer: New York, NY, USA, 2018; pp. 103–123.
- 49. Ungar, P.S. How teeth work. In *Evolution's Bite: A Story of Teeth, Diet, and Human Origins*; Ungar, P.S., Ed.; Princeton University Press: Princeton, NJ, USA, 2017; pp. 5–33.
- 50. Sims, M.E. Cranial morphology of five felids: *Acinonyx jubatus, Panthera onca, Panthera pardus, Puma concolor, Uncia uncia. Russ. J. Theriol.* **2012**, *11*, 157–170. [CrossRef]
- 51. Roy, J.; Singh, A.; Rohith, M.M.; Sharma, L.K.; Johnson, A.; Joshi, H.; Chinnadurai, V.; Chandra, K.; Thakur, M. Cranio-dental signature of three big cats of India: Implications in wildlife forensics. *Proc. Zool. Soc.* **2022**, *75*, 57–64. [CrossRef]
- 52. Hillson, S. Teeth, 2nd ed.; Cambridge University Press: Cambridge, UK, 2005.
- 53. Halstead, P.; Collins, P.; Isaakidou, V. Sorting the sheep from the goats: Morphological distinctions between the mandibles and mandibular teeth of adult *Ovis* and *Capra. J. Archaeol. Sci.* **2002**, *29*, 545–553. [CrossRef]
- 54. Whitenack, L.B.; Gottfried, M.D. A morphometric approach for addressing tooth-based species delimitation in fossil make sharks, *Isurus* (Elasmobranchii: Lamniformes). *J. Vertebr. Paleontol.* **2010**, *30*, 17–25. [CrossRef]
- 55. Boroni, N.L.; Lobo, L.S.; Romano, P.S.R.; Lessa, G. Taxonomic identification using geometric morphometric approach and limited data: An example using the upper molars of two sympatric species of *Calomys* (Cricetidae: Rodentia). *Zoologia* **2017**, *34*, e19864. [CrossRef]
- 56. Chuang, R.; Bonhomme, V. Rethinking the dental morphological differences between domestic equids. *J. Archaeol. Sci.* **2019**, 101, 140–148. [CrossRef]
- 57. Hulme-Beaman, A.; Claude, J.; Chaval, Y.; Evin, A.; Morand, S.; Vigne, J.D.; Dobney, K.; Cucchi, T. Dental shape variation and phylogenetic signal in the Rattini tribe species of Mainland Southeast Asia. *J. Mammal. Evol.* **2019**, *26*, 435–446. [CrossRef]

Animals 2022, 12, 2038 23 of 30

58. Miele, V.; Dussert, G.; Cucchi, T.; Renaud, S. Deep learning for species identification of modern and fossil rodent molars. *BioRxiv* 2020. [CrossRef]

- 59. Toledo-González, V.; Ortega-Ojeda, F.; Fonseca, G.M.; García-Ruiz, C.; Navarro-Cáceres, P.; Pérez-Lloret, P.; Marín-García, M.D.P. A morphological and morphometric dental analysis as a forensic tool to identify the Iberian wolf (*Canis lupus signatus*). *Animals* **2020**, *10*, 975. [CrossRef]
- 60. Plavcan, J.M.; Cope, D.A. Metric variation and species recognition in the fossil record. *Evol. Anthropol.* **2001**, 10, 204–222. [CrossRef]
- 61. Simpson, G.G.; Roe, A.; Lewontin, R.C. Quantitative Zoology; Harcourt, Brace and Company: New York, NY, USA, 1960.
- 62. Murmann, D.C.; Brumit, P.C.; Schrader, B.A.; Senn, D.R. A comparison of animal jaws and bite mark patterns. *J. Forensic. Sci.* **2006**, *51*, 846–860. [CrossRef]
- 63. Cucchi, T.; Mohaseb, A.; Peigné, S.; Debue, K.; Orlando, L.; Mashkour, M. Detecting taxonomic and phylogenetic signals in equid cheek teeth: Towards new palaeontological and archaeological proxies. *R. Soc. Open Sci.* **2017**, *4*, 160997. [CrossRef]
- 64. Berthaume, M.A.; Lazzari, V.; Guy, F. The landscape of tooth shape: Over 20 years of dental topography in primates. *Evol. Anthropol.* **2020**, 29, 245–262. [CrossRef]
- 65. Da Silva, R.F.; Mendes, S.D.S.C.; Marinho, D.E.A.; Júnior, A.F.R.; Guimarães, M.A. Importance of the comparative anatomy in Forensic Anthropology—Case report. *RSBO* **2013**, *10*, 193–197.
- 66. Courtenay, L.A.; Herranz-Rodrigo, D.; González-Aguilera, D.; Yravedra, J. Developments in data science solutions for carnivore tooth pit classification. *Sci. Rep.* **2021**, *11*, 10209. [CrossRef] [PubMed]
- 67. Yravedra, J.; Maté-González, M.Á.; Courtenay, L.A.; González-Aguilera, D.; Fernández-Fernández, M. The use of canid tooth marks on bone for the identification of livestock predation. *Sci. Rep.* **2019**, *9*, 16301. [CrossRef] [PubMed]
- 68. Courtenay, L.A.; Yravedra, J.; Huguet, R.; Aramendi, J.; Maté-González, M.Á.; González-Aguilera, D.; Arriaza, M.C. Combining machine learning algorithms and geometric morphometrics: A study of carnivore tooth marks. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **2019**, 522, 28–39. [CrossRef]
- 69. Whittaker, D.K.; Rawle, L.W. The effect of conditions of putrefaction on species determination in human and animal teeth. *Forensic Sci. Int.* **1987**, 35, 209–212. [CrossRef]
- 70. Balasse, M.; Ambrose, S.H. Distinguishing sheep and goats using dental morphology and stable carbon isotopes in C<sub>4</sub> grassland environments. *J. Archaeol. Sci.* **2005**, *32*, 691–702. [CrossRef]
- 71. Kim, T.W.; Lee, H.J.; Kim, Y.K.; Oh, H.S.; Han, S.H. Genetic identification of prey species from teeth in faeces from the Endangered leopard cat *Prionailurus bengalensis* using mitochondrial *cytochrome b* gene sequence. *Mitochondrial DNA A DNA Mapp. Seq. Anal.* **2018**, 29, 170–174. [CrossRef]
- 72. Minakshi, P.; Shafiq, M.; Alisha, A.; Ranjan, K.; Kumar, P.; Anshul, B.; Singh, R.; Basanti, S.; Prasad, G. Animal forensics and applications. In *Biotechnology: Prospects and Applications*; Salar, R.K., Gahlawat, S.K., Siwach, P., Duhan, J.S., Eds.; Springer: New Delhi, India, 2013; pp. 265–286. [CrossRef]
- 73. Buddhachat, K.; Klinhom, S.; Siengdee, P.; Brown, J.L.; Nomsiri, R.; Kaewmong, P.; Thitaram, C.; Mahakkanukrauh, P.; Nganvongpanit, K. Elemental analysis of bone, teeth, horn and antler in different animal species using non-invasive handheld X-ray fluorescence. *PLoS ONE* **2016**, *11*, e0155458. [CrossRef]
- 74. Nganvongpanit, K.; Brown, J.L.; Buddhachat, K.; Somgird, C.; Thitaram, C. Elemental analysis of Asian elephant (*Elephas maximus*) teeth using X-ray fluorescence and a comparison to other species. *Biol. Trace Elem. Res.* **2016**, *170*, 94–105. [CrossRef]
- 75. Nganvongpanit, K.; Buddhachat, K.; Klinhom, S.; Kaewmong, P.; Thitaram, C.; Mahakkanukrauh, P. Determining comparative elemental profile using handheld X-ray fluorescence in humans, elephants, dogs, and dolphins: Preliminary study for species identification. *Forensic Sci. Int.* **2016**, 263, 101–106. [CrossRef]
- 76. Irfanita, N.; Jaswir, I.; Mirghani, M.E.S.; Sukmasari, S.; Ardini, Y.D.; Lestari, W. Rapid detection of gelatin in dental materials using Attenuated Total Reflection Fourier Transform Infrared spectroscopy (ATR-FTIR). J. Phys. Conf. Ser. 2017, 884, 012090. [CrossRef]
- 77. Wei, X.; Yu, K.; Wu, H.; Shen, C.; Li, H.; Liu, R.; Sun, Q.; Wang, Z. Species identification of teeth of human and non-human. *Forensic Sci. Int.* **2022**, 333, 111205. [CrossRef] [PubMed]
- 78. Caniglia, R.; Fabbri, F.; Mastrogiuseppe, L.; Randia, E. Who is who? Identification of livestock predators using forensic genetic approaches. *Forensic Sci. Int. Genet.* **2013**, *7*, 397–404. [CrossRef]
- 79. Toledo-González, V.; Ortega-Ojeda, F.; Fonseca, G.M.; García-Ruiz, C.; Pérez-Lloret, P. Analysis of tooth mark patterns on bone remains caused by wolves (*Canis lupus*) and domestic dogs (*Canis lupus familiaris*) for taxonomic identification: A scoping review focused on their value as a forensic tool. *Appl. Anim. Behav. Sci.* 2021, 240, 105356. [CrossRef]
- 80. McPherson, F.J.; Chenoweth, P.J. Mammalian sexual dimorphism. Anim. Reprod. Sci. 2012, 131, 109–122. [CrossRef]
- 81. Lough-Stevens, M.; Schultz, N.G.; Dean, M.D. The baubellum is more developmentally and evolutionarily labile than the baculum. *Ecol. Evol.* **2017**, *8*, 1073–1083. [CrossRef] [PubMed]
- 82. Szuma, E. Geography of sexual dimorphism in the tooth size of the red fox *Vulpes vulpes* (Mammalia, Carnivora). *J. Zool. Syst. Evol. Res.* **2008**, *46*, 73–81. [CrossRef]
- 83. Weckerly, F.W. Sexual-size dimorphism: Influence of mass and mating systems in the most dimorphic mammals. *J. Mammal.* **1998**, 79, 33–52. [CrossRef]
- 84. Gittleman, J.L.; Valkenburgh, B.V. Sexual dimorphism in the canines and skulls of carnivores: Effects of size, phylogeny, and behavioural ecology. *J. Zool.* **1997**, 242, 97–117. [CrossRef]

Animals 2022, 12, 2038 24 of 30

85. Gingerich, P.D. Variation, sexual dimorphism, and social structure in the early Eocene horse *Hyracotherium* (Mammalia, Perissodactyla). *Paleobiology* **1981**, *7*, 443–455. [CrossRef]

- 86. Lüps, P.; Roper, T.J. Tooth size in the European badger (*Meles meles*) with special reference to sexual dimorphism, diet and intraspecific aggression. *Acta Theriol.* **1988**, *33*, 21–33. [CrossRef]
- 87. Mayer, J.J.; Brisbin, I.L., Jr. Sex identification of Sus scrofa based on canine morphology. J. Mammal. 1988, 69, 408–412. [CrossRef]
- 88. Lowry, M.S.; Folk, R.L. Sex determination of the California sea lion (*Zalophus californianus*) from canine teeth. *Mar. Mammal Sci.* 1990, *6*, 25–31. [CrossRef]
- 89. Valkenburgh, B.V.; Ruff, C.B. Canine tooth strength and killing behaviour in large carnivores. *J. Zool.* **1987**, 212, 379–397. [CrossRef]
- 90. Turner, A. Dental sex dimorphism in European lions (*Panthera leo* L.) of the Upper Pleistocene: Palaeoecological and palaeoethological implications. *Ann. Zool. Fenn.* **1984**, *21*, 1–8.
- 91. Christiansen, P.; Harris, J.M. Variation in craniomandibular morphology and sexual dimorphism in pantherines and the sabercat *Smilodon fatalis*. *PLoS ONE* **2012**, 7, e48352. [CrossRef]
- 92. Szuma, E. Variation and correlation patterns in the dentition of the red fox from Poland. Ann. Zool. Fenn. 2000, 37, 113–127.
- 93. Kurtén, B. Sex dimorphism and size trends in the cave bear, Ursus spelaeus Rosenmüller and Heinroth. Acta Zool. Fenn. 1955, 90, 1–48.
- 94. Larsen, T. Sexual dimorphism in the molar rows of the polar bear. J. Wildl. Manag. 1971, 35, 374–377. [CrossRef]
- 95. Harvey, P.H.; Kavanagh, M.; Clutton-Brock, T.H. Sexual dimorphism in primate teeth. J. Zool. Lond. 1978, 186, 474–485. [CrossRef]
- 96. Beauchamp, G. Canine tooth size variability in primates. Folia Primatol. 1989, 52, 148–155. [CrossRef] [PubMed]
- 97. Lucas, P.W.; Corlett, R.T.; Luke, D.A. Sexual dimorphism of tooth size in anthropoids. Hum. Evol. 1986, 1, 23–39. [CrossRef]
- 98. Plavcan, J.M. Sexual dimorphism in primate evolution. Am. J. Phys. Anthropol. 2001, 116 (Suppl. S33), 25–53. [CrossRef] [PubMed]
- 99. Schwartz, G.T.; Dean, C. Ontogeny of canine dimorphism in extant hominoids. *Am. J. Phys. Anthropol.* **2001**, *115*, 269–283. [CrossRef]
- 100. Plavcan, J.M. Sexual size dimorphism, canine dimorphism, and male-male competition in primates: Where do humans fit in? *Hum. Nat.* **2012**, 23, 45–67. [CrossRef]
- 101. Garn, S.; Kerewsky, R.; Swindler, D. Canine "field" in sexual dimorphism of tooth size. Nature 1966, 212, 1501–1502. [CrossRef]
- 102. Gingerich, P.D.; Ryan, A.S. Dental and cranial variation in living Indriidae. Primates 1979, 20, 141–159. [CrossRef]
- 103. Gingerich, P.D.; Schoeninger, M.J. Patterns of tooth size variability in the dentition of primates. *Am. J. Phys. Anthropol.* **1979**, *51*, 457–465. [CrossRef]
- 104. Plavcan, J.M.; van Schaik, C.P. Intrasexual competition and canine dimorphism in anthropoid primates. *Am. J. Phys. Anthropol.* **1992**, *87*, 461–477. [CrossRef]
- 105. Swindler, D.R. Appendix 1: Odontometry. In *Primate Dentition: An Introduction to the Teeth of Non-Human Primates*; Swindler, D.R., Ed.; Cambridge University Press: Cambridge, UK, 2002; pp. 165–219.
- 106. Sriraman, P.K. Forensic necropsy. In Wildlife Necropsy and Forensics; Sriraman, P.K., Ed.; CRC Press: Oxon, UK, 2021; pp. 152–199.
- 107. Cooper, J.E.; Cooper, M.E. Clinical work. In *Introduction to Veterinary and Comparative Forensic Medicine*; Cooper, J.E., Cooper, M.E., Eds.; Blackwell Publishing Ltd.: Oxford, UK, 2007; pp. 117–167.
- 108. Mendes-Oliveira, A.C.; Maria, S.L.S.; Lima, R.C.S.; Fernandes, A.S.; Almeida, P.C.R.; Montag, L.F.; Cavalho, O., Jr. Testing simple criteria for de age estimation of six hunted mammal species in Brazilian Amazon. *Mastozool. Neotrop.* **2012**, *19*, 105–116.
- 109. Cooper, J.E.; Cooper, M.E. Importance and application of animal law. In *Introduction to Veterinary and Comparative Forensic Medicine*; Cooper, J.E., Cooper, M.E., Eds.; Blackwell Publishing Ltd.: Oxford, UK, 2007; pp. 42–60.
- 110. Modina, S.C.; Andreis, M.E.; Moioli, M.; Di Giancamillo, M. Age assessment in puppies: Coming to terms with forensic requests. *Forensic Sci. Int.* **2019**, 297, 8–15. [CrossRef] [PubMed]
- 111. Park, K.; Ahn, J.; Kang, S.; Lee, E.; Kim, S.; Park, S.; Park, S.; Noh, H.; Seo, K. Determining the age of cats by pulp cavity/tooth width ratio using dental radiography. *J. Vet. Med. Sci.* **2014**, *15*, 557–561. [CrossRef] [PubMed]
- 112. Roccaro, M.; Peli, A. Age determination in dog puppies by teeth examination: Legal, health and welfare implications, review of the literature and practical considerations. *Vet. Ital.* **2020**, *56*, 149–162. [CrossRef] [PubMed]
- 113. Cooper, J.E.; Cooper, M.E. Pathology and post-mortem examinations. In *Introduction to Veterinary and Comparative Forensic Medicine*; Cooper, J.E., Cooper, M.E., Eds.; Blackwell Publishing Ltd.: Oxford, UK, 2007; pp. 168–224.
- 114. Mbizah, M.M.; Steenkamp, G.; Groom, R.J. Evaluation of the applicability of different age determination methods for estimating age of the endangered African wild dog (*Lycaon pictus*). *PLoS ONE* **2016**, *11*, e0164676. [CrossRef]
- 115. Sutton, L.K.; Byrd, J.H.; Brooks, J.W. Age determination in dogs and cats. In *Veterinary Forensic Pathology*; Brooks, J.W., Ed.; Springer: Cham, Switzerland, 2018; Volume 2, pp. 151–163.
- 116. Adserias-Garriga, J. Evolution of methods and state-of-the-art in dental age estimation. In *Age Estimation*; Adserias-Garriga, J., Ed.; Academic Press: London, UK, 2019; pp. 77–87.
- 117. O'Connor, T.P. Vertebrate demography by numbers: Age, sex, and zooarchaeological practice. In *Recent Advances in Ageing and Sexing Animal Bones*; Ruscillo, D., Ed.; Oxbow Books: Oxford, UK, 2006; pp. 1–8.
- 118. Lemons, M.; Beebe, D. Oral anatomy and physiology. In *Wiggs's Veterinary Dentistry: Principles and Practice*, 2nd ed.; Lobprise, H.B., Dodd, J.R., Eds.; Wiley-Blackwell: Hoboken, NJ, USA, 2019; pp. 1–24.
- 119. Spinage, C.A. A review of age determination of mammals by means of teeth, with special reference to Africa. *East Afr. Wildl. J.* **1973**, *11*, 165–187. [CrossRef]

Animals 2022, 12, 2038 25 of 30

120. Silver, I.A. The ageing of domestic animals. In *Science in Archaeology: A Comprehensive Survey of Progress and Research*, 2nd ed.; Brothwell, D., Higgs, E., Eds.; Basic Books: New York, NY, USA, 1963; pp. 250–268.

- 121. Lorászkó, G.; Rácz, B.; Ózsvári, L. Changes in the dentition of small dogs up to 4 months of age. Animals 2022, 12, 1417. [CrossRef]
- 122. Kelley, J.; Schwartz, G.T.; Smith, T.M. Age at first molar emergence in *Pan troglodytes* verus and variation in the timing of molar emergence among free-living chimpanzees. *J. Hum. Evol.* 2020, 145, 102823. [CrossRef]
- 123. Mahoney, P. Root growth and dental eruption in modern human deciduous teeth with preliminary observations on great apes. *J. Hum. Evol.* **2019**, 129, 46–53. [CrossRef]
- 124. Holly Smith, B.; Crummett, T.L.; Brandt, K.L. Ages of eruption of primate teeth: A compendium for aging individuals and comparing life histories. *Am. J. Phys. Anthropol.* **1994**, *37*, 177–231. [CrossRef]
- 125. Smith, T.D.; Muchlinski, M.N.; Jankord, K.D.; Progar, A.J.; Bonar, C.J.; Evans, S.; Williams, L.; Vinyard, C.J.; Deleon, V.B. Dental maturation, eruption, and gingival emergence in the upper jaw of newborn primates. *Anat. Rec.* 2015, 298, 2098–2131. [CrossRef]
- 126. Greenfield, H.J.; Arnold, E.R. Absolute age and tooth eruption and wear sequences in sheep and goat: Determining age-at-death in zooarchaeology using a modern control sample. *J. Archaeol. Sci.* 2008, 35, 836–849. [CrossRef]
- 127. Moran, N.C.; O'Connor, T.P. Age attribution in domestic sheep by skeletal and dental maturation: A pilot study of available sources. *Int. J. Osteoarchaeol.* **1994**, *4*, 267–285. [CrossRef]
- 128. Andrews, A.H. The relationship of bovine mandibular cheek tooth development to age determined by post-mortem radiographic examination of cattle aged between 12 and 24 months. *J. Agric. Sci.* **1982**, *98*, 109–117. [CrossRef]
- 129. Oroian, T.E.; Oroian, R.G.; Pasca, I.; Oroian, E.; Covrig, L. Methods of age estimation by dentition in *Sus scrofa ferus* sp. *Bull. Univ. Agric. Sci. Vet. Med.* 2010, 67, 291–295. [CrossRef]
- 130. Payne, S. Kill-off patterns in sheep and goats: The mandibles from Aşvan Kale. Anatol. Stud. 1973, 23, 281–303. [CrossRef]
- 131. Grant, A. The use of tooth wear as a guide to the age of domestic ungulates. In *Ageing and Sexing Animal Bones from Archaeological Sites*; Wilson, B., Grigson, C., Payne, S., Eds.; British Archaeological Reports: Oxford, UK, 1982; pp. 91–108.
- 132. Spinage, C.A. Age estimation of zebra. East Afr. Wildl. J. 1972, 10, 273–277. [CrossRef]
- 133. Levine, M.A. The use of crown height measurements and eruption-wear sequences to age horse teeth. In *Ageing and Sexing Animal Bones from Archaeological Sites*; Wilson, B., Grigson, C., Payne, S., Eds.; British Archaeological Reports: Oxford, UK, 1982; pp. 223–250.
- 134. Klein, R.G.; Wolf, C.; Freeman, L.G.; Allwarden, K. The use of dental crown heights for constructing age profiles of red deer and similar species in archaeological samples. *J. Archaeol. Sci.* **1981**, *8*, 1–31. [CrossRef]
- 135. Lieberman, D.E. The biological basis for seasonal increments in dental cementum and their application to archaeological research. *J. Archaeol. Sci.* **1994**, *21*, 525–539. [CrossRef]
- 136. Veiberg, V.; Nilsen, E.B.; Rolandsen, C.M.; Heim, M.; Andersen, R.; Holmstrøm, F.; Meisingset, E.L.; Solberg, E.J. The accuracy and precision of age determination by dental cementum annuli in four northern cervids. *Eur. J. Wildl. Res.* **2020**, *66*, 91. [CrossRef]
- 137. Wolfe, M.L. Age determination in moose from cemental layers of molar teeth. J. Wildl. Manag. 1969, 33, 428–431. [CrossRef]
- 138. Low, W.A.; Cowan, I.M.T. Age determination of deer by annular structure of dental cementum. *J. Wildl. Manag.* **1963**, 27, 466–471. [CrossRef]
- 139. Stoneberg, R.P.; Jonkel, C.J. Age determination of black bears by cementum layers. J. Wildl. Manag. 1966, 30, 411-414. [CrossRef]
- 140. Medill, S.; Derocher, A.E.; Stirling, I.; Lunn, N.; Moses, R.A. Estimating cementum annuli width in polar bears: Identifying sources of variation and error. *J. Mammal.* **2009**, *90*, 1256–1264. [CrossRef]
- 141. Christensen-Dalsgaard, S.N.; Aars, J.; Andersen, M.; Lockyer, C.; Yoccoz, N.G. Accuracy and precision in estimation of age of Norwegian Arctic polar bears (*Ursus maritimus*) using dental cementum layers from known-age individuals. *Polar Biol.* **2010**, *33*, 589–597. [CrossRef]
- 142. Mansfield, A.W.; Fisher, H.D. Age determination in the harbour seal, *Phoca vitulina* L. *Nature* 1960, 186, 92–93. [CrossRef]
- 143. Stewart, R.E.A.; Stewart, B.E.; Stirling, I.; Street, E. Counts of growth layer groups in cementum and dentine in ringed seals (*Phoca hispida*). *Mar. Mammal Sci.* **1996**, *12*, 383–401. [CrossRef]
- 144. Crowe, D.M.; Strickland, M.D. Dental annulation in the American badger. J. Mammal. 1975, 56, 269–272. [CrossRef]
- 145. Clarke, C.M.H.; Dzieciolowski, R.M.; Batcheler, D.; Frampton, C.M. A comparison of tooth eruption and wear and dental cementum techniques in age determination of New Zealand feral pigs. *Wildl. Res.* **1992**, *19*, 769–777. [CrossRef]
- 146. Nomokonova, T.; Losey, R.J.; McLachlin, K.; Bachura, O.P.; Gusev, A.V.; Kosintsev, P.A.; Fedorovad, N.V.; Sablin, M.V. Age estimation of archaeological dogs using pulp cavity closure ratios. *J. Archaeol. Sci.* **2020**, *123*, 105252. [CrossRef]
- 147. Knowlton, F.F.; Whittemore, S.L. Pulp cavity-tooth width ratios from known-age and wild-caught coyotes determined by radiography. *Wildl. Soc. Bull.* **2001**, 29, 239–244.
- 148. White, P.A.; Ikanda, D.; Ferrante, L.; Chardonnet, P.; Mesochina, P.; Cameriere, R. Age estimation of African lions *Panthera leo* by ratio of tooth Areas. *PLoS ONE* **2016**, *11*, e0153648. [CrossRef] [PubMed]
- 149. Hopkins, S.S.B. Estimation of body size in fossil mammals. In *Methods in Paleoecology: Reconstructing Cenozoic Terrestrial Environments and Ecological Communities*; Croft, D.A., Su, D.F., Simpson, S.W., Eds.; Springer: Berlin/Heidelberg, Germany, 2018; pp. 7–22. [CrossRef]
- 150. Christiansen, F.; Sironi, M.; Moore, M.J.; Di Martino, M.; Ricciardi, M.; Warick, H.A.; Irschick, D.J.; Gutierrez, R.; Uhart, M.M. Estimating body mass of free-living whales using aerial photogrammetry and 3D volumetrics. *Methods Ecol. Evol.* **2019**, *10*, 2034–2044. [CrossRef]

Animals 2022, 12, 2038 26 of 30

151. Iskjaer, C.; Slade, N.A.; Childs, J.E.; Glass, G.E.; Korch, G.W. Body mass as a measure of body size in small mammals. *J. Mammal.* 1989, 70, 662–667. [CrossRef]

- 152. Chrószcz, A.; Janeczek, M.; Onar, V.; Staniorowski, P.; Pospieszny, N. The shoulder height estimation in dogs based on the internal dimension of cranial cavity using mathematical formula. *Anat. Histol. Embryol.* **2007**, *36*, 269–271. [CrossRef]
- 153. Chrószcz, A.; Janeczek, M.; Pasicka, E.; Klećkowska-Nawrot, J. Height at the withers estimation in the horses based on the internal dimension of cranial cavity. *Folia Morphol.* **2014**, *73*, 143–148. [CrossRef]
- 154. Gingerich, P.D. Prediction of body mass in mammalian species from long bone lengths and diameters. In *Contributions from the Museum of Paleontology*; The University of Michigan: Ann Arbor, MI, USA, 1990; Volume 28, pp. 79–92.
- 155. Campione, N.E.; Evans, D.C. A universal scaling relationship between body mass and proximal limb bone dimensions in quadrupedal terrestrial tetrapods. *BMC Biol.* **2012**, *10*, 60. [CrossRef]
- 156. Ruff, C.B. Body mass and hindlimb bone cross-sectional and articular dimensions in anthropoid primates. In *Body Size in Mammalian Paleobiology: Estimation and Biological Implications*; Damuth, J., MacFadden, B.J., Eds.; Cambridge University Press: Cambridge, UK, 1990; pp. 119–150.
- 157. Copes, L.E.; Schwartz, G.T. The scale of it all: Postcanine tooth size, the taxon-level effect, and the universality of Gould's scaling law. *Paleobiology* **2010**, *36*, 188–203. [CrossRef]
- 158. Mendoza, M.; Janis, C.M.; Palmqvist, P. Estimating the body mass of extinct ungulates: A study on the use of multiple regression. *J. Zool.* **2006**, *270*, 90–101. [CrossRef]
- 159. Saarinen, J.; Cirilli, O.; Strani, F.; Meshida, K.; Bernor, R.L. Testing equid body mass estimate equations on modern zebras—With implications to understanding the relationship of body size, diet, and habitats of *Equus* in the Pleistocene of Europe. *Front. Ecol. Evol.* **2021**, *9*, 622412. [CrossRef]
- 160. Gould, S.J. On the scaling of tooth size in mammals. Am. Zool. 1975, 15, 351–362. [CrossRef]
- 161. McNab, B. Complications inherent in scaling the basal rate of metabolism in mammals. *Q. Rev. Biol.* **1988**, *63*, 25–54. [CrossRef] [PubMed]
- 162. Foster, J.R. Preliminary body mass estimates for mammalian genera of the Morrison Formation (Upper Jurassic, North America). *PaleoBios* **2009**, *28*, 114–122.
- 163. Losey, R.J.; Osipov, B.; Sivakumaran, R.; Nomokonova, T.; Kovychev, E.V.; Diatchina, N.G. Estimating body mass in dogs and wolves using cranial and mandibular dimensions: Application to Siberian canids. *Int. J. Osteoarchaeol.* **2015**, 25, 946–959. [CrossRef]
- 164. Freudenthal, M.; Martín-Suárez, E. Estimating body mass of fossil rodents. Scr. Geol. 2013, 145, 1–130.
- 165. Gingerich, P.D.; Smith, B.H.; Rosenberg, K. Allometric scaling in the dentition of primates and prediction of body weight from tooth size in fossils. *Am. J. Phys. Anthropol.* **1982**, *58*, 81–100. [CrossRef] [PubMed]
- 166. Valkenburgh, B.V.; Wang, X.; Damuth, J. Cope's rule, hypercarnivory, and extinction in North American canids. *Science* **2004**, *306*, 101–104. [CrossRef]
- 167. MacFadden, B. Fossil horses from "Eohippus" (*Hyracotherium*) to *Equus*: Scaling, Cope's Law, and the evolution of body size. *Paleobiology* **1986**, 12, 355–369. [CrossRef]
- 168. Gordon, C.L. A first look at estimating body size in dentally conservative marsupials. J. Mamm. Evol. 2003, 10, 1–21. [CrossRef]
- 169. Creighton, G.K. Static allometry of mammalian teeth and the correlation of tooth size and body size in contemporary mammals. *J. Zool.* 1980, 191, 435–443. [CrossRef]
- 170. Legendre, S. Analysis of mammalian communities from the late Eocene and Oligocene of southern France. *Palaeovertebrata* **1986**, *16*, 191–212.
- 171. Legendre, S.; Roth, C. Correlation of carnassial tooth size and body weight in recent carnivores (Mammalia). *Hist. Biol.* **1988**, 1, 85–98. [CrossRef]
- 172. Morris, B.; Mead, A.J. Body mass estimates from bone and tooth measurements in white-tailed deer, *Odocoileus virginianus*. *Ga. J. Sci.* **2016**, *74*, 18.
- 173. Gingerich, P.D. Size variability of the teeth in living mammals and the diagnosis of closely related sympatric fossil species. *J. Paleontol.* **1974**, *48*, 895–903.
- 174. Miller, E.H.; Mahoney, S.P.; Kennedy, M.L.; Kennedy, P.K. Variation, sexual dimorphism, and allometry in molar size of the black bear. *J. Mammal.* **2009**, *90*, 491–503. [CrossRef]
- 175. Hopkins, S.S.B. Reassessing the mass of exceptionally large rodents using toothrow length and area as proxies for body mass. *J. Mammal.* 2008, 89, 232–243. [CrossRef]
- 176. Moncunill-Solé, B.; Jordana, X.; Marín-Moratalla, N.; Moyà-Solà, S.; Köhler, M. How large are the extinct giant insular rodents? New body mass estimations from teeth and bones. *Integr. Zool.* **2014**, *9*, 197–212. [CrossRef]
- 177. Dagosto, M.; Terranova, C.J. Estimating the body size of Eocene primates: A comparison of results from dental and postcranial variables. *Int. J. Primatol.* **1992**, *13*, 307. [CrossRef]
- 178. Lauer, C. The relationship of tooth size to body size in a population of rhesus monkeys (*Macaca mulatta*). *Am. J. Phys. Anthropol.* **1975**, 43, 333–339. [CrossRef]
- 179. Perez, V.J.; Leder, R.M.; Badaut, T. Body length estimation of Neogene macrophagous lamniform sharks (*Carcharodon* and *Otodus*) derived from associated fossil dentitions. *Palaeontol. Electron.* **2021**, *4*, a09. [CrossRef]

Animals **2022**, 12, 2038 27 of 30

180. Rubenstein, D.R.; Hobson, K.A. From birds to butterflies: Animal movement patterns and stable isotopes. *Trends Ecol. Evol.* **2004**, 19, 256–263. [CrossRef]

- 181. Hobson, K. Tracing origins and migration of wildlife using stable isotopes: A review. *Oecologia* **1999**, *120*, 314–326. [CrossRef] [PubMed]
- 182. Reitsema, L.J. Laboratory and field methods for stable isotope analysis in human biology. *Am. J. Hum. Biol.* **2015**, *27*, 593–604. [CrossRef] [PubMed]
- 183. Schuba, J.; Südekum, K.H.; Pfeffer, E.; Jayanegara, A. Excretion of faecal, urinary urea and urinary non-urea nitrogen by four ruminant species as influenced by dietary nitrogen intake: A meta-analysis. *Livest. Sci.* **2017**, *198*, 82–88. [CrossRef]
- 184. Lourenço, P.M.; Granadeiro, J.P.; Guilherme, J.L.; Catry, T. Turnover rates of stable isotopes in avian blood and toenails: Implications for dietary and migration studies. *J. Exp. Mar. Biol. Ecol.* **2015**, 472, 89–96. [CrossRef]
- 185. Bearhop, S.; Furness, R.W.; Hilton, G.M.; Votier, S.C.; Waldron, S. A forensic approach to understanding diet and habitat use from stable isotope analysis of (avian) claw material. *Funct. Ecol.* **2003**, *17*, 270–275. [CrossRef]
- 186. Ayliffe, L.K.; Cerling, T.E.; Robinson, T.; West, A.G.; Sponheimer, M.; Passey, B.H.; Hammer, J.; Roeder, B.; Dearing, M.D.; Ehleringer, J.R. Turnover of carbon isotopes in tail hair and breath CO<sub>2</sub> of horses fed an isotopically varied diet. *Oecologia* **2004**, 139, 11–22. [CrossRef]
- 187. Evans-Ogden, L.J.; Hobson, K.A.; Lank, D.B. Blood isotopic ( $\delta^{13}$ C and  $\delta^{15}$ N) turnover and diet-tissue fractionation factors in captive Dunlin (*Calidris Alpina Pacifica*). *Auk* **2004**, *121*, 170–177. [CrossRef]
- 188. Bauchinger, U.; McWilliams, S. Carbon turnover in tissues of a passerine bird: Allometry, isotopic clocks, and phenotypic flexibility in organ size. *Physiol. Biochem. Zool.* **2009**, *82*, 787–797. [CrossRef]
- 189. Madigan, D.J.; Snodgrass, O.E.; Hyde, J.R.; Dewar, H. Stable isotope turnover rates and fractionation in captive California yellowtail (*Seriola dorsalis*): Insights for application to field studies. *Sci. Rep.* **2021**, *11*, 4466. [CrossRef]
- 190. Hedges, R.E.; Clement, J.G.; Thomas, C.D.; O'Connell, T.C. Collagen turnover in the adult femoral mid-shaft: Modeled from anthropogenic radiocarbon tracer measurements. *Am. J. Phys. Anthropol.* **2007**, *133*, 808–816. [CrossRef]
- 191. Hobson, K.A.; Clark, R.G. Assessing avian diets using stable isotopes I: Turnover of <sup>13</sup>C in tissues. *Condor* **1992**, 94, 181–188. [CrossRef]
- 192. Matsubayashi, J.; Tayasu, I. Collagen turnover and isotopic records in cortical bone. J. Archaeol. Sci. 2019, 106, 37-44. [CrossRef]
- 193. Gage, J.P.; Francis, M.J.O.; Triffit, J.T. Collagen and Dental Matrices; Butterworth-Heinemann: London, UK, 1989.
- 194. Hoppe, K.A.; Koch, P.L.; Furutani, T.T. Assessing the preservation of biogenic strontium in fossil bones and tooth enamel. *Int. J. Osteoarchaeol.* **2003**, *13*, 20–28. [CrossRef]
- 195. Meier-Augenstein, W. Chapter III.5: Provenancing people. In *Stable Isotope Forensics: Methods and Forensic Applications of Stable Isotope Analysis*, 2nd ed.; Meier-Augenstein, W., Ed.; Wiley: Chichester, UK, 2018; pp. 333–400.
- 196. Beard, B.L.; Johnson, C.M. Strontium isotope composition of skeletal material can determine the birth place and geographic mobility of humans and animals. *J. Forensic Sci.* **2000**, *45*, 1049–1061. [CrossRef] [PubMed]
- 197. Hale, C.; Ogden, R.; Ciavaglia, S.A.; Cook, G.T.; Clarke, G.; Ogle, S.; Webster, L.M.I. Investigating the origins of ivory recovered in the United Kingdom. *Forensic Sci. Int. Anim. Environ.* **2021**, *1*, 100027. [CrossRef]
- 198. Wyatt, T.; Johnson, K.; Hunter, L.; George, R.; Gunter, R. Corruption and wildlife trafficking: Three case studies involving Asia. *Asian J. Criminol.* **2018**, *13*, 35–55. [CrossRef]
- 199. Alexander, J.; Downs, C.T.; Butler, M.; Woodborne, S.; Symes, C.T. Stable isotope analyses as a forensic tool to monitor illegally traded African grey parrots. *Anim. Conserv.* **2019**, 22, 134–143. [CrossRef]
- 200. Ziegler, S.; Merker, S.; Streit, B.; Boner, M.; Jacob, D.E. Towards understanding isotope variability in elephant ivory to establish isotopic profiling and source-area determination. *Biol. Conserv.* **2016**, *197*, 154–163. [CrossRef]
- 201. Van der Merwe, N.; Lee-Thorp, J.; Thackeray, J.; Hall-Martin, A.; Kruger, F.J.; Coetzee, H.; Bell, R.H.V.; Lindeque, M. Source-area determination of elephant ivory by isotopic analysis. *Nature* **1990**, *346*, 744–746. [CrossRef]
- 202. Coutu, A.N.; Lee-Thorp, J.; Collins, M.J.; Lane, P.J. Mapping the elephants of the 19th century East African ivory trade with a multi-isotope approach. *PLoS ONE* **2016**, *11*, e0163606. [CrossRef]
- 203. Coutu, A.N.; Whitelaw, G.; le Roux, P.; Sealy, J. Earliest evidence for the ivory trade in Southern Africa: Isotopic and ZooMS analysis of seventh–tenth century AD ivory from KwaZulu-Natal. *Afr. Archaeol. Rev.* **2016**, *33*, 411–435. [CrossRef]
- 204. Morcatty, T.Q.; Bausch-Macedo, J.C.; Nekaris, K.A.; Ni, Q.; Durigan, C.C.; Svensson, M.S.; Nijman, V. Illegal trade in wild cats and its link to Chinese-led development in Central and South America. *Conserv. Biol.* **2020**, *34*, 1525–1535. [CrossRef] [PubMed]
- 205. Nuñez, A.M.; Aliaga-Rossel, E. Jaguar fangs trafficking by Chinese in Bolivia. CAT News 2017, 65, 50-51.
- 206. Arias, M.; Hinsley, A.; Nogales-Ascarrunz, P.; Negroes, N.; Glikman, J.A.; Milner-Gulland, E.J. Prevalence and characteristics of illegal jaguar trade in north-western Bolivia. *Conserv. Sci. Pract.* **2021**, *3*, e444. [CrossRef]
- 207. Charity, S.; Ferreira, M. Wildlife trade in the Brazilian Amazon. In *Wildlife Trafficking in Brazil*; Charity, S., Ferreira, M., Eds.; TRAFFIC International: Cambridge, UK, 2020; pp. 59–93.
- 208. Munro, R.; Munro, H.M.C. (Eds.) Estimation of time since death. In *Animal Abuse and Unlawful Killing: Forensic Veterinary Pathology*; Saunders Elsevier: London, UK, 2008; pp. 88–93.
- 209. Brooks, J.W. Postmortem changes in animal carcasses and estimation of the postmortem interval. *Vet. Pathol.* **2016**, 53, 929–940. [CrossRef]

Animals 2022, 12, 2038 28 of 30

210. Brooks, J.W.; Sutton, L. Postmortem changes and estimating the postmortem interval. In *Veterinary Forensic Pathology*; Brooks, J.W., Ed.; Springer: Cham, Switzerland, 2018; pp. 43–63.

- 211. Erlandsson, M.; Munro, R. Estimation of the post-mortem interval in beagle dogs. Sci. Justice 2007, 47, 150–154. [CrossRef]
- 212. Brooks, J.W. Postmortem changes and the estimation of time since death. In *Veterinary Forensics: Investigation, Evidence Collection, and Expert Testimony*; Rogers, E., Stern, A.W., Eds.; CRC Press: Boca Raton, FL, USA, 2018; pp. 225–250.
- 213. Merck, M.D.; Miller, D.M. Postmortem changes and the postmortem interval. In *Veterinary Forensics: Animal Cruelty Investigations*, 2nd ed.; Merck, M.D., Ed.; Wiley-Blackwell: Chichester, UK, 2013; pp. 255–286.
- 214. Probst, C.; Gethmann, J.; Amendt, J.; Lutz, L.; Teifke, J.P.; Conraths, F.J. Estimating the postmortem interval of wild boar carcasses. *Vet. Sci.* **2020**, *7*, *6*. [CrossRef]
- 215. Ugrappa, S.; Jain, A. An emergence of dental tissues in the forensic medicine for the postmortem interval estimation: A scoping review. *J. Forensic Sci. Med.* **2021**, *7*, 54–60. [CrossRef]
- 216. Akbulut, N.; Çetin, S.; Bilecenoğlu, B.; Altan, A.; Akbulut, S.; Ocak, M.; Orhan, K. The micro-CT evaluation of enamel-cement thickness, abrasion, and mineral density in teeth in the postmortem interval (PMI): New parameters for the determination of PMI. *Int. J. Leg. Med.* **2020**, *134*, 645–653. [CrossRef]
- 217. Granrud, M.A.; Dabbs, G.R. A preliminary study of incisor exfoliation as an estimator of the postmortem interval using accumulated degree days. *Forensic Sci. Int.* **2012**, 220, e29–e32. [CrossRef]
- 218. Mehendiratta, M.; Jain, K.; Boaz, K.; Bansal, M.; Manaktala, N. Estimation of time elapsed since the death from identification of morphological and histological time-related changes in dental pulp: An observational study from porcine teeth. *J. Forensic Dent. Sci.* 2015, 7, 95–100. [CrossRef] [PubMed]
- 219. Young, S.T.; Wells, J.D.; Hobbs, G.R.; Bishop, C.P. Estimating postmortem interval using RNA degradation and morphological changes in tooth pulp. *Forensic Sci. Int.* **2013**, 229, 163.e1–163.e6. [CrossRef] [PubMed]
- 220. Schulz, I.; Schneider, P.M.; Olek, K.; Rothschild, M.A.; Tsokos, M. Examination of postmortem animal interference to human remains using cross-species multiplex PCR. *Forensic Sci. Med. Pathol.* **2006**, *2*, 95–101. [CrossRef]
- 221. Ubelaker, D.H.; DeGaglia, C.M. The impact of scavenging: Perspective from casework in forensic anthropology. *Forensic Sci. Res.* **2020**, *5*, 32–37. [CrossRef]
- 222. Bernstein, M. The nature of bitemarks. In *Bitemark Evidence: A Color Atlas and Text*, 2nd ed.; Dorion, R., Ed.; CRC Press: Boca Raton, FL, USA, 2011; pp. 53–65.
- 223. Binford, L.R. Bones: Ancient Men and Modern Myths; Academic Press: New York, NY, USA, 1981.
- 224. Tanga, C. Gli Inumati di San Pelino e Sant'Alessandro, Corfinio (AQ): Osteobiografia dei Resti Umani. Bachelor's Thesis, 'G. d'Annunzio' of Chieti-Pescara, Chieti, Italy, December 2016.
- 225. Moraitis, K.; Spiliopoulou, C. Forensic implications of carnivore scavenging on human remains recovered from outdoor locations in Greece. *J. Forensic Leg. Med.* **2010**, *17*, 298–303. [CrossRef]
- 226. Tsokos, M.; Schulz, F. Indoor postmortem animal interference by carnivores and rodents: Report of two cases and review of the literature. *Int. J. Leg. Med.* **1999**, *112*, 115–119. [CrossRef]
- 227. Keyes, C.A.; Myburgh, J.; Brits, D. Taphonomic bone trauma caused by southern African scavengers. *Int. J. Leg. Med.* **2020**, 134, 1227–1238. [CrossRef]
- 228. Colard, T.; Delannoy, Y.; Naji, S.; Gosset, D.; Hartnett, K.; Bécart, A. Specific patterns of canine scavenging in indoor settings. *J. Forensic Sci.* 2015, 60, 495–500. [CrossRef]
- 229. Young, A.; Márquez-Grant, N.; Stillman, R.; Smith, M.J.; Korstjens, A.H. An investigation of red fox (*Vulpes vulpes*) and Eurasian badger (*Meles meles*) scavenging, scattering, and removal of deer Remains: Forensic implications and applications. *J. Forensic Sci.* **2015**, *60*, S39–S55. [CrossRef]
- 230. Young, A.; Stillman, R.; Smith, M.J.; Korstjens, A.H. Scavenger species-typical alteration to bone: Using bite mark dimensions to identify scavengers. *J. Forensic Sci.* **2015**, *60*, 1426–1435. [CrossRef]
- 231. Young, A.; Stillman, R.; Smith, M.J.; Korstjens, A.H. Applying knowledge of species-typical scavenging behavior to the search and recovery of mammalian skeletal remains. *J. Forensic Sci.* **2016**, *61*, 458–466. [CrossRef] [PubMed]
- 232. Bellis, C.; Ashton, K.J.; Freney, L.; Blair, B.; Griffiths, L.R. A molecular genetic approach for forensic animal species identification. *Forensic Sci. Int.* **2003**, *134*, 99–108. [CrossRef]
- 233. Jones, A. Animal Scavengers as Agents of Decomposition: The Postmortem Succession of Louisiana Wildlife. Master's Thesis, Louisiana State University and Agricultural and Mechanical College, Baton Rouge, LA, USA, 2011.
- 234. Mansegosa, D.A.; Marchiori, J.I.; Giannotti, P.S. Dislocation, consumption and marks in human bodies produced by carnivores: A comparated study with forensic cases of the central-west of Argentina. *J. Forensic Anthropol. Odont.* **2020**, *3*, 6–17.
- 235. Steadman, D.W.; Dautartas, A.; Kenyhercz, M.W.; Jantz, L.M.; Mundorff, A.; Vidoli, G.M. Differential scavenging among pig, rabbit, and human subjects. *J. Forensic Sci.* **2018**, *63*, 1684–1691. [CrossRef]
- 236. Krenzer, U. Compendio de Métodos Antropológico Forenses Para la Reconstrucción del Perfil Osteo-Biológico; CAFCA, Centro de Análisis Forense y Ciencias Aplicadas: Ciudad de Guatemala, Guatemala, 2006; Volume 7.
- 237. Cantu, M. Animal Scavenging on Human Skeletal Remains in the Southwest United States: A Preliminary Model. Master's Thesis, Louisiana State University and Agricultural and Mechanical College, Baton Rouge, LA, USA, 2014.
- 238. Erkol, Z.; Hösükler, E. Postmortem animal attacks on human corpses. In *Post Mortem Examination and Autopsy: Current Issues from Death to Laboratory Analysis*; Dogan, K.H., Ed.; InTech: London, UK, 2018.

Animals 2022, 12, 2038 29 of 30

239. Andrés, M.; Gidna, A.O.; Yravedra, J.; Domínguez-Rodrigo, M. A study of dimensional differences of tooth Marks (pits and scores) on bones modified by small and large carnivores. *Archaeol. Anthropol. Sci.* **2012**, *4*, 209–219. [CrossRef]

- 240. Klippel, W.E.; Synstelien, J.A. Rodents as taphonomic agents: Bone gnawing by brown rats and gray squirrels. *J. Forensic Sci.* **2007**, 52, 765–773. [CrossRef]
- 241. Tsokos, M.; Matschke, J.; Gehl, A.; Koops, E.; Püschel, K. Skin and soft tissue artifacts due to postmortem damage caused by rodents. *Forensic Sci. Int.* **1999**, *104*, 47–57. [CrossRef]
- 242. Brothwell, D. Further evidence of bone chewing by ungulates: The sheep of North Ronaldsay, Orkney. *J. Archaeol. Sci.* **1976**, 3, 179–182. [CrossRef]
- 243. Kierdorf, U. Fork formation and other signs of osteophagia on a long bone swallowed by a red deer stag (*Cervus elaphus*). *Int. J. Osteoarchaeol.* **1993**, *3*, 37–40. [CrossRef]
- 244. Meckel, L.A.; McDaneld, C.P.; Wescott, D.J. White-tailed deer as a taphonomic agent: Photographic evidence of white-tailed deer gnawing on human Bone. *J. Forensic Sci.* 2018, 63, 292–294. [CrossRef]
- 245. Tumram, N.K.; Ambade, V.N.; Dixit, P.G. Human fatalities caused by animal attacks: A six-year autopsy study. *Med. Leg. J.* **2017**, 85, 194–199. [CrossRef] [PubMed]
- 246. Kiuchi, T.; Matoba, K.; Terazawa, K. Unusual injuries on the right hand and forearm caused by unidentified wild animals. *Leg. Med.* 2008, *10*, 301–305. [CrossRef] [PubMed]
- 247. Byard, R.W.; James, R.A.; Gilbert, J.D. Diagnostic problems associated with cadaveric trauma from animal activity. *Am. J. Forensic Med. Pathol.* **2002**, 23, 238–244. [CrossRef] [PubMed]
- 248. Fonseca, G.; Mora, E.; Lucena, J.; Cantin, M. Forensic studies of dog attacks on humans: A focus on bite mark analysis. *Res. Rep. Forensic Med. Sci.* **2015**, *5*, 39–51. [CrossRef]
- 249. De Munnynck, K.; Van de Voorde, W. Forensic approach of fatal dog attacks: A case report and literature review. *Int. J. Leg. Med.* **2002**, *116*, 295–300. [CrossRef]
- 250. Pomara, C.; D'Errico, S.; Jarussi, V.; Turillazzi, E.; Fineschi, V. Cave canem: Bite mark analysis in a fatal dog pack attack. *Am. J. Forensic Med. Pathol.* **2011**, 32, 50–54. [CrossRef]
- 251. Haj Salem, N.; Belhadj, M.; Aissaoui, A.; Mesrati, M.A.; Chadly, A. Multidisciplinary approach to fatal dog attacks: A forensic case study. *J. Forensic Leg. Med.* **2013**, *20*, 763–766. [CrossRef]
- 252. Le Garff, E.; Mesli, V.; Delannoy, Y.; Pollard, J.; Becart, A.; Hedouin, V. Domestic predation of an elder: A fatal dog attack case. *J. Forensic Sci.* 2017, 62, 1379–1382. [CrossRef]
- 253. Santoro, V.; Smaldone, G.; Lozito, P.; Smaldone, M.; Introna, F. A forensic approach to fatal dog attacks. A case study and review of the literature. *Forensic Sci. Int.* **2011**, 206, e37–e42. [CrossRef]
- 254. Marko, P.B. Sympatry. In *Encyclopedia of Ecology*; Jørgensen, S.E., Fath, B.D., Eds.; Academic Press: Amsterdam, The Netherlands, 2008; pp. 3450–3458.
- 255. Ohana, A.J.S. Intercanine Distance Used as a Measure to Rule Out Biters in Bite Mark Forensics. Master's Thesis, University of Tennessee: Knoxville, TN, USA, 2016.
- Põdra, M.; Gómez, A.; Palazón, S. Do American mink kill European mink? Cautionary message for future recovery efforts. Eur. J. Wildl. Res. 2013, 59, 431–440. [CrossRef]
- 257. Verzuh, T.; Bergman, D.L.; Bender, S.C.; Dwire, M.; Breck, S.W. Intercanine width measurements to aid predation investigations: A comparison between sympatric native and non-native carnivores in the Mexican wolf recovery area. *J. Mammal.* 2018, 99, 1405–1410. [CrossRef]
- 258. Ratz, H.; Moller, H. *Identification of Predators of Royal Albatross Chicks at Taiaroa Head in February* 1994; Conservation Advisory Science Notes No. 138; Department of Conservation: Wellington, New Zealand, 1997.
- 259. Elbroch, M.; McFarland, C. (Eds.) Interpreting prey remains. In *Mammal Tracks and Sign: A Guide to North American Species*, 2nd ed.; Stackpole Books: Guilford, CT, USA, 2019; pp. 307–376.
- 260. Knüsel, C.; Robb, J. Funerary taphonomy: An overview of goals and methods. J. Archaeol. Sci. Rep. 2016, 10, 655–673. [CrossRef]
- 261. Cyriac, V.P.; Kodandaramaiah, U. Digging their own macroevolutionary grave: Fossoriality as an evolutionary dead end in snakes. *J. Evol. Biol.* **2018**, *31*, 587–598. [CrossRef] [PubMed]
- 262. Morlan, R.E. Dialogue: Rodent bones in archaeological sites. J. Can. D'archéologie 1994, 18, 135–142.
- 263. Camarós, E.; Cueto, M.; Lorenzo, C.; Villaverde, V.; Rivals, F. Large carnivore attacks on hominins during the Pleistocene: A forensic approach with a Neanderthal example. *Archaeol. Anthropol. Sci.* **2016**, *8*, 635–646. [CrossRef]
- 264. Wolverton, S.; Nagaoka, L. Zooarcheology: Investigating past interactions between humans and other Animals. In *Ethnozoology: Animals in Our Lives*; Nóbrega-Alves, R.R., Albuquerque, U.P., Eds.; Academic Press: London, UK, 2018; pp. 25–43. [CrossRef]
- 265. Domínguez-Rodrigo, M.; Piqueras, A. The use of tooth pits to identify carnivore taxa in tooth-marked archaeofaunas and their relevance to reconstruct hominid carcass processing behaviours. *J. Archaeol. Sci.* **2003**, *30*, 1385–1391. [CrossRef]
- 266. Selvaggio, M.M.; Wilder, J. Identifying the involvement of multiple carnivore taxa with archaeological bone assemblages. *J. Archaeol. Sci.* **2001**, *28*, 465–470. [CrossRef]
- 267. Brain, C.K. New Finds at the Swartkrans Australopithecine Site. Nature 1970, 225, 1112–1119. [CrossRef]
- 268. Pobiner, B. Paleoecological information in predator tooth marks. J. Taphon. 2008, 6, 373–397.
- 269. Haynes, G. Evidence of carnivore gnawing on Pleistocene and recent mammalian bones. Paleobiology 1980, 6, 341–351. [CrossRef]

Animals 2022, 12, 2038 30 of 30

270. Haynes, G. A guide for differentiating mammalian carnivore taxa responsible for gnaw damage to herbivore limb bones. *Paleobiology* **1983**, *9*, 164–172. [CrossRef]

- 271. Pante, M.C.; Blumenschine, R.J.; Capaldo, S.D.; Scott, R.S. Validation of bone surface modification models for inferring fossil hominin and carnivore feeding interactions, with reapplication to FLK 22, Olduvai Gorge, Tanzania. *J. Hum. Evol.* 2012, 63, 395–407. [CrossRef] [PubMed]
- 272. Egeland, C.P.; Domínguez-Rodrigo, M. Taphonomic perspectives on hominid site use and foraging strategies during Bed II times at Olduvai Gorge, Tanzania. *J. Hum. Evol.* **2008**, *55*, 1031–1052. [CrossRef]
- 273. Njau, J.K.; Blumenschine, R.J. Crocodylian and mammalian carnivore feeding traces on hominid fossils from FLK 22 and FLK NN 3, Plio-Pleistocene, Olduvai Gorge, Tanzania. *J. Hum. Evol.* 2012, 63, 408–417. [CrossRef]
- 274. Daujeard, C.; Geraads, D.; Gallotti, R.; Lefèvre, D.; Mohib, A.; Raynal, J.-P.; Hublin, J.-J. Pleistocene hominins as a resource for carnivores: A c. 500,000-year-old human femur bearing tooth-marks in North Africa (Thomas Quarry I, Morocco). *PLoS ONE* **2016**, *11*, e0152284. [CrossRef] [PubMed]
- 275. Holloway, R.L.; Broadfield, D.C.; Yuan, M.S. The Human Fossil Record; Wiley-Liss: Hoboken, NJ, USA, 2004; Volume 3.
- 276. Singh, V.; Shukla, S.; Sing, A. The principal factors responsible for biodiversity loss. Open J. Plant. Sci. 2021, 6, 11–14. [CrossRef]