

Review



# Extranodal NK/T-Cell Lymphoma, Nasal Type: Genetic, Biologic, and Clinical Aspects with a Central Focus on Epstein–Barr Virus Relation

Miki Takahara <sup>1,\*</sup>, Takumi Kumai <sup>1,2</sup>, Kan Kishibe <sup>1</sup>, Toshihiro Nagato <sup>3</sup> and Yasuaki Harabuchi <sup>1</sup>

- <sup>1</sup> Department of Otolaryngology-Head and Neck Surgery, Asahikawa Medical University, Asahikawa 078-8510, Japan; t-kumai@asahikawa-med.ac.jp (T.K.); kkisibe@asahikawa-med.ac.jp (K.K.); hyasu@asahikawa-med.ac.jp (Y.H.)
- <sup>2</sup> Department of Innovative Head & Neck Cancer Research and Treatment (IHNCRT), Asahikawa Medical University, Asahikawa 078-8510, Japan
- <sup>3</sup> Department of Pathology, Asahikawa Medical University, Asahikawa 078-8510, Japan; rijun@asahikawa-med.ac.jp
- \* Correspondence: miki@asahikawa-med.ac.jp; Tel.: +81-166-68-2554; Fax: +81-166-68-2559

Abstract: Extranodal NK/T-Cell Lymphoma, nasal type (ENKTL-NT) has some salient aspects. The lymphoma is commonly seen in Eastern Asia, has progressive necrotic lesions in the nasal cavity, makes midfacial destructive lesions, and shows poor prognosis. The lymphoma cell is originated from either NK- or  $\gamma\delta$  T-cells, which express CD56. Since the authors first demonstrated the existence of Epstein-Barr virus (EBV) DNA and EBV oncogenic proteins in lymphoma cells, ENKTL-NT has been recognized as an EBV-associated malignancy. Because the angiocentric and polymorphous lymphoma cells are mixed with inflammatory cells on a necrotic background, the diagnosis of ENKTL-NT requires CD56 immunostaining and EBER in situ hybridization. In addition, serum the EBV DNA level is useful for the diagnosis and monitoring of ENKTL-NT. Although ENKTL-NT is refractory lymphoma, the prognosis is improved by the development of therapies such as concomitant chemoradiotherapy. The basic research reveals that a wide variety of intracellular/cell surface molecules, cytokines, chemokines, and micro RNAs are involved in lymphomagenesis, and some of them are related to EBV. Understanding lymphoma behavior introduces new therapeutic strategies, such as the usage of immune checkpoint inhibitors, peptide vaccines, and molecular targeting therapy. This review addresses recent advances in basic and clinical aspects of ENKTL-NT, especially its relation to EBV features.

**Keywords:** extranodal NK/T-cell lymphoma; nasal type; Epstein–Barr virus (EBV); EBV-encoded small nuclear early region (EBER)-1; latent membrane protein (LMP) 1; EBV DNA; MPVIC-P

# 1. Introduction

Extranodal NK/T-cell lymphoma, nasal type (ENKTL-NT) has some salient aspects. Patients with ENKTL-NT are commonly seen in Eastern Asia [1–4] and Latin America [4,5] but less frequently in the United States and Europe [6–8]. ENKTL-NT usually develops progressive necrotic granulation in the nasal cavity and shows a poor prognosis [2,9]. Histologically, ENKTL-NT is composed of angiocentric and polymorphous lymphoreticular infiltrate, previously called "polymorphic reticulosis" [10,11]. Original cells of ENKTL-NT are either NK- or  $\gamma\delta$  T-cells, both of which express CD56 [2,8,12–16]. In 1990, the authors first discovered the presence of Epstein–Barr virus (EBV) DNA and protein in the cells [1], and then found the EBV latency pattern and the clonality of the EBV genome [2,16,17]. In the present day, ENKTL-NT is recognized as EBV-related lymphoma [16]. This characteristic is used for diagnosis and monitoring after therapy for ENKTL-NT. For

**Citation:** Takahara, M.; Kumai, T.; Kishibe, K.; Nagato, T.; Harabuchi, Y. Extranodal NK/T-Cell Lymphoma, Nasal Type: Genetic, Biologic, and Clinical Aspects with a Central Focus on Epstein–Barr Virus Relation. *Microorganisms* **2021**, *9*, 1381. https://doi.org/10.3390/ microorganisms9071381

Academic Editor: Mario Clerici

Received: 12 June 2021 Accepted: 21 June 2021 Published: 25 June 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/). instance, the in situ hybridization of the EBV-encoded small nuclear early region (EBER)-1 is necessary for the pathological diagnosis of ENKTL-NT [18], and circulating EBV DNA levels are of great utility as highly sensitive tumor markers [19]. Although the prognosis of ENKTL-NT is poor, recent advancements in therapies, including concurrent chemoradiotherapy [20] and intra-arterial infusion chemoradiotherapy [21], improved the outcome of ENKTL-NT, especially in the early stages. Since two cell lines were established from ENKTL-NT tissues [13], basic research of ENKTL-NT has developed rapidly, and now we know that various intracellular/cell surface molecule, cytokines, chemokines, and micro RNAs are involved in the development of ENKTL-NT. More importantly, some growth-related factors could be a target of further therapies. Because EBV affects the development of ENKTL-NT via these factors, EBV proteins, such as latent membrane protein (LMP) 1, will be the target also. In this review article, the authors introduce the current understandings of ENKTL-NT with a particular focus on its relation to EBV.

#### 2. Historical context

In 1897, McBride et al. [22] first mentioned a disease condition of progressing necrotic granuloma in the nasal cavity resulting in a rapid invasion of the nose and face (midline). The patients followed an aggressive and lethal clinical course, and the disease was initially named "lethal midline granuloma" from the clinical characteristics [23,24]. Conversely, based on the pathological characteristics, the disease was called "polymorphic reticulosis" [11] or "angiocentric lymphoma" [10], which was characterized by diffuse infiltrates of pleomorphic and atypical large and small lymphoid cells with frequent mitosis admixing with a large number of inflammatory cells, such as granulocytes, macrophages, and plasma cells, with ischemic necrosis and angiocentric and/or angioinvasive infiltrates in the lesions. In the late 20th century, phenotypic studies [9,25,26] revealed that the tumor cells had NK-cell (CD56) and T-cell (CD3) markers. Accordingly, this lymphoma was identified as ENKTL-NT [25].

Etiologically, we [1] first reported the presence of EBV DNA and EBV-determined nuclear antigen 1 (EBNA1) in the lymphoma cells from 5 Japanese patients. EBV involvement was subsequently reported in China [27], the United States [28], France [8], and other western countries [29]. Thus, ENKTL-NT is now recognized as an EBV-associated malignancy [16].

#### 3. Epidemiology

There is a clear regional difference in ENKTL-NT prevalence. In Asia and South America, ENKTL-NT makes up 3–10% of non-Hodgkin lymphomas, whereas it makes up less than 1% in Western countries [30–33]. In addition, in the United States, it is estimated that ENKTL-NT represents approximately 1–2% of all T/NK-cell lymphomas and approximately 0.2% of all non-Hodgkin lymphomas [34]. The reason for the difference may be explained by a difference in race. The mongoloid race, which is popular in Asia and South America, may have the susceptibility gene of ENKTL-NT. Although the susceptible gene has not been discovered yet, relations to HLA loci have been reported in other EBV-associated malignancies, of which prevalence varies by region. For instance, HLA-A0207, which is common among Chinese people, is consistently associated with nasopharyngeal carcinoma (NPC) in Taiwan [35]. Moreover, meta-analyses in 13 published studies showed positive associations between NPC and the HLA-A2, B14, and B46 [36]. In EBV-positive Hodgkin lymphoma, HLA-A1 is reported to increase, and HLA-A2 decreases the risk of development [37].

Another explanation of the difference could be a relation to EBV. The existence of specific EBV strains or variants that electively infect NK/T-cells and stay in the cells by evading immunological surveillance may cause geographical distributions. However, there are few reports showing relationship-specific strain variants or mutants along with ENKTL-NT [18]. Unlike type 1 EBV, type 2 EBV can infect T-cells more effectively [38]; however, type 2 is not popular in East Asia, where T-/NK-cell malignancies are commonly

seen [39]. In a previous study, we [40,41] showed that the EBV gene extracted from ENKTL-NT tissues had several missense mutations of amino acids recognized by CD8<sup>+</sup> T-cells. Therefore, lymphoma cells infected with the mutated EBV may escape from cytolysis by immune cells.

Environmental factors are also possibly involved in the difference. Our group [42] has shown that exposure to pesticides and chemical solvents increased incidences of ENKTL-NT. In addition, Kojya et al. [43] reported familial ENKTL-NT with exposure to pesticides.

# 4. Clinical Features

The literature-based clinicopathological features are summarized in Table 1 [3,18,44– 47]. The peak of the incidences of lymphoma occurs at the middle age of 40–50 years old, and the male-to-female ratio is approximately 2:1. More than 70% of the patients are diagnosed at an early stage, based on the Ann Arbor classification. The lesion is initially found as a necrotic granuloma and ulceration in the nasal cavity, which is an extranodal site [1,2] (Figure 1a,b). The tumor easily invades surrounding tissues, including the palate (Figure 1c), the nasopharynx (Figure 1d), facial skin (Figure 1e,f), the paranasal sinus, and the orbits, resulting in the destruction of the midline facial structure [1,22,24].

The most frequent symptoms are nasal obstruction and bloody nasal discharge [2,18,45]. In addition, prolonged fevers, as systemic symptoms, are also commonly seen [2,45,48]. In our study with 62 patients with ENKTL-NT [18], nasal obstruction, bloody nasal discharge, and prolonged fevers were seen in 49 (70%), 29 (47%), and 32 (52%) patients, respectively (Table 1).

. .

able 1. Clinicopathological features of extrahodal NK/1-Cell lymphoma, hasal typ	e.

Country	Japan	Japan	China	Korea	Korea	Brazil
Year	2019	2010	2008	2006	2005	2011
Authors	Harabuchi et al [18]	Suzuki et al [44]	Wu et al [45]	Lee et al [3]	Kim et al [46]	Gualco et al [47]
Case number	62	123	115	262	114	122
Age						
Range (mean)	20-85 (53)	14-89 (52)			(47)	9-89 (45)
>60	22 (35%)		20 (18%)	55 (21%)	20 (18%)	
Sex						
Male/Female	43/19	81/42	78/29	170/92	72/42	85/37
Clinical stage						
I + II (%)	57 (92%)	84 (68%)	87 (76%)	200 (76%)	114 (100%)	25 (81%)
I/II/III/IV	44/13/1/4	55/29/8/31	61/26/8/12		83/31/0/0	23/2/2/4
Symptom						
Nasal obstruction	49 (70%)		84 (73%)			97 (80%)
Bloody rhinorrhea	29 (47%)		50 (44%)			
B symptom	32 (52%)	56 (46%)	57 (53%)	92 (35%)	35 (31%)	
Involved tissues						
Nasal cavity	60 (97%)	111 (90%)	115 (100%)		73 (64%)	97 (80%)
Hard plate	11 (18%)		8 (7%)		15 (13%)	
Facial skin	13 (21%)	19 (15%)				
Pharynx	13 (21%)	28 (23%)	27 (23%)		21 (18%)	
Lymph nodes	10 (16%)	31 (25%)	21 (18%)			
Skin	9 (15%)					
Lung/Liver	10 (16%)	10 (8%)		4 (2%)		
Digestive tracts	5 (8%)			10 (4%)		
Bone marrow	3 (5%)	9 (7%)	3 (3%)	16 (6%)		
Pathologic findings (Positive/Total cases)						
CD3	25/47 (53%)	68/86 (79%)	105/108 (97%)		104 (98%)	116/122 (95%)
CD43	31/35 (89%)	15/17 (88%)				
CD45RO	25/35 (71%)	44/49 (90%)	103/110 (94%)		61/62 (98%)	
CD20	0/59 (0%)	1/14 (7%)	0/115 (0%)		0/106 (0%)	
CD56	61/62 (98%)	115/120 (96%)	95/105 (91%)	262 (100%)	94/106 (89%)	103/122 (84%)
CD16	5/11 (45%)	9/40 (23%)				
EBER	59/62 (95%)	93/94 (99%)	106/110 (96%)	262 (100%)	46/61 (75%)	74/74 (100%)
LMP1	25/53 (47%)					10/122 (8%)
Gene rearrangement (Positive/Total cases)						

B cell receptor	0/34 (0%)	
T cell receptor	12/34 (35%)	7/74 (10%)



**Figure 1.** The representative local findings of extranodal NK/T-cell lymphoma, nasal type. (**a**) Granulation in the nasal cavity; (**b**) necrotic tissue in the nasal cavity; (**c**) ulceration of the hard palate; (**d**) necrotic granulation in the nasopharynx; (**e**,**f**) infiltration of the nasal skin.

# 5. Pathology

Pathologic findings of ENKTL-NT were known as diffuse infiltration of pleomorphic large and small lymphoma cells (Figure 2a), mixed with various inflammatory cells on the necrotic background [2,10,11]. The lymphoma cells express T-cell markers, such as cytoplasmic CD3 (CD3ε), as well as NK-cell marker CD56 (Figure 2b) [2,8,9,12–16]. Perforin, Fas ligand, and intercellular adhesion molecule-1 (ICAM-1) are also expressed in the ENKTL-NT cells [49].

In regard to the original cells of ENKTL-NT, there are two lineages: NK and T-cell [14,15]. We [2] and other groups [50,51] showed that T-cell receptor (TCR) gene rearrangement was proved in ENKTL-NT tissues in some patients. According to our investigation, ENKTL-NT tissue from 12 (35%) out of 34 patients had TCR gene rearrangement [18]. In fact, Nagata et al. [13] reported that two ENKTL-NT cell lines established from the tissues have different lineages: NK and T-cell.



**Figure 2.** Pathological findings of extranodal NK/T-cell lymphoma, nasal type. (**a**) Hematoxylin and eosin staining; (**b**) immunohistochemical staining of CD56; (**c**) in situ hybridization of EBV DNA; (**d**) double fluorescence staining with CD2 (red) and EBV-encoded nuclear antigen (EBNA) 1 (green); (**e**) immunohistochemical staining of EBV-encoded latent membrane protein (LMP) 1; (**f**) In situ hybridization of EBV-encoded small nuclear early region (EBER) 1.

#### 6. EBV Status

In 1990, we first showed the presence of EBV DNA and EBV nuclear antigen (EBNA1) in the lymphoma cells from 5 patients with ENKTL-NT [1] (Figure 2c,d). According to the knowledge of that time, EBV-related lymphomas were of B-cell lineage, such as Burkitt lymphoma. Therefore, the surprising result that EBV could infect NK or T-cells propounded a new concept of EBV-related malignancies. In the tissue section, the cells stained by EBNA1 were also stained with CD2, suggesting that EBV-infected cells were not surrounding B-cells (Figure 2d). After this report, the same findings were reported from Chinese and Western countries [8,27–29]. Our investigation, using in situ hybridization, revealed that EBER-1, which was generally expressed in cells infected with EBV, was detected in the section in 59 (95%) out of 62 patients with ENKTL-NT [18] (Figure 2f, Table 1). These results suggest that ENKTL-NT is an EBV-related lymphoma.

Proof of the clonotypic EBV genome [2] is important for the verification that EBV is infected with the original cells before tumorigenic transformation. The lymphoma cells express EBNA 1 (Figure 2d) and LMP1 (Figure 2e); this indicates that ENKTL-NT is categorized as a type II latency infection of EBV [2,52]. In regard to the expression of LMP1, we found LMP 1 mRNA in the tissues of all examined patients with ENKTL-NT, but the LMP1 protein in only half of the patients. This discrepancy may be explained by the methylation of LMP coding sequences [2,17,48]. Additionally, ENKTL-NT cells expressed LMP1 protein by immunohistochemical staining in 25 (47%) out of 53 patients with ENKTL-NT (Figure 2e, Table 1) [18].

In regard to the characteristics of EBV infecting the ENKTL-NT cells, the 30-bp deletion in the codon 343–352 of LMP1, which is famous for the B95-8 strain, was detected in the tissues of the vast majority of the patients [41,53]. Moreover, we detected several missense mutations in the epitope of the LMPs recognized by HLA-A2-restricted CTL [40,41]. Therefore, the infected EBV may have an ability to escape immune surveillance by CTLs, resulting in playing a role in lymphomagenesis.

#### 7. EBV Infection of T or NK Cells

In primary infection, EBV in saliva infects B-cells by binding gp350/220 to CD21 and gH/gL/gp42 to HLA class II molecules [54]. Because both CD21 and HLA class II are expressed on the surface of B cells, B cells are the main hosts of EBV. Moreover, EBV could also infect the mucosal epithelium of the oropharynx [55]. This infection requires attachments between BMRF2 or the gH/gL of EBV and integrins on the epithelial cells [56].

In regard to NK/T-cells, there are some reports showing that EBV infects them. EBERpositive NK/T-cells were found in tonsils with acute infectious mononucleosis [57,58]. However, the mechanism with which EBV infects NK/T-cells remains unclear. Basically, neither CD21 nor HLA class II express in NK/T-cells. On the other hand, NK/T-cells express some integrins, which may act as the EBV-receptors in NK/T-cells as well as epithelial cells [39]. Because CD21 is expressed in premature T-cells and common lymphoid progenitors [59], EBV can infect premature T-cells during intra-thymic maturation and human immature T-cell lines in vitro [60,61]. Interestingly, the fact that the infection of both CD4<sup>+</sup> and CD8<sup>+</sup> T-cells [62], as well as both T-cells and NK-cells [63], were seen in patients with chronic active EBV infection (CAEBV) supports the hypothesis that EBV may infect premature T-cells. More recently, Smith et al. [64] reported that CD21 was detected in mature peripheral CD3<sup>+</sup> T-cells by the anti-CD21 monoclonal antibody clone HB5 instead of the popular antibody clone Bly4. Moreover, a second minor strain of EBV, EBV type 2, could infect mature peripheral T-cells through HB5 antibody-detected CD21[64].

Another possible mechanism is that B cells or epithelial cells infected with EBV may contribute to the infection of NK/T-cells in a cell-to-cell contact manner. NK cells attacked autologous EBV-infected B cells, activated, and acquired the B-cell membrane, including CD21 molecules, by synaptic transfer [65]. Interestingly, the transferred CD21 has the capacity to catch a viral particle of EBV on the NK cell. However, Lee et al. [66] reported that EBV failed to infect an EBV-negative NK cell line through synaptic transferred CD21. They also showed that EBV genes but not RNA were detected in the NK cells by only treatment of the EB-viral supernatant. This condition is known as latency stage 0, in which viral gene expression is mostly suppressed [67]. Along with B-cells primarily infected with EBV, the condition may be necessary for the avoidance of immune surveillance and the persistence of EBV in NK cells.

#### 8. Gene Mutations

Genetic alternations, which also have pathogenic importance, have been reported in ENKTL-NT. For instance, deletion of the chromosome 6q21-25 was frequently seen in lymphoma tissues [68–71]. Gene mutations of apoptosis-related cell surface receptor Fas (Apo-1/CD95) were detected in the tissues of more than half of the patients [72,73]. Our group and others examined the mutations of major tumor-related genes, such as p53, K-ras, and c-kit [48,74–77]. According to the results, the p53 mutations were detected in 20–50% of patients; however, the K-ras, c-kit, and  $\beta$ -catenin mutations were rare. In addition, we [48] found that the p53 missense mutation was a factor that could predict poor survival.

# 9. Diagnosis by Using EBV Infection

It is difficult to diagnose ENKTL-NT by pathological examination with standard staining, such as hematoxylin-eosin, because of numerous necrotic backgrounds and mixtures of inflammatory cells [2]. Therefore, additional immunostaining, such as CD2, cytoplasmic CD3 $\varepsilon$ , CD56, cytotoxic molecules (perforin, granzyme B, and T-cell intracellular antigen 1) is needed for the diagnosis [2,18,78]. Of these molecular markers, CD56 is the most trustable marker for diagnosis because of a high positivity rate in the patients with ENKTL-NT (Table 1) [3,18,44–47].

Another promising method for diagnosis of the tissue sections is EBER in-situ hybridization. In the pathology laboratory of a general hospital, this procedure must be done by using commercial detection kits, which can be used for clinical diagnosis. Moreover, clear staining is usually obtained for the sections compared to immunohistochemical staining. The absence of EBV in the lymphoma cells excludes the diagnosis of ENKTL-NT according to a high positivity rate in the patients with ENKTL-NT (Table 1) [3,18,44–47]. Therefore, at least CD56 immunostaining and EBER in situ hybridization are required for diagnosis.

The differential diagnosis of ENKTL-NT can be judged by the common location of extranodal sites, especially the nasal cavity, the presence of EBER and CD56, and an elevated EBV DNA level, which will be mentioned later. It should be distinguished from other NK/T-cell lymphoproliferative diseases, such as blastoid NK-cell lymphomas, cutaneous NK/T-cell lymphomas, aggressive NK cell lymphomas, or chronic lymphoproliferative disorder. However, ENKTL-NT is originally a distinguishing disease, and we rarely have difficulty with a differential diagnosis.

#### 10. Staging

Computed tomography (CT) and magnetic resonance imaging (MRI) scans are used for the assessment of local lesions, the involvement of the lymph nodes, and distal metastasis. In addition, a bone marrow biopsy and a gastric fiberscope are needed for the evaluation of bone marrow and gastric involvement, respectively. According to these findings, stage stratification has been performed by the Ann Arbor classification. Currently, instead of the Ann Arbor classification, the Lugano classification is recommended for the staging system [79]. Because ENKTL-NT is thought to be an FDG-avid nodal lymphoma, PET/CT imaging is also required for the Lugano classification [79].

The Ann Arbor classification subdivides patients according to the absence (A) or presence (B) of disease-related symptoms, such as prolonged fever. As mentioned above, half of the patients with ENKTL-NT had prolonged fever and were subcategorized into the presence (B) category. However, this subdividing is excluded, except for Hodgkin lymphoma, in the Lugano classification [79]. In addition, in the Lugano classification, a bone marrow biopsy is no longer indicated for the routine staging of Hodgkin lymphoma and most diffuse, large B-cell lymphomas [79]. However, in other lymphomas, a conventional biopsy is allowed because of inadequate evidence [79].

# 11. Monitoring by Using EBV Infection

Circulating EBV DNA levels measured by RT-PCR are known as a sensitive tumor marker of EBV-associated malignancies, such as nasopharyngeal carcinoma [80]. Therefore, we measured the serum EBV DNA levels of BamHI W fragments and LMP1 in 20 patients with ENKTL-NT by quantitative real-time PCR [19]. Serum EBV DNA levels were detected at high levels in all patients, but the levels were under the limit of detection in all healthy controls. The levels decreased according to the treatment and increased at relapse. The Kaplan–Meier method and univariate analyses revealed that high DNA levels of BamHI W and LMP1 at pre-treatment and high BamHI W DNA levels at post-treatment were associated with short disease-free survival and overall survival. After our report, Suzuki, et al. [81] examined the relationship between pre-treatment plasma EBV DNA levels and several clinical factors, and showed that detectable plasma EBV DNA was associated with a higher clinical stage, the presence of B symptoms, worse performance status, and higher serum soluble IL-2 receptor levels. Moreover, they showed the clinical stage and pre-treatment plasma EBV DNA were significant prognostic factors by multivariate analysis. These data suggest that the periodic measurement of serum levels of EBV DNAs is useful for diagnosis, disease monitoring, and the prediction of prognosis.

Micro-RNAs (miR) are small noncoding RNAs that control gene expression [82], and some miRs are known to be released from the cells [83]. EBV has the ability to encode circulating viral miR. We [84] examined the availability of the serum EBV miR levels as biomarkers for ENKTL-NT. Accordingly, the serum levels of miR-BART2-5p, miR-BART7-3p, miR-BART13-3p, and miR-BART1-5p were higher in patients with ENKTL-NT and significantly decreased after treatment. Moreover, a high miR-BART2-5p level correlated with a poor prognosis. Thus, circulating EBV miRs, particularly miR-BART2-5p, may be another candidate for useful diagnostic and prognostic biomarkers in ENKTL-NT patients.

#### 12. Proliferation and Invasion Factors

Since the establishment of two EBV-positive ENKTL-NT cell lines SNK-6 and SNT-8 by Nagata et al. [13], the foundational investigation of ENKTL-NT has been developing rapidly. We [85–87] performed comprehensive array analyses in order to examine the gene expression patterns of these ENKTL-NT cell lines. Accordingly, several interesting molecules, such as the intracellular/cell surface molecule, cytokines, chemokines, and micro RNAs, were up- or downregulated, and some molecules were directly related to the lymphoma proliferation and invasion by additional in vivo and in vitro examination. These findings are schematically presented in Figure 3.

For example, IL-9 [85], soluble intercellular adhesion molecule-1 (sICAM) -1 [88], and hepatocyte growth factor (HGF) [89] were overproduced by SNK-6 and SNT-8 cells, and act as an autocrine growth factor. On the other hand, interferon-gamma-inducible protein-10 (IP-10) served as an autocrine invasion factor [86]. Cyclin-dependent kinase (CDK)1 and survivin were highly expressed in the cells and transmitted cell proliferation signals [90]. Conversely, micro RNA (miR)-15a was less expressed in the cells, and reduced antiproliferative signals [87]. In addition, other molecules were indirectly related to lymphoma proliferation in a paracrine manner. For example, hyper-produced IL-10 [91] increased the expression of CD25 (IL-2 receptor alpha) on the cells, resulting in an increased sensibility of IL-2 provided by the surrounding cells. CD70 [92] was highly expressed in the cells and induced the proliferation signal by the binding of the soluble CD27 (CD70 ligand) from bystander cells. Over-produced endogenous CCL2 and CCL22 [93], as well as IP-10 [86], attracted monocytes, which expressed membrane-bound IL-15 and induced a proliferation signal into SNK-6 in a cell-to-cell contact manner [94].



**Figure 3.** The biological characteristics of extranodal NK/T-cell lymphoma, nasal type cells. A wide variety of intracellular/cell surface molecules, cytokines, chemokines, and micro RNAs were involved in the lymphomagenesis of ENKTL-NT.

#### 13. Involvement of EBV in Proliferation and Invasion Factors

Some of these molecules were suggested to be influenced by EBV. We showed that the knockdown of LMP1 in SNK-6 and SNT-8 cells induced the downregulation of CDK1 and survivin [90]. The treatment of several CDK1 and survivin inhibitors inhibited cell proliferation of the cells in a dose-dependent manner. Moreover, the Sp1 inhibitor mithramycin, one of the CDK1 and survivin inhibitors, significantly suppressed the growth of established ENKTL-NT in a murine xenograft model. On the other hand, the knockdown induced the upregulation of miR-15a in SNK-6 and SNT-8 cells [87]. miRs are small noncoding RNAs that inhibit gene expression by ligating target mRNAs to repress translation, and they play a role in various biological processes, including development, differentiation, apoptosis, and cell proliferation [82]. MYB and cyclin D1, which are validated targets of miR-15a, were highly expressed in the cells by quantitative PCR and Western blot analysis. The forced expression of a precursor miR-15a in the cells leads to decreased expressions, resulting in the inhibition of the G1 = S transition and cell proliferation. Because immunohistochemical studies revealed that CDK, survivin, MYB, and cyclin D1 were expressed in ENKTL-NT cells in the tissue section, these machineries may take place in vivo. These findings suggest that LMP1 plays an important role in cell proliferation via CDK1, survivin, and miR-15a.

IL-9 is a multifunctional cytokine mainly produced by activated Th2 lymphocytes [95]. SNK-6 and SNT-8 produced IL-9 and expressed IL-9 receptors on the cell surfaces [85]. An anti-IL-9 neutralizing antibody inhibited the growth of the cells, whereas recombinant human IL-9 enhanced their growth, suggesting that an autocrine loop of IL-9 was involved in the cell proliferation [85]. Importantly, IL-9 mRNA was not expressed in other EBV-negative NK-cell and T-cell lymphoma/leukemia cell lines, suggesting that EBV may be related to the IL-9 expression of SNK-6 and SNT-8 [85]. In fact, EBV infection of MT-2 cell, a human T-cell line, reportedly enhanced IL-9 mRNA expression, and IL-9 promoter-luciferase reporter assay revealed that EBER was responsible for IL-9 expression [96].

ICAM-1 is known as a classic cell adhesion molecule and a natural ligand of lymphocyte function-associated antigen-1 (LFA-1) [97]. sICAM-1 is a soluble form of ICAM-1, and IFN- $\gamma$  induces a release of sICAM-1 by shedding membrane-bound ICAM-1[98]. We have already shown that serum sICAM-1 levels were higher in patients with ENKTL-NT than in patients with other lymphomas [49]. Both ICAM-1 and LFA-1 were expressed in several NK-cell lines regardless of EBV infection; however, sICAM-1 was detected in culture supernatant of only EBV-positive NK-cell lines, including SNK-6 [88]. As well as IL-9, cell proliferation assay under the treatment of sICAM-1 or anti-LFA-1 antibodies revealed that sICAM-1 increased the proliferation of SNK-6 in an autocrine manner [88]. Because LMP1 induced NF- $\kappa$ B-dependent IFN- $\gamma$  secretion in lymphoblastoid cell lines [99], LMP1 might enhance sICAM-1 release via IFN- $\gamma$ -induced proteolytic cleavage. This hypothesis is supported by the finding that SNK-6 cells produced a large amount of IFN- $\gamma$  [91], and that serum sICAM-1 levels were higher in patients with LMP1-positive ENKTL-NT than in those with LMP1-negative.

IP-10 is a chemokine that attracts human monocytes, activated T-cells, and NK cells expressing CXCR3 on the cell surface [100]. SNK-6 and SNT-8 produced IP-10 and expressed CXCR3 [86]. The treatment of anti-IP-10 neutralizing antibodies and recombinant IP-10 affected the cell invasion, and this showed that an autocrine loop of IP-10 was involved in the cell invasion [86]. The treatment did not affect the cell proliferation; how-ever, surrounding monocytes enhanced the proliferation of SNK-6 and SNT-8 cells in a cell-to-cell contact manner [94], suggesting that IP-10 also took a part in the cell proliferation, indirectly. IP-10 was not produced by EBV-negative NK-cell lines, and Vockerodt et al. [101] showed that LMP1 was sufficient for inducing IP-10 expression in an examination of LMP1-transfected Burkitt's and Hodgkin's lymphoma cell lines.

#### 14. Environmental Factors Affecting EBV Status

LMP1 is known to be important for the EBV-mediated transformation of B-lymphocytes [102]. Moreover, LMP1 acts as an oncoprotein because it can induce the transformation of rodent fibroblast cell lines [103]. According to our previous findings, as described above [85–88,90,94], LMP1 is thought to have a pivotal role in the lymphomagenesis of ENKTL-NT as well. At the point of regulation of LMP1 expression, EBNA-2 enhanced the expression by activating the LMP1 promotor in the type III latency cells, such as EBV immortalized B-lymphocytes [104]; however, the regulation system had unclear type II latency cell levels, which did not express EBNA-2. Recently, Kis, et al. [105] showed that the external stimuli, such as CD40-ligand and IL-4, could induce LMP1 in a Hodgkin lymphoma-derived cell line infected with EBV, without the expression of EBNA-2. Therefore, we examined whether LMP1 expression in SNK-6 was affected by the external stimuli. Accordingly, IFN- $\gamma$ , IL-2, IL-4, IL-10, and IL-15 increased the LMP1 expression without the induction of EBNA-2 [91]. In these cytokines, IL-10 enhanced the LMP1 expression the most strongly and quickly. IFN- $\gamma$  and IL-10 were detected in the supernatant of SNK-6 culture, and the treatment of blocking antibodies against these cytokines showed the downregulation of LMP1 expression. These findings suggest that IFN- $\gamma$  and IL-10 sustained the LMP1 expression of SNK-6 in an autocrine manner.

The external stimuli enhancing the LMP1 expression in ENKTL-NT cells are not only these cytokines. We co-cultured SNK-6 with granulocytes and monocytes and examined whether proliferation and the LMP1 expression of the cells changed. The proliferation and LMP1 expression of SNK-6 were enhanced by co-cultured monocytes but not by granulocytes [94]. On the other hand, these enhancements were not found when monocytes were placed in a separate chamber, suggesting that cell-to-cell contact was required for these behaviors. As a key surface molecule responsible for these behaviors, we focused on membrane-bound IL-15, which is known to express in the monocytes [106] because exogenous IL-15 enhanced the proliferation and LMP1 expression of SNK-6, as described above [91], and monocytic cells reportedly activated peripheral blood NK-cells through membranebound IL-15 [107]. In fact, co-cultured monocytes expressed membrane-bound IL-15 on the cell surface. Moreover, the treatment of an antibody against IL-15 inhibited the monocyte-inducible proliferation and LMP1 expression of SNK-6. Immunohistochemical analysis revealed that CD14-positive monocytes preferentially colocalized with CD56-positive lymphoma cells, and therefore, the interaction between the surrounding monocytes and the ENKTL-NT cells may take place in vivo.

#### 15. Therapy for Early Stage Extranodal NK/T-Cell Lymphoma, Nasal Type

For early-stage ENKTL-NT, DeVIC chemotherapy (dexamethasone, etoposide, ifosfamide, and carboplatin), concomitant with local radiotherapy (RT-2/3DeVIC), was conducted as a phase I/II trial (JCOG0211) in Japan and showed a good clinical outcome for ENKTL-NT [20]. A reason why ifosfamide and carboplatin are included in this regimen is that they are independent drugs of multidrug resistance (MDR) genes 1, which are expressed in ENKTL-NT cells [108]. Etoposide has the effect of avoiding the development of virus-associated hemophagocytic syndrome (VAHS) [109]. The disease prognosis was improved by the intervention of RT-2/3DeVIC, and the 2-year and 5-year overall survival (OS) rates were 78% [110] and 70% [111], respectively. According to these results, RT-2/3DeVIC is recognized as a standard therapy for early-stage ENKTL-NT in Japan [112]. Moreover, RT-2/3DeVIC is recommended in the NCCN (National Comprehensive Cancer Network) guideline as the preferred regimen of concurrent chemoradiation therapy [113]. As another candidate, the guideline also mentions CCRT-VIDL (concurrent cisplatin chemoradiation followed by etoposide, ifosfamide, dexamethasone, and l-asparaginase chemotherapy) [114] as another recommended regimen of concurrent chemoradiation therapy [113]. CCRT-VIDL achieved a 60% rate of 5-years of overall survival [114].

# 16. Therapy for Advanced Stage Extranodal NK/T-Cell Lymphoma, Nasal Type

For advanced-stage ENKTL-NT, the SMILE (steroid, methotrexate, ifosfamide, L-asparaginase, and etoposide) regimen was developed in Japan and East Asia [110]. L-asparaginase was thought to be essential in the regimen for the control of aggressive disease progression in spite of high toxicities [114]. In a phase II trial of SMILE [115] for patients who had newly diagnosed stage IV disease, complete remission (CR) and the 1-year OS rate were 45% and 55%, respectively, and superior to those of existing therapies [112]. According to these results, SMILE is recognized as a standard therapy for advanced-stage ENKTL-NT in Japan [112]. The NCCN guideline also suggests the AspaMetDex (L-asparaginase, methotrexate, and dexamethasone) regimen [116] as well as SMILE as a combination chemotherapy regimen [113]. A result of the AspaMetDex phase II study [116] showed around a 40% rate of 2-year OS.

Hematopoietic stem cell transplantation (HSCT) is another approach to treat advanced-stage NNKTL-NT. However, it is unclear which type of HSCT is the most appropriate [117]. The guidelines by the American Society for Blood and Marrow Transplantation support the use of both autologous and allogeneic HSCT for relapsed localized ENKL or as a front-line consolidation therapy for disseminated ENKTL-NT [118]. The NCCN guideline shows that HSCT is mainly suitable for the patients in remission of advancedstage NNKTL-NT after first-line therapy, but it does not indicate which HSCT is better because of poor evidence [113].

The age restrictions of each therapy for early and advanced stage ENKTL-NT are controversial. RT-2/3DeVIC and SMILE are reported to be indicated for patients under 70 years old [117]. This restriction may be referred from each clinical trial [110,119]. Hematology practical guidelines from the Society of Japan have no mention of the restriction [112]. The NCCN guideline shows that a new clinical trial or radiotherapy alone is recommended in patients unfit for chemotherapy with early-stage ENKTL-NT [113]. Although the definition of "unfit patient" is not shown [113], radiotherapy alone is thought to be realistic therapy for elderly patients with early-stage ENKTL-NT. A therapy for elderly patients with advanced-stage ENKTL-NT is less certain, and we should select better therapy that fits each patient, including the best supportive care.

# 17. Arterial Infusion Chemotherapy with Concomitant Radiotherapy

Recently, we [21] have reported a novel arterial infusion chemotherapy via a superficial temporal artery with concomitant radiotherapy for patients with early-stage ENKTL-NT. The regimen was composed of methotrexate, peplomycin, etoposide, ifosfamide, carboplatin, and prednisolone (MPVIC-P), which are independent of MDR 1, as well as a DeVIC regimen. In this report, 12 Japanese patients with stage I–II were enrolled, and all patients achieved complete remission and survived without relapse. Detailed therapeutic protocols and outcomes were described previously [21].

At present, 18 patients underwent this therapy [120]. The patients' information is summarized in Table 2. The patients' ages ranged from 21 to 79 years (a median of 64 years), and the number of males and females were 16 and 2, respectively. There were 6 patients who had systemic symptoms, including fevers, night sweats, and weight loss. Serum lactase dehydrogenase levels and serum soluble IL-2 receptor levels ranged from 143 to 626 IU/L, with a median level of 193 IU/L, and from 237 to 990 U/mL, with a median level of 420 U/mL, respectively. In 13 (72%) of the 18 patients, serum EBV DNA copy numbers (100–270,000 copies/mL) had been detected; the levels in these 13 patients were undetectable after the therapy. With regard to the course of therapy, one patient died with systemic relapse 30 months after the therapy, and one patient survived but suffered a relapse in the larynx 13 months after the therapy after the relapse, resulting in the disappearance of relapsed disease. The remaining 16 patients survived without relapse during the observation period from 26 to 111 months after the therapy (median: 73 months). For

all 18 patients, the 5-year overall and disease-free survival rates are 94% and 89%, respectively. Thus, intra-arterial infusion MPVIC-P chemoradiotherapy is an effective therapy for early-stage ENKTL-NT.

Case	2	Carl	SystemicP	erformanc	eClinica	1	LDH .			EBV-DNA (Copy/mL)		Observation Period	Outeene	
No Age		eGender	Symtom	Status	Stage	Lesioi	<sup>n</sup> (IU/l) <sup>s</sup>	IL-2K (U/ml)I	Kadiation (Gy)Kespons	Respons	e Before Treatmen	tAfter Treatmen	(Months)	Outcome
1	48	Female	+	1	Ι	NC	205	346	56	CR	391	<100	111	Disease free
2	60	Male	-	0	Ι	NC	236	345	56	CR	149	<100	107	Disease free
3	64	Male	-	0	Ι	NC	162	290	54	CR	120	<100	107	Disease free
4	48	Male	+	1	Ι	NC	176	447	54	CR	1640	<100	103	Disease free
5	40	Female	+	1	Ι	NC	144	604	54	CR	160	<100	100	Disease free
6	70	Male	-	0	Ι	NC	152	528	54	CR	100	<100	89	Disease free
7	21	Male	+	0	Ι	NC	177	529	54	CR	100	<100	73	Disease free
8	63	Male	-	0	Ι	NC	151	530	54	CR	<100	<100	72	Disease free
9	58	Male	-	0	II	NC	765	2410	54	CR	270,000	<100	68	Disease free
10	47	Male	-	0	Ι	NC	164	298	54	CR	<100	<100	48	Disease free
11	67	Male	+	0	Ι	NC	626	990	54	CR	62000	<100	42	Disease free
12	21	Male	+	0	II	NC	205	406	54	CR	550	<100	39	Disease free
13	67	Male	-	0	Ι	NC	293	580	54	CR	790	<100	84	Disease free
14	79	Male	-	0	Ι	NC	193	452	54	CR	<100	<100	30	Died with disease
15	68	Male	-	0	Ι	NC	143	237	54	CR	150	<100	48	Disease free
16	71	Male	-	0	Ι	NC	144	268	54	CR	<100	<100	40	Alive with disease
17	79	Male	-	0	Ι	NC	168	633	54	CR	<100	<100	36	Disease free
18	66	Male	-	0	Ι	NC	209	420	54	CR	450	<100	26	Disease free

**Table 2.** Overview of the 18 patients with extranodal NK/T-cell lymphoma, nasal type treated with arterial infusion MPVIC-P chemoradiotherapy.

NC: Nasal Cavity, CR: Complete Remission.

#### 18. Prospective Therapies

Understanding the proliferation signals in ENKTL-NT cells may make it possible to develop new therapies. Simvastatin, an inhibitor of HMG CoA reductase, is known to block the binding of ICAM-1 to LFA-1. In fact, we confirmed that simvastatin reduced the number of viable SNK-6 cells in vivo [88]. Mithramycin, an antibiotic with anti-tumor properties, downregulates both CDK1 and survivin. We confirmed that mithramycin significantly suppressed the growth of established ENKTL-NT in a murine xenograft model [90].

Highly expressed CD70 in SNK-6[92] can be a target of immunotherapy. We have shown that the anti-CD70 antibody mediated the effective complement-dependent killing of SNK-6 [92]. The antibody may have an additional effect by inhibiting the proliferation signal mediated by the CD27–CD70 interaction. The anti-CCR4 antibody mogamulizumab has antitumor activity against cutaneous T-cell lymphoma by antibody-dependent cell killing [121], and has been already applied in a clinical setting. We [93] have shown that CCR4 was expressed in the SNK-6 and ENKTL-NT cells in the tissues section. The anti-CCR4 antibody may be useful for therapy of ENKTL-NT as well as cutaneous T-cell lymphoma.

Programmed cell death-1 (PD-1) inhibitors elicit tumor inhibitory effects by the reduction of negative immunoregulating activity through the inhibition of the attachment of PD-1 to PD-L1. We have already used some, such as Nivolumab and Pembrolizumab, against head and neck cancer in health insurance treatment. Because we found the expression of PD-L1 on ENKTL-NT cells in the tissues section [122], the inhibitors might have the same effect on ENKTL-NT as well as head and neck cancer. In fact, Kwong et al. reported that pembrolizumab was highly effective for a patient with refractory ENKTL-NT [123].

HGF is an autocrine growth factor of SNK-6, as described above [89], and its receptor, c-Met, is known as a tumor-associated antigen (TAA) for CD8<sup>+</sup> cytotoxic T-cells (CTLs) [124]. Because CD4<sup>+</sup> Helper T-cells (HTLs) are important for the induction of efficacious antitumor immunity [125], we examined whether c-Met on SNK-6 acts as a TAA for HTLs as well as CTLs. C-Met contained several epitope peptides, which could induce various HLA-DR-restricted specific HTLs, and these peptide-induced HTL lines have a cytolytic ability to SNK-6 [89]. In addition, we found that c-Met inhibitor ARQ197 enhanced HTL recognition by decreasing the TGF- $\beta$  production by SNK-6. These results suggest that the combination of c-Met-targeted therapy and immunotherapy is a promising therapy for ENKTL-NT.

# 19. Prospective Therapies by Targeting EBV

Because EBV-related malignancies express non-self viral antigens recognized by immune cells [126], a peptide vaccine is a prospective therapy for ENKTL-NT. Demachi-Okamura et al. [127] made LMP1-specific CTLs from a healthy donor by using 43-amino acid N-terminal deletion mutant LMP1 (DeltaLMP1)-expressing APC. The CTL clone recognized a peptide of LMP1 presented by HLA-A\*0206 molecules. An EBV-infected NK cell line derived from a patient with chronic active EBV infection (CAEBV) was specifically lysed by the CTL.

We [128] previously found an epitope peptide, which could bind to promiscuous MHC Class II (HLA-DR9, HLA-DR53, or HLA-DR15), by a computer-based peptide algorithm from LMP1. This peptide was naturally processed and expressed in EBV-positive NK-cell lines including SNK-6 and could elicit peptide-specific HTL, which displayed the Th1 phenotype and cytotoxic activity against the cells. Because this LMP1 epitope peptide overlaps with an HLA-A2-restricted CTL epitope, this peptide might have the ability to simultaneously induce antitumor CTL and HTL cells against ENKTL-NT cells.

LMP2A could also be a target of immunotherapy in ENKTL-NT. Although ENKTL-NT cells do not express conventional LMP2A proteins and transcripts, Fox et al. [129] reported that novel LMP2 mRNA initiated from within the EBV terminal repeats was expressed in EBV-positive NK cell lines and that LMP2-specific CTLs recognized and killed the cells. The novel LMP2 mRNA was also detected in ENKTL-NT biopsy samples. Overall, these data suggest that immunotherapy targeting LMP against ENKTL-NT may serve as an alternative therapeutic modality.

# 20. Conclusions

We described the clinical picture, diagnosis, therapy, and future prospects taken from basic and translational research studies from the standpoint of ENKTL-NT as an EBV-related lymphoma. We are optimizing EBV for the diagnosis and monitoring of ENKTL-NT. Although the therapeutic approach for ENKTL-NT has improved, the outcome, especially in advanced stages, is still unsatisfactory. Therefore, the prospective treatments mentioned above, including EBV-targeting therapy, should be developed to the next stage for clinical use, and successive research studies are also required for the discovery of new treatment strategies. We believe that further investigation will allow for ENKTL-NT to be a curable disease.

**Author Contributions:** Conceptualization, M.T. and Y.H.; methodology, K.K.; software, K.K.; validation, T.N., T.K. and M.T.; formal analysis, M.T.; investigation, M.T., T.N., K.K. and T.K.; resources, M.T.; data curation, T.N.; writing–original draft preparation, T.N.; writing–review and Editing, T.K.; visualization, T.K.; supervision, Y.H.; project administration, Y.H.; funding acquisition, M.T., T.N., K.K., T.K. and Y.H. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was supported by JSPS KAKENHI, grant numbers 18H02948 (Yasuaki Harabuchi), 20K09745 (Miki Takahara), and 20K09724 (Takumi Kumai).

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Institutional Review Board of Asahikawa medical university (no 1332, 18 February 2013).

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: Data sharing is not applicable to this article.

Acknowledgments: We appreciate T.H. and A.K. for their encouragement and spiritual support, S.U., K.Y. and Y.K. for their support of many experimental trials, and all members belonging to the Department of Otolaryngology—Head and Neck Surgery, Asahikawa Medical University.

Conflicts of Interest: The authors declare no conflict of interest.

#### References

- Harabuchi, Y.; Yamanaka, N.; Kataura, A.; Imai, S.; Kinoshita, T.; Mizuno, F.; Osato, T. Epstein-Barr virus in nasal T-cell lymphomas in patients with lethal midline granuloma. *Lancet* 1990, 335, 128–130.
- Harabuchi, Y.; Imai, S.; Wakashima, J.; Hirao, M.; Kataura, A.; Osato, T.; Kon, S. Nasal T-cell lymphoma causally associated with Epstein-Barr virus: Clinicopathologic, phenotypic, and genotypic studies. *Cancer* 1996, 77, 2137–2149, doi:10.1002/(SICI)1097-0142(19960515)77:10<2137::AID-CNCR27>3.0.CO;2-V.
- Lee, J.; Suh, C.; Park, Y.H.; Ko, Y.H.; Bang, S.M.; Lee, J.H.; Lee, D.H.; Huh, J.; Oh, S.Y.; Kwon, H.C.; et al. Extranodal natural killer T-cell lymphoma, nasal-type: A prognostic model from a retrospective multicenter study. J. Clin. Oncol. 2006, 24, 612–618.
- 4. Aozasa, K.; Ohsawa, M.; Tajima, K.; Sasaki, R.; Maeda, H.; Matsunaga, T.; Friedmann, I. Nation-wide study of lethal mid-line granuloma in Japan: Frequencies of Wegener's granulomatosis, polymorphic reticulosis, malignant lymphoma and other related conditions. *Int. J. Cancer* **1989**, *44*, 63–66.
- Altemani, A.; Barbosa, A.C.; Kulka, M.; Takahashi, T.; Endo, L.; Vassallo, J.; Lorand-Metze, I. Characteristics of nasal T/NK-cell lymphoma among Brazilians. *Neoplasma* 2002, 49, 55–60.
- Gaal, K.; Sun, N.C.; Hernandez, A.M.; Arber, D.A. Sinonasal NK/T-cell lymphomas in the United States. *Am. J. Surg. Pathol.* 2000, 24, 1511–1517.
- Vidal, R.W.; Devaney, K.; Ferlito, A.; Rinaldo, A.; Carbone, A. Sinonasal malignant lymphomas: A distinct clinicopathological category. *Ann. Otol. Rhinol. Laryngol.* 1999, 108, 411–419.

- Kanavaros, P.; Lescs, M.C.; Briere, J.; Divine, M.; Galateau, F.; Joab, I.; Bosq, J.; Farcet, J.P.; Reyes, F.; Gaulard, P. Nasal T-cell lymphoma: A clinicopathologic entity associated with peculiar phenotype and with Epstein-Barr virus. *Blood* 1993, *81*, 2688– 2695.
- Yamanaka, N.; Kataura, A.; Sambe, S.; Minase, T.; Ishii, Y. Midfacial T cell lymphoma: Characterization by monoclonal antibodies. Ann. Otol. Rhinol. Laryngol. 1985, 94, 207–211.
- Harris, N.L.; Jaffe, E.S.; Stein, H.; Banks, P.M.; Chan, J.K.; Cleary, M.L.; Delsol, G.; De Wolf-Peeters, C.; Falini, B.; Gatter, K.C.; et al. A revised European-American classification of lymphoid neoplasms: A proposal from the International Lymphoma Study Group. *Blood* 1994, 84, 1361–1392.
- 11. Eichel, B.S.; Harrison, E.G.; Devine, K.D.; Scanlon, P.W.; Brown, H.A. Primary lymphoma of the nose including a relationship to lethal midline granuloma. *Am. J. Surg.* **1966**, *112*, 597–605.
- 12. Emile, J.F.; Boulland, M.L.; Haioun, C.; Kanavaros, P.; Petrella, T.; Delfau-Larue, M.H.; Bensussan, A.; Farcet, J.P.; Gaulard, P. CD5-CD56+ T-cell receptor silent peripheral T-cell lymphomas are natural killer cell lymphomas. *Blood* **1996**, *87*, 1466–1473.
- Nagata, H.; Konno, A.; Kimura, N.; Zhang, Y.; Kimura, M.; Demachi, A.; Sekine, T.; Yamamoto, K.; Shimizu, N. Characterization
  of novel natural killer (NK)-cell and gammadelta T-cell lines established from primary lesions of nasal T/NK-cell lymphomas
  associated with the Epstein-Barr virus. *Blood* 2001, 97, 708–713.
- 14. Kinney, M.C. The role of morphologic features, phenotype, genotype, and anatomic site in defining extranodal T-cell or NK-cell neoplasms. *Am. J. Clin. Pathol.* **1999**, *111*, S104–S118.
- Suzuki, R.; Takeuchi, K.; Ohshima, K.; Nakamura, S. Extranodal NK/T-cell lymphoma: Diagnosis and treatment cues. *Hematol.* Oncol. 2008, 26, 66–72, doi:10.1002/hon.847.
- Harabuchi, Y.; Takahara, M.; Kishibe, K.; Moriai, S.; Nagato, T.; Ishii, H. Nasal natural killer (NK)/T-cell lymphoma: Clinical, histological, virological, and genetic features. *Int. J. Clin. Oncol.* 2009, 14, 181–190, doi:10.1007/s10147-009-0882-7.
- Minarovits, J.; Hu, L.; Imai, S.; Harabuchi, Y.; Kataura, A.; Minarovits-Kormuta, S.; Osato, T.; Klein, G. Clonality, expression and methylation patterns of the Epstein-Barr virus genomes in lethal midline granulomas classified as peripheral angiocentric T cell lymphomas. *J. Gen. Virol.* 1994, 75, 77–84.
- Harabuchi, Y.; Takahara, M.; Kishibe, K.; Nagato, T.; Kumai, T. Extranodal Natural Killer/T-Cell Lymphoma, Nasal Type: Basic Science and Clinical Progress. Front. Pediatr. 2019, 7, 141, doi:10.3389/fped.2019.00141.
- Ishii, H.; Ogino, T.; Berger, C.; Kochli-Schmitz, N.; Nagato, T.; Takahara, M.; Nadal, D.; Harabuchi, Y. Clinical usefulness of serum EBV DNA levels of BamHI W and LMP1 for Nasal NK/T-cell lymphoma. J. Med. Virol. 2007, 79, 562–572, doi:10.1002/jmv.20853.
- Yamaguchi, M.; Ogawa, S.; Nomoto, Y. Treatment outcome of nasal NK-cell lymphoma: A case report of 12 consecutivelydiagnosed cases and a review of the literature. J. Clin. Exp. Hematopathol. 2001, 41, 93–99.
- Takahara, M.; Nagato, T.; Kishibe, K.; Ueda, S.; Komabayashi, Y.; Yamashina, M.; Takahashi, K.; Harabuchi, Y. Novel treatment for early-stage nasal natural killer/T-cell lymphoma: Intra-maxillary arterial infusion chemotherapy with concomitant radiotherapy. *Hematol. Oncol.* 2017, 35, 158–162, doi:10.1002/hon.2273.
- 22. McBride, P. Photographs of a case of rapid destruction of the nose and face. J. Laryngol. Otol. 1897, 12, 64-66.
- Williams, H.L. Lethal granulomatous ulceration, involving midline facial tissues. *Ann. Otol. Rhino. Laryngol.* 1949, *58*, 1013–1055.
   Harabuchi, Y.; Kataura, A.; Kobayashi, K.; Yamamoto, T.; Yamanaka, N.; Hirao, M.; Onodera, K.; Kon, S. Lethal midline granuloma (peripheral T-cell lymphoma) after lymphomatoid papulosis. *Cancer* 1992, *70*, 835–839.
- 25. Ng, C.S.; Chan, J.K.; Lo, S.T. Expression of natural killer cell markers in non-Hodgkin's lymphomas. *Hum. Pathol.* **1987**, *18*, 1257–1262.
- 26. Ishii, Y.; Yamanaka, N.; Ogawa, K.; Yoshida, Y.; Takami, T.; Matsuura, A.; Isago, H.; Kataura, A.; Kikuchi, K. Nasal T-cell lymphoma as a type of so-called "lethal midline granuloma". *Cancer* **1982**, *50*, 2336–2344.
- 27. Ho, F.C.; Srivastava, G.; Loke, S.L.; Fu, K.H.; Leung, B.P.; Liang, R.; Choy, D. Presence of Epstein-Barr virus DNA in nasal lymphomas of B and 'T' cell type. *Hematol. Oncol.* **1990**, *8*, 271–281.
- 28. Medeiros, L.J.; Jaffe, E.S.; Chen, Y.Y.; Weiss, L.M. Localization of Epstein-Barr viral genomes in angiocentric immunoproliferative lesions. *Am. J. Surg. Pathol.* **1992**, *16*, 439–447.
- Weiss, L.M.; Gaffey, M.J.; Chen, Y.-Y.; Frierson, H.F. Frequency of Epstein-Barr viral DNA in "Western" sinonasal and Waldeyer's ring non-Hodgkin's lymphoma. *Am. J. Surg. Pathol.* 1992, *16*, 156–162.
- 30. Liu, J.; Song, B.; Fan, T.; Huang, C.; Xie, C.; Li, J.; Zhong, W.; Li, S.; Yu, J. Pathological and clinical characteristics of 1,248 non-Hodgkin's lymphomas from a regional cancer hospital in Shandong, China. *Asian Pac. J. Cancer Prev.* **2011**, *12*, 3055–3061.
- 31. Vose, J.; Armitage, J.; Weisenburger, D.; International, T.C.L.P. International peripheral T-cell and natural killer/T-cell lymphoma study: Pathology findings and clinical outcomes. *J. Clin. Oncol.* **2008**, *26*, 4124–4130, doi:10.1200/JCO.2008.16.4558.
- Rudiger, T.; Weisenburger, D.D.; Anderson, J.R.; Armitage, J.O.; Diebold, J.; MacLennan, K.A.; Nathwani, B.N.; Ullrich, F.; Muller-Hermelink, H.K.; Non-Hodgkin's Lymphoma Classification, P. Peripheral T-cell lymphoma (excluding anaplastic large-cell lymphoma): Results from the Non-Hodgkin's Lymphoma Classification Project. *Ann. Oncol.* 2002, *13*, 140–149.
- 33. Lima, M. Aggressive mature natural killer cell neoplasms: From epidemiology to diagnosis. Orphanet J. Rare Dis. 2013, 8, 95, doi:10.1186/1750-1172-8-95.

- 34. Haverkos, B.M.; Pan, Z.; Gru, A.A.; Freud, A.G.; Rabinovitch, R.; Xu-Welliver, M.; Otto, B.; Barrionuevo, C.; Baiocchi, R.A.; Rochford, R.; et al. Extranodal NK/T Cell Lymphoma, Nasal Type (ENKTL-NT): An Update on Epidemiology, Clinical Presentation, and Natural History in North American and European Cases. *Curr. Hematol. Malig. Rep.* **2016**, *11*, 514–527, doi:10.1007/s11899-016-0355-9.
- Hildesheim, A.; Apple, R.J.; Chen, C.J.; Wang, S.S.; Cheng, Y.J.; Klitz, W.; Mack, S.J.; Chen, I.H.; Hsu, M.M.; Yang, C.S.; et al. Association of HLA class I and II alleles and extended haplotypes with nasopharyngeal carcinoma in Taiwan. *J. Natl. Cancer Inst.* 2002, 94, 1780–1789, doi:10.1093/jnci/94.23.1780.
- Goldsmith, D.B.; West, T.M.; Morton, R. HLA associations with nasopharyngeal carcinoma in Southern Chinese: A meta-analysis. *Clin. Otolaryngol. Allied Sci.* 2002, 27, 61–67.
- Niens, M.; Jarrett, R.F.; Hepkema, B.; Nolte, I.M.; Diepstra, A.; Platteel, M.; Kouprie, N.; Delury, C.P.; Gallagher, A.; Visser, L.; et al. HLA-A\*02 is associated with a reduced risk and HLA-A\*01 with an increased risk of developing EBV+ Hodgkin lymphoma. *Blood* 2007, *110*, 3310–3315, doi:10.1182/blood-2007-05-086934.
- Coleman, C.B.; Wohlford, E.M.; Smith, N.A.; King, C.A.; Ritchie, J.A.; Baresel, P.C.; Kimura, H.; Rochford, R. Epstein-Barr virus type 2 latently infects T cells, inducing an atypical activation characterized by expression of lymphotactic cytokines. *J. Virol.* 2015, *89*, 2301–2312, doi:10.1128/jvi.03001-14.
- 39. Kimura, H. EBV in T-/NK-Cell Tumorigenesis. Adv. Exp. Med. Biol. 2018, 1045, 459–475, doi:10.1007/978-981-10-7230-7\_21.
- Nagamine, M.; Kishibe, K.; Takahara, M.; Nagato, T.; Ishii, H.; Bandoh, N.; Ogino, T.; Harabuchi, Y. Selected amino acid change encoding Epstein-Barr virus-specific T cell epitope of the LMP2A gene in Japanese nasal NK/T cell lymphoma patients. *Intervirology* 2007, *50*, 319–322, doi:10.1159/000106462.
- 41. Nagamine, M.; Takahara, M.; Kishibe, K.; Nagato, T.; Ishii, H.; Bandoh, N.; Ogino, T.; Harabuchi, Y. Sequence variations of Epstein-Barr virus LMP1 gene in nasal NK/T-cell lymphoma. *Virus Genes* **2007**, *34*, 47–54, doi:10.1007/s11262-006-0008-5.
- 42. Xu, G.; Wang, H.; Xie, K.; He, G.; Du, Z. Analysis of clinicopathological features and prognostic factors of 62 nasal NK/T-cell lymphomas. *J. Clin. Otorhinolaryngol.* **2007**, *21*, 932–934.
- 43. Kojya, S.; Matsumura, J.; Ting, L.; Hongyo, T.; Inazawa, J.; Kirihata, M.; Aozasa, K. Familial nasal NK/T-cell lymphoma and pesticide use. *Am. J. Hematol.* **2001**, *66*, 145–147, doi:10.1002/1096-8652(200102)66:2<145::aid-ajh1033>3.0.co;2-v.
- Suzuki, R.; Suzumiya, J.; Yamaguchi, M.; Nakamura, S.; Kameoka, J.; Kojima, H.; Abe, M.; Kinoshita, T.; Yoshino, T.; Iwatsuki, K.; et al. Prognostic factors for mature natural killer (NK) cell neoplasms: Aggressive NK cell leukemia and extranodal NK cell lymphoma, nasal type. *Ann. Oncol.* 2010, *21*, 1032–1040, doi:10.1093/annonc/mdp418.
- 45. Wu, X.; Li, P.; Zhao, J.; Yang, X.; Wang, F.; Yang, Y.Q.; Fang, F.; Xu, Y.; Zhang, H.; Wang, W.Y.; et al. A clinical study of 115 patients with extranodal natural killer/T-cell lymphoma, nasal type. *Clin. Oncol.* **2008**, *20*, 619–625, doi:10.1016/j.clon.2008.05.011.
- Kim, T.M.; Park, Y.H.; Lee, S.Y.; Kim, J.H.; Kim, D.W.; Im, S.A.; Kim, T.Y.; Kim, C.W.; Heo, D.S.; Bang, Y.J.; et al. Local tumor invasiveness is more predictive of survival than International Prognostic Index in stage I(E)/II(E) extranodal NK/T-cell lymphoma, nasal type. *Blood* 2005, 106, 3785–3790, doi:10.1182/blood-2005-05-2056.
- Gualco, G.; Domeny-Duarte, P.; Chioato, L.; Barber, G.; Natkunam, Y.; Bacchi, C.E. Clinicopathologic and molecular features of 122 Brazilian cases of nodal and extranodal NK/T-cell lymphoma, nasal type, with EBV subtyping analysis. *Am. J. Surg. Pathol.* 2011, 35, 1195–1203, doi:10.1097/PAS.0b013e31821ec4b5.
- 48. Takahara, M.; Kishibe, K.; Bandoh, N.; Nonaka, S.; Harabuchi, Y. P53, N- and K-Ras, and beta-catenin gene mutations and prognostic factors in nasal NK/T-cell lymphoma from Hokkaido, Japan. *Hum. Pathol* **2004**, *35*, 86–95.
- 49. Harabuchi, Y.; Kataura, A.; Imai, K. Circulating intercellular adhesion molecule-1 and its cellular expression in head and neck non-Hodgkin's lymphomas, including lethal midline granuloma. *Ann. Otol. Rhinol. Laryngol.* **1996**, *105*, 634–642.
- Gaulard, P.; Henni, T.; Marolleau, J.P.; Haioun, C.; Henni, Z.; Voisin, M.C.; Divine, M.; Goossens, M.; Farcet, J.P.; Reyes, F. Lethal midline granuloma (polymorphic reticulosis) and lymphomatoid granulomatosis. Evidence for a monoclonal T-cell lymphoproliferative disorder. *Cancer* 1988, 62, 705–710.
- 51. Yoon, T.Y.; Lee, H.T.; Chang, S.H. Nasal-type T/natural killer cell angiocentric lymphoma, Epstein-Barr virus-associated, and showing clonal T-cell receptor gamma gene rearrangement. *Br. J. Dermatol.* **1999**, *140*, 505–508.
- 52. Chiang, A.K.; Tao, Q.; Srivastava, G.; Ho, F.C. Nasal NK- and T-cell lymphomas share the same type of Epstein-Barr virus latency as nasopharyngeal carcinoma and Hodgkin's disease. *Int. J. Cancer* **1996**, *68*, 285–290.
- 53. Kim, J.E.; Kim, Y.A.; Jeon, Y.K.; Park, S.S.; Heo, D.S.; Kim, C.W. Comparative analysis of NK/T-cell lymphoma and peripheral T-cell lymphoma in Korea: Clinicopathological correlations and analysis of EBV strain type and 30-bp deletion variant LMP1. *Pathol. Int.* **2003**, *53*, 735–743.
- 54. Hutt-Fletcher, L.M. Epstein-Barr virus entry. J. Virol. 2007, 81, 7825–7832, doi:10.1128/jvi.00445-07.
- 55. Borza, C.M.; Hutt-Fletcher, L.M. Alternate replication in B cells and epithelial cells switches tropism of Epstein-Barr virus. *Nat. Med.* **2002**, *8*, 594–599, doi:10.1038/nm0602-594.
- 56. Tugizov, S.M.; Berline, J.W.; Palefsky, J.M. Epstein-Barr virus infection of polarized tongue and nasopharyngeal epithelial cells. *Nat. Med.* **2003**, *9*, 307–314, doi:10.1038/nm830.
- Anagnostopoulos, I.; Hummel, M.; Kreschel, C.; Stein, H. Morphology, immunophenotype, and distribution of latently and/or productively Epstein-Barr virus-infected cells in acute infectious mononucleosis: Implications for the interindividual infection route of Epstein-Barr virus. *Blood* 1995, *85*, 744–750.
- 58. Hudnall, S.D.; Ge, Y.; Wei, L.; Yang, N.P.; Wang, H.Q.; Chen, T. Distribution and phenotype of Epstein-Barr virus-infected cells in human pharyngeal tonsils. *Mod. Pathol.* **2005**, *18*, 519–527, doi:10.1038/modpathol.3800369.

- 59. Fischer, E.; Delibrias, C.; Kazatchkine, M.D. Expression of CR2 (the C3dg/EBV receptor, CD21) on normal human peripheral blood T lymphocytes. *J. Immunol.* **1991**, *146*, 865–869.
- Paterson, R.L.; Kelleher, C.; Amankonah, T.D.; Streib, J.E.; Xu, J.W.; Jones, J.F.; Gelfand, E.W. Model of Epstein-Barr virus infection of human thymocytes: Expression of viral genome and impact on cellular receptor expression in the T-lymphoblastic cell line, HPB-ALL. *Blood* 1995, 85, 456–464.
- Fischer, E.M.; Mouhoub, A.; Maillet, F.; Frémeaux-Bacchi, V.; Krief, C.; Gould, H.; Berrih-Aknin, S.; Kazatchkine, M.D. Expression of CD21 is developmentally regulated during thymic maturation of human T lymphocytes. *Int. Immunol.* 1999, 11, 1841–1849, doi:10.1093/intimm/11.11.1841.
- Endo, R.; Yoshioka, M.; Ebihara, T.; Ishiguro, N.; Kikuta, H.; Kobayashi, K. Clonal expansion of multiphenotypic Epstein-Barr virus-infected lymphocytes in chronic active Epstein-Barr virus infection. *Med. Hypotheses* 2004, 63, 582–587, doi:10.1016/j.mehy.2004.03.012.
- Ohga, S.; Ishimura, M.; Yoshimoto, G.; Miyamoto, T.; Takada, H.; Tanaka, T.; Ohshima, K.; Ogawa, Y.; Imadome, K.; Abe, Y.; et al. Clonal origin of Epstein-Barr virus (EBV)-infected T/NK-cell subpopulations in EBV-positive T/NK-cell lymphoproliferative disorders of childhood. *J. Clin. Virol.* 2011, *51*, 31–37, doi:10.1016/j.jcv.2011.01.014.
- 64. Smith, N.A.; Coleman, C.B.; Gewurz, B.E.; Rochford, R. CD21 (Complement Receptor 2) Is the Receptor for Epstein-Barr Virus Entry into T Cells. J. Virol. 2020, 94, doi:10.1128/jvi.00428-20.
- Tabiasco, J.; Vercellone, A.; Meggetto, F.; Hudrisier, D.; Brousset, P.; Fournié, J.J. Acquisition of viral receptor by NK cells through immunological synapse. J. Immunol. 2003, 170, 5993–5998, doi:10.4049/jimmunol.170.12.5993.
- Lee, J.H.; Choi, J.; Ahn, Y.O.; Kim, T.M.; Heo, D.S. CD21-independent Epstein-Barr virus entry into NK cells. Cell Immunol. 2018, 327, 21–25, doi:10.1016/j.cellimm.2018.01.011.
- 67. Joly, E.; Hudrisier, D. What is trogocytosis and what is its purpose? Nat. Immunol. 2003, 4, 815, doi:10.1038/ni0903-815.
- Tien, H.F.; Su, I.J.; Tang, J.L.; Liu, M.C.; Lee, F.Y.; Chen, Y.C.; Chuang, S.M. Clonal chromosomal abnormalities as direct evidence for clonality in nasal T/natural killer cell lymphomas. *Br. J. Haematol.* 1997, 97, 621–625.
- 69. Wong, K.F.; Chan, J.K.; Kwong, Y.L. Identification of del(6)(q21q25) as a recurring chromosomal abnormality in putative NK cell lymphoma/leukaemia. *Br. J. Haematol.* **1997**, *98*, 922–926.
- Siu, L.L.; Chan, V.; Chan, J.K.; Wong, K.F.; Liang, R.; Kwong, Y.L. Consistent patterns of allelic loss in natural killer cell lymphoma. *Am. J. Pathol.* 2000, 157, 1803–1809, doi:10.1016/s0002-9440(10)64818-3.
- Sun, H.S.; Su, I.J.; Lin, Y.C.; Chen, J.S.; Fang, S.Y. A 2.6 Mb interval on chromosome 6q25.2-q25.3 is commonly deleted in human nasal natural killer/T-cell lymphoma. *Br. J. Haematol.* 2003, 122, 590–599.
- 72. Takakuwa, T.; Dong, Z.; Nakatsuka, S.; Kojya, S.; Harabuchi, Y.; Yang, W.I.; Nagata, S.; Aozasa, K. Frequent mutations of Fas gene in nasal NK/T cell lymphoma. *Oncogene* **2002**, *21*, 4702–4705, doi:10.1038/sj.onc.1205571.
- Shen, L.; Liang, A.C.; Lu, L.; Au, W.Y.; Kwong, Y.L.; Liang, R.H.; Srivastava, G. Frequent deletion of Fas gene sequences encoding death and transmembrane domains in nasal natural killer/T-cell lymphoma. *Am. J. Pathol.* 2002, 161, 2123–2131, doi:10.1016/s0002-9440(10)64490-2.
- 74. Li, T.; Hongyo, T.; Syaifudin, M.; Nomura, T.; Dong, Z.; Shingu, N.; Kojya, S.; Nakatsuka, S.; Aozasa, K. Mutations of the p53 gene in nasal NK/T-cell lymphoma. *Lab. Investig.* **2000**, *80*, 493–499.
- Hoshida, Y.; Hongyo, T.; Jia, X.; He, Y.; Hasui, K.; Dong, Z.; Luo, W.J.; Ham, M.F.; Nomura, T.; Aozasa, K. Analysis of p53, K-ras, c-kit, and beta-catenin gene mutations in sinonasal NK/T cell lymphoma in northeast district of China. *Cancer Sci.* 2003, 94, 297–301.
- 76. Quintanilla-Martinez, L.; Kremer, M.; Keller, G.; Nathrath, M.; Gamboa-Dominguez, A.; Meneses, A.; Luna-Contreras, L.; Cabras, A.; Hoefler, H.; Mohar, A.; et al. p53 Mutations in nasal natural killer/T-cell lymphoma from Mexico: Association with large cell morphology and advanced disease. *Am. J. Pathol.* 2001, *159*, 2095–2105, doi:10.1016/s0002-9440(10)63061-1.
- 77. Hongyo, T.; Hoshida, Y.; Nakatsuka, S.; Syaifudin, M.; Kojya, S.; Yang, W.I.; Min, Y.H.; Chan, H.; Kim, C.H.; Harabuchi, Y.; et al. p53, K-ras, c-kit and beta-catenin gene mutations in sinonasal NK/T-cell lymphoma in Korea and Japan. *Oncol. Rep.* **2005**, *13*, 265–271.
- Tse, E.; Kwong, Y.L. The diagnosis and management of NK/T-cell lymphomas. J. Hematol. Oncol. 2017, 10, 85, doi:10.1186/s13045-017-0452-9.
- Cheson, B.D.; Fisher, R.I.; Barrington, S.F.; Cavalli, F.; Schwartz, L.H.; Zucca, E.; Lister, T.A. Recommendations for initial evaluation, staging, and response assessment of Hodgkin and non-Hodgkin lymphoma: The Lugano classification. *J. Clin. Oncol.* 2014, *32*, 3059–3068, doi:10.1200/jco.2013.54.8800.
- 80. Lei, K.I.; Chan, L.Y.; Chan, W.Y.; Johnson, P.J.; Lo, Y.M. Quantitative analysis of circulating cell-free Epstein-Barr virus (EBV) DNA levels in patients with EBV-associated lymphoid malignancies. *Br. J. Haematol.* **2000**, *111*, 239–246.
- Suzuki, R.; Yamaguchi, M.; Izutsu, K.; Yamamoto, G.; Takada, K.; Harabuchi, Y.; Isobe, Y.; Gomyo, H.; Koike, T.; Okamoto, M.; et al. Prospective measurement of Epstein-Barr virus-DNA in plasma and peripheral blood mononuclear cells of extranodal NK/T-cell lymphoma, nasal type. *Blood* 2011, *118*, 6018–6022, doi:10.1182/blood-2011-05-354142.
- 82. Bartel, D.P. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* **2004**, *116*, 281–297, doi:10.1016/s0092-8674(04)00045-5.
- Valadi, H.; Ekström, K.; Bossios, A.; Sjöstrand, M.; Lee, J.J.; Lötvall, J.O. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat. Cell Biol.* 2007, *9*, 654–659, doi:10.1038/ncb1596.

- 84. Komabayashi, Y.; Kishibe, K.; Nagato, T.; Ueda, S.; Takahara, M.; Harabuchi, Y. Circulating Epstein-Barr virus-encoded micro-RNAs as potential biomarkers for nasal natural killer/T-cell lymphoma. *Hematol. Oncol.* **2017**, *35*, 655–663, doi:10.1002/hon.2360.
- Nagato, T.; Kobayashi, H.; Kishibe, K.; Takahara, M.; Ogino, T.; Ishii, H.; Oikawa, K.; Aoki, N.; Sato, K.; Kimura, S.; et al. Expression of interleukin-9 in nasal natural killer/T-cell lymphoma cell lines and patients. *Clin. Cancer Res.* 2005, 11, 8250–8257, doi:10.1158/1078-0432.CCR-05-1426.
- Moriai, S.; Takahara, M.; Ogino, T.; Nagato, T.; Kishibe, K.; Ishii, H.; Katayama, A.; Shimizu, N.; Harabuchi, Y. Production of interferon-{gamma}-inducible protein-10 and its role as an autocrine invasion factor in nasal natural killer/T-cell lymphoma cells. *Clin. Cancer Res.* 2009, 15, 6771–6779, doi:10.1158/1078-0432.CCR-09-1052.
- Komabayashi, Y.; Kishibe, K.; Nagato, T.; Ueda, S.; Takahara, M.; Harabuchi, Y. Downregulation of miR-15a due to LMP1 promotes cell proliferation and predicts poor prognosis in nasal NK/T-cell lymphoma. *Am. J. Hematol.* 2014, *89*, 25–33, doi:10.1002/ajh.23570.
- Takahara, M.; Nagato, T.; Komabayashi, Y.; Yoshino, K.; Ueda, S.; Kishibe, K.; Harabuchi, Y. Soluble ICAM-1 secretion and its functional role as an autocrine growth factor in nasal NK/T cell lymphoma cells. *Exp. Hematol.* 2013, 41, 711–718, doi:10.1016/j.exphem.2013.03.009.
- Kumai, T.; Matsuda, Y.; Ohkuri, T.; Oikawa, K.; Ishibashi, K.; Aoki, N.; Kimura, S.; Harabuchi, Y.; Celis, E.; Kobayashi, H. c-Met is a novel tumor associated antigen for T-cell based immunotherapy against NK/T cell lymphoma. *Oncoimmunology* 2015, 4, e976077, doi:10.4161/2162402x.2014.976077.
- Nagato, T.; Ueda, S.; Takahara, M.; Kishibe, K.; Komabayashi, Y.; Kumai, T.; Ohara, K.; Hirata-Nozaki, Y.; Harabuchi, S.; Hayashi, R.; et al. Cyclin-dependent kinase 1 and survivin as potential therapeutic targets against nasal natural killer/T-cell lymphoma. *Lab. Investig.* 2019, 99, 612–624, doi:10.1038/s41374-018-0182-9.
- Takahara, M.; Kis, L.L.; Nagy, N.; Liu, A.; Harabuchi, Y.; Klein, G.; Klein, E. Concomitant increase of LMP1 and CD25 (IL-2-receptor alpha) expression induced by IL-10 in the EBV-positive NK lines SNK6 and KAI3. *Int. J. Cancer* 2006, 119, 2775–2783, doi:10.1002/ijc.22139.
- Yoshino, K.; Kishibe, K.; Nagato, T.; Ueda, S.; Komabayashi, Y.; Takahara, M.; Harabuchi, Y. Expression of CD70 in nasal natural killer/T cell lymphoma cell lines and patients; its role for cell proliferation through binding to soluble CD27. *Br. J. Haematol.* 2013, *160*, 331–342, doi:10.1111/bjh.12136.
- Kumai, T.; Nagato, T.; Kobayashi, H.; Komabayashi, Y.; Ueda, S.; Kishibe, K.; Ohkuri, T.; Takahara, M.; Celis, E.; Harabuchi, Y. CCL17 and CCL22/CCR4 signaling is a strong candidate for novel targeted therapy against nasal natural killer/T-cell lymphoma. *Cancer Immunol. Immunother.* 2015, 64, 697–705, doi:10.1007/s00262-015-1675-7.
- Ishii, H.; Takahara, M.; Nagato, T.; Kis, L.L.; Nagy, N.; Kishibe, K.; Harabuchi, Y.; Klein, E. Monocytes enhance cell proliferation and LMP1 expression of nasal natural killer/T-cell lymphoma cells by cell contact-dependent interaction through membranebound IL-15. *Int. J. Cancer* 2012, 130, 48–58, doi:10.1002/ijc.25969.
- 95. Demoulin, J.B.; Renauld, J.C. Interleukin 9 and its receptor: An overview of structure and function. *Int. Rev. Immunol.* **1998**, *16*, 345–364, doi:10.3109/08830189809043001.
- Yang, L.; Aozasa, K.; Oshimi, K.; Takada, K. Epstein-Barr virus (EBV)-encoded RNA promotes growth of EBV-infected T cells through interleukin-9 induction. *Cancer Res.* 2004, 64, 5332–5337.
- Marlin, S.D.; Springer, T.A. Purified intercellular adhesion molecule-1 (ICAM-1) is a ligand for lymphocyte function-associated antigen 1 (LFA-1). *Cell* 1987, *51*, 813–819, doi:10.1016/0092-8674(87)90104-8.
- Becker, J.C.; Dummer, R.; Hartmann, A.A.; Burg, G.; Schmidt, R.E. Shedding of ICAM-1 from human melanoma cell lines induced by IFN-gamma and tumor necrosis factor-alpha. Functional consequences on cell-mediated cytotoxicity. *J. Immunol.* 1991, 147, 4398–4401.
- Najjar, I.; Baran-Marszak, F.; Le Clorennec, C.; Laguillier, C.; Schischmanoff, O.; Youlyouz-Marfak, I.; Schlee, M.; Bornkamm, G.W.; Raphaël, M.; Feuillard, J.; et al. Latent membrane protein 1 regulates STAT1 through NF-kappaB-dependent interferon secretion in Epstein-Barr virus-immortalized B cells. J. Virol. 2005, 79, 4936–4943, doi:10.1128/jvi.79.8.4936-4943.2005.
- 100. Taub, D.D.; Lloyd, A.R.; Conlon, K.; Wang, J.M.; Ortaldo, J.R.; Harada, A.; Matsushima, K.; Kelvin, D.J.; Oppenheim, J.J. Recombinant human interferon-inducible protein 10 is a chemoattractant for human monocytes and T lymphocytes and promotes T cell adhesion to endothelial cells. J. Exp. Med. 1993, 177, 1809–1814, doi:10.1084/jem.177.6.1809.
- Vockerodt, M.; Pinkert, D.; Smola-Hess, S.; Michels, A.; Ransohoff, R.M.; Tesch, H.; Kube, D. The Epstein-Barr virus oncoprotein latent membrane protein 1 induces expression of the chemokine IP-10: Importance of mRNA half-life regulation. *Int. J. Cancer* 2005, 114, 598–605, doi:10.1002/ijc.20759.
- Kaye, K.M.; Izumi, K.M.; Kieff, E. Epstein-Barr virus latent membrane protein 1 is essential for B-lymphocyte growth transformation. Proc. Natl. Acad. Sci. USA 1993, 90, 9150–9154, doi:10.1073/pnas.90.19.9150.
- 103. Wang, D.; Liebowitz, D.; Kieff, E. An EBV membrane protein expressed in immortalized lymphocytes transforms established rodent cells. *Cell* **1985**, *43*, 831–840, doi:10.1016/0092-8674(85)90256-9.
- 104. Sjöblom, A.; Nerstedt, A.; Jansson, A.; Rymo, L. Domains of the Epstein-Barr virus nuclear antigen 2 (EBNA2) involved in the transactivation of the latent membrane protein 1 and the EBNA Cp promoters. *J. Gen. Virol.* **1995**, *76 Pt 11*, 2669–2678, doi:10.1099/0022-1317-76-11-2669.
- 105. Kis, L.L.; Nishikawa, J.; Takahara, M.; Nagy, N.; Matskova, L.; Takada, K.; Elmberger, P.G.; Ohlsson, A.; Klein, G.; Klein, E. In vitro EBV-infected subline of KMH2, derived from Hodgkin lymphoma, expresses only EBNA-1, while CD40 ligand and IL-4 induce LMP-1 but not EBNA-2. *Int. J. Cancer* 2005, *113*, 937–945, doi:10.1002/ijc.20654.

- Musso, T.; Calosso, L.; Zucca, M.; Millesimo, M.; Ravarino, D.; Giovarelli, M.; Malavasi, F.; Ponzi, A.N.; Paus, R.; Bulfone-Paus, S. Human monocytes constitutively express membrane-bound, biologically active, and interferon-gamma-upregulated interleukin-15. *Blood* 1999, 93, 3531–3539.
- 107. González-Alvaro, I.; Domínguez-Jiménez, C.; Ortiz, A.M.; Núñez-González, V.; Roda-Navarro, P.; Fernández-Ruiz, E.; Sancho, D.; Sánchez-Madrid, F. Interleukin-15 and interferon-gamma participate in the cross-talk between natural killer and monocytic cells required for tumour necrosis factor production. *Arthritis Res. Ther.* 2006, *8*, R88, doi:10.1186/ar1955.
- 108. Yamaguchi, M.; Kita, K.; Miwa, H.; Nishii, K.; Oka, K.; Ohno, T.; Shirakawa, S.; Fukumoto, M. Frequent expression of P-glycoprotein/MDR1 by nasal T-cell lymphoma cells. *Cancer* **1995**, *76*, 2351–2356.
- 109. Imashuku, S. Advances in the management of hemophagocytic lymphohistiocytosis. Int. J. Hematol. 2000, 72, 1–11.
- 110. Yamaguchi, M.; Suzuki, R.; Kwong, Y.L.; Kim, W.S.; Hasegawa, Y.; Izutsu, K.; Suzumiya, J.; Okamura, T.; Nakamura, S.; Kawa, K.; et al. Phase I study of dexamethasone, methotrexate, ifosfamide, L-asparaginase, and etoposide (SMILE) chemotherapy for advanced-stage, relapsed or refractory extranodal natural killer (NK)/T-cell lymphoma and leukemia. *Cancer Sci.* 2008, 99, 1016–1020, doi:10.1111/j.1349-7006.2008.00768.x.
- 111. Yamaguchi, M.; Tobinai, K.; Oguchi, M.; Ishizuka, N.; Kobayashi, Y.; Isobe, Y.; Ishizawa, K.; Maseki, N.; Itoh, K.; Usui, N.; et al. Concurrent chemoradiotherapy for localized nasal natural killer/T-cell lymphoma: An updated analysis of the Japan Clinical Oncology Group study JCOG0211. J. Clin. Oncol. 2012, 30, 4044–4046, doi:10.1200/jco.2012.45.6541.
- 112. Yamaguchi, M.; Suzuki, R. JSH practical guidelines for hematological malignancies, 2018: II. Lymphoma-9. Extranodal NK/Tcell lymphoma, nasal type (ENKL). *Int. J. Hematol.* 2019, 109, 371–376, doi:10.1007/s12185-019-02609-x.
- 113. NCCN Guidelines T-Cell Lymphomas Version 1.2021. Available online: www.nccn.org/patients (accessed on 29 May 2021).
- 114. Kim, T.M.; Kim, D.W.; Kang, Y.K.; Chung, J.; Song, H.S.; Kim, H.J.; Kim, B.S.; Lee, J.S.; Kim, H.; Yang, S.H.; et al. A phase II study of ifosfamide, methotrexate, etoposide, and prednisolone for previously untreated stage I/II extranodal natural killer/T-cell lymphoma, nasal type: A multicenter trial of the Korean Cancer Study Group. Oncologist 2014, 19, 1129–1130, doi:10.1634/theoncologist.2014-0305.
- 115. Yamaguchi, M.; Kwong, Y.L.; Kim, W.S.; Maeda, Y.; Hashimoto, C.; Suh, C.; Izutsu, K.; Ishida, F.; Isobe, Y.; Sueoka, E.; et al. Phase II study of SMILE chemotherapy for newly diagnosed stage IV, relapsed, or refractory extranodal natural killer (NK)/Tcell lymphoma, nasal type: The NK-Cell Tumor Study Group study. J. Clin. Oncol. 2011, 29, 4410–4416, doi:10.1200/jco.2011.35.6287.
- 116. Jaccard, A.; Gachard, N.; Marin, B.; Rogez, S.; Audrain, M.; Suarez, F.; Tilly, H.; Morschhauser, F.; Thieblemont, C.; Ysebaert, L.; et al. Efficacy of L-asparaginase with methotrexate and dexamethasone (AspaMetDex regimen) in patients with refractory or relapsing extranodal NK/T-cell lymphoma, a phase 2 study. *Blood* 2011, 117, 1834–1839, doi:10.1182/blood-2010-09-307454.
- 117. Yamaguchi, M.; Suzuki, R.; Oguchi, M. Advances in the treatment of extranodal NK/T-cell lymphoma, nasal type. *Blood* 2018, 131, 2528–2540, doi:10.1182/blood-2017-12-791418.
- 118. Kharfan-Dabaja, M.A.; Kumar, A.; Ayala, E.; Hamadani, M.; Reimer, P.; Gisselbrecht, C.; d'Amore, F.; Jantunen, E.; Ishida, T.; Bazarbachi, A.; et al. Clinical practice recommendations on indication and timing of hematopoietic cell transplantation in mature T CELL and NK/T cell lymphomas: An International collaborative effort on behalf of the guidelines committee of the American Society for Blood and Marrow Transplantation. *Biol. Blood Marrow Transplant.* 2017, 23, 1826–1838, doi:10.1016/j.bbmt.2017.07.027.
- 119. Yamaguchi, M.; Tobinai, K.; Oguchi, M.; Ishizuka, N.; Kobayashi, Y.; Isobe, Y.; Ishizawa, K.; Maseki, N.; Itoh, K.; Usui, N.; et al. Phase I/II study of concurrent chemoradiotherapy for localized nasal natural killer/T-cell lymphoma: Japan Clinical Oncology Group study JCOG0211. J. Clin. Oncol. 2009, 27, 5594–5600, doi:10.1200/jco.2009.23.8295.
- 120. Takahara, M. nasal NK/T-cell lymphoma. Jpn. J. Rhinool. 2019, 58, 85–87, doi:10.7248/jjrhi.58.85.
- 121. Kim, Y.H.; Bagot, M.; Pinter-Brown, L.; Rook, A.H.; Porcu, P.; Horwitz, S.M.; Whittaker, S.; Tokura, Y.; Vermeer, M.; Zinzani, P.L.; et al. Mogamulizumab versus vorinostat in previously treated cutaneous T-cell lymphoma (MAVORIC): An international, open-label, randomised, controlled phase 3 trial. *Lancet Oncol.* 2018, 19, 1192–1204, doi:10.1016/S1470-2045(18)30379-6.
- 122. Nagato, T.; Ohkuri, T.; Ohara, K.; Hirata, Y.; Kishibe, K.; Komabayashi, Y.; Ueda, S.; Takahara, M.; Kumai, T.; Ishibashi, K.; et al. Programmed death-ligand 1 and its soluble form are highly expressed in nasal natural killer/T-cell lymphoma: A potential rationale for immunotherapy. *Cancer Immunol. Immunother.* 2017, *66*, 877–890, doi:10.1007/s00262-017-1987-x.
- 123. Kwong, Y.L.; Chan, T.S.Y.; Tan, D.; Kim, S.J.; Poon, L.M.; Mow, B.; Khong, P.L.; Loong, F.; Au-Yeung, R.; Iqbal, J.; et al. PD1 blockade with pembrolizumab is highly effective in relapsed or refractory NK/T-cell lymphoma failing l-asparaginase. *Blood* 2017, 129, 2437–2442, doi:10.1182/blood-2016-12-756841.
- 124. Schag, K.; Schmidt, S.M.; Müller, M.R.; Weinschenk, T.; Appel, S.; Weck, M.M.; Grünebach, F.; Stevanovic, S.; Rammensee, H.G.; Brossart, P. Identification of C-met oncogene as a broadly expressed tumor-associated antigen recognized by cytotoxic T-lymphocytes. *Clin. Cancer Res.* 2004, *10*, 3658–3666, doi:10.1158/1078-0432.Ccr-03-0640.
- 125. Mortenson, E.D.; Park, S.; Jiang, Z.; Wang, S.; Fu, Y.X. Effective anti-neu-initiated antitumor responses require the complex role of CD4+ T cells. *Clin. Cancer Res.* 2013, *19*, 1476–1486, doi:10.1158/1078-0432.Ccr-12-2522.
- 126. Kumai, T.; Kobayashi, H.; Harabuchi, Y.; Celis, E. Peptide vaccines in cancer-old concept revisited. *Curr. Opin. Immunol.* 2017, 45, 1–7, doi:10.1016/j.coi.2016.11.001.
- 127. Demachi-Okamura, A.; Ito, Y.; Akatsuka, Y.; Tsujimura, K.; Morishima, Y.; Takahashi, T.; Kuzushima, K. Epstein-Barr virus (EBV) latent membrane protein-1-specific cytotoxic T lymphocytes targeting EBV-carrying natural killer cell malignancies. *Eur. J. Immunol.* 2006, *36*, 593–602, doi:10.1002/eji.200535485.

- 128. Kobayashi, H.; Nagato, T.; Takahara, M.; Sato, K.; Kimura, S.; Aoki, N.; Azumi, M.; Tateno, M.; Harabuchi, Y.; Celis, E. Induction of EBV-latent membrane protein 1-specific MHC class II-restricted T-cell responses against natural killer lymphoma cells. *Cancer Res.* 2008, *68*, 901–908, doi:10.1158/0008-5472.can-07-3212.
- 129. Fox, C.P.; Haigh, T.A.; Taylor, G.S.; Long, H.M.; Lee, S.P.; Shannon-Lowe, C.; O'Connor, S.; Bollard, C.M.; Iqbal, J.; Chan, W.C.; et al. A novel latent membrane 2 transcript expressed in Epstein-Barr virus-positive NK- and T-cell lymphoproliferative disease encodes a target for cellular immunotherapy. *Blood* **2010**, *116*, 3695–3704, doi:10.1182/blood-2010-06-292268.