



Supplementary Materials: The PTS Components in *Klebsiella pneumoniae* Affect Bacterial Capsular Polysaccharide Production and Macrophage Phagocytosis Resistance

Novaria Sari Dewi Panjaitan ^{1,†} Yu-Tze Horng ^{1,†}, Chih-Ching Chien ², Hung-Chi Yang ³, Ren-In You ¹ and Po-Chi Soo ^{1,*}

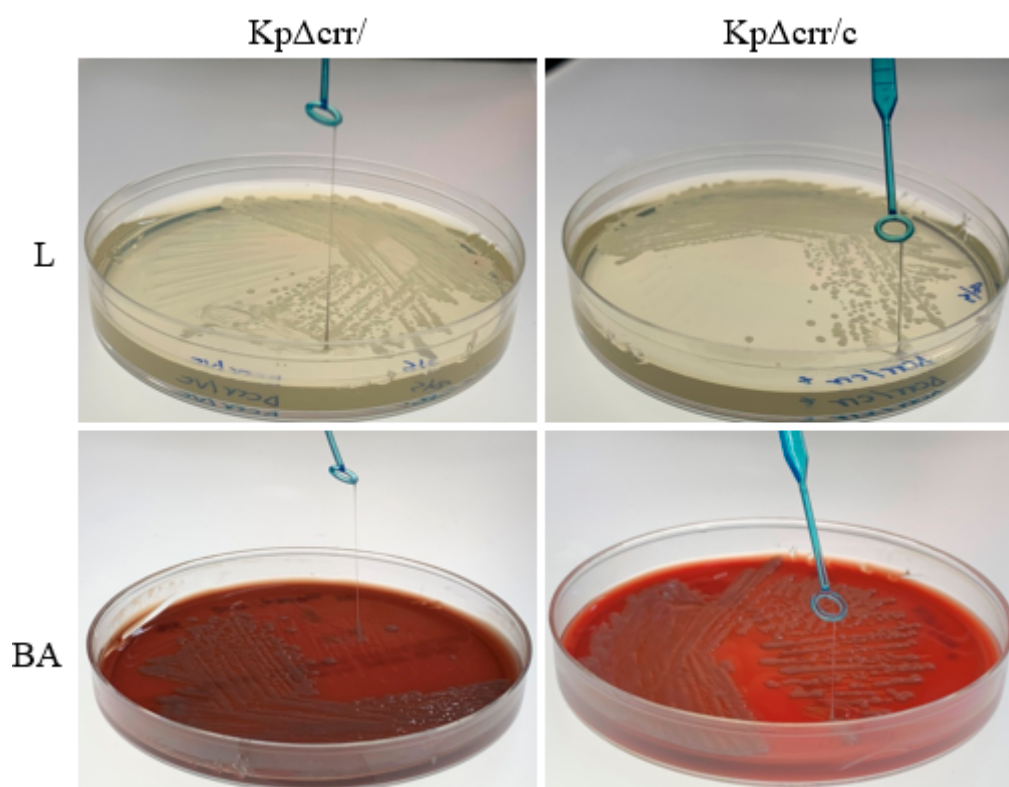


Figure S1. Hypermucoviscosity phenotype of *crr* complemented strain observed by the string test. LB: LB agar plate. BAP: 5% sheep blood agar plate. KpΔcrr/vc: *K. pneumoniae crr* mutant carrying pBAD33 (vector control). KpΔcrr/crr: *K. pneumoniae crr* mutant carrying pBAD33::crr (*crr* complemented strain).

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Table S1. Bacterial strains and plasmids used in this study.

Strain	Relevant genotype and phenotype	Reference or source
<i>E. coli</i>		
DH5α	F [−] , ϕ80dlacZ1M15 (<i>lacZYA-argF</i>) U169, <i>deoR</i> , <i>recA1</i> , <i>endA1</i> , <i>hsdR17</i> (rk [−] , mk ⁺), <i>phoA</i> , <i>supE44</i> , λ [−] , <i>thi-1</i> , <i>gyrA96</i> , <i>relA1</i>	Invitrogen (Thermo Fisher Scientific, USA)
S17-1 λpir	λ-pir lysogen of S17-1 [<i>thi pro hsdR[−] hsdM⁺ recA</i> RP4 2-Tc::Mu-Km::Tn7 (TpR. SmR.)]. Permissive host able to transfer suicide plasmids requiring the Pir protein by conjugation to recipient cells	a
<i>K. pneumoniae</i>		
STU1	Laboratory-maintained strain, Amp ^r	National Taiwan University. b
Δ <i>crr</i>	Deletion of <i>crr</i> gene in STU1	b
Δ <i>etcABC</i>	In frame deletion of <i>etcABC</i> genes in STU1	b
Δ <i>crr</i> Δ <i>etcABC</i>	In frame deletion of <i>etcABC</i> genes in Δ <i>crr</i>	b
Δ <i>crr/crr</i>	<i>crr</i> complement strain: <i>K. pneumoniae crr</i> mutant carrying pBAD33::crr	This work
Plasmid		
pBlueScript SK+ (pBSK)	Cloning vector containing <i>lac</i> promoter, pUC <i>ori</i> , Amp ^r	Stratagene (Stratagene California, USA)
pBSK-Gm	pBSK derivative carrying gentamicin resistance gene at the <i>ScaI</i> site, Gm ^r	c
pBSK-Gm::Km	pBSK-Gm derivative carrying kanamycin resistance gene at the <i>Sall</i> site, Gm ^r , Km ^r	This work
pBSK-Gm::Km::etcABC	pBSK-Gm::Km derivative carrying <i>etcABC</i> genes at the <i>ScaI</i> site, Gm ^r , Km ^r	This work
pBAD33	Expression vector utilizing P _{BAD} promoter, pACYC184 <i>ori</i> , Cm ^r	d
pBAD33::crr	pBAD33 carrying <i>crr</i> at the <i>XbaI</i> and <i>HindIII</i> sites with poly-His tag (6x His) fused to N-terminus of Crr.	This work
pMV261::ZsGreen	pMV261 carrying fluorescent reporter gene, <i>ZsGreen</i> , Amp ^r	kind gift from assistant Prof. Yih-Yuan Chen
pBSK-Km	pBSK derivative carrying kanamycin resistance gene at the <i>Sall</i> site, Km ^r	This work
pBSK-Km::ZsGreen	pBSK-Km carrying <i>ZsGreen</i> gene at the <i>Sall</i> and <i>BamHI</i> sites. <i>ZsGreen</i> gene was cloned from pMV261::ZsGreen, Km ^r .	This work
pW18mobsacB	Conjugative vector, R6K <i>ori</i> , RP4 <i>mob</i> , <i>sacB</i> , Km ^r , suicide plasmid in <i>K. pneumoniae</i> .	b

Note:

a R. Simon, U. Priefer., A. Pühler. Bio/Technology 1 784–791, 1983, <https://doi.org/10.1038/nbt1183-784>.b N. S. D. Panjaitan, Y. T. Horng, S. W. Cheng, W. T. Chung, P. C. Soo, Front Microbiol 10: 1558, 2019, <https://doi.org/10.3389/fmicb.2019.01558>. eCollection 2019c Y. T. Horng, C. J. Wang, W. T. Chung, H. J. Chao, Y. Y. Chen, P. C. Soo, J Microbiol Immunol Infect 51: 174–183, 2018, <https://doi.org/10.1016/j.jmii.2017.01.007>. Epub 2017 Jun 29.d L. M. Guzman, D. Belin, M. J. Carson, J. Beckwith, 177 4121–4130, 1995, <https://doi.org/10.1128/jb.177.14.4121-4130.1995>.**Table S2.** Oligonucleotide primers used in this study.

Primer	Sequences (5' → 3')	Target / purpose
16S rRNA qPCR FP	GCACAGAGAGCTTGC	16S rRNA
16S rRNA qPCR RP	CACTTTGGTCTTGCGA	16S rRNA
16S rRNA Probe	/56-FAM/ATGTCTGGG/ZEN/ AAACTGCCTGATGGA/3IABkFQ/	16S rRNA

recA qPCR FP	CCGCTTTCTCAATCAGCTTC	<i>recA</i> mRNA / RT-qPCR
recA qPCR RP	TTAAACAGGCCGAATTCCAG	<i>recA</i> mRNA / RT-qPCR
recA Probe	/56-FAM/TCGCCGTAG/ZEN/ AAGTTGATGCCTTCG/3IABkFQ/	<i>recA</i> mRNA / RT-qPCR
etcA qPCR FP	CGCCGAATATGTCAACGAGA	<i>etcA</i> mRNA / RT-qPCR
etcA qPCR RP	ATATGACTGTCAGCATCGGC	<i>etcA</i> mRNA / RT-qPCR
etcA Probe	/56-FAM/TTATCACCA/ZEN/ CAGCCACGCCAATCT/3IABkFQ/	<i>etcA</i> mRNA / RT-qPCR
etcB qPCR FP	GTAACGGTATTGGCAGCTCA	<i>etcB</i> mRNA / RT-qPCR
etcB qPCR RP	CAGCAATTTTCATCAGCGCA	<i>etcB</i> mRNA / RT-qPCR
etcB Probe	/56-FAM/ACAGTGGGC/ZEN/ CGGAAATTGATTTCG/3IABkFQ/	<i>etcB</i> mRNA / RT-qPCR
etcC qPCR FP	ACCGACCAAAACATGCTGAT	<i>etcC</i> mRNA / RT-qPCR
etcC qPCR RP	GAGCTGATGAATCCCACCAC	<i>etcC</i> mRNA / RT-qPCR
etcC Probe	/56-FAM/TGTTCTTCA/ZEN/ CCTTTAGTCCCACCGC/3IABkFQ/	<i>etcC</i> mRNA / RT-qPCR
galF qPCR FP	CAAAGGCAATTCCAAAGGAG	<i>galF</i> mRNA / RT-qPCR
galF qPCR RP	CCAGCTCGTAGGAGGTATCG	<i>galF</i> mRNA / RT-qPCR
galF Probe	/56-FAM/CGAGGAGTG/ZEN/CGTCACCAGAACAAT/3IABkFQ/	<i>galF</i> mRNA / RT-qPCR
wzi qPCR FP	TGACGGCCAGAACTACC	<i>wzi</i> mRNA / RT-qPCR
wzi qPCR RP	GACAACGACTTGCGTAACGA	<i>wzi</i> mRNA / RT-qPCR
gnd qPCR FP	TTCACCATCTTCAGCACGAG	<i>gnd</i> mRNA / RT-qPCR
gnd qPCR RP	ATGGTGGCAACACCTTCTTC	<i>gnd</i> mRNA / RT-qPCR
crr FP	CCCGGGGACGAGTTAATGACGCTGGTT	Amplification of <i>crr</i> for Crr complementation
crr RP	AAGCTTGTGCTATCGACCGGTAACCT	Amplification of <i>crr</i> for Crr complementation
crr up FP	GGCGGCCGCCCCGGGAAATTG AAGCGCTGCGTAGT	Construction of <i>crr</i> deletion used suicide plasmid
crr up RP	GGGGATCCGCATCTCGTGGATTAGCAGA	Construction of <i>crr</i> deletion used suicide plasmid
crr down FP	GGGGATCCCGGTAATCCGCATCAAGAAG	Construction of <i>crr</i> deletion used suicide plasmid
crr down RP	GGGAGCTCCCCGGGACGCCTATCGTCAGCACCT	Construction of <i>crr</i> deletion used suicide plasmid
KO crr 1HR FP	TTGCGATGGATCGTAAAGAG	PCR amplification for confirmation <i>crr</i> deletion
KO crr 1HR RP	GGTCACCACCGGCTATATG	PCR amplification for confirmation <i>crr</i> deletion