



Supplementary material

Table S1. Bacterial strains and plasmids used in this study.

Strains or plasmids	Genotype and/or characteristics	Reference or source
ATCC BAA-894	<i>Cronobacter sakazakii</i>	
$\Delta ompF$	WT	ATCC
<i>cpompF</i>	$\Delta ompF::km^r$	This study
	$\Delta ompF$ with pACYC184-413	This study
DH5 α	<i>Escherichia coli</i>	
	$\gamma\text{-}\Phi 80\Delta lacZ\Delta M15\Delta(lacZYA\text{-}argF)U169\text{ }recA1\text{ }endA1\text{ }hsdR17(rk^-\text{ }mk^-)$	Kim et al.
	<i>supE44 thi-1 gyrA relA1</i>	
	Plasmids	
pKD46	<i>oriR101 repA101(Ts) Amp^r araADpgam-bet-exo</i>	Kim et al.
pMDKUD	pMD18-T	This study
pACYC184	p15A <i>ori</i> Cm ^r Tet ^r	Kim et al.
pACYC413	pACYC184- <i>ompF</i>	This study

Table S2. Primers used in this study.

Gene amplified	Primers	Primer sequences (5' to 3')	Amplicon size (bp)	Note
		Mutant construction		
kana-F	Km ^r cassette	CGGATCCGAGGTATGTAGGCGGTGC	26	<i>Bam</i> HI
kana-R		CGCGTCGACATATGTATCCGCTCATGAATT	30	<i>Sal</i> I
ompF5-F	Upstream of <i>ompF</i>	GGGGTACCGCGTTGTGCCTGTAGCC	25	<i>Kpn</i> I
ompF5-R		CGGGATCCTGTCTGTCTGGCATCTTTCC	28	<i>Bam</i> HI
ompF3-F	Downstream of <i>ompF</i>	CGCGTCGACGAAATCACAATGGAACCTCGTC	31	<i>Sal</i> I
ompF3-R		CCCAAGCTTTCCCGTCTGCTGGTTTCG	27	<i>Hind</i> III
		Complementation		
Cp413-F	<i>ompF</i> gene	CGGGATCCTTCTATTACGGTTTCACGG	27	<i>Bam</i> HI
Cp413-R	sequence	CGCGTCGACACGAGGTTCCATTGTGATT	28	<i>Sal</i> I

Table S3. Raman intensities of different wavenumbers.

Wave numbers	Raman intensity (a.u.)		
	WT	$\Delta ompF$	<i>cpompF</i>
852.63	49803.93 ^a	12332.86 ^b	22702.15 ^c
1002.94	46893.56 ^a	10611.55 ^b	21559.08 ^c
1126.76	48777.15 ^a	11321.138 ^b	22094.28 ^c
1287.73	52595.36 ^a	16261.23 ^b	28445.75 ^c
1451.74	51277.15 ^a	13301.40 ^b	24290.73 ^c

^{a, b, c} in the same raw with different letters indicate significant difference at the 0.01 level according to one-way ANOVA.