



Supplementary Figure S1. Gel electropherogram of virulence-related gene amplicons from the 14 *Aeromonas* isolates used in this study. Virulence genes showing positive amplicons are only shown and red numbers indicate positive amplicons, respectively. *Aeromonas* strains were arranged in the following order: 1, *A. hydrophila* KN-Mc-1R1; 2, *A. hydrophila* KN-Mc-1R2; 3, *A. cavieae* KN-Mc-1R3; 4, *A. hydrophila* KN-Mc-2R1; 5, *A. cavieae* KN-Mc-3R1; 6, *A. hydrophila* KN-Mc-4N1; 7, *A. hydrophila* KN-Mc-5R2; 10, *A. hydrophila* KN-Mc-6U21; 11, *A. dhakensis* KN-Mc-6U2; 12, *A. hydrophila* KN-Mc-6U22; 13, *A. hydrophila* KN-Mc-10N1; 14, *A. rivipollensis* KN-Mc-11N1; N, negative control; M, SiZer[™]-100 plus DNA marker (Intron Biotech, Seongnam, Republic of Korea).

Gene	Primer sequence (5' to 3')	Ref	
Bacterial identification (Product size (bp))			
16C DNIA (14/4)	F(27F): AGAGTTTGATCMTGGCTCAG	Universal	
165 TKINA (1466)	R(1492R): TACGGYTACCTTGTTACGACTT	primer	
·····R (-1100)	F(gyrB3F): TCCGGCGGTCTGCACGGCGT	[1]	
gyrb (≈1100)	R(gyrB14R): TTGTCCGGGTTGTACTCGTC	[1]	
$m_0 P(F(0))$	F(Pasrpob-L): GCAGTGAAAGARTTCTTTGGTTC	[2]	
гров (380)	R(Rpob-R): GTTGCATGTTNGNACCCAT	[2]	
Virulence-associated genes (Product size (bp))			
A at (121)	F: CCTATGGCCTGAGCGAGAAG	[2]	
Act (451)	R: CCAGTTCCAGTCCCACCACT	[3]	
aorT(525)	F: GGCGCTTGGGCTCTACAC	[4]	
uex 1 (555)	R: GAGCCCGCGCATCTTCAG	[4]	
<i>Alt</i> (320)	F: AAAGCGTCTGACAGCGAAGT	[5]	
All (520)	R: AGCGCATAGGCGTTCTCTT	[0]	
accV(710)	F: ATGGACGGCGCCATGAAGTT	[6]	
usev (710)	R: TATTCGCCTTCACCCATCCC	[0]	
asn A (350)	F: CACCGAAGTATTGGGTCAGG	[3]	
<i>uspr</i> (000)	R: GGCTCATGCGTAACTCTGGT	[5]	
Ast (504)	F: ATCGTCAGCGACAGCTTCTT	[5]	
13t (30 1)	R: CTCATCCCTTGGCTTGTTGT	[0]	
BfnA (251)	F: CCGCAGGTGTGATGTTTTAC	[7]	
	R: TGCGGTGTTATTGTTTGCT	[7]	
BfnC(233)	F: ATGCCAAAGCTGACTGGTCT	[7]	
<i>Djp</i> G (200)	R: GACATGATTCCCGTTATAAA	[7]	
fla A (608)	F: TCCAACCGTYTGACCTC	[8]	
<i>Juu</i> 1 (000)	R: GMYTGGTTGCGRATGGT	[0]	
lafA (737)	F: CCAACTTYGCYTCYMTGACC	[5]	
	R: TCTTGGTCATRTTGGTGCTY	[0]	
str-1 (180)	F: ATAAATTGCCATTCGTTGACTAC	[9]	
<i>ow</i> 1 (100)	R: AGAACGCCCACTGAGATCATC	[2]	

Supplementary Table S1. List of PCR primers used in this study.

	stx-2 (255)	F: GGCACTGCTTGAAACTGCTCC	[9]					
		R: TCGCCAGTTATCTGACATTCTG	[-]					
	vasH (1.652)	F: GCTCTAGACCGGTGAACCCATCAAGCGCGTCCACT	[10]					
		R: TCCCCCCGGGCTGGTGGCCAGCAGCAGAGGCAATA	[10]					
Anti	microbial-resistance genes (Product size (bp))							
	tet A (211)	F(tetAF): GCTACATCCTGCTTGCCTTC						
		R(tetAR): GCATAGATCGCCGTGAAGAG						
	tetB (391)	F(ClassB tetAF): TCATTGCCGATACCACCTCAG						
T*		R(ClassB tetAR): CCAACCATCATGCTATTCCATCC						
	tet((897)	F(ClassC tetAF): CTGCTCGCTTCGCTACTTG	[11]					
	<i>(0)</i>	R(ClassC tetAR): GCCTACAATCCATGCCAACC	[11]					
	tatD(844)	F(ClassD tetAF): TGTGCTGTGGATGTTGTATCTC						
	leiD (044)	R(ClassD tetAR): CAGTGCCGTGCCAATCAG						
	tatE (714)	F(ClassE tetAF): ATGAACCGCACTGTGATGATG						
	letL (744)	R(ClassE tetAR): ACCGACCATTACGCCATCC						
	$\alpha_{\rm H} = \Lambda (662)$	F(ASGYRA1): CCATGAGCGTGATCGTAGGA						
	89774 (003)	R(ASGYRA2): CTTTGGCACGCACATAGACG	[12]					
	man C (419)	F(ASPARC3): CAGCGGCGCATCATCTAC	[12]					
	<i>parc</i> (418)	R(ASPARC4): GGATATCGGTGGCCATGC						
01	A1 ha AC (E80)	F(qnrAm-F): AGAGGATTTCTCACGCCAGG						
Q	<i>qnrA1</i> to <i>qnrA6</i> (380)	R(qnrAm-R): TGCCAGGCACAGATCTTGAC						
	$\mathbf{P}(1) = \mathbf{P}(1) \mathbf{P}(1)$	F(qnrBm-F): GGMATHGAAATTCGCCACTG	[10]					
	qnrB1 to qnrB6 (264)	R(qnrBm-R): TTTGCYGYYCGCCAGTCGAA	[13]					
	aux C1 to aux C2 (128)	F(qnrSm-F): GCAAGTTCATTGAACAGGGT						
	qnr51 to qnr52 (428)	R(qnrSm-R): TCTAAACCGTCGAGTTCGGCG						
T*		F(5'-CS): GGCATCCAAGCAGCAAG	[1.4]					
1	Class 1 integron (0.7~3.0 k)	R(3'-CS): AAGCAGACTTGACCTGA	[14]					
	CTV M 1 ((89))	F(CTXGp1-F): TTAGGAARTGTGCCGCTGYA						
	C1X-M-1 group (688)	R(CTXGp1-R): CGATATCGTTGGTGGTRCCAT						
	CTV M 2 means (404)	F(CTXGp2-F): CGTTAACGGCACGATGAC						
	C1X-M-2 group (404)	R(CTXGp2-R): CGATATCGTTGGTGGTRCCAT						
	(T_{1})	F(CTXGp9-F): TCAAGCCTGCCGATCTGGT						
	C1X-M-9 group (561)	R(CTXGp9-R): TGATTCTCGCCGCTGAAG	[4 =]					
		F(TEM-F): CATTTCCGTGTCGCCCTTATTC	[15]					
	1 EM (800)	R(TEM-R): CGTTCATCCATAGTTGCCTGAC						
		F(SHV-F): AGCCGCTTGAGCAAATTAAAC						
	SHV (713)	R(SHV-R): ATCCCGCAGATAAATCACCAC						
		F(OXAA-F): GGCACCAGATTCAACTTTCAAG						
Di	OXA-A variants (564)	R(OXAA-R): GACCCCAAGTTTCCTGTAAGTG						
B.	MOX-1, MOX-2, CMY-1, CMY-8 to -11 (520)	F(MOXMF): GCTGCTCAAGGAGCACAGGAT						
		R(MOXMR): CACATTGACATAGGTGTGGTGC						
	LAT-1 to -4, CMY-2 to -7, BIL-1 (462)	F(CITMF): TGGCCAGAACTGACAGGCAAA						
		R(CITMR)· TTTCTCCTGAACGTGGCTGGC						
	DHA-1 to -2 (405)	F(DHAMF): AACTTTCACAGGTGTGCTGGGT						
		R(DHAMR): CCGTACGCATACTGGCTTTGC						
	ACC (346)	F(ACCMF): AACAGCCTCAGCAGCCGGTTA	[16]					
		R(ACCMR): TTCGCCGCAATCATCCCTAGC						
	MIR-1, ACT-1 (302)	F(EBCMF): TCGGTAAAGCCGATGTTGCGG						
	, , , ,	R(EBCMR): CTTCCACTGCGGCTGCCAGTT						
	FOX-1 to -5b (190)	F(FOXMF): AACATGGGGTATCAGGGAGATG						
	× /	R(FOXMR): CAAAGCGCGTAACCGGATTGG						
	1 4 (=====		[4 [2]]					

* T, tetracycline; Q, quinolones; I, integrons; B, β-lactams; C, carbapenems.

	Strains													
	KN-Mc- 1R1	KN-Mc- 1R2	KN-Mc- 1R3	KN-Mc- 2R1	KN-Mc- 3R1	KN-Mc- 4N1	KN-Mc- 4N3	KN-Mc- 5R1	KN-Mc- 5R2	KN-Mc- 6U2	KN-Mc- 6U21	KN-Mc- 6U22	KN-Mc- 10N1	KN-Mc- 11N1
ONPG	+	+	+	+	+	+	+	-	-	+	+	+	+	+
ADH	+	+	+	+	+	+	+	+	+	+	+	+	+	+
LDC	+	+	-	+	-	+	+	+	+	+	+	+	+	-
ODC	-	-	-	-	-	-	-	-	-	-	-	-	-	-
CIT	-	-	-	-	-	-	-	-	-	-	-	-	-	-
H_2S	-	-	-	-	-	-	-	-	-	-	-	-	-	-
URE	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TDA	+	+	+	+	+	+	+	+	+	+	+	+	+	+
IND	+	+	+	+	+	+	+	+	+	+	+	+	+	+
VP	+	+	-	+	-	+	+	+	+	+	+	+	+	-
GEL	+	+	+	+	+	+	+	+	+	+	+	+	+	-
GLU	+	+	+	+	+	+	+	+	+	+	+	+	+	-
MAN	+	+	+	+	+	+	+	+	+	+	+	+	+	+
INO	-	-	-	-	-	-	-	-	-	-	-	-	-	-
SOR	-	-	-	-	-	-	-	-	-	-	-	-	-	-
RHA	-	-	-	-	-	-	-	-	-	+	-	-	-	-
SAC	+	+	+	+	+	+	+	+	+	+	+	-	+	+
MEL	-	-	-	-	-	-	-	-	-	-	-	-	-	-
AMY	-	-	+	-	+	+	+	-	-	-	-	-	-	-
ARA	+	+	+	+	+	+	+	+	+	+	-	+	+	+

Supplementary Table S2. Biochemical characterization of *Aeromonas* spp. used in this study.

ONPG; β-galactosidase, ADH; arginine dihydrolase, LDC; lysine decarboxylase, ODC; ornithine decarboxylase, CIT; citrate utilization, H₂S; H₂S production, URE; urease, TDA; tryptophane deaminase, IND; indole production, VP; Voges–Proskauer, GEL; gelatinase, GLU; glucose, MAN; mannitol, INO; inositol, SOR; sorbitol, RHA; rhamnose, SAC; saccharose, MEL; melibiose, AMY; amygdalin, ARA; arabinose.

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