Supplementary Materials

Nutrient Excess Triggers the Expression of the Penicillium chrysogenum Antifungal Protein PAFB

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Supplementary methods

1. Purification of Native PAFB from the P. chrysogenum wt

2×10⁸ conidia of *P. chryosogenum wt* and the mutant *pafB*^{paf_promoter} were inoculated in 5×200 mL fourfold concentrated PcMM medium or 5×200 mL 1× PcMM medium, respectively. Cultures were grown at 25 °C for 96 h at 210 rpm. Mycelium was removed and the cell-free supernatant was ultra-filtered (Ultracell 30 kDa, Millipore, Billerica, MA, USA). The ultrafiltrate of the *wt* strain was diluted in a ratio of 1:4 with water before applied to a CM-Sepharose (Fast Flow, GE Healthcare Life Sciences, Little Chalfont, UK) column. The ultrafiltrate of the mutant *pafB*^{paf_promoter} was directly applied to the column, which was equilibrated in phosphate buffer (10 mM NaPO₄, 25 mM NaCl, 0.15 mM EDTA, pH 6.6).

Protein was eluted applying 0.1-0.6 M NaCl. Fractions were analyzed for the presence of PAFB and PAF by Western blot experiments. Those fractions containing only PAFB were pooled and dialyzed (3.5 K MWCO, Thermo Fisher Scientific, Waltham, MA, USA) against ultra-pure ddH₂O. Protein concentrations were determined spectrophotometrically (A₂₈₀) and the purity was checked by SDS-PAGE by Silver staining.

2. P. chrysogenum surface cultures

A conidial suspension $(2 \times 10^5/\text{mL})$ was point inoculated in 5 µL aliquots on 1× PcMM or 4× PcMM agar and grown for 72-96 h at 25 °C. The colonies were removed from the agar plates using sterile tweezers and used for total RNA extraction as indicated in the main text (**Material and Methods**).

Supplementary tables

Culture medium	Composition	
P. chrysogenum minimal	0.3% NaNO ₃ , $0.05%$ MgSO ₄ × 7H ₂ O, $0.05%$ KCl, $0.005%$ FeSO ₄ × 7H ₂ O, $2%$ D(+)-	
medium (PcMM)	sucrose (w/v), 25mM KPO ₄ -buffer (pH = 5.8), 0.1% trace elements A (v/v)	
Trace elements A	0.1% FeSO ₄ × 7H ₂ O, 0.9% ZnSO ₄ × 7H ₂ O, 0.04% CuSO ₄ × 5H ₂ O, 0.01% MnSO ₄ × H ₂ O,	
	0.01% H ₃ BO ₃ , 0.01% Na ₂ MoO ₄ × 2H ₂ O (<i>w/v</i>)	
Complete medium (CM)	2.0% salt solution A, 0.1% trace elements B (v/v), 2.0% D(+)-glucose (w/v), 0.2%	
	bacteriological peptone, 0.1% yeast extract,	
	0.1% NZ-Amine (w/v), pH = 6.5	
Salt solution A	2.6 % KCl, 2.6% MgSO ₄ × 7H ₂ O, 7.6% KH ₂ PO ₄ (<i>w/v</i>), 0.2% chloroform (<i>v/v</i>)	
Trace elements B	1.3% ZnSO ₄ × 7H ₂ O, 0.07% CuSO ₄ × 5H ₂ O, 0.1% MnSO ₄ × H ₂ O,	
	0.006% Na ₂ B ₄ O ₇ × 10H ₂ O, 0.13% Na ₂ MoO ₄ × 2H ₂ O, 0.23% FeSO ₄ × 7H ₂ O (<i>w/v</i>)	
Lysogeny broth medium (LB) 1.0% NaCl, 1.0% bacteriological peptone, 0.5% yeast extract	1.0% NaCl, 1.0% bacteriological peptone, 0.5% yeast extract (w/v)	

Table S1. Media Used in This Study.

Table S2. Microbial Strains Used in This Study.

Strain	Source
Penicillium chrysogenum Q176 (wild type strain)	ATCC 10002

P. chrysogenum paf	Sonderegger et al. 2016 [1]
P. chrysogenum pafB	Huber et al. 2018 [2]
Penicillium chrysogenum pafB ^{paf_promoter}	This study
Penicillium chrysogenum pafB ^{paf_terminator}	This study
Penicillium chrysogenum pafB ^{xylP_promoter}	This study
Bacillus subtilis	ATCC 6633

Table S3. Oligonucleotides Used in This Study.

Oligo name	Sequence 5´3´
pafB3´NotI_rev	CAGGATGCGGCCGCTCGGTATCTTCGATAATTC
<i>pafB_Bgl</i> II_fw	AGATCTATGCATATTACTAGCATTGCCATTGTC
Onat1	CGCCGGTACGCGTGGATCGC
Onat2	AGGCACTGGATGGGTCCTTCAC
pafB5_ <i>BamH</i> I_rev	GGTGGAGGATCCGTCTTGTAGAGGATTGCGG
pafB5_PstI_fw	CAAGGACTGCAGCCGTTGACTAGACCTACACGC
5 pafb fl nested_fw	GCCAAGTTGCTCTTCTGATCTTCCG
3 pafb flanken nested_rev	GTGTCGGAAGTCGGGGAGC
Xylp_XbaI_rev_B	ATCTTCTAGACGACGGAAGCGCGCAGTCGG
Xylp_ <i>Bgl</i> II_fw_B	ACATAGATCTGGTTGGTTCTTCGAGTCGATG
pafB_SmaI_rev	CCCGGGTCAAACTGGGGTCTGGCAG
3´paf_ <i>BamH</i> I_rev	GCCGCTGGATCCCTAGTGCAGCAGTTTGATAG
pafB3_XbaI_fw	ACCAATTCTAGAGTTCCGCAAGAAACAGAGTCC
opafB_without prepro_fw	CTTAGTAAATTCGGAGGA
opafB_rev2	TCAAACTGGGGTCTGGCAG
opaf_without prepro_fw	CTGGCCAAATACACCGGA
opaf_rev	GATCGGATCCCTAGTCACAATCGACAGC
opafb_fw	ATGCATATTACTAGCATTGC
40S rib_protein_S6_rev	GAAGCTCAACATCTCCTACCCG
40S rib_protein_S6_fw	TACGCAGCATCACGGGCAGTC
40S rib_protein_S5_rev	CAAGGAGGTTCTGGCCGAGCA
40S rib_protein_S5_fw	TTGGCGGCGTTGATCAGCTC

Supplementary figures



Figure S1. Scheme of construction and verification of the mutants (a) pafB^{*paf_promoter*} (b) pafB^{*xyIP_promoter*} (c) pafB^{*paf_terminator*} in comparison to the *P. chrysogenum wt*. Left: The beige and grey boxes represent the *pafB* 5' or 3'UTR, respectively. The light green arrow and the blue arrow reflect the nourseothricin-acetyltransferase gene (*nat1*) and the *pafB* gene. The yellow, orange and dark green boxes represent the *paf-promoter*, *xyIP-promoter* and *paf-terminator*, respectively. The black lines A, B, C, D and E represent the transformation fragments. The crosses show regions involved in homologous recombination. The scissors indicate the position of digestion by respective restriction enzymes used for Southern blot analysis. The dashed blue and green lines represent the expected fragments detected in Southern blot analysis by the use of a *pafB-specific* or a *nat1-specific* DIG-probe, respectively. Right: Southern blot analysis of the respective mutants compared to the *P. chrysogenum wt*. Results for *pafB-specific* and *nat1-specific* probe are indicated in blue or green, respectively.



Figure S2. Verification of penicillin production in the culture broth of *P. chrysogenum*. Penicillin (2.5 μ g and 1.2 μ g) and 250 μ L of 120 h old *P. chrysogenum* conditioned cell-free culture broth were used in an inhibition zone assay using *B. subtilis*. Samples treated for 1 h with 1000 U penicillinase. As controls, untreated samples and 1000 U penicillinase alone were included in the assay.



Figure S3. Phenotypical characterization of the *P. chrysogenum pafBpaf_promoter* strain in comparison to the *P. chrysogenum wt*. (a) Mycelial dry weight [DW g/L] of fungal biomass grown in liquid 1× PcMM or 4× PcMM analyzed after 48, 72 and 96 h of cultivation. (b) *pafB* mRNA-expression in surface cultures of *P. chrysogenum wt*. (c) Phenotype of *P. chrysogenum wt* or *pafBpaf_promoter* on 1× PcMM or 4× PcMM agar after 72 h or 96 h incubation at 25°C. (d) Number of conidia per mL. Strains were grown on 1× PcMM or 4× PcMM or 4× PcMM agar for 96 h and conidia were harvested and counted. Values are given as mean ± SD (n=3). P-values in (d) were determined to compare the number of conidia generated under 1× PcMM and 4× PcMM conditions, for the *wt* and the *pafBpaf_promoter* strain, respectively. ** $p \le 0.005$, *** $p \le 0.0005$.

References

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