Supplementary Table S1 List of primers for detection of resistance genes

Gene	Primer sequence (5'-3')	Product size (bp)
vanA	F-GCGCGGTCCACTTGTAGATA	314
	R-TGAGCAACCCCCAAACAGTA	
vanB	F-AGACATTCCGGTCGAGGAAC	220
	R-GCTGTCAATTAGTGCGGGAA	
vanC1	F-ATCCAAGCTATTGACCCGCT	402
	R-TGTGGCAGGATCGTTTTCAT	
vanC2/3	F-CTAGCGCAATCGAAGCACTC	582
	R-GTAGGAGCACTGCGGAACAA	
gyrA	F-AAATCTGCCCGTGTCGTTGGT	343
	R-GCCATACCTACGGCGATACC	
parC	F-AAAAATCAGCGCGTACAGTG	327
	R-CGAGAGTTTGGCTTCGGTAT	
tetA	F-GCTACATCCTGCTTGCCTTC	210
	R-CATAGATCGCCGTGAAGAGG	
tetB	F-TTGGTTAGGGGCAAGTTTTG	659
	R-GTAATGGGCCAATAACACCG	
tetC	F-CTTGAGAGCCTTCAACCCAG	418
	R-ATGGTCGTCATCTACCTGCC	
tetD	F-AAACCATTACGGCATTCTGC	787
	R-GACCGGATACACCATCCATC	
tetE	F-AAACCACATCCTCCATACGC	278
	R-AAATAGGCCACAACCGTCAG	
tetG	F-GCTCGGTGGTATCTCTGCTC	468
	R-AGCAACAGAATCGGGAACAC	
tetK	F-TCGATAGGAACAGCAGTA	169
	R-CAGCAGATCCTACTCCTT	
tetL	F-TCGTTAGCGTGCTGTCATTC	267
	R-GTATCCCACCAATGTAGCCG	
tetM	F-GTATCCCACCAATGTAGCCG	406
	R-CGGTAAAGTTCGTCACACAC	
tetO	F-AACTTAGGCATTCTGGCTCAC	515
	R-TCCCACTGTTCCATATCGTCA	
tetS	F-CATAGACAAGCCGTTGACC	667
	R-ATGTTTTTGGAACGCCAGAG	
tetA(P)	F-CTTGGATTGCGGAAGAAGAG	676
	R-ATATGCCCATTTAACCACGC	
tetQ	F-TTATACTTCCTCCGGCATCG	904

	R-ATCGGTTCGAGAATGTCCAC	
tetX	F-CAATAATTGGTGGTGGACCC	468
	R-TTCTTACCTTGGACATCCCG	
emeA	F-GTGACAGCCTTTGTGGCAGAT	687
	R-TAGTCCGTTGATGGTTCCTTG	

In this study, the cocktail mixture consisted of 12.5 µL of PCR master mix (DreamTag MM Thermo Fisher), 20 µM for the primers, water, and 5 µL of template DNA, resulting in a total reaction volume of 25 µL. The amplification conditions involved an initial enzyme activation/denaturation at 94ºC for 5 minutes, followed by 35 cycles, with each cycle comprising 45 seconds at 94ºC, 60 s at 54ºC, and 90 s at 72ºC. A final elongation step was conducted for 15 minutes at 72ºC. The resulting amplicons were separated through electrophoresis in a 2% agarose gel at 110 V for 45 min, and the interpretation of results was based on the band sizes using a 100 bp ladder. with primer concentrations of 1.25 µM, 1.0 µM, 0.5 µM, 1.25 µM, and 0.5 µM, respectively. Group IV included primers for *tetA*(P) (1.25 µM), *tet*(Q) (1.25 µM) and *tet*(X) (1.25 µM) each). This was done in a reaction volume of 25 µL consisting of 12.5 µL master mix (OneTag Quick Load Master mix with standard buffer, New England BioLabs), and 5 µL of DNA template. Cycling conditions was as follows;

An initial denaturation at 94°C for 5 minutes was succeeded by 35 cycles, each consisting of 1 minute at 94°C, 1 minute at 55°C, and 1.5 minutes at 72°C.

Supplementary Table S2: Primers for the detection of virulence genes

Gene	**Primer sequence (5'-3')**	**Product size (bp)**
ace	F- AAAGTAGAATTAGATCCACAC	320
	R- TCTATCACATTCGGTTGCG	
asa1	F-GCACGCTATTACGAACTATGA	375
	R-TAAGAAAGAACATCACCACGA	
clyA	F- ACTCGGGGATTGATAGGC	688
	R- GCTGCTAAAGCTGCGCTT	

efaA	F- CGTGAGAAAGAAATGGAGGA	499
	R- CTACTAACACGTCACGAATG	
esp	F-AGATTTCATCTTTGATTCTTGG	510
	R-AATTGATTCTTTAGCATCTGG	
hyl	F- ACAGAAGAGCTGCAGGAAATG	276
	R- GACTGACGTCCAAGTTTCCAA	
gelE	F-TATGACAATGCTTTTTGGGAT	213
	R-AGATGCACCCGAAATAATATA	

The PCR reaction volume, set at 20 µL, comprised 10 µL of OneTag rapid Load Master Mix (New England Bio Labs), 1 µL each of primers, and 5 µL of DNA template. For *gyrA*, the protocol included an initial template denaturation at 95°C for 1 minute, followed by 36 cycles of denaturation at 95°C for 30 seconds, annealing at 53°C for 30 seconds, and extension at 72°C for 2 minutes. A final elongation step was performed at 72°C for 10 minutes. Regarding *parC*, the conditions consisted of an initial template denaturation at 95°C for 2 minutes, with the next being 36 cycles of denaturation at 95°C for 60 s, annealing at 60°C for 60 s, and extension at 72°C for 120 s, with a final extension at 72°C for 10 minutes.

In a 20 µL reaction mixture, 10 µL of Quick Load Master Mix, 0.5 µL each of forward and reverse primers, 4 µL of DNA-free water, and 5 µL of DNA template were combined. The cycling conditions included an enzyme activation and initial denaturation at 94°C for 5 minutes, with the next being 30 cycles of denaturation at 94°C for 45 seconds, annealing at 57°C for 60 seconds, and extension at 72°C for 90 seconds, with a final elongation at 72°C for 10 minutes [14]. Electrophoresis of 5 µL of the PCR product on a 2% agarose gel with 1x TAE buffer was performed, dyed with Gel Red, and visualized using a UV transilluminator (BIORAD). The 100-bp ladder was utilized