



Supplementary Materials

Crude Oil Degradation in Temperatures Below the Freezing Point by Bacteria from Hydrocarbon-Contaminated Arctic Soils and the Genome Analysis of *Sphingomonas* sp. AR_OL41

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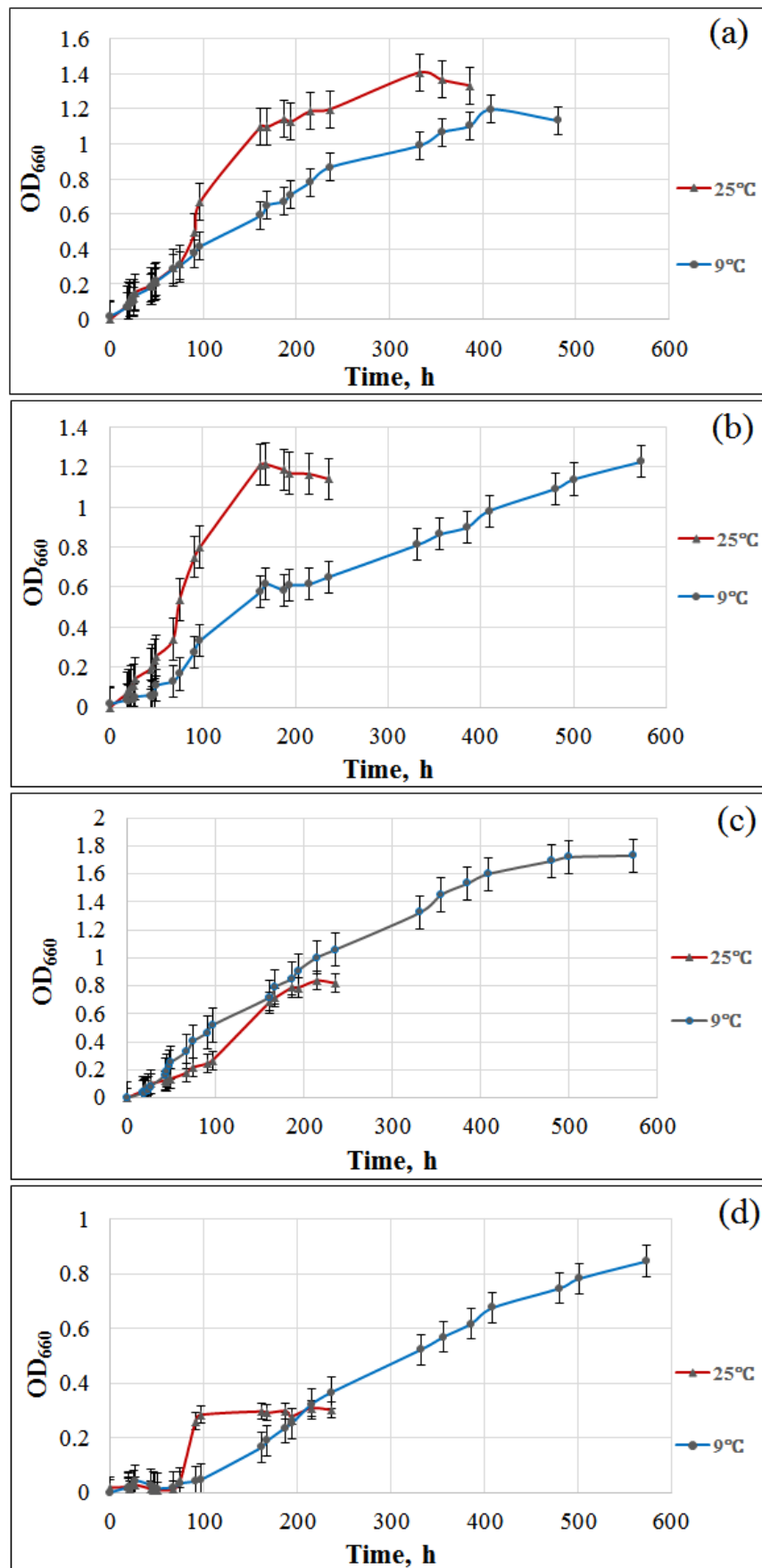


Figure S1. Growth curves of the strains *Pseudomonas frederiksbergensis* Ar-K7 (a), *Rhodococcus yunnanensis* Ar-K9 (b), *Arthrobacter alpinus* Ar-K10 (c), and *Sphingomonas* sp. AR_OL41 (d) in the TEG liquid medium at 9 °C and 25 °C.

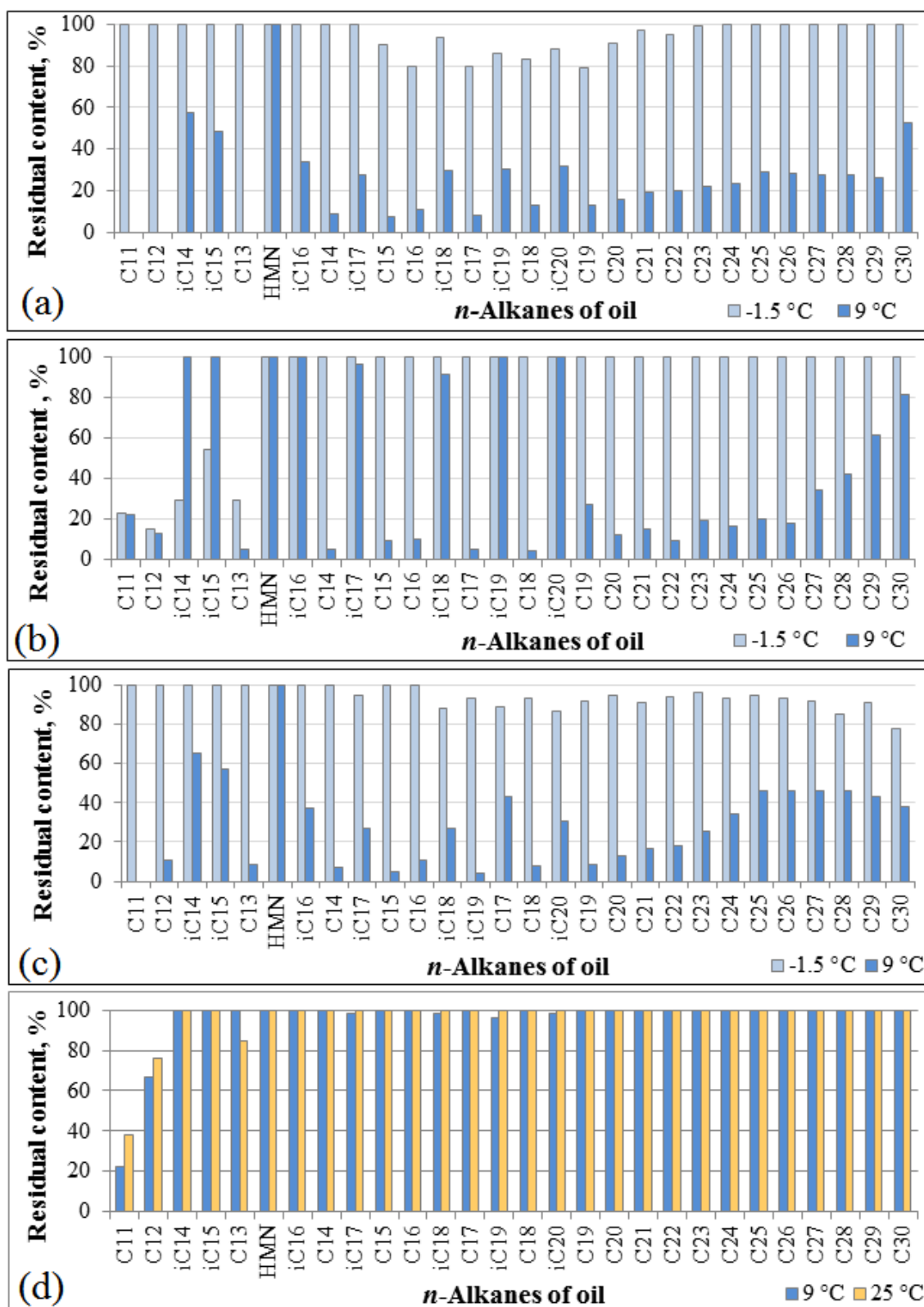


Figure S2. Residual content of *n*-alkanes in crude oil biodegraded by *Pseudomonas frederiksbergensis* Ar-K7 (a), *Rhodococcus yunnanensis* Ar-K9 (b), *Arthrobacter alpinus* Ar-K10 (c), and *Sphingomonas* sp. AR_OL41 (d) at -1.5 °C, 9 °C, and 25 °C.

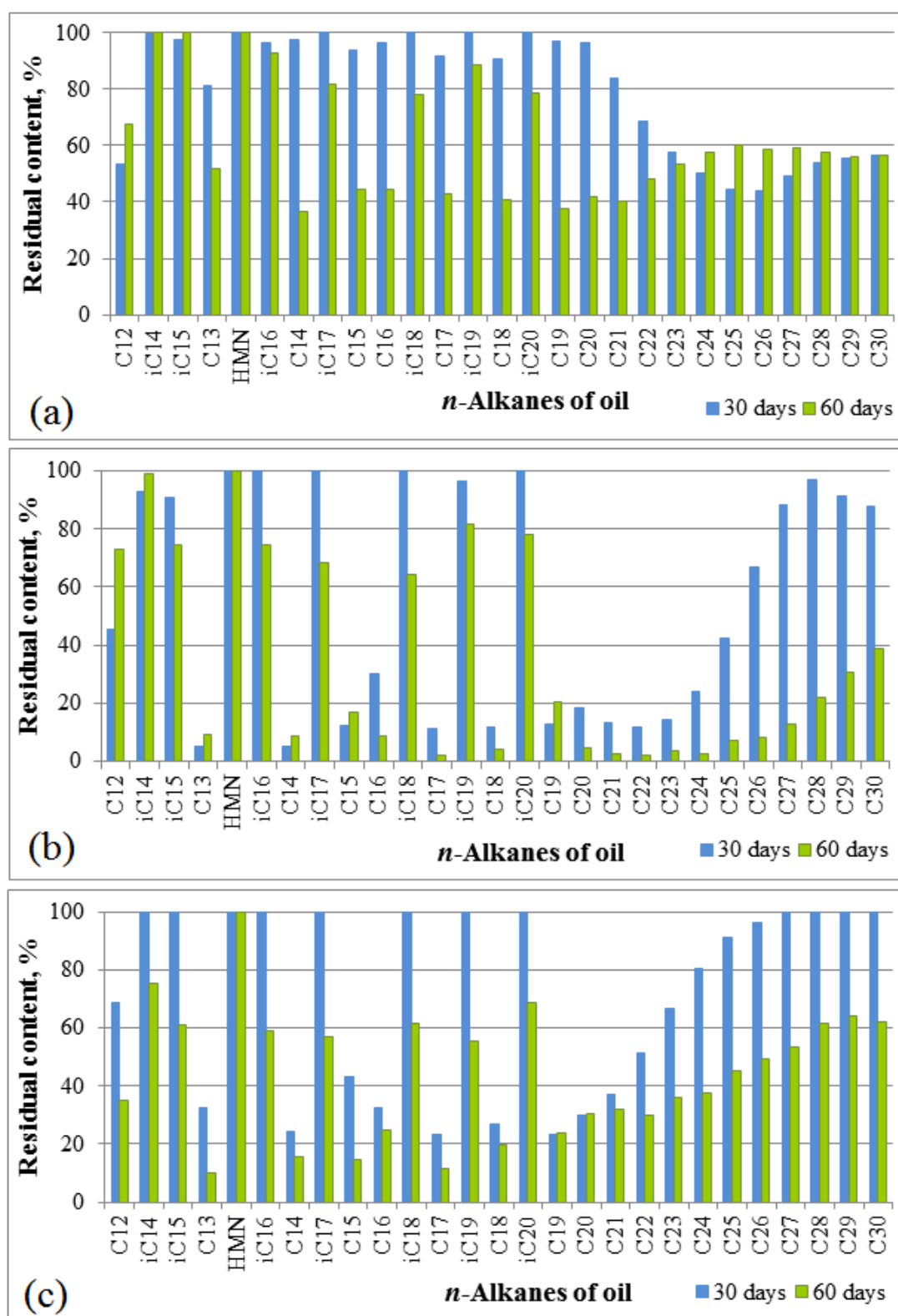


Figure S3. Residual content of *n*-alkanes (%) in crude oil biodegraded by *Pseudomonas frederiksbergensis* Ar-K7 (a), *Rhodococcus yunnanensis* Ar-K9 (b), and *Arthrobacter alpinus* Ar-K10 (c) during remediation of polluted sand.

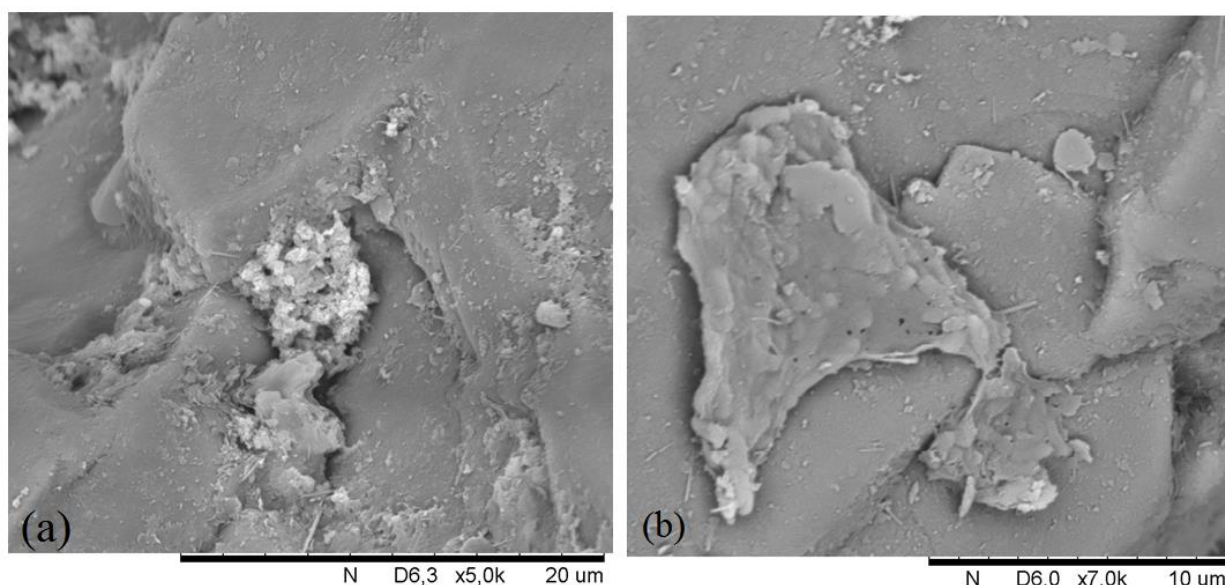


Figure S4. Scanning electron micrographs of biofilms of the strains *Pseudomonas frederiksbergensis* Ar-K7 (a) and *Rhodococcus yunnanensis* Ar-K9 (b) after 30 days of cultivation on oil-contaminated sand. The samples were examined under a scanning electron microscope TM3000 (Hitachi, Tokyo, Japan) with accelerating voltage 15 kV.

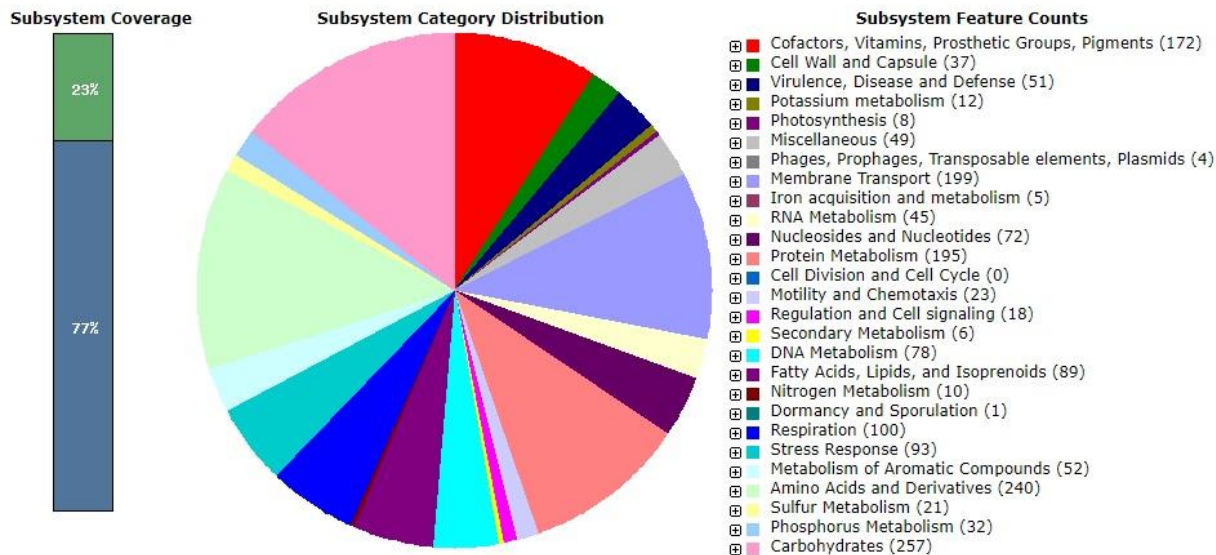


Figure S5. Subsystems of *Sphingomonas* sp. AR_OL41 based on RAST database.

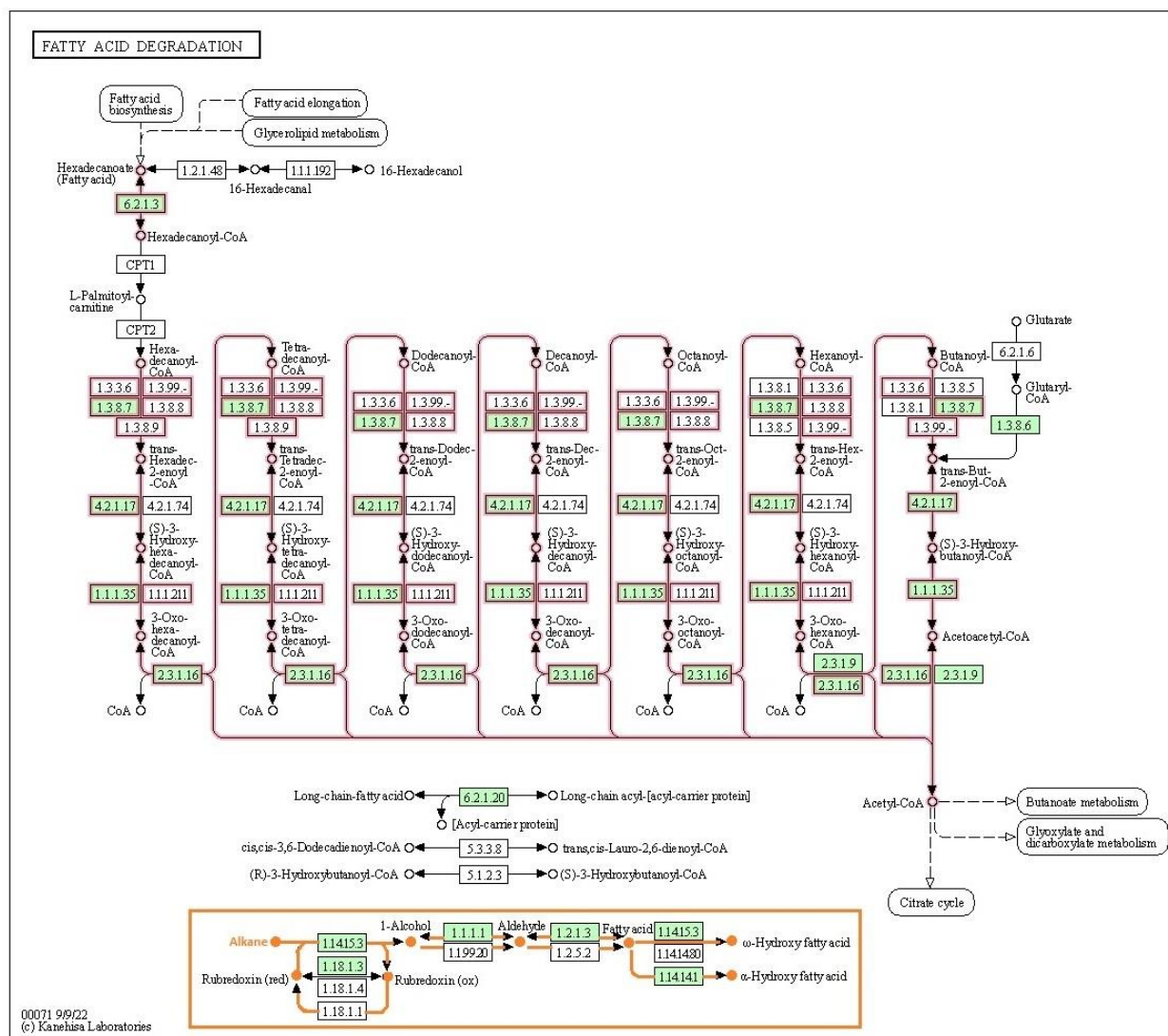


Figure S7. KEGG-map of degradation of fatty acids and *n*-alkanes (id 00071) based on genome analysis of the strain *Spingomonas* sp. AR_OL41.

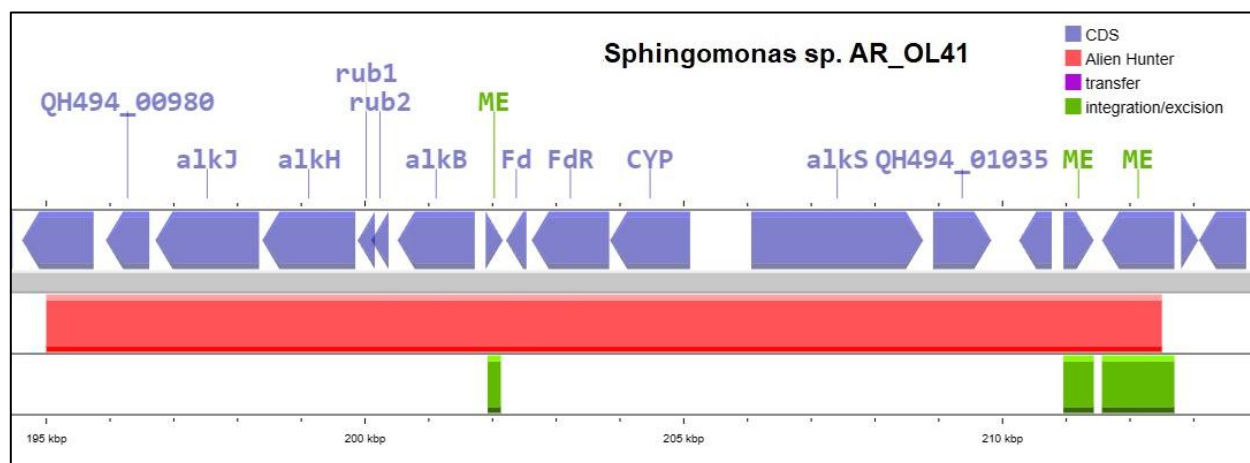


Figure S8. Localization of *n*-alkane degradation genes on the graphic map of the circular chromosome of the strain *Sphingomonas* sp. AR_OL41. The genes of mobile elements (ME) are highlighted in green. Designations: *alkB* – alkane-1 monooxygenase; *rub* – rubredoxin; *alkH* – aldehyde dehydrogenase; *alkJ* – alcohol dehydrogenase; Fd – ferredoxin; FdR – ferredoxin reductase; CYP – presumably cytochrome P450; *alkS* – transcriptional regulator; ME – proteins of mobile elements.

Table S1. Phenotypic characteristics of hydrocarbon-oxidizing bacteria *Pseudomonas frederiksbergensis* Ar-K7, *Rhodococcus yunnanensis* Ar-K9, *Arthrobacter alpinus* Ar-K10, and *Sphingomonas* sp. AR_OL41.

Characteristic	<i>Pseudomonas frederiksbergensis</i> Ar-K7	<i>Rhodococcus yunnanensis</i> Ar-K9	<i>Arthrobacter alpinus</i> Ar-K10	<i>Sphingomonas</i> sp. AR_OL41
Temperature range/ optimum, °C	−1.5...25/5	5...25/ 5–15	−1.5...25/ 10–25	9...25/20–25
NaCl range/optimum, % <i>w/v</i>	0–1.5/0.2	0–4/0.5	0–6/0–4	0–1.5/0
API® ZYM tests:				
Alkaline phosphatase	+	+	+	+
Esterase (C4)	+	W	+	+
Esterase lipase (C8)	+	+	+	+
Lipase (C14)	+	+	W	–
Leucine arylamidase	+	+	+	+
Valine arylamidase	+	+	+	+
Cystine arylamidase	+	+	+	+
Trypsin	–	+	+	–
α-Chymotrypsin	–	+	+	–
Acid phosphatase	+	+	+	+
Naphthol-AS-BI- phosphohydrolase	+	+	+	+
α-Galactosidase	–	–	+	–
β-Galactosidase	–	–	+	–
β-Glucuronidase	–	–	+	+
α-Glucosidase	–	–	+	+
β-Glucosidase	–	+	+	+
α-Mannosidase	–	–	+	–
API® 20E tests:				
β-Galactosidase	–	–	+	–
Arginine dihydrolase	+	–	–	–
Citrate utilization	+	–	+	–
Acid from glucose	+	+	–	–
Xylan	+	–	–	+
Chitin	+	+	+	+
Catalase	+	+	+	+
Oxidase	+	–	–	+

Results of the API® 20E tests for all studied strains were negative for lysine decarboxylase, ornithine decarboxylase, H₂S production, urease, tryptophane deaminase, indole production, acetoin production (Voges Proskauer), and gelatinase. In the API® ZYM tests, all four studied strains were negative for N-acetyl-β-glucosaminidase and α-fucosidase.

Table S2. Physiological growth parameters of *Pseudomonas frederiksbergensis* Ar-K7, *Rhodococcus yunnanensis* Ar-K9, and *Arthrobacter alpinus* Ar-K10 on oil-contaminated sand.

Sample	0 days		30 days				60 days				Oil decrease, mg/100 g of sand
	Cells number per g of sand	Biofilm density, OD ₅₄₀	Cells number per g of sand	Biofilm density, OD ₅₄₀	pH	Acetate/ ethanol, mg/kg of sand	Cells number per g of sand	Biofilm density, OD ₅₄₀	pH	Acetate/ ethanol, mg/kg of sand	
Ar-K7 (Control without oil)	10 ⁷	-	10 ⁷	0.32	7.7	81.6 / 11.0	10 ⁸	0.24	8.1	20.0 / 0.8	-
Ar-K7	10 ⁸	0.12	10 ⁷	0.35	7.7	22.1 / 5.1	10 ⁷	0.70	8.0	12.5 / 1.9	87±13
Ar-K9 (Control without oil)	10 ³	-	0	0.15	7.8	19.4 / 5.7	0	0.14	7.8	23.0 / 6.4	-
Ar-K9	10 ³	0.14	10 ⁶	0.64	7.7	15.6 / 6.5	10 ⁷	0.56	7.8	6.8 / 2.0	37±6
Ar-K10 (Control without oil)	10 ⁷	-	10 ⁶	0.20	7.8	18.1 / 5.3	10 ⁶	0.14	7.8	8.1 / 0.7	-
Ar-K10	10 ⁷	0.13	10 ⁷	0.30	7.8	34.9 / 6.0	10 ⁷	0.48	7.9	11.0 / 1.0	70±11

Table S3. Genes implicated in the adaptation to cold environments in the genome of *Sphingomonas* sp. AR_OL41.

Functional classification	Protein name	Gene Symbol	No. of genes	RefSeq Locus Tag
DNA replication	DNA gyrase subunit A (EC: 5.99.1.3)	<i>gyrA</i>	1	QH494_18230
	RecA protein	<i>gecA</i>	1	QH494_01255
	Chromosomal replication initiator protein DnaA	<i>dnaA</i>	1	QH494_20750
Nucleoid protein; DNA supercoiling	DNA-binding protein HU-beta		2	QH494_00455
				QH494_08860
RNA chaperones	Cold shock protein, CspA family		5	QH494_04405
				QH494_09435
				QH494_14005
				QH494_17445
				QH494_25730
Protein folding	Chaperone protein DnaK	<i>dnaK</i>	2	QH494_12850
				QH494_14160
	Chaperone protein DnaJ	<i>dnaJ</i>	1	QH494_12855
	Peptidyl-prolyl cis-trans isomerase (EC: 5.2.1.8)		3	QH494_11325
Protein biosynthesis	Translation initiation factor 1	<i>infA</i>	1	QH494_24975
	Ribosome-binding factor A	<i>rbfA</i>	1	QH494_27105
Unsaturation of membrane lipids	Fatty acid desaturase, type 2 Delta-9 fatty acid desaturase (EC: 1.14.19.1)		2	QH494_04100
			3	QH494_17510
				QH494_24800
				QH494_25035
Clustering-based subsystems	Exopolysaccharide biosynthesis protein		1	QH494_20245
Pyruvate metabolism II	Pyruvate dehydrogenase E1 component alpha subunit (EC: 1.2.4.1)	<i>pdhA</i>	2	QH494_02965
				QH494_02965
	Pyruvate dehydrogenase E1 component beta subunit (EC: 1.2.4.1)	<i>pdhB</i>	2	QH494_02960
				QH494_02960
	Pyruvate dehydrogenase E2 component (dihydrolipoyllysine-residue acetyltransferase) (EC: 2.3.1.12)	<i>pdhC</i>	1	QH494_06455