



Protection against Severe Illness versus Immunity—Redefining Vaccine Effectiveness in the Aftermath of COVID-19

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Abstract: Anti-SARS-CoV-2 vaccines have played a pivotal role in reducing the risk of developing severe illness from COVID-19, thus helping end the COVID-19 global public health emergency after more than three years. Intriguingly, as SARS-CoV-2 variants emerged, individuals who were fully vaccinated did get infected in high numbers, and viral loads in vaccinated individuals were as high as those in the unvaccinated. However, even with high viral loads, vaccinated individuals were significantly less likely to develop severe illness; this begs the question as to whether the main effect of anti-SARS-CoV-2 vaccines is to confer protection against severe illness or immunity against infection. The answer to this question is consequential, not only to the understanding of how anti-SARS-CoV-2 vaccines work, but also to public health efforts against existing and novel pathogens. In this review, we argue that immune system sensitization-desensitization rather than sterilizing immunity may explain vaccine-mediated protection against severe COVID-19 illness even when the SARS-CoV-2 viral load is high. Through the lessons learned from COVID-19, we make the case that in the disease's aftermath, public health agencies must revisit healthcare policies, including redefining the term "vaccine effectiveness."

Keywords: SARS-CoV-2; immunity; cytokines; infectious diseases; public health

1. Introduction

On 5 May 2023, the World Health Organization (WHO) declared that coronavirus disease 2019 (COVID-19), which is linked to infection with the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) [1] is no longer a public health emergency of international concern (PHEIC) [2–4]. The WHO cautioned that the end of COVID-19's PHEIC status was not a license to stop being vigilant against the disease, but rather that COVID-19 should now be managed by local public health agencies along with other common infectious diseases [2–4]. The WHO justified its decision on the basis of data, which indicate that the proportion of infections leading to severe illness and deaths from COVID-19 have satisfactorily declined since the PHEIC was first declared in January of 2020 [2–4]. This is indeed a cause for celebration, given how overwhelmed healthcare delivery systems were during the early waves of COVID-19 [5–13]. The accelerated development, testing, and deployment of anti-SARS-CoV-2 vaccines (a.k.a., COVID-19 vaccines), which were deemed safe and effective by various local regulatory bodies, and ratified by the WHO and other global public health agencies, have played a pivotal role in



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). reducing the likelihood of severe illness and death from COVID-19 [14,15]. Based on data from 185 countries, computational models suggest that during just the first year after the deployment of COVID-19 vaccines, there was about a 63% reduction in total deaths [16]. While celebrating the end of the COVID-19 PHEIC is warranted, it is important that the WHO and other health agencies critically assess the public health management of COVID-19 in order to more rapidly and effectively contain infectious disease outbreaks that might occur in the future [17].

In this review, we present two main lessons learned from the COVID-19 pandemic. First, we present information on the pathophysiology of COVID-19 and discuss how pathogens similar to SARS-CoV-2 might be causing severe illness and death. Second, we review the evidence on COVID-19 vaccines and make the case that immune system sensitization followed by desensitization to reduce severe outcomes (without necessarily reducing infections) might be an acceptable benchmark for vaccine effectiveness compared to sterilizing immunity (achieving an absolute reduction in infections).

2. SARS-CoV-2—Killer Virus or Just a Trigger for Kitchen Sink Inflammation?

SARS-CoV-2, which causes COVID-19, is a non-segmented, single-stranded, positivesense RNA virus from the β genus of coronaviruses [18,19]. SARS-CoV-2 infection was first identified in Wuhan, Hubei Province, China in December of 2019, and was initially characterized as virally-induced pneumonia by clinicians before it was finally isolated in bronchoalveolar lavage fluid from patients [19,20]. The clinical presentation of COVID-19 ranges from asymptomatic infection to mild respiratory symptoms to severe viral pneumonia [21].

While SARS-CoV-2 initially infects and compromises the respiratory system, it also induces multiorgan dysfunction and damage [22–24]. One of the main factors implicated in COVID-19 multiorgan failure is a massive release of proinflammatory cytokines—formally known as cytokine release syndrome (CRS) but better known by the colloquial term "cytokine storms" [25,26]. The inflammatory reaction in COVID-19 is due to the over-activation of multiple cellular subtypes in the human body [27]. Inhaled viral particles bind to epithelial cells in the nasal mucosa or travel down the nasopharyngeal tract to reach the more distal areas of the airway. The effect of the virus in eliciting an inflammatory response through different cell types is detailed below, and this gives us an idea of how the virus wreaks havoc on homeostasis in the host. Local and systemic inflammation followed by systemic disruption of homeostasis leads to multiorgan symptoms, multiorgan damage, and the high case fatality rate associated with SARS-CoV-2 infection (Figure 1).

2.1. Epithelial System

The epithelial cells are the first line of defense against invading pathogens [28]. The viral particles encounter different kinds of epithelial cells as they travel from the nose and mouth, which are the most common points of entry, all the way down to the alveolar sacs—the sites of gaseous exchange (Figure 1) [29]. The nasal mucosa has different cell types, such as ciliated epithelial cells, mucous cells, and basal cells [30]. These cells express angiotensin converting enzyme-2 (ACE2) and the transmembrane protein serine protease-2 (TMPRSS2) on their plasma membranes [31–33]. ACE2, TMPRSS2, and other plasma membrane proteins aid in the docking of SARS-CoV-2 onto host cells and facilitate endocytosis of the virus [32]. Once the virus has entered host cells, the virus releases its genetic material into the cytoplasm and hijacks the host's cellular machinery to produce more viral particles [34,35]—i.e., the cell is now infected with the virus. The release of viral particles from cells has been detected as early as one hour post-infection; however, 6-8 h post-infection is when significant viral load has been detected and infection of neighboring cells and loss of ciliated epithelium has been observed [36]. The loss of cilia on the epithelium results in reduced mucociliary clearance, which leads to increased local infection and progression of the disease [37,38]. This loss of clearance has been implicated in increased COVID-19 severity in individuals with pre-existing pulmonary inflammatory conditions like asthma [39], cystic fibrosis [40], and chronic obstructive pulmonary disease (COPD), where ciliary function is already impaired [41]. In nasal epithelial cells, there is a

cells

COVID-19 compared to mild cases [42–44]. Nasal Epithelium Ciliated epithelial cells SARS-CoV-2 Uninfected Resident/unactivated Epithelium macrophages Extravasating neutrophil Extravasating macrophage **Resting platelets** Damaged Activated platelets Epithelium **Blood Clot** Endothelial cell Alveolar Endothelium Epithelial cell Virus and Activated cytokine release Macrophage RBCs Alveolar Epithelium NETosis Neutrophil TNFa II -16 IFN-0 IL-8, CCL2, IL-1β Apoptotic

Figure 1. Pathophysiology of COVID-19 and "kitchen sink inflammation".

As the virus moves down the airway, it encounters the bronchial epithelial cells, the mucus cells, and the club cells [30] (Figure 1). In the middle and lower airway, ciliated epithelial cells that become infected lose their cilia and have a denuded appearance, and the levels of proinflammatory signals in these cells correlate with the disease severity [45].

When the virus finally reaches the alveoli (the distal-most regions of the airway) where the epithelium, endothelium, and blood cells interface and enable gas exchange [46], it activates numerous signaling cascades in multiple cell types, which cause the most damage and destruction [47] (Figure 1). In alveoli, the virus primarily infects alveolar type 2 epithelial cells (a.k.a., AT2 cells or type 2 pneumocytes), potentially due to the abundance of ACE2 on their surface [48]. Infection of type 2 pneumocytes seems to be the initial step that triggers a domino of inflammatory signals [49]. It has been shown that alveolar type 1 cells can also be infected, but to a lesser extent [50]. The infection and damage of alveolar pneumocytes results in a cascade of physiological changes, beginning with an increased production of proinflammatory cytokines, such as interleukin (IL)-1 (IL-1), IL-6, IL-8, tumor necrosis factor-alpha (TNF- α), and elevated levels of C-reactive protein (CRP) and D-dimer [51–55]. These inflammatory cytokines result in recruitment of immune cells-mainly neutrophils and macrophages to loci of inflammation. The recruitment of immune cells further exacerbates the situation, as it results in the loss of barrier function of the underlying endothelial layer [56,57]. Alveolar macrophages, which are recruited to sites of damage have been shown to produce various chemokines, such as CCL2, CCL3, CCL7, CCL8, CCL13, CCL20, and cytokines, such as CXCL1, CXCL3, and CXCL10 [58,59]. Among these chemokines, CCL2 and CCL3 attract more monocytes and macrophages to the alveoli and induce CXCR1 gene expression in them; this promotes the production of

delayed or a muted interferon response that is observed after infection. The levels of type I (interferon alpha, beta) and type III (interferon lambda) interferons are lower in severe COVID-19 compared to mild cases [42–44].

tissue-damaging, proinflammatory reactive oxygen species [60]. The sum effect is largescale destruction of epithelial tissue, which promotes the recruitment of neutrophils that try to resolve the situation by forming neutrophil extracellular traps (NET) [61]. The heightened cellular signaling and subsequent tissue damage described above leads to alveolar flooding with the interstitial fluid and the blood, which leads to progressive hypoxia. Thus, the replication of the virus in the alveoli results in the progression towards acute respiratory distress syndrome (ARDS), triggering an imbalance between pro-coagulation and anticoagulation (i.e., pro-fibrinolysis) pathways, plus stimulating complement activation, damage to hyaline membranes, and the formation of clots in the small and large blood vessels [62,63].

A. Nasal epithelium. The primary site of entry for SARS-CoV-2 is the nasal epithelium along with the oral mucosa. The virus binds to ciliated epithelial cells via ACE2 and is endocytosed, thus infecting the host cells. The virus uses host cell machinery to replicate and release more viral particles. The release of the viral particles results in the loss of epithelial barrier integrity, the release of inflammatory mediators, and the aggravation of symptoms. B. Alveolar epithelium. The Alveolar epithelium consists of ciliated type 2 alveolar epithelial cells (AT2) and type 1 non-ciliated (AT1) cells. Infection of the epithelial cells results in the production of a multitude of proinflammatory cytokines that promote extravasation/activation of the neutrophils and macrophages. This results in loss of barrier integrity, alveolar flooding, loss of functional surfactant, diffuse alveolar damage, and damage to the underlying endothelial cells. C. Endothelial cells. The loss of barrier integrity on the epithelial layer promotes further spread of damage to the endothelial layer. Secreted proinflammatory cytokines and chemokines recruit and activate a variety of immune cells (macrophages, neutrophils, and T and B cells). The exposed extracellular matrix also acts as a trigger for neutrophils, which undergo a process of cell death called NETosis; this leads to the aggregation and entanglement of platelets, thus promoting clot formation. Furthermore, T cells that are recruited to the site of injury can also aggravate endothelial injury. Figure created with BioRender.com.

2.2. Endothelial System

The endothelial cells line the innermost layers of the blood vessels (adjacent to the lumen) and tightly regulate the transport of nutrients, gases, metabolic wastes, bioactive molecules, and cells [64]. The endothelial cells lining the smaller capillaries express ACE2 and TMPRSS2, which as mentioned earlier are necessary for the host cell binding and internalization of SARS-CoV-2 [65]. The virus, on entering the endothelial cells, replicates and then disseminates through the blood stream to organs other than the lungs. The endothelial cells respond to the plethora of cytokines/chemokines that are produced by the epithelial cells upon infection. The predominant chemical signals that affect endothelial barrier function are IL-6 and TNF- α . Activation of the endothelial cells by IL-6 or TNF- α results in the production of IL-8 (a major chemoattractant for neutrophils) and monocyte chemoattractant protein (MCP-1) and in the activation of the C5a complement [66]. Indeed, plasma IL-6 levels in patients with COVID-19 correlate with the disease severity [67]. The recruitment of overly activated immune cells, a hallmark of COVID-19, can result in endothelial cell death, which promotes vascular leak and initiation of microfoci for clot formation [68,69]. Activated endothelial cells can release large amounts of von Willebrand factor (VWF) and factor VIII, which play an active role in clot formation [70]. In addition to stimulating the formation of fibrin, endothelial cells also secrete plasminogen activator inhibitor-1 (PAI-1), which inhibits clot dissolution [71]. During the initial stage of infection, the loss of contact with the surrounding cells results in the formation of circulating endothelial cells (CECs), which can travel from one tissue to another. During the course of infection, there is a loss of endothelial barrier integrity, which results in fluid accumulation in the alveolar and pleural spaces. This is a result of the upregulation of interleukin 2 receptor (IL-2R) on the endothelial cells and the increased release of IL-2 from the activated T cells [72]. Thus, direct viral infection or indirect activation of endothelial cells causes a loss of barrier function, which then results in fluid accumulation, increased extravasation

of immune cells, widespread microthrombosis, diffuse fibrin deposits, and secretion of PAI-1 that prevents clot dissolution—all working together to increase the likelihood of thromboembolisms [73,74] (Figure 1).

2.3. The Mismatch between Viral Load and Symptom Severity

In Section 2, we attempted to answer the question as to whether SARS-CoV-2 is a killer virus or just a trigger for "kitchen sink inflammation". The related expressions "to throw everything but the kitchen sink" or "to throw the kitchen sink" imply doing everything possible to address a problem, regardless of whether the solutions are likely to be good or bad [75,76]. Due to the novelty of SARS-CoV-2 to the human immune system, when people were first infected, the immune response was indeed comparable to throwing an inflammatory kitchen sink at the viral invader, resulting in cell-signaling storms [25,26,47,55,77–83]. The fact that some individuals with high viral loads presented with minimal symptoms, and some individuals with severe symptoms had low viral loads suggest that, SARS-CoV-2 by itself is not a killer virus, but rather, it is a nonspecific and overly aggressive inflammatory response, which causes the most harm [84–88]. This is not to diminish the fact that SARS-CoV-2 infection has been implicated in the deaths of nearly seven million people, chronic illness in 10-20% of the survivors, and about 800 million cumulative cases in just over three years [89,90]. However, the available data point towards severe illness and deaths being linked to an inflammatory response, which overwhelms the host, rather than widespread virus-mediated killing of cells in the respiratory system and other vital systems of the body [88]. Interestingly, an overly aggressive immune response, rather than the pathogen itself, might underlie severe illness in various infectious diseases [91]. Therefore, public health agencies must invest resources in finding safe, effective, practical, and standardized therapies, which can work to combat a wide range of existing and novel pathogens. In hindsight, one of the simplest public health strategies to blunt the impact of SARS-CoV-2 would have been to assume that it would behave just like its predecessor SARS-CoV (implicated in the SARS outbreak of the early 2000s) and amplify the importance of commonsense preventative healthcare measures (e.g., hand hygiene, mask wearing, and avoiding unnecessary social activities) until safe and effective pharmacological measures were available [92–94].

3. Calming the Inflammatory Storm by Conditioning the Immune System

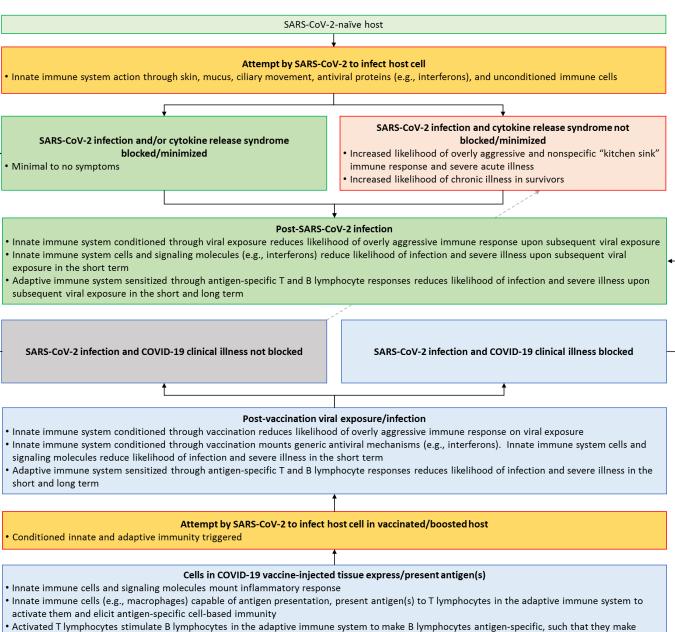
For over a century, vaccines have served as a beacon of hope against infectious diseases [95,96]. From mitigating the severity of various infectious diseases to completely eradicating certain diseases in populations, vaccines are easily one of mankind's greatest innovations in the realm of public health [97,98]. Vaccines come in a variety of forms such as live attenuated, inactivated, recombinant, toxoid, viral vector, and the recently developed messenger RNA (mRNA vaccines) [95,99]. These formulations all follow the same basic principle of exploiting the human immune system by exposing it to an innocuous form, portion, or product of a pathogen to induce long-lasting protection against the pathogen [100]. The benefits of this practice are reaped at an individual and population level [97,98]. The individual, upon exposure to the real pathogen post-vaccination, avoids severe disease outcomes due to his or her immune system being primed against it. On a population level there exists the goal of herd immunity, where entire communities are protected against a pathogen due to the high percentage of individuals who are vaccinated and/or have been exposed to natural infection [101,102]. If the fraction of the population that is vaccinated is low, then those who are not eligible for vaccination due to age, those who are immunocompromised, and those who might be particularly vulnerable to disease complications may not benefit from community-level protection. It is imperative that a proper understanding of vaccines is reached to maximize their effectiveness and attain herd immunity whenever possible for the maximum benefit to individuals and societies. This is especially relevant considering the recent COVID-19 pandemic, where vaccination has played a critical role in disease mitigation.

3.1. How the Most Common COVID-19 Vaccines Work

As highlighted earlier, severe cases of COVID-19 can trigger sepsis, hypoxemia, pneumonia, and tissue damage [21,103]. In line with what is expected of respiratory viruses, SARS-CoV-2 is transmitted mainly via the respiratory route by droplets and aerosolized viral particles and also possesses an incubation period of approximately 5–12 days, making the control of its spread challenging [18–20].

COVID-19 vaccines were in development within less than six months since the beginning of the outbreak, with public health agencies and industry partners around the world committing vast resources to vaccine development, testing, and deployment [104]. Out of these efforts emerged the novel mRNA vaccines, such as those produced by Moderna (Spikevax) and Pfizer-BioNTech (Comirnaty) [99,104]. These mRNA vaccines consist of a segment of mRNA, which has the genetic code to make the SARS-CoV-2's spike protein. Since naked mRNA is not stable, it was packaged in a lipid (a basic chemical component of fats and oils) coat to stabilize the structure and allow it to be readily delivered into cells [99,105–107]. Based on the established understanding of conserved cellular biological processes, we can assume that, once the mRNA-containing lipid spheres are inside a host cell, such as a muscle fiber in the deltoid muscle (the muscle that is most commonly injected), the mRNA information is used by the host cell ribosomes to assemble the amino acid sequence for the spike protein through a process called protein translation [106,107]. After translation, spike protein molecules are processed and presented as an antigen at the surface of the host cell, thereby activating both innate and adaptive immune responses, which confer immune memory [105–107] (Figure 2). Another anti-SARS-CoV-2 vaccine strategy that was widely employed, involved the use of DNA packaged in a harmless adenovirus [107,108]. DNA within the adenovirus contained the code needed for inoculated host cells to generate mRNA in their nuclei through DNA transcription; the mRNA would then exit the nuclei and bind to the ribosomes, and from that point on, the assembling and presentation of spike protein molecules was similar to that of mRNA vaccines [107].

The recognition of "self" from "non-self" is a fundamental aspect of immunity, and this function serves an important role in protecting the body from disease-causing elements [109]. Vaccination takes advantage of this by introducing a harmless version of the disease-causing pathogen, allowing the body to recognize and remember it. There are two phases of immunity that are activated upon infection or inoculation through vaccination: Innate immunity and adaptive immunity [110–112] (Figure 2). Innate immunity is the first line of defense and is activated when pattern recognition receptors detect foreign material or damage, which are known as pathogen- associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs) [110,111,113]. The result is the release of proinflammatory molecules, such as cytokines, which recruit more immune cells to the site of vaccination [110,111,113] (Figure 2). Shortly after this initial response, the adaptive immune system is activated (Figure 2). The adaptive immune system is critical for the control of viral infections, especially in the long term upon re-exposure to the same virus (Figure 2). The main players in the adaptive immune response are T and B lymphocytes, which possess a range of functions that include killing infected cells, activating other lymphocytes, producing antibodies, and generating memory cells [112] (Figure 2). With the production of memory cells and antibodies against SARS-CoV-2 through vaccination, the expected outcome in a vaccinated individual who was subsequently exposed to the actual virus, was a decreased likelihood of infection, decreased viral load if infected, reduced ability to transmit infection, and protection against severe illness (Figure 2). These predictions were indeed borne out by initial clinical trials, thus prompting emergency use authorization and approval of certain mRNA and DNA-adenovirus vaccines [108,114,115].



 Activated T lymphocytes stimulate B lymphocytes in the adaptive immune system to make B lymphocytes antigen-specific, such that they make antigen-specific antibodies

SARS-CoV-2-vaccinated/boosted host

Figure 2. The role of vaccine-induced immune system conditioning in reducing COVID-19 illness severity.

3.2. COVID-19 Vaccination—Sterilizing Immunity or Protection against Severe Illness?

As discussed earlier, vaccination against COVID-19 was predicted to confer sterilizing immunity—i.e., reduce the likelihood of infection and therefore transmission [116]. However, as real-world data from community vaccination programs emerged, it became clear that even those considered to be fully vaccinated (those who completed the required vaccination schedule for their age and health status) were still being infected in high numbers. Infections in individuals who were considered fully vaccinated were called "breakthrough cases" [117–119]. The incidence of breakthrough cases increased as new variants and subvariants of SARS-CoV-2 emerged and created waves of infections that swept across continents [120,121]. The first and earliest variant was identified in the UK and called Alpha, the second was identified in South Africa and called Beta, and the third was found in Brazil and called Gamma; Eta and Delta were the fourth and fifth variants, which were identified in the UK and India, respectively [120]. The Omicron variant was identified in Botswana and is the most highly mutated strain, with twice as many mutations than the Delta variant [121,122]. The incidence of breakthrough cases during the Delta variant wave was high; however, despite the Delta variant being characterized as more aggressive in its symptomatology than prior strains [123,124], the disease outcome in vaccinated individuals was better than in those who were unvaccinated [125–127]. Intriguingly, during the Delta variant wave of COVID-19, viral loads in fully vaccinated individuals were as high as those seen in unvaccinated individuals, indicating that vaccination was reducing the risk of severe illness without reducing host cell viral infection and without blunting the production of viral particles in infected individuals [117,118,128–131]. During the Delta wave, it was also humbling to learn that vaccinated individuals with breakthrough infections were just as likely as unvaccinated individuals to transmit the virus to others—disturbing news at a time when public health agencies in certain countries were relaxing nonpharmacological COVID-19 mitigation measures against a backdrop of vaccine inequity based on age, geography, economics, and other factors [117,132]. The conferring of protection against severe illness and death through vaccination without blocking infection has continued through subsequent waves of COVID-19 infections caused by less aggressive variants and subvariants (e.g., Omicron) [133]. These observations are likely linked to our earlier point about how a nonspecific and overly aggressive inflammatory response to SARS-CoV-2, rather than the extent of viral infection, might be the factor underlying disease severity [88]. Based on the lessons learned from COVID-19 vaccination programs, it is imperative that public health agencies reassess the benchmarks for vaccine effectiveness, since the benefits of vaccination have medical, sociological, and economic implications that extend far beyond the mere conferring of immunity as we know it [97,134]. While aiming to obtain sterilizing, lifelong immunity through vaccination is a fair goal, settling for vaccine-mediated protection against severe illness for fast-spreading diseases like COVID-19 would be an acceptable strategy, so that, the highest number of individuals around the world are rapidly protected and negative economic impact is reduced [102,135].

The flow chart summarizes how the innate and adaptive immune systems likely play vital roles in reducing the risk of severe acute COVID-19 illness through vaccination, natural infection, and natural infection following vaccination [28,136–139]. We make the case that being up to date on COVID-19 vaccination through updated SARS-CoV-2 vaccines/boosters acts as a type of "continuing education" for the immune system. This continuing education enables the immune system to remain conditioned, such that it mounts a calm and focused response to viral exposure and does not overwhelm the host through overly aggressive inflammation. We also indicate that in rare cases even vaccinated individuals might develop severe acute illness and chronic complications (the dotted grey arrow connecting the vaccination story in the flow chart (bottom) to the natural infection story in the flow chart (top)) [140,141].

3.3. Could Immune System Sensitization Followed by Desensitization Explain Vaccine-Induced Protection against Severe COVID-19?

Early in the COVID-19 pandemic, it was suggested that zoonotic transmission (animal to human) from bats was the most likely source of SARS-CoV-2 and caused the first set of infections in humans [142]. Viral genomes sharing > 90% similarity to human SARS-CoV-2 were found in bats and pangolins [143–146]. As the pandemic progressed, there were reports of other animals being infected with SARS-CoV-2, raising concerns that animals could act as viral reservoirs, leading to more human infections [142]. Similar to observations in certain vaccinated and unvaccinated humans, animals that test positive for SARS-CoV-2 may have mild to no symptoms but can still serve as reservoirs/vectors of the virus [147]. These observations also support the idea that severe illness does not usually ensue if the host does not mount a nonspecific and overly aggressive inflammatory response (Figures 1 and 2). Thus, it is likely that vaccination initially acts as a sensitizer to the immune system, which is naïve to SARS-CoV-2, but that through this sensitization, when

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inflammatory response that does not overwhelm the host [148–150] (Figure 2). Immune system desensitization is the process of gradually exposing an individual to an antigen so that their immune system becomes nonreactive [151]. Desensitization has been used for decades to improve clinical outcomes in people with allergies and asthma [151–153]. It is tempting to speculate if small doses of SARS-CoV-2 antigens that are updated to include new variants, might not only be able to act as a form of continuing education to the adaptive immune system, but also act as a way to maintain a conditioned and tolerant innate immune system, which does not overwhelm the host through inflammation when exposed to the actual pathogen [154] (Figure 2). Intriguingly, vaccine desensitization protocols have been successfully used to reduce the likelihood of allergic reactions in individuals who display signs of COVID-19 vaccine hypersensitivity [155–157].

3.4. Cutaneous Delivery of SARS-CoV-2 Vaccines as a Potential Strategy to Rapidly Vaccinate/Boost a Large Number of Individuals

In the preceding section, we presented evidence to suggest that COVID-19 vaccines offer protection to the host against severe illness and death even if they do not reduce the risk of infection. We then made the case that, if protection against severe illness rather than sterilizing immunity is an acceptable benchmark for vaccine effectiveness, it may be possible to achieve this by periodically exposing the immune system to small amounts of updated antigenic material. The logistics of administering updated vaccines to the entire global population on a semiannual or even annual basis is daunting [158]. It is in this context that cutaneous vaccine delivery systems are appealing.

The skin is a viable organ with a large concentration of antigen-presenting cells including the epidermal Langerhans cells and dermal dendritic cells [159]. The minimally invasive microneedle (MN) of cutaneous drug delivery systems reduces safety concerns associated with parenteral administration and increases patient compliance, thus making MN-enhanced transdermal delivery an attractive route for vaccine administration. Different types of antigens including inactivated whole virus, live attenuated virus, virus-like particles, recombinant bacteria, protein subunits, and plasmid DNA have been investigated to demonstrate immune responses following MN delivery. Many promising examples of MN-based vaccine delivery have appeared in recent publications [160,161], a few of which have been highlighted in this section. Antigens can be introduced into the skin using different MN approaches, including intradermal injection (similar to the Mantoux method), stratum corneum disruption by solid MN abrasion, antigen-coated MN, and dissolvable MN insertion [162].

Intradermal delivery using MNs can provide superior antigenicity, as demonstrated through studies with influenza vaccines. Using three different types of influenza vaccines (a whole inactivated influenza virus, a trivalent split-virion human vaccine, and a plasmid DNA encoding the influenza virus hemagglutinin), the MN-based system provided up to a 100-fold dose sparing in rats compared to intramuscular injection for the same induced immune response [163]. Recent clinical trials in adults showed that an MN-based delivery of influenza vaccine can induce immune responses with less antigen (9 μ g) at rates comparable to those with intramuscular (15 μ g) vaccination [164]. A stronger humoral immune response in the elderly was also induced with MN-based intradermal delivery compared to intramuscular injection [165]. A hollow 1.5 mm-long MN Becton Dickinson Soluvia prefilled microinjection system has recently been approved to deliver Fluzone intradermal vaccine (single 0.1 mL dose, Sanofi Pasteur) for influenza prophylaxis [94]. Other MN-based vaccines currently under investigation include a recombinant Bacillus anthracis vaccine formulated with aluminum and administered by hollow MNs [166], a live attenuated chimeric flavivirus vaccine delivered by skin microabrasion for the treatment of Japanese Encephalitis and yellow fever [167], and a smallpox DNA vaccine delivered by MN coating and skin electroporation [168]. It is therefore not surprising that MN-based delivery of COVID-19 vaccines is a topic of interest, which could potentially lead to a

paradigm shift in how large numbers of people could be safely and rapidly vaccinated with fewer logistical hurdles and reduced amounts of biowaste [169].

4. Conclusions

We started this review by highlighting the end of the COVID-19 global health emergency, which is undoubtedly a cause for celebration. We then went on to present two main lessons learned from the COVID-19 pandemic, and how these lessons prompt all public health stakeholders (i.e., the entire global human community) to examine our assumptions about how COVID-19 makes people sick and how COVID-19 vaccines work. Paying heed to these lessons could help improve public health and better prepare global communities for future health emergencies that may arise due to infectious diseases.

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References

- 1. Hu, B.; Guo, H.; Zhou, P.; Shi, Z.L. Characteristics of SARS-CoV-2 and COVID-19. *Nat. Rev. Microbiol.* 2021, 19, 141–154. [CrossRef]
- World Health Organization (WHO). Statement on the Fifteenth Meeting of the Ihr (2005) Emergency Committee on the COVID-19 Pandemic; World Health Organization (WHO): Geneva, Switzerland, 2005; Available online: https://www.who.int/news/ item/05-05-2023-statement-on-the-fifteenth-meeting-of-the-international-health-regulations-(2005)-emergency-committeeregarding-the-coronavirus-disease-(covid-19)-pandemic (accessed on 11 May 2023).
- 3. Wise, J. COVID-19: Who Declares End of Global Health Emergency. *Bmj* **2023**, *381*, 1041. [CrossRef]
- 4. Lenharo, M. Who Declares End to COVID-19's Emergency Phase. *Nature* 2023. [CrossRef]
- 5. Ranney, M.L.; Valerie Griffeth, M.P.H.; Jha, A.K. Critical Supply Shortages—The Need for Ventilators and Personal Protective Equipment during the COVID-19 Pandemic. *N. Engl. J. Med.* **2020**, *382*, e41. [CrossRef]
- Dey, A.K.; Bhan, N.; Rao, N.; Ghule, M.; Chatterji, S.; Raj, A. Factors Affecting Delayed and Non-Receipt of Healthcare during the COVID-19 Pandemic for Women in Rural Maharashtra, India: Evidence from a Cross-Sectional Study. *EClinicalMedicine* 2022, 53, 101741. [CrossRef]
- Rogers, E.H.; Porter, J.N.; Ren, J.J.; Battista, J.J.; Grewal, N.P.; Valentin, N.I. Experiences from the Epicenter: An Observational Report on Converting a Post Anesthesia Care Unit to a COVID-19 Intensive Care Unit. *Int. J. Surg. Glob. Health* 2020, 3, e20. [CrossRef]
- Davis, B.; Bankhead-Kendall, B.K.; Dumas, R.P. A Review of COVID-19's Impact on Modern Medical Systems from a Health Organization Management Perspective. *Health Technol.* 2022, 12, 815–824. [CrossRef]
- Parotto, E.; Lamberti-Castronuovo, A.; Censi, V.; Valente, M.; Atzori, A.; Ragazzoni, L. Exploring Italian Healthcare Facilities Response to COVID-19 Pandemic: Lessons Learned from the Italian Response to COVID-19 Initiative. *Front. Public Health* 2022, 10, 1016649. [CrossRef]
- 10. The Lancet Public, Health. COVID-19 in Spain: A Predictable Storm? Lancet Public Health 2020, 5, e568. [CrossRef]
- Sharma, A.; Gupta, P.; Jha, R. COVID-19: Impact on Health Supply Chain and Lessons to Be Learnt. J. Health Manag. 2020, 22, 248–261. [CrossRef]
- 12. Usher, A.D. Medical Oxygen Crisis: A Belated COVID-19 Response. Lancet 2021, 397, 868–869. [CrossRef]
- 13. Feinmann, J. How COVID-19 Revealed the Scandal of Medical Oxygen Supplies Worldwide. Bmj 2021, 373, n1166. [CrossRef]

- 14. Tenforde, M.W.; Self, W.H.; Adams, K.; Gaglani, M.; Ginde, A.A.; McNeal, T.; Ghamande, S.; Douin, D.J.; Talbot, H.K.; Casey, J.D.; et al. Association between Mrna Vaccination and COVID-19 Hospitalization and Disease Severity. *JAMA* 2021, *326*, 2043–2054. [CrossRef]
- 15. Moghadas, S.M.; Vilches, T.N.; Zhang, K.; Wells, C.R.; Shoukat, A.; Singer, B.H.; Meyers, L.A.; Neuzil, K.M.; Langley, J.M.; Fitzpatrick, M.C.; et al. The Impact of Vaccination on COVID-19 Outbreaks in the United States. *medRxiv* 2021. [CrossRef]
- Watson, O.J.; Barnsley, G.; Toor, J.; Hogan, A.B.; Winskill, P.; Ghani, A.C. Ghani. Global Impact of the First Year of COVID-19 Vaccination: A Mathematical Modelling Study. *Lancet Infect. Dis.* 2022, 22, 1293–1302. [CrossRef]
- 17. Torreele, E.; Kazatchkine, M.; Liu, J.; Dybul, M.; Cárdenas, M.; Singh, S.; Quigley, H.L.; McNab, C.; Sirleaf, E.J.; Mazzucato, M.; et al. Stopping Epidemics When and Where They Occur. *Lancet* **2023**, *401*, 324–328. [CrossRef]
- Samudrala, P.K.; Kumar, P.; Choudhary, K.; Thakur, N.; Wadekar, G.S.; Dayaramani, R.; Agrawal, M.; Alexander, A. Virology, Pathogenesis, Diagnosis and in-Line Treatment of COVID-19. *Eur. J. Pharmacol.* 2020, 883, 173375. [CrossRef]
- 19. Jin, Y.; Yang, H.; Ji, W.; Wu, W.; Chen, S.; Zhang, W.; Duan, G. Virology, Epidemiology, Pathogenesis, and Control of COVID-19. *Viruses* **2020**, *12*, 372. [CrossRef]
- Ciotti, M.; Ciccozzi, M.; Terrinoni, A.; Jiang, W.C.; Wang, C.B.; Bernardini, S. The COVID-19 Pandemic. Crit. Rev. Clin. Lab Sci. 2020, 57, 365–388. [CrossRef]
- Huang, H.; Cai, S.; Li, Y.; Li, Y.; Fan, Y.; Li, L.; Lei, C.; Tang, X.; Hu, F.; Li, F.; et al. Prognostic Factors for COVID-19 Pneumonia Progression to Severe Symptoms Based on Earlier Clinical Features: A Retrospective Analysis. *Front. Med.* 2020, 7, 557453. [CrossRef]
- 22. Morrow, A.J.; Sykes, R.; McIntosh, A.; Kamdar, A.; Bagot, C.; Bayes, H.K.; Blyth, K.G.; Briscoe, M.; Bulluck, H.; Carrick, D.; et al. A Multisystem, Cardio-Renal Investigation of Post-COVID-19 Illness. *Nat. Med.* **2022**, *28*, 1303–1313. [CrossRef]
- Gupta, A.; Madhavan, M.V.; Sehgal, K.; Nair, N.; Mahajan, S.; Sehrawat, T.S.; Bikdeli, B.; Ahluwalia, N.; Ausiello, J.C.; Wan, E.Y.; et al. Extrapulmonary Manifestations of COVID-19. *Nat. Med.* 2020, 26, 1017–1032. [CrossRef]
- 24. Mokhtari, T.; Hassani, F.; Ghaffari, N.; Ebrahimi, B.; Yarahmadi, A.; Hassanzadeh, G. COVID-19 and multiorgan failure: A narrative review on potential mechanisms. *Histochem. J.* **2020**, *51*, 613–628. [CrossRef]
- 25. Que, Y.; Hu, C.; Wan, K.; Hu, P.; Wang, R.; Luo, J.; Li, T.; Ping, R.; Hu, Q.; Sun, Y.; et al. Cytokine Release Syndrome in COVID-19: A Major Mechanism of Morbidity and Mortality. *Int. Rev. Immunol.* **2022**, *41*, 217–230. [CrossRef]
- Montazersaheb, S.; Khatibi, S.M.H.; Hejazi, M.S.; Tarhriz, V.; Farjami, A.; Sorbeni, F.G.; Farahzadi, R.; Ghasemnejad, T. Ghasemian Sorbeni, R. Farahzadi, and T. Ghasemnejad. COVID-19 Infection: An Overview on Cytokine Storm and Related Interventions. *Virol. J.* 2022, 19, 92. [CrossRef]
- 27. Mattoo SU, S.; Kim, S.J.; Ahn, D.G.; Myoung, J. Escape and over-Activation of Innate Immune Responses by SARS-CoV-2: Two Faces of a Coin. *Viruses* **2022**, *14*, 530. [CrossRef]
- Marshall, J.S.; Warrington, R.; Watson, W.; Kim, H.L. An Introduction to Immunology and Immunopathology. *Allergy Asthma Clin. Immunol.* 2018, 14, 49. [CrossRef]
- Murgolo, N.; Therien, A.G.; Howell, B.; Klein, D.; Koeplinger, K.; Lieberman, L.A.; Adam, G.C.; Flynn, J.; McKenna, P.; Swaminathan, G.; et al. SARS-CoV-2 tropism, entry, replication, and propagation: Considerations for drug discovery and development. *PLoS Pathog.* 2021, 17, e1009225. [CrossRef]
- Gengler, I.; Wang, J.C.; Speth, M.M.; Sedaghat, A.R. Sinonasal pathophysiology of SARS-CoV-2 and COVID-19: A systematic review of the current evidence. *Laryngoscope Investig. Otolaryngol.* 2020, *5*, 354–359. [CrossRef]
- Wruck, W.; Adjaye, J. SARS-CoV-2 receptor ACE2 is co-expressed with genes related to transmembrane serine proteases, viral entry, immunity and cellular stress. *Sci. Rep.* 2020, *10*, 21415. [CrossRef]
- Sungnak, W.; Huang, N.; Becavin, C.; Berg, M.; Queen, R.; Litvinukova, M.; Talavera-Lopez, C.; Maatz, H.; Reichart, D.; Sampaziotis, F.; et al. SARS-CoV-2 entry factors are highly expressed in nasal epithelial cells together with innate immune genes. *Nat. Med.* 2020, 26, 681–687. [CrossRef]
- 33. Collin, J.; Queen, R.; Zerti, D.; Dorgau, B.; Georgiou, M.; Djidrovski, I.; Hussain, R.; Coxhead, J.M.; Joseph, A.; Rooney, P.; et al. Co-expression of SARS-CoV-2 entry genes in the superficial adult human conjunctival, limbal and corneal epithelium suggests an additional route of entry via the ocular surface. *Ocul. Surf.* 2021, *19*, 190–200. [CrossRef]
- 34. V'kovski, P.; Kratzel, A.; Steiner, S.; Stalder, H.; Thiel, V. Coronavirus biology and replication: Implications for SARS-CoV-2. *Nat. Rev. Microbiol.* **2021**, *19*, 155–170. [CrossRef]
- 35. Hillen, H.S.; Kokic, G.; Farnung, L.; Dienemann, C.; Tegunov, D.; Cramer, P. Structure of replicating SARS-CoV-2 polymerase. *Nature* **2020**, *584*, 154–156. [CrossRef]
- Gamage, A.M.; Tan, K.S.; Chan, W.O.Y.; Liu, J.; Tan, C.W.; Ong, Y.K.; Thong, M.; Andiappan, A.K.; Anderson, D.E.; Wang, Y.; et al. Infection of human Nasal Epithelial Cells with SARS-CoV-2 and a 382-nt deletion isolate lacking ORF8 reveals similar viral kinetics and host transcriptional profiles. *PLoS Pathog.* 2020, *16*, e1009130. [CrossRef]
- 37. Zhu, N.; Wang, W.; Liu, Z.; Liang, C.; Wang, W.; Ye, F.; Huang, B.; Zhao, L.; Wang, H.; Zhou, W.; et al. Morphogenesis and cytopathic effect of SARS-CoV-2 infection in human airway epithelial cells. *Nat. Commun.* **2020**, *11*, 3910. [CrossRef]
- Ozturk, E.O.; Aslan, M.; Bayindir, T. The effect of COVID-19 on nasal mucociliary clearance. Acta Otolaryngol. 2022, 142, 329–332.
 [CrossRef]
- 39. Thomas, B.; Rutman, A.; Hirst, R.A.; Haldar, P.; Wardlaw, A.J.; Bankart, J.; Brightling, C.E.; O'Callaghan, C. Ciliary dysfunction and ultrastructural abnormalities are features of severe asthma. *J. Allergy Clin. Immunol.* **2010**, 126, 722–729.e2. [CrossRef]

- 40. Leung, H.M.; Birket, S.E.; Hyun, C.; Ford, T.N.; Cui, D.; Solomon, G.M.; Shei, R.J.; Adewale, A.T.; Lenzie, A.R.; Fernandez-Petty, C.M.; et al. Intranasal micro-optical coherence tomography imaging for cystic fibrosis studies. *Sci. Transl. Med.* **2019**, *11*, eaav3505. [CrossRef]
- Yaghi, A.; Zaman, A.; Cox, G.; Dolovich, M.B. Ciliary beating is depressed in nasal cilia from chronic obstructive pulmonary disease subjects. *Respir. Med.* 2012, 106, 1139–1147. [CrossRef]
- Fiege, J.K.; Thiede, J.M.; Nanda, H.A.; Matchett, W.E.; Moore, P.J.; Montanari, N.R.; Thielen, B.K.; Daniel, J.; Stanley, E.; Hunter, R.C.; et al. Single cell resolution of SARS-CoV-2 tropism, antiviral responses, and susceptibility to therapies in primary human airway epithelium. *PLoS Pathog.* 2021, 17, e1009292. [CrossRef]
- Galani, I.-E.; Rovina, N.; Lampropoulou, V.; Triantafyllia, V.; Manioudaki, M.; Pavlos, E.; Koukaki, E.; Fragkou, P.C.; Panou, V.; Rapti, V.; et al. Untuned antiviral immunity in COVID-19 revealed by temporal type I/III interferon patterns and flu comparison. *Nat. Immunol.* 2021, 22, 32–40. [CrossRef] [PubMed]
- Kim, Y.-M.; Shin, E.-C. Type I and III interferon responses in SARS-CoV-2 infection. *Exp. Mol. Med.* 2021, 53, 750–760. [CrossRef] [PubMed]
- Ziegler, C.G.; Miao, V.N.; Owings, A.H.; Navia, A.W.; Tang, Y.; Bromley, J.D.; Lotfy, P.; Sloan, M.; Laird, H.; Williams, H.B. Impaired local intrinsic immunity to SARS-CoV-2 infection in severe COVID-19. *Cell* 2021, *184*, 4713–4733.e4722. [CrossRef] [PubMed]
- 46. Lamers, M.M.; Haagmans, B.L. SARS-CoV-2 pathogenesis. Nat. Rev. Microbiol. 2022, 20, 270–284. [CrossRef]
- 47. Bosmuller, H.; Matter, M.; Fend, F.; Tzankov, A. The pulmonary pathology of COVID-19. *Virchows Arch.* **2021**, 478, 137–150. [CrossRef]
- Zou, X.; Chen, K.; Zou, J.; Han, P.; Hao, J.; Han, Z. Single-cell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable to 2019-nCoV infection. *Front. Med.* 2020, 14, 185–192. [CrossRef]
- Huang, J.; Hume, A.J.; Abo, K.M.; Werder, R.B.; Villacorta-Martin, C.; Alysandratos, K.D.; Beermann, M.L.; Simone-Roach, C.; Lindstrom-Vautrin, J.; Olejnik, J.; et al. SARS-CoV-2 Infection of Pluripotent Stem Cell-Derived Human Lung Alveolar Type 2 Cells Elicits a Rapid Epithelial-Intrinsic Inflammatory Response. *Cell Stem Cell* 2020, 27, 962–973.e967. [CrossRef]
- Rockx, B.; Kuiken, T.; Herfst, S.; Bestebroer, T.; Lamers, M.M.; Oude Munnink, B.B.; de Meulder, D.; van Amerongen, G.; van den Brand, J.; Okba, N.M.A.; et al. Comparative pathogenesis of COVID-19, MERS, and SARS in a nonhuman primate model. *Science* 2020, 368, 1012–1015. [CrossRef]
- Queiroz, M.A.F.; Neves, P.; Lima, S.S.; Lopes, J.D.C.; Torres, M.; Vallinoto, I.; Bichara, C.D.A.; Dos Santos, E.F.; de Brito, M.; da Silva, A.L.S.; et al. Cytokine Profiles Associated With Acute COVID-19 and Long COVID-19 Syndrome. *Front. Cell Infect. Microbiol.* 2022, 12, 922422. [CrossRef]
- Wilson, J.G.; Simpson, L.J.; Ferreira, A.-M.; Rustagi, A.; Roque, J.; Asuni, A.; Ranganath, T.; Grant, P.M.; Subramanian, A.; Rosenberg-Hasson, Y.; et al. Cytokine profile in plasma of severe COVID-19 does not differ from ARDS and sepsis. *JCI Insight* 2020, 5, 289. [CrossRef] [PubMed]
- 53. Huang, I.; Pranata, R.; Lim, M.A.; Oehadian, A.; Alisjahbana, B. C-reactive protein, procalcitonin, D-dimer, and ferritin in severe coronavirus disease-2019: A meta-analysis. *Ther. Adv. Respir. Dis.* **2020**, *14*, 1753466620937175. [CrossRef]
- Bello, S.; Lasierra, A.B.; López-Vergara, L.; de Diego, C.; Torralba, L.; de Gopegui, P.R.; Lahoz, R.; Abadía, C.; Godino, J.; Cebollada, A.; et al. IL-6 and cfDNA monitoring throughout COVID-19 hospitalization are accurate markers of its outcomes. *Respir. Res.* 2023, 24, 125. [CrossRef]
- 55. Frisoni, P.; Neri, M.; D'Errico, S.; Alfieri, L.; Bonuccelli, D.; Cingolani, M.; Di Paolo, M.; Gaudio, R.M.; Lestani, M.; Marti, M.; et al. Cytokine storm and histopathological findings in 60 cases of COVID-19-related death: From viral load research to immunohistochemical quantification of major players IL-1β, IL-6, IL-15 and TNF-α. *Forensic. Sci. Med. Pathol.* 2022, 18, 4–19. [CrossRef] [PubMed]
- 56. Meyer, N.J.; Gattinoni, L.; Calfee, C.S. Acute respiratory distress syndrome. Lancet 2021, 398, 622–637. [CrossRef]
- Millar, F.R.; Summers, C.; Griffiths, M.J.; Toshner, M.R.; Proudfoot, A.G. The pulmonary endothelium in acute respiratory distress syndrome: Insights and therapeutic opportunities. *Thorax* 2016, 71, 462–473. [CrossRef] [PubMed]
- Grant, R.A.; Morales-Nebreda, L.; Markov, N.S.; Swaminathan, S.; Querrey, M.; Guzman, E.R.; Abbott, D.A.; Donnelly, H.K.; Donayre, A.; Goldberg, I.A.; et al. Circuits between infected macrophages and T cells in SARS-CoV-2 pneumonia. *Nature* 2021, 590, 635–641. [CrossRef] [PubMed]
- Chua, R.L.; Lukassen, S.; Trump, S.; Hennig, B.P.; Wendisch, D.; Pott, F.; Debnath, O.; Thurmann, L.; Kurth, F.; Volker, M.T.; et al. COVID-19 severity correlates with airway epithelium-immune cell interactions identified by single-cell analysis. *Nat. Biotechnol.* 2020, *38*, 970–979. [CrossRef] [PubMed]
- 60. Rendeiro, A.F.; Ravichandran, H.; Bram, Y.; Chandar, V.; Kim, J.; Meydan, C.; Park, J.; Foox, J.; Hether, T.; Warren, S.; et al. The spatial landscape of lung pathology during COVID-19 progression. *Nature* **2021**, *593*, 564–569. [CrossRef] [PubMed]
- 61. Galani, I.E.; Andreakos, E. Neutrophils in viral infections: Current concepts and caveats. J. Leukoc. Biol. 2015, 98, 557–564. [CrossRef]
- Biswas, S.; Thakur, V.; Kaur, P.; Khan, A.; Kulshrestha, S.; Kumar, P. Blood clots in COVID-19 patients: Simplifying the curious mystery. *Med. Hypotheses* 2021, 146, 110371. [CrossRef] [PubMed]
- Tuculeanu, G.; Barbu, E.C.; Lazar, M.; Chitu-Tisu, C.E.; Moisa, E.; Negoita, S.I.; Ion, D.A. Coagulation Disorders in Sepsis and COVID-19— Two Sides of the Same Coin? A Review of Inflammation– Coagulation Crosstalk in Bacterial Sepsis and COVID-19. J. Clin. Med. 2023, 12, 601. [PubMed]

- 64. Bkaily, G.; Jacques, D. Morphological and Functional Remodeling of Vascular Endothelium in Cardiovascular Diseases. Int. J. Mol. Sci. 2023, 24, 1998. [CrossRef] [PubMed]
- Hoffmann, M.; Kleine-Weber, H.; Schroeder, S.; Kruger, N.; Herrler, T.; Erichsen, S.; Schiergens, T.S.; Herrler, G.; Wu, N.H.; Nitsche, A.; et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* 2020, *181*, 271–280.e278. [CrossRef]
- 66. Xu, S.-W.; Ilyas, I.; Weng, J.-P. Endothelial dysfunction in COVID-19: An overview of evidence, biomarkers, mechanisms and potential therapies. *Acta Pharmacol. Sin.* **2023**, *44*, 695–709. [CrossRef]
- Chen, X.; Zhao, B.; Qu, Y.; Chen, Y.; Xiong, J.; Feng, Y.; Men, D.; Huang, Q.; Liu, Y.; Yang, B.; et al. Detectable Serum Severe Acute Respiratory Syndrome Coronavirus 2 Viral Load (RNAemia) Is Closely Correlated With Drastically Elevated Interleukin 6 Level in Critically III Patients With Coronavirus Disease 2019. *Clin. Infect. Dis.* 2020, *71*, 1937–1942. [CrossRef]
- 68. Kaur, S.; Tripathi, D.M.; Yadav, A. The Enigma of Endothelium in COVID-19. Front. Physiol. 2020, 11, 989. [CrossRef]
- 69. Mosleh, W.; Chen, K.; Pfau, S.E.; Vashist, A. Endotheliitis and Endothelial Dysfunction in Patients with COVID-19: Its Role in Thrombosis and Adverse Outcomes. *J. Clin. Med.* **2020**, *9*, 1862. [CrossRef]
- 70. Mojzisch, A.; Brehm, M.A. The Manifold Cellular Functions of von Willebrand Factor. Cells 2021, 10, 2351. [CrossRef]
- 71. Huertas, A.; Montani, D.; Savale, L.; Pichon, J.; Tu, L.; Parent, F.; Guignabert, C.; Humbert, M. Endothelial cell dysfunction: A major player in SARS-CoV-2 infection (COVID-19)? *Eur. Respir. J.* **2020**, *56*, 2001634. [CrossRef]
- 72. Eurelings, L.E.M.; Miedema, J.R.; Dalm, V.; van Daele, P.L.A.; van Hagen, P.M.; van Laar, J.A.M.; Dik, W.A. Sensitivity and specificity of serum soluble interleukin-2 receptor for diagnosing sarcoidosis in a population of patients suspected of sarcoidosis. *PLoS ONE* **2019**, *14*, e0223897. [CrossRef]
- 73. Marini, J.J.; Gattinoni, L. Management of COVID-19 Respiratory Distress. JAMA 2020, 323, 2329–2330. [CrossRef]
- Suh, Y.J.; Hong, H.; Ohana, M.; Bompard, F.; Revel, M.P.; Valle, C.; Gervaise, A.; Poissy, J.; Susen, S.; Hekimian, G.; et al. Pulmonary Embolism and Deep Vein Thrombosis in COVID-19: A Systematic Review and Meta-Analysis. *Radiology* 2021, 298, E70–E80. [CrossRef] [PubMed]
- 75. Simpson, R.; Mendis, D. A Corpus-Based Study of Idioms in Academic Speech. TESOL Q. 2003, 37, 419–441. [CrossRef]
- Cambridge University Press & Assessment. Everything but the Kitchen Sink. Available online: https://dictionary.cambridge. org/us/dictionary/english/everything-but-the-kitchen-sink (accessed on 15 May 2023).
- 77. Roche, J.A.; Roche, R. A hypothesized role for dysregulated bradykinin signaling in COVID-19 respiratory complications. *FASEB J.* **2020**, *34*, 7265–7269. [CrossRef] [PubMed]
- 78. Wilczynski, S.A.; Wenceslau, C.F.; McCarthy, C.G.; Webb, R.C. A Cytokine/Bradykinin Storm Comparison: What Is the Relationship between Hypertension and COVID-19? *Am. J. Hypertens.* **2021**, *34*, 304–306. [CrossRef]
- 79. McCarthy, C.G.; Wilczynski, S.; Wenceslau, C.F.; Webb, R.C. A new storm on the horizon in COVID-19: Bradykinin-induced vascular complications. *Vasc. Pharmacol.* **2021**, *137*, 106826. [CrossRef] [PubMed]
- Garvin, M.R.; Alvarez, C.; Miller, J.I.; Prates, E.T.; Walker, A.M.; Amos, B.K.; Mast, A.E.; Justice, A.; Aronow, B.; Jacobson, D. A mechanistic model and therapeutic interventions for COVID-19 involving a RAS-mediated bradykinin storm. *eLife* 2020, *9*, e59177. [CrossRef]
- Hojyo, S.; Uchida, M.; Tanaka, K.; Hasebe, R.; Tanaka, Y.; Murakami, M.; Hirano, T. How COVID-19 induces cytokine storm with high mortality. *Inflamm. Regen.* 2020, 40, 37. [CrossRef]
- 82. Hu, B.; Huang, S.; Yin, L. The cytokine storm and COVID-19. J. Med. Virol. 2021, 93, 250–256. [CrossRef]
- Jiang, Y.; Rubin, L.; Peng, T.; Liu, L.; Xing, X.; Lazarovici, P.; Zheng, W. Cytokine storm in COVID-19: From viral infection to immune responses, diagnosis and therapy. *Int. J. Biol. Sci.* 2022, *18*, 459–472. [CrossRef] [PubMed]
- Rabaan, A.A.; Tirupathi, R.; Sule, A.A.; Aldali, J.; Mutair, A.A.; Alhumaid, S.; Muzaheed; Gupta, N.; Koritala, T.; Adhikari, R.; et al. Viral Dynamics and Real-Time RT-PCR Ct Values Correlation with Disease Severity in COVID-19. *Diagnostics* 2021, 11, 1091. [CrossRef] [PubMed]
- 85. Abdulrahman, A.; Mallah, S.I.; Alqahtani, M. COVID-19 viral load not associated with disease severity: Findings from a retrospective cohort study. *BMC Infect. Dis.* **2021**, *21*, 688. [CrossRef] [PubMed]
- Hasanoglu, I.; Korukluoglu, G.; Asilturk, D.; Cosgun, Y.; Kalem, A.K.; Altas, A.B.; Kayaaslan, B.; Eser, F.; Kuzucu, E.A.; Guner, R. Higher viral loads in asymptomatic COVID-19 patients might be the invisible part of the iceberg. *Infection* 2021, 49, 117–126. [CrossRef] [PubMed]
- Kim, Y.; Cheon, S.; Jeong, H.; Park, U.; Ha, N.-Y.; Lee, J.; Sohn, K.M.; Kim, Y.-S.; Cho, N.-H. Differential Association of Viral Dynamics with Disease Severity Depending on Patients' Age Group in COVID-19. *Front. Microbiol.* 2021, 12, 712260. [CrossRef] [PubMed]
- 88. Dorward, D.A.; Russell, C.D.; Um, I.H.; Elshani, M.; Armstrong, S.D.; Penrice-Randal, R.; Millar, T.; Lerpiniere, C.E.B.; Tagliavini, G.; Hartley, C.S.; et al. Tissue-Specific Immunopathology in Fatal COVID-19. *Am. J. Respir. Crit. Care Med.* 2020, 203, 192–201. [CrossRef]
- World Health Organization (WHO). WHO Coronavirus (COVID-19) Dashboard. Available online: https://covid19.who.int/ (accessed on 15 May 2023).
- 90. World Health Organization (WHO). Coronavirus Disease (COVID-19): Post COVID-19 Condition. Available online: https://www.who.int/news-room/questions-and-answers/item/coronavirus-disease-(covid-19)-post-covid-19-condition (accessed on 15 May 2023).

- Morris, G.; Bortolasci, C.C.; Puri, B.K.; Marx, W.; O'Neil, A.; Athan, E.; Walder, K.; Berk, M.; Olive, L.; Carvalho, A.F.; et al. The cytokine storms of COVID-19, H1N1 influenza, CRS and MAS compared. Can one sized treatment fit all? *Cytokine* 2021, 144, 155593. [CrossRef]
- Fung, T.S.; Liu, D.X. Similarities and Dissimilarities of COVID-19 and Other Coronavirus Diseases. *Annu. Rev. Microbiol.* 2021, 75, 19–47. [CrossRef]
- Hirano, T.; Murakami, M. COVID-19: A New Virus, but a Familiar Receptor and Cytokine Release Syndrome. *Immunity* 2020, 52, 731–733. [CrossRef]
- Bo, Y.; Guo, C.; Lin, C.; Zeng, Y.; Li, H.B.; Zhang, Y.; Hossain, M.S.; Chan, J.W.M.; Yeung, D.W.; Kwok, K.O.; et al. Effectiveness of non-pharmaceutical interventions on COVID-19 transmission in 190 countries from 23 January to 13 April 2020. *Int. J. Infect. Dis.* 2021, 102, 247–253. [CrossRef]
- 95. Plotkin, S. History of vaccination. Proc. Natl. Acad. Sci. USA 2014, 111, 12283–12287. [CrossRef] [PubMed]
- 96. Plotkin, S.A.; Plotkin, S.L. The development of vaccines: How the past led to the future. *Nat. Rev. Microbiol.* **2011**, *9*, 889–893. [CrossRef] [PubMed]
- Bloom, D.E.P.; Cadarette, D.; Ferranna, M.P. The Societal Value of Vaccination in the Age of COVID-19. *Am. J. Public Health* 2021, 111, 1049–1054. [CrossRef] [PubMed]
- Bloom, D.E.; Canning, D.; Weston, M. The value of vaccination. In *Fighting the Diseases of Poverty*; Routledge: Oxfordshire, UK, 2017; pp. 214–238.
- Gote, V.; Bolla, P.K.; Kommineni, N.; Butreddy, A.; Nukala, P.K.; Palakurthi, S.S.; Khan, W. A Comprehensive Review of mRNA Vaccines. Int. J. Mol. Sci. 2023, 24, 2700. [CrossRef] [PubMed]
- Irvine, D.J.; Aung, A.; Silva, M. Controlling timing and location in vaccines. Adv. Drug Deliv. Rev. 2020, 158, 91–115. [CrossRef]
 [PubMed]
- 101. Vanderslott, S.; Dattani, S.; Spooner, F.; Roser, M. Vaccination. Our World Data 2013.
- 102. Randolph, H.E.; Barreiro, L.B. Herd Immunity: Understanding COVID-19. Immunity 2020, 52, 737-741. [CrossRef]
- Kamal, M.; Abo Omirah, M.; Hussein, A.; Saeed, H. Assessment and characterisation of post-COVID-19 manifestations. *Int. J. Clin. Pract.* 2021, 75, e13746. [CrossRef]
- Li, Y.; Tenchov, R.; Smoot, J.; Liu, C.; Watkins, S.; Zhou, Q. A Comprehensive Review of the Global Efforts on COVID-19 Vaccine Development. ACS Cent. Sci. 2021, 7, 512–533. [CrossRef]
- 105. Schoenmaker, L.; Witzigmann, D.; Kulkarni, J.A.; Verbeke, R.; Kersten, G.; Jiskoot, W.; Crommelin, D.J.A. mRNA-lipid nanoparticle COVID-19 vaccines: Structure and stability. *Int. J. Pharm.* 2021, 601, 120586. [CrossRef]
- 106. Liang, Y.; Huang, L.; Liu, T. Development and Delivery Systems of mRNA Vaccines. Front. Bioeng. Biotechnol. 2021, 9, 718753. [CrossRef] [PubMed]
- Mascellino, M.T.; Di Timoteo, F.; De Angelis, M.; Oliva, A. Overview of the Main Anti-SARS-CoV-2 Vaccines: Mechanism of Action, Efficacy and Safety. *Infect. Drug Resist.* 2021, 14, 3459–3476. [CrossRef] [PubMed]
- 108. Voysey, M.; Clemens, S.A.C.; Madhi, S.A.; Weckx, L.Y.; Folegatti, P.M.; Aley, P.K.; Angus, B.; Baillie, V.L.; Barnabas, S.L.; Bhorat, Q.E.; et al. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: An interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. *Lancet* 2021, 397, 99–111. [CrossRef] [PubMed]
- 109. Schlee, M.; Hartmann, G. Discriminating self from non-self in nucleic acid sensing. *Nat. Rev. Immunol.* **2016**, *16*, 566–580. [CrossRef]
- 110. Murphy, K.; Weaver, C. Janeway's Immunobiology; Garland Science: New York, NY, USA, 2016.
- 111. Kasuga, Y.; Zhu, B.; Jang, K.J.; Yoo, J.S. Innate immune sensing of coronavirus and viral evasion strategies. *Exp. Mol. Med.* **2021**, *53*, 723–736. [CrossRef]
- 112. Sette, A.; Crotty, S. Adaptive immunity to SARS-CoV-2 and COVID-19. Cell 2021, 184, 861–880. [CrossRef]
- 113. Liu, G.; Gack, M.U. Distinct and Orchestrated Functions of RNA Sensors in Innate Immunity. Immunity 2020, 53, 26–42. [CrossRef]
- 114. Polack, F.P.; Thomas, S.J.; Kitchin, N.; Absalon, J.; Gurtman, A.; Lockhart, S.; Perez, J.L.; Perez Marc, G.; Moreira, E.D.; Zerbini, C.; et al. Safety and Efficacy of the BNT162b2 mRNA COVID-19 Vaccine. *N. Engl. J. Med.* **2020**, *383*, 2603–2615. [CrossRef]
- 115. Baden, L.R.; El Sahly, H.M.; Essink, B.; Kotloff, K.; Frey, S.; Novak, R.; Diemert, D.; Spector, S.A.; Rouphael, N.; Creech, C.B.; et al. Efficacy and Safety of the mRNA-1273 SARS-CoV-2 Vaccine. *N. Engl. J. Med.* **2020**, *384*, 403–416. [CrossRef]
- 116. Wahl, I.; Wardemann, H. Sterilizing immunity: Understanding COVID-19. Immunity 2022, 55, 2231–2235. [CrossRef]
- 117. Brown, C.M.; Vostok, J.; Johnson, H.; Burns, M.; Gharpure, R.; Sami, S.; Sabo, R.T.; Hall, N.; Foreman, A.; Schubert, P.L. Outbreak of SARS-CoV-2 infections, including COVID-19 vaccine breakthrough infections, associated with large public gatherings—Barnstable County, Massachusetts, July 2021. *Morb. Mortal. Wkly. Rep.* 2021, 70, 1059. [CrossRef] [PubMed]
- 118. Jung, J.; Kim, J.Y.; Park, H.; Park, S.; Lim, J.S.; Lim, S.Y.; Bae, S.; Lim, Y.J.; Kim, E.O.; Kim, J.; et al. Transmission and Infectious SARS-CoV-2 Shedding Kinetics in Vaccinated and Unvaccinated Individuals. *JAMA Netw. Open* 2022, 5, e2213606. [CrossRef] [PubMed]
- Kustin, T.; Harel, N.; Finkel, U.; Perchik, S.; Harari, S.; Tahor, M.; Caspi, I.; Levy, R.; Leshchinsky, M.; Ken Dror, S.; et al. Evidence for increased breakthrough rates of SARS-CoV-2 variants of concern in BNT162b2-mRNA-vaccinated individuals. *Nat. Med.* 2021, 27, 1379–1384. [CrossRef] [PubMed]
- Zhao, Y.; Huang, J.; Zhang, L.; Chen, S.; Gao, J.; Jiao, H. The global transmission of new coronavirus variants. *Environ. Res.* 2022, 206, 112240. [CrossRef] [PubMed]

- 121. He, X.; Hong, W.; Pan, X.; Lu, G.; Wei, X. SARS-CoV-2 Omicron variant: Characteristics and prevention. *MedComm* **2021**, *2*, 838–845. [CrossRef]
- 122. Tian, D.; Sun, Y.; Xu, H.; Ye, Q. The emergence and epidemic characteristics of the highly mutated SARS-CoV-2 Omicron variant. J. Med. Virol. 2022, 94, 2376–2383. [CrossRef]
- 123. Twohig, K.A.; Nyberg, T.; Zaidi, A.; Thelwall, S.; Sinnathamby, M.A.; Aliabadi, S.; Seaman, S.R.; Harris, R.J.; Hope, R.; Lopez-Bernal, J.; et al. Hospital admission and emergency care attendance risk for SARS-CoV-2 delta (B.1.617.2) compared with alpha (B.1.17) variants of concern: A cohort study. *Lancet Infect. Dis.* **2022**, *22*, 35–42. [CrossRef]
- 124. Suresh, M.R.; Dries, D.J. Delta and the Variants. Air Med. J. 2022, 41, 167–171. [CrossRef]
- 125. McDonald, I.; Murray, S.M.; Reynolds, C.J.; Altmann, D.M.; Boyton, R.J. Comparative systematic review and meta-analysis of reactogenicity, immunogenicity and efficacy of vaccines against SARS-CoV-2. *NPJ Vaccines* **2021**, *6*, 74. [CrossRef]
- 126. Novelli, G.; Biancolella, M.; Mehrian-Shai, R.; Colona, V.L.; Brito, A.F.; Grubaugh, N.D.; Vasiliou, V.; Luzzatto, L.; Reichardt, J.K. COVID-19 one year into the pandemic: From genetics and genomics to therapy, vaccination, and policy. *Hum. Genom.* 2021, 15, 1–13. [CrossRef]
- 127. Zhang, M.; Liang, Y.; Yu, D.; Du, B.; Cheng, W.; Li, L.; Yu, Z.; Luo, S.; Zhang, Y.; Wang, H.; et al. A systematic review of Vaccine Breakthrough Infections by SARS-CoV-2 Delta Variant. *Int. J. Biol. Sci.* **2022**, *18*, 889–900. [CrossRef]
- 128. Chia, P.Y.; Ong, S.W.X.; Chiew, C.J.; Ang, L.W.; Chavatte, J.M.; Mak, T.M.; Cui, L.; Kalimuddin, S.; Chia, W.N.; Tan, C.W.; et al. Virological and serological kinetics of SARS-CoV-2 Delta variant vaccine breakthrough infections: A multicentre cohort study. *Clin. Microbiol. Infect.* 2022, 28, 612.e611–612.e617. [CrossRef] [PubMed]
- 129. Singanayagam, A.; Hakki, S.; Dunning, J.; Madon, K.J.; Crone, M.A.; Koycheva, A.; Derqui-Fernandez, N.; Barnett, J.L.; Whitfield, M.G.; Varro, R.; et al. Community transmission and viral load kinetics of the SARS-CoV-2 delta (B.1.617.2) variant in vaccinated and unvaccinated individuals in the UK: A prospective, longitudinal, cohort study. *Lancet. Infect. Dis.* 2022, 22, 183–195. [CrossRef]
- 130. Acharya, C.B.; Schrom, J.; Mitchell, A.M.; Coil, D.A.; Marquez, C.; Rojas, S.; Wang, C.Y.; Liu, J.; Pilarowski, G.; Solis, L. No significant difference in viral load between vaccinated and unvaccinated, asymptomatic and symptomatic groups when infected with SARS-CoV-2 delta variant. *MedRxiv* 2021, 2021-09. [CrossRef]
- Riemersma, K.K.; Haddock, L.A., 3rd; Wilson, N.A.; Minor, N.; Eickhoff, J.; Grogan, B.E.; Kita-Yarbro, A.; Halfmann, P.J.; Segaloff, H.E.; Kocharian, A.; et al. Shedding of infectious SARS-CoV-2 despite vaccination. *PLoS Pathog.* 2022, 18, e1010876. [CrossRef] [PubMed]
- 132. Siddle, K.J.; Krasilnikova, L.A.; Moreno, G.K.; Schaffner, S.F.; Vostok, J.; Fitzgerald, N.A.; Lemieux, J.E.; Barkas, N.; Loreth, C.; Specht, I.; et al. Transmission from vaccinated individuals in a large SARS-CoV-2 Delta variant outbreak. *Cell* 2022, 185, 485–492.e410. [CrossRef]
- 133. Dimeglio, C.; Migueres, M.; Mansuy, J.M.; Saivin, S.; Miedougé, M.; Chapuy-Regaud, S.; Izopet, J. Antibody titers and breakthrough infections with Omicron SARS-CoV-2. J. Infect. 2022, 84, e13–e15. [CrossRef] [PubMed]
- 134. Tenforde, M.W.; Link-Gelles, R.; Patel, M.M. Long-term Protection Associated with COVID-19 Vaccination and Prior Infection. *JAMA* 2022, *328*, 1402–1404. [CrossRef]
- Schaeffer, B.; Taylor, B.; Bushman, M.; Hanage, W.P. The devil in the details: Herd immunity and pandemic response. *Cell Host Microbe* 2021, 29, 1048–1051. [CrossRef] [PubMed]
- 136. Rijkers, G.T.; Weterings, N.; Obregon-Henao, A.; Lepolder, M.; Dutt, T.S.; van Overveld, F.J.; Henao-Tamayo, M. Antigen Presentation of mRNA-Based and Virus-Vectored SARS-CoV-2 Vaccines. *Vaccines* 2021, 9, 848. [CrossRef]
- 137. Moss, P. The T cell immune response against SARS-CoV-2. Nat. Immunol. 2022, 23, 186–193. [CrossRef]
- 138. Chen, S.; Guan, F.; Candotti, F.; Benlagha, K.; Camara, N.O.S.; Herrada, A.A.; James, L.K.; Lei, J.; Miller, H.; Kubo, M.; et al. The role of B cells in COVID-19 infection and vaccination. *Front. Immunol.* **2022**, *13*, 988536. [CrossRef]
- 139. Liu, Y.; Zeng, Q.; Deng, C.; Li, M.; Li, L.; Liu, D.; Liu, M.; Ruan, X.; Mei, J.; Mo, R.; et al. Robust induction of B cell and T cell responses by a third dose of inactivated SARS-CoV-2 vaccine. *Cell Discov.* **2022**, *8*, 10. [CrossRef]
- 140. Amanatidou, E.; Gkiouliava, A.; Pella, E.; Serafidi, M.; Tsilingiris, D.; Vallianou, N.G.; Karampela, I.; Dalamaga, M. Breakthrough infections after COVID-19 vaccination: Insights, perspectives and challenges. *Metabol. Open* **2022**, *14*, 100180. [CrossRef]
- Al-Aly, Z.; Bowe, B.; Xie, Y. Long COVID after breakthrough SARS-CoV-2 infection. *Nat. Med.* 2022, 28, 1461–1467. [CrossRef]
 [PubMed]
- 142. Mahdy, M.A.A.; Younis, W.; Ewaida, Z. An Overview of SARS-CoV-2 and Animal Infection. Front. Vet. Sci. 2020, 7. [CrossRef]
- 143. Zhou, P.; Yang, X.L.; Wang, X.G.; Hu, B.; Zhang, L.; Zhang, W.; Si, H.R.; Zhu, Y.; Li, B.; Huang, C.L.; et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* **2020**, *579*, 270–273. [CrossRef] [PubMed]
- 144. Boni, M.F.; Lemey, P.; Jiang, X.; Lam, T.T.; Perry, B.W.; Castoe, T.A.; Rambaut, A.; Robertson, D.L. Evolutionary origins of the SARS-CoV-2 sarbecovirus lineage responsible for the COVID-19 pandemic. *Nat. Microbiol.* 2020, *5*, 1408–1417. [CrossRef] [PubMed]
- 145. Zhang, T.; Wu, Q.; Zhang, Z. Probable Pangolin Origin of SARS-CoV-2 Associated with the COVID-19 Outbreak. *Curr. Biol.* **2020**, *30*, 1346–1351.e1342. [CrossRef]
- 146. Haider, N.; Rothman-Ostrow, P.; Osman, A.Y.; Arruda, L.B.; Macfarlane-Berry, L.; Elton, L.; Thomason, M.J.; Yeboah-Manu, D.; Ansumana, R.; Kapata, N.; et al. COVID-19-Zoonosis or Emerging Infectious Disease? *Front. Public. Health* 2020, *8*, 596944. [CrossRef] [PubMed]

- 147. Prince, T.; Smith, S.L.; Radford, A.D.; Solomon, T.; Hughes, G.L.; Patterson, E.I. SARS-CoV-2 Infections in Animals: Reservoirs for Reverse Zoonosis and Models for Study. *Viruses* **2021**, *13*, 494. [CrossRef]
- 148. Fan, Q.; Shi, J.; Yang, Y.; Tang, G.; Jiang, M.; Li, J.; Tang, J.; Li, L.; Wen, X.; Zhang, L.; et al. Clinical characteristics and immune profile alterations in vaccinated individuals with breakthrough Delta SARS-CoV-2 infections. *Nat. Commun.* 2022, 13, 3979. [CrossRef]
- 149. Joseph, J. Trained Immunity as a Prospective Tool against Emerging Respiratory Pathogens. *Vaccines* **2022**, *10*, 1932.
- 150. Brueggeman, J.M.; Zhao, J.; Schank, M.; Yao, Z.Q.; Moorman, J.P. Trained Immunity: An Overview and the Impact on COVID-19. *Front. Immunol.* **2022**, *13*, 837524. [CrossRef] [PubMed]
- 151. Zachary, A.A.; Eng, H.S. Desensitization: Achieving immune detente. Tissue Antigens 2011, 77, 3–8. [CrossRef] [PubMed]
- 152. Yu, W.; Freeland, D.M.H.; Nadeau, K.C. Food allergy: Immune mechanisms, diagnosis and immunotherapy. *Nat. Rev. Immunol.* **2016**, *16*, 751–765. [CrossRef] [PubMed]
- Akdis, C.A.; Akdis, M. Mechanisms of allergen-specific immunotherapy and immune tolerance to allergens. *World Allergy Organ. J.* 2015, 8, 17. [CrossRef]
- Murphy, D.M.; Cox, D.J.; Connolly, S.A.; Breen, E.P.; Brugman, A.A.; Phelan, J.J.; Keane, J.; Basdeo, S.A. Trained immunity is induced in humans after immunization with an adenoviral vector COVID-19 vaccine. *J. Clin. Investig.* 2023, 133, e162581. [CrossRef]
- 155. Messina, M.R.; Crisciotti, C.; Pellegrini, L.; Nappi, E.; Racca, F.; Costanzo, G.; Del Moro, L.; Ferri, S.; Puggioni, F.; Canonica, G.W.; et al. Desensitization Protocols for Anti-SARS-CoV-2 Vaccines in Patients with High Risk of Allergic Reactions. *Vaccines* **2023**, *11*, 910. [CrossRef]
- 156. AlMuhizi, F.; Ton-Leclerc, S.; Fein, M.; Tsoukas, C.; Garvey, L.H.; Lee, D.; Ben-Shoshan, M.; Isabwe, G.A.C.; Copaescu, A.M. Successful Desensitization to mRNA COVID-19 Vaccine in a Case Series of Patients with a History of Anaphylaxis to the First Vaccine Dose. *Front. Allergy* 2022, *3*, 825164. [CrossRef]
- 157. Romantowski, J.; Kruszewski, J.; Solarski, O.; Bant, A.; Chciałowski, A.; Pietrzyk, I.; Sańpruch, P.; Górska, A.; Chełmińska, M.; Knurowska, A.; et al. Protocol of safe vaccination against COVID-19 in patients with high risk of allergic reactions. *Clin. Transl. Allergy* 2022, 12, e12152. [CrossRef]
- 158. Menni, C.; May, A.; Polidori, L.; Louca, P.; Wolf, J.; Capdevila, J.; Hu, C.; Ourselin, S.; Steves, C.J.; Valdes, A.M.; et al. COVID-19 vaccine waning and effectiveness and side-effects of boosters: A prospective community study from the ZOE COVID Study. *Lancet Infect. Dis.* **2022**, *22*, 1002–1010. [CrossRef]
- 159. Nguyen, A.V.; Soulika, A.M. The Dynamics of the Skin's Immune System. Int. J. Mol. Sci 2019, 20, 1811. [CrossRef]
- 160. Prausnitz, M.R.; Mikszta, J.A.; Cormier, M.; Andrianov, A.K. Microneedle-based vaccines. *Curr. Top Microbiol. Immunol.* **2009**, 333, 369–393. [CrossRef]
- Donnelly, R.F.; Singh, T.R.R.; Morrow, D.I.J.; Woolfson, A.D. Microneedle-mediated intradermal delivery. In *Microneedle-Mediated Transdermal and Intradermal Drug Delivery*, 2nd ed.; Donnelly, R.F., Singh, T.R.R., Morrow, D.I.J., Woolfson, A.D., Eds.; Wiley-Blackwell: Hoboken, NJ, USA, 2012; pp. 113–151.
- 162. Al-Zahrani, S.; Zaric, M.; McCrudden, C.; Scott, C.; Kissenpfennig, A.; Donnelly, R.F. Microneedle-mediated vaccine delivery: Harnessing cutaneous immunobiology to improve efficacy. *Expert Opin. Drug Deliv.* **2012**, *9*, 541–550. [CrossRef] [PubMed]
- Alarcon, J.B.; Hartley, A.W.; Harvey, N.G.; Mikszta, J.A. Preclinical evaluation of microneedle technology for intradermal delivery of influenza vaccines. *Clin. Vaccine Immunol.* 2007, 14, 375–381. [CrossRef] [PubMed]
- 164. Arnou, R.; Eavis, P.; Pardo, J.R.; Ambrozaitis, A.; Kazek, M.P.; Weber, F. Immunogenicity, large scale safety and lot consistency of an intradermal influenza vaccine in adults aged 18-60 years: Randomized, controlled, phase III trial. *Hum. Vaccine* 2010, 6, 346–354. [CrossRef] [PubMed]
- Lambert, P.H.; Laurent, P.E. Intradermal vaccine delivery: Will new delivery systems transform vaccine administration? *Vaccine* 2008, 26, 3197–3208. [CrossRef]
- 166. Mikszta, J.A.; Dekker, J.P., 3rd; Harvey, N.G.; Dean, C.H.; Brittingham, J.M.; Huang, J.; Sullivan, V.J.; Dyas, B.; Roy, C.J.; Ulrich, R.G. Microneedle-based intradermal delivery of the anthrax recombinant protective antigen vaccine. *Infect. Immun.* 2006, 74, 6806–6810. [CrossRef]
- 167. Dean, C.H.; Alarcon, J.B.; Waterston, A.M.; Draper, K.; Early, R.; Guirakhoo, F.; Monath, T.P.; Mikszta, J.A. Cutaneous delivery of a live, attenuated chimeric flavivirus vaccine against Japanese encephalitis (ChimeriVax)-JE) in non-human primates. *Hum. Vaccine* **2005**, *1*, 106–111. [CrossRef]
- 168. Hooper, J.W.; Golden, J.W.; Ferro, A.M.; King, A.D. Smallpox DNA vaccine delivered by novel skin electroporation device protects mice against intranasal poxvirus challenge. *Vaccine* 2007, 25, 1814–1823. [CrossRef]
- Hassan, J.; Haigh, C.; Ahmed, T.; Uddin, M.J.; Das, D.B. Potential of Microneedle Systems for COVID-19 Vaccination: Current Trends and Challenges. *Pharmaceutics* 2022, 14, 1066. [CrossRef] [PubMed]

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