

Supplementary Data

Transgenic microalgae expressing double-stranded RNA as potential feed supplements for controlling white spot syndrome in shrimp aquaculture

Patai Charoonnart^{1,2*}, Henry Nicolas Taunt³, Luyao Yang³, Conner Webb⁴, Colin Robinson⁴,
Vanvimon Saksmerprom^{1,2} & Saul Purton³

¹Center of Excellence for Shrimp Molecular Biology and Biotechnology (Centex Shrimp), Faculty of Science, Mahidol University, Bangkok 10400, Thailand

²National Center for Genetic Engineering and Biotechnology (BIOTEC), National Science and Technology Development Agency, Pathum Thani 12120, Thailand

³Department of Structural and Molecular Biology, University College London, London WC1E 6BT, UK

⁴Centre for Molecular Processing, School of Biosciences, University of Kent, Canterbury CT2 7NJ, UK

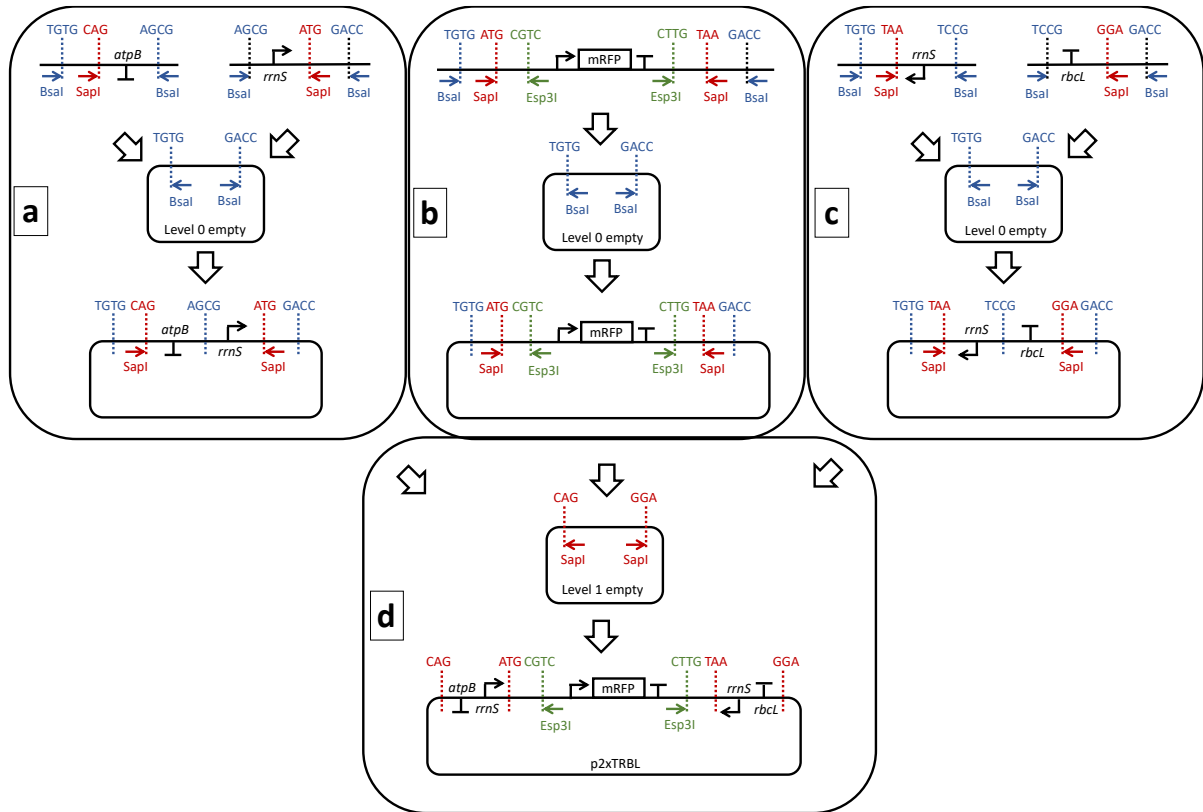


Figure S1 Golden Gate assembly of p2xTRBL. *C. reinhardtii* expression elements were amplified by PCR, with the addition of 5' extensions to allow *BsaI* cloning into the Level 0 empty vector (a, c). The mRFP cassette was synthesized *de novo*, also with the addition of Golden Gate cloning sequences (b). The three Level 0 cassettes were cloned into a Level 1 empty vector using *SapI* to generate p2xTRBL (d).

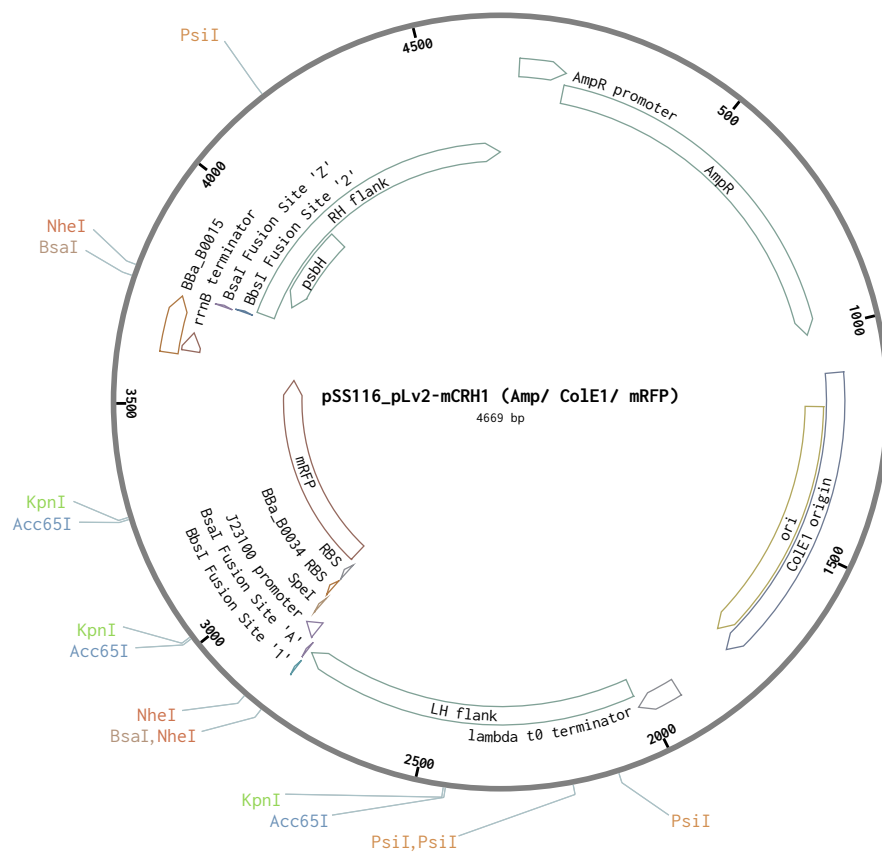


Figure S2 Map of plasmid pSS116, an integration vector for targeting transgenes into the *C. reinhardtii* chloroplast genome between *psbH* and *trnE2*.

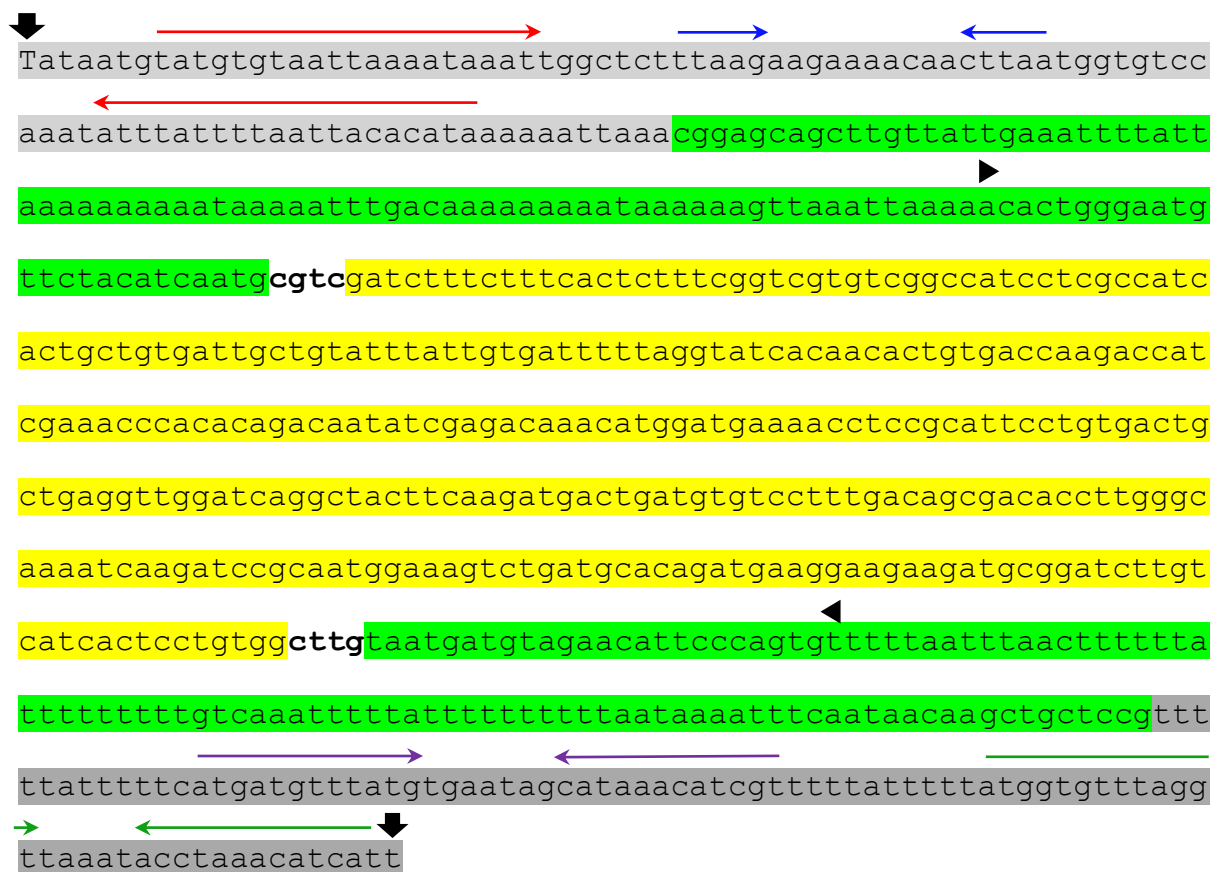


Figure S3 Sequence of the *VP28* transcription cassette. The *VP28* sequence is highlighted in **yellow**. *Esp31* fusion sites are in **bold**. The *rrnS* promoter elements are highlighted in **green** with the transcriptional start site indicated by a black triangle. The *atpB* terminator is highlighted in **light grey** and the *rbcL* terminator in **dark grey**. Inverted repeat regions are indicated by horizontal coloured arrows and the transcript termination sites by vertical black arrows.

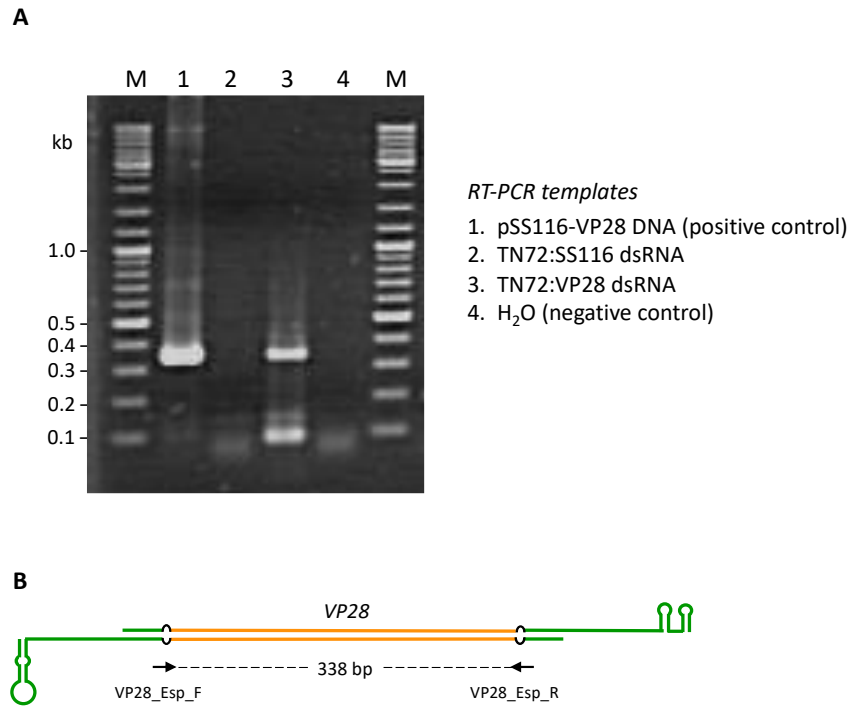


Figure S4 RT-PCR analysis of dsRNA from transformant TN72:VP28. **A.** Lane 1: PCR amplification of VP28 DNA from pSS116-VP28 plasmid. Lane 2: RT-PCR using dsRNA fraction from strain TN72:SS116. Lane 3: RT-PCR using dsRNA fraction from strain TN72:VP28. 4 RT-PCR using ddH₂O as template. **B.** Illustration of primers used for RT-PCR.

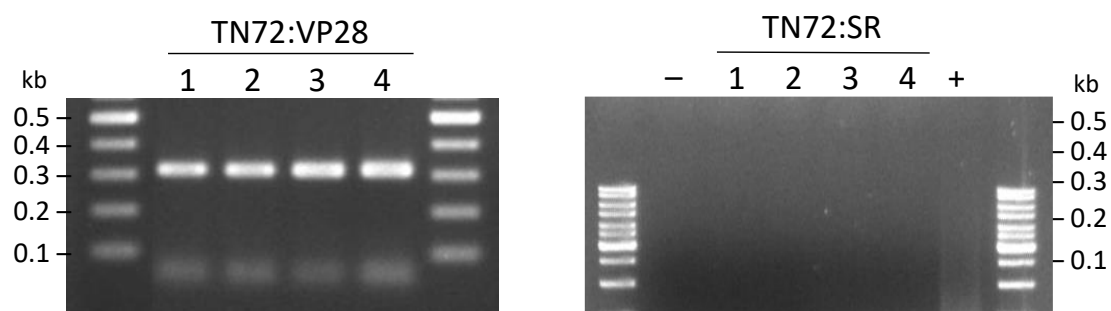


Figure S5 Detection of dsRNA from freeze-dried *C. reinhardtii*. Four replicate samples each representing 0.1 g of biomass from either TN72:VP28 (left panel) or TN72:SR (right panel) were used for dsRNA extraction and RT-PCR using specific primers VP28_Esp_F and VP28_Esp_R (See Figure S4). Controls are H₂O (–) or purified dsRNA (+) as template.

Table S1 Details of primers used

Primer	Sequence (5'–3')	Function	Reference
VP28_Esp_F	ATACG <u>CGTCTCT</u> CGTCGATCTTTCTTTCACTCTTTTCG	Generation of VP28 fragment for p2XTRBL cloning (<i>Esp</i> 31 sites underlined). RT-PCR analysis of dsRNA from <i>E. coli</i> and <i>C. reinhardtii</i> (PCR product of 338 bp)	This work
VP28_Esp_R	ATACG <u>CGTCTCT</u> CAAGCCACAGGAGTGATGACAA		
rbcL_F	GTCACCACCAGACATACGAAG	Internal control for chloroplast DNA extraction. (PCR product of 264 bp)	[18]
rbcL_R	GGTCACTACTTAAACGCTAC		
TN72_F	GTCATTGCGAAAATACTGGTGC	TN72 plastome-specific primers for assessing homoplasmic status. (PCR product of 0.88 kb)	[19]
TN72_R	CGGATGTAACTCAATCGGTAG		
WSSV447_F	ATGAGAATGAACTCCAACCTTAA	WSSV load assay.	[26]
WSSV447_R	CAGAGCCTAGTCTATCAATCAT		