

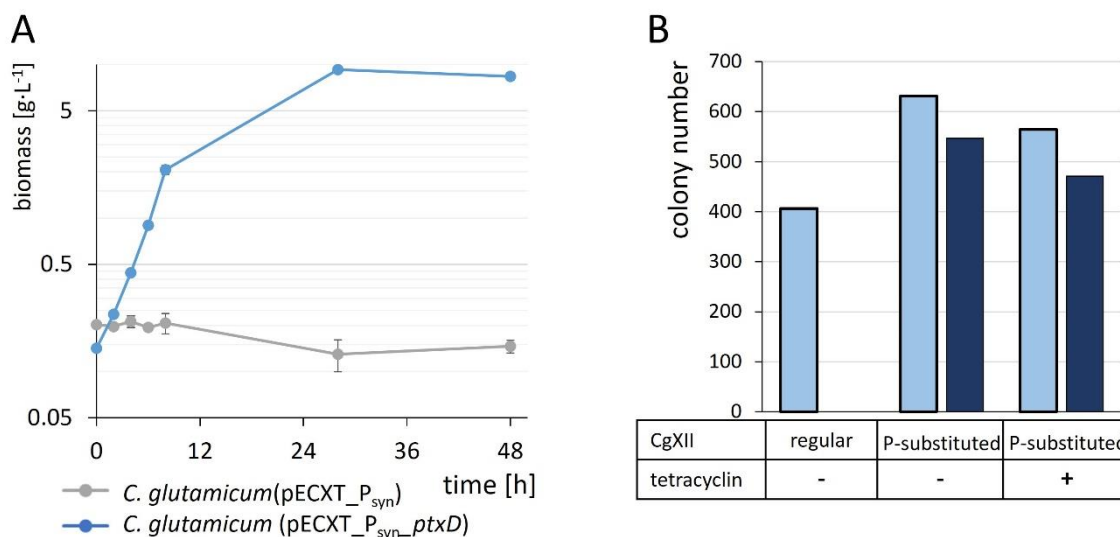
Supplementary material to

# Metabolic engineering of *Corynebacterium glutamicum* for sustainable production of the aromatic dicarboxylic acid dipicolinic acid

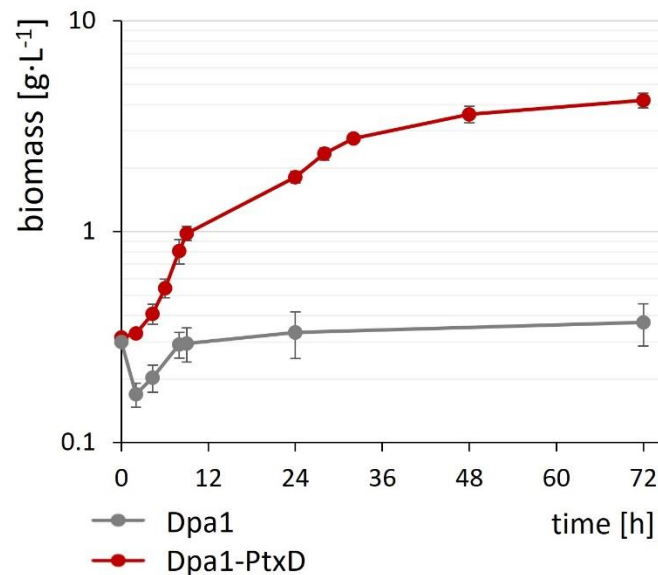
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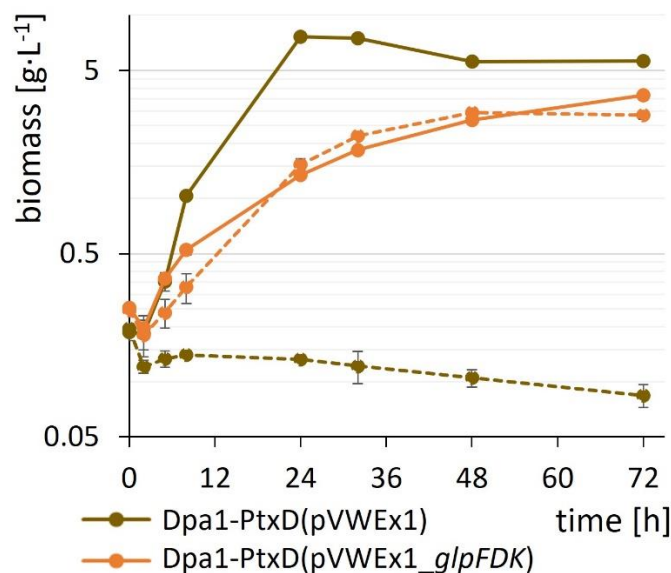
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**Figure S1.** Growth curves of *C. glutamicum*(pECXT\_P<sub>syn</sub>-ptxD) and control strain *C. glutamicum*(pECXT\_P<sub>syn</sub>). The strain expressing phosphite dehydrogenase from *Pseudomonas stutzeri* WM88 (blue) and the empty vector control strain (grey) were cultivated in P-substituted CgXII minimal medium, containing 40 g·L<sup>-1</sup> glucose and 13 mM di-sodium phosphite as sole phosphorous source. Values are given as means with standard deviations from technical triplicates (A). Colony numbers of *C. glutamicum*(pECXT\_P<sub>syn</sub>-ptxD) on LB (light blue) or LB-tetracycline agar plates, after cultivation for 17 serial dilutions in regular (phosphate as phosphorous source, no tetracycline) or in P-substituted CgXII minimal medium (13 mM di-sodium phosphite as phosphorous source) with or without tetracycline. After cultivation in P-substituted CgXII minimal medium with or without the addition of tetracycline, colony numbers were similar, whereas the plasmid was lost after cultivation in regular CgXII without tetracycline, demonstrating the aptitude of *ptxD* expression as selectable trait under non-sterile growth conditions (B).



**Figure S2.** Growth curves of *C. glutamicum* Dpa1-PtxD and control strain Dpa1. The DPA producing strain expressing phosphite dehydrogenase from *Pseudomonas stutzeri* WM88 (red) and the control strain (grey) were cultivated in P-substituted CgXII minimal medium, containing 40 g·L<sup>-1</sup> glucose and 13 mM di-sodium phosphite as sole phosphorous source. Values are given as means with standard deviations from technical triplicates.



**Figure S3.** Growth curves of *C. glutamicum* Dpa1-PtxD(pVWEx1\_glpFDK) control strain Dpa1-PtxD(pVWEx1). The DPA producing strain expressing phosphite dehydrogenase from *Pseudomonas stutzeri* WM88 (orange) and the control strain (brown) were cultivated in P-substituted CgXII minimal medium, containing 40 g·L<sup>-1</sup> glucose (solid lines) or 40 g·L<sup>-1</sup> glycerol (stacked lines) and 13 mM di-sodium phosphite as sole phosphorous source. Values are given as means with standard deviations from technical triplicates.

**Table S1.** Oligonucleotides used in this work.

Oligonucleotide	Sequence (5'-3')	Function
<i>ptxD</i> -fw-XbaI	GATTATCGAACGGTTTCATTCAGCATAGCTGAAAG GAGGCCCTTCAGATGCTGC	Amplification of <i>ptxD</i>
<i>ptxD</i> -rv-XbaI	GCTTGCATGCCTGCAGGTCGACTTTAATCCGCAGCT GGATTCGCC	Amplification of <i>ptxD</i>
<i>ptxD</i> -s1	CTTCAGGGCACAGCCGATAAC	Verification of <i>ptxD</i> sequence
<i>ptxD</i> -s2	GGAGGGTATCCGCGTTCAG	Verification of <i>ptxD</i> sequence
<i>dpaAB</i> -fw-XbaI	CAAAGGCCGAATCCAGCTGCGGATTAAGCTGAAAG GAGGCCCTTCAGATGCTTAC	Amplification of <i>dpaAB</i> for coexpression with <i>ptxD</i>
<i>dpaAB</i> -rv-XbaI	GCTTGCATGCCTGCAGGTCGACTCTATGCTGAATGA AACCGTTCCG	Amplification of <i>dpaAB</i> for coexpression with <i>ptxD</i>
<i>dpaAB</i> -fw	GGTTCCATGGAATTCGAGCTCGGTACCCGGGGAAA GGAGGCCCTTCAGATGCTTACTGGCATCAGGATCG	Amplification of <i>dpaAB</i>
<i>dpaAB</i> -rv	GCTTGCATGCCTGCAGGTCGACTCTAGAGCTATGCT GAATGAAACCGTTCCG	Amplification of <i>dpaAB</i>
<i>dpaAB</i> -rv- <i>dapA</i>	CTTAGCTGTAAACCTGTGCTCATCTGAAGGGCCTC CTTTCCTATGCTGAATGAAACCGTTCCG	Amplification of <i>dpaAB</i> for coexpression with <i>dapA</i>
<i>dpaAB</i> - <i>dapA</i> -fw	CGAACGGTTTCATTACAGCATAGGAAAGGAGGCCCT TCAGATGAGCACAGGTTTAACAGCTAAG	Amplification of <i>dapA</i> for coexpression with <i>dpaAB</i>
<i>dapA</i> -s	CAGGGACTCGGTTCCAACG	Verification of <i>dapA</i> sequence
<i>dapA</i> -rv	GCTTGCATGCCTGCAGGTCGACTCTAGAGTTATAGA ACTCCAGCTTTTTTCATGTCTTCTC	Verification of <i>dapA</i> sequence
pECXT_P <sub>syn</sub> -fw	TCAGTGAGCGAGGAAGC	Verification of pECXT_P <sub>syn</sub> plasmid sequences and transformants
pECXT99A_rv	TACTGCCGCCAGGCAAATTC	Verification of pECXT_P <sub>syn</sub> plasmid sequences and transformants
<i>amyA</i> -fw	GAGCTCGGTACCCGGGGATCTCAGCCGCGCCAGGT GTCGT	Amplification of <i>amyA</i>
<i>amyA</i> -rv	GAGCTCGGTACCCGGGGATCTCAGCCGCGCCAGGT GTCGT	Amplification of <i>amyA</i>
<i>dapB</i> -sg561-fw	GCCAAGCTTGCATGCCTGCAGCCCATCACAGCGTT AGGAG	Generation of sgRNA for <i>dapB</i> 561 knockdown
<i>dapB</i> -sg561-rv	GCTATTTCTAGCTCTAAAACCTCCTAACGCTGTGAT GGGC	Generation of sgRNA for <i>dapB</i> 561 knockdown
<i>dapB</i> -sg14-fw	GCCAAGCTTGCATGCCTGCAAAATGCTCAAGTCCTA CGACT	Generation of sgRNA for <i>dapB</i> 14 knockdown
<i>dapB</i> -sg14-rv	GCTATTTCTAGCTCTAAAACAGTCGTAGGACTTGAG CATT	Generation of sgRNA for <i>dapB</i> 14 knockdown
dCas9_s-fw	CCGCTTCTGCGTTCTGATTTAATCT	Verification of pS_dCas9 plasmid sequences and transformants
dCas9_s-rv	GTATGGCTGTGCAGGTCGTAAA	Verification of pS_dCas9 plasmid sequences and transformants

**Table S2.** DNA sequences used in this work.

DNA sequence of the used version of the *dpaAB* gene from *Paenibacillus sonchi* genomovar *riograndensis* SBR5 for construction of pECXT\_P<sub>syn</sub>-*dpaAB*.

ATGCTTACTGGCATCAGGATCGTGTTCTGGGCGGGGACGCGAGACAGATTGAAGTGATTGCGAAATGTGTGGAAATGGATG  
CTACGGTAAGCGCTGCCGGGTTGACAAGTGGGATGCCCCAAGCCCGGGGGTAAGCCTGGAACAAATGTCGGCAGAGCTGCT  
GAGCCGCGCGGATGTGCTGGTGTGCCAACGGTGGGGTGTGATGATGAAGGCAATATCAGTGCCCTCTTTCAACGGAGCGC  
CTGCAGCTGCTGGAGGAACATATCGCCGCGCTTCCGCCAAGCTGTATGGTCTATACCGGCATGGCCAAAAGCTACTTGCGCGG  
CCTGTGCGACAAATATCACTGAAGCTGGTGGAGCTGCTGAGCAGGGACGATGTGGCGATCTACAACTCCATCCCTACAGCAG  
AGGGAGCATTGGTTATGGCCATTGAGAATACGGATTTACAATCCATGGCTCTACCTCAATGGTTCTGGGCATGGGCAGAACAG  
GGTTTACCATGGCCAGAGTGTGTCAGGGACTCGGTTCCAACGTGAAGGTGGGAGTAAGAAAACAGGAGCATTACGCACGGGC  
CGAGGAGATGGGCTGGAAGCCCTTTATGACTGGAGAGCTGGTGGCGCATGCGCCGGAGGCGGATCTTATTTTCAACACCATCC  
CTAGTATGATTATCACCGCACAAAGTGTATCGCGTCTTCCAGACACTGTGTAATTATCGATCTGGCTTCCGCCCCGGGCGGGTG  
TGATTTCCGCTATGCGGAGAAACGCGGGATCAAGGCCATGTGGCACCGGGGACTGCCCGAGTCGTTGCTCCAAAAGCGCCG  
GAATTATTATGGCGGGCGCGCTGGTACAGTCGATATCGGACGAGACTTTTAAACAGGGGGGACGTTTAAATGGATTGGCATGGA  
AAAACAGTAGGTTATGCGGTGACCGGCTCACACTGCACGTTAGCCGAGGTTATGCCGAGATTACGCGGTTTATGGAGGGCGG  
AGCCAATGTGGTGCCGATTGTGTCGTCATCCGTGCTGAATACGACACCCGCTTTGGCACATCGGAAAATTGGCTAAAACAGTT  
GAAAGAAATAACAGGGAATGATATCATTCTACAATTGTTGAAGCGGAACCGCTGGGTCCTTCCAAGCTGCTGGATGTGCTGA  
CTATTGCACCCTGCACGGGGAATACGACAAGTAAATTGGCTAACGCCATGACCGACAGCCCCGTGCTGATGGCCGCCAAAGCG  
CAGCTGCGCAACAGCCGTCCGCTGGTGTGCAATCTCCACCAATGATGGACTTGGCCTGAATGCGGCGAATATCGCGAAGCT  
CCTGGTTGCGAAGAACATTTATTTTGTTCGTTCCGCCAGGATAATCCGCAGGGCAAGCCGAATTCGCTTGTGGCCAGATGGA  
CCTCATTCCCGAAGCCTGCTTTCGGGCTTTCAGGGCCAACAGCTGCAGCCGATGATTATCGAACGGTTTCATTAG

DNA sequence of codon-optimized version of the *ptxD* gene from *Pseudomonas stutzeri* WM88 for construction of pECXT\_P<sub>syn</sub>-*ptxD*.

ATGCTGCCAAAGCTCGTTATCACCCACCGGTTACGAAGAAATTTTGCAGCTGCTCGCACCACACTGCGAACTGATCACCAAC  
CAAACCGATTCTACCCTCACCCGTGAAGAAATTTTGCGCCGTTGTGCGGATGCACAGGCCATGATGGCTTTCATGCCAGATCGT  
GTGGATGCAGATTTCTGCAAGCCTGCCAGAACTCCGCGTTATCGGCTGTGCCCTGAAGGGTTTCGATAATTTGATGTTGAT  
GCTTGCACCGCGCGTGGCGTGTGGCTCACCTTCGTTCCAGATTTGCTGACCGTTCCAACCGCAGAACTGGCCATTGGTTTGGCT  
GTTGGCTTGGGTCGCCACCTGCGTGCAGCCGATGCGTTCGTTGCTCTGGCAAGTTCGTTGGTGGCAGCCACGCTTCTACGGC  
ACCGGTTTGGATAACGCAACCGTGGGCTTCTTGGGCATGGGTGCCATCGGTTTGGCTATGGCGGATCGCTGCAGGGCTGGG  
GTGCAACCCTGCAATACCACGAAGCAAAGGCCCTCGATACCCAGACCGAACAACGCCTGGGCCTCCGTCAAGTTGCATGCTCT  
GAATTGTTGCTCCTCTGATTTCACTTCTTGGCTTGGCACTGAACGCGGATACCCTCCACTTGGTTAATGCTGAATTGCTCGC  
ACTCGTTCGTCCAGGTGCTTTGCTGGTGAACCATGTCGCGGTTCCGTGGTTGATGAAGCTGCGGTGTTGGCAGCCCTGGAAC  
GTGGTCAGCTGGGCGGTTACGCTGCGGATGTTTTGAAATGGAAGATTGGGCTCGCGCGGATCGTCCACAGCAAATCGATCCA  
GCTCTCTTGGCGCACCCAAATACCCTTTCACCCACACATTGGCTCCGAGTTTCGCGCCGTGCGTTTGGAAATCGAACGCTGTG  
CAGCCGAGAACATTCTCAAGCATTGGCCGGTGAACGTCCAATCAACGCTGTGAATCGCCTCCAAAGGCGAATCCAGCTGCG  
GATTAA