

## Review

# Of Mycelium and Men: Inherent Human Susceptibility to Fungal Diseases

Donald C. Vinh 

- <sup>1</sup> Division of Infectious Diseases (Department of Medicine, McGill University Health Centre), Montreal, QC H4A 3J1, Canada; donald.vinh@mcgill.ca
- <sup>2</sup> Division of Medical Microbiology, Division of Molecular Genetics-Immunology (Department of OptiLab, McGill University Health Centre), Montreal, QC H4A 3J1, Canada
- <sup>3</sup> Centre of Excellence for Genetic Research in Infection and Immunity, Research Institute-McGill University Health Centre, Montreal, QC H4A 3J1, Canada

**Abstract:** In medical mycology, the main context of disease is iatrogenic-based disease. However, historically, and occasionally, even today, fungal diseases affect humans with no obvious risk factors, sometimes in a spectacular fashion. The field of “inborn errors of immunity” (IEI) has deduced at least some of these previously enigmatic cases; accordingly, the discovery of single-gene disorders with penetrant clinical effects and their immunologic dissection have provided a framework with which to understand some of the key pathways mediating human susceptibility to mycoses. By extension, they have also enabled the identification of naturally occurring auto-antibodies to cytokines that phenocopy such susceptibility. This review provides a comprehensive update of IEI and autoantibodies that inherently predispose humans to various fungal diseases.

**Keywords:** inborn errors of immunity; primary immunodeficiency; fungi; mycoses; invasive fungal disease; superficial fungal disease



**Citation:** Vinh, D.C. Of Mycelium and Men: Inherent Human Susceptibility to Fungal Diseases. *Pathogens* **2023**, *12*, 456. <https://doi.org/10.3390/pathogens12030456>

Academic Editor: Sudhir Gupta

Received: 28 January 2023

Revised: 9 March 2023

Accepted: 9 March 2023

Published: 14 March 2023



**Copyright:** © 2023 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Humans have lived intertwined with fungi since antiquity. Despite their virtues, fungi have also generated human diseases throughout history (antedating modern concepts of iatrogenically immunocompromised hosts), which were either mediated by toxins or manifesting as infection (mycoses). Invasive fungal disease (IFD) can either be superficial (i.e., invasive disease restricted to the mucosa, skin, and/or skin appendages) or deep-seated (invasive toward otherwise sterile tissue, e.g., visceral organs, fluid (e.g., blood), etc.). While the underlying basis for the host’s susceptibility to such fungal infections once remained obscure, a familial basis was observed in some cases, implying a hereditary component. The field of primary immunodeficiencies, now referred to as inborn errors of immunity (IEI), has enabled the discovery of some of these genetic elements. While fungal pathogenicity is the outcome of a complex interaction between a fungus, a host, and the environment, the field of medical mycology has focused on the former two in the context of iatrogenic immunosuppression, which is understandable because of its predominance in the clinical picture. On the other hand, defining the monogenic basis of human immunity to fungi in a natural state (i.e., in the absence of iatrogenic immunosuppression), which is performed through the field of IEI, can pinpoint critical molecular antifungal pathways. We review these disorders herein.

In the absence of anatomic defects, iatrogenic immunosuppression, or invasive procedures, fungal disease that is persistent, recurrent, and/or severe strongly suggests the presence of an underlying inborn error of immunity. However, the identification of this “fungal disease phenotype” is not a sufficient measure with which to label a person with an inborn error of immunity. In the purest sense, investigations identifying molecular

lesions, which are mechanistically causative of an infection through immunological dysfunction, should be pursued. Dissecting the defective immunobiological lesions, even in singular patients, permits the proposition of immune pathways that may be critical for host antifungal defenses. The addition of each new member of this patient group then adds to or refines such pathways or perhaps even leads to the proposition of alternative ones, eventually leading to the development of pathophysiological constructs of human immunity to various fungi.

## 2. Framework Ascribing Genetic Immunodeficiency to Fungal Susceptibility

The ultimate purpose of defining IEI causally associated with IFDs is to develop a molecular immunological framework of human immunity to mycosis; such information may even shed light on fungal susceptibility in the more complex secondary immunodeficiencies. To achieve this, stringent criteria need to be explicitly considered to attribute penetrant, genetic susceptibility to fungal disease, and such criteria are proposed herein:

- (1) The reported mycosis must have fulfilled standardized, diagnostic criteria of fungal disease, such as those defined by the European Organization for Research and Treatment of Cancer and Mycoses Study Group (EORTC-MSG) [1]. While initially formulated for IFD in iatrogenically immunocompromised patients, this framework provides provide a barometer for robust diagnosis. These criteria are intended to provide a strong deterrent against the misattribution of susceptibility to fungal disease based on colonization (rather than invasion), sample contamination, or non-specificity (e.g., based on certain serologic markers).
- (2) The mycosis must have occurred prior to any medical intervention. Some IEI require exogenous immunosuppression for autoinflammatory or autoimmune features or transplantation for failing organ function or hematopoietic reasons. Some patients with IEI may require indwelling catheters (e.g., for intravenous access), which may become contaminated and then serve as a portal of systemic entry. In addition, some patients with IEI may require surgical intervention (which may or may not be related to their IEI), but which may become a nidus for IFD (e.g., intestinal leak/perforation). These states, in themselves, can be associated with an elevated risk of IFD. A fungal infection that develops only after the use of these treatments or in transplanted states cannot be definitively attributed to the underlying IEI exclusively, although such iatrogenic cases are conducive to the generation of hypotheses (especially if observed repeatedly and/or supported by mechanistic investigations).
- (3) The mycosis must have occurred in an inborn error of immunity that was genetically identified and immunobiologically validated. This criterion is in place to avoid ascribing a fungal infection to a genetic variant of unproven significance.

The certainty of the association between a gene defect and fungal susceptibility is considered “strong” when a sufficient number of cases have been reported. Consistency in the clinical observation of susceptibility implies a persistent, inherent immunological basis. Given the rarity of IEI, and the potential ubiquitous or restricted geographic distribution of the implicated fungi with resulting variability in terms of individual exposure, a threshold of three unrelated cases is proposed and used herein. Alternatively, especially in cases where the associated inborn error of immunity or mycosis is extremely rare (e.g., with few cases or a single case reported), the demonstrable immunologic defect must be congruent with the immuno-pathophysiology for that specific mycosis previously shown in other IEI. Thus, a strong association is established when three or more unrelated cases have been reported or, in ultra-rare cases, there is pathophysiologic homogeneity for that mycosis in other IEI. Below this threshold, cases are considered “sporadic”, implying the need for additional reported cases or immuno-pathologic mechanisms to buttress the certainty of the association.

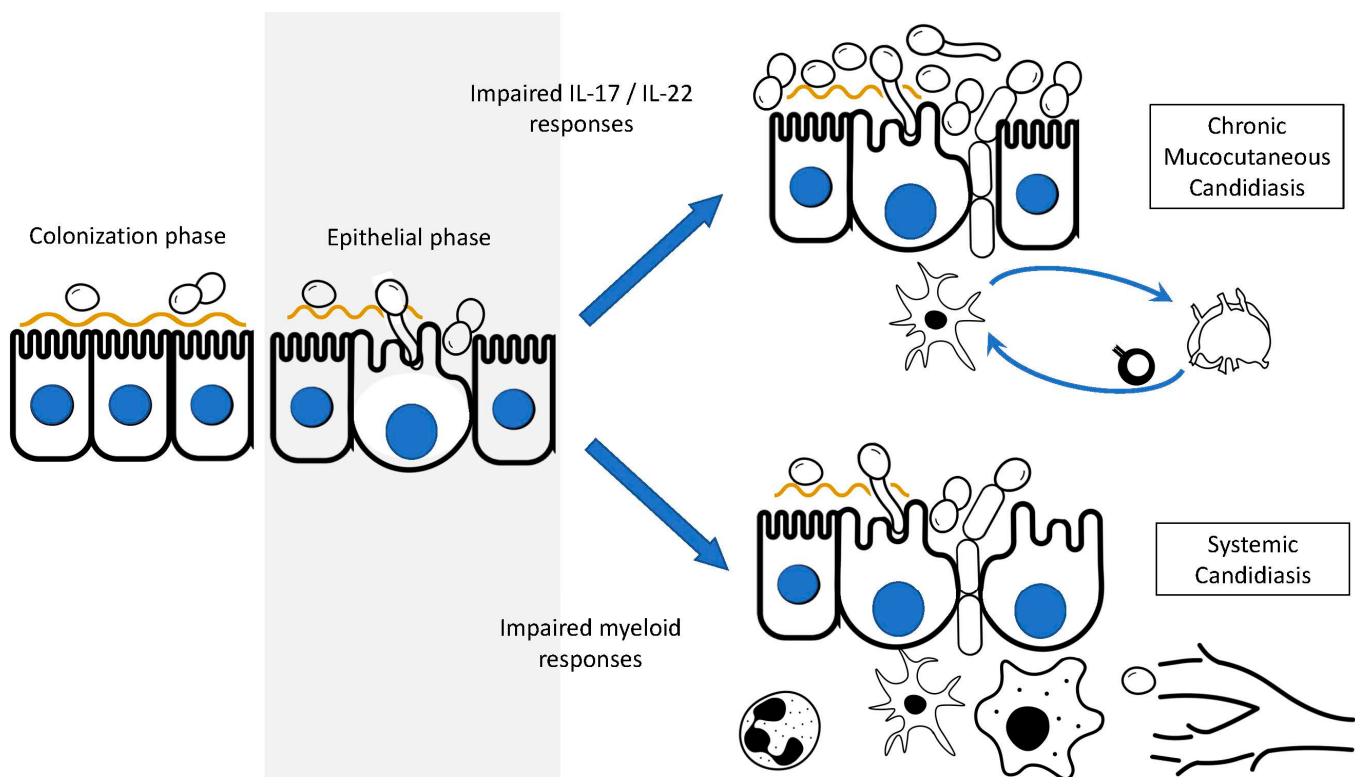
Collectively, these criteria aim to bolster the association of distinct mycoses with different IEI to more confidently identify underlying pathophysiological frameworks.

### 3. Candidiasis

Among the >200 defined *Candida* species, only a select few are specifically pathogenic to humans, with the foremost among these being *C. albicans* [2]. In a healthy state, *C. albicans* can be found as a commensal organism of the human microbiome, which, within the first year of life, colonizes the skin and gastrointestinal and/or genitourinary mucosae in as much as 70% of well individuals [3,4]. In this state, *C. albicans* likely reside in or above the dense layer of dead keratinocytes (stratum corneum) on the skin or in the mucus layer of mucosal surfaces. The acquisition of non-*albicans* species is less well characterized. Mucosal candidal colonization can be intermittent, with periods of loss of colonization, and can be influenced by environmental factors (e.g., tobacco), one's physiological status (e.g., pre-menopausal), medical conditions (e.g., diabetes, HIV status, etc.), or medications (e.g., antibiotics, steroids, etc.) [5,6].

#### 3.1. Chronic Mucocutaneous Candidiasis (CMC)

At these mucocutaneous barriers, *Candida* spp. can transition from colonizers to pathogens, leading to “superficial” disease, where invasion is restricted to the epithelial surfaces (Figure 1). Mucosal candidiasis manifests as one of several forms of oropharyngeal candidiasis (OPC, e.g., thrush/pseudomembranous OPC, erythematous lesions/atrophic OPC, and angular cheilitis), esophagitis, and/or vulvo-vaginitis. Cutaneous candidiasis can involve the skin (e.g., dermatitis, intertrigo, etc.) and/or nail structures (e.g., onychomycosis, paronychia, etc.). Mucocutaneous candidiasis can be acute or chronic. No specific time course formally distinguishes the “acute” from the “chronic” state; rather, clinical response to therapy, and thus, its recurrence or recalcitrance, is often used to guide the diagnosis of chronic mucocutaneous candidiasis (CMC).



**Figure 1.** Human immunological framework mediating susceptibility to candidiasis. Colonization of the epithelial surface results in a commensal state. Transition to virulent forms and the elaboration of candidalysin promotes epithelial invasion. “Type 17” immunity, along with IL-22 responses, are key for superficial defenses, while myeloid responses guard against systemic disease. See text for details.

The determinants of the transition from a commensal form to a pathogen at this interface include fungal virulence factors and alterations in the host defense mechanism. Selective defects in epithelial barrier integrity may permit CMC development based on its occurrence in the rare genodermatosis, keratitis, ichthyosis, and deafness (KID) syndrome [7–17]. KID syndrome, due to heterozygous mutations in the Gap junction beta-2 (*GJB2*) gene that encodes connexin-26 (Cx26), is complicated by chronic skin infections in approximately half of reported cases, with CMC being the most common [16,18,19]. Cx26 is a member of the connexins family, which oligomerize to form hemichannels in epidermal keratinocytes that either become functional channels at the plasma membrane or form gap junction channels by partnering with a hemichannel from an adjacent cell [20]. Cx26 controls dermal homeostasis by regulating keratinocyte differentiation and proliferation [21]. *GJB2* causal mutations precipitate the excessive opening of the Cx26 hemichannel, rendering it insensitive to the intra-epidermal variation in pH, and thus enabling the leakage of biomolecules that results in the increased proliferation of keratinocytes (hyperkeratosis) [20]. How such mutations confer susceptibility to CMC, whether at the level of the keratinocytes and/or through the impaired function of immunocytes, is unclear. Taki and colleagues have shown that human keratinocytes expressing a human mutation in Cx26 (p.D50N) demonstrated down-regulated expression of IL1A, IL-15, IL23R, CCL5, and TLR5 compared with wild-type keratinocytes, thus supporting the possibility that an immunologic defect intrinsic to epithelial cells may contribute to disease susceptibility [22]. On the other hand, older studies (prior to the discovery of mutant *GJB2* as the cause, and thus, in cases that were not genetically confirmed) have reported increased IgE levels [23,24], the absence of delayed-type hypersensitivity to *Candida* antigen despite CMC [25], impaired lymphocyte proliferative response to *Candida* in vitro [25], or defects in neutrophil chemotaxis [26]. Rerknimitr et al. reported a patient with CMC, KID syndrome (harboring the p.D50N mutation in *GJB2*), and the absence of S100<sup>+</sup> cells (representing Langerhans cells (LC) in the epidermis) [15]. Since LC have been shown to drive Th17 cell responses [27], this finding could pathophysiologically connect to KID syndrome, its CMC, and impaired IL-17 responses, although clearly more work is needed to confirm the paucity of LC in this disorder and mechanistically demonstrate “Type 17” immunodeficiency. Interestingly, no publications of other cases of genodermatosis (which were genetically confirmed) associated with an increased risk of CMC were identified. Thus, the hyperkeratosis of KID syndrome may contribute to candidal colonization, based on a similar phenomenon seen in psoriasis [28]; however, the absence of superficial candidiasis in the latter (along with psoriasis being an IL-17-potentiated process [29,30]) suggests that hyperkeratosis per se is insufficient to increase the risk of fungal disease. Intriguingly, KID syndrome can also predispose one to the dermatophyte *Trichophyton* sp. (see “Dermatophytosis” below).

The adhesion of *C. albicans* to epithelial cells triggers filamentous growth, i.e., the formation of pseudo-hyphae and hyphae, which is coupled with the elaboration of candidalysin, a fungal toxin that damages epithelial cell membranes [31]. The presence of candidalysin indicates a barrier breach, eliciting an epithelial host response consisting of antimicrobial peptides, chemokines, and cytokines, including IL-1 $\alpha$ , IL-6, IL-8, G-CSF, and GM-CSF [31]. IEI saliently marked by CMC have been instrumental in defining the importance of the IL-17 response in coordinating susceptibility to superficial candidiasis (Table 1). CMC has long been recognized as a feature of severe combined immunodeficiency (SCID), a heterogeneous group of inherited disorders marked by T cell deficiency, with the concomitant deficiency of B- and/or NK-cells. Other, acquired defects of T cells (e.g., HIV/AIDS, dysfunctions found in transplant recipients, etc.) were also known to be significantly affected by CMC. Collectively, these conditions pinpoint the role of T cells in susceptibility to CMC. The importance of the CD4<sup>+</sup> T helper cells subset characterized by the production of IL-17 (Th17 cells) was elucidated via dominant-negative STAT3 deficiency (DN-STAT3, previously called Autosomal-dominant hyper-IgE syndrome (AD-HIES) and originally termed “Job syndrome”), which is characterized by connective tissue anomalies (e.g., distinct facial features, hyperextensible joints, osteoporosis/bone fractures, scoliosis,

retained primary teeth, and coronary and cerebral aneurysms), “cold” skin abscesses (boils) due to *S. aureus*, and CMC. Heterozygous mutations in the transcription factor STAT3 causing molecular dominant inhibition were discovered in 2007 [32]. Soon after, these mutations were found to impair the development of the adaptive Th17 cells, providing a key breakthrough in the understanding of the importance of IL-17-mediated responses for anti-candidal mucosal immunity [33].

**Table 1.** IEI with increased susceptibility to CMC.

| Inborn Error of Immunity   | Gene    | Mode of Inheritance | Comment                                                                                     |
|----------------------------|---------|---------------------|---------------------------------------------------------------------------------------------|
| Decreased IL-17 production |         |                     |                                                                                             |
| DN-STAT3 (Job’s syndrome)  | STAT3   | AD                  | Decreased Th17 responses                                                                    |
| IL12p40 deficiency         | IL12B   | AR                  | MSMD                                                                                        |
| IL12Rβ1 deficiency         | IL12RB1 | AR                  | MSMD                                                                                        |
| ZNF341 deficiency          | ZNFR341 | AR                  | Phenocopy of DN-STAT3 (Job’s syndrome)                                                      |
| PGM3 deficiency            | PGM3    | AR                  | Congenital disorder of glycosylation                                                        |
| STAT1 GOF                  | STAT1   | AD                  | Associated with various infections and aneurysms, autoimmune disease, and/or tumours        |
| CARD9 deficiency           | CARD9   | AR                  | Variably decreased Th17 responses                                                           |
| Decreased IL-17 response   |         |                     |                                                                                             |
| IL-17F deficiency          | IL17F   | AD                  | Afflicted patients may also be at risk for <i>S. aureus</i> skin and soft tissue infections |
| IL-17RA deficiency         | IL17RA  | AR                  | Afflicted patients may also be at risk for <i>S. aureus</i> skin and soft tissue infections |
| IL-17RC deficiency         | IL17RC  | AR                  |                                                                                             |
| ACT1 deficiency            | ACT1    | AR                  | Afflicted patients may also be at risk for <i>S. aureus</i> skin and soft tissue infections |
| JNK1 deficiency            | JNK1    | AD                  | Syndrome with features of Ehler–Danlos-like connective tissue disorder                      |
| APECED                     | AIRE    | Typically AR        | Endocrinopathies.<br>Neutralizing auto-antibodies to IL-17 and IL-22                        |

STAT3 transduces the signaling effects of multiple cytokines, notably, IL-6, IL-10, and IL-23, thereby inducing the transcription of RORγt, which is critical for the development of Th17 cells, and the subsequent elaboration of IL-17 and IL-22. Loss-of-function (LOF) mutations in STAT3 thus impair Th17 differentiation, thereby crippling the IL-17 and IL-22 responses. Within the IL-17 family of cytokines, IL-17A and IL-17F have been best characterized in terms of fungal immunity; they function as homo- and hetero-dimers that signal through the IL-17 receptor (IL-17R), which is composed of IL-17RA and IL-17RC subunits [34,35]. IL-22 belongs to a distinct family of cytokines that acts on its cognate receptor (IL-20R), which is composed of the IL-22RA1 and IL-10RB subunits. IL-17R and IL-22R are expressed on keratinocytes. The distinct contributions of IL-17 and IL-22 to CMC immunity were shown in the elegant work conducted by the Gaffen lab [36]: IL-17A and IL-17F act on the supra-basal, post-mitotic epithelial cell layers (SEL), whereas IL-22 stimulates the basal, proliferative progenitor cells (BEL). IL-22’s activation of its cognate receptor on BEL and, consequently, its signaling through STAT3 lead to the replenishment of SEL that are responsive to IL-17. The IL-17 response, in turn, leads to a signature response of antimicrobial peptides, neutrophil-activating chemokines, and acute-phase response proteins [37].

Based on the compromised Th17 pathway in DN-STAT3 deficiency, the IL-17/IL-22 pathways were investigated in other IEI marked by CMC. Indeed, impaired IL-17 immunity emerged as a common theme in these disorders. Decreased IL-17 production was demonstrated in autosomal recessive (AR) IL-12p40 and IL12RB1 deficiency, which manifest principally with mycobacterial disease (see below), but in which 6–25% of patients are also affected with CMC [38–40]. In addition to diminished circulating IL-17-producing T cells, these patients also show decreased IL-22 production. Similarly, bi-allelic mutations in *ZNF341*, the transcription factor that regulates STAT3, lead to the loss of STAT3 and a phenocopy of AD-HIES/Job syndrome, including CMC with diminished IL-17 responses [41]. AR PGM3 deficiency is a congenital disorder of glycosylation that has occasionally been reported to manifest with CMC [42–44]. The precise immunological defect resulting from mutant PGM3 conferring susceptibility to CMC is not entirely understood, but slightly diminished Th17 levels have been described [42].

STAT1 gain-of-function (GOF) was originally discovered as a cause of CMC in 2011 for which poor production of IL-17 and IL-22 was observed [45,46]. It has since been identified as the most common genetic etiology of CMC. In addition to the latter infection, STAT1 GOF can also present with other mycoses (see below), non-fungal infections, and aneurysms, autoimmune diseases, and/or tumours [47]. Subsequently, AR ROR $\gamma$ t deficiency, due to biallelic mutations in *RORC* (the transcription factor essential for Th17 lineage development), was identified [48]. AR CARD9 deficiency can be associated with CMC, although its more flagrant presentation is one of invasive fungal disease (see later). Loss of CARD9 function has variably been found in association with diminished, albeit not abolished, Th17 responses [49–53].

In addition to the preceding disorders that decrease IL-17 (and selectively IL-22) production, the distinctive feature of CMC and the principal of physiologic homogeneity led to the discovery of IEI that cause impaired responses to IL-17 due to either autosomal dominant (AD) mutations in the IL-17F cytokine [54], AR mutations in the IL-17 receptor subunits (IL17RA or IL17RC) [54–58], AR mutations in the adaptor molecule required for IL-17 receptor signaling (ACT1) [59–63], or AD mutations in the kinase essential for its signaling (JNK1) [64]. To date, no mutations compromising IL-22 signaling have been identified.

Beyond genetic lesions that compromise the production of IL-17/IL-22 or the response to IL-17, CMC can also result from the neutralization of these two cytokines, as shown in autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED, also called Autoimmune polyglandular syndrome type 1 [APS-1]), which is typically due to AR mutations in *AIRE*, although dominant-negative mutations have also been reported [65,66]. APECED patients classically present with CMC and endocrinopathies (hypoparathyroidism and adrenal insufficiency) but can also occasionally present autoimmune disease (e.g., enteritis, pneumonitis, etc.) or dental anomalies. The anti-cytokine auto-antibodies in APECED are present due to the loss of *AIRE* function, which leads to the loss of thymic regulation and, consequently, the escape of auto-reactive T cells.

Although CMC is clearly associated with a loss of IL-17 (+/– IL-22) immunity, the decreased levels of circulating Th17 cells identified represent adaptive (antigen-specific) T lymphocytes. Murine data have been used to identify the presence of mucus-residing  $\gamma\delta$  T cells, invariant natural killer T (iNKT) cells, innate lymphoid cell type 3 (ILC3) cells, and TCR+ ‘natural’ Th17 cells (nTh17). All the cell populations listed are collectively referred to as “Type 17” cells, which are capable of producing IL-17 and IL-22 as well as GM-CSF [67,68]. Given their strategic location at the epithelial portal of entry, one or several of these cell types may be the immediate IL-17/IL-22 source in humans upon mucosal invasion by *Candida* sp. Further advances in characterizing tissue immunity on human mucocutaneous surfaces may help address this question and potentially guide the identification of novel genes responsible for these localized responses.

### 3.2. Invasive, “Deep-Seated” Candidiasis (IC)

Despite the persistent nature of CMC, with a clinically apparent fungal burden, CMC and IC are often mutually exclusive, implying the presence of distinct immunologic responses generated to control the respective disease and/or the compartmentalization of shared host defense pathways. In modern medicine, IC is most often the result of iatrogenic interventions or exogenous immunosuppression. In patients with acute leukemia, it has long been recognized that the risk of IC, as well as the severity of the fungal disease, correlate with the degree and duration of granulocytopenia, thus establishing circulating granulocyte levels as a proxy for risk of infection [69–71]. The risk also correlated with anti-leukemic agents and their toxicity to the gastrointestinal tract, with mucositis being an important determinant of invasive disease. This setting helped establish the importance of myeloid cells, particularly granulocytes (and primarily neutrophils at that), in the host defense against IC.

In distinction to the aforementioned paradigm, IC is rarely seen in IEI with myeloid defects. For example, “Congenital Neutropenia” (CN) refers to a heterogeneous group of disorders compromising neutrophil development. Despite the neutropenia (which can be profound), the risk of IC is not increased in these disorders [72]. Similarly, leukocyte adhesion deficiency (LAD) refers to a group of disorders in which mutant adhesion molecules (either integrins or selectins) impair leukocyte extravasation from blood vessels. Despite persistent neutrophilia, neutrophils are unable to mobilize to sites of infections, causing patients to develop recurrent/severe bacterial infections with a lack of pus formation. LAD patients have been considered at risk for fungal infections, but in reality, these manifest as intermittent episodes of mucosal candidiasis rather than IC [73–77]. Specific granule deficiency (SGD) is marked by neutrophils lacking secondary granule proteins along with an atypical nuclear morphology. SGD may be caused by both homozygous and heterozygous mutations in *CEBPE* [78]. Secondary (or specific) granules contain the antibacterial proteins lactoferrin, neutrophil gelatinase-associated lipocalin, cathelicidin, and lysozyme [79]. This functional defect results in recurrent bacterial infections but no reported increased risk for invasive fungal disease, including IC. Chronic granulomatous disease (CGD) is an inborn error of the phagocyte NADPH oxidase (phox) complex (see “Aspergillosis” section). The dysfunction of the phagocyte respiratory burst and the compromised generation of reactive oxygen species leads to recurrent bacterial and fungal infections as well as dysregulated granulomatous inflammation (e.g., colitis). Although patients with CGD are unambiguously at an increased risk for fungal infection, the mycoses tend to originate from mould (see the “Aspergillosis” section), and the risk of IC is remarkably rare. The rate of the latter may have been further influenced by the use of antifungal prophylaxis. Itraconazole prophylaxis was first shown to be effective in mitigating aspergillosis in 1994 [80,81]; subsequently, it was validated in a randomized clinical trial nearly a decade later [82]. Given that itraconazole prophylaxis is also effective against IC in other at-risk groups [83–86], it may have affected the modern risk of IC for patients with CGD. Nonetheless, IC was reported in CGD patients (diagnosed by nitroblue tetrazolium test) prior to the 1994 landmark findings [87–90] and subsequently [91,92]. Whether some of these cases may be iatrogenic (e.g., vascular-catheter-associated) is not clear, although in some reports, this does not seem to be the case. In distinction to IC, CGD is associated with an increased risk of systemic disease via infection with another yeast, *Trichosporon* [93–97]. Given the challenges in the antifungal management of trichosporonosis, biopsies for pan-microbiological culture should be performed. Collectively, these IEI indicate that myeloid defects do not automatically equate to an increased risk of IC.

In distinction to the absence of IC in the above IEI, there are rare reports of IC occurring in the absence of a known IEI. Specifically, there are historical descriptions of a fatal form of generalized moniliasis (the previous term for ‘candidiasis’) occurring in the absence of risk factors, including a fatal case involving the central nervous system [98]. Since then, several sporadic cases of spontaneous CNS candidiasis (sCNSc) have been reported, in which *Candida* sp. (primarily *C. albicans*) caused the infection of the brain and/or meninges without

a preceding clinical intervention, underlying immunocompromising disorder, or apparent immunosuppression. Interestingly, many reports involved French-Canadian [99–102] and French [103–105] patients. With medical mycology focusing on the more-common IC/candidemia occurring in hematologic malignancies, transplants, and iatrogenesis, this historical syndrome had almost been forgotten until the discovery of AR CARD9 deficiency [49]. Although the initial work highlighted the susceptibility of CARD9-deficient patients to CMC, the clinical description of presumed affected family members included one with candidal meningitis who developed seizures and hydrocephalus and then died, one with tumorous skull destruction, and one with candida meningo-encephalitis [49]. Drewniak and colleagues also reported a case of *C. dubliniensis* meningoencephalitis due to CARD9 deficiency [50]. The similarities of these CARD9-mutant cases to the historical sCNSc reports, along with the identification of a French-Canadian patient with sCNSc due to a c.T271C (p.Y91H) mutation [51], led to the recognition that biallelic mutations in CARD9 underlie sCNSc.

sCNSc is a form of chronic (subacute), disseminated candidiasis that is often the subject of clinical attention because of the involvement of the CNS (brain, meninges, and/or spinal cord) or related structures (e.g., the eye). However, non-neurological structures can also be affected, including the colon, liver, and bones [106–109]. The exact mechanism underlying susceptibility to invasive infection remains to be deciphered. However, one mechanism contributing to disease severity may be related to the impaired recruitment of neutrophils to sites of infection and/or impaired neutrophil clearance of yeast cells [50,110]. Mononuclear cell responses may also be defective (D. Vinh, manuscript in progress). These myeloid defects likely underlie the indolently progressive nature of reported cases, which can span years (presuming that these reports represent a single fungal clone that relapses). This presentation is in gross distinction from the acute forms of candidiasis seen in classical at-risk groups, which can be fulminant and is more reminiscent of the chronic, disseminated (hepatosplenic) candidiasis accompanying myeloid reconstitution following intensive cytoreductive chemotherapy [111,112]. Antifungal therapy may be effective in controlling sCNSc disease, although not invariably so. The cessation of antifungal therapy may be associated with a relapse of symptoms. It is unclear if the surgical resection of brain masses (often performed under the suspicion of a malignancy that is subsequently found to be an abscess) improves prognosis. Given that CARD9 deficiency has been associated with impaired peripheral blood mononuclear (PBMC) responses to *C. albicans* in vitro, including decreased fungus-induced GM-CSF production [51,52], and that large studies have shown that healthy controls distinctively produce GM-CSF in response to *C. albicans* (relative to other agonists) [113,114], adjuvant immunotherapy with recombinant human GM-CSF has been successfully used in some patients [51,52]. Similarly, given the associated myeloid defects, adjuvant G-CSF has also been effective [115,116]. Stem cell transplantation has also been attempted, wherein success led to clinical remission [117].

CARD9 deficiency has also been sporadically reported to predispose one to invasive disease caused by other yeast-like organisms, including *Saprochaete capitata* (previously known as *Geotrichum capitatum*) [118] and *Prototheca zopfii* (which is an achlorophyllous alga, not a fungi) [119]. Thus, it is important to carry out cultures and identification analyses to optimize antifungal therapy, and the reporting of such cases is informative.

#### 4. Thermally Dimorphic Endemic Mycoses

Thermally dimorphic endemic fungi (TDEF) are saprophytes characterized by temperature-dependent morphological variation (i.e., they exist in a mould form at ambient temperature and assume a yeast-like phase at body temperature) and geographical restriction. These fungi include *Blastomyces*, *Coccidioides*, *Emergomyces*, *Histoplasma*, *Paracoccidioides*, *Sporothrix*, and *Talaromyces*. Updates on the microbiology of these fungi have been recently reviewed [120–123]. In their geographic niches, these fungi are environmentally distributed, and, based on previous seroprevalence or delayed-type hypersensitivity-based investigations, exposure is both frequent and generally asymptomatic or symptomatically

self-limiting. On the other hand, some of these fungi are known to cause a variety of morbid syndromes in both overtly immunocompromised and apparently immunocompetent individuals. The field of IEI provides insight into the basis for the latter.

The first report of human blastomycosis in the Americas is attributed to Thomas Gilchrist, who presented the dermatological microscopic findings of a patient that was referred to him and, subsequently, detailed a second case of a chronically progressive, waxing-and-waning form of cutaneous blastomycosis [124]. Blastomycosis is most commonly caused by the *B. dermatitidis* complex (encompassing *B. dermatitidis* and *B. gilchristii*). Blastomycosis is said to be endemic to specific North American states and provinces that border the St. Lawrence River, the Great Lakes, the Mississippi River, and the Ohio River, although the geographic distribution of the causative fungi (and TDEF in general) is in flux owing to climate change and the human disruption of ecological niches [125–128]. Infection occurs via the inhalation of aerosolized conidia. The most common clinical syndromes of blastomycosis include acute pneumonia, chronic pneumonia, and extrapulmonary (disseminated) disease, occurring in 25–40% of patients, which can involve the skin, musculoskeletal system, genitourinary tract, and/or the central nervous system [129].

Coccidioidomycosis (also known as San Joaquin Valley Fever, Valley fever, or Desert fever) is caused by two morphologically similar but genetically distinct fungi, *C. immitis* and *C. posadasii*. It was first described in 1888 by Alejandro Posadas and his mentor, Robert Wernicke, after their evaluation of Domingo Ezcurra, who had suffered from an indolently progressive but ultimately fatal form of the disease, and whose head is enshrined in a museum in Argentina [130]. *Coccidioides* is predominantly endemic in the Southwestern USA and Central and South America. Infection occurs following inhalation of arthroconidial spores. Coccidioidomycosis can manifest as various lung diseases (e.g., acute pneumonia, chronic progressive fibrocavitary pneumonia, pulmonary nodules, or cavities) or, in <1 to 5% of those infected, as a disseminated disease; the most common sites of involvement of the latter are the CNS, skin, musculoskeletal system, and lymph nodes.

Emergomycosis is caused by *Emergomycetes* sp., whose genus was newly established in 2017 to include *Emmonsia*-like fungi [131]. Although disseminated disease has been reported in patients with advanced HIV or exogenous immunosuppression, no report of such mycosis associated with IEI has been identified.

In the Americas, histoplasmosis is typically caused by *H. capsulatum* (and its variants), whereas *H. duboisii* (restricted to Central and West Africa) causes African histoplasmosis. Although *H. capsulatum* was originally thought to be restricted to certain areas (e.g., central and eastern states surrounding the Ohio and Mississippi River Valleys, as well as the St. Lawrence River valley), it is now recognized that different species from the *Histoplasma* complex can be found worldwide [132]. Indeed, the first description of histoplasmosis by Samuel Darling in 1905 was of a disseminated, fatal form in a 27-year-old Martinique man presenting febrile vomiting and whose autopsy showed leptomeningitis with the involvement of the spleen and marrow [133]. Infection occurs via inhalation of small hyphal fragments and microconidia. Histoplasmosis can manifest as an acute pulmonary disease (varying from a mild/pauci-symptomatic form to severe/life-threatening forms); as complications associated with acute pulmonary disease that are thought to be immunologically mediated (e.g., erythema nodosum, arthritis, pericarditis, granulomatous, or fibrosing mediastinitis); as chronic cavitary pulmonary disease; or as disseminated disease [134].

Paracoccidioidomycosis (PCM), also known as South American blastomycosis, was first described by Adolpho Lutz in Brazil in 1908 who identified the disease in two patients with oral mucosal lesions and cervical lymphadenopathy [135]. Although PCM was attributed to *P. brasiliensis*, phylogenetic studies have identified various species within the *Paracoccidioides* species complex, although no distinction in clinical manifestations have been definitively established [136]. Infection is acquired by the inhalation of conidia. PCM is classified into distinct forms: The regressive form is a-/pauci-symptomatic, involving the lungs. The progressive disease can be acute/subacute (primarily with lymphadenopathy,

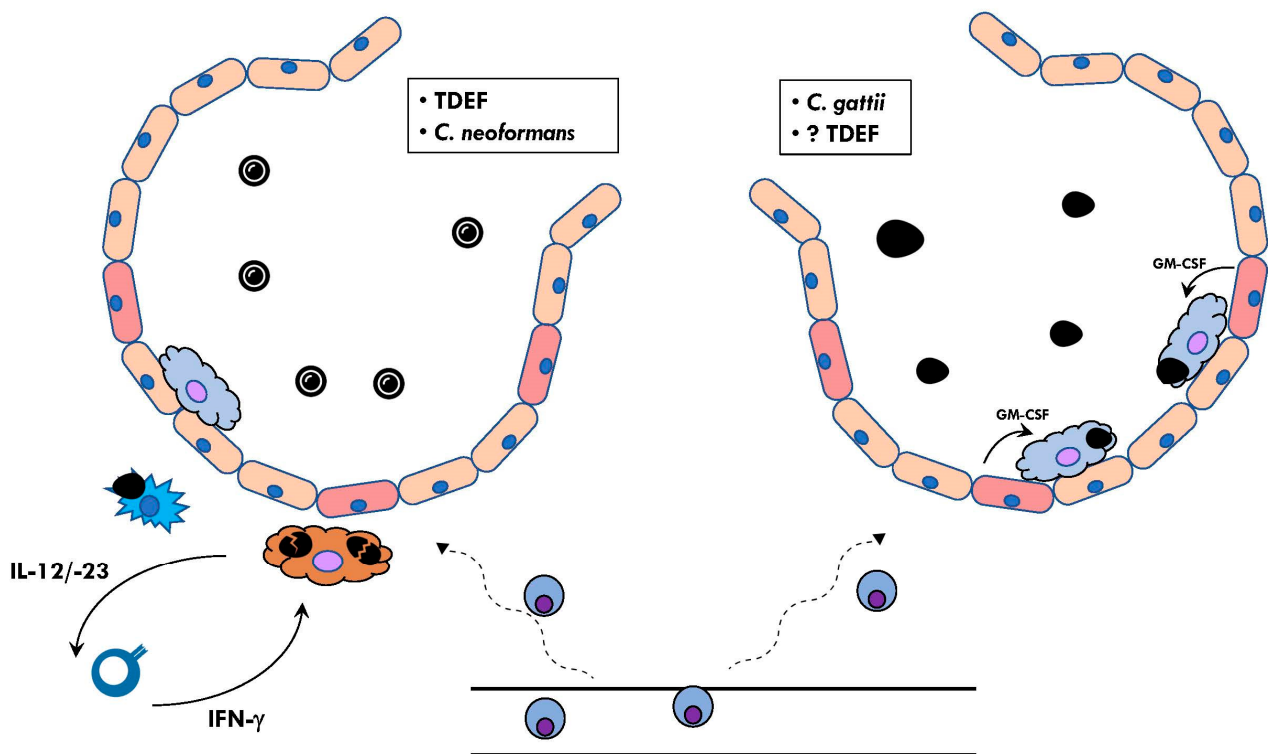
hepatomegaly, and/or splenomegaly) or chronic (involving the lungs, and, usually, the skin, mucosa, and/or viscera, such as the adrenal glands or CNS) [136].

Sporotrichosis was first reported by Benjamin Schenck in 1898 who identified the disease through multiple refractory skin ulcers along the forearm lymphatics [137]. Unlike other TDEF, sporotrichosis more commonly occurs following the soft tissue traumatic implantation of infectious propagules from an environmental source; inhalation is a less frequent route of acquisition. Caused by members of the *S. schenckii* complex, sporotrichosis patients can present skin disease (cutaneous-lymphatic or nodular lymphangitis), disseminated disease (with visceral involvement), or extracutaneous presentations (e.g., CNS, musculoskeletal, or pulmonary) [138].

Talaromycosis (formerly Penicilliosis) is caused by *T. marneffei* (previously known as *P. marneffei*), the only TDEF within the *Talaromyces* genus that is pathogenic to humans. The fungus is notably endemic in South and Southeast Asia. It was first described by Gabriel Segretain and colleagues, who also reported the first human infected by the disease after accidental autoinoculation [139–141]. The first report of a naturally occurring infection was in the form of disseminated disease (wherein the fungus was isolated from a removed spleen) in a patient with Hodgkin's disease [142]. Jayanetra and colleagues reported extensive disseminated disease in five patients, three of whom had no underlying disorder, causing death in three of the patients [143]. Infection follows inhalation of spores from the environment; although the bamboo rat is a known natural reservoir of the fungus, the exact environmental source, and how the bamboo rat contributes to transmission to humans, is unclear [144]. Given the fact that talaromycosis is a neglected tropical disease [145], our understanding of its clinical manifestations is limited [146]; thus, the prevalence of asymptomatic exposure is unclear. Pulmonary talaromycosis can include mass lesions, consolidation, cavities, effusion, or mediastinal lymphadenopathy. Patients with disseminated disease can present with skin lesions (which may be progressive and disfiguring), hepato/spleno-megaly, visceral infection, and/or CNS involvement.

The general pathophysiology for these mycoses is similar to that for mycobacteria: the inhalation of the infectious propagules results in their deposition in the alveoli, where they are ingested by alveolar macrophages. Under the influence of body temperature, the propagules convert to their yeast-like form intracellularly, disseminating to regional lymph nodes. The combination of innate and adaptive immunity attempts to contain the infection, wherein the formation of granuloma occurs. T-cell-mediated immunity is critical for this containment. The failure of this process results in systematic, progressive dissemination. This similarity to the pathophysiology of mycobacteria has been central to identifying IEI associated with TDEF (Figure 2).

Genetic defects in the capacity to produce or respond to interferon-gamma (IFN- $\gamma$ ) have been the IEI most frequently reported associated with TDEF mycoses (Figure 2). Most of these fall in the collection of disorders referred to as Mendelian Susceptibility to Mycobacterial Disease (MSMD) [147–149]. While MSMDs were initially recognized for their distinct susceptibility to mycobacteria (particularly non-tuberculous (NTM) and occasionally *M. tuberculosis*) and *Salmonella* (predisposing victims to extra-intestinal salmonellosis) [38,39,150–154], they were also subsequently found in patients with otherwise unexplained disseminated TDEF infection, including histoplasmosis (IL12RB1 [153,155–157]; IFNGR1 [158]), coccidioidomycosis (IFNGR1 [159] and IL12RB1 [160]), and paracoccidioidomycosis (IL12RB1 [161]). These findings provided the framework of human immunity to TDEF, which was followed by the identification of other IEI congruent with defective IFN- $\gamma$ -mediated immunity, including CD40L deficiency [162–170] (discussed further in the section on “Pneumocystis”), GATA2 deficiency [171], and STAT1 GOF [168,169,172,173].



**Figure 2.** Immune pathways governing susceptibility to thermally dimorphic endemic fungi (TDEF) and cryptococci. Left: Inhalation of infectious propagules from TDEF or *C. neoformans* results in a cell-mediated immune response dependent on the production of IFN- $\gamma$  and driven by IL-12/-23. Right: Host defense against *C. gattii* also relies on GM-CSF immunity. See text for details.

GATA2 deficiency, due to heterozygous germline mutations in the GATA2 transcription factor, results in a wide spectrum of syndromes, including MonoMAC [174–176]; dendritic cell, monocyte, B, and NK lymphoid (DCML) deficiency [177]; familial myelodysplasia/acute myeloid leukemia (f MDS/AML) [178]; Emberger syndrome (entailing deafness, lymphedema, and MDS/AML) [179]; chronic neutropenia [180]; and Warts, Immunodeficiency, Lymphedema, and anogenital Dysplasia (WILD) syndrome [181,182]. The core feature of GATA2 deficiency involves defective myeloid homeostasis. Notably, the progressive development of monocytopenia and dendritic cell cytopenia, with variable degrees of lymphocytopenia, occurs. Intriguingly, despite the eventual (near)-complete absence of monocytes and DC, tissue macrophages and Langerhans cells (LC) are observed in this disease (e.g., in tissue biopsies). However, the development of opportunistic infections, such as disseminated disease with mycobacteria or with TDEF and the predisposition to pulmonary alveolar proteinosis (PAP, entailing the inability of alveolar macrophages to clear a surfactant from the airspace), suggests that such tissue macrophages/DC are functionally impaired. Whether that dysfunction is cell-intrinsic, as suggested in a mouse model [183], or due to a loss of cooperation from incoming monocytes/DC, or both, is not clear. One of the sentinel descriptions of GATA2 deficiency noted susceptibility to disseminated histoplasmosis, which was noted in several patients [174]. Subsequently, non-disseminated blastomycosis (pulmonary only, reportedly severe) was identified in a woman with GATA2 deficiency, although the trigger to identify this inborn error of immunity was not related to the fungal disease but rather the subsequent development of herpes-simplex-virus-induced Hemophagocytic lymphohistiocytosis (HLH) nine years later [171]. Variants in GATA2 have also been identified in a Brazilian cohort in association with histoplasmosis, paracoccidioidomycosis, and sporotrichosis [184]: Most of the variants were predicted in silico to be benign, while one was rare and classified as a variant of undetermined significance (minor allele frequency <0.01%; associated with paracoccid-

oidomycosis). Unfortunately, none of the variants were characterized molecularly or functionally. It is presumed that the progressive loss in the quantity and/or defective function of monocytes/DC compromise the effector arm of the IFN- $\gamma$  axis, resulting in the failure of the containment of the fungal propagules and, subsequently, dissemination.

Mutations in STAT1 can result in a plethora of IEI. On the one hand, biallelic loss-of-function (LOF) mutations cause AR MSMD syndrome, with either partial or complete functional deficiency, resulting in susceptibility to mycobacteria and viruses [185–187]. Heterozygous LOF mutations can cause AD MSMD [188]. On the other hand, heterozygous GOF mutations cause “STAT1 GOF”, an inborn error of immunity that is distinct from MSMD and marked by susceptibility to TDEF [168,169,172,173] and other fungi (see “Chronic mucocutaneous candidiasis” and “Cryptococcus”). STAT1 GOF is a molecular gain-of-function mutation, that is, it allows for the normal phosphorylation of STAT1 but it is maintained for an abnormally long period of time (i.e., a prolonged phosphorylated state), leading to a loss of cellular function via a loss of responsiveness (tachyphylaxis) to repeated IFN- $\gamma$  stimulation.

DN-STAT3 (see “Chronic Mucocutaneous Candidiasis” above) has been reported to be associated with TDEF. The resulting fungal disease in these patients, however, may be atypical in presentation (e.g., tongue, laryngeal, or gastrointestinal histoplasmosis) [189–191], although diffuse/visceral disease has also been reported (e.g., CNS coccidioidomycosis and disseminated talaromycosis) [192–194]. Whereas MSMD, CD40L, MSMD, and STAT1 GOF are unified by impaired IFN- $\gamma$  immunity, the mechanism by which DN-STAT3 results in disseminated TDEF disease is not as well established. Diminished IFN- $\gamma$  production has been identified, albeit with respect to *C. albicans* [195] or *A. fumigatus* [196]. Whether this defect is pertinent to TDEF remains to be demonstrated.

Intriguingly, mutations in T-cell-co-stimulatory molecules other than CD40L, notably those disrupting CD28/CD80 or CD86 (via recessive mutations in CD28 [197] or its essential signaling partners, RLTPR (also known as CARMIL2) [198–205] or CARD11 [206–210]) or ICOS/ICOSL interactions (via recessive mutations in either ICOS [211–216] or ICOSL [217,218]), have not been reported in association with TDEF. Whether this reflects stochasticity (i.e., a lack of exposure from geographic restriction among those with these rare diseases), or whether sufficient pathologic exposure has indeed occurred, but the respective molecules and T cell activation pathways are not critical for TDF host defenses, are questions that remain to be determined.

The recognition that defective production of/response to IFN- $\gamma$  underlies at least some forms of severe TDEF mycoses led to the idea that supplementation with IFN- $\gamma$ , at least for those with diminished production, may be a therapeutic strategy for the curing of infections. Indeed, IFN- $\gamma$  immunotherapy, used adjunctively with antifungals, has been successful as a salvage therapy for refractory coccidioidomycosis in two IEI patients [159]. It has also been used successfully for this mycosis in patients without identified IEI [219–221], although one case presented modest improvement with IFN- $\gamma$ , and the identification of a dysregulated Th2 response with the subsequent addition of dupilumab (to block IL-4- and IL-13-mediated Th2 responses) led to resolution [221]. Whether IFN- $\gamma$ , either alone or with an inhibitor of dysregulated Th2 responses, may be successful for other TDEF remains to be reported.

Given the importance of the IFN- $\gamma$ -immunity framework to TDEF, it was subsequently found that this pathway can be compromised by a distinct, non-monogenic process: the inherent presence of neutralizing auto-antibodies to IFN- $\gamma$  (aa-IFN- $\gamma$ ). Again, the discovery of this immunologic defect underlying susceptibility to an infectious disease was first found in patients with a mycobacterial disease, namely, tuberculosis [222]. Subsequently, high-titer aa-IFN- $\gamma$  concentrations were found in patients with no other immunocompromising condition in association with other severe disseminated infections, including NTM [223–227], and germane to TDEF, *H. capsulatum*, and *T.(P.) marneffei* [228–242] (as well as *Cryptococcus*, see below). aa-IFN- $\gamma$  is an emerging, adult-onset form of immunodeficiency that is typically recognized in the fourth to sixth decades of life. An ethnic predisposition

among Southeast Asians is noteworthy: these autoantibodies have been primarily reported among those of Filipino, Thai, Vietnamese, Japanese, and Chinese ancestry, with other ethnic groups less commonly affected [243,244]. A female predominance has been shown [241], as has a genetic association with the HLA alleles, DRB1\*16:01 and DQB1\*05:02 [245,246]. aa-IFN- $\gamma$  inhibit STAT1 phosphorylation and IL-12 production. Thus, in patients (particularly women of Southeast Asian descent) with TDEF disease, particularly if severe or disseminated, aa-IFN- $\gamma$  should be assessed. Of note, aa-IFN- $\gamma$  were originally discovered in patients living with HIV (albeit without any clear association with infection susceptibility) [247], implying that HIV status and aa-IFN- $\gamma$  are not mutually exclusive in terms of their existence and, possibly, compromising effects on immunity. The management of aa-IFN- $\gamma$  has been challenging; standard broad-spectrum immunosuppressants (e.g., steroids) or immunomodulatory dosing of IVIG have been unsuccessful. Cyclophosphamide has been used with some success [240,248]. Given that this immunodeficiency is due to the presence of rogue autoantibodies, drugs specifically depleting B cells, such as Rituximab (anti-CD20) or Daratumumab (anti-CD38), have been reported to be effective in terms of reducing aa-IFN- $\gamma$  levels and clearing infection [240,249].

### 5. Cryptococcosis

Cryptococcosis is principally caused by yeast members of the *C. neoformans* or *C. gattii* species complexes that have different geographic, microbiologically phenotypic, and genotypic characteristics [250,251]. Cryptococci also have a distinct host-range [252]: The *C. neoformans* species complex (herein referred to as “*C. neoformans*”) has traditionally been viewed as an opportunistic pathogen in various immunocompromising conditions, notably those of HIV and transplant recipients. Meanwhile, the *C. gattii* species complex (herein referred to as “*C. gattii*”) tends to cause disease in apparently immunocompetent people. The distinction between these species complexes is relatively recent and precludes interpretation of which cryptococci were identified in previous reports of IEI patients. Thus, the instances wherein *C. gattii* has been specifically identified from such cases will be indicated.

The molecularly defined IEI associated with severe or disseminated cryptococcosis strikingly overlap with those that predispose one to infection with TDEF, namely, MSMD [253–255], GATA2 [174,176], CD40L deficiency [163,256–263], STAT1 GOF [47,264,265], and DN-STAT3 [266,267]; these IEI have been detailed above. This overlap most likely reflects a comparable, if not homologous, immunopathophysiology (Figure 2). It also underlines a distinct feature of the genotype/phenotype correlations of IEI: the infection phenotype requires microbial exposure, which may be highly variable between individuals with the same lesioned gene, resulting in variable expressivity, at least of the mycoses. It should also be noted that cryptococcosis can be seen in “idiopathic CD4<sup>+</sup> lymphocytopenia” (ICL), an immunologic phenotype that likely reflects the convergence of heterogeneous causes; in the absence of a clear genetic etiology, ICL is not included among these IEI, although clinicians should be aware of this possibility during their investigations.

As with the TDEF, aa-IFN- $\gamma$  have also been found in patients with unexplained cryptococcosis [231,268,269]. The characteristics of aa-IFN- $\gamma$  are detailed above (see “Thermally dimorphic endemic mycoses”). Compared to HIV-infected patients, these patients with aa-IFN- $\gamma$  tended to be less likely to present with meningitis and were more likely to present with bone and/or joint and lung/pleura as well as skin infections [269].

In addition to aa-IFN- $\gamma$ , high-titer, neutralizing auto-antibodies to GM-CSF (aa-GM-CSF) have been found in patients who were not overtly immunocompromised but who developed cryptococcal meningo-encephalitis that, distinctly, was associated with *C. gattii* [270,271]. aa-GM-CSF were first identified in patients with pulmonary alveolar proteinosis (PAP), a disorder characterized by dyspnea, alveolar infiltrates (determined radiologically), and a risk for progressive respiratory failure [272,273]. PAP results from the unregulated accumulation of a surfactant in the airways due to defective alveolar macrophage clearance [272]. PAP had been previously reported to be occasionally complicated by opportunistic infections, notably cryptococcosis (manifesting as the pulmonary form alone [274]

or with systemic disease [275,276]), thus potentially linking aa-GM-CSF to cryptococcal susceptibility. GM-CSF is a pleiotropic cytokine involved in numerous functions, including the terminal differentiation of monocytes into mature alveolar macrophages, the differentiation of DC, and enhanced myeloid antimicrobial functions [277]. aa-GM-CSF inhibit the GM-CSF-induced STAT5 phosphorylation of peripheral blood mononuclear cells in vitro, although the exact pathophysiology of how they facilitate disseminated disease remains to be deciphered. Cryptococcosis (and other opportunistic infections) can precede, occur concomitantly with, or follow the clinical development of PAP. Intriguingly, the loss of the GM-CSF pathway is associated with susceptibility to another neurotropic infection (i.e., nocardiosis [278,279]), and the selective loss of GM-CSF response by PBMC to *C. albicans* may also contribute to candidal invasion of the CNS through CARD9 deficiency [51,52] (see “Invasive Candidiasis” above), suggesting a role for GM-CSF in systemic protection against CNS infection. The underlying basis for the development of aa-GM-CSF is unclear. In addition, while both aa-IFN- $\gamma$  and aa-GM-CSF underlie susceptibility to “cryptococcosis”, it is unclear from the current published reports whether these auto-antibodies segregate with mutually exclusive susceptibility to the species complex (e.g., aa-IFN- $\gamma$  to *C. neoformans* vs. aa-GM-CSF to *C. gattii* only) and whether these auto-antibodies can co-exist in patients.

Intriguingly, historical reports of patients with PAP complicated by histoplasmosis have also been reported [276,280], although no reports of aa-GM-CSF underlying human cases of histoplasmosis have been identified.

## 6. Pneumocystis

*Pneumocystis* was discovered in 1909 and, at the time, was thought to be a protozoan parasite (*Trypanasoma*); genomic analysis resulted in its subsequent re-classification as a fungus in 1988 [281]. Within this fungal genus are several species, each of which demonstrate strict target–host species specificity, that is, they are able to infect only one host species. Thus, humans can be infected with *Pneumocystis jirovecii* and are, consequently, the only source capable of infecting other humans, while the previously more popular term “*P. carinii*” is the formal name for the species that infects rats [282].

*P. jirovecii* pneumonia (Pjp) was initially reported in the late 1930s in Europe, with subsequent clusters reported in the 1940s and 1950s, as an unusual pulmonary infection characterized by a prominent infiltration of mononuclear cells thought to be plasma cells (hence the term “interstitial plasma cell pneumonia”) [283,284]. Strikingly, these epidemics affected premature and malnourished (“dystrophic”) infants, usually between 6 weeks and 4 months of age, for which mortality rates were as high as 50% and wherein autopsy series noted pneumothorax, mediastinal emphysema, and alveolar exudate filling the gray lungs. While the immunological cause of these Pjp outbreaks in infants remains unclear, Pjp was recognized as a potential cause of fatal infection in SCID through the Vetter family (with death of an older brother) and other reports [285–287], and clusters of Pjp in previously healthy adults in the 1980s heralded the HIV/AIDS epidemic [288–290]; this provided the first evidence that profound cell-mediated immunodeficiency was conducive to the development of disease.

Exposure to *P. jirovecii* seems to be common: based on serology data of healthy US children, two-thirds had detectable *Pneumocystis* antibodies by the age of 4 years [291]. Using molecular and serologic techniques, it was found that asymptomatic exposure was frequent in Chilean children [292]. Infection is thought to occur primarily following the person-to-person transmission of asci (cysts) via respiratory secretions. As with other fungi, *P. jirovecii* is not a professional pathogen, as asymptomatic colonization has been documented at high frequencies [293]. The adherence of trophozoites to alveolar type I pneumocytes initiates the infectious process, which progresses and may ultimately be fatal in immunocompromised patients. The exact immunopathophysiology of Pjp is a challenge to study owing to, among other things, the difficulty of culturing the organism. However,

the natural history of Pjp in IEI underscores the importance of T cell immunity with respect to mitigating disease.

Combined immunodeficiency (CID) refers to a diverse group of IEI marked by defective cellular and humoral immunity. In its most severe form (Severe CID (SCID)), manifestations occur in infancy and typically include failure to thrive, recurrent infections, and opportunistic infections. Pjp can be a life-threatening infection in these IEI: prior to the routine use of prophylaxis, the overall Pjp infection rate in SCID was at least 27%, an estimated attack rate of at least 0.34 episodes per patient-year, with a true infection rate of 42% [286]. In some cases, Pjp may even be the sentinel manifestation of SCID [294]. SCID is fatal unless corrected early in life with hematopoietic stem cell transplantation (HSCT) or gene therapy (GT). In distinction to SCID, CID is survivable beyond infancy. While the associated defects may not be as profound as those in SCID, CID may also be associated with an increased risk of Pjp. Following the proposed criteria, the most frequently reported ones are described below.

HLA class II deficiency is an AR CID marked by the absence or greatly reduced expression of major histocompatibility class II molecules on the cell surface due to mutations in the MHC class II transactivator (CIITA), regulatory factor X-associated protein (RFXAP), regulatory factor X-5 (RFX5), and ankyrin-repeat-containing regulatory factor X (RFXANK) [295]. The major function of HLA class II molecules is to present antigens to T cells; hence, their expression is primarily restricted to thymic epithelial cells as well as professional antigen-presenting cells, namely, monocytes/macrophages, dendritic cells (DC), and B cells. Consequently, the loss of HLA class II function disturbs CD4<sup>+</sup> T helper cell development and renders cell-mediated immunity defective. Similarly, Wiskott–Aldrich syndrome (WAS), which occurs due to X-linked recessive mutations in WAS that compromise the actin cytoskeleton of hematopoietic cells, is marked by CID, eczema, and microthrombocytopenia [296]. Mutant WAS protein (WASp) interferes with proper migration and antigen uptake, immunological synapse formation between APC and T cells, and T cell proliferation. Hyper-IgM syndrome (HIGM) is caused by various genetic lesions of the CD40/CD154 (also known as the CD40 ligand (CD40L)) axis. Since this axis enables cross-talk between activated CD4<sup>+</sup> T lymphocytes (expressing CD40L) and B cells (constitutively expressing CD40), the loss of this function results in abnormal proliferation, class-switch recombination, and the somatic hypermutation of the latter lymphocytes, resulting in the impaired formation of antibody and memory cells [297]. Hence, patients classically have high IgM but low serum levels of IgG and IgA, leading to an increased risk for bacterial infections. Since the CD40/CD40L pathway also mediates co-stimulation between activated CD4<sup>+</sup> T cells and CD40-expressing monocytes, macrophages, and dendritic cells, mutations in either CD40L (X-linked) or in CD40 (AR) also lead to opportunistic infections, notably Pjp [257,298–310]. Similarly, nuclear factor-kappa-B essential modulator (*NEMO/IKK $\gamma$* ) [311–313], inhibitor of kappa light chain gene enhancer in B cells, alpha (*I $\kappa$ B $\alpha$* ) [314], or nuclear factor kappa-B subunit 1 (*NFKB1*) [315,316] are involved in CD40/CD40L signaling, and, when mutated, interfere with CD4<sup>+</sup> T cell/APC co-stimulation, resulting in HIGM syndrome, for which there are reports of Pjp. On the other hand, several other genes involved in CD40 signaling but acting in a B-cell-intrinsic manner (e.g., activation-induced cytidine deaminase (*AICDA*), uracil–DNA glycosylase (*UNG*), phosphatidylinositol 3-kinase catalytic delta (*PIK3CD*), and phosphatidylinositol 3-kinase regulatory subunit 1 alpha (*PIK3R1*)) may cause HIGM but have not been reported in association with Pjp. IEI involving the Ikaros Zinc Finger (IkZF) family of transcription factors, specifically dominant negative mutations (DN) in *IKZF1* (encoding IKAROS) or in *IKZF3* (encoding AIOLOS), confer increased susceptibility to Pjp. IkZF transcriptionally regulates hematopoietic development. DN-IKZF1 deficiency impairs lymphocyte development, wherein CD4<sup>+</sup> T cells that are primarily naïve and possess impaired function are developed [317]. Moreover, due to this deficiency, monocytes are dysfunctional [317]. DN-AIOLOS deficiency disrupts CD40/CD40L signaling [318]. These disorders suggest that the T–B cell interface (and its accompanying humoral immunity) is not critical for Pjp

host defense. Rather, it suggests that an airway CD4<sup>+</sup> T cell–APC interaction, or perhaps an airway CD4<sup>+</sup> T cell–alveolar epithelial cell interaction, is central. As with the identification of Th17 defects and CMC, perhaps a novel Th subset (in the circulation or in the alveolar space) may be discovered that could shed light on Pjp immunopathology.

Pjp has been reported in individual cases of GATA2 [319], XLA [320,321], and DOCK8 [322] but without any mechanistic evidence attributing the mutant gene to disease. If these observations consistently recur, they may provide much-needed granular data on the mechanisms of susceptibility.

## 7. Aspergillosis

Aspergillosis refers to a group of infections caused by molds of the genus *Aspergillus* and can be divided into three broad categories [323,324]: allergic bronchopulmonary aspergillosis (ABPA); invasive aspergillosis (IA); and chronic pulmonary aspergillosis (CPA). CPA refers to a heterogeneous group of conditions that are either indolent (>3 months) in seemingly immunocompetent individuals who have underlying structural lung disease (including ‘fungus ball’/aspergilloma, an isolated aspergillosis nodule, or chronic cavitary pulmonary aspergillosis (CCPA)) or progressive (worsening < 3 months) in moderately immunocompromised patients (i.e., subacute invasive aspergillosis (SAIA), also called chronic necrotizing pulmonary aspergillosis (CNPA)). It has been estimated that humans inhale at least several hundred *Aspergillus* spores daily [325]. Thus, the inhalation of conidia into the lower respiratory tract explains why the lungs are the primary site of disease. However, extra-pulmonary disease can occur, either as a contiguous extension or dissemination from a pulmonary portal of entry or, presumably, by inoculation that is localized and/or of a high burden (e.g., in the sinuses, gastrointestinal tract, skin, and soft tissue). Among the genetic disorders associated with these aspergillosis syndromes, ABPA is primarily seen with CFTR mutations, while IA and CPA are largely seen in chronic granulomatous disease (CGD), autosomal dominant hyper-IgE (Job’s) syndrome, GATA2 deficiency, and CARD9 deficiency.

### 7.1. Allergic Broncho-Pulmonary Aspergillosis (ABPA)

ABPA is a hypersensitivity reaction precipitated by *A. fumigatus* in the tracheo-bronchial tree. ABPA is seen in patients with cystic fibrosis (CF), an autosomal recessive disorder caused by mutations in the cystic fibrosis transmembrane conductance regulatory (CFTR) gene [326]. Impaired CFTR function leads to chloride ion/water imbalance, resulting in the classical syndrome of excessive mucous viscosity at the respiratory epithelium, pancreatic insufficiency, and fertility issues, as well as an abnormally elevated sweat chloride test. Intriguingly, only 1–15% of CF patients develop ABPA [327], even though *Aspergillus* spp. is cultivable from sputum in as much as 57% of patients [327]. ABPA can also be seen in asthmatic patients. As in CF, only a subset of patients with asthma develop ABPA, and this selectivity seems to be associated with the mono-allelic carriage of a CFTR mutation, with an estimated odds ratio of ~10 [327]. Collectively, these findings suggest that ABPA is a CFTR-related disorder [328]. This intra-individual variability of ABPA has led to the identification of additional genes, in the context of underlying lung disease, that are associated with an increased risk of ABPA [329–331]. While the mechanism(s) by which defective CFTR function contributes to an exuberant allergic inflammatory response is not completely defined, it is possible that abnormal mucous composition from mono- or bi-allelic CFTR mutations trap *Aspergillus* conidia, allowing them to persist beyond the time period in which they would normally be cleared [332]. In addition, the CFTR protein is also expressed in lymphocytes, and CD4<sup>+</sup> T cells bearing mutant CFTR demonstrate cytokine dysregulation with skewing towards a T helper type 2 (Th2) phenotype [333,334]. On the other hand, ICI with aberrantly excessive Th2 diathesis, such as CARD11, CARD14, CARML2, DOCK8, IKBKG, IL6R, and TBX21, have not (yet) been reported to lead to the development of ABPA, suggesting that Th2 skewing alone is not sufficient for the development of disease.

## 7.2. Invasive Aspergillosis and Chronic Pulmonary Aspergillosis

IA can be pulmonary, extra-pulmonary, or both. In reality, pulmonary disease spans the tempo spectrum of acute disease, SAIA, and CPA presented above, and these can all occur within the same patient, thereby discouraging the notion that they are mutually exclusive conditions or prototypical of one immunodeficiency compared to another.

Chronic granulomatous disease (CGD) is a primary phagocyte defect characterized by recurrent, life-threatening bacterial and fungal infections as well as dysregulated inflammation typically resulting in tissue granuloma formation. ‘Classical’ CGD is transmitted in either an X-linked (via *CYBB* encoding NOX2, previously known as gp91<sup>phox</sup>) or autosomal recessive (*NCF1*/p47<sup>phox</sup>; *NCF2*/p67<sup>phox</sup>; and *CYBA*/p22<sup>phox</sup>) manner [335]. Mutations in any of these subunits of the nicotinamide adenine dinucleotide phosphate (NAPDH) oxidase (phox) complex impair phagocyte respiratory burst and the generation of reactive oxygen species (ROS). Among IEI, CGD patients are distinctly susceptible to IA; at least 25–35% of these patients are affected [336–338]. IA in CGD can cause pulmonary disease. The clinical course can be chronic, spread in a contiguous fashion across anatomic planes, and is marked by the difficulty with it can be controlled with antifungals (the 3 “Cs”). Some of these features may be due to discrete species within the genus. For example, while *A. fumigatus* is considered the most common etiological agent of IA, it is not a singular species. Rather, moulds previously recognized as *A. fumigatus* now constitute a complex that is designated *Aspergillus* section *fumigati* subgenus *fumigate*. These sub-species (also referred to as “cryptic species”), in conjunction with host characteristics, may contribute to differences in clinical manifestations [336,339–342]. Additionally, CGD can lead to extra-pulmonary disease (e.g., paravertebral abscess, osteomyelitis, and CNS) [343–345]. Despite the severity of fungal disease, CGD patients may be a- or pauci-symptomatic, which is attributable to a lack of an inflammatory response, and may only be detected during screening exams (e.g., chest X-ray) or with a high index of clinical suspicion of non-specific symptoms.

CGD has some distinctive features of IA that are important. Firstly, CGD has unique susceptibility to infection with *A. nidulans*. Like *A. fumigatus*, *A. nidulans* is not recognized as representing a single species but rather a species complex; however, the impact of sub-species on the clinical trajectory of CGD is unclear [346]. Herein, they will be collectively referred to as the type species. *A. nidulans* is a poorly pathogenic mould that is better known for its use in studying genetics and cell biology or in the industrial synthesis of enzymes. However, in CGD, it is the second leading cause of IA, capable of causing both pulmonary and extra-pulmonary disease [347]. While the reason for its pathogenicity in CGD has long been enigmatic, the N-acetyl-galactosamine (GalNAc) content of this fungus’s exopolysaccharide, galactosaminogalactan (GAG), provides some insight [348]: the NADPH respiratory burst triggers the production of neutrophil extracellular traps (NETs), leading to hyphal damage and fungal killing. GAG confers mould resistance to NETs. *A. fumigatus* has abundant GAG expression, contributing to its resistance to neutrophil-mediated killing and thus its virulence. On the other hand, *A. nidulans* has greatly reduced GAG expression, owing to decreased GalNAc production, rendering it especially sensitive to the antifungal activity of NETs. The loss of the phagocyte oxidase complex in CGD leads to the loss of NET activity and the loss of *A. nidulans*’ non-pathogenicity. However, why *A. nidulans* is the second most common aspergilli causing disease in CGD [347], as opposed to other *Aspergillus* species, remains unclear; whether this simply reflects differences in environmental exposure (with *A. nidulans* being more ubiquitous than other species), or whether additional factors are at play, should be identified. A second distinguishing feature of IA in CGD is that serum galactomannan assays are insensitive in terms of diagnosing this fungal disease [349,350]; thus, a negative result does not exclude the possibility of IA (or moulds with cross-reactive galactomannans). Thirdly, in addition to CPA syndrome, CGD patients are also at risk for a fulminant syndrome called “mulch pneumonitis”, which is characterized by severe, acute, bilateral pneumonia with hypoxemia and respiratory failure and occurs following exposure to high levels of fungal spores from outdoor gardening

activities [351]. Despite the administration of antifungals, anti-inflammatory steroids, and critical care support (e.g., mechanical ventilation, extracorporeal membrane oxygenation, etc.), mortality can be high, but not invariably so [351–354]. Of note, in some patients, ‘mulch pneumonitis’ was the sentinel presentation of CGD [351].

The different genotypes underlying CGD predispose one to different risk levels for IA, and the residual activity of the encoded NADPH-oxidase complex (rather than the genotype per se) determines this risk [355]. Interestingly, ROS production may also be associated with risk for IA in a non-CGD, hematopoietic stem cell transplant patient population; this risk relates to a genomic SNP that is involved in ROS production but apparently not involved in the phagocyte NADPH-oxidase complex [356]. On the other hand, IEI that are partial phenocopies of CGD have been identified, such as p40<sup>phox</sup> deficiency (due to AR mutations in *NCF4*) [357] and protein kinase C  $\delta$  (PKC $\delta$  deficiency due to AR mutations in *PRKCD*) [358–360]. Regarding p40<sup>phox</sup> deficiency, patients have bacterial infections and hyperinflammation but no invasive fungal disease. The mutations underlying p40<sup>phox</sup> deficiency are LOF or hypomorphic, and they variably impair ROS production in neutrophils and monocytes (ranging from normal to decreased, but at higher levels than in CGD) and normal ROS production in monocyte-derived macrophages (moDM). PKC $\delta$  deficiency is primarily clinically marked by recurrent bacterial infections (involving the lungs, lymph nodes, and GI tract); no IA has yet been reported. In PKC $\delta$  deficiency, ROS production by neutrophils and monocytes mirrors that of p40<sup>phox</sup> deficiency, although in distinction to the latter, PKC $\delta$  deficient-moDM have very low levels of ROS production. On the other hand, MSMD due to hypomorphic mutation in *CYBB* (MSMD<sup>CYBB</sup>) manifests as an X-linked susceptibility to mycobacteria exclusively rather than the range of bacterial and fungal infections of CGD [361]. The hypomorphic mutations of MSMD<sup>CYBB</sup> molecularly distinguish it from the LOF *CYBB* mutations causing CGD. Moreover, at the cellular level, MSMD<sup>CYBB</sup> is characterized by normal respiratory burst in neutrophils or monocytes ex vivo, but impaired respiratory burst selectively in moDM and patient-derived EBV-immortalized lymphoblastoid cell lines (LCL). Collectively, these IEI indicate that cell-specific ROS generation in a fungal morphotype-specific manner may contribute to susceptibility to IA. When focused on the respiratory-epithelial interface, inhaled conidia are thought to initially encounter alveolar macrophages, which use (at least) the phagocyte NADPH oxidase complex to clear the spores. In reality, the cellular components of the alveoli include type I and II pneumocytes as well as alveolar macrophages, and the alveolar surface is primarily covered by type I pneumocytes (~90%) and type II pneumocytes (5–10%) [362]. Thus, the host cells that the inhaled conidia will most likely encounter first will be pneumocytes; indeed, in rats, such cells express the components of the phagocyte NADPH oxidase complex, with substantially higher levels of NOX2 (gp91<sup>phox</sup>), p22<sup>phox</sup>, p67<sup>phox</sup>, p47<sup>phox</sup>, and p40<sup>phox</sup> subunits as well as Rac1 in type I pneumocytes than in type II [363]. Although human bronchial epithelial cells generate ROS in response to *A. fumigatus* conidia [364], whether the phagocyte NADPH-oxidase complex is operational in pneumocytes’ defenses against aspergilli has not, to the best of our knowledge, been studied. However, if it were operational, it would be defective in ‘classic CGD’, owing to the germline nature of the mutations, but variably so—if not at all—in p40<sup>phox</sup> deficiency, PKC $\delta$  deficiency, and MSMD<sup>CYBB</sup>. It is potentially this junction that is critical in aspergilli pathogenesis. Failure to clear conidia (ostensibly by pneumocytes) results in their swelling and germination in the alveoli. Alveolar macrophages patrolling the airspace attempt to phagocytose and destroy the conidia. Local inflammatory responses recruit myeloid cells, particularly neutrophils, that are able to phagocytose resting and swollen conidia and kill mould hyphae (e.g., with NETs). When the phagocyte NADPH-oxidase complex is simultaneously defective in each of these components (pneumocytes, macrophages, and neutrophils), IA ensues. However, in IEI where there is a limited, cell-restricted defect in respiratory burst, IA does not transpire. Lastly, primary ciliary dyskinesia (PCD) is a heterogeneous group of inborn errors of respiratory cilia marked by recurrent airway infections and, occasionally, with airway colonization by *Aspergillus* and the development of

ABPA [365–368]. Strikingly, PCD patients have not shown increased rates of IA, suggesting that defective mucociliary clearance does not sufficiently impair anti-aspergilli immunity.

The high rate of IA in CGD patients led to the initial study on antifungal prophylaxis with itraconazole, demonstrating a reduction in rate of IA to one-third [80]. This effect was later confirmed in a randomized clinical trial [82]. Despite this, a breakthrough fungal disease had been observed, which, in some cases, was attributable to pharmacologic issues (e.g., adherence, absorption, etc.) [369]. Additionally, CGD patients are at risk for infection with other moulds, albeit less frequently, including other hyalohyphomycetes (e.g., *Pae-cilomyces* [370–372] and the morphologically similar *Rasamsonia* [373] and *Fusarium* [374]) and basidiomycete mushrooms (e.g., *Phellinus* [375–377]). Thus, some patients may receive voriconazole or posaconazole prophylaxis. The diversity of moulds capable of causing disease in CGD should prompt the need for tissue sampling for microbiological investigations. Owing to the refractory nature of IA or some of the other mycoses, granulocyte infusions have been attempted successfully [378–380], although there may be some reporting bias. Granulocyte infusions can also be given as a bridge to HSCT [381,382], although there is a risk of allo-immunization with subsequent graft failure [383], or given post-HSCT as a bridge to myeloid reconstitution [384]. HSCT with concomitant antifungal therapy, without granulocyte infusions, has also been successful [385].

Other IEI with a consistently reported risk for IA include DN-STAT3 (AD-HIES, or Job’s syndrome), IL6ST deficiency, CARD9 deficiency, and GATA2 deficiency. In DN-STAT3 (see the “Candidiasis” section presented above), IA is typically restricted to pulmonary disease in people with abnormal lung parenchyma (e.g., pneumatoceles, cysts, and bronchiectasis) [192,386–392]. This observation, along with the finding that myeloid cells from such patients equally inhibit *A. fumigatus* in vitro, suggests that, in this syndrome, STAT3 mutations confer susceptibility to IA through defects at the level of the lung epithelium [192]. Whether this reflects a defect in alveolar epithelial homeostasis (e.g., STAT3 regulates pneumocyte regeneration [393]) and/or a pneumocyte-intrinsic host defense defect remains to be fully elucidated. The IA of DN-STAT3 tends to behave in a manner similar to that of CPA, although lung erosion with pleural involvement, hemoptysis, or the dysfunction of lung function are serious concerns. Unfortunately, despite prolonged antifungal therapy and/or surgical resection, the clinical outcomes are sub-optimal [192,391]. Heterozygous, dominant negative mutations in IL6ST (DN-IL6ST) can also cause AD-HIES syndrome [394]. As with DN-STAT3, DN-IL6ST is associated with IA, manifesting primarily as a CPA, in the context of abnormal lung parenchyma. *IL6ST* encodes GP130, the shared transmembrane signalling subunit of the IL-6-receptor (IL6R) family. Intriguingly, two other IEI, AR IL6ST deficiency [395–397] and AR IL6R deficiency [398], can lead to clinical syndromes with phenotypic overlaps with that of DN-STAT3 or DN-IL6ST. However, IA has not been reported in the two former IEI; this may be due to the censorship of data, as these patients were diagnosed at a younger age than DN-STAT3 and with fewer pulmonary parenchyma abnormalities (in some patients, no anomalies were reported). In CARD9 deficiency (see the “Candidiasis” section presented above), both CPA [399] and extra-pulmonary disease without concomitant lung involvement (e.g., intra-abdominal, CNS, and cutaneous) [108,400] have been observed. As with sCNSc (see the “Candidiasis” section presented above), the extra-pulmonary aspergillosis of CARD9 deficiency is associated with a paucity of neutrophils on tissue histopathology, despite visible fungus, contributing to development of disease, although the underlying susceptibility at these unusual anatomical sites remains obscure. GATA2 deficiency has been reported in association with IA both prior to chemotherapy or HSCT for MDS/AML and, more commonly, during or following the execution of these treatment modalities [174,180,401,402]; as explained in the proposed criteria, the latter state renders the distinction between infection due to iatrogenesis vs. inherent susceptibility inconclusive. It may also be that GATA2 mutations in the context of these treatment modalities confer a heightened risk for IA relative to those who do not harbor mutations [403]. In the pre-chemotherapy/HSCT state, pulmonary aspergillosis can be seen, and the risk is particularly increased if there is pulmonary alveolar proteinosis

(PAP) [404]. Whether the IA reflects the impaired clearance of conidia due to accumulated proteinaceous surfactants or signifies impaired alveolar host defenses remains unclear.

While other IEI, particularly those implicating neutrophil dysfunction, may be considered to dispose one to the risk of IA, few reports fulfilling the above criteria have been identified. For example, previous reviews reported that congenital neutropenia or leukocyte adhesion deficiency predispose one to IA, and this has been reiterated in recent reviews. However, when probing the cited references to focus on published clinical cases, there is a dearth of such reports, which spans various IEI/neutrophil dysfunctions [405–407], or the data are published as registries [408] without the granularity of details to address the issues raised in the proposed criteria. Similarly, IA has been reported in STAT1 GOF, but with no details provided [47]. Thus, additional reports with relevant information are strongly needed to revisit these associations. Nonetheless, if these IEI are associated with IA (or, in the case of IEI marked by neutrophil dysfunction, with other IFD), such an association appears to occur relatively infrequently.

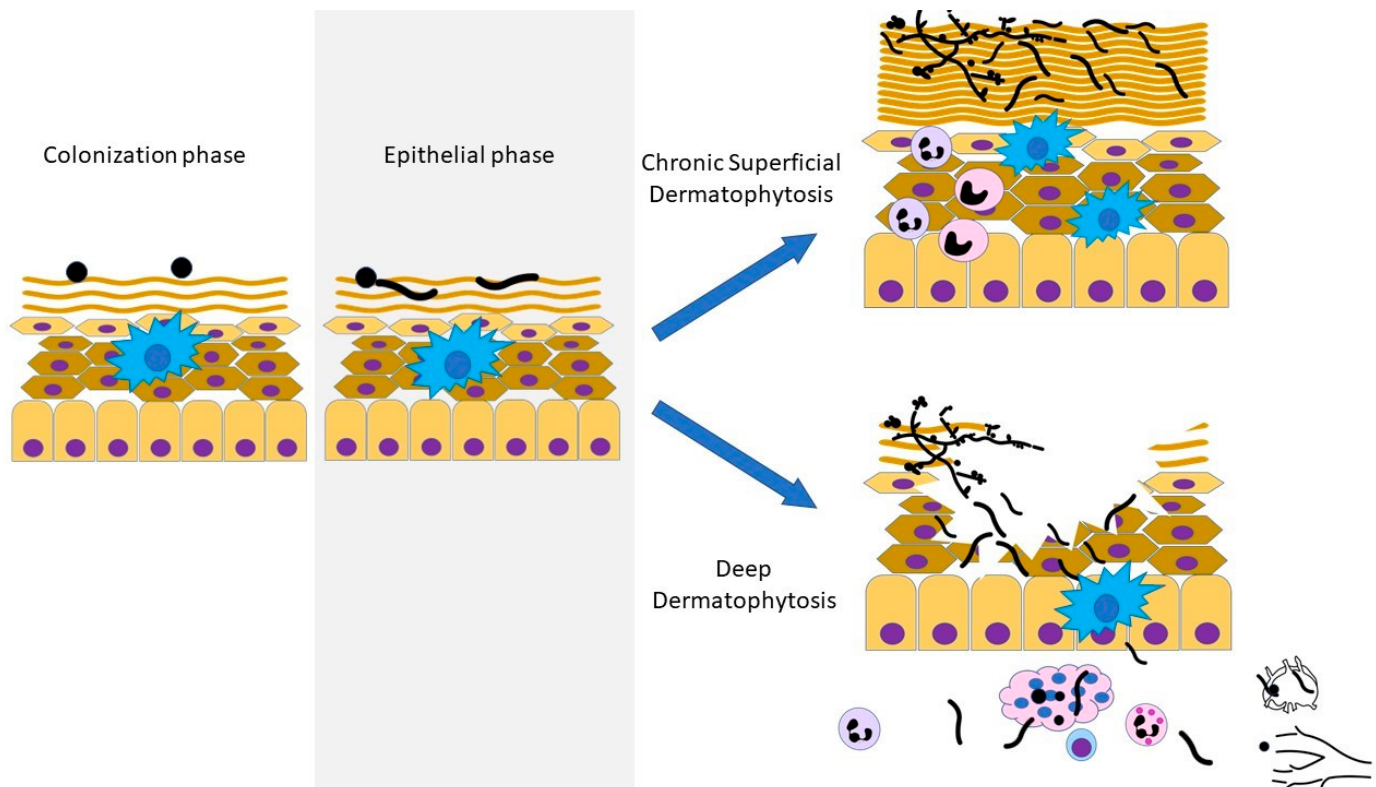
## 8. Dermatophytosis

Dermatophytes are hyalinated moulds of the Arthrodermataceae family with the capacity to invade keratinized tissue (i.e., skin, hair, and nails), causing “dermatophytosis” (colloquially referred to as “ringworm” or “tinea”). Dermatophytes were traditionally classified into three genera: *Trichophyton*, *Epidermophyton*, and *Microsporum*. Phylogenomic studies have since revised the taxonomy into seven genera: *Trichophyton*, *Epidermophyton*, and *Microsporum*, as well as *Paraphyton*, *Lophophyton*, *Arthroderma*, and *Nannizzia* [409,410]. Infection is acquired by the deposition of spores from humans, animals, or soil, accounting for anthrophilic, zoophilic, or geophilic spread, respectively. In immunocompetent individuals, infection is restricted to the stratum corneum, the outermost layer of the epidermis that contains a thickened layer of dead keratinocytes.

Despite their broad distribution and their affinity for a ubiquitous compound (keratin), dermatophytes are not professional pathogens. Previous serial sampling data have shown that these moulds can be isolated from humans for prolonged periods of time (months to years) in the absence of symptomatic infection [411]. Similarly, intra-familial transmission has been noted to be non-uniform. Thus, dermatophytes, like other microbes, demonstrate a spectrum of symptoms, ranging from asymptomatic infection and active disease to chronic disease. Chronic dermatophytosis has been previously defined as disease that is refractory to therapy, relapsing once treatment is stopped, and/or progressive [412]. Based on the genetic disorders described below, one could further separate ‘chronic dermatophytosis’ into two distinct chronic syndromes: one that manifests as a relapsing or recalcitrant, superficial disease (herein termed “chronic superficial dermatophytosis” [CSD]), and the other marked by superficial disease associated with invasion into deeper, non-keratinized tissue (termed “deep dermatophytosis” (DD)) (Figure 3).

As with CMC, genetic disorders of epithelial integrity predispose one to CSD. Although numerous cases of various ichthyosis disorders (e.g., ichthyosis vulgaris, lamellar ichthyosis, etc.) associated with dermatophytosis have been reported, dermatological diagnoses have been made clinically or, occasionally, via skin biopsy and without genetic confirmation [413–427]; following the proposed criteria above, these cases are not considered as definitive in this review. On the other hand, KID syndrome (due to dominant mutations in *GJB2*—which encodes Cx26—and conferring susceptibility to CMC (see above)) has also been associated with an increased risk for extensive CSD [10,12,19,428]. Some forms of palmoplantar keratoderma (PPK; also called ‘tylosis’) were also noted to be associated with increased rates of CSD [412,429]. PPK is a heterogeneous group of disorders characterized by the thickening (hyperkeratosis) of the palmar and plantar skin. Autosomal-dominant diffuse PPK can occur due to dominant negative mutations in keratin 9 (*KRT9*) [430], keratin 1 (*KRT1*) [431], or aquaporin 5 (*AQP5*) [432]. In previous studies from Norbotten, Sweden, the reported prevalence of associated dermatophytosis varied between 35 to 65% [433]. Keratins are a multi-gene family of proteins that form

intermediate filaments, which are the major cytoskeletal proteins in epithelial cells. Mutations in *KRT9*'s function through dominant negative interference with the assembly of these filaments and their specific contributions to PPK are due to the exclusive expression of KRT9 in the epidermal keratinocytes of the palms and soles [434]. It has been proposed that mutations in *KRT1* interfere with keratin intermediate filament interactions with other structures (e.g., desmosomes) or with the cross-linking of the cornified envelope [431]. AQP5 is expressed in the granular layer of the skin; PPK-causing GOF mutations disrupt keratinocyte water barrier function in the epidermis of the palms and soles [432,435]. Thus, selective defects in the barrier functions of the skin may confer susceptibility to CSD.



**Figure 3.** Host defense against dermatophytes. Dermatophytes can colonize keratinized surfaces. Disorders of keratin homeostasis may be associated with increased risk of developing chronic superficial dermatophytosis, wherein disease is limited to the skin. In rare instances, *CARD9* deficiency allows dermatophytes to permeate deeper structures, wherein systemic dissemination can also occur. See text for details.

CSD, which primarily occurs due to *Trichophyton* spp. or *Microsporon* spp., has also been reported in *STAT1* GOF [45,47]. This inborn error of immunity also predisposes one to CMC (see above). Whether this reflects the convergence of host defense pathways to mitigate these two superficial mycoses, or whether *STAT1* GOF has a distinct mechanism of susceptibility that implicates skin barrier function, remains to be explored.

Sentinel descriptions of chronic, progressively invasive dermatophytosis were made in the late 19th century when Majocchi described granulomatous dermal disease caused by *Trichophyton* [436]. Subsequently, rare cases were reported of chronic generalized trichophytosis involving the lymph nodes, muscles, bones, brain, or other viscera, including familial cases. One of the earliest documented cases of a family with fatal dermatophytosis was reported in 1975 involving a young Algerian girl who had extensive cutaneous disease at 14 years of age that progressed over 8 years [437]. She had two brothers, each of whom died at 11 and 13 years old, respectively, from a dermatophytic disease involving the skin, hair, and viscera that was caused by *T. schönleini*, which was also identified from her skin

lesions. Two other brothers were being treated for dermatophytosis (no further details were provided in this case), leading to the suspicion of a genetic immunodeficiency with susceptibility to dermatophytes. Further immunologic investigations at the time revealed intact humoral immunity but variably depressed cell-mediated immunity [438]. Indeed, Marill observed three key characteristics of what he termed “maladie dermatophytique” (French for “dermatophytic disease”) [439]: familial predisposition; deficiency of cell-mediated immunity; and unusual severity, with one patient developing a cerebral abscess and the other patients having died from the illness.

The breakthrough in understanding the underlying basis for this disorder came with the discovery of biallelic mutations in *CARD9* that were found in 17 patients of Algerian, Tunisian, or Moroccan descent with deep dermatophytosis (DD) [440]. Thus, AR *CARD9* deficiency not only predisposes one to CMC or extra-pulmonary aspergillosis (discussed above) but to DD as well. The vast majority of cases are caused by *Trichophyton* sp. (e.g., *T. rubrum*, *T. violaceum*, and *T. mentagrophytes*), although *Microsporum* sp. [441] may occasionally be the culprit. DD can be quite destructive, with painful, non-healing cutaneous ulcers that often fail to resolve with antifungal therapy or risk relapsing once antifungal therapy is stopped. Importantly, DD can also manifest with lymphadenitis or visceral abscesses, from which fungal elements can be seen on biopsy and the dermatophyte is cultivable from tissue.

The mechanism(s) by which *CARD9* mutations lead to seemingly disparate fungal diseases is not clear. Although *CARD9* LOF mutations do impair IL-17 responses, the absence of DD in other IEI marked by IL-17 deficiency (which manifest with CMC, as presented above), other than a singular report of DD in a patient harboring a DN-STAT3 mutation [442], implies that more specific immunologic mechanisms need to be deciphered to understand how *CARD9* mediates dermatophytic disease.

## 9. Pheohyphomycosis (Dematiaceous Fungi)

Pheohyphomycetes (dematiaceous fungi) are environmentally distributed moulds that are darkly pigmented due to the presence of melanin in the fungal cell wall. The host–fungal interaction can result in three human syndromes: eumycotic mycetoma, chromoblastomycosis, and phaeohyphomycosis [443]. Eumycotic mycetoma is a chronic cutaneous/subcutaneous infection characterized by the triad of ‘tumefaction’ (i.e., the indolent appearance of inflammatory nodules/mass with fibrosis); the penetration of sinuses or fistulae into deep tissue; and presence of granules in tissue or discharge [444]. Chromoblastomycosis is a chronic cutaneous/subcutaneous infection characterized by the presence of fungal muriform cells (also called ‘sclerotic bodies’ or ‘copper pennies’) in tissue. Pheohyphomycosis refers to infections caused by these dematiaceous fungi that are histologically distinct from the two other syndromes due to the presence of yeast-like cells and filamentous growth in tissue; additionally, disease is not necessarily restricted to cutaneous/subcutaneous organs but can be disseminated (including with the involvement of CNS). Importantly, the manifestation of the clinical syndrome resulting from the infection with these fungi is driven by the immune status of the patient.

The identification of dematiaceous fungi in a diagnostic microbiology laboratory traditionally involves the assessment of the sample fungus’s growth rate, its colony and microscopic morphological characteristics, and the execution of biochemical tests. Since different medium and culture conditions can lead to different morphologic forms for a given fungus, this approach has led to some confusion about nomenclature. Genomically based identification has emerged as a more accurate method of identification.

Superficial disease due to dematiaceous fungi is commonly restricted to global regions of warmer climates and lower latitudes. Unfortunately, this has hampered our understanding of underlying fungal pathophysiology. The classical paradigm has been that these fungi are widespread in the environment, including in soil and plants, and that trauma, even minor forms, account for the ensuing disease. Of course, this explanation is both simplistic and overlooks a clear knowledge gap: minor trauma must occur quite frequently in the

respective socio-economic context, yet disease does not. A single case of CARD9 deficiency causing chromoblastomycosis due to *Phialophora expansa* (within the *P. verrucosa* complex) has been reported [445]. Although an individual case, the authors elegantly buttressed this report with a mouse model replicating the disease and strongly suggested that CARD9 should be assessed in more cases of chromoblastomycosis.

Pheohyphomycosis has also been known for a long time but has remained enigmatic. Indeed, Marill was among the first to describe a “deep dematiaceous disease” due to *Hendersonula toruloidea* (a fungus at the time known to cause disease in fruit trees, which is now called *Neoscytalidium dimidiatum*) in an Algerian 30-year-old male who had, intriguingly, been diagnosed with a “deep dermatophytosis” at the age of 15 year and treated intermittently with griseofulvin [446]. Based on his investigations at the time, Marill suspected an immunological disorder. Nearly 35 years later, the first evidence to support his hypothesis was confirmed with the identification of AR CARD9 deficiency in four patients with pheohyphomycosis due to *P. verrucosa* [447]. This inborn error of immunity has since been identified in pheohyphomycosis due to *Exophiala* [448–451], other *Phialophora* species [452], *Aureobasidium* [453], *Alternaria* [454,455], and *Exserohilum* [456], as well as more esoteric dematiaceous fungi, including *Pallidocercospora crystallina*, *Ochroconis musae*, and *Corynespora cassiicola* [449,457–459]. Intriguingly, disseminated infection with *C. cassiicola* (with extension into the CNS) has also been reported in a patient with compound heterozygous mutations in *CLEC7A* (encoding Dectin-1, which is a fungal pattern recognition receptor that signals through CARD9) [460]. Other pheohyphomyces have also been reported with single heterozygous variants in *CLEC7A* or CARD9, although with distinct fungal etiologies than those for AR CARD9 deficiency [460]. Whether this distinction ostensibly reflects other genetic loci capable of influencing susceptibility to infection or severity to dematiaceous fungi or reflects inherent differences in fungal pathogenicity, or both, remains to be determined.

The management of pheohyphomycosis associated with AR CARD9 deficiency is quite heterogeneous, and clinical outcomes are not consistently reported. Various antifungals, either administered alone or in combination, have been used either as a sole therapy or in conjunction with surgical resection. This fungal disease is not invariably fatal, although relapses may occur. No reports of adjunctive cytokine therapy, as in AR CARD9 deficiency with invasive candidiasis (see above section), have been identified.

## 10. Mucormycosis

Fungi of the Mucorales order have aseptate (or pauci-septated), ribbon-like hyphae. The classical, acquired predisposing factors for infection with this fungus include neutropenia associated with chemotherapy for hematological malignancy, diabetes mellitus (particularly with poor glucose control and ketoacidosis), trauma, and iron chelation therapy. On the other hand, Mucormycosis has not been stereotypic or pathognomonic with respect to any single IEI. In CGD, spontaneous mucormycosis is exceedingly rare [343,461]. Most reported cases only occurred in patients receiving significant immunosuppression for several weeks [462], following HSCT [463,464], complicating anti-TNF- $\alpha$  treatment [465], or following invasive procedures [466,467]. However, an isolated case has been reported in the absence of immunosuppression [468]. The aggregate of these cases would suggest that defects in the phagocyte NADPH-oxidase system are not sufficient in terms of susceptibility to these moulds. There have also been reports of mucormycosis linked to AR CARD9 deficiency, wherein every case corresponded to exclusively cutaneous disease and was caused by *Mucor irregularis* (formerly *Rhizomucor variabilis* var. *variabilis*) [469–471]. Interestingly, *M. irregularis* is distinct among the Mucorales due to its tendency to cause chronic cutaneous/subcutaneous infection without angio-invasion and deeper dissemination, although the disease can be mutilating [472]. What is equally interesting is that all three reported CARD9 deficiency cases occurred in Chinese patients, and cases of the chronic skin disease caused by *M. irregularis* have exclusively been reported in Asia, particularly China [472], perhaps reflecting some aspect of selective evolutionary pressures. Mucormycosis has also

been singularly reported in association with GATA2 deficiency (although it is not clear if this case occurred during chemotherapy or HSCT) [473] and STAT1 GOF [474]. Clearly, more reports of native mucormycosis in IEL would significantly guide our understanding of human immunity to these moulds.

## 11. Conclusions

Fungal diseases reflect an underlying immunological deficiency. In the absence of an obvious medical condition or exogenous immunosuppression, patients with persistent superficial mycoses or those with invasive fungal disease should be evaluated for an underlying inborn error of immunity. Genomically based investigations combined with the functional validation of identified rare genetic variants and the characterization of their impacts on immunological functions and host–fungal interactions will further our understanding of human immunity against fungi. These insights will identify molecular targets that will pave the way for novel anti-fungal immunotherapies.

**Funding:** DCV is supported by the Senior Clinician-Scientist scholar program of the Fonds de Recherche du Québec-Santé (FRQS).

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** All data pertaining to the report are included within the manuscript.

**Conflicts of Interest:** None related to the manuscript. DCV has received clinical trial support from CSL Behring, Cidara Therapeutics, and Moderna; has received honoraria for advisory board consultations or speaker presentations from Astra Zeneca, CSL Behring, Merck Canada, Moderna, Novartis Canada, Qu biologics, and UCB Biosciences GmbH; and has a patent application pending (Electronic Filing System ID: 40101099) and a report of invention submitted to McGill University (Track code: D2021-0043).

## References

1. Bassetti, M.; Azoulay, E.; Kullberg, B.-J.; Ruhnke, M.; Shoham, S.; Vazquez, J.; Giacobbe, D.R.; Calandra, T. EORTC/MSGERC Definitions of Invasive Fungal Diseases: Summary of Activities of the Intensive Care Unit Working Group. *Clin. Infect. Dis.* **2021**, *72*, S121–S127. [\[CrossRef\]](#)
2. Begum, N.; Lee, S.; Portlock, T.J.; Pellon, A.; Nasab, S.D.S.; Nielsen, J.; Uhlen, M.; Moyes, D.L.; Shoaie, S. Integrative functional analysis uncovers metabolic differences between *Candida* species. *Commun. Biol.* **2022**, *5*, 1013. [\[CrossRef\]](#)
3. Marks, M.; Marks, S.; Brazeau, M. Yeast colonization in hospitalized and nonhospitalized children. *J. Pediatr.* **1975**, *87*, 524–527. [\[CrossRef\]](#)
4. Lay, K.M.; Russel, C. *Candida* species and yeasts in mouths of infants from a special care unit of a maternity hospital. *Arch. Dis. Child.* **1977**, *52*, 794–796. [\[CrossRef\]](#) [\[PubMed\]](#)
5. Ohmit, S.E.; Sobel, J.D.; Schuman, P.; Duerr, A.; Mayer, K.; Rompalo, A.; Klein, R.S. HIV Epidemiology Research Study (HERS) Group. Longitudinal Study of Mucosal *Candida* Species Colonization and Candidiasis among Human Immunodeficiency Virus (HIV)–Seropositive and At-Risk HIV–Seronegative Women. *J. Infect. Dis.* **2003**, *188*, 118–127. [\[CrossRef\]](#) [\[PubMed\]](#)
6. D’Enfert, C.; Kaune, A.-K.; Alaban, L.-R.; Chakraborty, S.; Cole, N.; Delavy, M.; Kosmala, D.; Marsaux, B.; Fróis-Martins, R.; Morelli, M.; et al. The impact of the Fungus–Host–Microbiota interplay upon *Candida albicans* infections: Current knowledge and new perspectives. *FEMS Microbiol. Rev.* **2020**, *45*, fuaa060. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Richard, G.; Rouan, F.; Willoughby, C.; Brown, N.; Chung, P.; Ryyänen, M.; Jabs, E.; Bale, S.J.; DiGiovanna, J.J.; Uitto, J.; et al. Missense Mutations in GJB2 Encoding Connexin-26 Cause the Ectodermal Dysplasia Keratitis–Ichthyosis–Deafness Syndrome. *Am. J. Hum. Genet.* **2002**, *70*, 1341–1348. [\[CrossRef\]](#)
8. Yotsumoto, S.; Hashiguchi, T.; Chen, X.; Ohtake, N.; Tomitaka, A.; Akamatsu, H.; Matsunaga, K.; Shiraishi, S.; Miura, H.; Adachi, J.; et al. Novel mutations in GJB2 encoding connexin-26 in Japanese patients with keratitis–ichthyosis–deafness syndrome. *Br. J. Dermatol.* **2003**, *148*, 649–653. [\[CrossRef\]](#)
9. Janecke, A.R.; Hennies, H.C.; Günther, B.; Gansl, G.; Smolle, J.; Messmer, E.M.; Utermann, G.; Rittinger, O. GJB2 mutations in keratitis–ichthyosis–deafness syndrome including its fatal form. *Am. J. Med Genet. Part A* **2005**, *133*, 128–131. [\[CrossRef\]](#)
10. Bygum, A.; Betz, R.; Kragballe, K.; Steiniche, T.; Peeters, N.; Wuyts, W.; Nöthen, M. KID Syndrome: Report of a Scandinavian Patient with Connexin-26 Gene Mutation. *Acta Derm.-Venereol.* **2005**, *85*, 152–155. [\[CrossRef\]](#)
11. Haruna, K.; Suga, Y.; Oizumi, A.; Mizuno, Y.; Endo, H.; Shimizu, T.; Hasegawa, T.; Ikeda, S. Severe form of keratitis–ichthyosis–deafness (KID) syndrome associated with septic complications. *J. Dermatol.* **2010**, *37*, 680–682. [\[CrossRef\]](#) [\[PubMed\]](#)

12. Lazic, T.; Li, Q.; Frank, M.; Uitto, J.; Zhou, L.H. Extending the Phenotypic Spectrum of Keratitis-Ichthyosis-Deafness Syndrome: Report of a Patient with GJB2 (G12R) Connexin 26 Mutation and Unusual Clinical Findings. *Pediatr. Dermatol.* **2012**, *29*, 349–357. [\[CrossRef\]](#)
13. Mazereeuw-Hautier, J.; Chiaverini, C.; Jonca, N.; Bieth, E.; Dreyfus, I.; Maza, A.; Cardot-Leccia, N.; Perrin, C.; Lacour, J. Lethal Form of Keratitis-Ichthyosis-Deafness Syndrome Caused by the GJB2 Mutation p.Ser17Phe. *Acta Derm.-Venereol.* **2014**, *94*, 591–592. [\[CrossRef\]](#)
14. Reinholz, M.; Hermans, C.; Dietrich, A.; Ruzicka, T.; Braun-Falco, M. A case of cutaneous vegetating candidiasis in a patient with Keratitis-Ichthyosis-Deafness Syndrome. *J. Eur. Acad. Dermatol. Venereol.* **2015**, *30*, 537–539. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Rerknimitr, P.; Tanizaki, H.; Otsuka, A.; Miyachi, Y.; Kabashima, K. Diminution of Langerhans cells in keratitis, ichthyosis and deafness (KID) syndrome patient with recalcitrant cutaneous candidiasis. *J. Eur. Acad. Dermatol. Venereol.* **2015**, *30*, e47–e49. [\[CrossRef\]](#)
16. Calderón-Castrat, X.; Vega-Zuñiga, J.; Velásquez, F.; Ballona, R. Vegetating Candidiasis: A Mimicker of Squamous Cell Carcinoma in Keratitis Ichthyosis Deafness Syndrome. *Pediatr. Dermatol.* **2017**, *34*, 201–203. [\[CrossRef\]](#) [\[PubMed\]](#)
17. Bartenstein, D.; Chung, H.J.; Hussain, S. Two Cases of Chronic Candidiasis in Keratitis-Ichthyosis-Deafness Syndrome. *Am. J. Dermatopathol.* **2018**, *40*, e138–e141. [\[CrossRef\]](#)
18. Miteva, L. Keratitis, ichthyosis, and deafness (KID) syndrome. *Pediatr. Dermatol.* **2002**, *19*, 513–516. [\[CrossRef\]](#)
19. Cogshall, K.; Farsani, T.; Ruben, B.; McCalmont, T.H.; Berger, T.G.; Fox, L.P.; Shinkai, K. Keratitis, ichthyosis, and deafness (KID) syndrome: A review of infectious and neoplastic complications. *J. Am. Acad. Dermatol.* **2013**, *69*, 127–134.e3. [\[CrossRef\]](#)
20. Lilly, E.; Sellitto, C.; Milstone, L.M.; White, T.W. Connexin channels in congenital skin disorders. *Semin. Cell Dev. Biol.* **2016**, *50*, 4–12. [\[CrossRef\]](#)
21. Van Campenhout, R.; Gomes, A.R.; De Groof, T.W.M.; Muyldermans, S.; Devoogdt, N.; Vinken, M. Mechanisms Underlying Connexin Hemichannel Activation in Disease. *Int. J. Mol. Sci.* **2021**, *22*, 3503. [\[CrossRef\]](#)
22. Taki, T.; Takeichi, T.; Sugiura, K.; Akiyama, M. Roles of aberrant hemichannel activities due to mutant connexin26 in the pathogenesis of KID syndrome. *Sci. Rep.* **2018**, *8*, 12824. [\[CrossRef\]](#)
23. Langer, K.; Konrad, K.; Wolff, K. Keratitis/ichthyosis and deafness (KID)-syndrome: Report of three cases and a review of the literature. *Br. J. Dermatol.* **1990**, *122*, 689–697. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Harms, M.; Gilardi, S.; Levy, P.M.; Saurat, J.H. KID Syndrome (Keratitis, Ichthyosis, and Deafness) and Chronic Mucocutaneous Candidiasis: Case Report and Review of the Literature. *Pediatr. Dermatol.* **1984**, *2*, 1–7. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Shiraishi, S.; Murakami, S.; Miki, Y. Oral fluconazole treatment of fungating candidiasis in the keratitis, ichthyosis and deafness (KID) syndrome. *Br. J. Dermatol.* **1994**, *131*, 904–907. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Pincus, S.H.; Thomas, I.T.; Clark, R.A.; Ochs, H.D. Defective neutrophil chemotaxis with variant ichthyosis, hyperimmunoglobulinemia E, and recurrent infections. *J. Pediatr.* **1975**, *87*, 908–911. [\[CrossRef\]](#)
27. Kashem, S.W.; Igyártó, B.Z.; Gerami-Nejad, M.; Kumamoto, Y.; Mohammed, J.; Jarrett, E.; Drummond, R.A.; Zurawski, S.M.; Zurawski, G.; Berman, J.; et al. Candida albicans Morphology and Dendritic Cell Subsets Determine T Helper Cell Differentiation. *Immunity* **2015**, *42*, 356–366. [\[CrossRef\]](#)
28. Patini, R.; Gioco, G.; Rupe, C.; Contaldo, M.; Serpico, R.; Giuliani, M.; Lajolo, C. Oral Candida and psoriasis: Is there association? A systematic review and trial sequential analysis. *Oral Dis.* **2022**. [\[CrossRef\]](#)
29. Alwan, W.; Nestle, F.O. Pathogenesis and treatment of psoriasis: Exploiting pathophysiological pathways for precision medicine. *Clin. Exp. Rheumatol.* **2015**, *33* (Suppl. 93), S2–S6.
30. Armstrong, A.W.; Read, C. Pathophysiology, Clinical Presentation, and Treatment of Psoriasis: A Review. *JAMA* **2020**, *323*, 1945–1960. [\[CrossRef\]](#)
31. Moyes, D.L.; Wilson, D.; Richardson, J.P.; Mogavero, S.; Tang, S.X.; Wernecke, J.; Höfs, S.; Gratacap, R.L.; Robbins, J.; Runglall, M.; et al. Candidalysin is a fungal peptide toxin critical for mucosal infection. *Nature* **2016**, *532*, 64–68. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Minegishi, Y.; Saito, M.; Tsuchiya, S.; Tsuge, I.; Takada, H.; Hara, T.; Kawamura, N.; Ariga, T.; Pasic, S.; Stojkovic, O.; et al. Dominant-negative mutations in the DNA-binding domain of STAT3 cause hyper-IgE syndrome. *Nature* **2007**, *448*, 1058–1062. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Holland, S.M.; Vinh, D.C. Yeast Infections—Human Genetics on the Rise. *N. Engl. J. Med.* **2009**, *361*, 1798–1801. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Richardson, J.P.; Moyes, D.L.; Ho, J.; Naglik, J.R. Candida innate immunity at the mucosa. *Semin. Cell Dev. Biol.* **2019**, *89*, 58–70. [\[CrossRef\]](#)
35. Huppler, A.R.; Verma, A.H.; Conti, H.R.; Gaffen, S.L. Neutrophils Do Not Express IL-17A in the Context of Acute Oropharyngeal Candidiasis. *Pathogens* **2015**, *4*, 559–572. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Aggor, F.E.Y.; Break, T.J.; Trevejo-Núñez, G.; Whibley, N.; Coleman, B.M.; Bailey, R.D.; Kaplan, D.H.; Naglik, J.R.; Shan, W.; Shetty, A.C.; et al. Oral epithelial IL-22/STAT3 signaling licenses IL-17-mediated immunity to oral mucosal candidiasis. *Sci. Immunol.* **2020**, *5*, eaba0570. [\[CrossRef\]](#)
37. Aggor, F.E.; Bertolini, M.; Zhou, C.; Taylor, T.C.; Abbott, D.A.; Musgrove, J.; Bruno, V.M.; Hand, T.W.; Gaffen, S.L. A gut-oral microbiome-driven axis controls oropharyngeal candidiasis through retinoic acid. *J. Clin. Invest.* **2022**, *7*. [\[CrossRef\]](#)

38. de Beaucoudrey, L.; Samarina, A.; Bustamante, J.; Cobat, A.; Boisson-Dupuis, S.; Feinberg, J.; Al-Muhsen, S.; Janni re, L.; Rose, Y.; de Suremain, M.; et al. Revisiting Human IL-12R 1 Deficiency. *Medicine* **2010**, *89*, 381–402. [\[CrossRef\]](#)
39. Prando, C.; Samarina, A.; Bustamante, J.; Boisson-Dupuis, S.; Cobat, A.; Picard, C.; AlSum, Z.; Al-Jumaah, S.; Al-Hajjar, S.; Frayha, H.; et al. Inherited IL-12p40 Deficiency. *Medicine* **2013**, *92*, 109–122. [\[CrossRef\]](#)
40. Okada, S.; Puel, A.; Casanova, J.-L.; Kobayashi, M. Chronic mucocutaneous candidiasis disease associated with inborn errors of IL-17 immunity. *Clin. Transl. Immunol.* **2016**, *5*, e114. [\[CrossRef\]](#)
41. B ziat, V.; Li, J.; Lin, J.-X.; Ma, C.S.; Li, P.; Bousfiha, A.; Pellier, I.; Zoghi, S.; Baris, S.; Keles, S.; et al. A recessive form of hyper-IgE syndrome by disruption of ZNF341-dependent STAT3 transcription and activity. *Sci. Immunol.* **2018**, *3*, eaat4956. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Sassi, A.; Lazaroski, S.; Wu, G.; Haslam, S.M.; Fliegau, M.; Mellouli, F.; Patiroglu, T.; Unal, E.; Ozdemir, M.A.; Jouhadi, Z.; et al. Hypomorphic homozygous mutations in phosphoglucomutase 3 (PGM3) impair immunity and increase serum IgE levels. *J. Allergy Clin. Immunol.* **2014**, *133*, 1410–1419.e13. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Winslow, A.; Jalazo, E.R.; Evans, A.; Winstead, M.; Moran, T. A De Novo Cause of PGM3 Deficiency Treated with Hematopoietic Stem Cell Transplantation. *J. Clin. Immunol.* **2022**, *42*, 691–694. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Fallahi, M.; Jamee, M.; Enayat, J.; Abdollahimajd, F.; Mesdaghi, M.; Khoddami, M.; Segarra-Roca, A.; Frohne, A.; Dmytrus, J.; Keramatipour, M.; et al. Novel PGM3 mutation in two siblings with combined immunodeficiency and childhood bullous pemphigoid: A case report and review of the literature. *Allergy Asthma Clin. Immunol.* **2022**, *18*, 111. [\[CrossRef\]](#)
45. van de Veerdonk, F.L.; Plantinga, T.S.; Hoischen, A.; Smeekens, S.P.; Joosten, L.A.; Gilissen, C.; Arts, P.; Rosentul, D.C.; Carmichael, A.J.; der Graaf, C.A.S.-V.; et al. STAT1 Mutations in Autosomal Dominant Chronic Mucocutaneous Candidiasis. *N. Engl. J. Med.* **2011**, *365*, 54–61. [\[CrossRef\]](#)
46. Liu, L.; Okada, S.; Kong, X.-F.; Kreins, A.Y.; Cypowyj, S.; Abhyankar, A.; Toubiana, J.; Itan, Y.; Audry, M.; Nitschke, P.; et al. Gain-of-function human STAT1 mutations impair IL-17 immunity and underlie chronic mucocutaneous candidiasis. *J. Exp. Med.* **2011**, *208*, 1635–1648. [\[CrossRef\]](#)
47. Toubiana, J.; Okada, S.; Hiller, J.; Oleastro, M.; Gomez, M.L.; Becerra, J.C.A.; Ouach e-Chardin, M.; Fouyssac, F.; Girisha, K.; Etzioni, A.; et al. Heterozygous STAT1 gain-of-function mutations underlie an unexpectedly broad clinical phenotype. *Blood* **2016**, *127*, 3154–3164. [\[CrossRef\]](#)
48. Okada, S.; Markle, J.G.; Deenick, E.K.; Mele, F.; Averbuch, D.; Lagos, M.; Alzahrani, M.; Al-Muhsen, S.; Halwani, R.; Ma, C.S.; et al. Impairment of immunity to Candida and Mycobacterium in humans with bi-allelic RORC mutations. *Science* **2015**, *349*, 606–613. [\[CrossRef\]](#)
49. Glocker, E.-O.; Hennigs, A.; Nabavi, M.; Sch ffer, A.A.; Woellner, C.; Salzer, U.; Pfeifer, D.; Veelken, H.; Warnatz, K.; Tahami, F.; et al. A Homozygous CARD9 Mutation in a Family with Susceptibility to Fungal Infections. *N. Engl. J. Med.* **2009**, *361*, 1727–1735. [\[CrossRef\]](#)
50. Drewniak, A.; Gazendam, R.P.; Tool, A.T.J.; van Houdt, M.; Jansen, M.H.; van Hamme, J.L.; van Leeuwen, E.M.M.; Roos, D.; Scalais, E.; de Beaufort, C.; et al. Invasive fungal infection and impaired neutrophil killing in human CARD9 deficiency. *Blood* **2013**, *121*, 2385–2392. [\[CrossRef\]](#)
51. Gavino, C.; Cotter, A.; Lichtenstein, D.; Lejtenyi, D.; Fortin, C.; Legault, C.; Alirezaie, N.; Majewski, J.; Sheppard, D.C.; Behr, M.A.; et al. CARD9 Deficiency and Spontaneous Central Nervous System Candidiasis: Complete Clinical Remission With GM-CSF Therapy. *Clin. Infect. Dis.* **2014**, *59*, 81–84. [\[CrossRef\]](#) [\[PubMed\]](#)
52. Gavino, C.; Hamel, N.; Bin Zeng, J.; Legault, C.; Guiot, M.-C.; Chankowsky, J.; Lejtenyi, D.; Lemire, M.; Alarie, I.; Dufresne, S.; et al. Impaired RASGRF1/ERK-mediated GM-CSF response characterizes CARD9 deficiency in French-Canadians. *J. Allergy Clin. Immunol.* **2015**, *137*, 1178–1188.e7. [\[CrossRef\]](#)
53. Corvilain, E.; Casanova, J.-L.; Puel, A. Inherited CARD9 Deficiency: Invasive Disease Caused by Ascomycete Fungi in Previously Healthy Children and Adults. *J. Clin. Immunol.* **2018**, *38*, 656–693. [\[CrossRef\]](#)
54. Puel, A.; Cypowyj, S.; Bustamante, J.; Wright, J.F.; Liu, L.; Lim, H.K.; Migaud, M.; Israel, L.; Chrabieh, M.; Audry, M.; et al. Chronic Mucocutaneous Candidiasis in Humans with Inborn Errors of Interleukin-17 Immunity. *Science* **2011**, *332*, 65–68. [\[CrossRef\]](#)
55. Fellmann, F.; Angelini, F.; Wassenberg, J.; Perreau, M.; Arenas-Ramirez, N.; Simon, G.; Boyman, O.; Demaria, O.; Christen-Zaech, S.; Hohl, D.; et al. IL-17 receptor A and adenosine deaminase 2 deficiency in siblings with recurrent infections and chronic inflammation. *J. Allergy Clin. Immunol.* **2015**, *137*, 1189–1196.e2. [\[CrossRef\]](#)
56. L vy, R.; Okada, S.; B ziat, V.; Moriya, K.; Liu, C.; Chai, L.Y.A.; Migaud, M.; Hauck, F.; Al Ali, A.; Cyrus, C.; et al. Genetic, immunological, and clinical features of patients with bacterial and fungal infections due to inherited IL-17RA deficiency. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E8277–E8285. [\[CrossRef\]](#) [\[PubMed\]](#)
57. Frede, N.; Rojas-Restrepo, J.; de Oteyza, A.C.G.; Buchta, M.; H bscher, K.; G mez-D az, L.; Proietti, M.; Saghabi, S.; Chavoshzadeh, Z.; Soler-Palacin, P.; et al. Genetic Analysis of a Cohort of 275 Patients with Hyper-IgE Syndromes and/or Chronic Mucocutaneous Candidiasis. *J. Clin. Immunol.* **2021**, *41*, 1804–1838. [\[CrossRef\]](#)
58. Ling, Y.; Cypowyj, S.; Aytekin, C.; Galicchio, M.; Camcioglu, Y.; Nepesov, S.; Ikinogullari, A.; Dogu, F.; Belkadi, A.; Levy, R.; et al. Inherited IL-17RC deficiency in patients with chronic mucocutaneous candidiasis. *J. Exp. Med.* **2015**, *212*, 619–631. [\[CrossRef\]](#) [\[PubMed\]](#)

59. Boisson, B.; Wang, C.; Pedergrana, V.; Wu, L.; Cypowyj, S.; Rybojad, M.; Belkadi, A.; Picard, C.; Abel, L.; Fieschi, C.; et al. An ACT1 Mutation Selectively Abolishes Interleukin-17 Responses in Humans with Chronic Mucocutaneous Candidiasis. *Immunity* **2013**, *39*, 676–686. [\[CrossRef\]](#)
60. Bhattad, S.; Dinakar, C.; Pinnamaraju, H.; Ganapathy, A.; Mannan, A. Chronic Mucocutaneous Candidiasis in an Adolescent Boy Due to a Novel Mutation in TRAF3IP2. *J. Clin. Immunol.* **2019**, *39*, 596–599. [\[CrossRef\]](#)
61. Shafer, S.; Yao, Y.; Comrie, W.; Cook, S.; Zhang, Y.; Yesil, G.; Karakoç-Aydiner, E.; Baris, S.; Cokugras, H.; Aydemir, S.; et al. Two patients with chronic mucocutaneous candidiasis caused by TRAF3IP2 deficiency. *J. Allergy Clin. Immunol.* **2020**, *148*, 256–261.e2. [\[CrossRef\]](#) [\[PubMed\]](#)
62. Marujo, F.; Pelham, S.J.; Freixo, J.; Cordeiro, A.I.; Martins, C.; Casanova, J.-L.; Lei, W.-T.; Puel, A.; Neves, J.F. A Novel TRAF3IP2 Mutation Causing Chronic Mucocutaneous Candidiasis. *J. Clin. Immunol.* **2020**, *41*, 1376–1379. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Lobo, P.B.; Lei, W.; Pelham, S.J.; Hernández, P.G.; Villaoslada, I.; de Felipe, B.; Lucena, J.M.; Casanova, J.; Olbrich, P.; Puel, A.; et al. Biallelic TRAF3IP2 variants causing chronic mucocutaneous candidiasis in a child harboring a STAT1 variant. *Pediatr. Allergy Immunol.* **2021**, *32*, 1804–1812. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Li, J.; Ritelli, M.; Ma, C.S.; Rao, G.; Habib, T.; Corvilain, E.; Bougarn, S.; Cypowyj, S.; Grodecká, L.; Lévy, R.; et al. Chronic mucocutaneous candidiasis and connective tissue disorder in humans with impaired JNK1-dependent responses to IL-17A/F and TGF- $\beta$ . *Sci. Immunol.* **2019**, *4*, eaax7965. [\[CrossRef\]](#) [\[PubMed\]](#)
65. Oikonomou, V.; Break, T.J.; Gaffen, S.L.; Moutsopoulos, N.M.; Lionakis, M.S. Infections in the monogenic autoimmune syndrome APECED. *Curr. Opin. Immunol.* **2021**, *72*, 286–297. [\[CrossRef\]](#)
66. Philippot, Q.; Casanova, J.-L.; Puel, A. Candidiasis in patients with APS-1: Low IL-17, high IFN- $\gamma$ , or both? *Curr. Opin. Immunol.* **2021**, *72*, 318–323. [\[CrossRef\]](#)
67. Naglik, J.R.; König, A.; Hube, B.; Gaffen, S.L. Candida albicans–epithelial interactions and induction of mucosal innate immunity. *Curr. Opin. Microbiol.* **2017**, *40*, 104–112. [\[CrossRef\]](#)
68. Gaffen, S.L.; Moutsopoulos, N.M. Regulation of host-microbe interactions at oral mucosal barriers by type 17 immunity. *Sci. Immunol.* **2020**, *5*, eaau4594. [\[CrossRef\]](#)
69. Baker, R.D. Leukopenia and Therapy in Leukemia as Factors Predisposing to Fatal Mycoses: Mucormycosis, Aspergillosis, and Cryptococcosis. *Am. J. Clin. Pathol.* **1962**, *37*, 358–373. [\[CrossRef\]](#)
70. Bodey, G.P.; Buckley, M.; Sathe, Y.S.; Freireich, E.J. Quantitative Relationships Between Circulating Leukocytes and Infection in Patients with Acute Leukemia. *Ann. Intern. Med.* **1966**, *64*, 328–340. [\[CrossRef\]](#)
71. Bodey, G.P. Fungal infections complicating acute leukemia. *J. Chronic Dis.* **1966**, *19*, 667–687. [\[CrossRef\]](#)
72. Donadieu, J.; Beaupain, B.; Fenneteau, O.; Bellanné-Chantelot, C. Congenital neutropenia in the era of genomics: Classification, diagnosis, and natural history. *Br. J. Haematol.* **2017**, *179*, 557–574. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Anderson, D.C.; Springer, T.A. LEUKOCYTE ADHESION DEFICIENCY: An Inherited Defect in the Mac-1, LFA-1, and p150,95 Glycoproteins. *Annu. Rev. Med.* **1987**, *38*, 175–194. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Harris, E.S.; Shigeoka, A.O.; Li, W.; Adams, R.H.; Prescott, S.M.; McIntyre, T.M.; Zimmerman, G.A.; Lorant, D.E. A novel syndrome of variant leukocyte adhesion deficiency involving defects in adhesion mediated by  $\beta$ 1 and  $\beta$ 2 integrins. *Blood* **2001**, *97*, 767–776. [\[CrossRef\]](#)
75. Engel, M.E.; Hickstein, D.D.; Bauer, T.R.; Calder, C.; Manes, B.; Frangoul, H. Matched unrelated bone marrow transplantation with reduced-intensity conditioning for leukocyte adhesion deficiency. *Bone Marrow Transplant.* **2006**, *37*, 717–718. [\[CrossRef\]](#)
76. Cox, D.P.; Weathers, D.R. Leukocyte adhesion deficiency type 1: An important consideration in the clinical differential diagnosis of prepubertal periodontitis. A case report and review of the literature. *Oral Surgery Oral Med. Oral Pathol. Oral Radiol. Endodontology* **2008**, *105*, 86–90. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Vasconcelos, D.D.M.; Beitler, B.; Martinez, G.A.; Pereira, J.; Filho, J.U.A.; Klautau, G.; Lian, Y.C.; Della Negra, M.; Duarte, A.J.D.S. CD18 deficiency evolving to megakaryocytic (M7) acute myeloid leukemia: Case report. *Blood Cells Mol. Dis.* **2014**, *53*, 180–184. [\[CrossRef\]](#)
78. Serwas, N.K.; Huemer, J.; Dieckmann, R.; Mejsstrikova, E.; Garnarcz, W.; Litzman, J.; Hoeger, B.; Zapletal, O.; Janda, A.; Bennett, K.L.; et al. CEBPE-Mutant Specific Granule Deficiency Correlates With Aberrant Granule Organization and Substantial Proteome Alterations in Neutrophils. *Front. Immunol.* **2018**, *9*, 588. [\[CrossRef\]](#)
79. Borregaard, N.; Sørensen, O.E.; Theilgaard-Mönch, K. Neutrophil granules: A library of innate immunity proteins. *Trends Immunol.* **2007**, *28*, 340–345. [\[CrossRef\]](#)
80. Mouy, R.; Veber, F.; Blanche, S.; Donadieu, J.; Brauner, R.; Levron, J.-C.; Griscelli, C.; Fischer, A. Long-term itraconazole prophylaxis against Aspergillus infections in thirty-two patients with chronic granulomatous disease. *J. Pediatr.* **1994**, *125*, 998–1003. [\[CrossRef\]](#)
81. Petropoulou, T.; Liese, J.; Tintelnot, K.; Gahr, M.; Belohradsky, B.H. Long-term treatment of patients with itraconazole for the prevention of Aspergillus infections in patients with chronic granulomatous disease (CGD). *Mycoses* **1994**, *37* (Suppl. 2), 64–69. [\[PubMed\]](#)
82. Gallin, J.I.; Alling, D.W.; Malech, H.L.; Wesley, R.; Koziol, D.; Marciano, B.; Eisenstein, E.M.; Turner, M.L.; DeCarlo, E.S.; Starling, J.M.; et al. Itraconazole to Prevent Fungal Infections in Chronic Granulomatous Disease. *N. Engl. J. Med.* **2003**, *348*, 2416–2422. [\[CrossRef\]](#)

83. Vardakas, K.Z.; Michalopoulos, A.; Falagas, M.E. Fluconazole versus itraconazole for antifungal prophylaxis in neutropenic patients with haematological malignancies: A meta-analysis of randomised-controlled trials. *Br. J. Haematol.* **2005**, *131*, 22–28. [[CrossRef](#)] [[PubMed](#)]
84. Robenshtok, E.; Gafter-Gvili, A.; Goldberg, E.; Weinberger, M.; Yeshurun, M.; Leibovici, L.; Paul, M. Antifungal Prophylaxis in Cancer Patients After Chemotherapy or Hematopoietic Stem-Cell Transplantation: Systematic Review and Meta-Analysis. *J. Clin. Oncol.* **2007**, *25*, 5471–5489. [[CrossRef](#)]
85. Wang, J.; Zhan, P.; Zhou, R.; Xu, J.; Shao, X.; Yang, Y.; Ouyang, J. Prophylaxis with itraconazole is more effective than prophylaxis with fluconazole in neutropenic patients with hematological malignancies: A meta-analysis of randomized-controlled trials. *Med. Oncol.* **2009**, *27*, 1082–1088. [[CrossRef](#)] [[PubMed](#)]
86. Ethier, M.; Science, M.; Beyene, J.; Briel, M.; Lehnbecher, T.; Sung, L. Mould-active compared with fluconazole prophylaxis to prevent invasive fungal diseases in cancer patients receiving chemotherapy or haematopoietic stem-cell transplantation: A systematic review and meta-analysis of randomised controlled trials. *Br. J. Cancer* **2012**, *106*, 1626–1637. [[CrossRef](#)] [[PubMed](#)]
87. Khoo, B.; Cho, C.; Smith, S.; Dudding, B. Cervical lymphadenitis due to *Candida albicans* infection. *J. Pediatr.* **1975**, *86*, 812–813. [[CrossRef](#)]
88. Orson, J.M.; Greco, R.G. Death from Torulopsis in chronic granulomatous disease. *J. Pediatr.* **1975**, *87*, 333. [[CrossRef](#)]
89. Cohen, M.S.; Isturiz, R.E.; Malech, H.; Root, R.K.; Wilfert, C.M.; Gutman, L.; Buckley, R.H. Fungal infection in chronic granulomatous disease: The importance of the phagocyte in defense against fungi. *Am. J. Med.* **1981**, *71*, 59–66. [[CrossRef](#)]
90. Fleischmann, J.; Church, J.A.; Lehrer, R.I. Case Report: Primary *Candida* Meningitis Chronic Granulomatous Disease. *Am. J. Med. Sci.* **1986**, *291*, 334–341. [[CrossRef](#)]
91. Levy, O.; Bourquin, J.-P.; McQueen, A.; Cantor, A.B.; Lachenauer, C.; Malley, R. Fatal disseminated *Candida lusitanae* infection in an infant with chronic granulomatous disease. *Pediatr. Infect. Dis. J.* **2002**, *21*, 262–264. [[CrossRef](#)] [[PubMed](#)]
92. Estrada, B.; Mancao, M.Y.; Polski, J.M.; Figarola, M.S. *Candida lusitanae* and Chronic Granulomatous Disease. *Pediatr. Infect. Dis. J.* **2006**, *25*, 758–759. [[CrossRef](#)]
93. Piwoz, J.A.; Stadtmauer, G.J.; Bottone, E.J.; Weitzman, I.; Shlasko, E.; Cunningham-Rundles, C. Trichosporon inkin lung abscesses presenting as a penetrating chest wall mass. *Pediatr. Infect. Dis. J.* **2000**, *19*, 1025–1027. [[CrossRef](#)]
94. Wynne, S.M.; Kwon-Chung, K.J.; Shea, Y.R.; Filie, A.C.; Varma, A.; Lupo, P.; Holland, S.M. Invasive infection with *Trichosporon inkin* in 2 siblings with chronic granulomatous disease. *J. Allergy Clin. Immunol.* **2004**, *114*, 1418–1424. [[CrossRef](#)] [[PubMed](#)]
95. Kenney, R.T.; Kwon-Chung, K.J.; Witebsky, F.G.; Melnick, D.A.; Malech, H.L.; Gallin, J.I. Invasive Infection with *Sarcinospore inkin* in a Patient with Chronic Granulomatous Disease. *Am. J. Clin. Pathol.* **1990**, *94*, 344–350. [[CrossRef](#)] [[PubMed](#)]
96. Mussa, A.Y.; Singh, V.K.; Randhawa, U.S.; Khan, Z.U. Disseminated fatal trichosporonosis: First case due to *Trichosporon inkin*. *J. Mycol. Médicale* **1998**, *8*, 196–199.
97. Hajjar, J.; Restrepo, A.; Javeri, H.; Wiederhold, N.P.; Papanastassiou, A.M.; Patterson, T.F. Multiple Brain Abscesses Caused by *Trichosporon inkin* in a Patient with X-Linked Chronic Granulomatous Disease (CGD) Successfully Treated with Antifungal Therapy. *J. Clin. Immunol.* **2017**, *37*, 519–523. [[CrossRef](#)]
98. Parrillo, O.J. Disseminated mycotic disease. *J. Am. Med. Assoc.* **1950**, *144*, 747–749. [[CrossRef](#)] [[PubMed](#)]
99. Morris, A.A.; Kalz, G.G.; Lotspeich, E.S. Ependymitis and meningitis due to *Candida* (monilia) *albicans*. *Arch. Neurol. Psychiatry* **1945**, *54*, 361–366. [[CrossRef](#)]
100. Bélisle, G.; Lachance, W.; Leblanc, G. Meningitis caused by *Candida albicans*. Report of a case and discussion. *L'union Med. Can.* **1968**, *97*, 710–715.
101. Black, J.T. Cerebral candidiasis: Case report of brain abscess secondary to *Candida albicans*, and review of literature. *J. Neurol. Neurosurg. Psychiatry* **1970**, *33*, 864–870. [[CrossRef](#)] [[PubMed](#)]
102. Germain, M.; Gourdeau, M.; Hébert, J. Familial Chronic Mucocutaneous Candidiasis Complicated by Deep *Candida* Infection. *Am. J. Med. Sci.* **1994**, *307*, 282–283. [[CrossRef](#)] [[PubMed](#)]
103. Borha, A.; Parienti, J.-J.; Emery, E.; Coskun, O.; Khouri, S.; Derlon, J.-M. Granulome cérébral à *Candida albicans* chez un patient immunocompétent. Cas clinique. *Neurochirurgie* **2009**, *55*, 57–62. [[CrossRef](#)]
104. Dalliere, C.C.; Thouvenot, E.; Baptista, G.; Le Moing, V.; Charif, M. Méningomyéloradiculite à *Candida albicans* chez un sujet immunocompétent. *Rev. Neurol.* **2010**, *166*, 741–744. [[CrossRef](#)]
105. Hoarau, G.; Kerdraon, O.; Lagree, M.; Vinchon, M.; François, N.; Dubos, F.; Sendid, B. Detection of (1,3)- $\beta$ -D-glucans in situ in a *Candida albicans* brain granuloma. *J. Infect.* **2013**, *67*, 622–624. [[CrossRef](#)] [[PubMed](#)]
106. Lanternier, F.; Mahdavian, A.; Barbat, E.; Chaussade, H.; Koumar, Y.; Levy, R.; Denis, B.; Brunel, A.-S.; Martin, S.; Loop, M.; et al. Inherited CARD9 deficiency in otherwise healthy children and adults with *Candida* species-induced meningoencephalitis, colitis, or both. *J. Allergy Clin. Immunol.* **2015**, *135*, 1558–1568.e2. [[CrossRef](#)]
107. Jones, N.; Garcez, T.; Newman, W.; Denning, D. Endogenous *Candida* endophthalmitis and osteomyelitis associated with CARD9 deficiency. *BMJ Case Rep.* **2016**, *2016*, bcr2015214117. [[CrossRef](#)]
108. Rieber, N.; Gazendam, R.P.; Freeman, A.F.; Hsu, A.P.; Collar, A.L.; Sugui, J.A.; Drummond, R.A.; Rongkavilit, C.; Hoffman, K.; Henderson, C.; et al. Extrapulmonary *Aspergillus* infection in patients with CARD9 deficiency. *J. Clin. Investig.* **2016**, *1*, e89890. [[CrossRef](#)]
109. Quan, C.; Li, X.; Shi, R.; Zhao, X.; Xu, H.; Wang, B.; Wang, X.; Hu, W.; Cao, H.; Zheng, J. Recurrent fungal infections in a Chinese patient with CARD 9 deficiency and a review of 48 cases. *Br. J. Dermatol.* **2018**, *180*, 1221–1225. [[CrossRef](#)]

110. Drummond, R.; Collar, A.; Swamydas, M.; Rodriguez, C.A.; Lim, J.K.; Mendez, L.M.; Fink, D.L.; Hsu, A.P.; Zhai, B.; Karauzum, H.; et al. CARD9-Dependent Neutrophil Recruitment Protects against Fungal Invasion of the Central Nervous System. *PLoS Pathog.* **2015**, *11*, e1005293. [\[CrossRef\]](#)
111. Martino, R.; Sureda, A.; Brunet, S. Disseminated Candidiasis in Patients with Acute Leukemia. *Clin. Infect. Dis.* **1998**, *26*, 245–246. [\[CrossRef\]](#)
112. Pagano, L.; Mele, L.; Fianchi, L.; Melillo, L.; Martino, B.; D’Antonio, D.; Tosti, M.E.; Posteraro, B.; Sanguinetti, M.; Trapè, G.; et al. Chronic disseminated candidiasis in patients with hematologic malignancies. Clinical features and outcome of 29 episodes. *Haematologica* **2002**, *87*, 535–541.
113. Duffy, D.; Rouilly, V.; Libri, V.; Hasan, M.; Beitz, B.; David, M.; Urrutia, A.; Bisiaux, A.; LaBrie, S.T.; Dubois, A.; et al. Functional Analysis via Standardized Whole-Blood Stimulation Systems Defines the Boundaries of a Healthy Immune Response to Complex Stimuli. *Immunity* **2014**, *40*, 436–450. [\[CrossRef\]](#) [\[PubMed\]](#)
114. Smeekens, S.P.; Ng, A.; Kumar, V.; Johnson, M.D.; Plantinga, T.S.; van Diemen, C.; Arts, P.; Verwiel, E.T.P.; Gresnigt, M.S.; Fransen, K.; et al. Functional genomics identifies type I interferon pathway as central for host defense against *Candida albicans*. *Nat. Commun.* **2013**, *4*, 1342. [\[CrossRef\]](#)
115. Celmeli, F.; Oztoprak, N.; Turkkahraman, D.; Seyman, D.; Mutlu, E.; Frede, N.; Köksoy, S.; Grimbacher, B. Successful Granulocyte Colony-stimulating Factor Treatment of Relapsing *Candida albicans* Meningoencephalitis Caused by CARD9 Deficiency. *Pediatr. Infect. Dis. J.* **2016**, *35*, 428–431. [\[CrossRef\]](#) [\[PubMed\]](#)
116. Du, B.; Shen, N.; Hu, J.; Tao, Y.; Mo, X.; Cao, Q. Complete clinical remission of invasive *Candida* infection with CARD9 deficiency after G-CSF treatment. *Comp. Immunol. Microbiol. Infect. Dis.* **2020**, *70*, 101417. [\[CrossRef\]](#) [\[PubMed\]](#)
117. Queiroz-Telles, F.; Mercier, T.; Maertens, J.; Sola, C.B.S.; Bonfim, C.; Lortholary, O.; Constantino-Silva, R.M.N.; Schrijvers, R.; Hagen, F.; Meis, J.F.; et al. Successful Allogeneic Stem Cell Transplantation in Patients with Inherited CARD9 Deficiency. *J. Clin. Immunol.* **2019**, *39*, 462–469. [\[CrossRef\]](#)
118. Erman, B.; Firtina, S.; Aksoy, B.A.; Aydogdu, S.; Genç, G.E.; Doğan, Ö.; Bozkurt, C.; Fışgın, T.; Çipe, F.E. Invasive *Saprochaete capitata* Infection in a Patient with Autosomal Recessive CARD9 Deficiency and a Review of the Literature. *J. Clin. Immunol.* **2020**, *40*, 466–474. [\[CrossRef\]](#)
119. Sari, S.; Dalgic, B.; Muehlenbachs, A.; DeLeon-Carnes, M.; Goldsmith, C.S.; Ekin, O.; Jain, D.; Keating, M.K.; Vilarinho, S. *Prototheca zopfii* Colitis in Inherited CARD9 Deficiency. *J. Infect. Dis.* **2018**, *218*, 485–489. [\[CrossRef\]](#)
120. Lockhart, S.; Toda, M.; Benedict, K.; Caceres, D.; Litvintseva, A. Endemic and Other Dimorphic Mycoses in The Americas. *J. Fungi* **2021**, *7*, 151. [\[CrossRef\]](#)
121. Rodrigues, A.M.; Hagen, F.; Puccia, R.; Hahn, R.C.; de Camargo, Z.P. *Paracoccidioides* and *Paracoccidioidomycosis* in the 21st Century. *Mycopathologia* **2023**, 1–5. [\[CrossRef\]](#) [\[PubMed\]](#)
122. Wang, F.; Han, R.; Chen, S. An Overlooked and Underrated Endemic Mycosis—*Talaromyces marneffei*. *Clin. Microbiol. Rev.* **2023**, e00051–22. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Kirkland, T.N.; Fierer, J. *Coccidioides immitis* and *posadasii*; A review of their biology, genomics, pathogenesis, and host immunity. *Virulence* **2018**, *9*, 1426–1435. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Gilchrist, T.C.; Stokes, W.R. A case of pseudo-lupus vulgaris caused by a blastomyces. *J. Exp. Med.* **1898**, *3*, 53–78. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Kaffenberger, B.H.; Shetlar, D.; Norton, S.A.; Rosenbach, M. The effect of climate change on skin disease in North America. *J. Am. Acad. Dermatol.* **2016**, *76*, 140–147. [\[CrossRef\]](#)
126. Hernandez, H.; Martinez, L.R. Relationship of environmental disturbances and the infectious potential of fungi. *Microbiology* **2018**, *164*, 233–241. [\[CrossRef\]](#)
127. Brown, E.; McTaggart, L.R.; Dunn, D.; Pszczolko, E.; Tsui, K.G.; Morris, S.K.; Stephens, D.; Kus, J.V.; Richardson, S.E. Epidemiology and Geographic Distribution of Blastomycosis, Histoplasmosis, and Coccidioidomycosis, Ontario, Canada, 1990–2015. *Emerg. Infect. Dis.* **2018**, *24*, 1257–1266. [\[CrossRef\]](#)
128. Gnat, S.; Łagowski, D.; Nowakiewicz, A.; Dylag, M. A global view on fungal infections in humans and animals: Infections caused by dimorphic fungi and dermatophytes. *J. Appl. Microbiol.* **2021**, *131*, 2688–2704. [\[CrossRef\]](#)
129. Chapman, S.W.; Dismukes, W.E.; Proia, L.A.; Bradsher, R.W.; Pappas, P.G.; Threlkeld, M.G.; Kauffman, C.A. Clinical Practice Guidelines for the Management of Blastomycosis: 2008 Update by the Infectious Diseases Society of America. *Clin. Infect. Dis.* **2008**, *46*, 1801–1812. [\[CrossRef\]](#)
130. Garcia, S.C.G.; Alanis, J.C.S.; Flores, M.G.; Gonzalez, S.E.G.; Cabrera, L.V.; Candiani, J.O. Coccidioidomycosis and the skin: A comprehensive review. *An. Bras. Dermatol.* **2015**, *90*, 610–619. [\[CrossRef\]](#)
131. Dukik, K.; Muñoz, J.F.; Jiang, Y.; Feng, P.; Sigler, L.; Stielow, J.B.; Freeke, J.; Jamalian, A.; Ende, B.G.V.D.; McEwen, J.G.; et al. Novel taxa of thermally dimorphic systemic pathogens in the *Ajellomycetaceae* (*Onygenales*). *Mycoses* **2017**, *60*, 296–309. [\[CrossRef\]](#)
132. Taylor, M.L.; Reyes-Montes, M.D.R.; Estrada-Bárcenas, D.A.; Zancopé-Oliveira, R.M.; Rodríguez-Arellanes, G.; Ramírez, J.A. Considerations about the Geographic Distribution of *Histoplasma* Species. *Appl. Environ. Microbiol.* **2022**, *88*, e02010–21. [\[CrossRef\]](#) [\[PubMed\]](#)
133. Darling, S.T. A protozoön general infection producing pseudotubercles in the lungs and focal necroses in the liver, spleen and lymphnodes. *J. Am. Med. Assoc.* **1906**, *46*, 1283–1285. [\[CrossRef\]](#)

134. Vinh, D.C.; Embil, J.M. Histoplasmosis, Blastomycosis, Coccidioidomycosis, and Other Dimorphic Fungi. In *Principles and Practice of Hospital Medicine*, 2nd ed.; McKean, S.C., Ross, J.J., Dressler, D.D., Scheurer, D.B., Eds.; McGraw-Hill Education: New York, NY, USA, 2017.
135. Furtado, T.A.; Wilson, J.W.; Plunkett, O.A. South american blastomycosis or paracoccidioidomycosis. *AMA Arch. Dermatol. Syphilol.* **1954**, *70*, 166–180. [[CrossRef](#)] [[PubMed](#)]
136. Hahn, R.C.; Hagen, F.; Mendes, R.P.; Burger, E.; Nery, A.F.; Siqueira, N.P.; Guevara, A.; Rodrigues, A.M.; de Camargo, Z.P. Paracoccidioidomycosis: Current Status and Future Trends. *Clin. Microbiol. Rev.* **2022**, *35*, e00233–21. [[CrossRef](#)]
137. Sporotrichosis. *JAMA* **1909**, *15*, 1192. [[CrossRef](#)]
138. De Carolis, E.; Posteraro, B.; Sanguinetti, M. Old and New Insights into *Sporothrix schenckii* Complex Biology and Identification. *Pathogens* **2022**, *11*, 297. [[CrossRef](#)]
139. Capponi, M.; Segretain, G.; Sureau, P. Penicilliosis from *Rhizomys sinensis*. *Bull. Soc. Pathol. Exot.* **1956**, *49*, 418–421.
140. Segretain, G. *Penicillium Marneffe* N. Sp., Agent D'une Mycose Du Système Réticulo-Endothélial. *Mycopathologia* **1959**, *11*, 327–353. [[CrossRef](#)]
141. Pruksaphon, K.; Nosanchuk, J.D.; Ratanabanangkoon, K.; Youngchim, S. *Talaromyces marneffe* Infection: Virulence, Intracellular Lifestyle and Host Defense Mechanisms. *J. Fungi* **2022**, *8*, 200. [[CrossRef](#)]
142. DiSalvo, M.A.F.; Fickling, A.M.; Ajello, L. Infection Caused by *Penicillium marneffe*: Description of First Natural Infection in Man. *Am. J. Clin. Pathol.* **1973**, *60*, 259–263. [[CrossRef](#)]
143. Jayanetra, P.; Prajaktam, R.; Ajello, L.; Sathaphatayavongs, B.; Lolekha, S.; Padhye, A.A.; Vathesatogit, P.; Atichartakarn, V.; Nitiyanant, P. Penicilliosis marneffe in Thailand: Report of Five Human Cases. *Am. J. Trop. Med. Hyg.* **1984**, *33*, 637–644. [[CrossRef](#)] [[PubMed](#)]
144. Li, Y.; Luo, H.; Fan, J.; Lan, X.; Liu, G.; Zhang, J.; Zhong, X.; Pang, Y.; Wang, J.; He, Z. Genomic analysis provides insights into the transmission and pathogenicity of *Talaromyces marneffe*. *Fungal Genet. Biol.* **2019**, *130*, 54–61. [[CrossRef](#)] [[PubMed](#)]
145. Narayanasamy, S.; Dat, V.Q.; Thanh, N.T.; Ly, V.T.; Chan, J.F.-W.; Yuen, K.-Y.; Ning, C.; Liang, H.; Li, L.; Chowdhary, A.; et al. A global call for talaromycosis to be recognised as a neglected tropical disease. *Lancet Glob. Health* **2021**, *9*, e1618–e1622. [[CrossRef](#)]
146. Cao, C.; Xi, L.; Chaturvedi, V. Talaromycosis (Penicilliosis) Due to *Talaromyces* (*Penicillium*) *marneffe*: Insights into the Clinical Trends of a Major Fungal Disease 60 Years After the Discovery of the Pathogen. *Mycopathologia* **2019**, *184*, 709–720. [[CrossRef](#)]
147. Döffinger, R.; Dupuis, S.; Picard, C.; Fieschi, C.; Feinberg, J.; Barcenas-Morales, G.; Casanova, J.-L. Inherited disorders of IL-12 and IFN $\gamma$ -mediated immunity: A molecular genetics update. *Mol. Immunol.* **2002**, *38*, 903–909. [[CrossRef](#)]
148. Rosain, J.; Kong, X.; Martinez-Barricarte, R.; Oleaga-Quintas, C.; Ramirez-Alejo, N.; Markle, J.; Okada, S.; Boisson-Dupuis, S.; Casanova, J.; Bustamante, J. Mendelian susceptibility to mycobacterial disease: 2014–2018 update. *Immunol. Cell Biol.* **2018**, *97*, 360–367. [[CrossRef](#)]
149. Kerner, G.; Rosain, J.; Guérin, A.; Al-Khabaz, A.; Oleaga-Quintas, C.; Rapaport, F.; Massaad, M.J.; Ding, J.-Y.; Khan, T.; Al Ali, F.; et al. Inherited human IFN- $\gamma$  deficiency underlies mycobacterial disease. *J. Clin. Invest.* **2020**, *130*, 3158–3171. [[CrossRef](#)] [[PubMed](#)]
150. van de Vosse, E.; Haverkamp, M.H.; Ramirez-Alejo, N.; Martinez-Gallo, M.; Blancas-Galicia, L.; Metin, A.; Garty, B.Z.; Sun-Tan, Ç.; Broides, A.; de Paus, R.A.; et al. IL-12R $\beta$ 1 Deficiency: Mutation Update and Description of the IL12RB1 Variation Database. *Hum. Mutat.* **2013**, *34*, 1329–1339. [[CrossRef](#)]
151. Alaki, E.M.; Aljobair, F.; Alaklobi, F.; Al Shamrani, M.; Al-Zahim, F.; Dongues, A.; Casanova, J.-L. Chronic Disseminated Salmonellosis in a Patient With Interleukin-12p40 Deficiency. *Pediatr. Infect. Dis. J.* **2018**, *37*, 90–93. [[CrossRef](#)]
152. Mahdavian, S.A.; Mansouri, D.; Jamee, M.; Zaki-Dizaji, M.; Aghdam, K.R.; Mortaz, E.; Khorasanizadeh, M.; Eskian, M.; Movahedi, M.; Ghaffaripour, H.; et al. Mendelian Susceptibility to Mycobacterial Disease (MSMD): Clinical and Genetic Features of 32 Iranian Patients. *J. Clin. Immunol.* **2020**, *40*, 872–882. [[CrossRef](#)]
153. Vicuña, A.K.P.; Nakashimada, M.Y.; Lara, X.L.; Flores, E.M.; Núñez, M.E.N.; Lona-Reyes, J.C.; Nieto, L.H.; Vázquez, M.G.R.; Santos, J.B.; Iñiguez, L.; et al. Mendelian Susceptibility to Mycobacterial Disease: Retrospective Clinical and Genetic Study in Mexico. *J. Clin. Immunol.* **2023**, *43*, 123–135. [[CrossRef](#)]
154. Tanir, G.; Dogu, F.; Tuygun, N.; İkinciogullari, A.; Aytakin, C.; Aydemir, C.; Yuksek, M.; Boduroglu, E.C.; De Beaucoudrey, L.; Fieschi, C.; et al. Complete deficiency of the IL-12 receptor  $\beta$ 1 chain: Three unrelated Turkish children with unusual clinical features. *Eur. J. Pediatr.* **2006**, *165*, 415–417. [[CrossRef](#)] [[PubMed](#)]
155. Rosain, J.; Oleaga-Quintas, C.; Deswarte, C.; Verdin, H.; Marot, S.; Syridou, G.; Mansouri, M.; Mahdavian, S.A.; Venegas-Montoya, E.; Tsolia, M.; et al. A Variety of Alu-Mediated Copy Number Variations Can Underlie IL-12R $\beta$ 1 Deficiency. *J. Clin. Immunol.* **2018**, *38*, 617–627. [[CrossRef](#)] [[PubMed](#)]
156. León-Lara, X.; Hernández-Nieto, L.; Zamora, C.V.; Rodríguez-D'Cid, R.; Gutiérrez, M.E.C.; Espinosa-Padilla, S.; Bustamante, J.; Puel, A.; Blancas-Galicia, L. Disseminated Infectious Disease Caused by *Histoplasma capsulatum* in an Adult Patient as First Manifestation of Inherited IL-12R $\beta$ 1 Deficiency. *J. Clin. Immunol.* **2020**, *40*, 1051–1054. [[CrossRef](#)]
157. de Souza, T.L.; Fernandes, R.C.D.S.C.; Da Silva, J.A.; Júnior, V.G.A.; Coelho, A.G.; Faria, A.C.S.; Simão, N.M.M.S.; Filho, J.T.S.; Deswarte, C.; Boisson-Dupuis, S.; et al. Microbial Disease Spectrum Linked to a Novel IL-12R $\beta$ 1 N-Terminal Signal Peptide Stop-Gain Homozygous Mutation with Paradoxical Receptor Cell-Surface Expression. *Front. Microbiol.* **2017**, *8*, 616. [[CrossRef](#)]
158. Zerbe, C.S.; Holland, S.M. Disseminated Histoplasmosis in Persons with Interferon- Receptor 1 Deficiency. *Clin. Infect. Dis.* **2005**, *41*, 38–41. [[CrossRef](#)] [[PubMed](#)]

159. Vinh, D.; Masannat, F.; Dzioba, R.B.; Galgiani, J.N.; Holland, S.M. Refractory Disseminated Coccidioidomycosis and Mycobacteriosis in Interferon- $\gamma$  Receptor 1 Deficiency. *Clin. Infect. Dis.* **2009**, *49*, e62–e65. [\[CrossRef\]](#)
160. Vinh, D.; Schwartz, B.; Hsu, A.; Miranda, D.J.; Valdez, P.A.; Fink, D.; Lau, K.P.; Long-Priel, D.; Kuhns, D.B.; Uzel, G.; et al. Interleukin-12 Receptor 1 Deficiency Predisposing to Disseminated Coccidioidomycosis. *Clin. Infect. Dis.* **2011**, *52*, e99–e102. [\[CrossRef\]](#)
161. De Moraes-Vasconcelos, D.; Grumach, A.; Yamaguti, A.; Andrade, M.E.B.; Fieschi, C.; De Beaucoudrey, L.; Casanova, J.-L.; Duarte, A.J.S. Paracoccidioides brasiliensis Disseminated Disease in a Patient with Inherited Deficiency in the 1 Subunit of the Interleukin (IL)-12/IL-23 Receptor. *Clin. Infect. Dis.* **2005**, *41*, 31–37. [\[CrossRef\]](#)
162. Pedroza, L.A.; Guerrero, N.; Stray-Pedersen, A.; Tafur, C.; Macias, R.; Muñoz, G.; Akdemir, Z.C.; Jhangiani, S.N.; Watkin, L.B.; Chinn, I.K.; et al. First Case of CD40LG Deficiency in Ecuador, Diagnosed after Whole Exome Sequencing in a Patient with Severe Cutaneous Histoplasmosis. *Front. Pediatr.* **2017**, *5*, 17. [\[CrossRef\]](#) [\[PubMed\]](#)
163. Winkelstein, J.A.; Marino, M.C.; Ochs, H.; Fuleihan, R.; Scholl, P.R.; Geha, R.; Stiehm, E.R.; Conley, M.E. The X-Linked Hyper-IgM Syndrome. *Medicine* **2003**, *82*, 373–384. [\[CrossRef\]](#) [\[PubMed\]](#)
164. Danielian, S.; Oleastro, M.; Rivas, M.E.; Cantisano, C.; Zelazko, M. Clinical Follow-Up of 11 Argentinian CD40L-Deficient Patients with 7 Unique Mutations Including the So-Called “Milder” Mutants. *J. Clin. Immunol.* **2007**, *27*, 455–459. [\[CrossRef\]](#)
165. Cabral-Marques, O.; Schimke, L.-F.; Pereira, P.V.S.; Falcai, A.; de Oliveira, J.B.; Hackett, M.J.; Errante, P.R.; Weber, C.W.; Ferreira, J.F.; Kuntze, G.; et al. Expanding the Clinical and Genetic Spectrum of Human CD40L Deficiency: The Occurrence of Paracoccidioidomycosis and Other Unusual Infections in Brazilian Patients. *J. Clin. Immunol.* **2011**, *32*, 212–220. [\[CrossRef\]](#)
166. Marques, O.C.; Arslanian, C.; Ramos, R.N.; Morato, M.; Schimke, L.; Pereira, P.V.S.; Jancar, S.; Ferreira, J.F.; Weber, C.W.; Kuntze, G.; et al. Dendritic cells from X-linked hyper-IgM patients present impaired responses to Candida albicans and Paracoccidioides brasiliensis. *J. Allergy Clin. Immunol.* **2012**, *129*, 778–786. [\[CrossRef\]](#)
167. Du, X.; Tang, W.; Chen, X.; Zeng, T.; Wang, Y.; Chen, Z.; Xu, T.; Zhou, L.; Tang, X.; An, Y.; et al. Clinical, genetic and immunological characteristics of 40 Chinese patients with CD40 ligand deficiency. *Scand. J. Immunol.* **2019**, *90*, e12798. [\[CrossRef\]](#)
168. Lee, P.P.; Lao-Araya, M.; Yang, J.; Chan, K.-W.; Ma, H.; Pei, L.-C.; Kui, L.; Mao, H.; Yang, W.; Zhao, X.; et al. Application of Flow Cytometry in the Diagnostics Pipeline of Primary Immunodeficiencies Underlying Disseminated Talaromyces marneffeii Infection in HIV-Negative Children. *Front. Immunol.* **2019**, *10*, 2189. [\[CrossRef\]](#) [\[PubMed\]](#)
169. Wang, L.; Luo, Y.; Li, X.; Li, Y.; Xia, Y.; He, T.; Huang, Y.; Xu, Y.; Yang, Z.; Ling, J.; et al. Talaromyces marneffeii Infections in 8 Chinese Children with Inborn Errors of Immunity. *Mycopathologia* **2022**, *187*, 455–467. [\[CrossRef\]](#) [\[PubMed\]](#)
170. Fan, H.; Huang, L.; Yang, D.; Zhang, C.; Zeng, Q.; Yin, G.; Lu, G.; Shen, K. Respiratory infections in X-linked hyper-IgM syndrome with CD40LG mutation: A case series of seven children in China. *BMC Pediatr.* **2022**, *22*, 675. [\[CrossRef\]](#)
171. Spinner, M.A.; Ker, J.P.; Stoudenmire, C.J.; Fadare, O.; Mace, E.; Orange, J.S.; Hsu, A.; Holland, S.M. GATA2 deficiency underlying severe blastomycosis and fatal herpes simplex virus-associated hemophagocytic lymphohistiocytosis. *J. Allergy Clin. Immunol.* **2015**, *137*, 638–640. [\[CrossRef\]](#)
172. Sampaio, E.P.; Hsu, A.P.; Pechacek, J.; Bax, H.I.; Dias, D.L.; Paulson, M.L.; Chandrasekaran, P.; Rosen, L.B.; Carvalho, D.S.; Ding, L.; et al. Signal transducer and activator of transcription 1 (STAT1) gain-of-function mutations and disseminated coccidioidomycosis and histoplasmosis. *J. Allergy Clin. Immunol.* **2013**, *131*, 1624–1634. [\[CrossRef\]](#) [\[PubMed\]](#)
173. Chen, K.; Tan, J.; Qian, S.; Wu, S.; Chen, Q. Case Report: Disseminated Talaromyces marneffeii Infection in a Patient With Chronic Mucocutaneous Candidiasis and a Novel STAT1 Gain-of-Function Mutation. *Front. Immunol.* **2021**, *12*, 682350. [\[CrossRef\]](#)
174. Vinh, D.; Patel, S.; Uzel, G.; Anderson, V.L.; Freeman, A.F.; Olivier, K.N.; Spalding, C.; Hughes, S.; Pittaluga, S.; Raffeld, M.; et al. Autosomal dominant and sporadic monocytopenia with susceptibility to mycobacteria, fungi, papillomaviruses, and myelodysplasia. *Blood* **2010**, *115*, 1519–1529. [\[CrossRef\]](#) [\[PubMed\]](#)
175. Calvo, K.R.; Vinh, D.C.; Maric, I.; Wang, W.; Noel, P.; Stetler-Stevenson, M.; Arthur, D.C.; Raffeld, M.; Dutra, A.; Pak, E.; et al. Myelodysplasia in autosomal dominant and sporadic monocytopenia immunodeficiency syndrome: Diagnostic features and clinical implications. *Haematologica* **2011**, *96*, 1221–1225. [\[CrossRef\]](#)
176. Hsu, A.; Sampaio, E.P.; Khan, J.; Calvo, K.; Lemieux, J.E.; Patel, S.; Frucht, D.M.; Vinh, D.; Auth, R.D.; Freeman, A.F.; et al. Mutations in GATA2 are associated with the autosomal dominant and sporadic monocytopenia and mycobacterial infection (MonoMAC) syndrome. *Blood* **2011**, *118*, 2653–2655. [\[CrossRef\]](#) [\[PubMed\]](#)
177. Dickinson, R.E.; Griffin, H.R.; Bigley, V.; Reynard, L.N.; Hussain, R.; Haniffa, M.; Lakey, J.; Rahman, T.; Wang, X.-N.; McGovern, N.; et al. Exome sequencing identifies GATA-2 mutation as the cause of dendritic cell, monocyte, B and NK lymphoid deficiency. *Blood* **2011**, *118*, 2656–2658. [\[CrossRef\]](#)
178. Hahn, C.; Chong, C.E.; Carmichael, C.; Wilkins, E.J.; Brautigan, P.J.; Li, X.-C.; Babic, M.; Lin, M.; Carmagnac, A.; Lee, Y.K.; et al. Heritable GATA2 mutations associated with familial myelodysplastic syndrome and acute myeloid leukemia. *Nat. Genet.* **2011**, *43*, 1012–1017. [\[CrossRef\]](#)
179. Ostergaard, P.; Simpson, M.; Connell, F.C.; Steward, C.; Brice, G.; Woollard, W.J.; Dafou, D.; Kilo, T.; Smithson, S.; Lunt, P.; et al. Mutations in GATA2 cause primary lymphedema associated with a predisposition to acute myeloid leukemia (Emberger syndrome). *Nat. Genet.* **2011**, *43*, 929–931. [\[CrossRef\]](#)
180. Pasquet, M.; Bellanné-Chantelot, C.; Tavitian, S.; Prade, N.; Beaupain, B.; LaRochelle, O.; Petit, A.; Rohrlisch, P.S.; Ferrand, C.; Neste, E.V.D.; et al. High frequency of GATA2 mutations in patients with mild chronic neutropenia evolving to MonoMAC syndrome, myelodysplasia, and acute myeloid leukemia. *Blood* **2013**, *121*, 822–829. [\[CrossRef\]](#)

181. Dorn, J.M.; Patnaik, M.S.; Van Hee, M.; Smith, M.J.; Lagerstedt, S.A.; Newman, C.C.; Boyce, T.G.; Abraham, R.S. WILD syndrome is GATA2 deficiency: A novel deletion in the GATA2 gene. *J. Allergy Clin. Immunol. Pract.* **2017**, *5*, 1149–1152.e1. [\[CrossRef\]](#)
182. Kuriyama, Y.; Hattori, M.; Mitsui, T.; Nakano, H.; Oikawa, D.; Tokunaga, F.; Ishikawa, O.; Shimizu, A. Generalized verrucosis caused by various human papillomaviruses in a patient with GATA2 deficiency. *J. Dermatol.* **2018**, *45*, e108–e109. [\[CrossRef\]](#) [\[PubMed\]](#)
183. Lasbury, M.E.; Tang, X.; Durant, P.J.; Lee, C.-H. Effect of Transcription Factor GATA-2 on Phagocytic Activity of Alveolar Macrophages from *Pneumocystis carinii*-Infected Hosts. *Infect. Immun.* **2003**, *71*, 4943–4952. [\[CrossRef\]](#) [\[PubMed\]](#)
184. Mendes-De-Almeida, D.P.; Andrade, F.G.; dos Santos-Bueno, F.V.; Freitas, D.F.S.; Soares-Lima, S.C.; Zancopé-Oliveira, R.M.; Pombo-De-Oliveira, M.S. GATA2 variants in patients with non-tuberculous mycobacterial or fungal infections without known immunodeficiencies. *Hematol. Transfus. Cell Ther.* **2022**. [\[CrossRef\]](#) [\[PubMed\]](#)
185. Chapgier, A.; Boisson-Dupuis, S.; Jouanguy, E.; Vogt, G.; Feinberg, J.; Prochnicka-Chalufour, A.; Casrouge, A.; Yang, K.; Soudais, C.; Fieschi, C.; et al. Novel STAT1 Alleles in Otherwise Healthy Patients with Mycobacterial Disease. *PLoS Genet.* **2006**, *2*, e131. [\[CrossRef\]](#) [\[PubMed\]](#)
186. Chapgier, A.; Kong, X.-F.; Boisson-Dupuis, S.; Jouanguy, E.; Averbuch, D.; Feinberg, J.; Zhang, S.-Y.; Bustamante, J.; Vogt, G.; Lejeune, J.; et al. A partial form of recessive STAT1 deficiency in humans. *J. Clin. Investig.* **2009**, *119*, 1502–1514. [\[CrossRef\]](#) [\[PubMed\]](#)
187. Boisson-Dupuis, S.; Jouanguy, E.; Al-Hajjar, S.; Fieschi, C.; Al-Mohsen, I.Z.; Al-Jumaah, S.; Yang, K.; Chapgier, A.; Eidsenck, C.; Eid, P.; et al. Impaired response to interferon- $\alpha/\beta$  and lethal viral disease in human STAT1 deficiency. *Nat. Genet.* **2003**, *33*, 388–391. [\[CrossRef\]](#)
188. Bustamante, J. Mendelian susceptibility to mycobacterial disease: Recent discoveries. *Hum. Genet.* **2020**, *139*, 993–1000. [\[CrossRef\]](#)
189. Alberti-Flor, J.J.; Granda, A. Ileocecal Histoplasmosis Mimicking Crohn's Disease in a Patient with Job's Syndrome. *Digestion* **1986**, *33*, 176–180. [\[CrossRef\]](#)
190. Cappell, M.S.; Manzione, N.C. Recurrent colonic histoplasmosis after standard therapy with amphotericin B in a patient with Job's syndrome. *Am. J. Gastroenterol.* **1991**, *86*, 119–120.
191. Desai, K.; Huston, D.P.; Harriman, G.R. Previously undiagnosed hyper-IgE syndrome in an adult with multiple systemic fungal infections. *J. Allergy Clin. Immunol.* **1996**, *98*, 1123–1124. [\[CrossRef\]](#)
192. Vinh, D.C.; Sugui, J.A.; Hsu, A.P.; Freeman, A.F.; Holland, S.M. Invasive fungal disease in autosomal-dominant hyper-IgE syndrome. *J. Allergy Clin. Immunol.* **2010**, *125*, 1389–1390. [\[CrossRef\]](#) [\[PubMed\]](#)
193. Odio, C.D.; Milligan, K.L.; McGowan, K.; Spergel, A.K.R.; Bishop, R.; Boris, L.; Urban, A.; Welch, P.; Heller, T.; Kleiner, D.; et al. Endemic mycoses in patients with STAT3-mutated hyper-IgE (Job) syndrome. *J. Allergy Clin. Immunol.* **2015**, *136*, 1411–1413.e2. [\[CrossRef\]](#)
194. Lee, P.P.W.; Chan, K.-W.; Ho, M.H.-K.; Chen, X.-Y.; Li, C.-H.; Chu, K.-M.; Zeng, H.-S.; Lau, Y.L. Penicilliosis in Children without HIV Infection—Are They Immunodeficient? *Clin. Infect. Dis.* **2011**, *54*, e8–e19. [\[CrossRef\]](#)
195. Netea, M.G.; Kullberg, B.J.; van Der Meer, J.W.M. Severely impaired IL-12/IL-18/IFN $\gamma$  axis in patients with hyper IgE syndrome. *Eur. J. Clin. Investig.* **2005**, *35*, 718–721. [\[CrossRef\]](#)
196. Danion, F.; Aïmanianda, V.; Bayry, J.; Duréault, A.; Wong, S.; Bougnoux, M.-E.; Tcherakian, C.; Alyanakian, M.-A.; Guegan, H.; Puel, A.; et al. Aspergillus fumigatus Infection in Humans With STAT3-Deficiency Is Associated With Defective Interferon-Gamma and Th17 Responses. *Front. Immunol.* **2020**, *11*, 38. [\[CrossRef\]](#) [\[PubMed\]](#)
197. Béziat, V.; Rapaport, F.; Hu, J.; Titeux, M.; Claustres, M.B.D.; Bourgey, M.; Griffin, H.; Bandet, E.; Ma, C.S.; Sherkat, R.; et al. Humans with inherited T cell CD28 deficiency are susceptible to skin papillomaviruses but are otherwise healthy. *Cell* **2021**, *184*, 3812–3828.e30. [\[CrossRef\]](#) [\[PubMed\]](#)
198. Wang, Y.; Ma, C.S.; Ling, Y.; Bousfiha, A.; Camcioglu, Y.; Jacquot, S.; Payne, K.; Crestani, E.; Roncagalli, R.; Belkadi, A.; et al. Dual T cell- and B cell-intrinsic deficiency in humans with biallelic RLTPR mutations. *J. Exp. Med.* **2016**, *213*, 2413–2435. [\[CrossRef\]](#) [\[PubMed\]](#)
199. Alazami, A.M.; Al-Helale, M.; Alhissi, S.; Al-Saud, B.; Alajlan, H.; Monies, D.; Shah, Z.; Abouelhoda, M.; Arnaout, R.; Al-Dhekri, H.; et al. Novel CARMIL2 Mutations in Patients with Variable Clinical Dermatitis, Infections, and Combined Immunodeficiency. *Front. Immunol.* **2018**, *9*, 203. [\[CrossRef\]](#) [\[PubMed\]](#)
200. Kurolap, A.; Adiv, O.E.; Konnikova, L.; Werner, L.; Gonzaga-Jauregui, C.; Steinberg, M.; Mitsialis, V.; Mory, A.; Nunberg, M.Y.; Wall, S.; et al. A Unique Presentation of Infantile-Onset Colitis and Eosinophilic Disease without Recurrent Infections Resulting from a Novel Homozygous CARMIL2 Variant. *J. Clin. Immunol.* **2019**, *39*, 430–439. [\[CrossRef\]](#)
201. Marangi, G.; Garcovich, S.; Sante, G.; Orteschi, D.; Frangella, S.; Scaldaferrì, F.; Genuardi, M.; Peris, K.; Gurrieri, F.; Zollino, M. Complex Muco-cutaneous Manifestations of CARMIL2-associated Combined Immunodeficiency: A Novel Presentation of Dysfunctional Epithelial Barriers. *Acta Derm.-Venereol.* **2020**, *100*, 1–2. [\[CrossRef\]](#) [\[PubMed\]](#)
202. Maccari, M.E.; Speckmann, C.; Heeg, M.; Reimer, A.; Casetti, F.; Has, C.; Ehl, S.; Castro, C.N. Profound immunodeficiency with severe skin disease explained by concomitant novel CARMIL2 and PLEC1 loss-of-function mutations. *Clin. Immunol.* **2019**, *208*, 108228. [\[CrossRef\]](#)
203. Atschekzei, F.; Jacobs, R.; Wetzke, M.; Sogkas, G.; Schröder, C.; Ahrenstorf, G.; Dhingra, A.; Ott, H.; Baumann, U.; Schmidt, R.E. A Novel CARMIL2 Mutation Resulting in Combined Immunodeficiency Manifesting with Dermatitis, Fungal, and Viral Skin Infections As Well as Selective Antibody Deficiency. *J. Clin. Immunol.* **2019**, *39*, 274–276. [\[CrossRef\]](#)

204. Kolukisa, B.; Baser, D.; Akcam, B.; Danielson, J.; Eltan, S.B.; Haliloglu, Y.; Sefer, A.P.; Babayeva, R.; Akgun, G.; Charbonnier, L.; et al. Evolution and long-term outcomes of combined immunodeficiency due to CARMIL2 deficiency. *Allergy* **2021**, *77*, 1004–1019. [\[CrossRef\]](#)
205. Lévy, R.; Gothe, F.; Momenilandi, M.; Magg, T.; Materna, M.; Peters, P.; Raedler, J.; Philippot, Q.; Rack-Hoch, A.L.; Langlais, D.; et al. Human CARMIL2 deficiency underlies a broader immunological and clinical phenotype than CD28 deficiency. *J. Exp. Med.* **2023**, *220*, e20220275. [\[CrossRef\]](#) [\[PubMed\]](#)
206. Stepensky, P.; Keller, B.; Buchta, M.; Kienzler, A.-K.; Elpeleg, O.; Somech, R.; Cohen, S.; Shachar, I.; Miosge, L.A.; Schlesier, M.; et al. Deficiency of caspase recruitment domain family, member 11 (CARD11), causes profound combined immunodeficiency in human subjects. *J. Allergy Clin. Immunol.* **2013**, *131*, 477–485.e1. [\[CrossRef\]](#)
207. Greil, J.; Rausch, T.; Giese, T.; Bandapalli, O.R.; Daniel, V.; Bekerredjian-Ding, I.; Stütz, A.M.; Drees, C.; Roth, S.; Ruland, J.; et al. Whole-exome sequencing links caspase recruitment domain 11 (CARD11) inactivation to severe combined immunodeficiency. *J. Allergy Clin. Immunol.* **2013**, *131*, 1376–1383.e3. [\[CrossRef\]](#)
208. Snow, A.L.; Xiao, W.; Stinson, J.R.; Lu, W.; Chaigne-Delalande, B.; Zheng, L.; Pittaluga, S.; Matthews, H.F.; Schmitz, R.; Jhavar, S.; et al. Congenital B cell lymphocytosis explained by novel germline CARD11 mutations. *J. Exp. Med.* **2012**, *209*, 2247–2261. [\[CrossRef\]](#)
209. Brohl, A.; Stinson, J.R.; Su, H.C.; Badgett, T.; Jennings, C.D.; Sukumar, G.; Sindiri, S.; Wang, W.; Kardava, L.; Moir, S.; et al. Germline CARD11 Mutation in a Patient with Severe Congenital B Cell Lymphocytosis. *J. Clin. Immunol.* **2014**, *35*, 32–46. [\[CrossRef\]](#)
210. Lu, H.Y.; Biggs, C.M.; Blanchard-Rohner, G.; Fung, S.-Y.; Sharma, M.; Turvey, S.E. Germline CBM-opathies: From immunodeficiency to atopy. *J. Allergy Clin. Immunol.* **2019**, *143*, 1661–1673. [\[CrossRef\]](#)
211. Grimbacher, B.; Hutloff, A.; Schlesier, M.; Glocker, E.; Warnatz, K.; Dräger, R.; Eibel, H.; Fischer, B.; Schäffer, A.A.; Mages, H.W.; et al. Homozygous loss of ICOS is associated with adult-onset common variable immunodeficiency. *Nat. Immunol.* **2003**, *4*, 261–268. [\[CrossRef\]](#)
212. Salzer, U.; Maul-Pavicic, A.; Cunningham-Rundles, C.; Urschel, S.; Belohradsky, B.H.; Litzman, J.; Holm, A.; Franco, J.L.; Plebani, A.; Hammarstrom, L.; et al. ICOS deficiency in patients with common variable immunodeficiency. *Clin. Immunol.* **2004**, *113*, 234–240. [\[CrossRef\]](#)
213. Bossaller, L.; Burger, J.; Draeger, R.; Grimbacher, B.; Knoth, R.; Plebani, A.; Durandy, A.; Baumann, U.; Schlesier, M.; Welcher, A.A.; et al. ICOS Deficiency Is Associated with a Severe Reduction of CXCR5+CD4 Germinal Center Th Cells. *J. Immunol.* **2006**, *177*, 4927–4932. [\[CrossRef\]](#)
214. Takahashi, N.; Matsumoto, K.; Saito, H.; Nanki, T.; Miyasaka, N.; Kobata, T.; Azuma, M.; Lee, S.-K.; Mizutani, S.; Morio, T. Impaired CD4 and CD8 Effector Function and Decreased Memory T Cell Populations in ICOS-Deficient Patients. *J. Immunol.* **2009**, *182*, 5515–5527. [\[CrossRef\]](#)
215. Chou, J.; Massaad, M.J.; Cangemi, B.; Bainter, W.; Platt, C.; Badran, Y.R.; Raphael, B.P.; Kamin, D.S.; Goldsmith, J.D.; Pai, S.-Y.; et al. A novel mutation in ICOS presenting as hypogammaglobulinemia with susceptibility to opportunistic pathogens. *J. Allergy Clin. Immunol.* **2015**, *136*, 794–797.e1. [\[CrossRef\]](#)
216. Abolhassani, H.; El-Sherbiny, Y.M.; Arumugakani, G.; Carter, C.; Richards, S.; Lawless, D.; Wood, P.; Buckland, M.; Heydarzadeh, M.; Aghamohammadi, A.; et al. Expanding Clinical Phenotype and Novel Insights into the Pathogenesis of ICOS Deficiency. *J. Clin. Immunol.* **2010**, *40*, 277–288. [\[CrossRef\]](#)
217. Roussel, L.; Landekic, M.; Golizeh, M.; Gavino, C.; Zhong, M.-C.; Chen, J.; Faubert, D.; Blanchet-Cohen, A.; Dansereau, L.; Parent, M.-A.; et al. Loss of human ICOSL results in combined immunodeficiency. *J. Exp. Med.* **2018**, *215*, 3151–3164. [\[CrossRef\]](#)
218. Loo, V.G.; Roussel, L.; Bernier, S.; Perez, A.; Vinh, D.C. Late-Onset Combined Immunodeficiency with Refractory CMV Disease due to ICOSL Deficiency. *J. Clin. Immunol.* **2022**, *42*, 206–209. [\[CrossRef\]](#)
219. Kuberski, T.T.; Servi, R.J.; Rubin, P.J. Successful Treatment of a Critically Ill Patient with Disseminated Coccidioidomycosis, Using Adjunctive Interferon- $\gamma$ . *Clin. Infect. Dis.* **2004**, *38*, 910–912. [\[CrossRef\]](#)
220. Duplessis, C.A.; Tilley, D.; Bavaro, M.; Hale, B.; Holland, S.M. Two cases illustrating successful adjunctive interferon- $\gamma$  immunotherapy in refractory disseminated coccidioidomycosis. *J. Infect.* **2011**, *63*, 223–228. [\[CrossRef\]](#)
221. Tsai, M.; Thauland, T.J.; Huang, A.Y.; Bun, C.; Fitzwater, S.; Krogstad, P.; Douine, E.D.; Nelson, S.F.; Lee, H.; Garcia-Lloret, M.I.; et al. Disseminated Coccidioidomycosis Treated with Interferon- $\gamma$  and Dupilumab. *N. Engl. J. Med.* **2020**, *382*, 2337–2343. [\[CrossRef\]](#)
222. Madariaga, L.; Amurrio, C.; Martín, G.; García-Cebrian, F.; Bicandi, J.; Lardelli, P.; Suarez, M.D.; Cisterna, R. Detection of anti-interferon-gamma autoantibodies in subjects infected by Mycobacterium tuberculosis. *Int. J. Tuberc. Lung Dis.* **1998**, *2*, 62–68.
223. Doffinger, R.; Helbert, M.R.; Morales, G.B.; Yang, K.; Dupuis, S.; Ceron-Gutierrez, L.; Espitia-Pinzon, C.; Barnes, N.; Bothamley, G.; Casanova, J.; et al. Autoantibodies to Interferon- $\gamma$  in a Patient with Selective Susceptibility to Mycobacterial Infection and Organ-Specific Autoimmunity. *Clin. Infect. Dis.* **2004**, *38*, e10–e14. [\[CrossRef\]](#)
224. Höflich, C.; Sabat, R.; Rosseau, S.; Temmesfeld, B.; Slevogt, H.; Döcke, W.D.; Grütz, G.; Meisel, C.; Halle, E.; Göbel, U.B.; et al. Naturally occurring anti-IFN- $\gamma$  autoantibody and severe infections with Mycobacterium chelonae and Burkholderia cocovenenans. *Blood* **2004**, *103*, 673–675. [\[CrossRef\]](#)

225. Kampmann, B.; Hemingway, C.; Stephens, A.; Davidson, R.; Goodsall, A.; Anderson, S.; Nicol, M.; Schölvinck, E.; Relman, D.; Waddell, S.; et al. Acquired predisposition to mycobacterial disease due to autoantibodies to IFN- $\gamma$ . *J. Clin. Investig.* **2005**, *115*, 2480–2488. [\[CrossRef\]](#) [\[PubMed\]](#)
226. Patel, S.Y.; Ding, L.; Brown, M.R.; Lantz, L.; Gay, T.; Cohen, S.; Martyak, L.A.; Kubak, B.; Holland, S.M. Anti-IFN- $\gamma$  Autoantibodies in Disseminated Nontuberculous Mycobacterial Infections. *J. Immunol.* **2005**, *175*, 4769–4776. [\[CrossRef\]](#) [\[PubMed\]](#)
227. Koya, T.; Tsubata, C.; Kagamu, H.; Koyama, K.-I.; Hayashi, M.; Takada, T.; Gejyo, F.; Kuwabara, K.; Itoh, T.; Tanabe, Y. Anti-interferon- $\gamma$  autoantibody in a patient with disseminated Mycobacterium avium complex. *J. Infect. Chemother.* **2009**, *15*, 118–122. [\[CrossRef\]](#) [\[PubMed\]](#)
228. Browne, S.K.; Burbelo, P.D.; Chetchotisakd, P.; Suputtamongkol, Y.; Kiertiburanakul, S.; Shaw, P.A.; Kirk, J.L.; Jutivorakool, K.; Zaman, R.; Ding, L.; et al. Adult-Onset Immunodeficiency in Thailand and Taiwan. *N. Engl. J. Med.* **2012**, *367*, 725–734. [\[CrossRef\]](#) [\[PubMed\]](#)
229. Wongkulab, P.; Wipasa, J.; Chaiwarith, R.; Supparatpinyo, K. Autoantibody to Interferon-gamma Associated with Adult-Onset Immunodeficiency in Non-HIV Individuals in Northern Thailand. *PLoS ONE* **2013**, *8*, e76371. [\[CrossRef\]](#)
230. Chan, J.F.; Lau, S.K.; Yuen, K.Y.; Woo, P.C. Talaromyces (Penicillium) marneffeii infection in non-HIV-infected patients. *Emerg. Microbes Infect.* **2016**, *5*, 1–9. [\[CrossRef\]](#)
231. Wipasa, J.; Chaiwarith, R.; Chawansuntati, K.; Praparattanapan, J.; Rattanathammethee, K.; Supparatpinyo, K. Characterization of anti-interferon- $\gamma$  antibodies in HIV-negative immunodeficient patients infected with unusual intracellular microorganisms. *Exp. Biol. Med.* **2018**, *243*, 621–626. [\[CrossRef\]](#)
232. Su, S.-S.; Zhang, S.-N.; Ye, J.-R.; Xu, L.-N.; Lin, P.-C.; Xu, H.-Y.; Wu, Q.; Li, Y.-P. Disseminated Talaromyces marneffeii And Mycobacterium avium Infection Accompanied Sweet's Syndrome In A Patient With Anti-Interferon- $\gamma$  Autoantibodies: A Case Report. *Infect. Drug Resist.* **2019**, *12*, 3189–3195. [\[CrossRef\]](#)
233. Zeng, W.; Qiu, Y.; Tang, S.; Zhang, J.; Pan, M.; Zhong, X. Characterization of Anti-Interferon- $\gamma$  Antibodies in HIV-Negative Patients Infected With Disseminated Talaromyces marneffeii and Cryptococcosis. *Open Forum Infect. Dis.* **2019**, *6*, ofz208. [\[CrossRef\]](#)
234. Wongkamhla, T.; Chongtrakool, P.; Jitmuang, A. A case report of Talaromyces marneffeii Oro-pharyngo-laryngitis: A rare manifestation of Talaromycosis. *BMC Infect. Dis.* **2019**, *19*, 1034. [\[CrossRef\]](#)
235. Liang, X.; Si, L.; Li, Y.; Zhang, J.; Deng, J.; Bai, J.; Li, M.; He, Z. Talaromyces marneffeii infection relapse presenting as osteolytic destruction followed by suspected nontuberculous mycobacterium infection during 6 years of follow-up: A case update. *Int. J. Infect. Dis.* **2020**, *93*, 208–210. [\[CrossRef\]](#)
236. Jin, W.; Liu, J.; Chen, K.; Shen, L.; Zhou, Y.; Wang, L. Coinfection by Talaromyces marneffeii and Mycobacterium abscessus in a human immunodeficiency virus-negative patient with anti-interferon- $\gamma$  autoantibody: A case report. *J. Int. Med. Res.* **2021**, *49*, 0300060520976471. [\[CrossRef\]](#) [\[PubMed\]](#)
237. Chen, Z.-M.; Li, Z.-T.; Li, S.-Q.; Guan, W.-J.; Qiu, Y.; Lei, Z.-Y.; Zhan, Y.-Q.; Zhou, H.; Lin, S.; Wang, X.; et al. Clinical findings of Talaromyces marneffeii infection among patients with anti-interferon- $\gamma$  immunodeficiency: A prospective cohort study. *BMC Infect. Dis.* **2021**, *21*, 587. [\[CrossRef\]](#) [\[PubMed\]](#)
238. Lin, F.; Yang, Z.; Qiu, Y.; Zeng, W.; Liu, G.; Zhang, J. Talaromyces Marneffeii Infection in Lung Cancer Patients with Positive AIGAs: A Rare Case Report. *Infect. Drug Resist.* **2021**, *14*, 5005–5013. [\[CrossRef\]](#) [\[PubMed\]](#)
239. Qiu, Y.; Pan, M.; Yang, Z.; Zeng, W.; Zhang, H.; Li, Z.; Zhang, J. Talaromyces marneffeii and Mycobacterium tuberculosis co-infection in a patient with high titer anti-interferon- $\gamma$  autoantibodies: A case report. *BMC Infect. Dis.* **2022**, *22*, 98. [\[CrossRef\]](#) [\[PubMed\]](#)
240. Qiu, Y.; Fang, G.; Ye, F.; Zeng, W.; Tang, M.; Wei, X.; Yang, J.; Li, Z.; Zhang, J. Pathogen spectrum and immunotherapy in patients with anti-IFN- $\gamma$  autoantibodies: A multicenter retrospective study and systematic review. *Front. Immunol.* **2022**, *13*, 1051673. [\[CrossRef\]](#) [\[PubMed\]](#)
241. Hong, G.H.; Ortega-Villa, A.M.; Hunsberger, S.; Chetchotisakd, P.; Anunnatsiri, S.; Mootsikapun, P.; Rosen, L.B.; Zerbe, C.S.; Holland, S.M. Natural History and Evolution of Anti-Interferon- $\gamma$  Autoantibody-Associated Immunodeficiency Syndrome in Thailand and the United States. *Clin. Infect. Dis.* **2020**, *71*, 53–62. [\[CrossRef\]](#) [\[PubMed\]](#)
242. Tang, B.S.-F.; Chan, J.F.-W.; Chen, M.; Tsang, O.T.-Y.; Mok, M.Y.; Lai, R.W.-M.; Lee, R.; Que, T.-L.; Tse, H.; Li, I.W.-S.; et al. Disseminated Penicilliosis, Recurrent Bacteremic Nontyphoidal Salmonellosis, and Burkholderiosis Associated with Acquired Immunodeficiency Due to Autoantibody against Gamma Interferon. *Clin. Vaccine Immunol.* **2010**, *17*, 1132–1138. [\[CrossRef\]](#)
243. Kampitak, T.; Suwanpimolkul, G.; Browne, S.; Suankratay, C. Anti-interferon- $\gamma$  autoantibody and opportunistic infections: Case series and review of the literature. *Infection* **2010**, *39*, 65–71. [\[CrossRef\]](#) [\[PubMed\]](#)
244. Chan, J.F.-W.; Yee, K.-S.; Tang, B.S.-F.; Cheng, V.C.-C.; Hung, I.F.-N.; Yuen, K.-Y. Adult-onset immunodeficiency due to anti-interferon-gamma autoantibody in mainland Chinese. *Chin. Med. J.* **2014**, *127*, 1189–1190. [\[PubMed\]](#)
245. Chi, C.-Y.; Chu, C.-C.; Liu, J.-P.; Lin, C.-H.; Ho, M.-W.; Lo, W.-J.; Lin, P.-C.; Chen, H.-J.; Chou, C.-H.; Feng, J.-Y.; et al. Anti-IFN- $\gamma$  autoantibodies in adults with disseminated nontuberculous mycobacterial infections are associated with HLA-DRB1\*16:02 and HLA-DQB1\*05:02 and the reactivation of latent varicella-zoster virus infection. *Blood* **2013**, *121*, 1357–1366. [\[CrossRef\]](#)
246. Pithukpakorn, M.; Roothumnong, E.; Angkasekwinai, N.; Suktitipat, B.; Assawamakin, A.; Luangwedchakarn, V.; Umrod, P.; Thongnoppakhun, W.; Foongladda, S.; Suputtamongkol, Y. HLA-DRB1 and HLA-DQB1 Are Associated with Adult-Onset Immunodeficiency with Acquired Anti-Interferon-Gamma Autoantibodies. *PLoS ONE* **2015**, *10*, e0128481. [\[CrossRef\]](#)

247. Caruso, A.; Foresti, I.; Gribaudo, G.; Bonfanti, C.; Pollara, P.; Dolei, A.; Landolfo, S.; Turano, A. Anti-interferon-gamma antibodies in sera from HIV infected patients. *J. Biol. Regul. Homeost. Agents* **1989**, *3*, 8–12. [\[PubMed\]](#)
248. Zeng, W.; Tang, M.; Yang, M.; Fang, G.; Tang, S.; Zhang, J. Intravenous Cyclophosphamide Therapy for Anti-IFN- $\gamma$  Autoantibody-Associated *Talaromyces marneffei* Infection. *Open Forum Infect. Dis.* **2022**, *9*, ofac612. [\[CrossRef\]](#)
249. Ochoa, S.; Ding, L.; Kreuzburg, S.; Treat, J.; Holland, S.M.; Zerbe, C.S. Daratumumab (Anti-CD38) for Treatment of Disseminated Nontuberculous Mycobacteria in a Patient With Anti-Interferon- $\gamma$  Autoantibodies. *Clin. Infect. Dis.* **2021**, *72*, 2206–2208. [\[CrossRef\]](#)
250. Kwon-Chung, K.J.; Bennett, J.E.; Wickes, B.L.; Meyer, W.; Cuomo, C.A.; Wollenburg, K.R.; Bicanic, T.A.; Castañeda, E.; Chang, Y.C.; Chen, J.; et al. The Case for Adopting the “Species Complex” Nomenclature for the Etiologic Agents of Cryptococcosis. *mSphere* **2017**, *2*, e00357-16. [\[CrossRef\]](#)
251. Hagen, F.; Lumbsch, H.T.; Arsenijevic, V.A.; Badali, H.; Bertout, S.; Billmyre, R.B.; Bragulat, M.R.; Cabañes, F.J.; Carbia, M.; Chakrabarti, A.; et al. Importance of Resolving Fungal Nomenclature: The Case of Multiple Pathogenic Species in the *Cryptococcus* Genus. *mSphere* **2017**, *2*, e00238-17. [\[CrossRef\]](#)
252. Yang, D.-H.; England, M.R.; Salvator, H.; Anjum, S.; Park, Y.-D.; Marr, K.A.; Chu, L.A.; Govender, N.P.; Lockhart, S.R.; Desnos-Ollivier, M.; et al. *Cryptococcus gattii* Species Complex as an Opportunistic Pathogen: Underlying Medical Conditions Associated with the Infection. *Mbio* **2021**, *12*, e02708-21. [\[CrossRef\]](#) [\[PubMed\]](#)
253. Rezai, M.S.; Khotael, G.; Kheirkhah, M.; Hedayat, T.; Geramishoar, M.M.; Mahjoub, F. Cryptococcosis and Deficiency of Interleukin12r. *Pediatr. Infect. Dis. J.* **2008**, *27*, 673. [\[CrossRef\]](#) [\[PubMed\]](#)
254. Jirapongsananuruk, O.; Luangwedchakarn, V.; Niemela, J.E.; Pacharn, P.; Visitsunthorn, N.; Thepthai, C.; Vichyanond, P.; Piboonpocanun, S.; Fleisher, T.A. Cryptococcal osteomyelitis in a child with a novel compound mutation of the IL12RB1 gene. *Asian Pac. J. Allergy Immunol.* **2012**, *30*, 79. [\[PubMed\]](#)
255. Sood, V.; Ramachandran, R.; Paliana, R.; Prabhakar, A.; Inamdar, N.; Pattanashetti, N.; Joshi, V.; Sharma, M.; Rawat, A.; Kohli, H.; et al. Disseminated Cryptococcosis in a Deceptively Immunocompetent Adolescent. *Int. J. Rare Dis. Disord.* **2019**, *2*, 013.
256. Jo, E.K.; Kim, H.S.; Lee, M.Y.; Iseki, M.; Lee, J.H.; Song, C.H.; Park, J.K.; Hwang, T.J.; Kook, H. X-linked Hyper-IgM Syndrome Associated with *Cryptosporidium parvum* and *Cryptococcus neoformans* Infections: The First Case with Molecular Diagnosis in Korea. *J. Korean Med. Sci.* **2002**, *17*, 116–120. [\[CrossRef\]](#)
257. Levy, J.; Espanol-Boren, T.; Thomas, C.; Fischer, A.; Tovo, P.; Bordigoni, P.; Resnick, I.; Fasth, A.; Baer, M.; Gomez, L.; et al. Clinical spectrum of X-linked hyper-IgM syndrome. *J. Pediatr.* **1997**, *131*, 47–54. [\[CrossRef\]](#)
258. Mitsui-Sekinaka, K.; Imai, K.; Sato, H.; Tomizawa, D.; Kajiwarra, M.; Nagasawa, M.; Morio, T.; Nonoyama, S. Clinical features and hematopoietic stem cell transplantations for CD40 ligand deficiency in Japan. *J. Allergy Clin. Immunol.* **2015**, *136*, 1018–1024. [\[CrossRef\]](#)
259. França, T.T.; Leite, L.F.B.; Maximo, T.A.; Lambert, C.G.L.; Zurro, N.B.; Forte, W.C.N.; Condino-Neto, A. A Novel de Novo Mutation in the CD40 Ligand Gene in a Patient With a Mild X-Linked Hyper-IgM Phenotype Initially Diagnosed as CVID: New Aspects of Old Diseases. *Front. Pediatr.* **2018**, *6*, 130. [\[CrossRef\]](#)
260. Romani, L.; Williamson, P.R.; Di Cesare, S.; Di Matteo, G.; De Luca, M.; Carsetti, R.; Figà-Talamanca, L.; Cancrini, C.; Rossi, P.; Finocchi, A. Cryptococcal Meningitis and Post-Infectious Inflammatory Response Syndrome in a Patient With X-Linked Hyper IgM Syndrome: A Case Report and Review of the Literature. *Front. Immunol.* **2021**, *12*, 2757. [\[CrossRef\]](#)
261. Malheiro, L.; Lazzara, D.; Xerinda, S.; Pinheiro, M.D.; Sarmiento, A. Cryptococcal meningoencephalitis in a patient with hyper immunoglobulin M (IgM) syndrome: A case report. *BMC Res. Notes* **2014**, *7*, 1–5. [\[CrossRef\]](#)
262. Al-Banyan, M.; Alqahtani, A.; Sheikh, F.; Khaliq, A.M.R.; Al Rayes, H.; Al-Tarifi, A.; Al-Saud, B.; Arnaout, R.K. Cryptococcal Meningoencephalitis in a Patient with Hyper IgM Syndrome Due to CD40 Deficiency: Case Report and Literature Review. *Am. J. Infect. Dis.* **2019**, *15*, 24–28. [\[CrossRef\]](#)
263. Françoise, U.; Lafont, E.; Suarez, F.; Lanternier, F.; Lortholary, O. Disseminated Cryptococcosis in a Patient with CD40 Ligand Deficiency. *J. Clin. Immunol.* **2022**, *42*, 1622–1625. [\[CrossRef\]](#) [\[PubMed\]](#)
264. Nemoto, K.; Kawanami, T.; Hoshina, T.; Ishimura, M.; Yamasaki, K.; Okada, S.; Kanegane, H.; Yatera, K.; Kusuhashi, K. Impaired B-Cell Differentiation in a Patient With STAT1 Gain-of-Function Mutation. *Front. Immunol.* **2020**, *11*, 557521. [\[CrossRef\]](#)
265. Zhang, W.; Chen, X.; Gao, G.; Xing, S.; Zhou, L.; Tang, X.; Zhao, X.; An, Y. Clinical Relevance of Gain- and Loss-of-Function Germline Mutations in STAT1: A Systematic Review. *Front. Immunol.* **2021**, *12*, 654406. [\[CrossRef\]](#)
266. Grimbacher, B.; Holland, S.M.; Gallin, J.I.; Greenberg, F.; Hill, S.C.; Malech, H.L.; Miller, J.A.; O’Connell, A.C.; Puck, J.M. Hyper-IgE Syndrome with Recurrent Infections—An Autosomal Dominant Multisystem Disorder. *N. Engl. J. Med.* **1999**, *340*, 692–702. [\[CrossRef\]](#)
267. Holland, S.; DeLeo, F.R.; Elloumi, H.Z.; Hsu, A.P.; Uzel, G.; Brodsky, N.; Freeman, A.F.; Demidowich, A.; Davis, J.; Turner, M.L.C.; et al. STAT3 Mutations in the Hyper-IgE Syndrome. *N. Engl. J. Med.* **2007**, *357*, 1608–1619. [\[CrossRef\]](#)
268. Rujirachun, P.; Sangwongwanich, J.; Chayakulkeeree, M. Triple infection with *Cryptococcus*, varicella-zoster virus, and *Mycobacterium abscessus* in a patient with anti-interferon-gamma autoantibodies: A case report. *BMC Infect. Dis.* **2020**, *20*, 232. [\[CrossRef\]](#)
269. Chetchotisakd, P.; Anunnatsiri, S.; Nithichanon, A.; Lertmemongkolchai, G. Cryptococcosis in Anti-Interferon-Gamma Autoantibody-Positive Patients: A Different Clinical Manifestation from HIV-Infected Patients. *Jpn. J. Infect. Dis.* **2017**, *70*, 69–74. [\[CrossRef\]](#)

270. Rosen, L.B.; Freeman, A.F.; Yang, L.M.; Jutivorakool, K.; Olivier, K.N.; Angkasekwinai, N.; Suputtamongkol, Y.; Bennett, J.E.; Pyrgos, V.; Williamson, P.R.; et al. Anti-GM-CSF Autoantibodies in Patients with Cryptococcal Meningitis. *J. Immunol.* **2013**, *190*, 3959–3966. [\[CrossRef\]](#) [\[PubMed\]](#)
271. Saijo, T.; Chen, J.; Chen, S.C.A.; Rosen, L.B.; Yi, J.; Sorrell, T.; Bennett, J.E.; Holland, S.M.; Browne, S.K.; Kwon-Chung, K.J. Anti-Granulocyte-Macrophage Colony-Stimulating Factor Autoantibodies Are a Risk Factor for Central Nervous System Infection by *Cryptococcus gattii* in Otherwise Immunocompetent Patients. *Mbio* **2014**, *5*, e00912–14. [\[CrossRef\]](#) [\[PubMed\]](#)
272. Trapnell, B.C.; Nakata, K.; Bonella, F.; Campo, I.; Griesse, M.; Hamilton, J.; Wang, T.; Morgan, C.; Cottin, V.; McCarthy, C. Pulmonary alveolar proteinosis. *Nat. Rev. Dis. Prim.* **2019**, *5*, 16. [\[CrossRef\]](#)
273. Rosen, S.H.; Castleman, B.; Liebow, A.A.; Enzinger, F.M.; Hunt, R.T.N. Pulmonary Alveolar Proteinosis. *N. Engl. J. Med.* **1958**, *258*, 1123–1142. [\[CrossRef\]](#) [\[PubMed\]](#)
274. Sunderland, W.A.; Campbell, R.A.; Edwards, M.J. Pulmonary alveolar proteinosis and pulmonary cryptococcosis in an adolescent boy. *J. Pediatr.* **1972**, *80*, 450–456. [\[CrossRef\]](#)
275. Lee, Y.C.G.; Chew, G.T.; Robinson, B.W.S. Pulmonary and meningeal cryptococcosis in pulmonary alveolar proteinosis. *Aust. N. Z. J. Med.* **1999**, *29*, 843–844. [\[CrossRef\]](#) [\[PubMed\]](#)
276. Punatar, A.D.; Kusne, S.; Blair, J.E.; Seville, M.T.; Vikram, H.R. Opportunistic infections in patients with pulmonary alveolar proteinosis. *J. Infect.* **2012**, *65*, 173–179. [\[CrossRef\]](#)
277. Ataya, A.; Knight, V.; Carey, B.C.; Lee, E.; Tarling, E.J.; Wang, T. The Role of GM-CSF Autoantibodies in Infection and Autoimmune Pulmonary Alveolar Proteinosis: A Concise Review. *Front. Immunol.* **2021**, *12*, 4958. [\[CrossRef\]](#) [\[PubMed\]](#)
278. Berthou, C.; Mailhe, M.; Vély, F.; Gauthier, C.; Mège, J.-L.; Lagier, J.-C.; Melenotte, C. Granulocyte Macrophage Colony-Stimulating Factor-Specific Autoantibodies and Cerebral Nocardia With Pulmonary Alveolar Proteinosis. *Open Forum Infect. Dis.* **2021**, *8*, ofaa612. [\[CrossRef\]](#)
279. Rosen, L.B.; Pereira, N.R.; Figueiredo, C.; Fiske, L.C.; Ressler, R.A.; Hong, J.C.; Gregg, K.S.; Henry, T.L.; Pak, K.J.; Baumgarten, K.L.; et al. Nocardia-Induced Granulocyte Macrophage Colony-Stimulating Factor Is Neutralized by Autoantibodies in Disseminated/Extrapulmonary Nocardiosis. *Clin. Infect. Dis.* **2015**, *60*, 1017–1025. [\[CrossRef\]](#)
280. Garrido, L.; Mata-Essayag, S.; de Capriles, C.H.; Landaeta, M.E.; Pacheco, I.; Fuentes, Z. Pulmonary histoplasmosis: Unusual histopathologic findings. *Pathol.-Res. Pract.* **2006**, *202*, 373–378. [\[CrossRef\]](#)
281. Edman, J.C.; Kovacs, J.A.; Masur, H.; Santi, D.V.; Elwood, H.J.; Sogin, M.L. Ribosomal RNA sequence shows *Pneumocystis carinii* to be a member of the Fungi. *Nature* **1988**, *334*, 519–522. [\[CrossRef\]](#)
282. Cissé, O.H.; Hauser, P.M. Genomics and evolution of *Pneumocystis* species. *Infect. Genet. Evol.* **2018**, *65*, 308–320. [\[CrossRef\]](#) [\[PubMed\]](#)
283. Van der Meer, G.; Brug, S.L. Infection à *Pneumocystis* chez l’homme et chez les animaux. *Ann. Soc. Belg Med. Trop.* **1942**, *22*, 301–309.
284. Sternberg, S.D.; Rosenthal, J.H. Interstitial plasma cell pneumonia. *J. Pediatr.* **1955**, *46*, 380–393. [\[CrossRef\]](#) [\[PubMed\]](#)
285. Williamson, A.P.; Montgomery, J.R.; South, M.A.; Wilson, R. A Special Report: Four-year Study of a Boy with Combined Immune Deficiency Maintained in Strict Reverse Isolation from Birth. *Pediatr. Res.* **1977**, *11*, 63–64. [\[CrossRef\]](#) [\[PubMed\]](#)
286. Leggiadro, R.J.; Winkelstein, J.A.; Hughes, W.T. Prevalence of *Pneumocystis carinii* pneumonitis in severe combined immunodeficiency. *J. Pediatr.* **1981**, *99*, 96–98. [\[CrossRef\]](#) [\[PubMed\]](#)
287. Lauzon, D.; Delage, G.; Brochu, P.; Michaud, J.; Jasmin, G.; Joncas, J.H.; Lapointe, N. Pathogens in children with severe combined immune deficiency disease or AIDS. *Can. Med. Assoc. J.* **1986**, *135*, 33–38.
288. Kaposi’s sarcoma and *Pneumocystis* pneumonia among homosexual men—New York City and California. *MMWR Morb. Mortal. Wkly. Rep.* **1981**, *30*, 305–308.
289. Gottlieb, M.S.; Schroff, R.; Schanker, H.M.; Weisman, J.D.; Fan, P.T.; Wolf, R.A.; Saxon, A. *Pneumocystis carinii* Pneumonia and Mucosal Candidiasis in Previously Healthy Homosexual Men. *N. Engl. J. Med.* **1981**, *305*, 1425–1431. [\[CrossRef\]](#)
290. Siegal, F.P.; Lopez, C.; Hammer, G.S.; Brown, A.E.; Kornfeld, S.J.; Gold, J.; Hassett, J.; Hirschman, S.Z.; Cunningham-Rundles, C.; Adelsberg, B.R.; et al. Severe Acquired Immunodeficiency in Male Homosexuals, Manifested by Chronic Perianal Ulcerative Herpes Simplex Lesions. *N. Engl. J. Med.* **1981**, *305*, 1439–1444. [\[CrossRef\]](#)
291. Pifer, L.L.; Hughes, W.T.; Stagno, S.; Woods, D. *Pneumocystis carinii* Infection: Evidence for High Prevalence in Normal and Immunosuppressed Children. *Pediatrics* **1978**, *61*, 35–41. [\[CrossRef\]](#)
292. Vargas, S.L.; Hughes, W.T.; Santolaya, M.E.; Ulloa, A.V.; Ponce, C.A.; Cabrera, C.E.; Cumsille, F.; Gigliotti, F. Search for Primary Infection by *Pneumocystis carinii* in a Cohort of Normal, Healthy Infants. *Clin. Infect. Dis.* **2001**, *32*, 855–861. [\[CrossRef\]](#) [\[PubMed\]](#)
293. Vera, C.; Rueda, Z.V. Transmission and Colonization of *Pneumocystis jirovecii*. *J. Fungi* **2021**, *7*, 979. [\[CrossRef\]](#) [\[PubMed\]](#)
294. Berrington, J.; Flood, T.J.; Abinun, M.; Galloway, A.; Cant, A.J. Unsuspected *Pneumocystis carinii* pneumonia at presentation of severe primary immunodeficiency. *Arch. Dis. Child.* **2000**, *82*, 144–147. [\[CrossRef\]](#) [\[PubMed\]](#)
295. Hanna, S.; Etzioni, A. MHC class I and II deficiencies. *J. Allergy Clin. Immunol.* **2014**, *134*, 269–275. [\[CrossRef\]](#)
296. Hsu, A.P. Not too little, not too much: The impact of mutation types in Wiskott-Aldrich syndrome and RAC2 patients. *Clin. Exp. Immunol.* **2023**. [\[CrossRef\]](#)
297. Hirbod-Mobarakeh, A.; Aghamohammadi, A.; Rezaei, N. Immunoglobulin class switch recombination deficiency type 1 or CD40 ligand deficiency: From bedside to bench and back again. *Expert Rev. Clin. Immunol.* **2014**, *10*, 91–105. [\[CrossRef\]](#)

298. Cetin, E.; Lee, E.Y. *Pneumocystis carinii* pneumonia in an infant with hypogammaglobulinemia. *Pediatr. Radiol.* **2006**, *37*, 329. [\[CrossRef\]](#)
299. Zhang, F.Z.; Yuan, J.X.; Qin, L.; Tang, L.F. Pulmonary Alveolar Proteinosis Due to *Pneumocystis carinii* in Type 1 Hyper-IgM Syndrome: A Case Report. *Front. Pediatr.* **2020**, *8*, 264. [\[CrossRef\]](#)
300. Elenga, N.; Dulorme, F.; Basile, G.D.S.; Mahamat, A. *Pneumocystis jirovecii* Pneumonia Revealing De Novo Mutation Causing X-Linked Hyper-IgM Syndrome in an Infant Male. The First Case Reported From French Guiana. *J. Pediatr. Hematol.* **2012**, *34*, 528–530. [\[CrossRef\]](#)
301. Leone, V.; Tommasini, A.; Andolina, M.; Runti, G.; De Vonderweid, U.; Campello, C.; Notarangelo, L.; Ventura, A. Elective bone marrow transplantation in a child with X-linked hyper-IgM syndrome presenting with acute respiratory distress syndrome. *Bone Marrow Transplant.* **2002**, *30*, 49–52. [\[CrossRef\]](#)
302. Huang, S.H.; Meng, X.Y.; Bai, Z.J.; Li, Y.; Wu, S.Y. X-Linked Hyper IgM Syndrome Manifesting as Recurrent *Pneumocystis jirovecii* Pneumonia: A Case Report. *J. Trop. Pediatr.* **2020**, *66*, 648–654. [\[CrossRef\]](#) [\[PubMed\]](#)
303. Kim, D.; Shin, J.A.; Han, S.B.; Chung, N.-G.; Jeong, D.C. *Pneumocystis jirovecii* pneumonia as an initial manifestation of hyper-IgM syndrome in an infant. *Medicine* **2019**, *98*, e14559. [\[CrossRef\]](#)
304. Li, H.; Cao, Y.; Ma, J.; Li, C. X-linked hyper IgM syndrome with severe eosinophilia: A case report and review of the literature. *BMC Pediatr.* **2022**, *22*, 178. [\[CrossRef\]](#)
305. Li, J.; Miao, H.; Wu, L.; Fang, Y. Interstitial pneumonia as the initial presentation in an infant with a novel mutation of CD40 ligand-associated X-linked hyper-IgM syndrome. *Medicine* **2020**, *99*, e20505. [\[CrossRef\]](#)
306. Cunningham, C.K.; Bonville, C.A.; Ochs, H.D.; Seyama, K.; John, P.A.; Rotbart, H.A.; Weiner, L.B. Enteroviral meningoencephalitis as a complication of X-linked hyper IgM syndrome. *J. Pediatr.* **1999**, *134*, 584–588. [\[CrossRef\]](#)
307. Milledge, J.; Kakakios, A.; Gillis, J.; Fitzgerald, D.A. *Pneumocystis carinii* pneumonia as a presenting feature of X-linked hyper-IgM syndrome. *J. Paediatr. Child Health* **2003**, *39*, 704–706. [\[CrossRef\]](#)
308. Thomas, C.; Basile, G.D.S.; Le Deist, F.; Theophile, D.; Benkerrou, M.; Haddad, E.; Blanche, S.; Fischer, A. Correction of X-Linked Hyper-IgM Syndrome by Allogeneic Bone Marrow Transplantation. *N. Engl. J. Med.* **1995**, *333*, 426–429. [\[CrossRef\]](#)
309. Lee, W.-I.; Torgerson, T.R.; Schumacher, M.J.; Yel, L.; Zhu, Q.; Ochs, H.D. Molecular analysis of a large cohort of patients with the hyper immunoglobulin M (IgM) syndrome. *Blood* **2005**, *105*, 1881–1890. [\[CrossRef\]](#)
310. Pickell, J.A.; Gallagher, J.L.; Chang, Y.; Patel, N.C. Normal CD40L Expression in an Infant with X-Linked Hyper IgM Syndrome By Gene Sequencing. *J. Allergy Clin. Immunol.* **2015**, *135*, AB12. [\[CrossRef\]](#)
311. Dupuis-Girod, S.; Corradini, N.; Hadj-Rabia, S.; Fournet, J.-C.; Faivre, L.; Le Deist, F.; Durand, P.; Döfninger, R.; Smahi, A.; Israel, A.; et al. Osteopetrosis, Lymphedema, Anhidrotic Ectodermal Dysplasia, and Immunodeficiency in a Boy and Incontinentia Pigmenti in His Mother. *Pediatrics* **2002**, *109*, e97. [\[CrossRef\]](#) [\[PubMed\]](#)
312. Salt, B.H.; Niemela, J.E.; Pandey, R.; Hanson, E.P.; Deering, R.P.; Quinones, R.; Jain, A.; Orange, J.S.; Gelfand, E.W. IKBKG (nuclear factor- $\kappa$ B essential modulator) mutation can be associated with opportunistic infection without impairing Toll-like receptor function. *J. Allergy Clin. Immunol.* **2008**, *121*, 976–982. [\[CrossRef\]](#)
313. Carlberg, V.M.; Lofgren, S.M.; Mann, J.A.; Austin, J.P.; Nolt, D.; Shereck, E.B.; Saldaña, B.D.; Zonana, J.; Krol, A.L. Hypohidrotic Ectodermal Dysplasia, Osteopetrosis, Lymphedema, and Immunodeficiency in an Infant with Multiple Opportunistic Infections. *Pediatr. Dermatol.* **2014**, *31*, 716–721. [\[CrossRef\]](#) [\[PubMed\]](#)
314. Janssen, R.; Van Wengen, A.; Hoeve, M.A.; Dam, M.T.; Van Der Burg, M.; Van Dongen, J.; van de Vosse, E.; Van Tol, M.; Bredius, R.; Ottenhoff, T.H.; et al. The Same I $\kappa$ B $\alpha$  Mutation in Two Related Individuals Leads to Completely Different Clinical Syndromes. *J. Exp. Med.* **2004**, *200*, 559–568. [\[CrossRef\]](#) [\[PubMed\]](#)
315. Maffucci, P.; Fillion, C.A.; Boisson, B.; Itan, Y.; Shang, L.; Casanova, J.-L.; Cunningham-Rundles, C. Genetic Diagnosis Using Whole Exome Sequencing in Common Variable Immunodeficiency. *Front. Immunol.* **2016**, *7*, 220. [\[CrossRef\]](#)
316. Mandola, A.B.; Sharfe, N.; Nagdi, Z.; Dadi, H.; Vong, L.; Merico, D.; Ngan, B.; Reid, B.; Roifman, C.M. Combined immunodeficiency caused by a novel homozygous NFKB1 mutation. *J. Allergy Clin. Immunol.* **2021**, *147*, 727–733.e2. [\[CrossRef\]](#)
317. Boutboul, D.; Kuehn, H.S.; Van De Wyngaert, Z.; Niemela, J.E.; Callebaut, I.; Stoddard, J.; Lenoir, C.; Barlogis, V.; Farnarier, C.; Vely, F.; et al. Dominant-negative IKZF1 mutations cause a T, B, and myeloid cell combined immunodeficiency. *J. Clin. Investig.* **2018**, *128*, 3071–3087. [\[CrossRef\]](#) [\[PubMed\]](#)
318. Yamashita, M.; Morio, T. AIOLOS Variants Causing Immunodeficiency in Human and Mice. *Front. Immunol.* **2022**, *13*, 1502. [\[CrossRef\]](#)
319. González-Lara, M.F.; Wisniewski-Yáñez, A.; Pérez-Patrigéon, S.; Hsu, A.P.; Holland, S.M.; Cuellar-Rodríguez, J.M. *Pneumocystis jirovecii* pneumonia and GATA2 deficiency: Expanding the spectrum of the disease. *J. Infect.* **2017**, *74*, 425–427. [\[CrossRef\]](#)
320. Gaspar, H.B.; Ferrando, M.; Caragol, I.; Hernandez, M.; Bertran, J.M.; De Gracia, X.; Lester, T.; Kinnon, C.; Ashton, E.; Espanol, T. Kinase mutant Btk results in atypical X-linked agammaglobulinemia phenotype. *Clin. Exp. Immunol.* **2000**, *120*, 346–350. [\[CrossRef\]](#)
321. Jongco, A.M.; Gough, J.D.; Sarnataro, K.; Rosenthal, D.W.; Moreau, J.; Ponda, P.; Bonagura, V.R. X-linked agammaglobulinemia presenting as polymicrobial pneumonia, including *Pneumocystis jirovecii*. *Ann. Allergy Asthma Immunol.* **2014**, *112*, 74–75.e2. [\[CrossRef\]](#)
322. Brunet, B.A.; Rodriguez, R. Unusual presentation of combined immunodeficiency in a child with homozygous DOCK8 mutation. *Ann. Allergy Asthma Immunol.* **2017**, *119*, 294–295. [\[CrossRef\]](#)

323. Denning, D.W.; Cadranel, J.; Beigelman-Aubry, C.; Ader, F.; Chakrabarti, A.; Blot, S.; Ullmann, A.J.; Dimopoulos, G.; Lange, C.; Dimopoulos, C.L. on behalf of the European Society for Clinical Microbiology and Infectious Diseases and European Respiratory Society. Chronic pulmonary aspergillosis: Rationale and clinical guidelines for diagnosis and management. *Eur. Respir. J.* **2016**, *47*, 45–68. [\[CrossRef\]](#)
324. Ullmann, A.J.; Aguado, J.M.; Arikan-Akdoglu, S.; Denning, D.W.; Groll, A.H.; Lagrou, K.; Lass-Flörl, C.; Lewis, R.E.; Munoz, P.; Verweij, P.E.; et al. Diagnosis and management of Aspergillus diseases: Executive summary of the 2017 ESCMID-ECMM-ERS guideline. *Clin. Microbiol. Infect.* **2018**, *24*, e1–e38. [\[CrossRef\]](#) [\[PubMed\]](#)
325. Latgé, J.-P. *Aspergillus fumigatus* and Aspergillosis. *Clin. Microbiol. Rev.* **1999**, *12*, 310–350. [\[CrossRef\]](#)
326. Li, B.C.; Huh, S.M.; Prieto, M.D.; Hong, G.; Schwarz, C.; Moss, R.B.; Quon, B.S. Biomarkers for the Diagnosis of Allergic Bronchopulmonary Aspergillosis in Cystic Fibrosis: A Systematic Review and Meta-Analysis. *J. Allergy Clin. Immunol. Pract.* **2021**, *9*, 1909–1930.e4. [\[CrossRef\]](#) [\[PubMed\]](#)
327. Agarwal, R.; Khan, A.; Aggarwal, A.N.; Gupta, D. Link between CFTR mutations and ABPA: A systematic review and meta-analysis. *Mycoses* **2011**, *55*, 357–365. [\[CrossRef\]](#) [\[PubMed\]](#)
328. Lebecque, P.; Pepermans, X.; Marchand, E.; Leonard, A.; Leal, T. ABPA in adulthood: A CFTR-related disorder. *Thorax* **2011**, *66*, 540–541. [\[CrossRef\]](#) [\[PubMed\]](#)
329. Vaid, M.; Kaur, S.; Sambatakou, H.; Madan, T.; Denning, D.W.; Sarma, P.U. Distinct alleles of mannose-binding lectin (MBL) and surfactant proteins A (SP-A) in patients with chronic cavitary pulmonary aspergillosis and allergic bronchopulmonary aspergillosis. *Clin. Chem. Lab. Med.* **2007**, *45*, 183–186. [\[CrossRef\]](#)
330. Carvalho, A.; Pasqualotto, A.C.; Pitzurra, L.; Romani, L.; Denning, D.W.; Rodrigues, F. Polymorphisms in Toll-Like Receptor Genes and Susceptibility to Pulmonary Aspergillosis. *J. Infect. Dis.* **2008**, *197*, 618–621. [\[CrossRef\]](#) [\[PubMed\]](#)
331. Overton, N.L.D.; Brakhage, A.A.; Thywissen, A.; Denning, D.W.; Bowyer, P. Mutations in EEA1 are associated with allergic bronchopulmonary aspergillosis and affect phagocytosis of *Aspergillus fumigatus* by human macrophages. *PLoS ONE* **2018**, *13*, e0185706. [\[CrossRef\]](#)
332. Chaudhary, N.; Datta, K.; Askin, F.B.; Staab, J.F.; Marr, K.A. Cystic Fibrosis Transmembrane Conductance Regulator Regulates Epithelial Cell Response to *Aspergillus* and Resultant Pulmonary Inflammation. *Am. J. Respir. Crit. Care Med.* **2012**, *185*, 301–310. [\[CrossRef\]](#)
333. Hartl, D.; Griesse, M.; Kappler, M.; Zissel, G.; Reinhardt, D.; Rebhan, C.; Schendel, D.J.; Krauss-Etschmann, S. Pulmonary TH2 response in *Pseudomonas aeruginosa*-infected patients with cystic fibrosis. *J. Allergy Clin. Immunol.* **2006**, *117*, 204–211. [\[CrossRef\]](#) [\[PubMed\]](#)
334. Knutsen, A.P.; Hutchinson, P.S.; Albers, G.M.; Consolino, J.; Smick, J.; Kurup, V.P. Increased sensitivity to IL-4 in cystic fibrosis patients with allergic bronchopulmonary aspergillosis. *Allergy* **2004**, *59*, 81–87. [\[CrossRef\]](#) [\[PubMed\]](#)
335. Roos, D. Chronic Granulomatous Disease. *Methods Mol. Biol.* **2019**, *1982*, 531–542. [\[CrossRef\]](#) [\[PubMed\]](#)
336. Vinh, D.C.; Shea, Y.R.; Sugui, J.A.; Parrilla-Castellar, E.R.; Freeman, A.F.; Campbell, J.W.; Pittaluga, S.; Jones, P.A.; Zelazny, A.; Kleiner, D.; et al. Invasive Aspergillosis Due to *Neosartorya udagawae*. *Clin. Infect. Dis.* **2009**, *49*, 102–111. [\[CrossRef\]](#)
337. Hasui, M.; The Study Group of Phagocyte Disorders of Japan. Chronic granulomatous disease in Japan: Incidence and natural history. *Pediatr. Int.* **1999**, *41*, 589–593. [\[CrossRef\]](#)
338. Soler-Palacin, P.; Margareto, C.; Llobet, P.; Asensio, O.; Hernandez-Gonzalez, M.; Caragol, I.; Español, T. Chronic granulomatous disease in pediatric patients: 25 years of experience. *Allergol. Immunopathol.* **2007**, *35*, 83–89. [\[CrossRef\]](#)
339. Vinh, D.C.; Shea, Y.R.; Jones, P.A.; Freeman, A.F.; Zelazny, A.; Holland, S.M. Chronic Invasive Aspergillosis caused by *Aspergillus viridinutans*. *Emerg. Infect. Dis.* **2009**, *15*, 1292–1294. [\[CrossRef\]](#)
340. Sabino, R.; Ferreira, J.A.; Moss, R.B.; Valente, J.; Verissimo, C.; Carolino, E.; Clemons, K.V.; Everson, C.; Banaei, N.; Penner, J.; et al. Molecular epidemiology of *Aspergillus* collected from cystic fibrosis patients. *J. Cyst. Fibros.* **2015**, *14*, 474–481. [\[CrossRef\]](#)
341. Talbot, J.J.; Barrs, V.R. One-health pathogens in the *Aspergillus viridinutans* complex. *Med. Mycol.* **2018**, *56*, 1–12. [\[CrossRef\]](#)
342. Seyedmousavi, S.; Lionakis, M.S.; Parta, M.; Peterson, S.W.; Kwon-Chung, K.J. Emerging *Aspergillus* Species Almost Exclusively Associated with Primary Immunodeficiencies. *Open Forum Infect. Dis.* **2018**, *5*, ofy213. [\[CrossRef\]](#)
343. Winkelstein, J.A.; Marino, M.C.; Johnston, R.B.; Boyle, J.; Curnutte, J.; Gallin, J.I.; Malech, H.; Holland, S.M.; Ochs, H.; Quie, P.; et al. Chronic Granulomatous Disease: Report on a National Registry of 368 Patients. *Medicine* **2000**, *79*, 155–169. [\[CrossRef\]](#) [\[PubMed\]](#)
344. Kobayashi, S.; Murayama, S.; Takanashi, S.; Takahashi, K.; Miyatsuka, S.; Fujita, T.; Ichinohe, S.; Koike, Y.; Kohagizawa, T.; Mori, H.; et al. Clinical features and prognoses of 23 patients with chronic granulomatous disease followed for 21 years by a single hospital in Japan. *Eur. J. Pediatr.* **2008**, *167*, 1389–1394. [\[CrossRef\]](#)
345. Martire, B.; Rondelli, R.; Soresina, A.; Pignata, C.; Broccoletti, T.; Finocchi, A.; Rossi, P.; Gattorno, M.; Rabusin, M.; Azzari, C.; et al. Clinical features, long-term follow-up and outcome of a large cohort of patients with Chronic Granulomatous Disease: An Italian multicenter study. *Clin. Immunol.* **2008**, *126*, 155–164. [\[CrossRef\]](#)
346. Tavakoli, M.; Rivero-Menendez, O.; Abastabar, M.; Hedayati, M.T.; Sabino, R.; Siopi, M.; Zarrinfar, H.; Nouripour-Sisakht, S.; Lee, H.; Valadan, R.; et al. Genetic diversity and antifungal susceptibility patterns of *Aspergillus nidulans* complex obtained from clinical and environmental sources. *Mycoses* **2020**, *63*, 78–88. [\[CrossRef\]](#) [\[PubMed\]](#)
347. Segal, B.H.; DeCarlo, E.S.; Kwon-Chung, K.J.; Malech, H.; Gallin, J.I.; Holland, S.M. *Aspergillus nidulans* Infection in Chronic Granulomatous Disease. *Medicine* **1998**, *77*, 345–354. [\[CrossRef\]](#) [\[PubMed\]](#)

348. Lee, M.J.; Liu, H.; Barker, B.M.; Snarr, B.D.; Gravelat, F.N.; Al Abdallah, Q.; Gavino, C.; Baistrocchi, S.R.; Ostapska, H.; Xiao, T.; et al. The Fungal Exopolysaccharide Galactosaminogalactan Mediates Virulence by Enhancing Resistance to Neutrophil Extracellular Traps. *PLoS Pathog.* **2015**, *11*, e1005187. [[CrossRef](#)]
349. Verweij, P.E.; Weemaes, C.M.; Curfs, J.H.A.J.; Bretagne, S.; Meis, J.F.G.M. Failure to Detect Circulating *Aspergillus* Markers in a Patient with Chronic Granulomatous Disease and Invasive Aspergillosis. *J. Clin. Microbiol.* **2000**, *38*, 3900–3901. [[CrossRef](#)]
350. Blumental, S.; Mouy, R.; Mahlaoui, N.; Bougnoux, M.-E.; Debré, M.; Beauté, J.; Lortholary, O.; Blanche, S.; Fischer, A. Invasive Mold Infections in Chronic Granulomatous Disease: A 25-Year Retrospective Survey. *Clin. Infect. Dis.* **2011**, *53*, e159–e169. [[CrossRef](#)]
351. Siddiqui, S.; Anderson, V.L.; Hilligoss, D.M.; Abinun, M.; Kuijpers, T.W.; Masur, H.; Witebsky, F.G.; Shea, Y.R.; Gallin, J.I.; Malech, H.; et al. Fulminant Mulch Pneumonitis: An Emergency Presentation of Chronic Granulomatous Disease. *Clin. Infect. Dis.* **2007**, *45*, 673–681. [[CrossRef](#)]
352. Ameratunga, R.; Woon, S.-T.; Vyas, J.; Roberts, S. Fulminant Mulch Pneumonitis in Undiagnosed Chronic Granulomatous Disease: A Medical Emergency. *Clin. Pediatr.* **2010**, *49*, 1143–1146. [[CrossRef](#)]
353. Maaloul, I.; Ben Ameer, S.; Chabchoub, I.; Kolsi, R.; Bahloul, M.; Kamoun, T.; Bouaziz, M.; Hachicha, M. Fulminant mulch pneumonitis in a previously healthy child. *Arch. Pediatr.* **2018**, *25*, 495–496. [[CrossRef](#)]
354. Cowan, J.; Pakhale, S.; Angel, J.B. Mulch pneumonitis in a patient with chronic granulomatous disease. *Cmaj* **2022**, *194*, E1510. [[CrossRef](#)] [[PubMed](#)]
355. Marciano, B.E.; Spalding, C.; Fitzgerald, A.; Mann, D.; Brown, T.; Osgood, S.; Yockey, L.; Darnell, D.N.; Barnhart, L.; Daub, J.; et al. Common Severe Infections in Chronic Granulomatous Disease. *Clin. Infect. Dis.* **2015**, *60*, 1176–1183. [[CrossRef](#)] [[PubMed](#)]
356. Matzaraki, V.; Beno, A.; Jaeger, M.; Gresnigt, M.S.; Keur, N.; Boahen, C.; Cunha, C.; Gonçalves, S.M.; Leite, L.; Lacerda, J.F.; et al. Genetic determinants of fungi-induced ROS production are associated with the risk of invasive pulmonary aspergillosis. *Redox Biol.* **2022**, *55*, 102391. [[CrossRef](#)]
357. Van De Geer, A.; Nieto-Patlán, A.; Kuhns, D.B.; Tool, A.T.; Arias, A.A.; Bouaziz, M.; De Boer, M.; Franco, J.L.; Gazendam, R.P.; Van Hamme, J.L.; et al. Inherited p40phox deficiency differs from classic chronic granulomatous disease. *J. Clin. Investig.* **2018**, *128*, 3957–3975. [[CrossRef](#)] [[PubMed](#)]
358. Belot, A.; Kasher, P.R.; Trotter, E.W.; Foray, A.-P.; Debaud, A.-L.; Rice, G.I.; Szykiewicz, M.; Zabot, M.-T.; Rouvet, I.; Bhaskar, S.S.; et al. Protein Kinase C $\delta$  Deficiency Causes Mendelian Systemic Lupus Erythematosus With B Cell-Defective Apoptosis and Hyperproliferation. *Arthritis Rheum.* **2013**, *65*, 2161–2171. [[CrossRef](#)]
359. Neehus, A.-L.; Moriya, K.; Nieto-Patlán, A.; Le Voyer, T.; Lévy, R.; Özen, A.; Karakoc-Aydiner, E.; Baris, S.; Yildiran, A.; Altundag, E.; et al. Impaired respiratory burst contributes to infections in PKC $\delta$ -deficient patients. *J. Exp. Med.* **2021**, *218*, e20210501. [[CrossRef](#)]
360. Neehus, A.-L.; Tuano, K.; Le Voyer, T.; Nandiwada, S.L.; Murthy, K.; Puel, A.; Casanova, J.-L.; Chinen, J.; Bustamante, J. Chronic Granulomatous Disease-Like Presentation of a Child with Autosomal Recessive PKC $\delta$  Deficiency. *J. Clin. Immunol.* **2022**, *42*, 1244–1253. [[CrossRef](#)] [[PubMed](#)]
361. Bustamante, J.; Arias, A.A.; Vogt, G.; Picard, C.; Galicia, L.B.; Prando, C.; Grant, A.V.; Marchal, C.C.; Hubeau, M.; Chapgier, A.; et al. Germline CYBB mutations that selectively affect macrophages in kindreds with X-linked predisposition to tuberculous mycobacterial disease. *Nat. Immunol.* **2011**, *12*, 213–221. [[CrossRef](#)]
362. Lowe, J.S.; Anderson, P.G. Chapter 10-Respiratory System. In *Stevens & Lowe's Human Histology*, 4th ed.; Lowe, J.S., Anderson, P.G., Eds.; Mosby: Philadelphia, PA, USA, 2015; pp. 166–185. [[CrossRef](#)]
363. Takemura, Y.; Goodson, P.; Bao, H.F.; Jain, L.; Helms, M.N. Rac1-mediated NADPH oxidase release of O $_2^-$  regulates epithelial sodium channel activity in the alveolar epithelium. *Am. J. Physiol. Cell Mol. Physiol.* **2010**, *298*, L509–L520. [[CrossRef](#)]
364. Sun, W.-K.; Lu, X.; Li, X.; Sun, Q.-Y.; Su, X.; Song, Y.; Sun, H.-M.; Shi, Y. Dectin-1 is inducible and plays a crucial role in *Aspergillus*-induced innate immune responses in human bronchial epithelial cells. *Eur. J. Clin. Microbiol. Infect. Dis.* **2012**, *31*, 2755–2764. [[CrossRef](#)] [[PubMed](#)]
365. Zysman-Colman, Z.N.; Kaspy, K.R.; Alizadehfar, R.; Nykamp, K.R.; Zariwala, M.A.; Knowles, M.R.; Vinh, D.C.; Shapiro, A.J. Nasal Nitric Oxide in Primary Immunodeficiency and Primary Ciliary Dyskinesia: Helping to Distinguish Between Clinically Similar Diseases. *J. Clin. Immunol.* **2019**, *39*, 216–224. [[CrossRef](#)]
366. Sehgal, I.S.; Dhooria, S.; Bal, A.; Agarwal, R. Allergic bronchopulmonary aspergillosis in an adult with Kartagener syndrome. *BMJ Case Rep.* **2015**, *2015*, bcr2015211493. [[CrossRef](#)] [[PubMed](#)]
367. Allaer, L.; Lejeune, S.; Mordacq, C.; Deschildre, A.; Thumerelle, C. Primary ciliary dyskinesia and fungal infections: Two cases of allergic bronchopulmonary aspergillosis in children. *Pediatr. Pulmonol.* **2022**, *57*, 1809–1813. [[CrossRef](#)] [[PubMed](#)]
368. Shah, A.; Shoemark, A.; MacNeill, S.J.; Bhaludin, B.; Rogers, A.; Bilton, D.; Hansell, D.M.; Wilson, R.; Loebinger, M.R. A longitudinal study characterising a large adult primary ciliary dyskinesia population. *Eur. Respir. J.* **2016**, *48*, 441–450. [[CrossRef](#)]
369. Jones, L.B.K.R.; McGrogan, P.; Flood, T.J.; Gennery, A.R.; Morton, L.; Thrasher, A.; Goldblatt, D.; Parker, L.; Cant, A.J. Special Article: Chronic granulomatous disease in the United Kingdom and Ireland: A comprehensive national patient-based registry. *Clin. Exp. Immunol.* **2008**, *152*, 211–218. [[CrossRef](#)]
370. Silliman, C.C.; Lawellin, D.W.; Lohr, J.A.; Rodgers, B.M.; Donowitz, L.G. Paecilomyces lilacinus infection in a child with chronic granulomatous disease. *J. Infect.* **1992**, *24*, 191–195. [[CrossRef](#)]

371. Wang, S.-M.; Shieh, C.-C.; Liu, C.-C. Successful treatment of *Paecilomyces variotii* splenic abscesses: A rare complication in a previously unrecognized chronic granulomatous disease child. *Diagn. Microbiol. Infect. Dis.* **2005**, *53*, 149–152. [\[CrossRef\]](#)
372. Lemaigre, C.; Suarez, F.; Martellosio, J.-P.; Barbarin, C.; Brunet, K.; Chomel, J.C.; Hainaut, E.; Rammaert, B.; Roblot, F.; Torregrosa-Diaz, J.M. Late Onset of Chronic Granulomatous Disease Revealed by *Paecilomyces lilacinus* Cutaneous Infection. *J. Clin. Immunol.* **2022**, *42*, 60–63. [\[CrossRef\]](#)
373. De Ravin, S.S.; Challipalli, M.; Anderson, V.; Shea, Y.R.; Marciano, B.; Hilligoss, D.; Marquesen, M.; DeCastro, R.; Liu, Y.-C.; Sutton, D.A.; et al. *Geosmithia argillacea*: An Emerging Cause of Invasive Mycosis in Human Chronic Granulomatous Disease. *Clin. Infect. Dis.* **2011**, *52*, e136–e143. [\[CrossRef\]](#)
374. Rawat, A.; Vignesh, P.; Sharma, A.; Shandilya, J.K.; Sharma, M.; Suri, D.; Gupta, A.; Gautam, V.; Ray, P.; Rudramurthy, S.M.; et al. Infection Profile in Chronic Granulomatous Disease: A 23-Year Experience from a Tertiary Care Center in North India. *J. Clin. Immunol.* **2017**, *37*, 319–328. [\[CrossRef\]](#) [\[PubMed\]](#)
375. De Ravin, S.S.; Parta, M.; Sutton, D.A.; Wickes, B.L.; Thompson, E.H.; Wiederhold, N.P.; Nakasone, K.K.; Alimchandani, M.; Oconnell, A.; Notarangelo, L.; et al. Paravertebral Mushroom: Identification of a Novel Species of *Phellinus* as a Human Pathogen in Chronic Granulomatous Disease. *J. Clin. Microbiol.* **2014**, *52*, 2726–2729. [\[CrossRef\]](#) [\[PubMed\]](#)
376. Shigemura, T.; Nakazawa, Y.; Amano, Y.; Sudo, A.; Watanabe, M.; Kobayashi, M.; Koike, K.; Agematsu, K.; Nishimura, K. Subcutaneous abscess due to the basidiomycete *Phellinus mori* in a patient with chronic granulomatous disease. *Infection* **2015**, *43*, 371–375. [\[CrossRef\]](#) [\[PubMed\]](#)
377. Haidar, G.; Zerbe, C.S.; Cheng, M.; Zelazny, A.M.; Holland, S.M.; Sheridan, K.R. *Phellinus* species: An emerging cause of refractory fungal infections in patients with X-linked chronic granulomatous disease. *Mycoses* **2017**, *60*, 155–160. [\[CrossRef\]](#) [\[PubMed\]](#)
378. Hek, L.G.V.; Verweij, P.E.; Weemaes, C.M.; van Dalen, R.; Yntema, J.-B.; Meis, J.F. Successful Treatment with Voriconazole of Invasive Aspergillosis in Chronic Granulomatous Disease. *Am. J. Respir. Crit. Care Med.* **1998**, *157*, 1694–1696. [\[CrossRef\]](#)
379. Emmendorffer, A.; Lohmann-Matthes, M.-L.; Roesler, J. Kinetics of transfused neutrophils in peripheral blood and BAL fluid of a patient with variant X-linked chronic granulomatous disease. *Eur. J. Haematol.* **1991**, *47*, 246–252. [\[CrossRef\]](#)
380. Marciano, B.E.; Allen, E.S.; Conry-Cantilena, C.; Kristosturyan, E.; Klein, H.G.; Fleisher, T.A.; Holland, S.M.; Malech, H.; Rosenzweig, S.D. Granulocyte transfusions in patients with chronic granulomatous disease and refractory infections: The NIH experience. *J. Allergy Clin. Immunol.* **2017**, *140*, 622–625. [\[CrossRef\]](#)
381. Bielorai, B.; Toren, A.; Wolach, B.; Mandel, M.; Golan, H.; Neumann, Y.; Kaplinisky, C.; Weintraub, M.; Keller, N.; Amariglio, N.; et al. Successful treatment of invasive aspergillosis in chronic granulomatous disease by granulocyte transfusions followed by peripheral blood stem cell transplantation. *Bone Marrow Transplant.* **2000**, *26*, 1025–1028. [\[CrossRef\]](#)
382. Borge, J.P.D.; Theobald, M.N.; DeCastro, N.R.; Malech, H.L.; Leitman, S.; Kang, E.M. Successful Control of Preexistent Active Infection by Granulocyte Transfusions During Conditioning Induced Cytopenia In Patients with Chronic Granulomatous Disease Undergoing Hematopoietic Stem Cell Transplant. *Blood* **2010**, *116*, 1329–1339. [\[CrossRef\]](#)
383. Arnold, D.E.; Chellapandian, D.; Parikh, S.; Mallhi, K.; Marsh, R.A.; Heimall, J.R.; Grossman, D.; Chitty-Lopez, M.; Murguia-Favela, L.; Gennery, A.R.; et al. Granulocyte Transfusions in Patients with Chronic Granulomatous Disease Undergoing Hematopoietic Cell Transplantation or Gene Therapy. *J. Clin. Immunol.* **2022**, *42*, 1026–1035. [\[CrossRef\]](#)
384. Ozsahin, H.; von Planta, M.; Müller, I.; Steinert, H.C.; Nadal, D.; Lauener, R.; Tuchscheid, P.; Willi, U.V.; Ozsahin, M.; Crompton, N.E.; et al. Successful treatment of invasive aspergillosis in chronic granulomatous disease by bone marrow transplantation, granulocyte colony-stimulating factor-mobilized granulocytes, and liposomal amphotericin-B. *Blood* **1998**, *92*, 2719–2724. [\[CrossRef\]](#) [\[PubMed\]](#)
385. Dedieu, C.; Landwehr-Kenzel, S.; Thee, S.; Oevermann, L.; Voigt, S.; Marggraf, K.; Schulte, J.; Lau, S.; Roesler, J.; Kuehl, J.-S.; et al. Hematopoietic Stem Cell Transplantation Cures Therapy-refractory Aspergillosis in Chronic Granulomatous Disease. *Pediatr. Infect. Dis. J.* **2021**, *40*, 649–654. [\[CrossRef\]](#)
386. Shamberger, R.C.; Wohl, M.E.; Perez-Atayde, A.; Hendren, W.H. Pneumatocele complicating hyperimmunoglobulin E syndrome (Job's syndrome). *Ann. Thorac. Surg.* **1992**, *54*, 1206–1208. [\[CrossRef\]](#) [\[PubMed\]](#)
387. Hall, R.A.; Salhany, K.E.; Lebel, E.; Bavaria, J.E.; Kaiser, L.R. Fungal pulmonary abscess in an adult secondary to hyperimmunoglobulin E (Job's) syndrome. *Ann. Thorac. Surg.* **1995**, *59*, 759–761. [\[CrossRef\]](#) [\[PubMed\]](#)
388. Santambrogio, L.; Nosotti, M.; Pavoni, G.; Harte, M.; Pietogrande, M.C. Pneumatocele Complicated by Fungal Lung Abscess in Job's Syndrome: Successful Lobectomy with the Aid of Videothoracoscopy. *Scand. Cardiovasc. J.* **1997**, *31*, 177–179. [\[CrossRef\]](#)
389. Van Der Meer, J.W.M.; Bont, L.; Verhage, J. Aspergillus Infection in Patients with Hyperimmunoglobulin E Syndrome. *Clin. Infect. Dis.* **1998**, *27*, 1337. [\[CrossRef\]](#)
390. Chandesris, M.-O.; Melki, I.; Natividad, A.; Puel, A.; Fieschi, C.; Yun, L.; Thumerelle, C.; Oksenhendler, E.; Boutboul, D.; Thomas, C.; et al. Autosomal Dominant STAT3 Deficiency and Hyper-IgE Syndrome. *Medicine* **2012**, *91*, e1–e19. [\[CrossRef\]](#)
391. Duréault, A.; Tcherakian, C.; Poiree, S.; Catherinot, E.; Danion, F.; Jouvion, G.; Bougnoux, M.E.; Mahlaoui, N.; Givel, C.; Castelle, M.; et al. Spectrum of Pulmonary Aspergillosis in Hyper-IgE Syndrome with Autosomal-Dominant STAT3 Deficiency. *J. Allergy Clin. Immunol. Pract.* **2019**, *7*, 1986–1995.e3. [\[CrossRef\]](#)
392. Kasuga, K.; Nakamoto, K.; Doi, K.; Kurokawa, N.; Saraya, T.; Ishii, H. Chronic pulmonary aspergillosis in a patient with hyper-IgE syndrome. *Respirol. Case Rep.* **2022**, *10*, e0887. [\[CrossRef\]](#)

393. Paris, A.J.; Hayer, K.E.; Oved, J.H.; Avgousti, D.C.; Toulmin, S.A.; Zepp, J.A.; Zacharias, W.J.; Katzen, J.B.; Basil, M.C.; Kremp, M.M.; et al. STAT3–BDNF–TrkB signalling promotes alveolar epithelial regeneration after lung injury. *Nature* **2020**, *22*, 1197–1210. [[CrossRef](#)] [[PubMed](#)]
394. Béziat, V.; Tavernier, S.J.; Chen, Y.-H.; Ma, C.S.; Materna, M.; Laurence, A.; Staal, J.; Aschenbrenner, D.; Roels, L.; Worley, L.; et al. Dominant-negative mutations in human IL6ST underlie hyper-IgE syndrome. *J. Exp. Med.* **2020**, *217*, e20191804. [[CrossRef](#)] [[PubMed](#)]
395. Schwerd, T.; Twigg, S.R.; Aschenbrenner, D.; Manrique, S.; Miller, K.A.; Taylor, I.B.; Capitani, M.; McGowan, S.J.; Sweeney, E.; Weber, A.; et al. A biallelic mutation in IL6ST encoding the GP130 co-receptor causes immunodeficiency and craniosynostosis. *J. Exp. Med.* **2017**, *214*, 2547–2562. [[CrossRef](#)] [[PubMed](#)]
396. Shahin, T.; Aschenbrenner, D.; Cagdas, D.; Bal, S.K.; Conde, C.D.; Garncarz, W.; Medgyesi, D.; Schwerd, T.; Karaatmaca, B.; Cetinkaya, P.G.; et al. Selective loss of function variants in *IL6ST* cause Hyper-IgE syndrome with distinct impairments of T-cell phenotype and function. *Haematologica* **2019**, *104*, 609–621. [[CrossRef](#)]
397. Chen, Y.-H.; Zastrow, D.B.; Metcalfe, R.D.; Gartner, L.; Krause, F.; Morton, C.J.; Marwaha, S.; Fresard, L.; Huang, Y.; Zhao, C.; et al. Functional and structural analysis of cytokine-selective IL6ST defects that cause recessive hyper-IgE syndrome. *J. Allergy Clin. Immunol.* **2021**, *148*, 585–598. [[CrossRef](#)]
398. Spencer, S.; Bal, S.K.; Egner, W.; Allen, H.L.; Raza, S.I.; Ma, C.A.; Gürel, M.; Zhang, Y.; Sun, G.; Sabroe, R.A.; et al. Loss of the interleukin-6 receptor causes immunodeficiency, atopy, and abnormal inflammatory responses. *J. Exp. Med.* **2019**, *216*, 1986–1998. [[CrossRef](#)]
399. Perez, L.; Messina, F.; Negroni, R.; Arechavala, A.; Bustamante, J.; Oleastro, M.; Migaud, M.; Casanova, J.-L.; Puel, A.; Santiso, G. Inherited CARD9 Deficiency in a Patient with Both Exophiala spinifera and Aspergillus nomius Severe Infections. *J. Clin. Immunol.* **2020**, *40*, 359–366. [[CrossRef](#)]
400. Zhang, Y.; Huang, C.; Song, Y.; Ma, Y.; Wan, Z.; Zhu, X.; Wang, X.; Li, R. Primary Cutaneous Aspergillosis in a Patient with CARD9 Deficiency and Aspergillus Susceptibility of Card9 Knockout Mice. *J. Clin. Immunol.* **2021**, *41*, 427–440. [[CrossRef](#)]
401. Bortnick, R.; Wlodarski, M.; de Haas, V.; De Moerloose, B.; Dworzak, M.; Hasle, H.; Masetti, R.; Starý, J.; Turkiewicz, D.; Ussowicz, M.; et al. Hematopoietic stem cell transplantation in children and adolescents with GATA2-related myelodysplastic syndrome. *Bone Marrow Transplant.* **2021**, *56*, 2732–2741. [[CrossRef](#)]
402. Vila, A.; Dapás, J.I.; Rivero, C.V.; Bocanegra, F.; Furnari, R.F.; Hsu, A.P.; Holland, S.M. Multiple Opportunistic Infections in a Woman with GATA2 Mutation. *Int. J. Infect. Dis.* **2016**, *54*, 89–91. [[CrossRef](#)]
403. Vedula, R.S.; Cheng, M.P.; Ronayne, C.E.; Farmakiotis, D.; Ho, V.T.; Koo, S.; Marty, F.M.; Lindsley, R.C.; Bold, T.D. Somatic GATA2 mutations define a subgroup of myeloid malignancy patients at high risk for invasive fungal disease. *Blood Adv.* **2021**, *5*, 54–60. [[CrossRef](#)]
404. Marciano, B.E.; Olivier, K.N.; Folio, L.R.; Zerbe, C.S.; Hsu, A.P.; Freeman, A.F.; Filie, A.C.; Spinner, M.A.; Sanchez, L.A.; Lovell, J.P.; et al. Pulmonary Manifestations of GATA2 Deficiency. *Chest* **2021**, *160*, 1350–1359. [[CrossRef](#)]
405. Fischer, A.; Descamps-Latscha, B.; Gerota, I.; Scheinmetzler, C.; Virelizier, J.; Trung, P.; Lisowska-Grosppierre, B.; Perez, N.; Durandy, A.; Griscelli, C. Bone-marrow transplantation for inborn error of phagocytic cells associated with defective adherence, chemotaxis, and oxidative response during opsonised particle phagocytosis. *Lancet* **1983**, *322*, 473–476. [[CrossRef](#)] [[PubMed](#)]
406. Mates, M.; Kirkpatrick, C.H. A man with multiple infections with unusual organisms. *Ann. Allergy Asthma Immunol.* **1999**, *82*, 17–22. [[CrossRef](#)] [[PubMed](#)]
407. Kuijpers, T.W.; Van Bruggen, R.; Kamerbeek, N.; Tool, A.T.J.; Hicsonmez, G.; Gurgey, A.; Karow, A.; Verhoeven, A.J.; Seeger, K.; Sanal, O.; et al. Natural history and early diagnosis of LAD-1/variant syndrome. *Blood* **2007**, *109*, 3529–3537. [[CrossRef](#)] [[PubMed](#)]
408. Bellanné-Chantelot, C.; Clauin, S.; Leblanc, T.; Cassinat, B.; Rodrigues-Lima, F.; Beaufils, S.; Vaury, C.; Barkaoui, M.; Fenneteau, O.; Maier-Redelsperger, M.; et al. Mutations in the ELA2 gene correlate with more severe expression of neutropenia: A study of 81 patients from the French Neutropenia Register. *Blood* **2004**, *103*, 4119–4125. [[CrossRef](#)] [[PubMed](#)]
409. Zhan, P.; Dukik, K.; Li, D.; Sun, J.; Stielow, J.; Ende, B.G.V.D.; Brankovics, B.; Menken, S.; Mei, H.; Bao, W.; et al. Phylogeny of dermatophytes with genomic character evaluation of clinically distinct *Trichophyton rubrum* and *T. violaceum*. *Stud. Mycol.* **2018**, *89*, 153–175. [[CrossRef](#)] [[PubMed](#)]
410. Gupta, C.; Das, S.; Gaurav, V.; Singh, P.K.; Rai, G.; Datt, S.; Tigga, R.A.; Pandhi, D.; Bhattacharya, S.N.; Ansari, M.A.; et al. Review on host-pathogen interaction in dermatophyte infections. *J. Med. Mycol.* **2022**, *33*, 101331. [[CrossRef](#)]
411. Abdel-Rahman, S.M. Genetic Predictors of Susceptibility to Dermatophytoses. *Mycopathologia* **2017**, *182*, 67–76. [[CrossRef](#)]
412. Hay, R. Chronic dermatophyte infections. I. Clinical and mycological features. *Br. J. Dermatol.* **1982**, *106*, 1–7. [[CrossRef](#)]
413. Shelley, E.D.; Shelley, W.B.; Schafer, R.L. Generalized *Trichophyton rubrum* infection in congenital ichthyosiform erythroderma. *J. Am. Acad. Dermatol.* **1989**, *20*, 1133–1134. [[CrossRef](#)] [[PubMed](#)]
414. Sheetz, K.; Lynch, P.J. Ichthyosis and dermatophyte fungal infection. *J. Am. Acad. Dermatol.* **1991**, *24*, 321. [[CrossRef](#)] [[PubMed](#)]
415. Moreno-Giménez, J. Infections by *Trichophyton rubrum*. *J. Am. Acad. Dermatol.* **1991**, *24*, 323–324. [[CrossRef](#)] [[PubMed](#)]
416. Agostini, G.; Geti, V.; Difonzo, E.M.; Giannotti, B. Dermatophyte infection in ichthyosis vulgaris. *Mycoses* **1992**, *35*, 197–199. [[CrossRef](#)] [[PubMed](#)]
417. Sentamilselvi, G.; Kamalam, A.; Ajithadas, K.; Janaki, C.; Thambiah, A. Scenario of chronic dermatophytosis: An Indian study. *Mycopathologia* **1997**, *140*, 129–135. [[CrossRef](#)] [[PubMed](#)]

418. Hoetzenecker, W.; Schanz, S.; Schaller, M.; Fierlbeck, G. Generalized tinea corporis due to *Trichophyton rubrum* in ichthyosis vulgaris. *J. Eur. Acad. Dermatol. Venereol.* **2007**, *21*, 1129–1131. [\[CrossRef\]](#)
419. Grahovac, M.; Budimčić, D. Unrecognized dermatophyte infection in ichthyosis vulgaris. *Acta Derm. Croat* **2009**, *17*, 127–130.
420. De Freitas, C.F.N.P.; Mulinari-Brenner, F.; Fontana, H.R.; Gentili, A.C.; Hammerschmidt, M. Ichthyosis associated with widespread tinea corporis: Report of three cases. *An. Bras. Dermatol.* **2013**, *88*, 627–630. [\[CrossRef\]](#)
421. Schøsler, L.; Andersen, L.K.; Arendrup, M.C.; Sommerlund, M. Recurrent terbinafine resistant *Trichophyton rubrum* infection in a child with congenital ichthyosis. *Pediatr. Dermatol.* **2018**, *35*, 259–260. [\[CrossRef\]](#)
422. Szlávicz, E.; Németh, C.; Szepes, É.; Gyömörei, C.; Gyulai, R.; Lengyel, Z. Congenital ichthyosis associated with *Trichophyton rubrum* tinea, imitating drug hypersensitivity reaction. *Med. Mycol. Case Rep.* **2020**, *29*, 15–17. [\[CrossRef\]](#)
423. Steele, L.; Hong, A.; Balogh, P.; O'Toole, E.A.; Harwood, C.A.; Maruthappu, T. Disseminated tinea incognita in a patient with ichthyosis vulgaris and eczema. *Clin. Exp. Dermatol.* **2021**, *46*, 210–212. [\[CrossRef\]](#)
424. Agrawal, M.; Yadav, P.; Yadav, J.; Chander, R. Clear zone phenomenon: A rare phenomenon in ichthyosis with co-existing superficial fungal infection. *Indian J. Dermatol. Venereol. Leprol.* **2021**, *87*, 103–105. [\[CrossRef\]](#)
425. Ibsen, H.H.W.; Brandrup, F. Tinea corporis due to *Trichophyton verrucosum* in recessive, X-linked ichthyosis. *Mycoses* **2009**, *36*, 319–320. [\[CrossRef\]](#) [\[PubMed\]](#)
426. Kamalam, A.; Thambiah, A.S. Genetic Ichthyosis and *Trichophyton rubrum* Infection in Infants: Genetische Ichthyosen und *Trichophyton-rubrum*-Infektionen bei kleinen Kindern. *Mycoses* **1982**, *25*, 281–283. [\[CrossRef\]](#) [\[PubMed\]](#)
427. Scheers, C.; Andre, J.; Thompson, C.; Rebuffat, E.; Harag, S.; Kolivras, A. Refractory *Trichophyton rubrum* Infection in Lamellar Ichthyosis. *Pediatr. Dermatol.* **2013**, *30*, e200–e203. [\[CrossRef\]](#)
428. Machan, M.; Kestenbaum, T.; Fraga, G.R. Diffuse Hyperkeratosis in a Deaf and Blind 48-Year-Old Woman—Quiz Case. *Arch. Dermatol.* **2012**, *148*, 1199. [\[CrossRef\]](#) [\[PubMed\]](#)
429. Mikhail, G.R.; Babel, D. *Trichophyton rubrum* infection and keratoderma palmaris et plantaris. *Arch. Dermatol.* **1981**, *117*, 753–754. [\[CrossRef\]](#)
430. Reis, A.; Hennies, H.-C.; Langbein, L.; Digweed, M.; Mischke, D.; Drechsler, M.; Schröck, E.; Royer-Pokora, B.; Franke, W.W.; Sperling, K.; et al. Keratin 9 gene mutations in epidermolytic palmoplantar keratoderma (EPPK). *Nat. Genet.* **1994**, *6*, 174–179. [\[CrossRef\]](#)
431. Kimonis, V.; Yang, J.-M.; Doyle, S.Z.; Bale, S.J.; Compton, J.G.; DiGiovanna, J.J. A Mutation in the V1 End Domain of Keratin 1 in Non-Epidermolytic Palmar-Plantar Keratoderma. *J. Invest. Dermatol.* **1994**, *103*, 764–769. [\[CrossRef\]](#)
432. Blaydon, D.C.; Lind, L.K.; Plagnol, V.; Linton, K.J.; Smith, F.J.; Wilson, N.J.; McLean, W.I.; Munro, C.S.; South, A.P.; Leigh, I.M.; et al. Mutations in AQP5, Encoding a Water-Channel Protein, Cause Autosomal-Dominant Diffuse Nonepidermolytic Palmoplantar Keratoderma. *Am. J. Hum. Genet.* **2013**, *93*, 330–335. [\[CrossRef\]](#)
433. Nielsen, P.G. Dermatophyte Infections in Hereditary Palmo-Plantar Keratoderma. *Dermatology* **1984**, *168*, 238–241. [\[CrossRef\]](#)
434. Xiao, H.; Guo, Y.; Yi, J.; Xia, H.; Xu, H.; Yuan, L.; Hu, P.; Yang, Z.; He, Z.; Lu, H.; et al. Identification of a Novel Keratin 9 Missense Mutation in a Chinese Family with Epidermolytic Palmoplantar Keratoderma. *Cell Physiol. Biochem.* **2018**, *46*, 1919–1929. [\[CrossRef\]](#) [\[PubMed\]](#)
435. Krøigård, A.B.; Hetland, L.E.; Clemmensen, O.; Blaydon, D.; Hertz, J.M.; Bygum, A. The first Danish family reported with an AQP5 mutation presenting diffuse non-epidermolytic palmoplantar keratoderma of Bothnian type, hyperhidrosis and frequent *Corynebacterium* infections: A case report. *BMC Dermatol.* **2016**, *16*, 7. [\[CrossRef\]](#)
436. Majocchi, D. A new trichophytic granuloma: Clinical and mycological studies. *Bull. R. Acad. Med. Roma* **1883**, *9*, 220–223.
437. Marill, F.G.; Liautaud, B.; Hamra-Krouha, M.S. Fatal evolution of a dermatophytic disease due to *Trichophyton schönleini*. *Bull. Soc. Pathol. Exot. Fil.* **1975**, *68*, 450–456.
438. Brahmni, Z.; Liautaud, B.; Marill, F. Depressed cell-mediated immunity in chronic dermatophytic infections. *Ann. d'immunol.* **1980**, *131*, 143–153.
439. Liautaud, B.; Marill, F.G. Dermatophytic disease. Recent Algerian observations. *Bull. Soc. Pathol. Exot. Fil.* **1984**, *77*, 637–648.
440. Lanternier, F.; Pathan, S.; Vincent, Q.B.; Liu, L.; Cypowyj, S.; Prando, C.; Migaud, M.; Taibi, L.; Ammar-Khodja, A.; Stambouli, O.B.; et al. Deep Dermatophytosis and Inherited CARD9 Deficiency. *N. Engl. J. Med.* **2013**, *369*, 1704–1714. [\[CrossRef\]](#)
441. Zhang, Y.; Mijiti, J.; Huang, C.; Song, Y.; Wan, Z.; Li, R.; Kang, X.; Wang, X. Deep dermatophytosis caused by *Microsporium ferrugineum* in a patient with CARD 9 mutations. *Br. J. Dermatol.* **2019**, *181*, 1093–1095. [\[CrossRef\]](#)
442. Simpson, J.; Fröbel, P.; Seneviratne, S.; Brown, M.; Lowe, D.; Grimbacher, B.; Fliegauf, M.; Fearfield, L. Invasive dermatophyte infection with *Trichophyton interdigitale* is associated with prurigo-induced pseudoperforation and a signal transducer and activator of transcription 3 mutation. *Br. J. Dermatol.* **2018**, *179*, 750–754. [\[CrossRef\]](#)
443. Vinh, D.C. The molecular immunology of human susceptibility to fungal diseases: Lessons from single gene defects of immunity. *Expert Rev. Clin. Immunol.* **2019**, *15*, 461–486. [\[CrossRef\]](#)
444. McElroy, J.A.; Prestes, C.D.A.; Su, W.P. Mycetoma: Infection with tumefaction, draining sinuses, and “grains”. *Cutis* **1992**, *49*, 107–110.
445. Huang, C.; Deng, W.; Zhang, Y.; Zhang, K.; Ma, Y.; Song, Y.; Wan, Z.; Wang, X.; Li, R. CARD9 deficiency predisposing chromoblastomycosis: A case report and comparative transcriptome study. *Front. Immunol.* **2022**, *13*, 112. [\[CrossRef\]](#) [\[PubMed\]](#)
446. Mariat, F.; Liautaud, B.; Liautaud, M.; Marill, F.G. *Hendersonula toruloidea*, causative agent of a fungal verrucous dermatitis observed in Algeria. *Sabouraudia* **1978**, *16*, 133–140. [\[CrossRef\]](#) [\[PubMed\]](#)

447. Wang, X.; Wang, W.; Lin, Z.; Wang, X.; Li, T.; Yu, J.; Liu, W.; Tong, Z.; Xu, Y.; Zhang, J.; et al. CARD9 mutations linked to subcutaneous phaeohyphomycosis and TH17 cell deficiencies. *J. Allergy Clin. Immunol.* **2014**, *133*, 905–908.e3. [\[CrossRef\]](#) [\[PubMed\]](#)
448. Lanternier, F.; Barbati, E.; Meinzer, U.; Liu, L.; Pedergrana, V.; Migaud, M.; Héritier, S.; Chomton, M.; Frémond, M.-L.; Gonzales, E.; et al. Inherited CARD9 Deficiency in 2 Unrelated Patients With Invasive Exophiala Infection. *J. Infect. Dis.* **2015**, *211*, 1241–1250. [\[CrossRef\]](#)
449. Wang, X.; Zhang, R.; Wu, W.; Song, Y.; Wan, Z.; Han, W.; Li, R. Impaired Specific Antifungal Immunity in CARD9-Deficient Patients with Phaeohyphomycosis. *J. Investig. Dermatol.* **2018**, *138*, 607–617. [\[CrossRef\]](#)
450. Wang, C.; Xing, H.; Jiang, X.; Zeng, J.; Liu, Z.; Chen, J.; Wu, Y. Cerebral Phaeohyphomycosis Caused by Exophiala dermatitidis in a Chinese CARD9-Deficient Patient: A Case Report and Literature Review. *Front. Neurol.* **2019**, *10*, 938. [\[CrossRef\]](#)
451. Imanaka, Y.; Taniguchi, M.; Doi, T.; Tsumura, M.; Nagaoka, R.; Shimomura, M.; Asano, T.; Kagawa, R.; Mizoguchi, Y.; Karakawa, S.; et al. Inherited CARD9 Deficiency in a Child with Invasive Disease Due to Exophiala dermatitidis and Two Older but Asymptomatic Siblings. *J. Clin. Immunol.* **2021**, *41*, 975–986. [\[CrossRef\]](#)
452. Huang, C.; Zhang, Y.; Song, Y.; Wan, Z.; Wang, X.; Li, R. Phaeohyphomycosis caused by *Phialophora americana* with CARD9 mutation and 20-year literature review in China. *Mycoses* **2019**, *62*, 908–919. [\[CrossRef\]](#)
453. Gavino, C.; Mellinghoff, S.; Cornely, O.A.; Landekic, M.; Le, C.; Langelier, M.; Golizeh, M.; Proske, S.; Vinh, D.C. Novel bi-allelic splice mutations in CARD9 causing adult-onset *Candida* endophthalmitis. *Mycoses* **2018**, *61*, 61–65. [\[CrossRef\]](#) [\[PubMed\]](#)
454. Lai, S.H.; Duque, J.S.R.; Chung, B.H.-Y.; Chung, T.W.-H.; Leung, D.; Ho, R.S.-L.; Lee, R.; Poon, R.W.; Chua, G.T.; Cheong, K.-N.; et al. Invasive cerebral phaeohyphomycosis in a Chinese boy with CARD9 deficiency and showing unique radiological features, managed with surgical excision and antifungal treatment. *Int. J. Infect. Dis.* **2021**, *107*, 59–61. [\[CrossRef\]](#) [\[PubMed\]](#)
455. Paccoud, O.; Vignier, N.; Boui, M.; Migaud, M.; Vironneau, P.; Kania, R.; Méchaï, F.; Brun, S.; Alanio, A.; Tauziède-Espariat, A.; et al. Invasive Rhinosinusitis Caused by *Alternaria infectoria* in a Patient with Autosomal Recessive CARD9 Deficiency and a Review of the Literature. *J. Fungi* **2022**, *8*, 446. [\[CrossRef\]](#)
456. Kalantri, M.; Khopkar, U.; Shah, A.; Bargir, U.A.; Hule, G.; Madkaikar, M. A case of disseminated subcutaneous phaeohyphomycosis caused by *Exserohilum rostratum* with CARD9 mutation. *Indian J. Dermatol. Venereol. Leprol.* **2021**, *88*, 59–61. [\[CrossRef\]](#)
457. Yan, X.; Yu, C.; Fu, X.; Bao, F.; Du, D.; Wang, C.; Wang, N.; Wang, S.; Shi, Z.; Zhou, G.; et al. CARD 9 mutation linked to *Corynespora cassicola* infection in a Chinese patient. *Br. J. Dermatol.* **2016**, *174*, 176–179. [\[CrossRef\]](#) [\[PubMed\]](#)
458. Arango-Franco, C.A.; Moncada-Vélez, M.; Beltrán, C.P.; Berrío, I.; Mogollón, C.; Restrepo, A.; Trujillo, M.; Osorio, S.D.; Castro, L.; Gómez, L.V.; et al. Early-Onset Invasive Infection Due to *Corynespora cassicola* Associated with Compound Heterozygous CARD9 Mutations in a Colombian Patient. *J. Clin. Immunol.* **2018**, *38*, 794–803. [\[CrossRef\]](#)
459. Guo, Y.; Zhu, Z.; Gao, J.; Zhang, C.; Zhang, X.; Dang, E.; Li, W.; Qiao, H.; Liao, W.; Wang, G.; et al. The Phytopathogenic Fungus *Pallidocercospora crystallina*-Caused Localized Subcutaneous Phaeohyphomycosis in a Patient with a Homozygous Missense CARD9 Mutation. *J. Clin. Immunol.* **2019**, *39*, 713–725. [\[CrossRef\]](#)
460. Drummond, R.A.; Desai, J.V.; Hsu, A.P.; Oikonomou, V.; Vinh, D.C.; Acklin, J.A.; Abers, M.S.; Walkiewicz, M.A.; Anzick, S.L.; Swamydas, M.; et al. Human Dectin-1 deficiency impairs macrophage-mediated defense against phaeohyphomycosis. *J. Clin. Invest.* **2022**, *132*, 1–14. [\[CrossRef\]](#)
461. Liese, J.; Kloos, S.; Jendrossek, V.; Petropoulou, T.; Wintergerst, U.; Notheis, G.; Gahr, M.; Belohradsky, B.H. Long-term follow-up and outcome of 39 patients with chronic granulomatous disease. *J. Pediatr.* **2000**, *137*, 687–693. [\[CrossRef\]](#)
462. Vinh, D.C.; Freeman, A.F.; Shea, Y.R.; Malech, H.L.; Abinun, M.; Weinberg, G.A.; Holland, S.M. Mucormycosis in chronic granulomatous disease: Association with iatrogenic immunosuppression. *J. Allergy Clin. Immunol.* **2009**, *123*, 1411–1413. [\[CrossRef\]](#)
463. Layios, N.; Canivet, J.-L.; Baron, F.; Moutschen, M.; Hayette, M.-P. *Mortierella wolfii*—Associated Invasive Disease. *Emerg. Infect. Dis.* **2014**, *20*, 1591–1592. [\[CrossRef\]](#)
464. Winstead, M.; Ozolek, J.; Nowalk, A.; Williams, J.; Lugt, M.V.; Lin, P. Disseminated Lichtheimia ramosa Infection After Hematopoietic Stem Cell Transplantation in a Child With Chronic Granulomatous Disease. *Pediatr. Infect. Dis. J.* **2017**, *36*, 1222–1224. [\[CrossRef\]](#)
465. Conrad, A.; Neven, B.; Mahlaoui, N.; Suarez, F.; Sokol, H.; Ruemmele, F.M.; Rouzaud, C.; Moshous, D.; Lortholary, O.; Blanche, S.; et al. Infections in Patients with Chronic Granulomatous Disease Treated with Tumor Necrosis Factor Alpha Blockers for Inflammatory Complications. *J. Clin. Immunol.* **2020**, *41*, 185–193. [\[CrossRef\]](#) [\[PubMed\]](#)
466. Al-Otaibi, A.M.; Al-Shahrani, D.A.; Al-Idrissi, E.M.; Al-Abdely, H.M. Invasive mucormycosis in chronic granulomatous disease. *Saudi Med. J.* **2016**, *37*, 567–569. [\[CrossRef\]](#)
467. Nadeem, A.; Wahla, A.; Altarifi, A. Invasive Mediastinal Mucormycosis with Pulmonary and Cardiac Involvement in an Adult with Chronic Granulomatous Disease: Case Report and Review of the Literature. *Eur. J. Case Rep. Intern. Med.* **2021**, *8*, 002435. [\[CrossRef\]](#)
468. Wildenbeest, J.G.; Oomen, M.W.; Brüggemann, R.J.; de Boer, M.; Bijleveld, Y.; Berg, J.M.V.D.; Kuijpers, T.W.; Pakr, D. Rhizopus Oryzae Skin Infection Treated With Posaconazole in a Boy With Chronic Granulomatous Disease. *Pediatr. Infect. Dis. J.* **2010**, *29*, 578. [\[CrossRef\]](#) [\[PubMed\]](#)

469. Wang, X.; Wang, A.; Wang, X.; Li, R.; Yu, J. Cutaneous mucormycosis caused by *Mucor irregularis* in a patient with CARD 9 deficiency. *Br. J. Dermatol.* **2019**, *180*, 213–214. [[CrossRef](#)]
470. Wang, X.; Ding, H.; Chen, Z.; Zeng, X.; Sun, J.; Chen, H.; Fu, M. CARD9 Deficiency in a Chinese Man with Cutaneous Mucormycosis, Recurrent Deep Dermatophytosis and a Review of the Literature. *Mycopathologia* **2020**, *185*, 1041–1050. [[CrossRef](#)] [[PubMed](#)]
471. Zhang, L.; Huang, J.; Ma, Y.; Wan, Z.; Dai, H.; Li, R.; Gu, H.; Wang, X. Primary Cutaneous Mucormycosis, Candida Onychomycosis and Endophthalmitis in a Patient with CARD9 Mutation. *Mycopathologia* **2022**, *187*, 305–308. [[CrossRef](#)]
472. Lu, X.-L.; Najafzadeh, M.; Dolatabadi, S.; Ran, Y.-P.; Ende, A.G.V.D.; Shen, Y.-N.; Li, C.-Y.; Xi, L.-Y.; Hao, F.; Zhang, Q.-Q.; et al. Taxonomy and epidemiology of *Mucor irregularis*, agent of chronic cutaneous mucormycosis. *Persoonia-Mol. Phylogeny Evol. Fungi* **2013**, *30*, 48–56. [[CrossRef](#)]
473. Donadieu, J.; Lamant, M.; Fieschi, C.; De Fontbrune, F.S.; Caye, A.; Ouachee, M.; Beaupain, B.; Bustamante, J.; Poiriel, H.A.; Isidor, B.; et al. Natural history of GATA2 deficiency in a survey of 79 French and Belgian patients. *Haematologica* **2018**, *103*, 1278–1287. [[CrossRef](#)] [[PubMed](#)]
474. Kumar, N.; Hanks, M.E.; Chandrasekaran, P.; Davis, B.C.; Hsu, A.P.; Van Wagoner, N.J.; Merlin, J.S.; Spalding, C.; La Hoz, R.M.; Holland, S.M.; et al. Gain-of-function signal transducer and activator of transcription 1 (STAT1) mutation-related primary immunodeficiency is associated with disseminated mucormycosis. *J. Allergy Clin. Immunol.* **2014**, *134*, 236–239. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.