

Adaptive Evolution Compensated the plasmid Fitness Costs brought by specific Genetic Conflicts

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Table S4. Detailed function of non-synonymous SNP-genes in *E. coli* C600 carrying the pHN330 plasmid.

Gene	Product and function	Supplementary description
<i>ptrA</i>	Protease III; hydrolyzes oligopeptides and peptides; involved in the protein catabolic process of bacteria.	Swamy et al. found that lack of PtrA has no effect on bacterial growth [1].
<i>lpdA</i>	Lipoamide dehydrogenase; component of the pyruvate dehydrogenase complex, which catalyzes the conversion of pyruvate to acetyl-CoA.	Murarka et al. found that pyruvate dehydrogenase deficient <i>E. coli</i> had decreased growth and glucose metabolism rates [2].
<i>bamA</i>	Outer membrane protein assembly factor BamA; component of the BAM complex, which is essential to constructing and maintaining the outer membrane.	Lack of BamA causes defects in assembly of β -barrel protein complex, influencing the construction of the outer membrane [3].
<i>melA</i>	α -Galactosidase; hydrolyzes melibiose to glucose and galactose.	Melibiose is a common nutrient. Belyaeva et al. found that mutants in the <i>melA</i> gene affect the growth of bacteria with melibiose as the carbon source [4].
<i>gatA</i>	Galactitol-specific PTS enzyme IIA component; involved in galactinol phosphorylation and transport.	Galactinol is transported into the cytosol by the PTS system and is a carbon source that provides energy for bacterial growth. Lengeler et al. found that bacteria with a mutated <i>gatA</i> gene cannot transport galactinol [5].
<i>mdoB</i>	Phosphoglycerol transferase I; catalyzes phosphoglycerol residues from phosphatidylglycerol into membrane-derived oligosaccharide.	Membrane-derived oligosaccharide plays an important role in maintaining the stability of the bacterial membrane [6].
<i>yihV</i>	6-Deoxy-6-sulfofructose kinase; involved in the sulphoglycolysis pathway.	A mutation in the <i>yihV</i> gene influences the utilization of sulfoquinovose in bacteria [7].
<i>mntH</i>	Divalent metal cation transporter MntH; transports most metal ions, with high affinity for Mn (II).	Makui et al. found that inactivation of MntH did not affect bacterial growth in nutrient rich medium [8].

<i>trkA</i>	NAD-binding component of Trk potassium transporters; K ⁺ uptake mediated by Trk transport system with hypotonic conditions.	TrkA plays an important role in maintaining potassium ion homeostasis under hypotonic conditions [9].
<i>ydfK</i>	Cold shock protein; unknown function.	It has been reported that after a cold shock, the expression of YdfK is upregulated [10].

Table S5. Detailed functions of non-synonymous SNP-genes in *Escherichia coli* C600N500.

Gene	Product and function	Supplementary description
<i>bamA</i>	Outer membrane protein assembly factor BamA; BamA is a component of the BAM complex, which is essential to constructing and maintaining the outer membrane.	Lack of BamA causes defects in the assembly of the β -barrel protein complex, influencing the construction of the outer membrane [3].
<i>bcsG</i>	Cellulose phosphoethanolamine transferase; the BcsE-BcsF-BcsG pathway controls the modification of cellulose by direct protein–protein interactions.	Exopolysaccharide cellulose is the major component of the self-produced extracellular matrix in biofilms, which enhances antibiotic resistance. The <i>bcsG</i> gene is essential for the formation of cellulose; lack of the <i>bcsG</i> gene results in the inability to form a biofilm [11].
<i>lptF</i>	Lipopolysaccharide transport system protein LptF; involved in lipopolysaccharide transport.	Sherman et al. found that LptF is essential for the transport of LPS. LPS plays a vital role in cell mobility, biofilm formation, antibiotic resistance, intestinal colonization, and is responsible for protecting the bacterial cell [12].
<i>mreC</i>	Cell shape determining protein MreC; <i>mreC</i> gene is a component of the <i>mreBCD</i> operon; encodes for cell cytoskeletal proteins.	The function of the bacterial cytoskeleton is highly conserved. Kruse T et al. found that the bacterial cell will undergo morphological change then lysis when the MreBCD protein is depleted [13].
<i>sfmD</i>	Fimbrial usher protein SfmD	Involved in the functioning of pili—macromolecular structures on the surface of gram-negative bacteria; defects in SfmD will not affect the biofilm formation of <i>E. coli</i> [14].

<i>rfaF</i>	ADP-heptose–LPS heptosyltransferase 2; involved in the synthesis of the inner core of LPS.	The inner core of LPS is highly conserved and responsible for the function of the outer membrane barrier [15].
<i>gudD</i>	D-glucarate dehydratase; involved in the D-glucarate catabolic pathway.	D-glucarate is not a common product in glucose metabolism [16].
<i>araC</i>	DNA-binding transcriptional dual regulator AraC; in the presence of arabinose, AraC stimulates the transcription of arabinose utilization genes.	Generally, bacteria utilize glucose as a carbon source due to carbon catabolite repression [17].
<i>pepQ</i>	Xaa-Pro dipeptidase; PepQ is capable of X-proline dipeptide hydrolysis.	Mutants in the <i>pepQ</i> gene of <i>E. coli</i> have no observable change in phenotype [18].
<i>paaJ</i>	Beta-ketoadipyl-CoA thiolase; the <i>paa</i> gene cluster is involved in phenylacetic acid metabolism.	Phenylacetic acid is not a common energy source.
<i>dgt</i>	Deoxyguanosinetriphosphate triphosphohydrolase; catalytic hydrolysis of dGTP to deoxyguanosine and triphosphate.	dGTPase is involved in maintaining proper dNTP pools and plays a vital role in cell survival and DNA replication. A mutation in the <i>dgt</i> gene reduces the fidelity of DNA replication [19].
<i>mshA</i>	D-inositol 3-phosphate glycosyltransferase; involved in the synthesis of extracellular colanic acid.	Colanic acid, an extracellular polysaccharide secreted by <i>E. coli</i> , is thought to promote biofilm formation and is important for survival. Ren et al. knocked out four consecutive genes of the CA operon (<i>wza</i> , <i>wzb</i> , <i>wzc</i> , and <i>wcaA</i>) and found that the strain was unable to synthesize colanic acid [20].

Table S6. Detailed functions of non-synonymous SNP-genes in *E. coli* C600Y500.

Gene	Product and Function	Supplementary description
<i>fliY</i>	Cystine ABC transporter periplasmic binding protein; FliY together with the L-cysteine exporter EAMA constitute the L-cysteine/L-cystine shuttle system involved in oxidative stress resistance	Ohtsu et al. found that deletion of the <i>fliY</i> gene results in decreased <i>E. coli</i> growth in LB broth [21].
<i>bamA</i>	Outer membrane protein assembly factor BamA; BamA is a component of the BAM complex, which is essential to constructing and maintaining the outer membrane.	Lack of BamA causes defects in the assembly of the β -barrel protein complex, influencing the construction of the outer membrane [3].
<i>yghQ</i>	Putative transport protein YghQ	No study of the <i>yghQ</i> gene has yet been conducted.

<i>caiA</i>	Crotonobetainyl-CoA reductase; Convert crotonobetainyl to γ -butyrobetaine under anaerobic conditions.	The expression of <i>caiA</i> suppressed is suppressed under aerobic conditions [22].
<i>gloA</i>	Glyoxalase I; involved in the glutathione-dependent glyoxalase I pathway that metabolizes methylglyoxal.	Endogenous synthesis of methylglyoxal occurs in the glycolytic pathway; 300 μ M methylglyoxal causes growth arrest and higher concentrations cause cell death. The glutathione-dependent glyoxalase I pathway is the major pathway for methylglyoxal detoxification [23].
<i>acnA</i>	Aconitate hydratase A; catalyzes citrate and isocitrate via cisaconitate in the citric acid and glyoxylate cycles.	Mutations in the <i>acnA</i> gene has no effect on the growth of <i>E. coli</i> [24].
<i>rlmB</i>	23S rRNA methyltransferase RlmB; responsible for the methylation of Gm2251 in 23S rRNA	RlmB has no important role in ribosome assembly or function in <i>E. coli</i> [25].
<i>ydiF</i>	Acetate-CoA transferase; displays enzymatic activity with short-chain acyl-CoAs.	Rangarajan et al. found that YdiF is involved in metabolism of short-chain fatty acid [26].
<i>sbp</i>	Sulfate/thiosulfate ABC transporter periplasmic binding protein Sbp; regulates the composition of sulfate–thiosulfate transporters.	Mutants in the <i>sbp</i> gene do not affect thiosulfate and sulfate binding or uptake [27].
<i>hycF</i>	Formate hydrogenlyase subunit HycF; enables utilization of formic acid as a source of carbon in <i>E. coli</i> .	HycF functions only under anaerobic conditions [28].
<i>tfaE</i>	Putative tail fiber assembly protein TfaE.	No study of the <i>tfaE</i> gene has yet been conducted.
<i>ycjX</i>	Putative protein YcjX.	No study of the <i>ycjX</i> gene has yet been conducted.

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