insects

Article

# Morphological and Molecular Analysis of Australian Earwigs (Dermaptera) Points to Unique Species and Regional Endemism in the Anisolabididae Family 

Oliver P. Stuart ${ }^{1}$, Matthew Binns ${ }^{1,2}$, Paul A. Umina ${ }^{1,3}$, Joanne Holloway ${ }^{4}$, Dustin Severtson ${ }^{5}$, Michael Nash ${ }^{6,7}$, Thomas Heddle ${ }^{8}$, Maarten van Helden ${ }^{6,8}$ and Ary A. Hoffmann 1,*<br>${ }^{1}$ School of BioSciences, Bio21 Molecular Science and Biotechnology Institute, The University of Melbourne, Parkville, Victoria 3052, Australia; oliver.stuart93@gmail.com (O.P.S.); matthew.binns@csiro.au (M.B.); pumina@unimelb.edu.au (P.A.U.)<br>2 Agriculture and Food Business Unit, Commonwealth Scientific and Industrial Research Organisation, Black Mountain, Australian Capital Territory 2601, Australia<br>${ }^{3}$ cesar, 293 Royal Parade, Parkville, Victoria 3052, Australia<br>${ }^{4}$ New South Wales Department of Primary Industries, Wagga Wagga Agricultural Institute, Pine Gully Road, Charles Sturt University, New South Wales 2795, Australia; joanne.holloway@dpi.nsw.gov.au<br>5 Department of Primary Industries and Regional Development, South Perth, Western Australia 6151, Australia; dustin.severtson@dpird.wa.gov.au<br>6 School of Agriculture, Food and Wine, the University of Adelaide, Urrbrae, South Australia 5064, Australia; Maarten.VanHelden@sa.gov.au<br>7 School of Life Science, College of Science, Health and Engineering, La Trobe University, Bundoora, Victoria 3086, Australia; whatbugsyou@gmail.com<br>8 South Australian Research and Development Institute, Entomology, Waite Road, Waite, Urrbrae, South Australia 5064, Australia; thomas.heddle@sa.gov.au<br>* Correspondence: ary@unimelb.edu.au

Received: 1 February 2019; Accepted: 7 March 2019; Published: date

## Supplementary Materials

Table S1. Location of field sites from which adult Dermaptera were collected.

| $\#$ | Site ID | State | Latitude | Longitude |
| :---: | :---: | :---: | :---: | :---: |
| 1 | Muradup | WA | -33.8380 | 116.9522 |
| 2 | Frankland River | WA | -34.4123 | 117.0167 |
| 3 | Kojonup | WA | -33.8426 | 117.2405 |
| 4 | Mt. Barker | WA | -34.6348 | 117.5299 |
| 5 | Tenterden | WA | -34.3549 | 117.6573 |
| 6 | Amelup | WA | -34.3049 | 118.1039 |
| 7 | South Stirling | WA | -34.6245 | 118.2758 |
| 8 | Needilup | WA | -33.9553 | 118.7607 |
| 9 | Boxwood Hill | WA | -34.4059 | 118.8616 |
| 10 | Jacup | WA | -33.7822 | 119.1235 |
| 11 | Fitzgerald | WA | -33.6170 | 119.5841 |
| 12 | Wittenoom Hills | WA | -33.5208 | 122.2667 |
| 13 | Kapinnie | SA | -34.1155 | 135.5122 |
| 14 | Oleo Rd | SA | -34.3813 | 135.8071 |
| 15 | North Shield | SA | -34.6090 | 135.8298 |
| 16 | Yallunda Flat | SA | -34.3221 | 135.8397 |
| 17 | Warooka | SA | -34.9862 | 137.3193 |
| 18 | Petersville | SA | -34.3465 | 137.7843 |
| 19 | Manoora | SA | -33.9520 | 138.8513 |


| 20 | Milang | SA | -35.3834 | 138.9341 |
| :--- | :---: | :---: | :---: | :---: |
| 21 | Frayville | SA | -34.8256 | 139.2424 |
| 22 | Lake Hawdon South | SA | -37.2316 | 140.0259 |
| 23 | Bool Lagoon | SA | -37.2049 | 140.7111 |
| 24 | Inverleigh | VIC | -38.0862 | 143.9344 |
| 25 | Elmore | VIC | -36.4863 | 144.5513 |
| 26 | Finley | NSW | -35.6258 | 145.6040 |
| 27 | Devenish | VIC | -36.3327 | 145.8903 |
| 28 | Thoona | VIC | -36.3031 | 146.0989 |
| 29 | Traralgon | VIC | -38.2013 | 146.6533 |
| 30 | Grong Grong | NSW | -34.7250 | 146.7452 |
| 31 | Downside 1 | NSW | -34.9553 | 147.3101 |
| 32 | Downside 2 | NSW | -34.9574 | 147.3113 |
| 33 | Peak Hill | NSW | -32.6696 | 148.0390 |
| 34 | Wallendbeen | NSW | -34.5450 | 148.1110 |
| 35 | Canowindra | NSW | -33.4256 | 148.6678 |
| 36 | Tamworth | NSW | -31.1457 | 150.9742 |

Numbers refer to labels on Figure S1.

Table S2. Summary statistics for morphometric variables for the Anisolabididae morphospecies.

| Morphospecies | N | Paramere length | Paramere width | Maximum forceps length | Maximum forceps width | Forceps asymmetry index | Paramere length-width ratio | Basal width of forceps | Forceps lengthwidth ratio |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Anisolabis sp. 1 | 12 | 1.189 (0.092) | $\begin{gathered} \hline 0.396 \\ (0.044) \\ 0.302- \end{gathered}$ | 3.182 (0.501) | 2.476 (0.597) | 0.483 (0.196) | 3.027 (0.284) | 3.490 (0.627) | 1.320 (0.197) |
|  |  | 1.057-1.393 | 0.459 | 2.517-3.885 | 1.689-3.280 | 0.204-0.909 | 2.678-3.785 | 2.765-4.419 | 1.058-1.693 |
| Anisolabis sp. 2 | 18 | 1.087 (0.058) | $\begin{gathered} 0.283 \\ (0.027) \\ 0.229- \end{gathered}$ | 3.073 (0.302) | 1.499 (0.150) | 0.148 (0.045) | 3.862 (0.354) | 2.392 (0.177) | 2.064 (0.241) |
|  |  | 0.974-1.198 | 0.325 | 2.468-3.633 | 1.190-1.754 | 0.067-0.219 | 3.361-4.633 | 1.945-2.624 | 1.750-2.744 |
| Gonolabis forcipata | 23 | 0.644 (0.034) | $\begin{gathered} 0.415 \\ (0.024) \\ 0.370- \end{gathered}$ | 2.204 (0.260) | 1.471 (0.217) | 0.098 (0.040) | 1.558 (0.100) | 2.551 (0.280) | 1.513 (0.172) |
|  |  | 0.583-0.715 | 0.466 | 1.717-2.849 | 1.122-2.049 | 0.047-0.209 | 1.421-1.806 | 1.987-3.368 | 1.249-2.041 |
| Gonolabis nr. gilesi | 14 | 0.613 (0.043) | $\begin{gathered} 0.353 \\ (0.024) \\ 0.301- \end{gathered}$ | 1.810 (0.318) | 1.402 (0.175) | 0.176 (0.147) | 1.744 (0.149) | 2.344 (0.237) | 1.302 (0.242) |
|  |  | 0.533-0.709 | 0.398 | 1.506-2.733 | 1.240-1.914 | 0.014-0.609 | 1.481-1.937 | 2.012-2.760 | 1.000-1.995 |
| Gonolabis sp. 1 | 32 | 0.529 (0.032) | $\begin{gathered} 0.422 \\ (0.026) \\ 0.369- \end{gathered}$ | 3.434 (0.315) | 1.730 (0.272) | 0.051 (0.043) | 1.255 (0.074) | 2.927 (0.201) | 2.036 (0.394) |
|  |  | 0.472-0.590 | 0.484 | 2.859-4.116 | 1.276-2.257 | 0.001-0.174 | 1.129-1.463 | 2.579-3.320 | 1.498-2.868 |
| Gonolabis sp. 2 | 22 | 0.658 (0.039) | $\begin{gathered} 0.452 \\ (0.026) \\ 0.419- \end{gathered}$ | 2.469 (0.206) | 1.489 (0.162) | 0.078 (0.036) | 1.459 (0.086) | 2.827 (0.230) | 1.674 (0.190) |
|  |  | 0.549-0.721 | 0.521 | 2.121-2.968 | 1.138-1.739 | 0.020-0.148 | 1.286-1.675 | 2.403-3.378 | 1.343-2.104 |


| Gonolabis sp. $3 \quad 3 \quad$$0.616(0.084)$ 0.410 <br> $(0.031)$ <br> $0.381-$ $2.390(0.431)$ $1.332(0.341)$$\quad 0.072(0.011)$ |
| :--- |
| $0.520-0.677$ |

Table S3. Details of linear discriminant function analysis.

| Classification | Function | Variable |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Basal width of forceps | Forceps asymmetry index | Paramere length-width ratio | Forceps lengthwidth ratio |
| Genus | 1 (NA) | -1.568 | -3.061 | -4.107 | -0.572 |
| Anisolabis | 1 (NA) | -0.593 | 0.216 | 2.267 | 3.865 |
| Gonolabis | 1 (0.966) | 1.427 | -1.653 | -8.550 | 1.534 |
| Gonolabis (G. sp. 3 removed) | 2 (0.024) | -3.802 | -0.224 | -1.078 | 2.272 |
|  | 3 (0.009) | 0.910 | 14.878 | -1.398 | 0.568 |
|  | 4 (0.001) | 1.012 | -2.706 | 5.547 | 2.375 |
|  | 1 (0.975) | 1.245 | -1.720 | -8.600 | 1.588 |
|  | 2 (0.018) | -3.639 | 4.728 | -2.290 | 2.055 |
|  | 3 (0.007) | -2.266 | -13.008 | -0.513 | -0.477 |

[^0]Table S4. Uncorrected p-distances within and between earwig species on the Barcode of Life Data Systems public repository's cytochrome oxidase c subunit 1 barcodes (507-680 bp).

|  | N | Chelidura guentheri | Forficula auricularia A | Forficula auricularia B | Labidura riparia | Nala <br> lividipes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Chelidura guentheri | 6 | $\begin{array}{r} 0.001 \\ (0.001) \end{array}$ | 0.020 | 0.020 | 0.019 | 0.020 |
| Forficula auricularia A | 558 | 0.231 | $\begin{array}{r} <0.001 \\ (<0.001) \end{array}$ | 0.015 | 0.020 | 0.020 |
| Forficula auricularia B | 130 | 0.220 | 0.102 | $\begin{array}{r} 0.003 \\ (0.001) \end{array}$ | 0.019 | 0.020 |
| Labidura riparia | 6 | 0.189 | 0.207 | 0.205 | $\begin{array}{r} 0.006 \\ (0.002) \end{array}$ | 0.017 |
| Nala lividipes | 5 | 0.199 | 0.197 | 0.196 | 0.147 | $\begin{array}{r} 0.007 \\ (0.002) \\ \hline \end{array}$ |

Only records with species names attached and with sequences > 500 bp were retrieved. Interspecific means and variances are shown on the left and right of the diagonal. Intraspecific mean shown on diagonal with variance in brackets. Variances were calculated using 1000 bootstraps. N refers to total individuals retrieved. Forficula auricularia A and B refers to cryptic sibling species [1].


Figure S1. Map of field sites from which adult Dermaptera were collected. Numbers refer to sites listed in Table S1. Axes represent longitude and latitude.


Figure S2. Gene trees inferred from alignment of mitochondrial sequence data (1016 bp) for Anisolabididae morphospecies ( 658 bp from cytochrome $c$ oxidase subunit 1 and 358 bp from cytochrome $b$ ). Node labels represent bootstrap support from 1000 bootstraps ( $a$ and $b$ ) and posterior probability (c). (a) Best tree inferred under a maximum likelihood framework in RAxML using a general time reversible model of sequence evolution with a gamma-distributed rate parameter for site evolution estimated empirically; log-likelihood $=-5689.987$. (b) Tree inferred exactly as for (a) with a constraint tree representing the known morphospecies imposed on the search space; log-likelihood =
-5689.989. (c) Maximum clade credibility tree inferred using *BEAST, with models of site evolution estimated simultaneously to tree estimation using bModelTest and a birth-death process for the tree prior; 95\% HPDI of tree log-likelihood $=[-5276.756,-5254.856]$, mean log-likelihood $=-5265.388$.


Figure S3. Gene trees inferred from alignment of a fragment of $28 S$ ribosomal DNA ( 442 bp ) from Anisolabididae morphospecies. Node labels represent bootstrap support from 1000 bootstraps (a and b) and posterior probability (c). (a) Best tree inferred under a maximum likelihood framework in RAxML using a general time reversible model of sequence evolution with a gamma-distributed rate parameter for site evolution estimated empirically; log-likelihood $=-1087.959$. (b) Tree inferred exactly as for (a) with a constraint tree representing the known morphospecies imposed on the search space; log-likelihood $=-1087.959$. (c) Maximum clade credibility tree inferred using *BEAST, with models of site evolution estimated simultaneously to tree estimation using bModelTest and a birthdeath process for the tree prior; 95\% HPDI of tree log-likelihood $=[-1123.086,-1102.800]$, mean loglikelihood $=-1114.919$.


Figure S4. Gene trees inferred from alignment of a fragment of tubulin-alpha 1 (466 bp) from Anisolabididae morphospecies. Node labels represent bootstrap support from 1000 bootstraps (a and b) and posterior probability (c). (a) Best tree inferred under a maximum likelihood framework in RAxML using a general time reversible model of sequence evolution with a gamma-distributed rate parameter for site evolution estimated empirically; log-likelihood $=-1183.262$. (b) Tree inferred
exactly as for (a) with a constraint tree representing the known morphospecies imposed on the search space; log-likelihood $=-1184.068$. (c) Maximum clade credibility tree inferred using *BEAST, with models of site evolution estimated simultaneously to tree estimation using bModelTest and a birthdeath process for the tree prior; $95 \%$ HPDI of tree log-likelihood $=[-1086.380,-1054.702]$, mean loglikelihood $=-1066.792$.

## Reference

1. Wirth, T.; Le Guellec, R.; Vancassel, M.; Veuille, M. Molecular and reproductive characterisation of sibling species in the European earwig (Forficula auricularia). Evolution 1998, 52, 260-265.
© 2019 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).

[^0]:    "Classification" refers to the grouping variable used (i.e., "Genus" refers to the discrimination of the two genera, "Anisolabis" to that between Anisolabis spp., etc.). Function refers to the individual discriminant functions generated, of which there can be a maximum of $k-1$, where $k$ is the number of levels of the grouping variable (morphospecies or genera in this case). Functions are numbered in order of the proportion of variance explained by each, which is shown in brackets. "Forceps asymmetry" is an index calculated as the sum of the absolute differences between the heights and widths of a specimen's two forceps, "Paramere ratio" is the ratio of a specimen's right paramere's length to its width, and "Forceps ratio" is the ratio of the maximum forceps length to the maximum forceps width. These three measurements were dimensionless; the basal width of the forceps was in mm.

