

Perspective

Circuits and Biomarkers of the Central Nervous System Relating to Astronaut Performance: Summary Report for a NASA-Sponsored Technical Interchange Meeting

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Citation: Alwood, J.S.; Mulavara, A.P.; Iyer, J.; Mhatre, S.D.; Rosi, S.; Shelhamer, M.; Davis, C.; Jones, C.W.; Mao, X.W.; Desai, R.I.; et al. Circuits and Biomarkers of the Central Nervous System Relating to Astronaut Performance: Summary Report for a NASA-Sponsored Technical Interchange Meeting. *Life* **2023**, *13*, 1852. <https://doi.org/10.3390/life13091852>

Academic Editors: Larry D. Sanford and Richard A. Britten

Received: 15 June 2023

Revised: 24 August 2023

Accepted: 25 August 2023

Published: 31 August 2023



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Abstract: Biomarkers, ranging from molecules to behavior, can be used to identify thresholds beyond which performance of mission tasks may be compromised and could potentially trigger the activation of countermeasures. Identification of homologous brain regions and/or neural circuits related to operational performance may allow for translational studies between species. Three discussion groups were directed to use operationally relevant performance tasks as a driver when identifying biomarkers and brain regions or circuits for selected constructs. Here we summarize small-group discussions in tables of circuits and biomarkers categorized by (a) sensorimotor, (b) behavioral medicine and (c) integrated approaches (e.g., physiological responses). In total, hundreds of biomarkers have been identified and are summarized herein by the respective group leads. We hope the meeting proceedings become a rich resource for NASA's Human Research Program (HRP) and the community of researchers.

Keywords: biomarker; cognition; behavior; performance; brain circuit; astronaut; CNS

1. Introduction

Astronauts on long-duration space missions (e.g., transits to Mars) will experience the combined, potentially synergistic, impacts of simultaneous exposures to spaceflight hazards that affect the central nervous system (CNS) and operationally relevant behavior and performance [1]. While individual spaceflight hazards are often individually well quantified, in long-duration spaceflight, astronauts will experience multiple hazards simultaneously [2,3].

Parcelsus' famous dictum on dose effects of exposures [4] reinforces the importance of an integrated approach to systematically identify and investigate the relationships of how spaceflight exposures may synergistically interact to pose a risk to the astronauts and the mission. NASA developed the Combined Behavioral Stressors (CBS) project which integrates research topics across three high-impact spaceflight hazard exposures – space radiation, isolation & confinement, and altered gravity -- to inform performance outcome limits and permissible exposure limits, and to help identify and establish mitigation strategies. An integrated research approach is focused on identifying biomarker changes associated with exposures to the CBS-associated hazards to identify and develop effective monitoring, and apply countermeasures for mitigating risk to crew health and performance [5]. This is consistent with recent calls for more comprehensive and integrated biomarkers to better identify how different biomarkers can exert different causal effects between and among them [6].

The CBS Integrated Research Plan identifies biomarkers that are linked to in-flight and post-flight decrements in an astronaut's operational performance resulting from simultaneous exposures to the CBS-relevant spaceflight hazards. In this context, a biomarker is defined as a characteristic that is "objectively measured and evaluated as an indicator of normal biologic processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention" [7].

As sampling of in situ biomarkers in astronauts is not necessarily possible, translational models are useful. To promote the utility of translational models, NASA consistently updates the exposure levels in rodents as they relate to humans; for example, NASA recently adjusted their integrated research platforms involving animal exposures to expected levels of spaceflight radiation related to dose and duration [8]. It is, therefore, essential that biomarkers are useful for bi-directional translation of homologous human and animal measures, which is a cornerstone of the NASA's CBS project—allowing for the linking of the probability for performance decrements (during and/or after mission) to the level of exposure to a CBS relevant spaceflight hazard, such as radiation exposure.

This paper reviews the results of NASA's biomarker technical interchange meeting (TIM) that was focused on creating a comprehensive list of constructs, identifying underlying and related brain regions, neural circuits, and biomarkers for inclusion in predictive models to assess and validate changes in future astronaut risk status, as well as to identify changes in operationally relevant brain pathways (e.g., procedural memory) after exposures to varying types and amounts of potentially synergistically acting spaceflight hazards. The overall goals of this biomarker TIM were to (i) identify relevant brain regions, neural circuits, functions, and associated biomarkers, and relate them to operationally relevant performance, and (ii) identify any critical needs for new biomarker knowledge ("gaps") that can be filled by additional focused and translational animal experiments that include a plausible pathway toward eventual biomarker validation in humans.

2. Meeting Synopsis

Biomarkers—ranging from molecules to behavior—can be used to identify thresholds beyond which performance of mission tasks may be compromised and could potentially trigger the activation of countermeasures. Identification of homologous brain regions and/or neural circuits related to operational performance may allow for translational studies between species. Three discussion groups were asked to use operationally relevant performance tasks as drivers when identifying biomarkers and regions or circuits for the constructs listed in Appendix A. Participants are listed in Appendix B. Here, we summarize the discussions below across the three groups. In total, hundreds of biomarkers have been identified, with references provided mainly in the respective tables for each group. We hope the meeting proceedings become a rich resource for NASA's Human Research Program (HRP) and the community of researchers.

3. Summaries of Discussions and Recommendations from Each of the Breakout Sessions

3.1. Sensorimotor Influences on Operational Performance (Leads: S. Rosi, M. Shelhamer)

The goal of Group 1 was to create lists of biomarkers and brain regions and/or neural circuits related to operational performance for constructs that are prioritized in HRP's sensorimotor risk. Group 1 assessed the following 13 key constructs in Table 1: visual function, spatial orientation, vestibular, proprioception, hearing, motion sickness, smell and taste, postural control and balance, locomotion, fine motor control, perception, gaze, and pain. Note that the panel assessed translatability based on the existence of rodent models and did not suggest using non-human primates (NHPs), nor did they identify a construct that should be tested in NHPs.

3.1.1. Summary of Discussions

During discussion of each of the 13 constructs, 10 themes emerged. Although identification of themes was outside the scope of the panel, these themes were applicable to nearly all constructs discussed and, therefore, we define them here:

1. Connections between constructs. Distinctions between the constructs are, in many cases, artificial. Although segregated disciplinary expertise has achieved a great deal in the sensorimotor domain, the different constructs are so closely interconnected that it is hard to discuss them separately in a way that is true to the science and to the operational implications. As an example, vestibular function, gaze control, balance, and locomotion are very closely related, and yet they are often addressed as specific and separable. Another example is perception. Almost all sensorimotor constructs involve perception in some way; vestibular perception—perception of the upright—affects the ability to balance. Perception of upright is influenced by changes that occur in microgravity, which is a vestibular effect. Again, these specific constructs become tightly entangled and it is difficult to separate them in terms of biomarkers and operational relevance.

2. Many spaceflight stressors and sensorimotor effects occur simultaneously with different time courses. Not only do the different constructs interact, they do so with different time courses. The most overt and acute forms of vestibular adaptation (related to space motion sickness) occur over the course of a few days, whereas other vestibular-mediated functions (e.g., the sense of being truly comfortable with the three-dimensional aspects of motion in a weightless environment) develop over several weeks. Some adaptive sensorimotor changes in space occur with similar time courses as those seen in analogous environments on the ground. For example, the changing contributions of vestibular, proprioceptive, and efference copy information during recovery from labyrinthectomy in an animal model [9] have time courses that mimic recovery of motor control during locomotion after spaceflight [10]. Similarly, ground-based studies in animals show that development of efference copy over several weeks mimics the time course of the development of three-dimensional spatial sense in astronauts over the same time period. The similar time courses suggest that these may be aspects of the same underlying process. This might provide translational opportunities from ground-based animal models and may inform a process for preadaptation paradigms for spaceflight.

3. Multi-sensory integration. This is related to the theme of interacting constructs. Most sensorimotor behaviors and perceptions arise from the simultaneous activation of multiple sensory systems. An obvious example is the combination of visual and vestibular information for gaze control (vestibulo-ocular reflex (VOR)). Another is the prevalence of proprioceptive and kinesthetic influences, in addition to vestibular and visual influences, on posture and locomotion.

4. Stress. Spaceflight involves multiple simultaneous stressors—physiological, psychological, and environmental. These have widespread and sometimes unknown influences on sensorimotor function, and likely on the ability to adaptively alter sensorimotor function. The effects of stress on motor learning and on motion sickness are two examples: stress affects motor learning, which alters adaptation, which can change the ability to recover from motion sickness, which can increase stress.

5. Learning. Almost all the individual constructs exhibit adaptive behaviors to spaceflight and these adaptive behaviors may complicate the usefulness of the constructs as biomarkers because the response that is being assessed will change with adaptation to spaceflight. Of course, such adaptation is desirable and should be promoted, but it complicates the use of a biomarker to identify increased risk to astronaut health and performance. This would be especially true in missions of extended duration where the adaptive processes might not be understood. A specific biomarker for learning and adaptation would be desirable.

6. Some constructs might be easily measured but lack relevance. As an example, the angular VOR has been extensively studied and is easy to measure, but little or no evidence exists that it changes significantly due to spaceflight, or that any changes have an operational impact.

7. Neural circuits. Interpretation of neural circuitry is not always straightforward. There is not always a direct analogy between animals (where many circuits have been delineated) and humans; the neural circuitry is different in some cases, and there are also adaptive changes that make the definition of standard circuits difficult. Circuit function is implicitly assessed with behavioral measures, so knowledge of some circuit characteristics such as neurotransmitters and common pathways might aid in the interpretation of behavioral markers.

8. Vestibular Cognition. The relationship between cognition and the vestibular system, and the vestibular effects on cognition, is operationally relevant and directly connects cognition and sensorimotor functions. This connection is seen in many patients with vestibular problems. No specific construct exists for this, and it is difficult to conceive of a specific biomarker.

Overall, the sensorimotor issues of multi-sensory/multi-effector interactions and learning, and their relation to stress, are not yet sufficiently studied, and they likely greatly influence human performance in space. These do not yet lend themselves to direct biomarker identification.

3.1.2. Recommendations

The panel evaluated each specific construct to determine if a good biomarker exists that is operationally relevant for astronauts, and that translates from animal models. The panel also commented on gaps in each construct that would need to be filled to produce an effective biomarker.

1. Visual function is easily measured (acuity, visual fields, etc.), and these measures may help to parse out visual effects from motor effects when there is a functional deficit. Retinal remodeling can be assessed with optical coherence tomography (in flight), and is hence a biomarker. Translatability is clear because many of these aspects can be tested in rodents (e.g., visual acuity in mice and even real-time visual tracking). This is clearly a useful biomarker.

2. Spatial orientation is extremely important. The panel extensively discussed grid cells—the cells in the entorhinal cortex that underlie spatial orientation. The firing of grid cells provides information that can be used to assess spatial orientation as it adapts to alterations in gravity, which is further substantiated as a potential biomarker due to its translational potential as grid cells are present and accessible in rodents. Thus, neural circuits in the hippocampus and medial entorhinal cortex are important.

3. A great deal of information exists on vestibular function in spaceflight. Basic vestibular function is not significantly altered in the microgravity environment of space, although central processing and higher-level derived functions (e.g., spatial orientation, tilt-translation perception) often are. It is, however, important to consider vestibular changes in the context of the integrated spaceflight stressors. So, as noted, the VOR changes little in weightlessness, but it would be useful to assess VOR in the context of other stressors (e.g., radiation, fatigue, etc.); for example, what is the combined impact of multiple stressors? These aspects need to be elucidated, which can be accomplished through rodent studies (e.g., the narrow balance beam as a viable animal assessment). Taken together, vestibular change (e.g., VOR or balance beam performance) is a suitable biomarker.

4. Proprioception was identified as one of the most strongly interconnected constructs, exhibiting significant overlap with several other constructs. Little is known about the effects of (CBS risks) radiation or other stressors on the peripheral nervous system and, consequently, proprioception (this is a gap in knowledge). A rodent model would provide translational opportunities, as proprioception can be measured in that model (e.g., tape removal test, whisker test). Hence, measures of proprioception are suitable biomarkers.

5. Hearing loss is often a factor associated with spaceflight, perhaps due in part to fluid shifts, and hearing assessment in flight may help to parse out the effect of the fluid shift from noise-induced loss. However, the panel noted that these data are not particularly operationally relevant: hearing loss has not been a functional problem. As such, hearing loss is not a priority biomarker.

6. Motion sickness is a known problem that needs to be further assessed because it can have serious operational impacts [11,12], especially when first experiencing a gravity field after extended weightlessness. Motion sickness susceptibility is still unpredictable. This line of work might be revisited with more recent knowledge on learning and adaptation or might be investigated in relation to the impact on specific operational tasks. We do not know how motion sickness induces stress and how stress feeds back to motion sickness and the overall well-being of astronauts. The interaction of motion sickness, so-pite, stress, and crew performance has been studied in other contexts. This work should be reviewed; however, it may still be valuable to investigate these effects in the specific context of spaceflight, with its multiple simultaneous stressors and unique demands. Again, there are several overlapping biomarkers. A drawback in this area is translatability, because it is very difficult to measure motion sickness in rodents. This is a useful biomarker, albeit with some uncertainties as to translational aspects.

7. Smell and taste are particularly important for humans as social creatures and are also clearly important in space. These constructs overlap with the well-being and operational performance of astronauts. Smell and olfaction can be markers for neurodegeneration. Loss of olfaction (anosmia) is an early marker in COVID-19 and Alzheimer's disease, as examples, and is therefore a biomarker for neurodegeneration that can also easily be tested in rodents. This biomarker is rated highly.

8. Posture and balance are important operational issues. They are problematic as biomarkers because, again, their functions cannot be isolated to discrete neural circuits due to the overlap of several circuits for multi-sensory integration and motor control. Rodent models are somewhat problematic because of the difference between neural circuits and functions in organisms with four legs (rodents) relative to two legs (humans).

9. As with posture and balance, locomotion is operationally relevant and important, but good rodent models in spaceflight or microgravity environments are lacking. It might be useful to consider static/dynamic balance control as opposed to posture/locomotion.

10. Fine motor control is difficult to assess because of the large number of confounders. Related factors that can alter fine motor function include changes in proprioception, hand-eye coordination, and others. Although functionally important, it may not be particularly relevant for operational control tasks, and suitable rodent models are lacking. The many confounders alone make this problematic as a discrete biomarker.

11. Perception is in fact a component of almost all the other constructs because it can include spatial orientation, depth perception, vestibular orientation, time perception, and others. Understanding of this construct is important and would address many of the other constructs, but there are many overlaps. Proprioception may be altered and is a critical issue on its own, but it will be most important to address in the context of other stressors. Specific aspects of perception have been noted in spaceflight and can have operational impacts, and so it would also be beneficial to consider perception in this performance context. Nevertheless, parsing out perceptual effects per se remains difficult. Thus, this was not considered to be a good biomarker.

12. The panel did not rate gaze and pain highly as biomarkers. Gaze largely overlaps vestibular function (and has been studied almost as much), so gaze control can be subsumed under vestibular function. Pain per se is not a good biomarker because of confounders between the perception and the sensation of pain. Nociception can depend on sex and other individual factors. Although biomarkers of inflammation exist, these are associated with pain. Hence, pain itself is not a discrete biomarker.

Table 1. Circuits and biomarkers for sensorimotor domains.

Key Indicator/Construct	Human Performance Test (Details about the Actual Test/Assay)	Animal Performance Test (Details about the Actual Test/Assay)	Caveats/Notes/Related Functional Performance Tasks/Prediction of Behavioral Outcome in Humans	Brain Region	Human/NHP Neural Circuit/Pathways	Rodent Neural Circuit/Pathway	Biomarkers (Rodents/Humans/NHPs)		
							Inaccessible	Accessible (Translatable to Astronauts)	Gaps/Notes
Visual	(1) Visual field testing	(1) Visual field testing		Visual cortex (Occipital lobe of the primary cortex)	<u>Retino-geniculate-striate pathway (Conscious vision)</u> Dorsal pathway (spatial location and action): Retina → LGN → V1 → V2 → MT (parietal lobe) Ventral pathway (characteristics of objects): Retina → LGN → V1 → V2 → V4 (temporal lobe) [13]	Retina-Superior Colliculus-Lateral posterior nucleus-Visual cortex1 pathway [14]	Retinal markers- autopsy, superior colliculus pathway— neural circuitry, intracranial pressure in astronauts— lumbar puncture for pressure detection, retinal vasculature imaging— vessel length density and loss of photo receptor cells, role of endothelial structure or vasculature, acceleration of incident of cataract (on cornea, not CNS)	Imaging: Inflight CT, MRI imaging, ultrasound, OCT, visual field measurements, cataract as predictor Structural changes in eye, nerve, occipital cortex, pretectum, superior colliculus.	(1) Potential Optical/Eye damage in astronauts— could also be indicator of neurological symptoms. (2) Any imaging other than ultrasound is difficult to do in space. Difficult to get a gold standard test for

						and light flashes (post-flight and long-term issue), fluorescent imaging of the retinal vasculature.	Vision function test, sampling of tears [15], Intraocular pressure measurement, Saccades [16], Behavioral measures, Live pupil tracking	intracranial pressure in space. (3) Possibility of lumbar punctures in astronaut— intracranial pressure. (4) VR environments for complex sensory integration— Somatosensory component	
Spatial Orientation	1. Path integration-passive and active 2. Virtual maze perspective taking tests 3. Visual object	1. Changes in activity of head direction, grid, place cells 2. Morris water maze 3. Spatial navigation	- Test in higher animals: NHP - Spatial navigation	Hippocampus and parahippocampal regions, cerebellum, brain stem, Retrosplenial cortex (Grid cells, border cells, head direction	Vestibulospinal pathway	<u>Proposed head direction pathway 1:</u> Vestibular nuclei (VN) → Cerebellum → ventral lateral nucleus of thalamus (VLN) → parietal cortex → temporal	Hippocampal protein lysate: Afg3l1, Tpx2, Neuroligin-3, RB1-inducible coiled-coil 1, Mast3, Kif21a, DnaJ (Hsp40) homolog, SLIT-ROBO Rho GTPase-activating protein 2, Rasgrf1 [20]	Structural changes in hippocampus, anterior thalamus, subiculum. Electrodermal activity measured by wrist worn device [21],	(1) Virtual reality biomarker development for astronauts. (2) Spatial orientation during g-transitions

	learning (VOLT)	4. Touch screen cognitive testing [17].		cells—cortical regions-egocentric and allocentric reference frame) [18]		cortex → hippocampus? <u>Proposed head direction pathway 2:</u> Vestibular nuclei (VN) → hippocampus [19]	Optical coherence tomography (OCT), Illusionary experience, somatographic illusion— questionnaire	(3) Different species have varied responses. Need a model that would be most translatable.	
Vestibular	1. Drop test/Jump down test 2. VEMP 3. OVAR response (Sensorimotor component after 30 rpm) 4. Time constant or constant rotation 5. ocular counter roll (but noisy)	1. Balance beam test (narrow beam) (2) Righting reflex 2. VEMP (can be done in space and can help distinguish utricular and saccular functions) 3. OVAR response	Test in higher animals: NHP	Thalamus and cortex	Thalamocortical pathways Anterior vestibulothalamic pathway: Vestibular nuclei (VN) → Nucleus prepositus and supragenual nucleus (NPH/SGN) → Anterior dorsal thalamus (ADN) → Entorhinal cortex → Hippocampus Posterior vestibulothalamic pathway: Vestibular nuclei (VN) →	(1) Vestibular nucleus → Dorsal tegmental nucleus (DTN) → Lateral mammillary nucleus (LMN) → Anterodorsal nucleus (ADN) → Post-subiculum (PS) → Hippocampus (2) Vestibular nucleus → Pedunculopontine tegmental nucleus (PPTN)	Otopetrin1, Alpha 2 adrenergic receptors [23], Glutamate receptor expression [24], c-FOS, vestibular hair cells [25], cerebellar nodulus of adult rats [26–28], TEM of synaptic ribbons [29–33]	Nausea related—cardiac sensitivity to baroreceptor reflex; raised Heart rate; raised cortisol; reduced dominant power on EGG baseline, questionnaire [34,35], Serum: NSE	(1) Effects of stress on vestibular compensation and adaptation. (2) Social stress, performance anxiety, other psychological stress—will it impede recovery? (3) Stress impedes motor

	4. Active vs. Passive motion on vestibular nucleus neurons			Ventral posterior lateral nucleus (VPL) → vestibular cortical areas. [9] - Three neuron pathway Vestibulo-ocular reflex...	→ supramammillary nucleus SUM → Medial septum → Hippocampus (3) Vestibular nucleus → Thalamus → Parietal cortex → Entorhinal/Perirhinal cortices → Hippocampus [22].	and S100β learning in [36], Otolin-1 mice (Fragile X mice). vibration-induced nystagmus [38]
	5. VSEP (otolith function)			--> vestibular nuclei		
	6. Swimming test (for subtle deficits, screening test)			--> Vestibulo-ocular reflex and efferent (vestibular processing)		
Gaze	1. Gaze Holding/Gaze stability		Visual pathway, Frontal eye fields, vestibular nuclei, cerebellum, oculomotor system, parietal cortex, postcentral	<u>Horizontal vestibular-generated eye movement:</u> Horizontal semicircular canal → Vestibular nucleus (Vestibular ganglion) and cerebral cortex inputs (frontal eye field) →		Structural changes in cerebellum (conventional and mass-spec imaging), Diplopia, Blurring of vision, vestibulo-ocular reflex.
	2. Eye-head coordination	1. Gaze Holding	Test in higher animals: NHP			
	3. Redirecting gaze					

				gyrus, Entorhinal cortex neurons	Paramedian pontine reticular formation (PPRF or gaze center) → Medial longitudinal fasciculus (MLF) → ipsilateral lateral rectus muscle (eye) and contralateral medial rectus muscle (eye) [39].	Gaze holding/stability and ability to redirect the gaze with accuracy—integrative Biomarker
Locomotion	<p>1. Tandem Walking (=Beam Walking in Animal);</p> <p>2. Perturbation during walking</p> <p>3. Navigating obstacle course while walking (eg. Functional</p>	<p>1. Rotarod walking (=tandem walking);</p> <p>3. Actigraphy in animals;</p> <p>4. Open field Test directly in humans when possible.</p>	<p>Animal model tests should be developed:</p> <p>a. DigiGait 2.0 Analysis with perturbation, belt or surface perturbation (=human perturbation during walking);</p> <p>b. Dual task test (Catwalk); c. Rodent obstacle course (=FMT)</p>	<p>Mesencephalic locomotor region (MLR) in the midbrain</p>	<p>(1) Reticulospinal pathway: Motor cortex → Basal ganglia → Mesencephalic locomotor region → Pons/Medulla (Reticulospinal cells) → Spinal cord/Central pattern generator → Muscle [40].</p> <p>(2) Vestibulospinal pathway</p>	<p>(1) Reticulospinal pathway (major pathway for initiating locomotion): Motor cortex → Basal ganglia → Thalamus → Mesencephalic locomotor region → Pons/Medulla → Spinal cord/Central pattern generator network → Muscle</p> <p>Behavioral tests. Locomotion and gait as a biomarker associated with NDs</p> <p>(1) Can be nested in vestibular, posture, and gait construct</p> <p>(2) Static Vs. Dynamic postural control is important</p>

<p>Mobility Test) 4. Statistical modeling of actigraphy data</p>	<p>(2) <u>Vestibulospinal pathway</u> (3) <u>Rubriospinal pathway</u> [41]</p>						
<p>Postural control, Balance</p>	<p>1. CDP. 2. Get up From Fall Test 3. Induced stepping (hold and release) 4. Body sway test (non-parallel two-leg model). 5 Engaged leg model of body sway (uneven weight distribution)</p>	<p>(1) Rotarod (2) Zebrafish Active Posturography (Zap); (3) Floating Platform Tests— Postural sway— measured by Center of Pressure (COP) Assay (=COP) Test directly in humans when possible.</p>	<p>Animal model tests should be developed: (a) Floating Platform Test (b) Motion Capture Analysis (exists but advanced version can be developed)</p>	<p>Cerebellum, sensorimotor cortex, vestibular cortex, prefrontal cortex</p>	<p>Postural information → Vestibular/Visual/Somatosenory input → Brainstem, cerebellum, thalamus → Temporoparietal cortex (vestibular cortex/posteroparietal cortex) → primary sensory cortex → Supplementary motor area and premotor area (info. integration from hippocampus) → basal ganglia/cerebellum (corticovestibular</p>	<p><u>Posture-head stabilization:</u> Inner ear vestibular receptors → vestibular nerve → ipsilateral vestibular nuclei in brain stem → vestibulocerebellum/medial vestibulospinal fasciculus → ipsi/contralateral projections → motor neurons (neck muscle) <u>Locomotion coordination:</u> Inner ear vestibular</p>	<p>Rodents: Circling, body sway area, the barycenter, the support surface and the weight distribution of the rats when they were moving or stationary [43]. (1) Operationally relevant. Need to evaluate before EVA (2) Animal models not so useful (2 vs. 4 leg)</p>

projections) → receptors →
 Brain stem → vestibular nerve
 Spinal cord → ipsilateral
 (reticulospinal tract) vestibular nuclei
 → Muscle [42]. in brain stem →
 striatum
 (thalamic
 relay)/Lateral
 vestibulospinal
 fasciculus →
 ipsilateral
 projections →
 locomotor central
 pattern generator
 → motor neurons
 (trunk and leg
 muscles) [43].

Motion sickness	<p>1. Graybiel scale (comprehensive) 2. Nausea (0 to 10) 3. Eye strain (0–10)</p>	<p>Not reliable in rodent. Ferrets have vomiting response. squirrel monkey and rhesus monkey—</p>	<p>Brain stem and Cerebellum</p>	<p>Input (Visual, Vestibular labyrinth, proprioceptive) → vestibular nuclei → cerebellum → brainstem autonomic centers → vomiting center [44].</p>	<p>Structural changes in inner ear. Increased plasma glucose [45], Nausea related— cardiac sensitivity to</p>	<p>(1) Study the effects of stress, sleep deprivation, head-loading, oscillation vibrations, prolonged fixation, and</p>
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difficult to
test

baroreceptor motion
reflex; raised sickness
Heart rate; (2) There are
raised enormous
cortisol; differences in
reduced individual
dominant susceptibility,
power on with respect
EGG to both
baseline sensitivity
[34,35] and
adaptation/rapid decay of
stimulus. So,
in long term
space
missions like
to Mars-
should we
pre-screen the
astronauts?
But
predicting
susceptibility
is unclear.
(3) How
relevant is it

									to astronaut performance considering it affects only during g transitions (~1% of their time in a 3 year mission). (4) Sopite syndrome— can affect operational performance —Combined effect. (5) Translatability -ferret and mouse model, tricky to track
Proprioception	1. Force and joint position test; 2. Dysmetria (finger to nose) test +/-	1. Von Frey Fibers; 2. Static force von Frey 3. Two-choice	Animal model tests should be developed: a. Force and joint position test; b. No identified	Thalamus, Somatosensory cortex, cerebellum, vestibular cortex,	<u>Dorsal Column pathway:</u> Proprioceptors → Spinal cord → Nucleus cuneatus (Medulla) →	<u>Thalamo-insular pathway</u> [46] Proprioceptive signals from Jaw-closing muscle spindles (JCMSs)	Piezo2 [47], Erg3 transcript levels [48]. Transient receptors which are responsive to camphor, menthol, and capsaicin to	<u>fMRI and Diffusion tensor imaging (DTI):</u> structural	(1) Very little data from peripheral nervous system and spinal cord.

eyes closed; 3. Foot sensitivity via pressure algometry (provides objective measure) = Von Frey Fibers; 4. Thesiometry, vibration at different frequency ranges for slow or fast adapting sensors 5. Tendon tap test, tonic vibrations? complementi ng Hoffman reflexes	mechanosens ory assay 4. Cotton swab assay 5. Tail Clip assay 6. Tape response assay 7. Hargreaves assay 8. Randall- Selitto assay 9. Complete Freund's adjuvant with von Frey 10. Bradykinin with von Frey 11. Two temperature choice assay.	animal equivalent of dysmetria	prefrontal cortex, Right putamen, parietal cortex, mouse barrel cortex (homunculus)	Ventral Posterior lateral nucleus (Thalamus) → primary somatosensory cortex Spinocerebellar pathway (unconscious proprioception): Muscle → Spinal cord → cerebellum	→ the caudo- ventromedial edge (VPMcvm) of ventral posteromedial thalamic nucleus (VPM) → dorsal part of granular insular cortex rostroventrally adjacent to the rostral most part of the secondary somatosensory cortex (dGlrvs2) Proprioceptive signals → thalamus → cerebral cortex	stimulate the receptors and check the response.	differences within the right putamen [49]-not done in orbit	(2) Need to look at the effects of combined stressors
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- 12. Thesiometry testing— withdrawal responses
- 13. Coupling a Y maze in dark and add tape for tactile responses.
- 14. Barrel reception system
- 15. Whisker test coupled with NOR

Fine motor control	<ul style="list-style-type: none"> 1. Peg board; 2. Fine motor test (Holden iPad); 3. String/rope pull 4. Precision grip post-flight (JL) 	<ul style="list-style-type: none"> 1. String pull; 2. Spaghetti eating; 3. Lever manipulation 	<p>Animal model tests should be developed:</p> <ul style="list-style-type: none"> a. Peg board 	<p>Cerebellum, basal ganglia, thalamus, rubrospinal, sensorimotor cortex, prefrontal cortex, frontal lobe</p>	<p>Vestibular/Visual input → Brainstem, cerebellum, thalamus → Temporoparietal cortex (vestibular cortex and posterior parietal cortex) → S1 (Primary sensory cortex) → M1</p>	<p>Visual/Olfactory input → Sensorimotor cortex → Corticospinal tract (Motor and Sensory) → Cervical spinal cord → Sensory and Motor</p>	<p>Isometric pinch grip force between the thumb and index finger [51]</p>	<p>(1) Proprioception can be connected to the fine motor control. (2) Animals have fine motor control, but</p>
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				(Primary motor cortex) → Lateral corticospinal tract → Spinal cord → Muscle [42]	neurons → Muscle [50]	we need to standardize and develop a model
Perception	<p>1. Depth— Egocentric distance</p> <p>2. Motion illusions— Verbal reports of illusions when changing modules or looking outside</p> <p>3. Time— Duration estimates</p>	<p>1. Shape— Novel object recognition</p> <p>2. Depth— Cognitive Flexibility</p> <p>3. Time— Navigation and Foraging</p> <p>4. Visual— Food protection behavior</p>	<p><u>Time perception:</u></p> <p>Frontal cortex, basal ganglia, parietal cortex, cerebellum, and hippocampus, lateral and medial entorhinal cortex [52]</p>	<p><u>Dorsal stream pathway (where):</u></p> <p>Retina → Visual cortex (V1, V3) → Middle temporal area (V3A/MT/V5) and Medial superior temporal area → Intra-parietal area → Parieto-occipital area (PO/V6)</p>		<p>Structural changes in somatosensory cortex, Perception as a biomarker? —has many confounding factors</p> <p>(1) Adaptation following flight + return?</p> <p>(2) Some disagreement regarding the relevance of perception in performing operationally relevant tasks</p>
				<p><u>Ventral stream pathway (what):</u></p> <p>Retina → Visual cortex (V1) → Visual cortex (V2) → Visual cortex (V4) → Inferior-temporal cortex → Fusiform gyrus (Fusiform face area)</p>		

				and occipital face area) [53]				
Pain	<p>(1) Back pain (2) Skin sensitivity (3) Pain modulation while modulating vestibular sensitivity (4) Joint pain</p>	<p>Crew after one-year long duration mission had significant skin sensitivity for prolonged periods</p>	<p>Thalamus, Primary somatosensory cortex</p>	<p>Pain or Nociception Pathway: <u>Ascending:</u> Nociceptors in Skin → Spinal cord → medulla → midbrain → Thalamus → Primary somatosensory cortex. <u>Descending:</u> Amygdala → Hypothalamus → PAG → rostral ventromedial medulla → spinal cord → nociceptor [54,55]</p>	<p><u>Ascending pain pathway:</u> Nociception receptors → spinal cord dorsal horn → parabrachial nucleus (brain stem) → thalamus and amygdala → somatosensory cortex/prefrontal cortex/anterior cingulate [56]</p>	<p>Bilateral lesion in mPFC [57]</p>	<p>Blood: MFAP3, GNG7, CNTN1, LY9, CCDC144B, and GBP1 [58], sICAM-1 [59], fMRI based brain imaging [60], Autonomic nervous system markers: Pupil reflexes, Electrodermal activity, Peripheral pulsatile component of cardiac cycle, Heart rate, Blood</p>	<p>(1) Need to focus on peripheral nervous system and include and utilize blood markers. (2) Individual pain tolerance is variable</p>

								pressure [61]. Blood markers, miRNA markers, inflammatory factors and CCR2 receptor, Pain as biomarker (many confounders)
Smell and taste	1. University of Pennsylvania Smell/Taste identification Test in animals—odor is very important, social interactions, fear conditioning,	1. University of Pennsylvania Smell/Taste identification Test in animals—odor is very important, social interactions, fear conditioning, Smell and Taste has been hypothesized to be modified secondary to fluid shifts causing increase in salt and spice intake leading to dysregulation of body salt composition	Gustatory and olfactory cortex, Piriform cortex and homology to hippocampus . Olfactory epithelial, like hippocampus , has continual neurogenesis	<u>Gustatory pathway:</u> Tongue → solitary nucleus (medulla) → thalamic nucleus (ventral posterior medial nucleus) → gustatory cortex → hippocampus (identification) Olfactory receptors → olfactory bulb → olfactory cortex →	<u>Olfactory pathway:</u> input → olfactory sensory neurons in olfactory epithelium → olfactory bulb → hippocampus → amygdala → learning/behavioral input [62] Smell and hippocampal	Olfactory bulb volume [63]	<u>Nasal mucus</u> (1) Loss of (smell): Sonic hedgehog levels [64]; <u>Saliva</u> (taste)—Sonic hedgehog [65] <u>Blood</u> —miRNA panel	(1) Loss of smell impacts social interaction and can lead to depression. Loss of smell in long term missions can contribute to depression. (2) Smell can also have a

memory sequences of odor.

hippocampus (odor memory) Olfactory receptors → olfactory bulb → olfactory cortex → thalamus → orbitofrontal cortex (conscious perception of smell) circuits are similar ----> can be used to assess broader cognitive dysfunction

including mitochondrial stress markers. Smell test: Scratch and sniff test. Smell as a biomarker. downstream effect. Onset of smell precedes for many years in AD patients. (3) What about systemic response associated with smell deficits; can we have blood biomarkers for it? Mitochondria l functions are associated with olfactory pathways— can we test mitochondria ? can we identify miRNAs

									associated with olfactory issues?
									(1) Need to study combinatorial stressors (2) Effects of microgravity on hearing/auditory. (3) Largely ignored—as most of behavior test do not rely on hearing ability
Hearing	1. Otoacoustic emission 2. Auditory evoked potential analysis	1. Otoacoustic emission 2. Auditory evoked potential analysis	Test in higher animals: NHP	Auditory cortex	<u>Auditory pathway:</u> Ear → cochlea → cochlear nucleus (medulla) → superior olive (medulla) → inferior colliculus (midbrain) → medial geniculate (thalamus) → auditory cortex Lemniscal auditory pathway, olivo-cochlear system	<u>Ascending auditory pathway:</u> Ear → Cochlea → Cochlear nucleus → superior olive → inferior colliculus → medial geniculate nucleus (dorsal thalamic nucleus) → auditory cortex [66]		<u>Blood:</u> Prestin [67,68], Low frequency hearing loss	

3.2. Behavioral Medicine Influences on Operational Performance (Leads: C. Davis, David Dinges)

The goal of Group 2 was to create lists of biomarkers and brain regions and/or neural circuits that are related to operational performance for constructs that are prioritized in the HRP's Behavioral Medicine (BMed) risk. Group 2 assessed the following key constructs which are summarized below and in Table 2: memory, attention and dual tasking, executive function, working memory, learning and plasticity, social processes, individual behavioral states, arousal and regulatory, emotional regulation, risk taking/tolerance, and stress.

3.2.1. Summary of Discussions

Many of the themes that arose during this panel's discussion were also discussed by the sensorimotor group (Group 1), including learning and plasticity for assessing an astronaut's general level of adaptability. The panel also discussed the importance of studying individual differences in these different behaviors, in addition to various modifying factors, such as sex, age, the impact of stress, and immune status. The panel also highlighted the importance of general biomarkers that are not specific to any construct, behavior, or tissue, but could provide a more accurate reflection of overall behavioral health.

Behavior is a biomarker. One major theme that emerged from the discussion was the fact that behavior is an important biomarker. Although biomarkers and brain regions and neural circuits are important for understanding the biological basis of changes in operational performance, the behavior itself needs to be studied as an indicator of changes in operational performance. Variations in behavior, such as increases in variability of response and instability in performance, are often the most sensitive indicators of degradation of operational performance [69,70]. Furthermore, marked inter-individual differences exist in these domains, some of which appear to be phenotypic [70,71]. However, limited knowledge exists regarding the biological basis of these individual differences and how they are modulated by spaceflight stressors. For several constructs, the panel noted specific behavioral changes that should be considered as biomarkers and gave examples of potential neuroimaging modalities that could be used to investigate underlying brain regions and neural circuits. More studies of human behavior in spaceflight are needed. Behavioral tests with greater ethological relevance to animal models would most likely yield better translation of findings to human operational performance. The panel discussed similarities between attention tasks and dual tasking; performance instability, increases in the variability of responding, and increased impulsivity are all behavior markers indicating a problem [70,72,73]. These changes can be subtle, which highlights the importance of knowing the organism's baseline performance for a task, so that changes to that baseline will then indicate a problem. Finally, behavioral biomarkers can be used to determine when an organism—from rodents to humans—is unable to use new information in the environment to adapt their behavior; these results have been obtained primarily from reversal learning and extinction tasks that are highlighted under General Brain Plasticity below.

Common measurements for studying brain biomarkers. Various neuroimaging modalities were discussed for most of the constructs, and because the panel focused on measures that could be assessed during spaceflight and across species, electroencephalogram (EEG) and event-related potentials were regarded as valuable for identifying markers associated with several constructs, including memory, working memory, attention, dual tasking, and learning and plasticity. The use of whole-brain and region-specific EEGs were both considered useful, with whole-brain EEG being particularly important for learning and plasticity [74,75]. Region-specific EEGs were regarded as most useful when coupled with a behavioral task dependent on that region, such as frontal cortex activity and attention or performance on an adaptive N-back test to assess working memory.

Near-infrared spectroscopy (NIRS) and functional NIRS were also regarded as useful for assessing underlying neural targets during task performance during spaceflight.

Magnetic resonance electroencephalography and other frameworks for integrating multiple imaging modalities should also be investigated, such as joint imaging markers from simultaneous magnetic resonance imaging (MRI) and EEG (e.g., temporal volume, cortical thickness) that are associated with cognitive status in healthy individuals, pathophysiological changes in neurodegenerative diseases, and after traumatic brain injury [76–81]. The panel contended that these simultaneous recordings could provide a more accurate diagnosis of pathology than either modality alone.

Overlapping markers among constructs. The panel agreed that many biomarkers overlap among the constructs, such as the gastrointestinal (GI) microbiome, immune markers, and the influence of steroid hormones. As such, these markers could be general markers of behavioral health. For translational studies, most of these markers can be measured in animal models and have supporting preclinical evidence to demonstrate their relevance to human CNS function and disease.

- Immune markers. Several accessible biomarkers are common to various constructs, including inflammatory markers such as Tumor Necrosis Factor alpha (TNF-alpha), Interleukin 6 (IL-6), and Interleukin 8 (IL-8).
- Oxidative stress markers. The panel considered transthyretin (TTR) as a biomarker of neuronal stress that could be useful for assessing general CNS health, irrespective of a specific BMed construct. Although TTR is possibly inaccessible for spaceflight (e.g., choroid plexus TTR, lumbar puncture for cerebrospinal fluid), recent work suggests serum levels could be indicative of CNS pathology [82].
- Microbiome. The GI microbiome is connected to the brain through the gut–brain axis and the panel regarded this as an important system to assess potential biomarkers indicative of CNS pathology. Recent research demonstrates a vital role of the GI microbiome in CNS pathology and psychiatric disorders [83–85] and the microbiome has important implications for health during long-duration spaceflight [86,87].

Incorporate modifying factors into biomarker studies. The panel discussed additional factors important for spaceflight, and differences in many of the BMed constructs that were not included on the worksheet, such as sex, age, stress, immune status, steroid hormone levels, and prior experiences. The panel noted that any findings regarding the usefulness of the various biomarkers should also include tests of these biomarkers under these additional conditions to determine if the markers were relevant when these other factors are included. For example, a biomarker might be useful for males, but not females, or the menstrual cycle phase could impact the usefulness of the biomarker in females. Studying biomarkers under combined spaceflight factors in analog environments [88] was also viewed as being important to determine the usefulness of these biomarkers, given that individuals might respond differently to various spaceflight factors.

Default mode network (DMN). The panel discussed the importance of the DMN in both normal and pathophysiological processes as it relates to several of the BMed constructs, and they considered DMN to be a marker that might overlap among constructs (e.g., changes in DMN could indicate memory and attention problems, in addition to sensorimotor changes). The DMN is a brain system that is preferentially activated when the brain is at wakeful rest [89,90]. Core regions of the DMN include the medial prefrontal cortex, posterior cingulate cortex, and parts of the precuneus, as well as the hippocampus, retrosplenial cortex, and angular gyrus [91]. Changes in activation of the DMN have been associated with several psychiatric conditions, including post-traumatic stress disorder, Alzheimer’s disease, autism, depression, and chronic pain [92–96]. DMN activation can be modulated by different interventions and physiological processes, including physical activity and exercise, sleeping, resting wakefulness, sleep deprivation [97–99], and age [100]. The panel regarded the DMN as an important biomarker of brain function, and given its relationship to other cognitive functions (e.g., attention), they thought it could be

useful for understanding changes in operational performance. Because the DMN could be an important marker associated with multiple constructs (e.g., memory, working memory), the panel suggested it could also be an important marker for integration of these constructs and/or how modifying factors influence these constructs (e.g., sleep/wake and sleep deprivation). The DMN seems to be essential to the social understanding of others and could provide a biomarker for spaceflight-associated changes in social cognition and behavior.

3.2.2. Recommendations

The panel evaluated each specific construct to determine if a good biomarker exists that is operationally relevant for astronauts and that translates from animal models. The panel also commented on gaps in each construct that would need to be filled to produce an effective biomarker.

1. Attention. The panel identified several important behavioral markers from attention tests, primarily the psychomotor vigilance test, including increased variability in responses, decreased psychomotor speed, impulsivity, instability in performance, and lapses of attention. Several of these performance measures have been studied on the International Space Station (ISS) and in various analogs of the spaceflight environment [101,102].

2. Dual tasking. This construct overlaps BMed and sensorimotor effects and demonstrates the interconnectedness of numerous constructs relevant to operational performance. Furthermore, dual tasking is argued to be a useful behavioral method for assessing changes in cognitive reserve [103–105] during spaceflight and after g-transitions after landing [72,73]. Dual tasking measurements during long-duration spaceflight have identified long-term deficits in visuomotor performance and that cognitive reserve is reduced, possibly due to continued sensorimotor adaptation and stress [72]. Dual tasking measures could be useful behavioral biomarkers of how individuals adapt to the spaceflight environment.

3. Procedural memory. This form of memory [106] was not specifically identified in the two different memory constructs, but the panel felt that it is essential for operational performance and should be mentioned as a subheading under the memory construct.

4. General brain plasticity as an important biomarker of adaptability or lack of adaptability. Operational performance requires a brain that can adapt to stressors under various spaceflight conditions. As such, alterations in brain “adaptability” could be a useful biomarker indicating degradation in operational performance [107]. For example, simple adaptation to repetitive stimuli or general adaptation across multiple tasks (not only task-specific changes) might indicate how the nervous system is faring in a space-like environment (i.e., whether the brain is able to adapt to this new environment, and whether this adaptability is changing over time). This construct is important because it integrates across all measures, can be translated between rodents and humans, and clinical markers of brain damage exist that could be useful biomarkers (e.g., blood brain-derived neurotrophic factor [88]). In addition, learning and plasticity are constructs that have been tested in animal models relevant to astronaut performance (e.g., reversal learning, extinction learning), including after space radiation exposure [108,109].

5. Reversal learning is used extensively in animal models to assess cognitive flexibility and translates well between rodents and humans [110,111]. The panel suggested that reversal learning under stress or under multiple spaceflight stressors could be paired with neuroimaging (e.g., EEG) to identify factors that impair brain adaptability, and to allow translation from rodents to humans.

6. Although social processes were listed as a standalone construct, the panel noted that social interactions are important for the other constructs, and can be affected by the way individuals interact, the way the crew interacts, and how they perceive the interactions of others or the emotional states of others. This is not trivial and is not necessarily easy to assess, but it is integrated into all other constructs. These interactions highlight the

need to consider how these individual states impact the group, and the need to determine if there are biomarkers of these interactions, and/or if those interactions then change the individual biomarkers.

7. Inclusion of additional constructs. When the panel took a broad view of the worksheet, they concluded that additional constructs should be added. Although many of these additional constructs were embodied within some of the other constructs, the panel thought they should be discussed as discrete constructs and how they affect operational performance.

Emotion regulation. This includes dysregulation that is subclinical, but not psychiatric disorders such as depression or anxiety, because those are included in the individual behavioral states construct.

Executive function. Assays to measure executive function were included in the attention construct, but executive function, irrespective of attention, is important to operational performance.

Risk taking/tolerance. The Balloon Analog Risk task is included within the astronauts' Cognition Test Battery test, and the panel thought that risk taking/tolerance should be a discrete construct and not embedded within another construct. Risk taking/tolerance is also important for social interactions and group dynamics [112,113] and should be examined in animal models under different spaceflight stressors.

Stress. For example, astronauts' self-reported stress ratings increased during 6-month ISS missions [102] and these changes could have important implications for the usefulness of biomarkers throughout the mission.

The panel identified the following gaps in knowledge:

Lack of integrated approach. The panel noted several gaps that could be addressed by first taking an integrated approach to these different constructs. For example, sleep loss or stress will most likely affect all constructs on the list. The constructs are intertwined, and many things can affect them, and for this reason, our group suggested the use of more general biomarkers, instead of construct-specific biomarkers; for example, a "general health" biomarker or a "vulnerability" biomarker that would indicate an individual's status on some continuum of functioning within the spaceflight environment. What remains unknown is whether the biomarkers that have been identified are informative under all conditions, or if these markers will change as external stressors and internal conditions change.

Importance of stress. The panel noted several modifying factors, but stress emerged as a critical factor that probably deserves its own category on the worksheet.

Lack of sex differences or inclusion of sex. Sex needs to be considered throughout all the constructs. It was not included in any construct and could have important implications for determining what biomarkers are relevant and useful.

Inclusion of microbiome. This appears to be important to brain function, and as such, could affect the majority of the BMed constructs. A better understanding of the specific bacteria, dysbiosis, etc., and how they relate to cognition and the different performance constructs, would be useful for biomarker development.

Lack of measurements for individual differences. The panel noted the importance of inter-individual differences for these constructs and their likelihood of affecting operational performance. All individuals can be trained with the same techniques, but it is not known, nor can we currently predict, how each individual will continue to perform in the spaceflight environment. This is especially true when hazards such as radiation exposure and isolation are combined. Methods are required to measure these differences and to understand how they might impact operational performance.

Additional gaps. These include the need for better technology to quantify biomarkers during spaceflight, and greater understanding of the differences between diurnal humans and nocturnal animal models (e.g., rodents) and how this influences the biomarkers we identify and study.

Table 2. Circuits and biomarkers for behavioral medicine domains.

Key Indicator/Construct	Human Performance Test	Animal Performance Test	Caveats/Notes/Related Functional Performance Tasks/Prediction of Behavioral Outcome in Humans	Brain Region	Human/NHP Neural Circuit/Pathways	Rodent Neural Circuit/Pathway	Biomarkers (Rodents/Humans/NHPs)		Gaps/Notes
							Inaccessible	Accessible (Translatable to Astronauts)	
Memory	Mnemonic similarity test (MST) (BPSO)- this test includes Novel object recognition (NOR)	1. Object in place	- Needed for recall of training, what you did minutes, hours, days ago	Hippocampus and associated regions	<u>Excitatory trisynaptic circuit</u> Direct memory formation: Entorhinal cortex → Dentate gyrus → CA3 → CA1 → Entorhinal cortex V	Excitatory trisynaptic circuit	CSF: APOE, amyloid. Hippocampus: decreased BDNF, increased GFAP, inflammatory marker, synaptic marker, Arc	Imaging -CT, fMRI, PET, EEG, MEG, TMS scan for Default mode network activity, negative amplitude, hippocampal sharp wave ripples (rodents), no contrast for glymphatic system. Blood: APOE, TREM levels, d-cycloserine, neurofilament	(1) Study effects of Stress, immune system? (2) Study the effect of Combined stressors? (3) Sex Differences? (4) Resource constraints for spaceflight mission— development of readily
		2. Social Recognition	- Age-related cognitive decline; mild cognitive decline (MCI); neurodegenerative conditions and dementia						

								light chain, BBB breakdown. Behavior - fMRI, EEG and ERPs with behavioral test and stressor. GI microbiome. NIRS/fNIRS	accessible and implement able technology for biomarker quantification (5) Ethologically relevant animal tests that are relevant to human performance tests
Attention and dual tasking	1. Reaction time- PVT 2. Dual Task Test (e.g., cognitive-motor, divided attention): a. PVT b. Walking with distractors 3. Odd-ball stimulus	1. PVT 2. Attention set-shifting: 3. 5C-CPT 5 choice performance test (selective attention)	<ul style="list-style-type: none"> •Used operationally as go/no-go test; operational activities requiring high skill might get most affected; •PVT should be considered for performance under pressure with distractions 	Prefrontal cortex (lateral PFC) and anterior cingulate cortex	Selective attention: Visual cortex → Lateral intraparietal cortex or Middle intraparietal sulcus → prefrontal cortex [116,117]	sustained attention (PVT/CPT): pedunclopontine tegmental nucleus (PPTN) → substantia nigra pars compacta (SNc) → striatum and PFC → motor control (cholinergic output) [117]	Catecholamine— Noradrenaline, dopamine, mAChR and nAChR	Imaging: fMRI, PET, EEG scan [118,119], EEG of frontal cortex with behavioral task, pupil diameter, NIRS/fNIRS; Urine: norepinephrine, 3-methoxy-	(1) Correlation between attention, stress, immune dysfunction, and sleep. (2) Predictive validity of operationa

4- l
hydroxypheny performan
lglycol; ce in
Plasma: astronauts
monoamine —No data
oxidase, on that.
neuropeptide Also need
Y [120], Zinc, rodent and
ferritin; Saliva: human
cortisol, analogs.
Genetic and (3) Access
behavioral to
biomarkers, operationa
inflammation l task data
related and self-
systemic monitoring
markers. data
Behavioral (4)
markers— Wearable
Increase in devices for
variability of continuous
response, monitoring
impulsivity, of heart
instability in rate,
performance, sleep/wake
attention cycles, rest
lapses, dual activity
tasking (motor and other
control + autonomic
primary task). activities
ECG heart rate without
measurement, disrupting

autonomic measurements, and rest activity cycles with task performance GI microbiome; polysomnography (in sleep) and skin conductance/EDA
 other crew activities/adding crew time. (5) Continuous and close tracking of crew behavior. (6) Note the bias towards response and response strategy of an individual and its dependency towards individual's motivation.

Working Memory	1. Fractal 2 back 2. Object rotation in space 3. Spatial WM	1. Radial arm water maze-trials to criterion, latency is common	- Docking: Egress procedures and EVA-related;— Crew should stop with plans	Fronto-parietal brain regions, including the	Prefrontal cortex --> Visual component	PFC-hippocampus (dorsal)— visual component	Rodents-microglia activation in prefrontal cortex and	<u>Imaging:</u> CT, fMRI, PET, EEG, MEG, TMS scan for default mode network,	(1) Cross-cutting issue with immune markers?
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across studies, for can be completion/perf modified for ormance of task each with possible individual catastrophic animal, can be consequences if modified for not performed test-rests 2. correctly— modified Anxiolytic Barnes maze effects— Anti- (operant n- depressive back in effects— rodents lacks Exploratory stable behavior and baseline) measure of 3. NHP: anxiety in open touchscreen, areas saccades 4. Elevated plus maze and elevated zero maze 5 Forced swim test 6. Light-dark box without elevation 7. Tail suspension test 8. Puzzle box paradigm— adaptive	prefrontal, cingulate, and parietal cortices and mediodors al thalamus (rodent, [121]) Exploratory behavior and measure of anxiety in open areas	hippocamp Neuroimaging (2) us, Afg3l1, with adaptive Integrative Tpx2, N-back task, approach Neuroigin dopaminergic -3, RB1- system, whole inducible brain or coiled-coil targeted 1, Mast3, frontal, Kif21a, parietal, and DnaJ striatal region (Hsp40) <u>Blood</u> : cortisol homolog, levels, SLIT- immune ROBO Rho cytokine - GTPase- chemokine activating levels (TNFa, protein 2, IL8, IL-1ra, Rasgrf1 Tpo, VEGF, [20] CCL2, CCL4, and CXCL5) [122]. <u>Salivary</u> : immune markers. <u>Eye</u> : blink rate for indicator of dopamine sensitivity. GI microbiome, NIRS/fNIRS
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Walking with visuomotor adaptation (b) Split Belt Locomotion Test
 3. Whole body tasks (a) Ladder rung walk test
 4. Mismatch negativity (plasticity + perceptual learning, non-motor component, EEG measure)
 5. Barnes maze
 6. Extinction learning (Fear extinction).
 7. Reversal learning (under stress)
 8. Delayed matching to position (DMP)
 9. Radial arm maze

Social Processes (e.g., Socialization)	Socialization: Self-report survey, sociometric badge	<u>Socialization:</u> 1. Social fear 2. Social approach to a stranger mouse	Prefrontal cortex, Amygdala, Hypothalamus, striatum	Aggression: Sensory reception → Prefrontal Cortex → Amygdala → Hypothalamus → Periaqueductal grey (midbrain)/Ventral	Social attachment: Olfactory cues → Vomeronasal organ (VNO)/Main olfactory epithelium (MOE) → Accessory olfactory bulb (AOB) → Amygdala → Lateral	TRPc ko mice (loss of aggression) [126], reduced/lo	CT, fMRI, PET, EEG scan Blood-Vasopressin and oxytocin levels, 5-HT,	(1) Learning effects and sex difference
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<p>conflict, communication, bonding)</p>	<p><u>Conflict:</u> Self-report survey, journal analysis, observational ratings <u>Communication:</u> Self-report survey, communication recording analysis, observational ratings <u>Bonding:</u> Observational ratings</p>	<p>3. Reciprocal social interactions 4. Conditioned place preference to conspecifics 5. Social recognition 6. Juvenile play 7. Nesting patterns in home cage <u>Conflict (Aggression)</u> 1. Social Defeat 2. Resident intruder attack 3. Routine observation 4. Isolation-induced fighting 5. Tube test for social dominance <u>Communication</u> 1. Ultrasonic Vocalizations</p>	<p>Tegmental area → Aggressive behavior [124]</p>	<p>Septum → mPFC → Nucleus accumbens Olfactory cues → VNO/MOE → AOB → Amygdala (2) Social stimuli → mPFC → Nucleus accumbens/Hypothalamus/Amygdala/Ventral tegmental area./Dorsal raphe nucleus/hippocampus Aggression: Olfactory cues → VNO/MOE → AOB/Main olfactory bulb (MOB) → Amygdala → Hypothalamus/Bed nucleus of the stria terminalis (BNST)/Hippocampus (Hippocampus → Lateral Septum) [125]</p>	<p>ss of nNOS (increased aggression and reduced social investigation) [127], Neurologin-3, PSD95, parvalbumin, bone hormone-osteocalcin. Radiation studies in brain—CCL2, CD206, CD163, PSD-95 in PFC, Dopamine receptor levels</p>	<p>nNOS (male mice), testosterone (social regulation), cortisol, progesterone, cortisol to testosterone ratio, cortisol to oxytocin ratio. Imaging-Striatum and reward related brain regions. Psycho variables—heart rate, skin sensitivity. GI microbiome; NIRS/fNIRS; polysomnography (in sleep) and skin conductance/E DA. Behavior-eye gaze and eye tracking</p>	<p>(2) Behavior of one animal/astronaut would affect others behavior (3) Understanding the dynamic social interaction between the crew members, psychological ownership of the space, habitat size to social interaction and any areas that need mitigation.</p>
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emitted
during social
interactions
2. Response to
vocalizations
form
conspecifics
3. Deposition
of social
olfactory
pheromones
Bonding
1. Pair
Bonding
2.
Observation,
Grooming,
Inter/Intra-
Social
Interactions
3.
Oxytocin/Vaso
pressin levels
Social
Hierarchy
1. Hierarchal
testing/Social
stability
measurements
—convergent
testing like
tube testing

		2. Urine marking (sex should be considered)							
		3. Hotspot testing							
Individual Behavioral States (e.g., Stress, Depression, Mood and Anxiety)	Stress: Visual Analog Scale	<u>Stress</u> 1.					<u>Choroidal plexus:</u> TTR (independent of radiation exposure). CSF: Glutamate, GABA, Acetylcholine,	fMRI scan <u>Blood:</u> Glutamate, GABA, Norepinephrine, Dopamine, Serotonin, Vasopressin, Orexin, cortisol, corticosterone, <u>Immune markers:</u> IL6, B-cells, Cortisol, TNFa, IL4, IL5, IL-10 [122,129,130], CSF – TTR (lumbar puncture) <u>Saliva:</u> Cortisol; NIRS/fNIRS.	(1) How individual behavioral state will impact the others in the group (cohesion, behavioral state of the group). This relates to where the crew is in the craft and who interacts with whom, crew member who isolates themselves can be a
	Depression: Beck Depression Inventory Mood: Profile of mood states-short form, Zung self-rating depression scale, Hamilton Rating Scale for Anxiety, Beck Scale for suicide Ideation and Beck Hopelessness Scale, Quality of Life Enjoyment & Satisfaction Questionnaire	<u>Depression</u> 1. Forced swim test 2. Inescapable shock 3. Low sucrose preference (Anhedonia) 4. Tail suspension 5. Social defeat 6. Leaned helplessness 7. Novelty-Suppressed Feeding <u>Mood</u> 1. High elevated plus maze 2. High changing	Prefrontal cortex (PFC), subgenual cingulate cortex (Cg25), subcortical hippocampus, nucleus accumbens, amygdala, ventral tegmental area	<u>5HTergic/NEergic Depression pathway:</u> Locus coeruleus/Dorsal raphe → Amygdala/Hippocampus/Ventral tegmental area/Nucleus accumbens → Prefrontal cortex [128]	<u>5HTergic/NEergic Depression pathway:</u> Locus coeruleus/Dorsal raphe → Amygdala/Hippocampus/Ventral tegmental area/Nucleus accumbens → Prefrontal cortex [128]				

, reinforcement
 Psychological schedules
 General Well-Being Index, 3. High open field
 Pittsburgh Sleep Quality Index Anxiety
 Risk 1. Light-dark exploration
 Tolerance: 2. Vogel conflict test
 balloon 3. Marble buying
 analog task 4.
 Unpredictable chronic mild stress
Risk Tolerance
 1. Elevated plus maze (head dips),
 2. delayed reward task (impulsivity),
 3. Rat gambling task.
 4. Predator odor risk taking test

exposure),
 glial and
 synaptic
 dysfunction

behavior
 issue to be
 detected
 and dealt
 with.

Arousal and Regulatory (e.g., sleep,	Sleep duration and Architecture: Actigraphy	<u>Sleep duration and Architecture:</u> Actigraphy,	<u>Sleep duration and Architecture:</u> Actigraphy and	Hypothalamus, Brain stem, Spinal	<u>Sleep:</u> Retina (light) and metabolic inputs (peptidergic hormones, nutrient	<u>Circadian rhythm:</u> Retina → Retinohypothalamic tract → Suprachiasmatic nucleus → Paraventricular nucleus →	Brain Melatonin levels (not accurate	CT, fMRI, PET, EEG, polysomnography scan 6-	(1) Sex differences (2) Associatio
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circadian phase)	and EEG 1. PVT 2. Visual analog scale towards alertness— assessing sleep quality Circadian phase: Actigraphy (not good biomarker)	Sleep Island, EEG <u>Circadian phase:</u> Actigraphy (not a good biomarker)	EEG, PVT, sleep quality <u>Circadian phase:</u> Actigraphy (not good biomarker)	cord, Suprachias matic nucleus	signals) → Retinohypothalamic tract and Arcuate nucleus → suprachiasmatic nucleus → ventral subparaventricular zone → dorsomedial hypothalamus → ventrolateral preoptic nucleus → sleep <u>Wakefulness:</u> Retina (light) and metabolic inputs (peptidergic hormones, nutrient signals) → Retinohypothalamic tract and Arcuate nucleus → suprachiasmatic nucleus → lateral hypothalamic area (melanocyte concentrating hormone/orexin-producing neurons) → wakefulness [131]	Medial forebrain bundle → Intermediolateral cell column → Superior cervical ganglion → Nervi conarii → Pineal gland (Melanocyte—Melanin secretion) [132]	with rodents) nocturnal animals and light cycle and when the test is conducted (light or dark cycle) Sex difference	sulphatoxymelatonin (aMT6) collected every 2 to 8 h. over 24 to 48 h period, melatonin, Timeless, period 1–3, growth hormone (SOCS) [133] Actiwatch (sleep quality, duration), Urine: 6-sulphatoxymelatonin (good biomarker); Melatonin in blood and saliva (not accurate), core body temperature (susceptible to masking), GI microbiome, genotype changes—per3 polymorphisms (human),	ns between menstrual cycle phase, sleep need and circadian (major gap!) → actually, not only estrogen, but testosterone cycles too, so should consider both! Differences between nocturnal and diurnal species! Most rodents are nocturnal, but most behavioral tests on
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Dqb10602 gene (narcolepsy), Immune markers—IL6; behavioral tests; NIRS/fNIRS	rodents (in general, not sleep specific) are done in light. (4) New technology for measuring fluid shift and shift of brain in the cranial compartments. Tympanic membrane movement measurement (5) Sleep duration, quality, and continuity. Need to ensure that sleep is: not disturbed, adequate
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		<p>for operations, at appropriate circadian time, entrained by light, exercise etc. Sleep quality is an orthogonal component to stress and emotional status. (6) Diet and its contribution (7) Intersubject variability</p>
<p>Emotional regulation</p>	<p>Hippocampus, striatum, PFC</p>	<p>Psychology, subclinical –Facial expression, emotional</p>

regulation.
Regulation
of the
conflict.
Executive
functions.

3.3. Integrated Biomarker and Signaling-Pathway Approaches for Understanding Operational Performance (Leads: X.W. Mao, R. I. Desai)

The goal Group 3 was to use a systems-biology approach to generate lists of biomarkers and signaling pathways related to CNS circuitry and operational performance that will be important to monitor in astronauts during spaceflight and after return to Earth. To achieve this goal, the integrated approaches team (a) reviewed and identified a broad array of biomarkers of important mechanisms known from space research (i.e., what is known); this panel discussed research on biomarkers and signaling pathways in animals and humans that could be used to assess the effects of acute or long-duration exposure to spaceflight stressors on operationally relevant performance; (b) considered knowledge from other CNS-health studies that could be repurposed for assessing astronauts (e.g., aging, disorder, disease); and (c) documented open questions and research gaps in the knowledge base that connect genes and biological pathways to brain regions and neural circuits that link to operational performance (i.e., what is not known, needed experiments). Discussions are summarized below and in Tables 3 and 4. The goal of this integrated approaches team was to provide recommendations regarding the availability, validity, and limitations of biomarkers and signaling pathways to be examined in future research.

3.3.1. Summary of Discussions

It should be emphasized at the outset that the results of this integrated approaches exercise did not reveal any biomarker (or combination thereof) that was uniformly responsive across different regions of the brain to a single or given combination of spaceflight stressors. The panel raised the following distinct, yet overlapping questions:

1. Does the literature provide any useful insight regarding if or how combined exposure to spaceflight stressors might interact to alter (additive, synergize, diminish) biomarkers and signaling pathways involved in CNS function?
2. What experiments need to be performed to inform how these combined stressors interact and affect biomarkers and signaling pathways associated with CNS function?
3. What are the challenges that need to be addressed for data collection and storage?
4. What information do we need for successful biomarker repurposing?
5. What new experiments, analysis, and techniques are needed?
6. What information about biomarkers and signaling pathways is needed to identify and implement effective spaceflight countermeasures that will minimize CNS decrements associated with the long-duration spaceflight beyond Earth's protective magnetosphere?

Below is a summary of the key issues that were raised by the integrated approaches panel.

1. First and foremost, all group members recognized the need for standardizing certain aspects of the experimental protocol across laboratories; in particular, standardizing (a) factors related to the degree of exposure to a spaceflight stressor (e.g., space radiation (Galactic Cosmic Radiation simulation), dose, dose rate, and energy; isolation/confinement; altered gravitational levels (Mars, lunar or Earth)); (b) the type of animal models used (e.g., age, sex, strain, species; see below) and the time of tissue collection. This approach will permit meaningful comparisons and interpretations of data from different endpoints collected among investigators.
2. The panel overwhelmingly agreed that a paucity of information exists on how CNS-related neurocognitive performance is affected in laboratory animals that have been exposed to space-relevant radiation (e.g., a low-dose (<0.5Gy)/low-dose-rate of simulated galactic cosmic rays) and that such effects have not yet been systematically studied.

3. Although studies using several species (e.g., rats, mice) have provided important information about how spaceflight stressors may affect behavior and cognitive function, extrapolating data from rodents to humans is an imperfect science. Notably, the translational value of larger size animals (e.g., NHPs) used in various research domains, including neurobiological, neurobehavioral, and complex cognitive processes, has been validated and established over many decades. These successes are based on numerous factors including (1) the considerable overlap in the genetic, physiological, pharmacokinetic, neurobiology, and neurobehavioral effects in NHPs and humans; (2) the proven reliability of NHPs as subjects in long-duration (i.e., longitudinal) neurobehavioral and cognitive studies; and (3) the ability to use powerful within-subject designs that are similar to those used in human studies, which permit meaningful conclusions or inferences by evaluating all treatment effects in individuals as well as in groups. Considerations such as these suggest that NHPs are especially well-suited for ground-based study of the acute and long-term neurobehavioral effects induced by spaceflight stressors, either alone or in combination, and for translating effects to astronauts. Thus, there was considerable appreciation in the group that the use of appropriate animal models, especially targeted studies in NHPs to confirm or advance observations in rodents, should be carefully considered by NASA in future work.
4. The panel recognized that an integrated “omics” profiling strategy using technologies such as genomics, proteomics, and metabolomics is desperately needed to further expand understanding of the underlying brain systems/mechanisms that may be affected by exposure to spaceflight stressors. This multimodal approach will be highly beneficial to determine biomarker datasets of differentially expressed genes, proteins, or metabolomic/lipidomic signatures and the pathways that lead to pathological and possible degenerative changes in the brain. An omics-based molecular phenotyping approach for characterizing biosignatures associated with low-dose space radiation, simulated microgravity, and other space environmental stressors will provide a deeper understanding of the underlying mechanisms responsible for brain structure and pathophysiological changes. This approach will also provide critical information about how individual sensitivity (e.g., genetic, epigenetic, previous injury, age, and sex/gender) will influence how spaceflight stressors affect operational performance. However, as stated above, it will be critical for protocols and metadata from experiments in different laboratories to be standardized and processed on a uniform pipeline.
5. A need was identified for longitudinal studies that provide information about changes within the brain (i.e., acute to chronic). This is especially germane for determining if exposure to spaceflight stressors produces short- or long-term neurobiological (or degenerative) adaptations that affect operationally relevant behavioral and neurocognitive performance. A major complication associated with determining how the brain responds to stress insults is the latency between exposure and the expression of injury (e.g., cell loss or dysfunction). Thus, it is essential that longitudinal studies are conducted to meaningfully quantify the development and progression of the CNS injury response.
6. At present, few studies have examined the combined impact of spaceflight stressors on operational performance and/or associated neurobiological changes in the brain. Thus, it is critical that future studies use ground-based animal models that incorporate stressors that are inherent to the spaceflight environment, i.e., space-like radiation exposure and other spaceflight environment stressors including high pCO₂, fluid shifts, microgravity, environmental constraints, emotional stress, and circadian misalignment/sleep deprivation. This will permit data to be extrapolated more accurately to estimate potential risks encountered by astronauts during deep space missions. Ground-based studies to examine the impact of combined spaceflight

conditions and the underlying mechanism(s) of potential interaction on structural and functional deficits in the brain are very limited.

7. The panel overwhelmingly agreed that significant effort and resources are needed to develop new cutting-edge techniques to identify brain biomarkers that may indicate operationally relevant neurocognitive performance. Novel imaging techniques that provide an early detection of the subtle changes in the brain and identify the target population and biomarkers for intervention are essential. Thus, to improve knowledge about anatomical, physiological, and functional changes to the brain, especially for longitudinal evaluation, an effort is needed to develop advanced computerized tomography scan, functional magnetic resonance imaging (fMRI), positron emission tomography scan, EEG, magnetoencephalography, and transcranial magnetic stimulation scan imaging technologies.

The panel members agreed that a critical need exists to use data better and carefully from flown astronauts to evaluate the actual acute and long-term health risk of the spaceflight environment. Importantly, there was appreciation that human data could be better related to outcomes from animal studies, which may help characterize alterations in circadian rhythm and sleep, immune system, neurotransmitters, neurobiology (i.e., brain structure and function), and vasculature. If used carefully, follow-up analysis of omics, biochemistry, imaging, and a battery of behavior and neurocognitive testing will provide critical human data that may be used to evaluate the actual acute and long-term health risk of the space environment.

3.3.2. Recommendations

Table 3 highlights the major observations and points of discussion that were addressed by the integrated approaches panel. Although it is likely that exposure to combined spaceflight stressors will alter a wide range of biomarkers in different endpoints in animals and humans, ultimately, it is critical that these biomarkers are consistently and reliably linked with changes in operationally relevant behavior and neurocognitive performance. Evidence so far suggests that specific neurocognitive impairments may manifest under evolving mission scenarios (i.e., increased cognitive load) and, therefore, assessing the impact of spaceflight hazards on a wide range of operationally relevant behavioral and neurocognitive tasks is critical. Moreover, the panel suggested that NASA should explore both novel and trained paradigms with increased difficulty of determining the level of impairment. Finally, to promote translation between animal models and humans, parallel behavioral and neurocognitive testing paradigms exist between rodents ↔ NHPs ↔ humans that should be further exploited.

Table 3. The major observations and points discussed by the panel.

<u>Oxidative Stress</u>	<u>Neurotransmitters</u>
<p><u>Blood biomarkers:</u> 8-oxo-dG in immune cells, MDA, f2-isoprostane, Nitrotryosine; brain HNE, glutathione, lipid peroxidation, ROS, NFKb, MAPK activation, Xanthine oxidase</p> <ul style="list-style-type: none"> • Oxidative stress-associated mitochondrial dysfunction has been shown in many cells, tissue and organ system, their impacts have to be further investigated. • The role of diet in mitigating oxidative stress associated with spaceflight. • Epigenetic clock measurements in astronauts and related to time in space or deep space and 	<p><u>Behavioral biomarkers:</u> mood, depression, anxiety tests</p> <ul style="list-style-type: none"> • Limited in vitro data that are inconsistent across studies. Only one neurotransmitter examined at a time (e.g., DA, glutamate, 5-HT, or ACh). • Human studies with MRI spectroscopy are difficult to do in real-time. • Only invasive rodent assays are available. • Need studies that associate neurotransmitter changes with changes in lipids/metabolites. • Neurotransmitters provide a direct readout of CNS functionality at multiple levels: behavioral, emotional, systemic stress, endocrine, and electrophysiological.

their association with oxidative stress-induced aging.

- miRNA signatures and exosomes in identifying oxidative stress biomarkers and as novel biomarkers in brain pathogenesis.

- Cross-species correlates (chemical changes): rodents–NHP–Humans and should be translated to lipidomic and metabolomic findings.

Neuroinflammation

Blood biomarkers: COX-2, TREM, IL-4, TNF, BDNF, corticosterone; YKL-40, ICAM-1, VCAM-1, IL-15, and Flt-1 in CSF; **Behavioral biomarkers:** cognitive tests

- Specificity of blood biomarkers such as cytokines (variability with circadian changes and time of collection).
- Animal to human correlation (circadian and sleep system differences, rhythm differences, immune differences, white-matter differences, vasculature differences).
- Applying cell-free DNA and subsequent methylation analysis can give high sensitivity measurement of BBB integrity, cell breakdown and inflammation in the brain.

One-carbon metabolism

Blood biomarkers: folate, Vit. B-12, methylmalonic acid and homocysteine, MMPs; CSF: 5MTHF

- Difficult to correlate biomarker changes between CSF and plasma
- Genetic variations in folate-mediated one carbon metabolism predict risk of adverse effects in space flight—mechanisms are unknown
- Relation to white matter hyperintensity, inflammation, BBB permeability, inflammation, behavioral changes, e.g., Hyperhomocysteinemia, vascular dementia.

The panel identified the following gaps in knowledge:

- How can data be integrated across many biology scales for CNS endpoints?
- How can system biology approaches with new technologies – organ cultures, organs-on-a-chip made from normal human cells, integrated “omics” (genomics, proteomics, metabolomics) and cutting-edge brain imaging techniques—be used to estimate acute CNS risks to astronauts from space environment?
- How can knowledge of space environment-induced biomarkers/pathways in neuroinflammation, blood–brain barrier function, vasculature, glia activation be integrated towards better understanding of their impact on acute pathophysiological changes in the brain and late neurodegeneration?
- What is the likelihood of increases in the brain susceptibility to later development of neurological disorders as results of observed changes?
- What is the relationship between neurochemical biomarkers and operationally relevant performance?
- What are the temporal and regional differences in neurochemical biomarkers and their influence on operationally relevant performance? What is the right neurochemical balance?
- What CNS neurotransmitter metabolites can be measured peripherally? Can wearable devices/sensors be used instead of blood?
- Is personalized nutrition (i.e., B-vitamin supplementation) a viable SANS countermeasure?
- Do recurring cycles of sleep deprivation affect performance/vestibular/sensorimotor changes, recovery, and biomarkers?
- What is the role of individual susceptibility—genetic, epigenetic, previous injury, age, and sex/gender—in addressing CNS risk?

Information that is lacking includes astronaut data to monitor the level of DNA damage over time; miRNA signatures as neurodegeneration markers for acute/chronic injury; data from integrated phenotypic studies in models; and omics to identify molecular changes at the synaptic level.

Table 4. Circuits and biomarkers for integrated approaches/physiological responses.

Physiological Responses	Related Gene Ontology Terms	Biomarkers (Human/Rodent)		Associated Pathways/Signaling Cascade	CNS Health Risks	Human Behavioral Measure	Rodent/NHP Behavioral Measure	Open Questions/Gaps (How to Close?)	Notes/Limitations on Biomarkers
		Inaccessible	Accessible						
Neuroinflammation	Glial activation, neuron apoptotic process, BBB disruption, endothelial dysfunction, oxidative stress	CSF: YKL-40, ICAM-1, VCAM-1, IL-15, and Flt-1 [134], Brain lysates-CCR2 [135], Brain lysate-proteomics, IHC, IL21. CSF-cytokine (accurate for neuroinflammation)	Blood: COX-2, cytokines, TREM [136], IL4, TNF, BDNF [137], Corticosterone [138], c-reactive protein, IL-6 and TNFa, glial fibrillary acidic protein (GFAP), IL110, IL4 (variability due to circadian disruption or sleep deprivation), IL21	NFKB signaling, Chemokine signaling, TNF signaling, Calcium signaling, Serotonergic synapse, VEGF signaling, Autophagy, oxidative stress	Neurodegenerative disorders, meningitis	Cognition, Mathematical processing (MTH), Running memory continuous performance test (CPT), Delayed matching-to-sample (MTS), Code substitution (CDS)	Spontaneous new home behavior, Elevated plus maze, light/dark box, WMWM and fear conditionin g, contextual fear conditionin g, Morris water maze test, pass avoidance performanc e test, climbing pole test	(1) Longitudinal study of blood biomarker (e.g., cytokines) and correlating with individual’s biological clock (variability across individual of approx. 5 h.), clinical and medical history. (2) Flight deployable ELISA cytokine panel (3) Microfluidics based system	(1) Threshold? (2) Challenges for data collection and storage: (3) Unclear whether plasma will be collected and stored in space, then assessed on Earth, or are we looking for measures that can be done in real time in space? Some of these assays require special equipment and assays. Importance of storage consistency- Plasma biomarkers are very sensitive to processing and storage conditions,

that can be deployed, miniaturized microscope and flow cytometer. (4) For animal to human study correlation— Tissues can be harvested and animal study should be contextual to the question asked. Humanized mouse model—good for immunologica l study. (5) Leverage omics data. (6) Countermeas ure development requires living system.

including type of plastic for tubes, tube size and volume of aliquots. (4) Recommend many small aliquots to maximize potential for number of biomarkers that can be assessed, because freeze-thaw also significant influences measurement. (5) Specificity of blood biomarkers such as cytokines (variability with circadian changes). (6) Animal to human correlation (circadian and sleep system differences, rhythm differences, immune

								(7) Other animal model— Canine, pig, marmoset— reinventing the wheel?	differences, white-matter differences, vasculature differences).
Neurotransmitters	Neurotransmitter release and metabolism, cellular metabolism	Brain lysates:						(1). What is the relationship between brain neurochemistry and behavior?	<u>Limitations:</u> (1) Inconsistent data across studies: one neurotransmitter system examined (e.g., DA, glutamate, or 5-HT): comprehensive assessment needed. (2) Human studies with MRI spectroscopy are difficult to do in real-time. (3) Rodents' assays are invasive measures, lack less invasive techniques (4) Need studies that associate neurotransmitter
		Serotonin [139], Dopamine and dopamine regulating enzyme, COMPT [139,140], Ach [139], Norepinephrine, Epinephrine, Glutamate [141], Glutamate receptors (NMDAR2A/2B) [133], Stress hormones-cortisol, cortisol, oxytocin; Corticotrophin-releasing hormone (CRH);	Blood: Serotonin [139], Dopamine and dopamine regulating enzyme, COMPT [139,140], Ach [139], Norepinephrine, Epinephrine, Glutamate, GABA [141], Glutamate receptors (NMDAR2A/2B) [133], Stress hormones-cortisol Imaging: CT, fMRI, PET, EEG, MEG, TMS scan	Monoamine pathway: mesocorticolimbic; Hypothalamic-pituitary-adrenal (HPA) axis	Mood, Depression, Anxiety, Alzheimer's, schizophrenia, Parkinson's, other degenerative conditions; Social stress (Stress leading to social dominance)	Mnemonic similarity test (MST) (BPSO)- this test includes Novel object recognition (NOR), learning and motor tasks	Thigmotaxis, water maze, elevated maze, open field test, passive avoidance		

									are best measured peripherally? (5) Wearable devices/sensors to measure metabolites instead of blood tests	changes with changes in lipids and other metabolites <u>Strengths:</u> (1) Neurotransmitters provide a direct readout of CNS functionality at multiple levels. (2) Cross-species correlated (chemical changes) rodents—NHP—Humans. Should be translated to lipidomic and metabolomic findings.
One-carbon metabolism	SANS, BBB, endothelial dysfunction, CSF pressure, Bioenergetics	Brain: B-vitamin and 1C metabolite profiles, DNA strand breaks; uracil in genomic DNA and mitochondrial DNA (higher sensitivity)	Blood: serum and RBC, folate, vitamin B12, methylmalonic acid and homocysteine, MMPs, Met, AdoMet (P. Stover), Formate, one-carbon nutrients, and their methylation	Folate and methionine production, Epigenetic methylation, DNA synthesis and repair, Neurotransmitter metabolism,	SANS, Neurodegenerative disorder (AD), neurodevelopment, Depression	Cognition: Standardized Mini-Mental State Examination, simple reaction time (SRT), choice reaction time (CRT), digit vigilance task (DVT)		Cognitive tests (Morris water maze)	Is personalized nutrition (i.e., B-vitamin supplementation) a viable SANS countermeasure?	(1) Correlating biomarker changes between CSF and plasma? (2) Relation to white matter hyperintensity, inflammation, BBB permeability, inflammation, behavioral

			profiling (inputs towards one carbon metabolism pathway). Imaging: OCT for SANS, MRI for WMH; skin autofluorescence for AGE; Ultrasound Elastography (scleral stiffness), OCT angiography CSF: 5MTHF	Trans-sulfuration pathway, Bioenergetic crisis			changes, e.g., Hyperhomocysteinemia, vascular dementia.	
Oxidative stress	Autophagy, inflammation, Lipid peroxidation, Bioenergetics	Tissue: Glutathione, lipid peroxidation, ROS, NFKb, MAPK activation [143], Blood vessel-Xanthine oxidase [144]	Blood/Urine: Cytokines levels, HNE, MDA, f2-isoprostane, Nitrotyrosine levels [145], 8OHdG; reduced/total glutathione, total antioxidant capacity, superoxide dismutase, glutathione peroxidase, advanced glycation end products (AGEs), glycated albumin, 3-nitrotyrosine,	Oxidative phosphorylation, Mitochondrial dysfunction, NFR2-mediated oxidative stress response, Superoxide radicals' degradation, Neuroinflammation, apoptosis, necrosis, neurovascular impairments,	Neurodegenerative disorders, Cardiovascular disorders, affects multiple organs, Anxiety, Depression, Schizophrenia, Metabolic disorders, SANS.	Anxiety and depression related behavioral tests (Visual Analog Scale Depression: Beck Depression Inventory), psychomotor tests (Tandem Walking, Perturbation during walking, navigating obstacle course while walking (e.g., Functional Mobility Test)),	Anxiety related (Elevated plus maze, hole-board and open field tests), Psychomotor tests (Rod walking, wire suspension/wire hanging, plank walking, inclined screen, accelerating	(1) Can diet mitigate oxidative stress associated with space flight? (2) What are the relationships between oxidative stress, immune function during flight? (3) miRNA signatures? Antagonist-countermeasures

			oxidized LDL, miR383 (regulating AQP4), cell-free DNA (genetic and epigenetic changes) <u>Imaging:</u> CT, fMRI, PET, EEG scan, PET with 62Cu-ATSM [146]	Bioenergetic crisis		Cognitive tests (Mnemonic similarity test (MST) (BPSO)- this test includes Novel object recognition (NOR), Fractal 2B, object rotation in space)	rotarod), Cognitive tests (Morris water maze)	re, specificity, applicability? (4) Exosomes?	
Mitochondrial dysfunction								Plasma: Formate (mito one carbon metabolism) biomarker of mitochondrial function.	
Synaptic plasticity/Neurotrophic Factors	Regulation of synaptic plasticity, modulation of chemical synaptic transmission, neurotrophin receptor activity	Brain lysates: BDNF, Neurotrophin-3 [147], synaptophysin [148], CtBP2, Shank1a [29], 14-3-3 proteins (CSF marker of CNS degeneration), EEG markers, BDNF, c-Fos	<u>Imaging:</u> CT, fMRI, PET, EEG, MEG, TMS scan; Plasma: Neurofilament light (NfL), phospho-tau 181 (pTau181), beta-amyloid 40 and 42, BDNF; CSF: NfL, pTau181, beta-amyloid 40 and 42.	Ubiquitin-proteasome, lysophosphatidic acid (LPA), Calcium signaling (PI3K, PLC gamma), MAPK/ERK	Neurodegenerative disorders, schizophrenia	1.Sequence/procedural; 2. Eye-Head/Eye-Head-Hand adaptation tasks— (a) VOR adaptation test (b) Eye-Head-Hand- visuo-motor adaptation task 3. Whole body tasks (a) Walking with visuomotor adaptation	1. Odor sequence learning (non-motor) and Eye Head Hand tasks: (a) Nystagmus and compensati following labyrinthect	(1) Markers of neurodegeneration are missing. Acute and chronic injury can be tracked longitudinally with plasma NfL. (2) Lacks integration of phenotypic studies in models and omics.	Which biomarkers can we repurpose from terrestrial disorders to spaceflight? There have been huge advances in Alzheimer’s and vascular dementia blood-based biomarkers. While associated with aging, these markers can reflect neuronal and vascular

						(b) Split Belt Locomotion Test 4. Mismatch negativity	omy (b) Rodent VOR test 3. Whole body tasks (a) Ladder rung walk test 4. Mismatch negativity (plasticity + perceptual learning, non-motor component, EEG measure) 5. Mathematical processing (MTH)	(3) miRNA signatures are missing. (4) Identify molecular changes at the synaptic level (5) Relatively unexplored area	injury and later risk of cognitive problems. NFL is a marker of neuronal injury that is increased significantly in traumatic brain injury, many forms of dementia, and CTE.
Vestibular/Sensorimotor alterations	Vestibular reflex, vestibular hair cell stereocilium organization, vestibular receptor cell stereocilium organization	Otopetrin1, Alpha 2 adrenergic receptors [23], Glutamate receptor expression [24], c-FOS, vestibular hair cells [25],	Nausea related—cardiac sensitivity to baroreceptor reflex; raised Heart rate; raised cortisol; reduced dominant power on EGG baseline, questionnaire [34,35], Circadian	Motion sickness, Dizziness, Loss of Hearing, Postural imbalance, Vertigo	Cognition, Spatial memory, Graybiel scale, CDP, get up From Fall Test, Drop test/Jump down test, VEMP, OVAR response	Rotarod, Zebrafish Active Posturography (Zap); Floating Platform Tests—Postural sway—	(1) Robotic simulations (2) What happens in a more regular schedule? (3) What are the effects of recurring cycles of sleep	(1) Sleep loss and circadian changes affect the sensorimotor and cognitive function. (2) Caffeine + light – effective countermeasures. (3) Primary task is not affected	

		<p>cerebellar nodulus of adult rats [26–28], TEM of synaptic ribbons [29–33,149].</p>	<p>measurements Imaging: CT, fMRI, PET, EEG, MEG, TMS scan</p>			<p>measured by Center of Pressure (COP) Assay (=COP), Righting reflex, VEMP, OVAR response, Active vs. Passive motion on vestibular nucleus neurons, Mid-air righting reflex</p>	<p>deprivation? How do they recover? How does it affect performance? We need biomarkers for that.</p>	<p>during sleep loss but the secondary tasks are. This should be considered for effects on operational performance.</p>	
<p>DNA damage</p>	<p>DNA repair, DNA metabolic process, cellular response to DNA damage stimulus</p>	<p>Brain/other tissues: Staining with Anti-8-oxo-dg, 53bp1</p>	<p>Blood: DNA lesions via HPLC, 8-oxo dg, micronuclei, double strand DNA breaks, chromosomal aberrations/translocations, one carbon metabolites</p>	<p>Cell cycle checkpoint activation, DNA Repair, apoptosis,</p>	<p>Radiotherapy</p>	<p>Cognitive tests</p>	<p>Oxidative stress and inflammation related cognitive tests</p>	<p>Monitor the level of DNA damage over time- need astronaut data</p>	<p>(1) Since brain and neurons are not proliferative, DNA damage is might not be relevant in CNS. However, peripheral DNA damage is useful to studying the general diversity and individual differences of</p>

responses to radiation (again a surrogate, assuming that the brain will respond the same as the rest of the body). (2) Use baseline DNA damage as a predictor for responses to irradiation/spaceflight (astronaut panel pre/post flight). (3) Sleep deprivation exacerbates DNA damage in rats and humans. We cannot train/adapt to sleep deprivation. Note suggested markers for radiation dosage-bio-dosimetry: FLT3LG, SAA1, C3, VCAM

Blood brain barrier permeability	Inflammation, one	CSF: Albumin [150], Brain IHC—	Blood: Occludin, c-Fibronectin, Ubiquitin carboxyl-	Endothelial activation, Systemic	Inflammation, stroke, Alzheimer’s	<u>Stress</u> : Visual Analog Scale <u>Depression</u> : Beck	Locomotor activity, open field,	(1) Is BBB function altered in	(1) Circadian changes in astronauts (avg.
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carbon metabolism	Aquaporin 4 [151], IHC, MMP-9, long-term microglial activation, astrocyte morphology, Endothelial cells, Somatic mosaicism	terminal hydrolase isozyme L1, S100 calcium-binding protein B, Circulating brain microvascular endothelial cells ([150], stroke research), Corticosterone MMP-9, Cell free DNA Imaging: fMRI, PET scan, free water MRI; Epigenetic clock (accelerated aging).	inflammation, Kynurenine pathway, Tight junction damage, Oxidative stress, glial activation, MAPK pathway, PKC pathway, degradation of basal lamina and ECM.	Depression Inventory	hole-board, and grip strength tests, anxiety, and depressive behaviors	astronauts on ISS (or Artemis) missions? (2) Study the glymphatic system-removal of solutes from the brain across the BBB. (3) Need to understand the association of MMP9, occludins, S100, etc. with drainage of BBB. What is the physiological relevance? Glymphatic system is important for sleep as well. (4) Mutations, mosaicism etc. will affect the endothelial	sleep 6 h. though allocated 8–9 h) can add more stress. (2) Epigenetic and aging association [152]. Easily conducted. (3) DNA methylation observed in radiation and inhibition on global level can mitigate hypermethylation related cognitive deficits.
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cells and may cause BBB leakiness, leading to physiological effects. (5) Association of sleep with debris clearance. Amyloid clearance from the brain occurs during sleep → relevance to both sleep/circadian and glymphatic system. (6) Astrocyte morphology –unexplored. Astrocyte expressing AQP4 would be important for glymphatic system.

(7) Epigenetic clock measurements in astronauts and related to time in space etc. Or deep space to look at age acceleration
 (8) Development of rodent in vivo imaging technologies for BBB integrity.
 (9) Radiation induced senescence and functional readout in brain—glial cells, epithelial cells, somatic mosaicism

Vasculature	Blood vessel development, heart development	Adhesion molecules (VE-cadherin), tight junction proteins	Blood: Endothelial function markers (serum nitric oxide, tetra- and dihydrobiopterin	Adherens junction, Endothelial activation, systemic	Inflammation, stroke, Alzheimer’s	Stress: Visual Analog Scale Depression: Beck Depression Inventory	Locomotor activity, open field, hole-board, and grip	(1) What are the biochemical underpinnings of the	(1) Topological difference in vasculature and its susceptibility
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(Claudin 3, 5, 12, Occludin), Zo-1, MMPs	(BH4) and (BH2), soluble intercellular adhesion molecule-1 (sICAM-1), soluble vascular cell adhesion molecule-1 (sVCAM-1), endothelin-1, asymmetric dimethylarginine (ADMA), L-arginine, formate, and soluble E-selectin. Imaging: fMRI, PET scan. Noninvasive peripheral arterial tonometry (PAT) technology can be used to assess the reactive hyperemia index (RHI) and the augmentation index [153]; Vascular damage MRI measures: Cerebrovascular reactivity (CVR) (Pre and post flight): Present with CO2 challenge; Free water. Plasma:	inflammation, oxidative stress, hypoxia	strength tests, and depressive behaviors	thrombotic events seen inflight? (2) Also missing are chronic vascular injury markers. This biomarker has gained rapid adoption in many fields in the last few years. (3) Lack of cerebrovascular reactivity MRI data pre and post flight (4) Lack of 7T MRI for perivascular spaces (5) How do the biomarkers for vascular cognitive impairment change in astronauts?	towards the various stressors
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		Placental growth factor (PIGF), IL-8; VEGF-D; CSF: PIGF; IL-8				(6) Developing computational modeling of vascular changes?
miRNA regulation	Transcriptional regulation	Serum: miR-383-5p [154]	Transcriptional regulation	Cognitive tests	Cognitive tests	
Circadian Phase (sleep, sleepiness, performance impairment, immune function, endocrine function, bone metabolism, reproductive function)		Lipidomics, metabolomics, transcriptomics, proteomics	Accident, injury (short-term/immediate); cardiometabolic and neurological disorders, compromised immunity (long-term)	Cognitive tests	Cognitive tests	Candidates identified; operational validation required (1) Currently blood-borne but development of urinomics, saliva and breath matrices ongoing; (2) Can predict several days in advance; single vs. multiple samples. (3) Model organism—consideration of diurnal model over nocturnal. Marmoset? Indian palm squirrels?—restarting and reinventing the wheel? (4) Consistency in animal models and

		standardization in measurement. (5) Primary task is not affected during sleep loss but secondary tasks are. (Considered for operationally relevant performance)
Neuronal and brain Damage Markers	Blood: neurofilament, tau, abeta1-42, common pathology radiation and AD biomarkers (need to be explored)	Note suggested markers: NAA/Creatine ratio

4. Overall Summary and Recommendations

In total, hundreds of biomarkers have been identified and synthesized through this effort. Synthesizing across all three topical groups, the following common responses emerged as general themes:

- Biomarkers span all levels of data from molecules to behavior.
- Integrated stressors and integrated effects should be studied, including studies using multi-sensory approaches, for example, combined sleep and radiation exposure.
 - Note combined effects of HZE radiation exposure and sleep fragmentation in rodent models show dramatic effects specific to brain regions [109].
 - Integrated sensorimotor and cognition effects should be considered for study, e.g., olfaction and vestibular.
- The responses themselves will have multiple downstream impacts. Treatment may not be successful following a reductionist manner.
- Modifying factors should be identified and tracked throughout assessment, e.g., cognitive load, stress, circadian aspects, and sex, and their impacts on executive function and attention.
- Learning and plasticity were highlighted as critical areas to assess during spaceflight to determine the astronaut's general level of cognitive and sensorimotor adaptability.
- Biomarkers were recommended not just for immediate predictiveness, but also for long-term predictiveness of damage (late effects that can follow the initial injury by months or longer). As an example, some omics biomarkers may precede pathologies by months.
- Studying appropriate animal models in parallel with astronauts is extremely valuable for determining applicable constructs/responses, and to better understand the astronaut's condition.

We hope this effort yields usable knowledge and an effective tool for HRP and the CBS Project to improve monitoring and management of astronaut cognitive and behavioral health.

Author Contributions: All co-authors played a critical role in collecting and disseminating the knowledge in this article. Meeting organizers, discussion leads, participants, and facilitators are itemized in Appendix B. All authors have read and agreed to the published version of the manuscript.

Funding: This technical interchange meeting was funded by NASA Human Research Program, through the Human Factors and Behavioral Performance Element in conjunction with Space Radiation Element and the Human Health Countermeasures Element.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: We thank the MTSBI and CBS team members and Facilitators (Appendix B) for supporting all facets of the TIM and thank the Human Health Countermeasures, Space Radiation, and Human Factors & Behavioral Performance Elements of HRP and HRP Chief Scientist's Office for supporting this TIM. In particular, we thank David Dinges, Gregory Nelson, S. Robin Elgart, Janice Zawaski, and Scott J. Wood for expert consultation and support and Kerry George for critical manuscript review.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations/Acronyms

5-HT	5-hydroxytryptamine
5MTHF	L-Methylfolate
8-oxo-dG	8-Oxo-2'-deoxyguanosine
Ach	Acetylcholine
AOP	Adverse Outcome Pathways
AQP-4	Aquaporin-4
ARC	Ames Research Center
BBB	Blood Brain Barrier
BDNF	Brain-derived Neurotrophic Factor
BMed	Behavioral Medicine
CBS	Central Nervous System, Behavioral Medicine, and Sensorimotor
CNS	Central Nervous System
COX-2	Cyclooxygenase-2
CSF	Cerebrospinal Fluid
DA	Dopamine
DMN	Default Mode Network
EEG	Electroencephalogram
Flt-1	Fms Related Receptor Tyrosine Kinase 1
fMRI	Functional Magnetic Resonance Imaging
GFAP	Glial Fibrillary Acidic Protein
GI	Gastrointestinal
HNE	4-hydroxynonenal
HRP	Human Research Program
ICAM-1	Intercellular Adhesion Molecule 1
IL-15	Interleukin-15
IL-4	Interleukin-4
ISS	International Space Station
JSC	Johnson Space Center
MAPK	Mitogen-activated Protein Kinase
MDA	Malondialdehyde
MMP-9	Matrix Metalloproteinase 9
MMPs	Matrix metalloproteinase
MRI	Magnetic Resonance Imaging
MTSBI	Model Translation & Space Biology Integration
NFKb	Nuclear Factor kappa B
NHP	Non-human Primates
NIRS	Near-Infrared Spectroscopy
PI	Principal Investigator

ROS	Reactive Oxygen Species
S100b	S100 Calcium Binding Protein B
SM	Sensorimotor
TIM	Technical Interchange Meeting
TNF	Tumor Necrosis Factor
TREM	Triggering Receptor Expressed on Myeloid cells
TRR	Transthyretin
UCSF	University of California San Francisco
USRA	Universities Space Research Association
USUHS	Uniformed Services University of the Health Sciences
VCAM-1	Vascular Cell Adhesion Molecule 1
VOR	Vestibular-ocular Reflex
YKL-40	Chitinase-3-like protein 1
ZO-1	Zonula occludens-1

Appendix A. Agenda of Meeting

A NASA translational working group TIM titled Circuits and Biomarkers of the Central Nervous System Relating to Astronaut Performance (Biomarker TIM) was held virtually between 21–25 September 2020, and was supported by the NASA HRP’s Human Factors and Behavioral Performance Element in conjunction with Space Radiation Element and the Human Health Countermeasures Element. The goals of this Biomarker TIM were to (1) identify relevant brain regions, neural circuits, functions, and associated biomarkers that relate to operationally relevant performance and (2) identify any critical needs for new biomarker knowledge (“gaps”) that can be filled by additional focused and translational animal experiments that include a plausible pathway toward eventual biomarker validation in humans.

Deliverables addressing these goals may ultimately inform countermeasure strategies to maintain performance standards and identify performance limits for astronauts. To address the goals, 22 extramural experts from 19 academic institutions and 26 intramural experts from various NASA centers contributed to 15 talks reviewing findings from biomarker research on animals and humans in response to terrestrial and spaceflight stressors, and then participated in virtual thematic breakout sessions to systematically and qualitatively review biomarkers and associated brain circuits for 30 cognitive or behavioral constructs or physiological responses. The topics of the breakout sessions were sensorimotor influences (Group 1), behavioral medicine influences (Group 2), and integrated approaches to understanding operationally relevant performance (Group 3), and respective behavioral constructs listed in Table A1. Before the TIM, a portfolio of documents and scientific literature was shared with participants to frame the workshop and help the participants prepare.

Table A1. List of behavioral constructs for discussion groups.

Sensorimotor	Behavioral Medicine	Integrated Approaches: Physiological Responses
• Visual	• Memory	• Neuroinflammation
• Spatial Orientation	• Attention and Dual Tasking	• Neurotransmitters
• Vestibular	• Executive Function	• One-Carbon Metabolism
• Proprioception	• Working Memory	• Oxidative Stress

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|--------------------------------|--------------------------------|--|
| • Hearing | • Learning and Plasticity | • Synaptic Plasticity and Neurotrophic Factors |
| • Motion Sickness | • Social Processes | • Vestibular and Sensorimotor alterations |
| • Smell and Taste | • Individual Behavioral States | • DNA Damage |
| • Postural Control and Balance | • Arousal and Regulatory | • Blood Brain Barrier Permeability |
| • Locomotion | • Emotional Regulation | • Vasculature |
| • Fine Motor Control | • Risk Taking/Tolerance | • miRNA Regulation |
| • Perception | • Stress | • Circadian Phase |
| • Gaze | | • Neuronal Damage |
| • Pain | | |
-

Appendix B. Organizers & Participants

Lead Organizers

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James Lackner, PhD, Brandeis University

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Group 2: Behavioral Medicine Influences on Operational Performance (includes Cognition)

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References

1. Kiffer, F.; Alexander, T.; Anderson, J.E.; Groves, T.; Wang, J.; Sridharan, V.; Boerma, M.; Allen, A.R. Late Effects of 16O-Particle Radiation on Female Social and Cognitive Behavior and Hippocampal Physiology. *Radiat. Res.* **2019**, *191*, 278–294. <https://doi.org/10.1667/RR15092.1>.
2. NASA.; CBS. *Integrated Research Plan to Assess the Combined Effects of Space Radiation, Altered Gravity, and Isolation and Confinement on Crew Health and Performance: Problem Statement*; NASA: Washington, DC, USA, 2019.
3. Tu, D.; Basner, M.; Smith, M.G.; Williams, E.S.; Ryder, V.E.; Romoser, A.A.; Ecker, A.; Aeschbach, D.; Stahn, A.C.; Jones, C.W.; et al. Dynamic ensemble prediction of cognitive performance in spaceflight. *Sci. Rep.* **2022**, *12*, 11032. <https://doi.org/10.1038/s41598-022-14456-8>.
4. Grandjean, P. Paracelsus Revisited: The Dose Concept in a Complex World. *Basic. Clin. Pharmacol. Toxicol.* **2016**, *119*, 126–132. <https://doi.org/10.1111/bcpt.12622>.
5. Carnell, L.S. Spaceflight medical countermeasures: A strategic approach for mitigating effects from solar particle events. *Int. J. Radiat. Biol.* **2021**, *97*, S125–S131. <https://doi.org/10.1080/09553002.2020.1820603>.
6. Alfano, C.; Farina, L.; Petti, M. Networks as Biomarkers: Uses and Purposes. *Genes* **2023**, *14*, 429. <https://doi.org/10.3390/genes14020429>.

7. Institute of Medicine. *Evaluation of Biomarkers and Surrogate Endpoints in Chronic Disease*; Micheel, C.M., Ball, J.R., Eds.; The National Academies Press: Washington, DC, USA, 2010; ISBN 978-0-309-15129-0.
8. Simonsen, L.C.; Slaba, T.C.; Guida, P.; Rusek, A. NASA's first ground-based Galactic Cosmic Ray Simulator: Enabling a new era in space radiobiology research. *PLoS Biol.* **2020**, *18*, e3000669. <https://doi.org/10.1371/journal.pbio.3000669>.
9. Cullen, K.E. Vestibular processing during natural self-motion: Implications for perception and action. *Nat. Rev. Neurosci.* **2019**, *20*, 346–363. <https://doi.org/10.1038/s41583-019-0153-1>.
10. Mulavara, A.P.; Peters, B.T.; Miller, C.A.; Kofman, I.S.; Reschke, M.F.; Taylor, L.C.; Lawrence, E.L.; Wood, S.J.; Laurie, S.S.; Lee, S.M.C.; et al. Physiological and Functional Alterations after Spaceflight and Bed Rest. *Med. Sci. Sports Exerc.* **2018**, *50*, 1961–1980. <https://doi.org/10.1249/MSS.0000000000001615>.
11. Diamond, S.G.; Markham, C.H. Prediction of space motion sickness susceptibility by disconjugate eye torsion in parabolic flight. *Aviat. Space. Environ. Med.* **1991**, *62*, 201–205.
12. Markham, C.H.; Diamond, S.G. A predictive test for space motion sickness. *J. Vestib. Res.* **1993**, *3*, 289–295.
13. Bachatene, L.; Bharmauria, V.; Molotchnikoff, S. Adaptation and Neuronal Network in Visual Cortex. In *Visual Cortex—Current Status and Perspectives*; InTech: London, UK, 2012.
14. Fang, Q.; Chou, X.L.; Peng, B.; Zhong, W.; Zhang, L.I.; Tao, H.W. A Differential Circuit via Retino-Colliculo-Pulvinar Pathway Enhances Feature Selectivity in Visual Cortex through Surround Suppression. *Neuron* **2020**, *105*, 355–369.e6. <https://doi.org/10.1016/j.neuron.2019.10.027>.
15. Tamhane, M.; Cabrera-Ghayouri, S.; Abelian, G.; Viswanath, V. Review of Biomarkers in Ocular Matrices: Challenges and Opportunities. *Pharm. Res.* **2019**, *36*, 40. <https://doi.org/10.1007/s11095-019-2569-8>.
16. Hutton, S.B. Cognitive control of saccadic eye movements. *Brain Cogn.* **2008**, *68*, 327–340. <https://doi.org/10.1016/j.bandc.2008.08.021>.
17. Bussey, T.J.; Padain, T.L.; Skillings, E.A.; Winters, B.D.; Morton, A.J.; Saksida, L.M. The touchscreen cognitive testing method for rodents: How to get the best out of your rat. *Learn. Mem.* **2008**, *15*, 516–523. <https://doi.org/10.1101/lm.987808>.
18. Vesuna, S.; Kauvar, I.V.; Richman, E.; Gore, F.; Oskotsky, T.; Sava-Segal, C.; Luo, L.; Malenka, R.C.; Henderson, J.M.; Nuyujukian, P.; et al. Deep posteromedial cortical rhythm in dissociation. *Nature* **2020**, *586*, 87–94. <https://doi.org/10.1038/s41586-020-2731-9>.
19. Aitken, P.; Zheng, Y.; Smith, P.F. The modulation of hippocampal theta rhythm by the vestibular system. *J. Neurophysiol.* **2018**, *119*, 548–562. <https://doi.org/10.1152/jn.00548.2017>.
20. Dutta, S.M.; Hadley, M.M.; Peterman, S.; Jewell, J.S.; Duncan, V.D.; Britten, R.A. Quantitative Proteomic Analysis of the Hippocampus of Rats with GCR-Induced Spatial Memory Impairment. *Radiat. Res.* **2018**, *189*, 136–145. <https://doi.org/10.1667/RR14822.1>.
21. Tamura, A.; Iwamoto, T.; Ozaki, H.; Kimura, M.; Tsujimoto, Y.; Wada, Y. Wrist-worn electrodermal activity as a novel neurophysiological biomarker of autonomic symptoms in spatial disorientation. *Front. Neurol.* **2018**, *9*, 1056. <https://doi.org/10.3389/fneur.2018.01056>.
22. Smith, P.F.; Horii, A.; Russell, N.; Bilkey, D.K.; Zheng, Y.; Liu, P.; Kerr, D.S.; Darlington, C.L. The effects of vestibular lesions on hippocampal function in rats. *Prog. Neurobiol.* **2005**, *75*, 391–405. <https://doi.org/10.1016/j.pneurobio.2005.04.004>.
23. Dumas, R.; Mitton, D.; Laporte, S.; Dubouset, J.; Steib, J.P.; Lavaste, F.; Skalli, W. Explicit calibration method and specific device designed for stereoradiography. *J. Biomech.* **2003**, *36*, 827–834. [https://doi.org/10.1016/S0021-9290\(03\)00016-2](https://doi.org/10.1016/S0021-9290(03)00016-2).
24. Uno, Y.; Horii, A.; Uno, A.; Fuse, Y.; Fukushima, M.; Doi, K.; Kubo, T. Quantitative changes in mRNA expression of glutamate receptors in the rat peripheral and central vestibular systems following hypergravity. *J. Neurochem.* **2002**, *81*, 1308–1317. <https://doi.org/10.1046/j.1471-4159.2002.00933.x>.
25. Cohen, B.; Yakushin, S.B.; Holstein, G.R.; Dai, M.; Tomko, D.L.; Badakva, A.M.; Kozlovskaya, I.B. Vestibular Experiments in Space. *Adv. Space Biol. Med.* **2005**, *10*, 105–164. [https://doi.org/10.1016/S1569-2574\(05\)10005-7](https://doi.org/10.1016/S1569-2574(05)10005-7).
26. Krasnov, I.; D'yachkova, L. Ultrastructure of the cortex of the cerebellar nodulus in rats after a flight on the biosatellite Kosmos-1514. *Kosmicheskaya Biologiya i Aviakosmicheskaya Meditsina* **1986**, *20*, 45–48. <https://pubmed.ncbi.nlm.nih.gov/3784524/>.
27. Krasnov, I.; Dyachkova, L. The effect of space flight on the ultrastructure of the rat cerebellar and hemisphere cortex. *The Physiologist* **1990**, *33* (Suppl. 1), S29–S30. <https://pubmed.ncbi.nlm.nih.gov/2371337/>.
28. Holstein, G.; Kukielka, E.; Martinelli, G. Anatomical observations of the rat cerebellar nodulus after 24 h of spaceflight. *J. Gravitational Physiol.* **1999**, *6*, P47–P50.
29. Sultemeier, D.R.; Choy, K.R.; Schweizer, F.E.; Hoffman, L.F. Spaceflight-induced synaptic modifications within hair cells of the mammalian utricle. *J. Neurophysiol.* **2017**, *117*, 2163–2178. <https://doi.org/10.1152/jn.00240.2016>.
30. Ross, M.D. Morphological changes in rat vestibular system following weightlessness. *J. Vestib. Res.* **1993**, *3*, 241–251. <http://www.ncbi.nlm.nih.gov/pubmed/7903895>.
31. Ross, M.D. A spaceflight study of synaptic plasticity in adult rat vestibular maculas. *Acta Otolaryngol. Suppl.* **1994**, *516*, 3–14. <http://www.ncbi.nlm.nih.gov/pubmed/7976320>.
32. Ross, M.D. Changes in ribbon synapses and rough endoplasmic reticulum of rat utricular macular hair cells in weightlessness. *Acta Otolaryngol.* **2000**, *120*, 490–499. <https://doi.org/10.1080/000164800750045983>.
33. Ross, M.D.; Varelas, J. Synaptic ribbon plasticity, ribbon size and potential regulatory mechanisms in utricular and saccular maculae. *J. Vestib. Res.* **2005**, *15*, 17–30. <http://www.ncbi.nlm.nih.gov/pubmed/15908737>.

34. Zhang, L.L.; Wang, J.Q.; Qi, R.R.; Pan, L.L.; Li, M.; Cai, Y.L. Motion Sickness: Current Knowledge and Recent Advance. *CNS Neurosci. Ther.* **2016**, *22*, 15–24. <https://doi.org/10.1111/cns.12468>.
35. Ng, K.; Chua, Y.; Ban, V.F.; Gresty, M.; Coen, S.; Sanger, G.; Williams, S.; Barker, G.; Andrews, P.; Aziz, Q. Identifying human biomarkers of nausea for refining animal studies on emesis. *Gut* **2011**, *60*, A162–A162. <https://doi.org/10.1136/gut.2011.239301.344>.
36. Sohn, J.H.; Kim, C.H.; Lee, S.H.; Kim, J.H.; Lee, J.J. Diagnostic Value of Serum Biomarkers for Differentiating Central and Peripheral Causes of Acute Vertigo. *Front. Med.* **2020**, *7*, 84. <https://doi.org/10.3389/fmed.2020.00084>.
37. Wu, Y.; Han, W.; Yan, W.; Lu, X.; Zhou, M.; Li, L.; Guan, Q.; Fan, Z. Increased Otolin-1 in Serum as a Potential Biomarker for Idiopathic Benign Paroxysmal Positional Vertigo Episodes. *Front. Neurol.* **2020**, *11*, 367. <https://doi.org/10.3389/fneur.2020.00367>.
38. Hamann, K.F. Vibration-Induced Nystagmus: A Biomarker for Vestibular Deficits—A Synopsis. *ORL* **2017**, *79*, 112–120. <https://doi.org/10.1159/000455720>.
39. Osborne, D.; Theodorou, M.; Lee, H.; Ranger, M.; Hedley-Lewis, M.; Shawkat, F.; Harris, C.M.; Self, J.E. Supranuclear eye movements and nystagmus in children: A review of the literature and guide to clinical examination, interpretation of findings and age-appropriate norms. *Eye* **2019**, *33*, 261–273. <https://doi.org/10.1038/s41433-018-0216-y>.
40. Ryczko, D.; Dubuc, R. Dopamine and the Brainstem Locomotor Networks: From Lamprey to Human. *Front. Neurosci.* **2017**, *11*, 295. <https://doi.org/10.3389/fnins.2017.00295>.
41. Goulding, M. Circuits controlling vertebrate locomotion: Moving in a new direction. *Nat. Rev. Neurosci.* **2009**, *10*, 507–518. <https://doi.org/10.1038/nrn2608>.
42. Takakusaki, K. Functional Neuroanatomy for Posture and Gait Control. *J. Mov. Disord.* **2017**, *10*, 1–17. <https://doi.org/10.14802/jmd.16062>.
43. Marouane, E.; Rastoldo, G.; El Mahmoudi, N.; Péricat, D.; Chabbert, C.; Artzner, V.; Tighilet, B. Identification of New Biomarkers of Posturo-Locomotor Instability in a Rodent Model of Vestibular Pathology. *Front. Neurol.* **2020**, *11*, 470. <https://doi.org/10.3389/fneur.2020.00470>.
44. Gordon, C.R.; Shupak, A. Prevention and treatment of motion sickness. *Am. Fam. Physician* **2014**, *90*, 41–46. <http://www.ncbi.nlm.nih.gov/pubmed/25077505>.
45. Mo, F.-F.; Qin, H.-H.; Wang, X.-L.; Shen, Z.-L.; Xu, Z.; Wang, K.-H.; Cai, Y.-L.; Li, M. Acute hyperglycemia is related to gastrointestinal symptoms in motion sickness: An experimental study. *Physiol. Behav.* **2012**, *105*, 394–401. <https://doi.org/10.1016/j.physbeh.2011.08.024>.
46. Sato, F.; Uemura, Y.; Kanno, C.; Tsutsumi, Y.; Tomita, A.; Oka, A.; Kato, T.; Uchino, K.; Murakami, J.; Haque, T.; et al. Thalamo-insular pathway conveying orofacial muscle proprioception in the rat. *Neuroscience* **2017**, *365*, 158–178. <https://doi.org/10.1016/j.neuroscience.2017.09.050>.
47. Ranade, S.S.; Woo, S.-H.; Dubin, A.E.; Moshourab, R.A.; Wetzel, C.; Petrus, M.; Mathur, J.; Bégay, V.; Coste, B.; Mainquist, J.; et al. Piezo2 is the major transducer of mechanical forces for touch sensation in mice. *Nature* **2014**, *516*, 121–125. <https://doi.org/10.1038/nature13980>.
48. Oliveira Fernandes, M.; Tourtellotte, W.G. Egr3-dependent muscle spindle stretch receptor intrafusal muscle fiber differentiation and fusimotor innervation homeostasis. *J. Neurosci.* **2015**, *35*, 5566–5578. <https://doi.org/10.1523/JNEUROSCI.0241-15.2015>.
49. Goble, D.J.; Coxon, J.P.; Van Impe, A.; Geurts, M.; Van Hecke, W.; Sunaert, S.; Wenderoth, N.; Swinnen, S.P. The neural basis of central proprioceptive processing in older versus younger adults: An important sensory role for right putamen. *Hum. Brain Mapp.* **2012**, *33*, 895–908. <https://doi.org/10.1002/hbm.21257>.
50. Ueno, M.; Nakamura, Y.; Li, J.; Gu, Z.; Niehaus, J.; Maezawa, M.; Crone, S.A.; Goulding, M.; Baccei, M.L.; Yoshida, Y. Corticospinal Circuits from the Sensory and Motor Cortices Differentially Regulate Skilled Movements through Distinct Spinal Interneurons. *Cell Rep.* **2018**, *23*, 1286–1300.e7. <https://doi.org/10.1016/j.celrep.2018.03.137>.
51. Pradhan, S.D.; Brewer, B.R.; Carvell, G.E.; Sparto, P.J.; Delitto, A.; Matsuoka, Y. Assessment of fine motor control in individuals with Parkinson’s disease using force tracking with a secondary cognitive task. *J. Neurol. Phys. Ther.* **2010**, *34*, 32–40. <https://doi.org/10.1097/NPT.0b013e3181d055a6>.
52. Heys, J.G.; Wu, Z.; Allegra Mascaro, A.L.; Dombeck, D.A. Inactivation of the Medial Entorhinal Cortex Selectively Disrupts Learning of Interval Timing. *Cell Rep.* **2020**, *32*, 108163. <https://doi.org/10.1016/j.celrep.2020.108163>.
53. Milleret, C.; Bui Quoc, E. Beyond Rehabilitation of Acuity, Ocular Alignment, and Binocularity in Infantile Strabismus. *Front. Syst. Neurosci.* **2018**, *12*, 29. <https://doi.org/10.3389/fnsys.2018.00029>.
54. Steeds, C.E. The anatomy and physiology of pain. *Surgery* **2016**, *34*, 55–59. <https://doi.org/10.1016/j.mpsur.2015.11.005>.
55. Zjawiony, J.K.; Machado, A.S.; Menegatti, R.; Ghedini, P.C.; Costa, E.A.; Pedrino, G.R.; Lukas, S.E.; Franco, O.L.; Silva, O.N.; Fajemiroye, J.O. Cutting-Edge Search for Safer Opioid Pain Relief: Retrospective Review of Salvinorin A and Its Analogs. *Front. psychiatry* **2019**, *10*, 157. <https://doi.org/10.3389/fpsy.2019.00157>.
56. Graham, B.; Callister, R. Pain. In *The Mouse Nervous System*; Elsevier: Amsterdam, The Netherlands, 2012; pp. 589–606.
57. Pastoriza, L.N.; Morrow, T.J.; Casey, K.L. Medial frontal cortex lesions selectively attenuate the hot plate response: Possible nocifensive apraxia in the rat. *Pain* **1996**, *64*, 11–17. [https://doi.org/10.1016/0304-3959\(95\)00070-4](https://doi.org/10.1016/0304-3959(95)00070-4).
58. Niculescu, A.B.; Le-Niculescu, H.; Levey, D.F.; Roseberry, K.; Soe, K.C.; Rogers, J.; Khan, F.; Jones, T.; Judd, S.; McCormick, M.A.; et al. Towards precision medicine for pain: Diagnostic biomarkers and repurposed drugs. *Mol. Psychiatry* **2019**, *24*, 501–522. <https://doi.org/10.1038/s41380-018-0345-5>.

59. Luchting, B.; Hinske, L.C.G.; Rachinger-Adam, B.; Celi, L.A.; Kreth, S.; Azad, S.C. Soluble intercellular adhesion molecule-1: A potential biomarker for pain intensity in chronic pain patients. *Biomark. Med.* **2017**, *11*, 265–276. <https://doi.org/10.2217/bmm-2016-0246>.
60. Boissoneault, J.; Sevel, L.; Letzen, J.; Robinson, M.; Staud, R. Biomarkers for Musculoskeletal Pain Conditions: Use of Brain Imaging and Machine Learning. *Curr. Rheumatol. Rep.* **2017**, *19*, 5. <https://doi.org/10.1007/s11926-017-0629-9>.
61. Cowen, R.; Stasiowska, M.K.; Laycock, H.; Bantel, C. Assessing pain objectively: The use of physiological markers. *Anaesthesia* **2015**, *70*, 828–847. <https://doi.org/10.1111/anae.13018>.
62. Sakano, H. Developmental regulation of olfactory circuit formation in mice. *Dev. Growth Differ.* **2020**, *62*, 199–213. <https://doi.org/10.1111/dgd.12657>.
63. Latchney, S.E.; Rivera, P.D.; Mao, X.W.; Ferguson, V.L.; Bateman, T.A.; Stodieck, L.S.; Nelson, G.A.; Eisch, A.J. The effect of spaceflight on mouse olfactory bulb volume, neurogenesis, and cell death indicates the protective effect of novel environment. *J. Appl. Physiol.* **2014**, *116*, 1593–1604. <https://doi.org/10.1152/jappphysiol.01174.2013>.
64. Henkin, R.I.; Hosein, S.; Stateman, W.A.; Knöppel, A.B.; Abdelmeguid, M. Improved smell function with increased nasal mucosal sonic hedgehog in hyposmic patients after treatment with oral theophylline. *Am. J. Otolaryngol.* **2017**, *38*, 143–147. <https://doi.org/10.1016/j.amjoto.2016.11.010>.
65. Henkin, R.I.; Knöppel, A.B.; Abdelmeguid, M.; Stateman, W.A.; Hosein, S. Sonic hedgehog is present in parotid saliva and is decreased in patients with taste dysfunction. *J. Oral. Pathol. Med.* **2017**, *46*, 829–833. <https://doi.org/10.1111/jop.12541>.
66. Mueller, T. What is the Thalamus in Zebrafish? *Front. Neurosci.* **2012**, *6*, 64. <https://doi.org/10.3389/fnins.2012.00064>.
67. Sun, C.; Xuan, X.; Zhou, Z.; Yuan, Y.; Xue, F. A Preliminary Report on the Investigation of Prestin as a Biomarker for Idiopathic Sudden Sensorineural Hearing Loss. *Ear. Nose. Throat J.* **2020**, *99*, 528–531. <https://doi.org/10.1177/0145561319849949>.
68. Parham, K.; Dyhrfeld-Johnsen, J. Outer Hair Cell Molecular Protein, Prestin, as a Serum Biomarker for Hearing Loss: Proof of Concept. *Otol. Neurotol.* **2016**, *37*, 1217–1222. <https://doi.org/10.1097/MAO.0000000000001164>.
69. Basner, M.; Moore, T.M.; Hermosillo, E.; Nasrini, J.; Dinges, D.F.; Gur, R.C.; Johannes, B. Cognition Test Battery Performance Is Associated with Simulated 6df Spacecraft Docking Performance. *Aerosp. Med. Hum. Perform.* **2020**, *91*, 861–867. <https://doi.org/10.3357/amhp.5602.2020>.
70. Van Dongen, H.P.; Baynard, M.D.; Maislin, G.; Dinges, D.F. Systematic interindividual differences in neurobehavioral impairment from sleep loss: Evidence of trait-like differential vulnerability. *Sleep* **2004**, *27*, 423–433.
71. Tkachenko, O.; Dinges, D.F. Interindividual variability in neurobehavioral response to sleep loss: A comprehensive review. *Neurosci. Biobehav. Rev.* **2018**, *89*, 29–48. <https://doi.org/10.1016/j.neubiorev.2018.03.017>.
72. Bock, O.; Weigelt, C.; Bloomberg, J.J. Cognitive demand of human sensorimotor performance during an extended space mission: A dual-task study. *Aviat. Sp. Env. Med.* **2010**, *81*, 819–824. <https://doi.org/10.3357/asm.2608.2010>.
73. Moore, S.T.; Dilda, V.; Morris, T.R.; Yungheer, D.A.; MacDougall, H.G.; Wood, S.J. Long-duration spaceflight adversely affects post-standing operator proficiency. *Sci. Rep.* **2019**, *9*, 2677. <https://doi.org/10.1038/s41598-019-39058-9>.
74. Featherstone, R.E.; Melnychenko, O.; Siegel, S.J. Mismatch negativity in preclinical models of schizophrenia. *Schizophr. Res.* **2018**, *191*, 35–42. <https://doi.org/10.1016/j.schres.2017.07.039>.
75. Gonzalez-Castillo, J.; Hoy, C.W.; Handwerker, D.A.; Robinson, M.E.; Buchanan, L.C.; Saad, Z.S.; Bandettini, P.A. Tracking ongoing cognition in individuals using brief, whole-brain functional connectivity patterns. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 8762–8767. <https://doi.org/10.1073/pnas.1501242112>.
76. Edlow, B.L.; Chatelle, C.; Spencer, C.A.; Chu, C.J.; Bodien, Y.G.; O'Connor, K.L.; Hirschberg, R.E.; Hochberg, L.R.; Giacino, J.T.; Rosenthal, E.S.; et al. Early detection of consciousness in patients with acute severe traumatic brain injury. *Brain* **2017**, *140*, 2399–2414. <https://doi.org/10.1093/brain/awx176>.
77. Haufe, S.; DeGuzman, P.; Henin, S.; Arcaro, M.; Honey, C.J.; Hasson, U.; Parra, L.C. Elucidating relations between fMRI, ECoG, and EEG through a common natural stimulus. *Neuroimage* **2018**, *179*, 79–91. <https://doi.org/10.1016/j.neuroimage.2018.06.016>.
78. Itthipuripat, S.; Sprague, T.C.; Serences, J.T. Functional MRI and EEG Index Complementary Attentional Modulations. *J. Neurosci.* **2019**, *39*, 6162–6179. <https://doi.org/10.1523/JNEUROSCI.2519-18.2019>.
79. Nguyen, T.; Zhou, T.; Potter, T.; Zou, L.; Zhang, Y. The Cortical Network of Emotion Regulation: Insights From Advanced EEG-fMRI Integration Analysis. *IEEE Trans. Med. Imaging* **2019**, *38*, 2423–2433. <https://doi.org/10.1109/TMI.2019.2900978>.
80. Waser, M.; Benke, T.; Dal-Bianco, P.; Garn, H.; Mosbacher, J.A.; Ransmayr, G.; Schmidt, R.; Seiler, S.; Sorensen, H.B.D.; Jennum, P.J. Neuroimaging markers of global cognition in early Alzheimer's disease: A magnetic resonance imaging-electroencephalography study. *Brain Behav.* **2019**, *9*, e01197. <https://doi.org/10.1002/brb3.1197>.
81. Thatcher, R.; McAlaster, R.; Camacho, M.; Salazar, A.; Biver, C. Biophysical linkage between MRI and EEG amplitude in closed head injury. *Neuroimage* **1998**, *7*, 352–367.
82. Tien, Y.T.; Lee, W.J.; Liao, Y.C.; Wang, W.F.; Jhang, K.M.; Wang, S.J.; Fuh, J.L. Plasma Transthyretin as a Predictor of Amnesic Mild Cognitive Impairment Conversion to Dementia. *Sci. Rep.* **2019**, *9*, 18691. <https://doi.org/10.1038/s41598-019-55318-0>.
83. Dabrowski, W.; Siwicka-Gieroba, D.; Kotfis, K.; Zaid, S.; Terpilowska, S.; Robba, C.; Siwicki, A.K. The brain-gut axis—Where are we now and how can we modulate these connections? *Curr. Neuropharmacol.* **2020**, *19*, 1164–1177. <https://doi.org/10.2174/1570159x18666201119155535>.
84. Hattori, N.; Yamashiro, Y. The Gut-Brain Axis. *Ann. Nutr. Metab.* **2021**, *77* (Suppl. 2), 1–3. <https://doi.org/10.1159/000512226>.
85. Sun, M.; Ma, K.; Wen, J.; Wang, G.; Zhang, C.; Li, Q.; Bao, X.; Wang, H. A Review of the Brain-Gut-Microbiome Axis and the Potential Role of Microbiota in Alzheimer's Disease. *J. Alzheimers Dis.* **2020**, *73*, 849–865. <https://doi.org/10.3233/jad-190872>.

86. LaPelusa, M.; Donoviel, D.; Branzini, S.E.; Carlson, P.E., Jr.; Culler, S.; Cheema, A.K.; Kaddurah-Daouk, R.; Kelly, D.; de Cremoux, I.; Knight, R.; et al. Microbiome for Mars: Surveying microbiome connections to healthcare with implications for long-duration human spaceflight, virtual workshop, July 13, 2020. *Microbiome* **2021**, *9*, 2. <https://doi.org/10.1186/s40168-020-00951-5>.
87. Voorhies, A.A.; Mark Ott, C.; Mehta, S.; Pierson, D.L.; Crucian, B.E.; Feiveson, A.; Oubre, C.M.; Torralba, M.; Moncera, K.; Zhang, Y.; et al. Study of the impact of long-duration space missions at the International Space Station on the astronaut microbiome. *Sci. Rep.* **2019**, *9*, 9911. <https://doi.org/10.1038/s41598-019-46303-8>.
88. Stahn, A.C.; Gunga, H.C.; Kohlberg, E.; Gallinat, J.; Dinges, D.F.; Kühn, S. Brain Changes in Response to Long Antarctic Expeditions. *N. Engl. J. Med.* **2019**, *381*, 2273–2275. <https://doi.org/10.1056/NEJMc1904905>.
89. Raichle, M.E. The brain's default mode network. *Annu. Rev. Neurosci.* **2015**, *38*, 433–447. <https://doi.org/10.1146/annurev-neuro-071013-014030>.
90. Raichle, M.E.; MacLeod, A.M.; Snyder, A.Z.; Powers, W.J.; Gusnard, D.A.; Shulman, G.L. A default mode of brain function. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 676–682. <https://doi.org/10.1073/pnas.98.2.676>.
91. Buckner, R.L.; Andrews-Hanna, J.R.; Schacter, D.L. The brain's default network: Anatomy, function, and relevance to disease. *Ann. N. Y. Acad. Sci.* **2008**, *1124*, 1–38. <https://doi.org/10.1196/annals.1440.011>.
92. Baliki, M.N.; Geha, P.Y.; Apkarian, A.V.; Chialvo, D.R. Beyond feeling: Chronic pain hurts the brain, disrupting the default-mode network dynamics. *J. Neurosci.* **2008**, *28*, 1398–1403. <https://doi.org/10.1523/jneurosci.4123-07.2008>.
93. Broyd, S.J.; Demanuele, C.; Debener, S.; Helps, S.K.; James, C.J.; Sonuga-Barke, E.J. Default-mode brain dysfunction in mental disorders: A systematic review. *Neurosci. Biobehav. Rev.* **2009**, *33*, 279–296. <https://doi.org/10.1016/j.neubiorev.2008.09.002>.
94. Greicius, M.D.; Srivastava, G.; Reiss, A.L.; Menon, V. Default-mode network activity distinguishes Alzheimer's disease from healthy aging: Evidence from functional MRI. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 4637–4642. <https://doi.org/10.1073/pnas.0308627101>.
95. Kucyi, A.; Moayed, M.; Weissman-Fogel, I.; Goldberg, M.B.; Freeman, B.V.; Tenenbaum, H.C.; Davis, K.D. Enhanced medial prefrontal-default mode network functional connectivity in chronic pain and its association with pain rumination. *J. Neurosci.* **2014**, *34*, 3969–3975. <https://doi.org/10.1523/jneurosci.5055-13.2014>.
96. Whitfield-Gabrieli, S.; Ford, J.M. Default mode network activity and connectivity in psychopathology. *Annu. Rev. Clin. Psychol.* **2012**, *8*, 49–76. <https://doi.org/10.1146/annurev-clinpsy-032511-143049>.
97. Basner, M.; Rao, H.; Goel, N.; Dinges, D.F. Sleep deprivation and neurobehavioral dynamics. *Curr. Opin. Neurobiol.* **2013**, *23*, 854–863. <https://doi.org/10.1016/j.conb.2013.02.008>.
98. Picchioni, D.; Duyn, J.H.; Horovitz, S.G. Sleep and the functional connectome. *Neuroimage* **2013**, *80*, 387–396. <https://doi.org/10.1016/j.neuroimage.2013.05.067>.
99. Voss, M.W.; Soto, C.; Yoo, S.; Sodoma, M.; Vivar, C.; van Praag, H. Exercise and Hippocampal Memory Systems. *Trends Cogn. Sci.* **2019**, *23*, 318–333. <https://doi.org/10.1016/j.tics.2019.01.006>.
100. Sambataro, F.; Murty, V.P.; Callicott, J.H.; Tan, H.Y.; Das, S.; Weinberger, D.R.; Mattay, V.S. Age-related alterations in default mode network: Impact on working memory performance. *Neurobiol. Aging* **2010**, *31*, 839–852. <https://doi.org/10.1016/j.neurobiolaging.2008.05.022>.
101. Basner, M.; Nasrini, J.; Hermosillo, E.; McGuire, S.; Dinges, D.F.; Moore, T.M.; Gur, R.C.; Rittweger, J.; Mulder, E.; Wittkowski, M.; et al. Effects of -12° head-down tilt with and without elevated levels of CO₂ on cognitive performance: The SPACECOT study. *J. Appl. Physiol.* **2018**, *124*, 750–760. <https://doi.org/10.1152/jappphysiol.00855.2017>.
102. Jones, C.W.; Basner, M.; Mollicone, D.J.; Mott, C.M.; Dinges, D.F. Sleep deficiency in spaceflight is associated with degraded neurobehavioral functions and elevated stress in astronauts on six-month missions aboard the International Space Station. *Sleep* **2022**, *45*, zsac006. <https://doi.org/10.1093/sleep/zsac006>.
103. Barulli, D.; Stern, Y. Efficiency, capacity, compensation, maintenance, plasticity: Emerging concepts in cognitive reserve. *Trends Cogn. Sci.* **2013**, *17*, 502–509. <https://doi.org/10.1016/j.tics.2013.08.012>.
104. Clewett, D.V.; Lee, T.H.; Greening, S.; Ponzio, A.; Margalit, E.; Mather, M. Neuromelanin marks the spot: Identifying a locus coeruleus biomarker of cognitive reserve in healthy aging. *Neurobiol. Aging* **2016**, *37*, 117–126. <https://doi.org/10.1016/j.neurobiolaging.2015.09.019>.
105. Stern, Y. An approach to studying the neural correlates of reserve. *Brain Imaging Behav.* **2017**, *11*, 410–416. <https://doi.org/10.1007/s11682-016-9566-x>.
106. Gao, Z.; van Beugen, B.J.; De Zeeuw, C.I. Distributed synergistic plasticity and cerebellar learning. *Nat. Rev. Neurosci.* **2012**, *13*, 619–635. <https://doi.org/10.1038/nrn3312>.
107. McGregor, H.R.; Lee, J.K.; Mulder, E.R.; De Dios, Y.E.; Beltran, N.E.; Kofman, I.S.; Bloomberg, J.J.; Mulavara, A.P.; Seidler, R.D. Brain connectivity and behavioral changes in a spaceflight analog environment with elevated CO₂. *Neuroimage* **2021**, *225*, 117450. <https://doi.org/10.1016/j.neuroimage.2020.117450>.
108. Acharya, M.M.; Baulch, J.E.; Klein, P.M.; Baddour, A.A.D.; Apodaca, L.A.; Kramár, E.A.; Alikhani, L.; Garcia, C.; Angulo, M.C.; Batra, R.S.; et al. New Concerns for Neurocognitive Function during Deep Space Exposures to Chronic, Low Dose-Rate, Neutron Radiation. *eNeuro* **2019**, *6*, ENEURO.0094-19. <https://doi.org/10.1523/ENEURO.0094-19.2019>.
109. Britten, R.A.; Fesshaye, A.S.; Duncan, V.D.; Wellman, L.L.; Sanford, L.D. Sleep Fragmentation Exacerbates Executive Function Impairments Induced by Low Doses of Si Ions. *Radiat. Res.* **2020**, *194*, 116–123. <https://doi.org/10.1667/RADE-20-00080.1>.
110. Izquierdo, A.; Brigman, J.L.; Radke, A.K.; Rudebeck, P.H.; Holmes, A. The neural basis of reversal learning: An updated perspective. *Neuroscience* **2017**, *345*, 12–26. <https://doi.org/10.1016/j.neuroscience.2016.03.021>.

111. Nithianantharajah, J.; McKeachan, A.G.; Stewart, T.J.; Johnstone, M.; Blackwood, D.H.; St Clair, D.; Grant, S.G.; Bussey, T.J.; Saksida, L.M. Bridging the translational divide: Identical cognitive touchscreen testing in mice and humans carrying mutations in a disease-relevant homologous gene. *Sci. Rep.* **2015**, *5*, 14613. <https://doi.org/10.1038/srep14613>.
112. Chaumet, G.; Taillard, J.; Sagaspe, P.; Pagani, M.; Dinges, D.F.; Pavy-Le-Traon, A.; Bareille, M.P.; Rascol, O.; Philip, P. Confinement and sleep deprivation effects on propensity to take risks. *Aviat. Sp. Env. Med.* **2009**, *80*, 73–80. <https://doi.org/10.3357/ASEM.2366.2009>.
113. Dinges, D.F.; Basner, M.; Mollicone, D.; Ecker, A.; Jones, C. *Reaction Self Test on ISS: 6-Month Missions*; University of Pennsylvania, Philadelphia, PA, 2016.
114. Deng, W.; Aimone, J.B.; Gage, F.H. New neurons and new memories: How does adult hippocampal neurogenesis affect learning and memory? *Nat. Rev. Neurosci.* **2010**, *11*, 339–350. <https://doi.org/10.1038/nrn2822>.
115. Roy, D.S.; Kitamura, T.; Okuyama, T.; Ogawa, S.K.; Sun, C.; Obata, Y.; Yoshiki, A.; Tonegawa, S. Distinct Neural Circuits for the Formation and Retrieval of Episodic Memories. *Cell* **2017**, *170*, 1000–1012.e19. <https://doi.org/10.1016/j.cell.2017.07.013>.
116. Mueller, A.; Hong, D.S.; Shepard, S.; Moore, T. Linking ADHD to the Neural Circuitry of Attention. *Trends Cogn. Sci.* **2017**, *21*, 474–488. <https://doi.org/10.1016/j.tics.2017.03.009>.
117. Barkley, R.A. Behavioral inhibition, sustained attention, and executive functions: Constructing a unifying theory of ADHD. *Psychol. Bull.* **1997**, *121*, 65–94. <https://doi.org/10.1037/0033-2909.121.1.65>.
118. Mazaheri, A.; Coffey-Corina, S.; Mangun, G.R.; Bekker, E.M.; Berry, A.S.; Corbett, B.A. Functional disconnection of frontal cortex and visual cortex in attention-deficit/hyperactivity disorder. *Biol. Psychiatry* **2010**, *67*, 617–623. <https://doi.org/10.1016/j.biopsych.2009.11.022>.
119. Luck, S.J.; Ford, J.M.; Sarter, M.; Lustig, C. CNTRICS final biomarker selection: Control of attention. *Schizophr. Bull.* **2012**, *38*, 53–61. <https://doi.org/10.1093/schbul/sbr065>.
120. Faraone, S.V.; Bonvicini, C.; Scassellati, C. Biomarkers in the diagnosis of ADHD—promising directions. *Curr. Psychiatry Rep.* **2014**, *16*, 497. <https://doi.org/10.1007/s11920-014-0497-1>.
121. Bolkan, S.S.; Stujenske, J.M.; Parnaudeau, S.; Spellman, T.J.; Rauffenbart, C.; Abbas, A.I.; Harris, A.Z.; Gordon, J.A.; Kellendonk, C. Publisher Correction: Thalamic projections sustain prefrontal activity during working memory maintenance. *Nat. Neurosci.* **2018**, *21*, 1138. <https://doi.org/10.1038/s41593-018-0132-2>.
122. Crucian, B.E.; Zwart, S.R.; Mehta, S.; Uchakin, P.; Quiariarte, H.D.; Pierson, D.; Sams, C.F.; Smith, S.M. Plasma cytokine concentrations indicate that in vivo hormonal regulation of immunity is altered during long-duration spaceflight. *J. Interf. Cytokine Res.* **2014**, *34*, 778–786. <https://doi.org/10.1089/jir.2013.0129>.
123. Carey, L.; Nilsson, M.; Boyd, L. Learning following Brain Injury: Neural Plasticity Markers. *Neural Plast.* **2019**, *2019*, 4838159. <https://doi.org/10.1155/2019/4838159>.
124. Rosell, D.R.; Siever, L.J. The neurobiology of aggression and violence. *CNS Spectr.* **2015**, *20*, 254–279. <https://doi.org/10.1017/S109285291500019X>.
125. Ko, J. Neuroanatomical Substrates of Rodent Social Behavior: The Medial Prefrontal Cortex and Its Projection Patterns. *Front. Neural Circuits* **2017**, *11*, 41. <https://doi.org/10.3389/fncir.2017.00041>.
126. Freichel, M.; Vennekens, R.; Olausson, J.; Stolz, S.; Philipp, S.E.; Weissgerber, P.; Flockerzi, V. Functional role of TRPC proteins in native systems: Implications from knockout and knock-down studies. *J. Physiol.* **2005**, *567*, 59–66. <https://doi.org/10.1113/jphysiol.2005.092999>.
127. Trainor, B.C.; Workman, J.L.; Jessen, R.; Nelson, R.J. Impaired nitric oxide synthase signaling dissociates social investigation and aggression. *Behav. Neurosci.* **2007**, *121*, 362–369. <https://doi.org/10.1037/0735-7044.121.2.362>.
128. Nestler, E.J.; Barrot, M.; DiLeone, R.J.; Eisch, A.J.; Gold, S.J.; Monteggia, L.M. Neurobiology of depression. *Neuron* **2002**, *34*, 13–25. [https://doi.org/10.1016/s0896-6273\(02\)00653-0](https://doi.org/10.1016/s0896-6273(02)00653-0).
129. Crucian, B.; Stowe, R.; Quiariarte, H.; Pierson, D.; Sams, C. Monocyte phenotype and cytokine production profiles are dysregulated by short-duration spaceflight. *Aviat. Sp. Environ. Med.* **2011**, *82*, 857–862. <https://doi.org/10.3357/ASEM.3047.2011>.
130. Crucian, B.E.; Choukèr, A.; Simpson, R.J.; Mehta, S.; Marshall, G.; Smith, S.M.; Zwart, S.R.; Heer, M.; Ponomarev, S.; Whitmire, A.; et al. Immune system dysregulation during spaceflight: Potential countermeasures for deep space exploration missions. *Front. Immunol.* **2018**, *9*, 1437. <https://doi.org/10.3389/fimmu.2018.01437>.
131. Huang, W.; Ramsey, K.M.; Marcheva, B.; Bass, J. Circadian rhythms, sleep, and metabolism. *J. Clin. Invest.* **2011**, *121*, 2133–2141. <https://doi.org/10.1172/JCI46043>.
132. Moore, R.Y. Neural control of the pineal gland. *Behav. Brain Res.* **1996**, *73*, 125–130. [https://doi.org/10.1016/0166-4328\(96\)00083-6](https://doi.org/10.1016/0166-4328(96)00083-6).
133. Shang, X.; Xu, B.; Li, Q.; Zhai, B.; Xu, X.; Zhang, T. Neural oscillations as a bridge between glutamatergic system and emotional behaviors in simulated microgravity-induced mice. *Behav. Brain Res.* **2017**, *317*, 286–291. <https://doi.org/10.1016/j.bbr.2016.09.063>.
134. Janelidze, S.; Mattsson, N.; Stomrud, E.; Lindberg, O.; Palmqvist, S.; Zetterberg, H.; Blennow, K.; Hansson, O. CSF biomarkers of neuroinflammation and cerebrovascular dysfunction in early Alzheimer disease. *Neurology* **2018**, *91*, e867–e877. <https://doi.org/10.1212/WNL.0000000000006082>.
135. Raber, J.; Allen, A.R.; Rosi, S.; Sharma, S.; Dayger, C.; Davis, M.J.; Fike, J.R. Effects of (56)Fe radiation on hippocampal function in mice deficient in chemokine receptor 2 (CCR2). *Behav. Brain Res.* **2013**, *246*, 69–75. <https://doi.org/10.1016/j.bbr.2013.03.003>.

136. Aïd, S.; Bosetti, F. Targeting cyclooxygenases-1 and -2 in neuroinflammation: Therapeutic implications. *Biochimie* **2011**, *93*, 46–51. <https://doi.org/10.1016/j.biochi.2010.09.009>.
137. Derecki, N.C.; Cardani, A.N.; Yang, C.H.; Quinnes, K.M.; Crihfield, A.; Lynch, K.R.; Kipnis, J. Regulation of learning and memory by meningeal immunity: A key role for IL-4. *J. Exp. Med.* **2010**, *207*, 1067–1080. <https://doi.org/10.1084/jem.20091419>.
138. Guéguinou, N.; Bojados, M.; Jamon, M.; Derradji, H.; Baatout, S.; Tschirhart, E.; Fripiat, J.-P.; Legrand-Frossi, C. Stress response and humoral immune system alterations related to chronic hypergravity in mice. *Psychoneuroendocrinology* **2012**, *37*, 137–147. <https://doi.org/10.1016/j.psyneuen.2011.05.015>.
139. Kokhan, V.S.; Matveeva, M.I.; Bazyan, A.S.; Kudrin, V.S.; Mukhametov, A.; Shtemberg, A.S. Combined effects of antiorthostatic suspension and ionizing radiation on the behaviour and neurotransmitters changes in different brain structures of rats. *Behav. Brain Res.* **2017**, *320*, 473–483. <https://doi.org/10.1016/j.bbr.2016.10.032>.
140. Kulikova, E.A.; Kulikov, V.A.; Sinyakova, N.A.; Kulikov, A.V.; Popova, N.K. The effect of long-term hindlimb unloading on the expression of risk neurogenes encoding elements of serotonin-, dopaminergic systems and apoptosis; comparison with the effect of actual spaceflight on mouse brain. *Neurosci. Lett.* **2017**, *640*, 88–92. <https://doi.org/10.1016/j.neulet.2017.01.023>.
141. Wu, X.; Li, D.; Liu, J.; Diao, L.; Ling, S.; Li, Y.; Gao, J.; Fan, Q.; Sun, W.; Li, Q.; et al. Dammarane Sapogenins Ameliorates Neurocognitive Functional Impairment Induced by Simulated Long-Duration Spaceflight. *Front. Pharmacol.* **2017**, *8*, 315. <https://doi.org/10.3389/fphar.2017.00315>.
142. Newman, E.L.; Leonard, M.Z.; Arena, D.T.; de Almeida, R.M.M.; Miczek, K.A. Social defeat stress and escalation of cocaine and alcohol consumption: Focus on CRF. *Neurobiol. Stress.* **2018**, *9*, 151–165. <https://doi.org/10.1016/j.ynstr.2018.09.007>.
143. Wise, K.C.; Manna, S.K.; Yamauchi, K.; Ramesh, V.; Wilson, B.L.; Thomas, R.L.; Sarkar, S.; Kulkarni, A.D.; Pellis, N.R.; Ramesh, G.T. Activation of nuclear transcription factor-kappaB in mouse brain induced by a simulated microgravity environment. *In Vitro Cell. Dev. Biol. Anim.* **2005**, *41*, 118–123. <https://doi.org/10.1290/0501006.1>.
144. Delp, M.D.; Charvat, J.M.; Limoli, C.L.; Globus, R.K.; Ghosh, P. Apollo Lunar Astronauts Show Higher Cardiovascular Disease Mortality: Possible Deep Space Radiation Effects on the Vascular Endothelium. *Sci. Rep.* **2016**, *6*, 29901. <https://doi.org/10.1038/srep29901>.
145. Frijhoff, J.; Winyard, P.G.; Zarkovic, N.; Davies, S.S.; Stocker, R.; Cheng, D.; Knight, A.R.; Taylor, E.L.; Oettrich, J.; Ruskovska, T.; et al. Clinical Relevance of Biomarkers of Oxidative Stress. *Antioxid. Redox Signal.* **2015**, *23*, 1144–1170. <https://doi.org/10.1089/ars.2015.6317>.
146. Ikawa, M.; Okazawa, H.; Nakamoto, Y.; Yoneda, M. PET Imaging for Oxidative Stress in Neurodegenerative Disorders Associated with Mitochondrial Dysfunction. *Antioxidants* **2020**, *9*, 861. <https://doi.org/10.3390/antiox9090861>.
147. Sajdel-Sulkowska, E.M.; Xu, M.; Koibuchi, N. Cerebellar brain-derived neurotrophic factor, nerve growth factor, and neurotrophin-3 expression in male and female rats is differentially affected by hypergravity exposure during discrete developmental periods. *Cerebellum* **2009**, *8*, 454–462. <https://doi.org/10.1007/s12311-009-0122-8>.
148. Rudbeck, E.; Bellone, J.A.; Szücs, A.; Bonnicks, K.; Mehrotra-Carter, S.; Badaut, J.; Nelson, G.A.; Hartman, R.E.; Vlkolinský, R. Low-dose proton radiation effects in a transgenic mouse model of Alzheimer’s disease—Implications for space travel. *PLoS ONE* **2017**, *12*, e0186168. <https://doi.org/10.1371/journal.pone.0186168>.
149. Simpson, R.H.; Rodda, J.; Reinecke, C.J. Adrenoleukodystrophy in a mother and son. *J. Neurol. Neurosurg. Psychiatry* **1987**, *50*, 1165–1172. <https://doi.org/10.1136/jnnp.50.9.1165>.
150. Li, W.; Pan, R.; Qi, Z.; Liu, K.J. Current progress in searching for clinically useful biomarkers of blood-brain barrier damage following cerebral ischemia. *Brain Circ.* **2018**, *4*, 145–152. https://doi.org/10.4103/bc.bc_11_18.
151. Bellone, J.A.; Gifford, P.S.; Nishiyama, N.C.; Hartman, R.E.; Mao, X.W. Long-term effects of simulated microgravity and/or chronic exposure to low-dose gamma radiation on behavior and blood-brain barrier integrity. *NPJ Microgravity* **2016**, *2*, 16019. <https://doi.org/10.1038/npjimgrav.2016.19>.
152. Lu, A.T.; Quach, A.; Wilson, J.G.; Reiner, A.P.; Aviv, A.; Raj, K.; Hou, L.; Baccarelli, A.A.; Li, Y.; Stewart, J.D.; et al. DNA methylation GrimAge strongly predicts lifespan and healthspan. *Aging* **2019**, *11*, 303–327. <https://doi.org/10.18632/aging.101684>.
153. Brüll, V.; Burak, C.; Stoffel-Wagner, B.; Wolfram, S.; Nickenig, G.; Müller, C.; Langguth, P.; Altelheld, B.; Fimmers, R.; Stehle, P.; et al. Acute intake of quercetin from onion skin extract does not influence postprandial blood pressure and endothelial function in overweight-to-obese adults with hypertension: A randomized, double-blind, placebo-controlled, crossover trial. *Eur. J. Nutr.* **2017**, *56*, 1347–1357. <https://doi.org/10.1007/s00394-016-1185-1>.
154. Zhang, H.; Chen, J.; Wang, H.; Lu, X.; Li, K.; Yang, C.; Wu, F.; Xu, Z.; Nie, H.; Ding, B.; et al. Serum Metabolomics Associating With Circulating MicroRNA Profiles Reveal the Role of miR-383-5p in Rat Hippocampus Under Simulated Microgravity. *Front. Physiol.* **2020**, *11*, 939. <https://doi.org/10.3389/fphys.2020.00939>.

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