
Article

Comparison of *Auxenochlorella protothecoides* and *Chlorella* spp. Chloroplast Genomes: Evidence for Endosymbiosis and Horizontal Virus-like Gene Transfer

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Supplementary Figures

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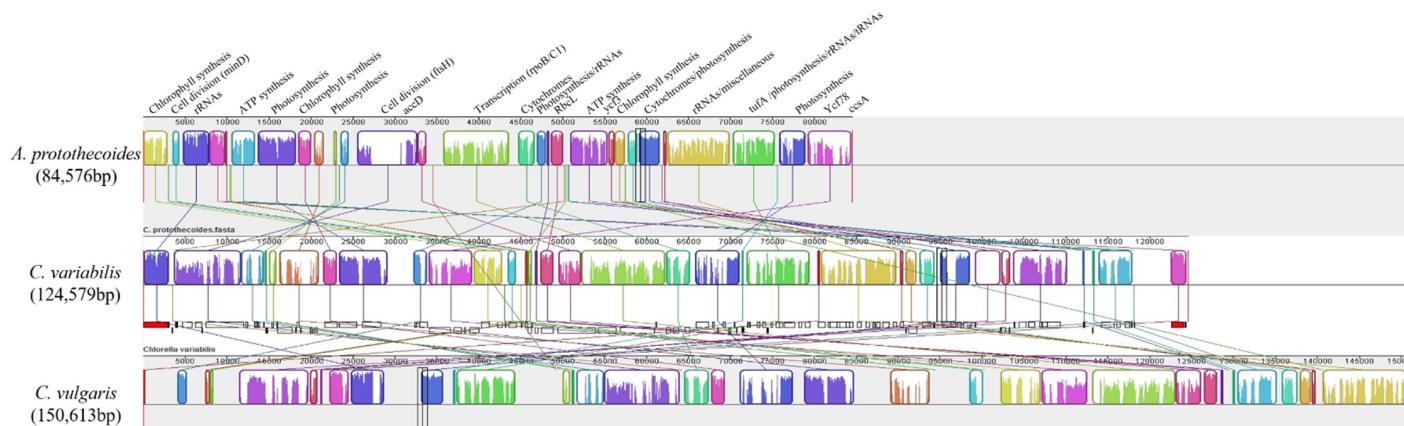


Figure S1. Global chloroplast genome sequence alignment of *Auxenochlorella protothecoides*, *Chlorella variabilis*, and *Chlorella vulgaris*. Local co-linear blocks (LCB) were identified as regions of homologous sequences shared by all three species. The lines connecting the genomes indicate orthologous gene clusters found in all three chloroplast genomes. The peaks in the LCB block indicate a similarity profile within each gene cluster. Areas that are completely white were not aligned and probably contain sequence elements specific to a particular genome. The multiple chloroplast genome alignment was conducted using Mauve software (<http://darlinglab.org/mauve/user-guide/introduction.html>).

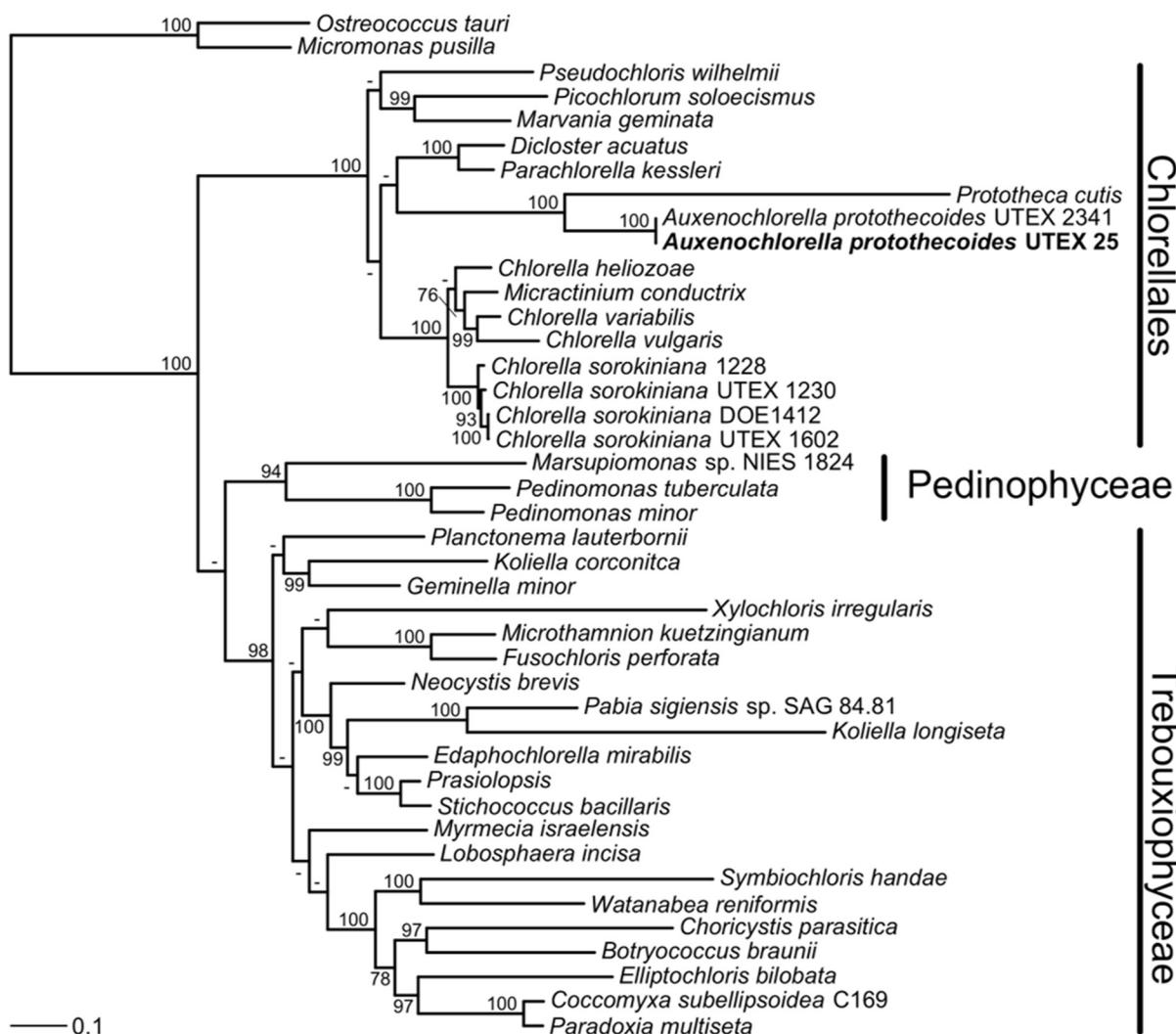


Figure S2. Phylogenetic relationships among the Trebouxiophyceae chloroplast genomes using Maximum Likelihood. Numbers placed at major nodes indicate bootstrap confidence values at $\geq 70\%$ for 1000 iterations.

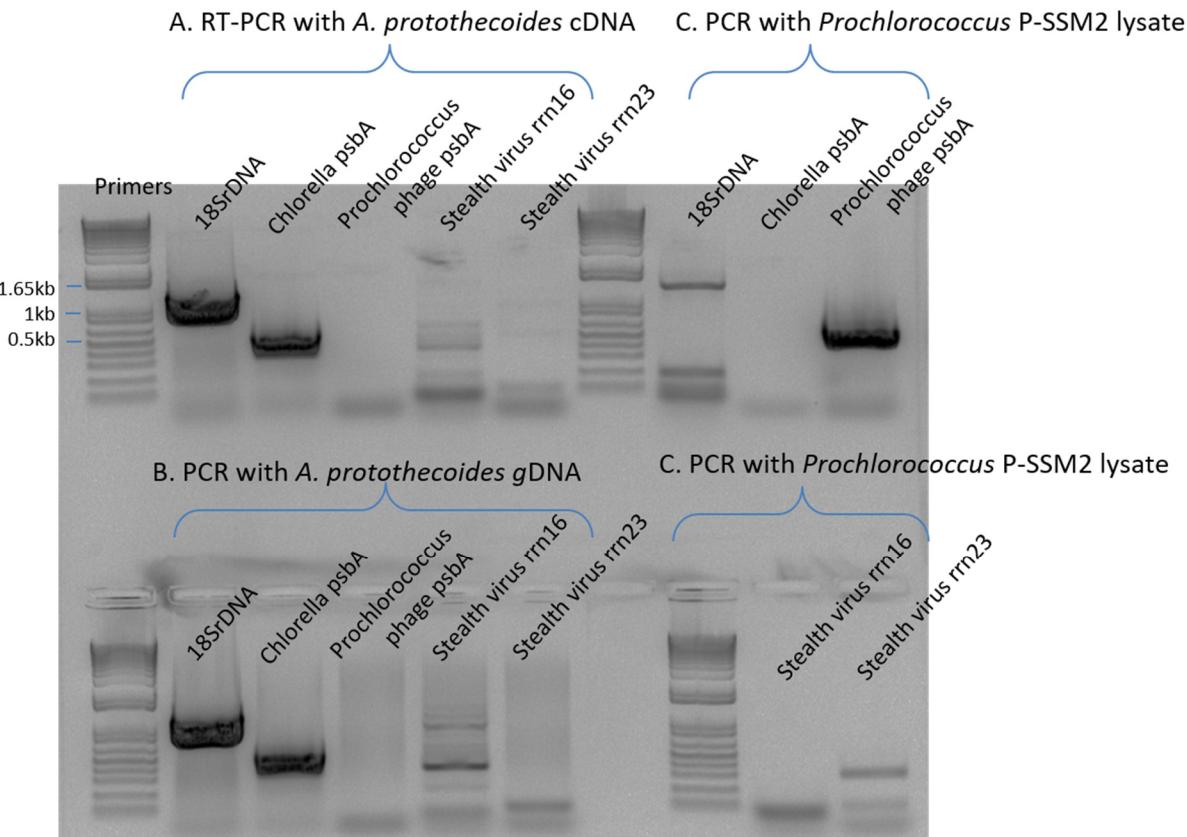


Figure S3. An original gel image used in Figure 6. Reverse Transcriptase-Polymerase Chain Reaction (RT-PCR) on *Auxenochlorella protothecoides* cDNA for examination of presence of viral genes (*Prochlorococcus* P-SSM2 *psbA*, Stealth virus *rrn16* and *rrn23*). 18S ribosomal DNA is used as internal control. A. RT-PCR was performed using gene specific primers of *A. protothecoides* *psbA*, *Prochlorococcus* phage *psbA* (588nt), Stealth virus *rrn16* (230nt) and Stealth virus *rrn23* (331nt). B. Conventional PCR was performed on *C. protothecoides* gDNA to show the presence of viral genes. C. Conventional PCR was performed on *Prochlorococcus* P-SSM2 lysate to confirm the presence of *psbA*.

Supplementary Tables

Table S1. Inventory of predicted genes within the *Auxenochlorella protothecoides*, *Chlorella variabilis*, and *Chlorella vulgaris* chloroplast genomes. trn: transfer RNA; rrn: ribosomal RNA; rpl: ribosomal protein large subunit; rrs: ribosomal protein small subunit; chl: chlorophyll biosynthesis, psa/psb: photosystem I/II subunits; rpo: RNA polymerase subunit; ycf: hypothetical chloroplast ORF; cysA: sulfate transport ATP-binding protein; ftsH/minE: plastid division; cemA: envelope membrane. †Gene including intron. *Not found in the other Chlorellaceae species in Table 1. § Partial sequences. The copy number for each is indicated parenthetically.

<i>A. protothecoides</i> UTEX 25 84,576 bp (114 genes) (GC: 30.8%)	<i>C. variabilis</i> 124,579 bp (113 genes) (GC: 34%)	<i>C. vulgaris</i> 150,613 bp (120 genes) (GC: 32%)
Photosynthesis		
Photosystem I		
<i>psaA</i> <i>psaB</i> <i>psaC</i> <i>psaI</i> <i>psaJ</i> <i>psaM</i>	<i>psaA</i> <i>psaB</i> <i>psaC</i> <i>psaI</i> <i>psaJ</i> <i>psaM</i>	<i>psaA</i> <i>psaB</i> <i>psaC</i> <i>psaI</i> <i>psaJ</i> <i>psaM</i>
Photosystem II		
<i>psbA</i> <i>psbB</i> <i>psbC</i> <i>psbD</i> <i>psbE</i> <i>psbF</i> <i>psbA</i> (2) <i>psbC</i> (3) <i>psbD</i> <i>psbE</i> <i>psbF</i> <i>psbH</i> <i>psbA</i> <i>psbC</i> <i>psbD</i> <i>psbE</i> <i>psbF</i> <i>psbH</i> <i>psbH</i> <i>psbI</i> <i>psbJ</i> <i>psbK</i> <i>psbL</i> <i>psbM</i> <i>psbI</i> <i>psbJ</i> <i>psbK</i> <i>psbL</i> <i>psbM</i> <i>psbN</i> <i>psbI</i> <i>psbJ</i> <i>psbK</i> <i>psbL</i> <i>psbM</i> <i>psbN</i> <i>psbT</i> <i>psbN</i> <i>psbT</i> <i>psbZ</i> <i>psbT</i> <i>psbZ</i> <i>psi_psbT</i> <i>psbZ</i> <i>psi_psbT</i>		
Cytochrome		
<i>petA</i> <i>petB</i> <i>petD</i> <i>petG</i> <i>petL</i>	<i>petA</i> <i>petB</i> <i>petD</i> <i>petG</i> <i>petL</i>	<i>petA</i> <i>petB</i> <i>petD</i> <i>petG</i> <i>petL</i>
ATP synthesis		
<i>atpA</i> <i>atpB</i> <i>atpE</i> <i>atpF</i> <i>atpH</i> <i>atpI</i>	<i>atpA</i> <i>atpB</i> <i>atpE</i> <i>atpF</i> <i>atpH</i> <i>atpI</i>	<i>atpA</i> <i>atpB</i> <i>atpE</i> <i>atpF</i> <i>atpH</i> <i>atpI</i>
Chlorophyll synthesis		
<i>chlB</i> <i>chlI</i> (2) <i>chlL</i> <i>chlN</i>	<i>chlB</i> <i>chlI</i> <i>chlL</i> † <i>chlN</i>	<i>chlB</i> <i>chlI</i> † <i>chlL</i> <i>chlN</i>
Rubisco		
<i>rbcL</i>	<i>rbcL</i>	<i>rbcL</i>
Transcription/translation		
RNA polymerase		
<i>rpoA</i> <i>rpoB</i> <i>rpoC1</i> <i>rpoC2</i>	<i>rpoA</i> <i>rpoB</i> <i>rpoC1</i> <i>rpoC2</i>	<i>rpoA</i> <i>rpoB</i> <i>rpoC1</i> <i>rpoC2</i> , § <i>rpoC2</i>
Translation factor		
<i>infA</i> <i>tufA</i>	<i>infA</i> <i>tufA</i>	<i>infA</i> <i>tufA</i>
Chloroplast division		
<i>minD</i> <i>ftsH</i>	<i>minD</i> <i>ftsH</i>	<i>minD</i> * <i>minE</i> <i>ftsH</i>
Ribosomal RNAs		
<i>rrn5</i> <i>rrn16</i> <i>rrn23</i> (2)	<i>rrn5</i> <i>rrn16</i> <i>rrn23</i> (2)	<i>rrn5</i> <i>rrn16</i> (4) <i>rrn23</i>
Ribosomal proteins		
Large subunits		
<i>rpl2</i> <i>rpl5</i> <i>rpl12</i> <i>rpl14</i> <i>rpl16</i> <i>rpl19</i> <i>rpl20</i> <i>rpl23</i> <i>rpl32</i> <i>rpl36</i>	<i>rpl2</i> <i>rpl5</i> <i>rpl12</i> <i>rpl14</i> <i>rpl16</i> <i>rpl19</i> <i>rpl20</i> <i>rpl23</i> <i>rpl32</i> <i>rpl36</i>	<i>rpl2</i> <i>rpl5</i> <i>rpl12</i> <i>rpl14</i> <i>rpl16</i> <i>rpl19</i> <i>rpl20</i> <i>rpl23</i> <i>rpl32</i> <i>rpl36</i>
Small subunits		
<i>rps2</i> <i>rps3</i> <i>rps4</i> <i>rps7</i> <i>rps8</i> <i>rps9</i> <i>rps11</i> <i>rps12</i> <i>rps12_5end</i> <i>rps14</i> <i>rps18</i>	<i>rps2</i> <i>rps3</i> <i>rps4</i> <i>rps7</i> <i>rps8</i> <i>rps9</i> <i>rps11</i> <i>rps2</i> <i>rps3</i> <i>rps4</i> <i>rps7</i> <i>rps8</i> <i>rps9</i> <i>rps11</i> <i>rps12</i> <i>rps12_3end</i> <i>rps14</i> <i>rps18</i> <i>rps19</i>	<i>rps2</i> <i>rps3</i> <i>rps4</i> <i>rps7</i> <i>rps8</i> <i>rps9</i> <i>rps11</i> <i>rps12</i> <i>rps12_3end</i> <i>rps14</i> <i>rps18</i> <i>rps19</i>
Transfer RNA		
<i>trnA-UGC</i> <i>trnC-GCA</i> <i>trnD-GUC</i> <i>trnE-UUC</i> <i>trnF-GAA</i> <i>trnfM-CAU</i>	<i>trnA-UGC</i> <i>trnC-GCA</i> (2) <i>trnD-GUC</i> <i>trnE-</i> <i>UUC</i> <i>trnF-GAA</i> <i>trnfM-CAU</i> <i>trnG-</i> <i>UUC</i> <i>trnF-GAA</i> <i>trnfM-CAU</i> <i>trnG-GCC</i> (2)	<i>trnA-UGC</i> <i>trnC-GCA</i> <i>trnD-GUC</i> <i>trnE-</i> <i>UUC</i> <i>trnF-GAA</i> <i>trnfM-CAU</i> <i>trnG-GCC</i> (2)

<i>trnG-GCC</i>	<i>trnG-UCC</i>	<i>trnH-GUG</i>	<i>GCC(2)</i>	<i>trnG-UCC</i>	<i>trnH-GUG</i>	<i>trnI-CAU</i>	<i>trnG-UCC</i>	<i>trnH-GUG</i>	<i>trnI-CAU</i>	<i>trnI-</i>
<i>trnH-GUG</i>	<i>trnI-CAU</i>	<i>trnI-GAU</i>	<i>trnK-</i>	<i>trnI-GAU</i>	<i>trnK-UUU</i>	<i>trnL-CAA</i>	<i>trnL-</i>	<i>GAU</i>	<i>trnK-UUU</i>	<i>trnL-CAA</i>
<i>trnL-UUU</i>	<i>trnL-GAG</i>	<i>trnL-UAA</i>	<i>trnL-UAG</i>	<i>GAG</i>	<i>trnL-UAG</i>	<i>trnM-CAU</i>	<i>trnN-GUU</i>	<i>trnL-UAG</i>	<i>trnM-CAU</i>	<i>trnN-GUU</i>
<i>trnM-CAU</i>	<i>trnN-GUU</i>	<i>trnP-GGG</i>	<i>trnP-GGG</i>	<i>trnP-UGG</i>	<i>trnQ-UUG</i>	<i>trnR-</i>	<i>trnP-GGG</i>	<i>trnP-UGG</i>	<i>trnQ-UUG</i>	<i>trnR-ACG</i>
<i>trnP-UGG</i>	<i>trnQ-UUG</i>	<i>trnR-ACG</i>	<i>ACG</i>	<i>trnR-CCG</i>	<i>trnR-UCU</i>	<i>trnS-GCU</i>	<i>trnR-CCG</i>	<i>trnR-UCU</i>	<i>trnS-GCU</i>	<i>trnS-</i>
<i>trnR-CCG</i>	<i>trnR-UCU</i>	<i>trnS-GCU</i>	<i>trnS-</i>	<i>trnS-GGA</i>	<i>trnS-UGA</i>	<i>trnT-GGU</i>	<i>trnT-</i>	<i>GGA</i>	<i>trnS-UGA</i>	<i>trnT-GGU</i>
<i>trnS-UGA</i>	<i>trnT-GGU</i>	<i>trnT-UGU</i>	<i>UGU</i>	<i>trnV-UAC</i>	<i>trnW-CCA</i>	<i>trnY-GUA</i>	<i>trnV-UAC</i>	<i>trnV-UAC</i>	<i>trnW-CCA</i>	<i>trnY-</i>
<i>trnV-UAC</i>	<i>trnW-CCA</i>	<i>trnY-GUA</i>								<i>GUA</i>

Fatty acid biosynthesis

	<i>accD</i>	<i>accD</i>	<i>accD</i>
Others			
<i>ccsA</i>	<i>cemA</i>	<i>clpP</i>	<i>cysT</i>
<i>ycf3</i>	<i>ycf4</i>	<i>ycf12</i>	<i>ycf78</i>

Table S2. Protein sequences ($n = 26$) identified based on the best-fitting partitioning ($n = 17$) and protein substitution models. The best-fitting partition and protein substitution scheme for 26 proteins, resulting in 17 partitions, determined by PartitionFinder version 2.1.1.

Partition number	Protein	Description	Number of sites	Substitution model
1	<i>atpB</i>	ATP synthase beta-subunit	598	LG4X
2	<i>psaA</i>	P700 chlorophyll a-apoprotein A1	1486	MTZOA+I+G+F
	<i>psaB</i>	P700 chlorophyll a-apoprotein A2		
3	<i>psaC</i>	subunit VII of photosystem I (Fe-S polypeptide)	556	LG4X
	<i>rbcL</i>	large subunit of Rubisco		
4	<i>psbC</i>	photosystem II CP43 apoprotein	488	LG4X
5	<i>RPL12</i>	ribosomal protein L12	136	LG+G+F
6	<i>RPL14</i>	ribosomal protein L14	217	LG4X
	<i>RPS19</i>	ribosomal protein S19		
7	<i>RPL16</i>	ribosomal protein L16	144	LG4X
8	<i>RPS4</i>	ribosomal protein S4	654	LG+I+G+F
	<i>RPS8</i>	ribosomal protein S8		
9	<i>RPL19</i>	ribosomal protein L19	494	LG4X
	<i>RPL5</i>	ribosomal protein L5		
10	<i>RPL2</i>	ribosomal protein L2	122	LG4X
	<i>RPL20</i>	ribosomal protein L20		
11	<i>RPL23</i>	ribosomal protein L23	737	JTTDCMUT+G+F
	<i>RPS18</i>	ribosomal protein S18		
12	<i>RPS2</i>	ribosomal protein S2		
	<i>RPL32</i>	ribosomal protein L32		
13	<i>RPS11</i>	ribosomal protein S11	136	LG4X
14	<i>RPS12</i>	ribosomal protein S12	127	CPREV+I+G
15	<i>RPS14</i>	ribosomal protein S14	102	LG4X
16	<i>RPS3</i>	ribosomal protein S3	425	MTREV+I+G+F
17	<i>RPS7</i>	ribosomal protein S7	157	LG4X
	<i>RPS9</i>	ribosomal protein S9	205	CPREV+I+G

Table S3. Results of BLASTn searches for the *Auxenochlorella protothecoides* UTEX 25 cpDNA against genome sequences of the Cyanobacteria (taxid:1117) available in the National Center for Biotechnology Information (NCBI) GenBank database (https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastn&PAGE_TYPE=BlastSearch&LINK_LOC=blasthome). For top hit selection, the following filtering criteria (percent identity: >86%, E-value: 0, and query coverage: >1%) were used.

NCBI GenBank Accession #	Cyanobacterium	Gene	Query Coverage	Percent Identity	Gaps
1 EU703214.1	Uncultured cyanobacterium	16S rRNA	1%	88.94%	54/1420(3%)
2 FJ901770.1	Uncultured cyanobacterium	16S rRNA	1%	89.49%	47/1313(3%)
3 CP060212.1	<i>Limnospira</i> sp. BM	23S rRNA	9%	86.55%	31/1323(2%)
4 MT863733.1	<i>Limnospira</i> sp. BM	23S rRNA	2%	86.48%	32/1324(2%)
5 AP012549.1	Cyanobacterium endosymbiont of <i>Epithemia turgida</i> isolate EtSB Lake Yunoko	23S rRNA	4%	87.98%	31/1190(2%)
6 NR076532.1	<i>Crocospaera subtropica</i> ATCC 51142	23S rRNA	2%	87.64%	29/1189(2%)
7 CP000806.1	<i>Crocospaera subtropica</i> ATCC 51142	23S rRNA	4%	87.64%	29/1189(2%)
8 AP018202.2	<i>Synechococcus vulgaris</i> NIES-2134	23S rRNA	4%	87.53%	29/1187(2%)
9 AP018341.1	Cyanobacterium endosymbiont of <i>Rhopalodia gibberula</i>	23S rRNA	4%	87.39%	31/1190(2%)
10 AY584507.1	<i>Euhalotheca</i> sp. BAA001	23S rRNA	2%	86.29%	28/1240(2%)
11 NR102529.1	<i>Stanieria cyanosphaera</i> PCC 7437	23S rRNA	2%	86.33%	24/1222(1%)
12 CP003653.1	<i>Stanieria cyanosphaera</i> PCC 7437	23S rRNA	6%	86.33%	24/1222(1%)
13 CP051167.1	<i>Oxynema</i> sp. AP17	23S rRNA	4%	86.81%	16/1183(1%)
14 NR076263.1	<i>Synechococcus elongatus</i> PCC 6301	23S rRNA	2%	86.75%	20/1185(1%)
15 CP000100.1	<i>Synechococcus elongatus</i> PCC 7942 = FACHB-805	23S rRNA	4%	86.75%	20/1185(1%)
16 LC455673.1	<i>Kamptonema laetevirens</i> NIES-31	23S rRNA	2%	86.51%	25/1201(2%)
17 X00512.1	<i>Synechococcus elongatus</i> PCC 6301	23S rRNA	2%	86.67%	22/1185(1%)
18 AP017375.1	<i>Stanieria</i> sp. NIES-3757	23S rRNA	8%	86.59%	19/1186(1%)
19 CP051168.1	<i>Phormidium</i> sp. ETS-05	23S rRNA	4%	86.59%	21/1186(1%)
20 CP000240.1	<i>Synechococcus</i> sp. JA-2-3B'a(2-13)	23S rRNA	4%	86.62%	26/1188(2%)
21 LC455660.1	<i>Planktothrix agardhii</i>	23S rRNA	2%	86.46%	20/1189(1%)
22 LC455656.1	<i>Planktothrix agardhii</i>	23S rRNA	2%	86.46%	20/1189(1%)
23 AP017991.1	<i>Planktothrix agardhii</i> NIES-204	23S rRNA	5%	86.46%	20/1189(1%)
24 LO018304.1	<i>Planktothrix agardhii</i>	23S rRNA	5%	86.46%	20/1189(1%)
25 CP000239.1	<i>Synechococcus</i> sp. JA-3-3Ab	23S rRNA	4%	86.48%	26/1191(2%)
26 NR076758.1	<i>Nostoc azollae</i> 0708	23S rRNA	2%	86.45%	23/1188(1%)
27 CP002059.1	<i>Nostoc azollae</i> 0708	23S rRNA	4%	86.45%	23/1188(1%)
28 AP018254.1	<i>Calothrix</i> sp. NIES-3974	23S rRNA	6%	86.29%	20/1189(1%)
29 LC455612.1	<i>Phormidium</i> sp. NIES-2121	23S rRNA	2%	86.18%	19/1187(1%)
30 CP016483.1	<i>Synechococcus</i> sp. PCC 8807	23S rRNA	4%	86.16%	20/1185(1%)
31 CP040360.1	<i>Synechococcus</i> sp. PCC 11901	23S rRNA	4%	86.08%	20/1185(1%)
32 CP013998.1	<i>Synechococcus</i> sp. PCC 73109	23S rRNA	4%	86.08%	20/1185(1%)
33 CP000951.1	<i>Synechococcus</i> sp. PCC 7002	23S rRNA	4%	86.08%	20/1185(1%)
34 NR102540.1	<i>Gloeocapsa</i> sp. PCC 7428	23S rRNA	2%	86.03%	22/1188(1%)
35 NR102530.1	<i>Cylindrospermum stagnale</i> PCC 7417	23S rRNA	2%	86.04%	23/1189(1%)
36 CP003646.1	<i>Gloeocapsa</i> sp. PCC 7428	23S rRNA	7%	86.03%	22/1188(1%)
37 CP003642.1	<i>Cylindrospermum stagnale</i> PCC 7417	23S rRNA	4%	86.04%	23/1189(1%)