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Therapeutic Interventions in Rat Models of Preterm Hypoxic Ischemic Injury: Effects of Hypothermia, Caffeine, and the Influence of Sex

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Abstract: Infants born prematurely have an increased risk of experiencing brain injury, specifically injury caused by Hypoxia Ischemia (HI). There is no approved treatment for preterm infants, in contrast to term infants that experience Hypoxic Ischemic Encephalopathy (HIE) and can be treated with hypothermia. Given this increased risk and lack of approved treatment, it is imperative to explore and model potential treatments in animal models of preterm injury. Hypothermia is one potential treatment, though cooling to current clinical standards has been found to be detrimental for preterm infants. However, mild hypothermia may prove useful. Caffeine is another treatment that is already used in preterm infants to treat apnea of prematurity, and has shown neuroprotective effects. Both of these treatments show sex differences in behavioral outcomes and neuroprotective effects, which are critical to explore when working to translate from animal to human. The effects and research history of hypothermia, caffeine and how sex affects these treatment outcomes will be explored further in this review article.

Keywords: preterm; hypoxia; ischemia; animal models; caffeine; hypothermia; sex differences

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1. Introduction

Approximately 11–12% of newborns in the US and 11% globally are premature, meaning born at less than 37 gestational weeks (GW) of prenatal development (aka preterm) [1–3]. These infants have increased mortality risk and are vulnerable to medical conditions that compromise cerebral oxygen supply and brain function, as well as impair long-term outcomes. Statistics indicate that more than 30% of premature infants have significant long-term disabilities [4,5]. Beyond stabilizing care, neonatologists have few medical options to protect surviving premature infants from long-term risks. There are no approved preterm neuroprotective treatments available, even for infants with severe intracranial bleeding or other forms of major encephalopathy. The high rates of premature birth, poor long-term outcomes, and lack of available treatments combine to make preterm brain injury a major health priority.

When cellular oxygen is reduced by hypoxia, ATP failure associated with inefficient anaerobic metabolism restricts neuronal activity. Complete oxygen restriction following ischemia or hypoxia-ischemia depletes high-energy metabolites even faster. This critical ATP loss leads to excess extracellular glutamate, prolonged neural depolarization, and elevated calcium and sodium influx. Maturity of glutamate receptors determines the rate of events [6,7], but eventually sodium over-load leads to cell swelling and necrotic cell death, while calcium over-load activates neuronal nitric oxide synthase (nNos) that accumulates and activates free radicals nitric oxide and peroxynitrate. Mitochondrial dysfunction and translocation of cytochrome-c from the mitochondria to the nucleus, caspase cleavage, chromatin condensation, and DNA fragmentation follow. This mitochondrial

dysfunction also increases the levels of reactive oxygen species, that can contribute to cellular death [8]. Simultaneous activation of poly (ADP-ribose) polymerase-1 (Parp-1; a DNA repair enzyme) leads to release of apoptotic factors [9–11]. Both pathways cause an increase in inflammation, microglial activation, cell death, and tissue loss. Two potential treatments, hypothermia and caffeine may help mediate these effects.

Although the mechanisms of action for caffeine and hypothermia are not precisely known, therapeutic effects of hypothermia are thought to be mediated by a preservation of energy metabolism, reduction in cytotoxic edema and free radicals, and reduction in immune and inflammatory response as well as apoptotic cell death. Caffeine acts as an adenosine antagonist (specifically at A2A and/or A1 receptors), and this interaction may attenuate calcium influx [12,13]. Caffeine may also reduce microglial activation [14] and/or reduce neuroinflammation either by decreasing the release of inflammatory proteins or blocking their action. [15].

Preterm brain injury and associated poor outcomes often originate in hypoxic-ischemic events (HI; decreased blood and/or oxygen supply) [16-19]. HI events often occur in the perinatal period. The associated preterm encephalopathies can follow from vascular fragility/immaturity and irregular blood pressure that together cause rupture and bleeding. Typical areas of injury involve the capillary-rich germinal matrix and/or peri-ventricular region (IVH hemorrhage). IVH still occurs in about 20-30% of severe preterm term infants despite now-routine antenatal corticosteroid use [16–18]. This injury particularly affects glial progenitors that are actively proliferating in the region, and are vulnerable to insult. Thus, intra-cranial hemorrhage in very preterm infants often damages white matter (e.g., motor tracts). This damage may be focal/cystic, or involve diffuse white matter loss (periventricular leukomalacia or PVL)-both are associated with cerebral palsy (CP) [17,18,20–27]. Perfusion failure/reperfusion injury can also result from falling blood pressure and capillary collapse, again with variable tissue loss. One of the most common causes of HI in the preterm brain is chronic hypoxemia secondary to respiratory insufficiency due to apnea, bradycardia and/or bronchopulmonary dysplasia (BPD). All of these conditions are very common in preterm infants. Resulting chronic low oxygen can cause diffuse brain injury in vascular "watershed" zones, and emergent PVL [19,21,28]. Although HI events are less common in late preterm infants, when they occur they typically lead to neurodegeneration associated with serious cognitive disabilities [17,18,29].

Other factors in the perinatal period that can affect susceptibility to HI include mothers experiencing chorioamnionitis, or preeclampsia, Preeclampsia is common in preterm mothers and leads to high blood pressure and protein in their urine. Infants born to preeclampsia mothers have a greater risk for periods of hypoxia and interventricular hemorrhage when compared to infants born prematurely but to mothers without preeclampsia [30]. Chorioamnionitis, an infection of the amniotic fluid and membranes surrounding the fetus, leads to an increase in inflammation for both the mother and infant fighting infection. Chorioamnionitis leads to an increased risk for PVL and IVH, and increased variability in brain oxygenation [31,32]. Events like placental abruption, which is more common in preterm delivery, increase instances of HIE in both term and preterm infants [33].

Poor outcomes associated with premature birth include blindness/deafness, cerebral palsy (CP), learning disability, ADHD, reduced neurodevelopmental scores (e.g., IQ and mental development index (MDI)) [34–54]. Specific statistics vary based on gestational age at birth, survival rates, medical complications, and criteria for study inclusion, but estimates suggest about 30% of surviving preterm infants (including those with and without HI events) experience cognitive delay or disability [55]. This rises to 40% for very preterm infants (< 32 GW), and exceeds 50% for extremely preterm infants (< 28 GW) [54]. Meta-analyses show that preterm children on average lose 10 IQ points, with specific difficulties in language acquisition and processing [45,56–60]. These language problems may relate to core underlying deficits in rapid auditory processing, such as the ability to process rapidly changing acoustic information in speech (e.g., /ba/ vs. /da/). Acoustic processing

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deficits are often seen in preterm infants, both with and without diagnosed brain injury [59,61–64]. Since these measures can be obtained pre-lingually (< 1 year) they offer excellent prognostic value, and in fact early rapid acoustic processing scores are highly predictive of later language outcomes [61,63–68]. Importantly, these skills are not only affected under pathologic conditions, but predict language abilities in typically developing children as well [61,63–69]. This link provides a strong rationale for inclusion of rapid acoustic processing in animal models of early brain injury as a translational marker for language-relevant outcomes in clinical populations. Premature infants show other cognitive problems as well, including deficits in learning, memory, and executive functioning [69–71], poor performance on spatial memory tasks [41,72], and impairments in visual attention with heightened incidence of attention-deficit disorders [34,36,39,43,73–79].

2. Animal Models of Preterm Brain Injury

To explore behavioral outcomes following HI injury, animal models of induced insult have been developed in various species including sheep, piglets, and rodents [62-64,66-74]. Large-animal models offer key insight to cellular pathology of HI-induced white matter injury since small animals have less myelin compared to humans [80,81], but have drawbacks such as limitations in behavioral assessment. Overall, the most widely employed neonatal HI model is one adapted from Levine by Rice & Vannucci [82]. This model involves HI injury induced in rats on postnatal day (P) 7 by unilateral ligation/cauterization of a carotid artery (typically right) combined with a period of exposure to a hypoxic environment (45–180 min at 8% O²). For many years, P7 rats were thought to simulate near-term injury in human neonates (~38 GW;) [83] based on comparisons of peak brain growth across species [84,85]. With more precise mapping of cross-species neurodevelopmental markers, it is now apparent that a P7 rat equates more closely to a late preterm infant (32–34 GW) [86,87], with P7 HI in rats simulating late preterm brain injury [87,88]. Other authors argue that P7 corresponds to 36 GW, which is still earlier than previously thought, with P10 being roughly equivalent to a term infant [89]. P7 HI male rats exhibit reliable deficits in rapid auditory processing as juveniles and adults, and deficits are persistent and stable within subjects [90-105]. This follows other rodent models of language-related disabilities [106-112], and supports the idea that rapid acoustic processing deficits in animal models can serve as a biomarker of neural disruptions linked to language impairments [61,63-68]. Deficits on spatial and working memory tasks have also been validated in P7 HI rats (Morris Water Maze task, Non-Spatial Maze, delayed-matchto-place maze tasks) [93,96,97,113–117], as well as deficits in visual attention [114,118,119]. Perhaps based on this extensive literature, most pre-clinical neuroprotective HI studies use a P7 HI rat model (Rice-Vannucci) [88,104,105,120-132].

Large animal models (lambs and piglets) have been subjected to HI injury during fetal life. Typically, *in utero* umbilical/placental occlusion is used to simulate preterm-like brain injury, and as a platform to assess neuroprotective strategies [132–136]. Only a few studies have assessed preterm-like (< P6) HI injuries in rodent models [92,101,126,137–140]. Importantly, even fewer of these studies report comprehensive long-term behavioral assessments following preterm-like injury, or investigations of therapeutic intervention applicable to preterm infants.

3. Therapeutic Hypothermia for Neonatal Brain Injury

Currently, the most widely employed and extensively studied intervention for full-term infants with moderate to severe hypoxic ischemic encephalopathy (HIE)—a form of post-HI injury associated with birth complications (e.g., umbilical occlusion) is hypothermia (head or whole-body cooling) [141,142]. Using head-cap or cooling blankets, affected infants are cooled by about 4 °C (33.5 °C core temperature) within 6 h of birth, and maintained for 72 h before re-warming. These accepted parameters are the result of extensive, randomized multi-site cooling trials that measured mortality, morbidity, motor impairments/CP, sensory impairments, and cognitive outcomes [47,143–145]. Cooling of full-

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term infants with moderate to severe HIE has been shown to reduce mortality and incidence of major disability by about 25% (depending on inclusion criteria and outcome measures) [142] and improves mental/cognitive outcomes. Importantly, however, this therapy remains approved only for term infants, \geq 36 GW. It should be noted that some clinicians are starting to use hypothermia at younger gestational ages. Despite being the most successful intervention used in term infants with HI (albeit with modest benefits), cooling has not been rigorously trialed or approved for < 36 GW infants. Reports are limited to a handful of late preterm case studies, and one clinical trial that proposed using a term hypothermia regime (4 °C for 72 h) in preterm infants of 32–36 GW. However, this trial was halted after recruitment of only 4 infants, and reported 50% adverse outcome and 25% mortality [146-148] (NCT00620711). Another study has shown similar adverse outcomes from preterm hypothermia, but as there was no control group of uncooled preterm infants, it is difficult to say if the cooling truly caused more harm [18,33]. Concerns about use of cooling therapies in preterm infants center on possible deleterious complications like hypoglycemia and coagulopathy [18,148]. Indeed, the 72 h regimen optimized to term HI infants may not be effective in preterm infants. Most pediatric associations agree that it should not be used in preterm infants, outside of a research setting, due to potential deleterious outcomes [20]. However, premature infants might benefit from an abbreviated and milder temperature reduction as shown below in work by Smith [103,104].

Potter [149] conducted a study using a hypothermia treatment modeled after the approved therapy for term infants with moderate or severe brain injury (as defined by prolonged core temperature reduction of 4 °C). They reduced core body temperature of P6 rat pups by 4 °C for 5 h following induction of HI (human GA = 32–35 week). Matched littermates received HI followed by normothermic conditions, or sham treatment with comparable hypothermia or normothermia. Results showed that this cooling intervention was not only ineffective, but it was also deleterious to both sham and P6 HI rats as measured by behavioral and neuroanatomical outcomes [129,149]. Specifically, cooled sham males had worse scores on a Silent Gap acoustic task compared to normothermic sham males, and significantly worse scores on Non-Spatial Water Maze (p < 0.01) [149]. These findings have important implications for therapeutic intervention in at-risk preterm human populations, and certainly promote caution in the application of existing hypothermia protocols to at-risk preterm infants. However, they did successfully use a more modest form of temperature reduction in P7 HI rats, specifically a 1.5 °C temperature reduction for 2 h (Figure 1). This intervention—in contrast to the 4 °C/5 h regimen described above was effective in mitigating some long-term behavioral deficits in both male and female rats with P7 HI injury and offered some protection from gross brain injury (Figure 1) [103,105]. Using a repeated measures ANOVA, cooled HI female and male rats showed improved performance compared to normothermic HI rats on a silent gap rapid auditory processing task (Females: p < 0.05, Males: p < 0.05) [104]. Cooled HI females and males also showed similar improvements in performance on the Morris Water Maze (females: *p* < 0.05, males: p < 0.05) compared to HI normothermic animals using a repeated measures ANOVA [104]. On the Non-spatial Water Maze a marginal treatment effect was seen in females, showing improved performance in the cooled HI group compared to normothermic HI animals (p = 0.08) [97]. Males on the same task did not show any overall treatment effects, but did show, through individual t-tests, normothermic rats doing significantly worse compared to shams (p = 0.05) [104].

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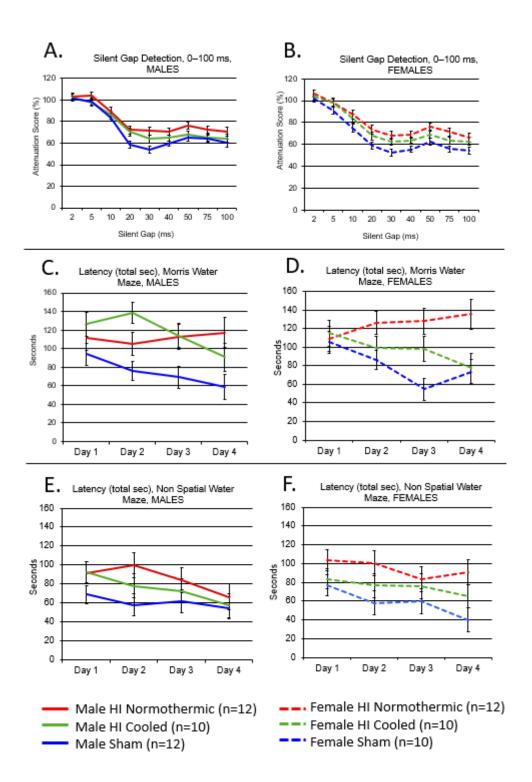


Figure 1. Cooling effects (Males and Females). Performance in P7 HI normothermic (red), HI cooled (green) and Sham normothermic (blue) male rats on: (**A**) silent gap 0–100 ms detection; (**C**) Morris Water Maze (MWM); and (**E**) Non-Spatial Water Maze (NSM). Performance in P7 HI normothermic (red), HI cooled (green) and Sham normothermic (blue) female rats on: (**B**) Silent Gap 0–100 ms detection; (**D**) Morris water maze (MWM); and (**F**) Non-Spatial Water Maze (NSM). Adapted with permission from Ref. [104]. 2015 by the authors.

4. Therapeutic Caffeine for Neonatal Brain Injury

To our knowledge the only therapeutic agents that have been explicitly trialed for neuroprotection in preterm infants are erythropoietin (PENUT trials) [150] and

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magnesium sulfate [122]. Magnesium sulfate has a drawback that treatment must be prophylactic, meaning the unnecessary treatment of many infants to achieve efficacy for those with injury. Moreover, magnesium only appears to improve motor issues. However, retrospective analyses of caffeine administered to preterm infants for respiratory stimulation suggest it may offer neuroprotection [151–155]. One of the most well-known prospective studies - caffeine for the treatment of apnea ("CAP") - randomized a large sample of preterm infants to caffeine or placebo treatment for respiratory stimulation. Results showed that preterm infants treated with caffeine citrate (20 mg/kg loading dose; 5–10 mg/kg daily maintenance) had better survival without neurodevelopmental disability, and lower rates of CP and cognitive delay at 18-21 months [156]. By 5 years of age caffeine was still associated with enhanced motor function, but not reduced rates of cerebral palsy or intellectual impairment [157]. However, other recent studies continue to support beneficial outcomes from caffeine, based primarily on retrospective assessment of caffeine administered for respiratory applications in preterm infants. Moreover, studies show that benefits from early/immediate caffeine treatment are more robust than later caffeine administration [151–155].

These clinical findings are consistent with animal work showing that treatment with caffeine citrate (50 mg/kg/day) from postnatal day 1 (P1) to P12 increased pyramidal neuronal growth in prefrontal cortex of rats at P35 and P70 [158]. Such growth-promoting effects could explain improved cognitive function reported in both children and rats treated with caffeine after brain injury. In another study, caffeine citrate (15–20 mg/kg/day) administered to rats from P2 to P6 reduced seizure susceptibility to some chemo-convulsants in both juvenile and adult rats [159]. Further, rat pups exposed to caffeine from P0–P12 or P1–7 via treated lactating dams and then subjected to induced injury showed enhanced myelination, with reduced ventriculomegaly and tissue loss relative to untreated injured pups [160,161]. Mice with neonatal HI were also shown to benefit from a single dose of caffeine [162]. These combined findings support rigorous testing of the therapeutic benefits of caffeine in preterm HI injury rat models.

Our lab conducted two published studies using caffeine treatment in moderate to late-preterm neonatal rat models of HI injury [149,163] and found a consistent benefit of caffeine treatment following HI. In the first study, we assessed effects of one-time caffeine in P7 HI male rats (late preterm), using a Rice-Vannucci injury (120 min hypoxia). Caffeine was administered immediately after injury. We found beneficial long-term effects of caffeine treatment on several tasks, including significant improvement on the Morris Water Maze task [93]. We also saw decreased injury-related brain pathology in caffeine-treated HI rats relative to untreated HI rats in adulthood. In the second study, we assessed the effects of an immediate plus 24 h-delayed dose of caffeine in male rats with HI induced on P6 [149]. Again, we found significant benefits of caffeine treatment on a Rotarod task, Silent Gap 0-100 ms acoustic detection task, and Non-Spatial Water Maze (though trends were seen on other tasks). We also found reduced injury-related pathology in caffeinetreated HI compared to untreated HI rats in adulthood. Importantly, a recent pilot study extended these beneficial effects to P6 HI-injured female rats, with improved performance on Silent Gap 0–100 ms and Morris Water Maze, and reduced neuropathologic indices in caffeine-treated P6 HI female rats compared to HI-saline female littermates [164].

We performed a *post hoc* pooled multi-variate ANOVA to test for differences between P6 and P7 rats. This also included analysis of a single vs. multiple dose regimen of therapy. Sex was not use as a variable. Results of a repeated measures ANOVA showed robust significant benefits of caffeine on outcome measures. We found no difference in the beneficial effect between P6 and P7. We continued to see superior behavioral effects with caffeine treated pups compared to non-treated on Silent Gap acoustic discrimination 0–100 ms (F(1,47) = 11.3, p < 0.01; Figure 2A); Morris Water Maze (F(1,47) = 4.2, p < 0.05; Figure 2B); and Non-spatial Water Maze (F(1,47) = 4.5, p < 0.05; Figure 2C). In all analyses, Day was significant (reflecting ongoing improvement) but did not interact with Treatment. Task was further included as a repeated-measures variable (3 levels), and here the effect

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size for the difference between HI saline and HI caffeine-treated rats was quite robust (0.93 effect size; F(1,47) = 10.5, p < 0.01), confirming that these three tasks together (SG-100, MWM and NSM) provide an excellent and sensitive tool for preliminary screening of therapeutic caffeine treatment parameters in a rat neonatal HI model.

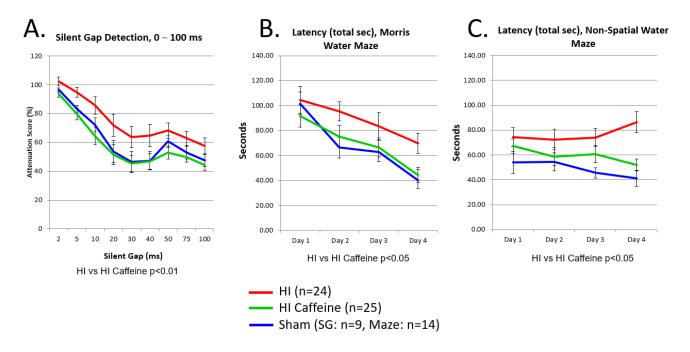


Figure 2. Caffeine Effects (Males). Performance in HI saline-treated (red), HI caffeine-treated (green) and sham-saline (blue) male rats on: (**A**) Silent Gap 0–100 ms detection; (**B**) spatial Morris Water Maze (MWM); and (**C**) Non-spatial water maze (NSM). Unpublished data re-analysis Adapted with permission from Refs. [93,149]. 2013 by the authors and 2018 by ISDN.

5. Mechanisms of Hypoxia/Ischemia, Cell Death, and Brain Injury of Prematurity

As mentioned in the introduction, the exact mechanisms of action of hypothermia and caffeine treatment are not fully understood. For hypothermia, it is thought that benefits are seen through preservation of energy metabolism and a reduction in cytotoxic edema, free radicals, inflammation, and apoptotic cell death. Caffeine, as a non-selective adenosine antagonist is thought to act at the A_1 and A_{2A} receptors to reduce calcium influx, and potentially reduce microglial activation or the release of inflammatory compounds [15–17].

We recently addressed this question in a study examining the effects of caffeine treatment following P6 HI injury in a rat model on microglial activation as measured 48 h postinjury. Both male and female rats were assigned to either a P6 Sham, HI injury followed by saline, or HI injury followed by caffeine treatment [165]. Results showed that both male and female rats with P6 HI injury and saline treatment exhibited significantly elevated chromatin condensation 48 h after injury in the right cortex (side of HI injury) as measured by concentrated DAPI staining. Male and female rats with P6 HI injury followed by caffeine treatment did not differ significantly from shams and had chromatin condensation values mid-way between sham and HI saline animals. The lack of significant chromatin condensation in HI caffeine rats relative to shams—despite a robust effect in HI rats treated only with saline, confirms behavioral evidence of therapeutic caffeine effects, as observed using behavioral measures shown above [165]. Interestingly, though we saw similar reductions in cell death between the sexes, when we looked at microglial activation as measured by soma-size of Iba-labelled cells, we saw trending microglial activation

differences in HI saline male rats versus HI caffeine male rats (p = 0.08, one-tail) specifically with less activation in caffeine treated subjects. A very different pattern was seen in females, where we saw no significant microglial activation in HI saline animals relative to shams, nor effects of caffeine on microglial activation. These results suggest that therapeutic effects of caffeine in neonatal males may occur at least in part via reductions in microglial activation, whereas female benefits from caffeine appear to be conferred via different protective mechanisms. This is despite the fact that, both sexes show structural and functional benefits following caffeine treatment (per cumulative studies).

6. Sex Differences in Neonatal HI and Therapeutic Intervention

Our overall findings combined with the existing literature highlight the importance of including sex differences in studies of perinatal brain injury in term and preterm infants as well as sex differences in response to therapeutic intervention [128,161,166–168]. While caffeine appears to exert similar levels of protection in both sexes, hypothermia appears to have different degrees of benefit in males and females [104,164]. Surprisingly few neonatal brain injury outcome studies consider sex as a variable, and none of the human trials provide outcome data for males and females separately. Yet, human research supports differences in the events surrounding HI in males and females. Male infants are 61% more likely to experience brain injury [169,170], and have a higher incidence of prematurity, anoxia, intraventricular hemorrhage, and mortality from prematurity [171-175]. Males are more likely to be diagnosed with developmental disorders including speech and language disorders and ADHD. Males develop CP at a higher frequency relative to females with similar brain injury [171,176-179]. The heightened susceptibility of males to HI insult is coupled with exacerbated cognitive/behavioral deficits following HI [48,173,180–183].Preterm males either with or without with neonatal HI injuries score lower than comparable females cognitive and developmental outcomes [38,39,43,49,171,175,176,180,184,185]. These differences were summarized in a meta-analysis performed by our lab showing significantly worse IQ outcomes for preterm male infants compared to matched females [91]. Despite important clinical implications, precise mechanisms underlying sex differences in outcomes are not well understood. Testosterone may exacerbate neural injury in the male or estrogen/progesterone could be protective in the female [88,140,186-189]. Other evidence suggests sex differences in cell death pathways may favor females [10,100,190–193] as well as sex differences in inflammation, microglial activation, and/or post-injury cell genesis [189,194,195]. Knowledge about the mechanisms of this female advantage could lead to novel therapeutic discovery. Moreover, males and females may respond differently to interventions, which is clinically important [128,166-168,176,180]. A classic illustration is the case of indomethacin, which was widely used in preterm infants to lower IVH risk, and later discovered to be effective only in males [196,197].

Overall results show that although caffeine reduces cell death in both male and female rats as measured by DAPI to index chromatin condensation, offering equivalent protection, the intermediary impact on microglial activation as a therapeutic mechanism may be quite different in the sexes. This could have critical implications for individual optimization of timing, dosing, and/or interactions between caffeine and other adjunct interventions in at-risk male versus female infants. Our results may also indicate that different mechanisms of cell death in neonatal males and females following HI may lend themselves to sex-specific next generation interventions.

7. Conclusions

Translational work and animal models looking at preterm HI are extremely important especially because preterm infants are at higher risk of brain injury and there are no approved treatments. This contrasts term infants, where injury risk is lower but approved treatment exists. It also appears from work in our lab and others that what is therapeutic for term infants may not be effective for preterm infants (e.g., hypothermic

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treatment) or what works/is beneficial for one sex may not be for the other (indomethacin, mild hypothermia, and caffeine). In addition, the effect of injury appears to have different outcomes in extremely preterm vs. mildly preterm infants [92]. Thus, extensive animal modeling helps to avoid later discoveries of adverse outcomes once treatment trials move to humans.

Current work from our lab and other studies show that mild hypothermia and caffeine treatment have great potential to be effective treatments for preterm HI. There are other potential treatments in the translational pipeline, such as metformin, exendin-4, and leptin [198–200]. Further work on potential treatment must model conditions of prematurity and sex. As seen above, sex and gestational age may affect the mechanism of protection as well as how effective the treatment is. Modeling injury in rodents < P7, reporting sex differences, and looking at a wider variety of induced brain injury (because there are many potential causes of brain injury in the preterm), would help to ensure we can better transition from pre-clinical animal models into human trials. To further help validate our models, there are several factors that may contribute to development of preterm brain injury that should also be considered (genetics, inflammatory conditions, other comorbidities, and recurrent hypoxia). These other variables must be included and addressed in research to ensure we are using the most valid modeling we can, which will enhance the transition into human studies.

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References

- Kochanek, K.D.; Kirmeyer, S.E.; Martin, J.A.; Strobino, D.M.; Guyer, B. Annual Summary of Vital Statistics: 2009. *Pediatrics* 2012, 129, 338–348.
- 2. Purisch, S.E.; Gyamfi-Bannerman, C. Epidemiology of preterm birth. Semin. Perinatol. 2017, 41, 387–391.
- 3. Walani, S.R. Global burden of preterm birth. Int. J. Gynecol. Obstet. 2020, 150, 31–33. https://doi.org/10.1002/ijgo.13195.
- 4. Boyle, C.A.; Boulet, S.; Schieve, L.A.; Cohen, R.A.; Blumberg, S.J.; Yeargin-Allsopp, M.; Visser, S.; Kogan, M.D. Trends in the Prevalence of Developmental Disabilities in US Children, 1997–2008. *Pediatrics* **2011**, 127, 1034–1042.
- 5. Platt, M.J. Outcomes in preterm infants. *Public Health* **2014**, 128, 399–403.
- 6. Cernada, M.; Cubells, E.; Torres-Cuevas, I.; Kuligowski, J.; Escobar, J.; Aguar, M.; Escrig, R.; Vento, M. Oxygen in the delivery room. *Early Hum. Dev.* **2013**, *89*, S11–S13.
- 7. Fields, R.D. Glutamate receptors: The cause or cure in perinatal white matter injury? *Neuron Glia Biol.* **2010**, *6*, 209–211.
- 8. Dan Dunn, J.; Alvarez, L.A.; Zhang, X.; Soldati, T. Reactive oxygen species and mitochondria: A nexus of cellular homeostasis. *Redox Biol.* **2015**, *6*, 472–485. https://doi.org/10.1016/j.redox.2015.09.005.
- 9. Ferrer, I.; Planas, A.M. Signaling of cell death and cell survival following focal cerebral ischemia: Life and death struggle in the penumbra. *J. Neuropathol. Exp. Neurol.* **2003**, *62*, 329–339.
- 10. Lang, J.T.; McCullough, L.D. Pathways to ischemic neuronal cell death: Are sex differences relevant? *J. Transl. Med.* **2008**, *6*, 1–10.
- 11. Zhu, C.; Xu, F.; Wang, X.; Shibata, M.; Uchiyama, Y.; Blomgren, K.; Hagberg, H. Different apoptotic mechanisms are activated in male and female brains after neonatal hypoxia-ischaemia. *J. Neurochem.* **2006**, *96*, 1016–1027.
- 12. Chen, X.; He, H.; Wang, G.; Yang, B.; Ren, W.; Ma, L.; Yu, Q. Stereospecific determination of cis- and trans-resveratrol in rat plasma by HPLC: Application to pharmacokinetic studies. *Biomed. Chromatogr.* **2007**, *21*, 257–265.
- 13. Costenla, A.R.; Cunha, R.A.; de Mendonça, A. Caffeine, adenosine receptors, and synaptic plasticity. *J. Alzheimers Dis.* **2010**, 20, S25–S34.

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14. Orr, A.G.; Orr, A.L.; Li, X.-J.; Gross, R.E.; Traynelis, S.F. Adenosine A2A receptor mediates microglial process retraction. *Nat. Neurosci.* **2009**, *12*, 872–878.

- 15. Rebola, N.; Simões, A.P.; Canas, P.M.; Tome, A.R.; Andrade, G.M.; Barry, C.E.; Agostinho, P.M.; Lynch, M.A.; Cunha, R.A. Adenosine A2A receptors control neuroinflammation and consequent hippocampal neuronal dysfunction. *J. Neurochem.* **2011**, 117, 100–111.
- 16. Barrett, R.D.; Bennet, L.; Davidson, J.; Dean, J.M.; George, S.; Emerald, B.S.; Gunn, A.J. Destruction and reconstruction: Hypoxia and the developing brain. *Birth Defects Ress. Part C Embryo Today Rev.* **2007**, *81*, 163–176.
- 17. Haynes, R.L.; Sleeper, L.A.; Volpe, J.J.; Kinney, H.C. Neuropathologic Studies of the Encephalopathy of Prematurity in the Late Preterm Infant. *Clin. Perinatol.* **2013**, *40*, 707–722.
- 18. Laptook, A.R. Birth Asphyxia and Hypoxic-Ischemic Brain Injury in the Preterm Infant. Clin. Perinatol. 2016, 43, 529–545.
- 19. Volpe, J.J. Brain injury in premature infants: A complex amalgam of destructive and developmental disturbances. *Lancet Neurol.* **2009**, *8*, 110–124.
- 20. de Vries, L.S.; Groenendaal, F. Patterns of neonatal hypoxic–ischaemic brain injury. Neuroradiology 2010, 52, 555–566.
- 21. Hankins, G.D.V.; Speer, M. Defining the Pathogenesis and Pathophysiology of Neonatal Encephalopathy and Cerebral Palsy. *Obstet. Gynecol.* **2003**, *102*, 628–636.
- 22. Perlman, J.M. White matter injury in the preterm infant: An important determination of abnormal neurodevelopment outcome. *Early Hum. Dev.* **1998**, *53*, 99–120.
- 23. Volpe, J.J. Neonatal encephalitis and white matter injury: More than just inflammation? Ann. Neurol. 2008, 64, 232–236.
- Volpe, J.J. The Encephalopathy of Prematurity Brain Injury and Impaired Brain Development Inextricably Intertwined. Semin. Pediatr. Neurol. 2009, 16, 167–178.
- 25. Volpe, J.J. Neonatal encephalopathy: An inadequate term for hypoxic-ischemic encephalopathy. Ann. Neurol. 2012, 72, 156–166.
- 26. Volpe, J.J. Perinatal brain injury: From pathogenesis to neuroprotection. Ment. Retard. Dev. Disabil. Res. Rev. 2001, 7, 56–64.
- 27. Volpe, J.J. Commentary: Cerebral White Matter Injury of the Premature Infant—More Common Than You Think. *Pediatrics* **2003**, *112*, 176–180.
- 28. Krageloh-Mann, I.; Toft, P.; Lunding, J.; Andresen, J.; Pryds, O.; Lou, H.C. Brain lesions in preterms: Origin, consequences and compensation. *Acta Paediatr.* **1999**, *88*, 897–908.
- 29. Du Plessis, A.J.; Volpe, J.J. Perinatal brain injury in the preterm and term newborn. Curr. Opin. Neurol. 2002, 15, 151–157.
- 30. Mendola, P.; Mumford, S.L.; Männistö, T.I.; Holston, A.; Reddy, U.M.; Laughon, S.K. Controlled Direct Effects of Preeclampsia on Neonatal Health After Accounting for Mediation by Preterm Birth. *Epidemiology* **2015**, 26, 17–26. https://doi.org/10.1097/ede.000000000000213.
- 31. Yanowitz, T.D.; Potter, D.M.; Bowen, A.; Baker, R.W.; Roberts, J.M. Variability in Cerebral Oxygen Delivery Is Reduced in Premature Neonates Exposed to Chorioamnionitis. *Pediatr. Res.* **2006**, *59*, 299–304. https://doi.org/10.1203/01.pdr.0000196738.03171.f1.
- 32. Paton, M.C.B.; McDonald, C.A.; Allison, B.J.; Fahey, M.C.; Jenkin, G.; Miller, S.L. Perinatal Brain Injury as a Consequence of Preterm Birth and Intrauterine Inflammation: Designing Targeted Stem Cell Therapies. *Front. Neurosci.* **2017**, *11*, 200. https://doi.org/10.3389/fnins.2017.00200.
- 33. Rao, R.; Trivedi, S.; Vesoulis, Z.; Liao, S.M.; Smyser, C.D.; Mathur, A.M. Safety and Short-Term Outcomes of Therapeutic Hypothermia in Preterm Neonates 34–35 Weeks Gestational Age with Hypoxic-Ischemic Encephalopathy. *J. Pediatr.* **2016**, *183*, 37–42. https://doi.org/10.1016/j.jpeds.2016.11.019.
- 34. Aarnoudse-Moens, C.S.; Weisglas-Kuperus, N.; Duivevoorden, H.J.; van Goudoever, J.B.; Oosterlaan, J. Executive function and IQ predict mathematical and attention problems in very preterm children. *PLoS ONE* **2013**, *8*, e55994.
- 35. Feldman, H.M.; Janosky, J.E.; Scher, M.S.; Wareham, N.L. Language abilities following prematurity, periventricular brain injury, and cerebral palsy. *J. Commun. Disord.* **1994**, 27, 71–90.
- 36. Aarnoudse-Moens, C.S.H.; Weisglas-Kuperus, N.; van Goudoever, J.B.; Oosterlaan, J. Meta-Analysis of Neurobehavioral Outcomes in Very Preterm and/or Very Low Birth Weight Children. *Pediatrics* **2009**, 124, 717–728.
- 37. Allen, M.C. Neurodevelopmental outcomes of preterm infants. Curr. Opin. Neurol. 2008, 21, 123–128.
- 38. Aylward, G.P. Neurodevelopmental Outcomes of Infants Born Prematurely. J. Dev. Behav. Pediatr. 2005, 26, 427-440.
- 39. Aylward, G.P. Cognitive and neuropsychological outcomes: More than IQ scores. *Ment. Retard. Dev. Disabil. Res. Rev.* **2002**, *8*, 234–240.
- 40. Badawi, N.; Keogh, J.M.; Dixon, G.; Kurinczuk, J.J. Developmental outcomes of newborn encephalopathy in the term infant.. *Indian J. Pediatr.* **2001**, *68*, 527–530.
- 41. Baron, I.S.; Brandt, J.; Ahronovich, M.D.; Baker, R.; Erickson, K.; Litman, F.R. Selective deficit in spatial location memory in extremely low birth weight children at age six: The PETIT Study. *Child Neuropsychol.* **2012**, *18*, 299–311.
- 42. Baron, I.S.; Erickson, K.; Ahronovich, M.D.; Coulehan, K.; Baker, R.; Litman, F.R. Visuospatial and verbal fluency relative deficits in 'complicated' late-preterm preschool children. *Early Hum. Dev.* **2009**, *85*, 751–754.
- Begega, A.; López, M.M.; De Iscar, M.J.; Cuesta-Izquierdo, M.; Solís, G.; Fernández-Colomer, B.; Álvarez, L.; Méndez, M.; Arias, J.L. Assessment of the global intelligence and selective cognitive capacities in preterm 8-year-old children. *Psicothema* 2010, 22, 648–653.
- 44. Bhutta, A.T.; Cleves, M.A.; Casey, P.H.; Cradock, M.M.; Anand, K.J.S. Cognitive and behavioral outcomes of children who were born preterm: A meta-analysis. *JAMA* **2002**, *288*, 728–737.

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45. Cserjesi, R.; Van Braeckel, K.N.; Butcher, P.R.; Kerstjens, J.M.; Reijneveld, S.A.; Bouma, A.; Geuze, R.H.; Bos, A.F. Functioning of 7-Year-Old Children Born at 32 to 35 Weeks' Gestational Age. *Pediatrics* **2012**, *130*, e838–e846.

- 46. Edwards, A.D.; Brocklehurst, P.; Gunn, A.J.; Halliday, H.; Juszczak, E.; Levene, M.; Strohm, B.; Thoresen, M.; Whitelaw, A.; Azzopardi, D. Neurological outcomes at 18 months of age after moderate hypothermia for perinatal hypoxic ischaemic encephalopathy: Synthesis and meta-analysis of injury in term neonatal encephalopathy. *Pediatr. Neurol.* **2010**, *40*, 215–226.
- 47. Fazzi, E.; Bova, S.; Giovenzana, A.; Signorini, S.; Uggetti, C.; Bianchi, P. Cognitive visual dysfunctions in preterm children with periventricular leukomalacia. *Dev. Med. Child Neurol.* **2009**, *51*, 974–981.
- 48. Kent, A.L.; Wright, I.M.; Abdel-Latif, M.E.; New South Wales and Australian Capital Territory Neonatal Intensive Care Units Audit Group. Mortality and adverse neurologic outcomes are greater in preterm male infants. *Pediatrics* **2012**, *129*, 124–131.
- 49. Leversen, K.T.; Sommerfelt, K.; Rønnestad, A.; Kaaresen, P.I.; Farstad, T.; Skranes, J.; Støen, R.; Elgen, I.B.; Rettedal, S.; Eide, G.E.; et al. Prediction of Neurodevelopmental and Sensory Outcome at 5 Years in Norwegian Children Born Extremely Preterm. *Pediatrics* **2011**, 127, e630–e638.
- 50. Perricone, G.; Morales, M.R.; Anzalone, G. Neurodevelopmental outcomes of moderately preterm birth: Precursors of attention deficit hyperactivity disorderat preschool age. *Springerplus* **2013**, *2*, 221.
- 51. Mercuri, E.; Haataja, L.; Guzzetta, A.; Anker, S.; Cowan, F.; Rutherford, M.; Andrew, R.; Braddick, O.; Cioni, G.; Dubowitz, L.; et al. Visual function in term infants with hypoxic-ischaemic insults: Correlation with neurodevelopment at 2 years of age. *Arch. Dis. Child. Fetal Neonatal Ed.* **1999**, *80*, F99–F104.
- 52. Mwaniki, M.K.; Atieno, M.; Lawn, J.E.; Newton, C.R. Long-term neurodevelopmental outcomes after intrauterine and neonatal insults: A systematic review. *Lancet* **2012**, *379*, 445–452.
- Perlman, M.; Shah, P.S. Hypoxic-Ischemic Encephalopathy: Challenges in Outcome and Prediction. J. Pediatr. 2011, 158, e51
 e54
- 54. Vohr, B.R. Neurodevelopmental Outcomes of Extremely Preterm Infants. Clin. Perinatol. 2014, 41, 241–255.
- 55. Edgin, J.O.; Inder, T.E.; Anderson, P.J.; Hood, K.M.; Clark, C.A.; Woodward, L.J. Executive functioning in preschool children born very preterm: Relationship with early white matter pathology. *J. Int. Neuropsychol. Soc.* **2007**, *14*, 90–101.
- 56. Briscoe, J.; Gathercole, S.E.; Marlow, N. Short-Term Memory and Language Outcomes After Extreme Prematurity at Birth. *J. Speech, Lang. Hear. Res.* **1998**, 41, 654–666.
- 57. Casiro, O.G.; Moddemann, D.M.; Stanwick, R.S.; Panikkar-Thiessen, V.K.; Cowan, H.; Cheang, M.S. Language development of very low birth weight infants and fullterm controls at 12 months of age. *Early Hum. Dev.* **1990**, 24, 65–77.
- 58. Jansson-Verkasalo, E.; Korpilahti, P.; Jäntti, V.; Valkama, M.; Vainionpää, L.; Alku, P.; Suominen, K.; Näätänen, R. Neurophysiologic correlates of deficient phonological representations and object naming in prematurely born children. *Clin. Neurophysiol.* **2004**, *115*, 179–187.
- 59. Jansson-Verkasalo, E.; Valkama, M.; Vainionpaa, L.; Paakko, E.; Ilkko, E.; Lehtihalmes, M. Language development in very low birth weight preterm children: A follow-up study. *Folia Phoniatr*. *Et Logop*. **2004**, *56*, 108–119.
- 60. Steinman, K.J.; Gorno-Tempini, M.L.; Glidden, D.V.; Kramer, J.H.; Miller, S.P.; Barkovich, A.J.; Ferriero, D.M. Neonatal Watershed Brain Injury on Magnetic Resonance Imaging Correlates with Verbal IQ at 4 Years. *Pediatrics* **2009**, 123, 1025–1030.
- 61. Downie, A.L.S.; Jakobson, L.S.; Frisk, V.; Ushycky, I. Auditory Temporal Processing Deficits in Children with Periventricular Brain Injury. *Brain Lang.* **2002**, *80*, 208–225.
- 62. Gallo, J.; Dias, K.Z.; Pereira, L.D.; Azevedo, M.F.; Sousa, E.C. Auditory processing evaluation in children born preterm. [Avaliacao do processamento auditivo em criancas nascidas pre-termo]. *J. Soc. Bras. Fonoaudiol.* **2011**, 23, 95–101.
- 63. Ortiz-Mantilla, S.; Choudhury, N.; Leevers, H.; Benasich, A.A. Understanding language and cognitive deficits in very low birth weight children. *Dev. Psychobiol.* **2008**, *50*, 107–126.
- 64. Benasich, A.A.; Tallal, P. Infant discrimination of rapid auditory cues predicts later language impairment. *Behav. Brain Res.* **2002**, 136, 31–49.
- 65. Downie, A.L.S.; Frisk, V.; Jakobson, L.S. The Impact of Periventricular Brain Injury on Reading and Spelling Abilities in the Late Elementary and Adolescent Years. *Child Neuropsychol.* **2005**, *11*, 479–495.
- 66. Benasich, A.A.; Choudhury, N.; Friedman, J.T.; Realpe-Bonilla, T.; Chojnowska, C.; Gou, Z. The infant as a prelinguistic model for language learning impairments: Predicting from event-related potentials to behavior. *Neuropsychologia* **2006**, 44, 396–411.
- 67. Benasich, A.A.; Thomas, J.J.; Choudhury, N.; Leppänen, P. The importance of rapid auditory processing abilities to early language development: Evidence from converging methodologies. *Dev. Psychobiol.* **2002**, 40, 278–292.
- 68. Choudhury, N.; Leppänen, P.H.; Leevers, H.J.; Benasich, A.A. Infant information processing and family history of specific language impairment: Converging evidence for RAP deficits from two paradigms. *Dev. Sci.* **2007**, *10*, 213–236.
- 69. Burnett, A.C.; Scratch, S.E.; Anderson, P.J. Executive function outcome in preterm adolescents. *Early Hum. Dev.* **2013**, *89*, 215–220.
- 70. Espy, K.A.; Stalets, M.M.; McDiarmid, M.M.; Senn, T.E.; Cwik, M.F.; Hamby, A. Executive Functions in Preschool Children Born Preterm: Application of Cognitive Neuroscience Paradigms. *Child Neuropsychol.* **2002**, *8*, 83–92.
- 71. Luu, T.M.; Ment, L.; Allan, W.; Schneider, K.; Vohr, B.R. Executive and Memory Function in Adolescents Born Very Preterm. *Pediatrics* **2011**, 127, e639–e646.
- 72. Curtis, W.J.; Zhuang, J.; Townsend, E.L.; Hu, X.; Nelson, C.A. Memory in Early Adolescents Born Prematurely: A Functional Magnetic Resonance Imaging Investigation. *Dev. Neuropsychol.* **2006**, *29*, 341–377.

73. de Kieviet, J.F.; van Elburg, R.M.; Lafeber, H.N.; Oosterlaan, J. Attention problems of very preterm children compared with agematched term controls at school-age. *J. Pediatr.* **2012**, *161*, 824–829.e1.

- 74. Getahun, D., Rhoads, G.G.; Demissie, K.; Lu, S.E.; Quinn, V.P.; Fassett, M.J.; Wing, D.A.; Jacobsen, S.J. In utero exposure to ischemic-hypoxic conditions and attention-deficit/hyperactivity disorder. *Pediatrics* **2013**, *131*, e53–e61.
- 75. Lindström, K.; Lindblad, F.; Hjern, A. Preterm Birth and Attention-Deficit/Hyperactivity Disorder in Schoolchildren. *Pediatrics* **2011**, 127, 858–865.
- 76. Lou, H.C. Etiology and pathogenesis of attention-deficit hyperactivity disorder (ADHD): Significance of prematurity and perinatal hypoxic-haemodynamic encephalopathy. *Acta Paediatr.* **1996**, *85*, 1266–1271.
- 77. Scott, M.N.; Taylor, H.G.; Fristad, M.A.; Klein, N.; Espy, K.A.; Minich, N.; Hack, M. Behavior Disorders in Extremely Preterm/Extremely Low Birth Weight Children in Kindergarten. *J. Dev. Behav. Pediatr.* **2012**, *33*, 202–213.
- 78. Shum, D.; Neulinger, K.; O'Callaghan, M.; Mohay, H. Attentional problems in children born very preterm or with extremely low birth weight at 7–9 years. *Arch. Clin. Neuropsychol.* **2008**, 23, 103–112.
- 79. Sun, J.; Buys, N. Early executive function deficit in preterm children and its association with neurodevelopmental disorders in childhood: A literature review. *Int. J. Adolesc. Med. Health* **2012**, *24*, 291–299.
- 80. Silbereis, J.C.; Huang, E.; Back, S.A.; Rowitch, D.H. Towards improved animal models of neonatal white matter injury associated with cerebral palsy. *Dis. Model. Mech.* **2010**, *3*, 678–688.
- 81. Jantzie, L.L.; Robinson, S. Preclinical Models of Encephalopathy of Prematurity. Dev. Neurosci. 2015, 37, 277–288.
- 82. Rice, J.E., 3rd; Vannucci, R.C.; Brierley, J.B. The influence of immaturity on hypoxic-ischemic brain damage in the rat. *Ann. Neurol.* **1981**, 9, 131–141.
- 83. Clancy, B.; Darlington, R.; Finlay, B. Translating developmental time across mammalian species. Neuroscience 2001, 105, 7–17.
- 84. Dobbing, J.; Sands, J. Quantitative growth and development of human brain. Arch. Dis. Child. 1973, 48, 757–767.
- 85. Dobbing, J.; Sands, J. Comparative aspects of the brain growth spurt. Early Hum. Dev. 1979, 3, 79–83.
- 86. Semple, B.D.; Blomgren, K.; Gimlin, K.; Ferriero, D.M.; Noble-Haeusslein, L.J. Brain development in rodents and humans: Identifying benchmarks of maturation and vulnerability to injury across species. *Prog. Neurobiol.* **2013**, *106*–*107*, 1–16.
- 87. Workman, A.D.; Charvet, C.J.; Clancy, B.; Darlington, R.B.; Finlay, B.L. Modeling transformations of neurodevelopmental sequences across mammalian species. *J. Neurosci.* **2013**, *33*, 7368–7383.
- 88. Patel, S.D.; Pierce, L.; Ciardiello, A.; Hutton, A.; Paskewitz, S.; Aronowitz, E.; Voss, H.U.; Moore, H.; Vannucci, S.J. Therapeutic hypothermia and hypoxia–ischemia in the term-equivalent neonatal rat: Characterization of a translational preclinical model. *Pediatr. Res.* **2015**, *78*, 264–271.
- 89. Mallard, C.; Vexler, Z.S. Modeling Ischemia in the Immature Brain: How Translational Are Animal Models? *Stroke* **2015**, *46*, 3006–3011. https://doi.org/10.1161/STROKEAHA.115.007776.
- 90. Alexander, M.; Garbus, H.; Smith, A.L.; Rosenkrantz, T.S.; Fitch, R.H. Behavioral and histological outcomes following neonatal HI injury in a preterm (P3) and term (P7) rodent model. *Behav. Brain Res.* **2013**, 259, 85–96.
- 91. Alexander, M.L.; Hill, C.A.; Rosenkrantz, T.; Fitch, R.H. Evaluation of the Therapeutic Benefit of Delayed Administration of Erythropoietin following Early Hypoxic-Ischemic Injury in Rodents. *Dev. Neurosci.* **2012**, *34*, 515–524.
- 92. Alexander, M.L.; Smith, A.; Rosenkrantz, T.; Garbus, H.; Fitch, R.H. The effects of P3 vs P7 HI injury on MGN morphology. *Int. J. Dev. Neurosci.* **2014**, 33, 1–7.
- 93. Alexander, M.L.; Smith, A.L.; Rosenkrantz, T.; Fitch, R.H. Therapeutic effect of caffeine treatment immediately following neonatal hypoxic-ischemic injury on spatial memory in male rats. *Brain Sci.* **2013**, *3*, 177–190.
- 94. McClure, M.M.; Peiffer, A.M.; Rosen, G.D.; Fitch, R.H. Auditory processing deficits in rats with neonatal hypoxic-ischemic injury. *Int. J. Dev. Neurosci.* **2005**, *23*, 351–362.
- 95. McClure, M.M.; Threlkeld, S.W.; Fitch, R.H. The effects of erythropoietin on auditory processing following neonatal hypoxic-ischemic injury. *Brain Res.* **2006**, *1087*, 190–195.
- 96. McClure, M.; Threlkeld, S.; Fitch, R.H. Auditory processing and learning/memory following erythropoietin administration in neonatally HI injured rats. *Brain Res.* **2006**, *1132*, 203–209.
- 97. McClure, M.M.; Threlkeld, S.W.; Rosen, G.D.; Fitch, R.H. Rapid auditory processing and learning deficits in rats with P1 versus P7 neonatal hypoxic-ischemic injury. *Behav. Brain Res.* **2006**, *172*, 114–121.
- 98. McClure, M.M.; Threlkeld, S.W.; Rosen, G.D.; Fitch, R.H. Auditory processing deficits in unilaterally and bilaterally injured hypoxic–ischemic rats. *NeuroReport* **2005**, *16*, 1309–1312.
- McCullough, L.D.; Hurn, P.D. Estrogen and ischemic neuroprotection: An integrated view. Trends Endocrinol. Metab. 2003, 14, 228–235.
- 100. McCullough, L.D.; Zeng, Z.; Blizzard, K.K.; Debchoudhury, I.; Hurn, P.D. Ischemic Nitric Oxide and Poly (ADP-Ribose) Polymerase-1 in Cerebral Ischemia: Male Toxicity, Female Protection. *J. Cereb. Blood Flow Metab.* **2005**, *25*, 502–512.
- 101. Smith, A.L.; Alexander, M.; Rosenkrantz, T.S.; Sadek, M.L.; Fitch, R.H. Sex differences in behavioral outcome following neonatal hypoxia ischemia: Insights from a clinical meta-analysis and a rodent model of induced hypoxic ischemic brain injury. *Exp. Neurol.* **2014**, 254, 54–67.
- 102. Smith, A.L.; Hill, C.A.; Alexander, M.; Szalkowski, C.E.; Chrobak, J.J.; Rosenkrantz, T.S.; Fitch, R.H. Spatial Working Memory Deficits in Male Rats Following Neonatal Hypoxic Ischemic Brain Injury Can Be Attenuated by Task Modifications. *Brain Sci.* **2014**, *4*, 240–272.

103. Smith, A.L.; Alexander, M.; Chrobak, J.J.; Rosenkrantz, T.S.; Fitch, R.H. Dissociation in the Effects of Induced Neonatal Hypoxia-Ischemia on Rapid Auditory Processing and Spatial Working Memory in Male Rats. *Dev. Neurosci.* **2015**, *37*, 440–452.

- 104. Smith, A.L.; Garbus, H.; Rosenkrantz, T.S.; Fitch, R.H. Sex Differences in Behavioral Outcomes Following Temperature Modulation During Induced Neonatal Hypoxic Ischemic Injury in Rats. *Brain Sci.* **2015**, *5*, 220–240.
- 105. Smith, A.L.; Rosenkrantz, T.S.; Fitch, R.H. Neuropathology and neural reorganization following temperature reduction during induced neonatal hypoxic ischemic brain injury in male and female rats. *Neural Plast.* **2016**, 2016, 2585230.
- 106. Fitch, R.H.; Alexander, M.L.; Threlkeld, S.W. Early neural disruption and auditory processing outcomes in rodent models: Implications for developmental language disability. *Front. Syst. Neurosci.* **2013**, 7, 58.
- 107. Fitch, R.H.; Threlkeld, S.W.; McClure, M.M.; Peiffer, A.M. Use of a modified prepulse inhibition paradigm to assess complex auditory discrimination in rodents. *Brain Res. Bull.* **2008**, *76*, 1–7.
- 108. Peiffer, A.M.; Dunleavy, C.K.; Frenkel, M.; Gabel, L.A.; LoTurco, J.J.; Rosen, G.D.; Fitch, R.H. Impaired detection of variable duration embedded tones in ectopic NZB/BINJ mice. *NeuroReport* **2001**, *12*, 2875–2879.
- 109. Peiffer, A.M.; Rosen, G.D.; Fitch, R.H. Sex differences in rapid auditory processing deficits in ectopic BXSB/MpJ mice. *NeuroReport* **2002**, *13*, 2277–2280.
- 110. Peiffer, A.M.; Rosen, G.D.; Fitch, R. Sex differences in rapid auditory processing deficits in microgyric rats. *Dev. Brain Res.* **2004**, 148, 53–57.
- 111. Threlkeld, S.W.; McClure, M.M.; Bai, J.; Wang, Y.; LoTurco, J.J.; Rosen, G.D.; Fitch, R.H. Developmental disruptions and behavioral impairments in rats following in utero RNAi of Dyx1c1. *Brain Res. Bull.* 2007, 71, 508–514. https://doi.org/10.1016/j.brainres-bull.2006.11.005.
- 112. Threlkeld, S.W.; McClure, M.M.; Rosen, G.D.; Fitch, R.H. Developmental timeframes for induction of microgyria and rapid auditory processing deficits in the rat. *Brain Res.* **2006**, *1109*, 22–31. https://doi.org/10.1016/j.brainres.2006.06.022.
- 113. Balduini, W.; De Angelis, V.; Mazzoni, E.; Cimino, M. Long-lasting behavioral alterations following a hypoxic/ischemic brain injury in neonatal rats. *Brain Res.* **2000**, *859*, 318–325.
- 114. Ikeda, T.; Mishima, K.; Yoshikawa, T.; Iwasaki, K.; Fujiwara, M.; Xia, Y.X.; Ikenoue, T. Selective and long-term learning impairment following neonatal hypoxic-ischemic brain insult in rats. *Behav. Brain Res.* **2001**, *118*, 17–25.
- 115. Arteni, N.S.; Salgueiro, J.; Torres, I.; Achaval, M.; Netto, C.A. Neonatal cerebral hypoxia–ischemia causes lateralized memory impairments in the adult rat. *Brain Res.* **2003**, *973*, 171–178.
- 116. Hill, C.A.; Threlkeld, S.W.; Fitch, R.H. Early testosterone modulated sex differences in behavioral outcome following neonatal hypoxia ischemia in rats. *Int. J. Dev. Neurosci.* **2011**, *29*, 381–388.
- 117. Young, R.S.; Kolonich, J.; Woods, C.L.; Yagel, S.K. Behavioral performance of rats following neonatal hypoxia-ischemia.. *Stroke* **1986**, *17*, 1313–1316.
- 118. Mishima, K.; Ikeda, T.; Yoshikawa, T.; Aoo, N.; Egashira, N.; Xia, Y.X.; Ikenoue, T.; Iwasaki, K.; Fujiwara, M. Effects of hypothermia and hyperthermia on attentional and spatial learning deficits following neonatal hypoxia-ischemic insult in rats. *Behav. Brain Res.* **2004**, *151*, 209–217.
- 119. Arteni, N.S.; Pereira, L.O.; Rodrigues, A.L.; Lavinsky, D.; Achaval, M.E.; Netto, C.A. Lateralized and sex-dependent behavioral and morphological effects of unilateral neonatal cerebral hypoxia-ischemia in the rat. *Behav. Brain Res.* **2010**, 210, 92–98.
- 120. Burchell, S.R.; Dixon, B.J.; Tang, J.; Zhang, J.H. Isoflurane Provides Neuroprotection in Neonatal Hypoxic Ischemic Brain Injury. J. Investig. Med. 2013, 61, 1078–1083.
- 121. Jiang, H.; Lei, J.-J.; Zhang, Y.-H. Protective effect of topiramate on hypoxic-ischemic brain injury in neonatal rat. *Asian Pac. J. Trop. Med.* **2014**, *7*, 496–500.
- 122. Juul, S.E.; Ferriero, D.M. Pharmacologic Neuroprotective Strategies in Neonatal Brain Injury. Clin. Perinatol. 2013, 41, 119–131.
- 123. Li, X.; Zhang, J.; Chai, S.; Wang, X. Progesterone alleviates hypoxic-ischemic brain injury via the Akt/GSK-3β signaling pathway. *Exp. Ther. Med.* **2014**, *8*, 1241–1246.
- 124. Ozyener, F.; Çetinkaya, M.; Alkan, T.; Gören, B.; Kafa, I.M.; Kurt, M.A.; Koksal, N. Neuroprotective effects of melatonin administered alone or in combination with topiramate in neonatal hypoxic-ischemic rat model. *Restor. Neurol. Neurosci.* **2012**, *30*, 435–444.
- 125. Palmer, C.; Towfighi, J.; Roberts, R.L.; Heitjan, D.F. Allopurinol administered after inducing hypoxia-ischemia reduces brain injury in 7-day-old rats. *Pediatr. Res.* **1993**, *33*, 405–411.
- 126. van de Looij, Y.; Chatagner, A.; Quairiaux, C.; Gruetter, R.; Huppi, P.S.; Sizonenko, S.V. Multi-Modal Assessment of Long-Term Erythropoietin Treatment after Neonatal Hypoxic-Ischemic Injury in Rat Brain. *PLoS ONE* **2014**, *9*, e95643.
- 127. Villapol, S.; Fau, S.; Renolleau, S.; Biran, V.; Charriaut-Marlangue, C.; Baud, O. Melatonin Promotes Myelination by Decreasing White Matter Inflammation After Neonatal Stroke. *Pediatr. Res.* **2011**, *69*, 51–55.
- 128. Wen, T.C.; Rogido, M.; Peng, H.; Genetta, T.; Moore, J.; Sola, A. Gender differences in long-term beneficial effects of erythropoietin given after neonatal stroke in postnatal day-7 rats. *Neuroscience* **2006**, *139*, 803–811.
- 129. Wood, T.; Osredkar, D.; Puchades, M.; Maes, E.; Falck, M.; Flatebø, T.; Walløe, L.; Sabir, H.; Thoresen, M. Treatment temperature and insult severity influence the neuroprotective effects of therapeutic hypothermia. *Nature* **2016**, *6*, 1–20.
- 130. Xie, C.; Zhou, K.; Wang, X.; Blomgren, K.; Zhu, C. Therapeutic Benefits of Delayed Lithium Administration in the Neonatal Rat after Cerebral Hypoxia-Ischemia. *PLoS ONE* **2014**, *9*, e107192.

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131. Yang, T.; Zhuang, L.; Fidalgo, A.M.R.; Petrides, E.; Terrando, N.; Wu, X.; Sanders, R.D.; Robertson, N.J.; Johnson, M.R.; Maze, M.; et al. Xenon and Sevoflurane Provide Analgesia during Labor and Fetal Brain Protection in a Perinatal Rat Model of Hypoxia-Ischemia. *PLoS ONE* **2012**, *7*, e37020.

- 132. Zhao, P.; Ji, G.; Xue, H.; Yu, W.; Zhao, X.; Ding, M.; Yang, Y.; Zuo, Z. Isoflurane postconditioning improved long-term neurological outcome possibly via inhibiting the mitochondrial permeability transition pore in neonatal rats after brain hypoxia-ischemia. *Neuroscience* **2014**, *280*, 193–203.
- 133. Back, S.A.; Riddle, A.; Dean, J.; Hohimer, A.R. The Instrumented Fetal Sheep as a Model of Cerebral White Matter Injury in the Premature Infant. *Neurotherapeutics* **2012**, *9*, 359–370.
- 134. Davidson, J.O.; Fraser, M.; Naylor, A.S.; Roelfsema, V.; Gunn, A.J.; Bennet, L. The effect of cerebral hypothermia on cortisol and ACTH responses after umbilical cord occlusion in preterm fetal sheep. *Pediatr. Res.* **2008**, *63*, 51e5.
- 135. Koome, M.E.; Davidson, J.O.; Drury, P.P.; Mathai, S.; Booth, L.C.; Gunn, A.J.; Bennet, L. Antenatal Dexamethasone after Asphyxia Increases Neural Injury in Preterm Fetal Sheep. *PLoS ONE* **2013**, *8*, e77480.
- 136. Welin, A.-K.; Svedin, P.; Lapatto, R.; Sultan, B.; Hagberg, H.; Gressens, P.; Kjellmer, I.; Mallard, C. Melatonin Reduces Inflammation and Cell Death in White Matter in the Mid-Gestation Fetal Sheep Following Umbilical Cord Occlusion. *Pediatr. Res.* **2007**, *61*, 153–158.
- 137. Huang, Z.; Liu, J.; Cheung, P.-Y.; Chen, C. Long-term cognitive impairment and myelination deficiency in a rat model of perinatal hypoxic-ischemic brain injury. *Brain Res.* **2009**, *1301*, 100–109.
- 138. Carty, M.L.; Wixey, J.A.; Colditz, P.B.; Buller, K.M. Post-insult minocycline treatment attenuates hypoxia-ischemia-induced neuroinflammation and white matter injury in the neonatal rat: A comparison of two different dose regimens. *Int. J. Dev. Neurosci.* 2008, 26, 477–485.
- 139. Li, W.-J.; Mao, F.-X.; Chen, H.-J.; Qian, L.-H.; Buzby, J.S. Treatment with UDP-glucose, GDNF, and memantine promotes SVZ and white matter self-repair by endogenous glial progenitor cells in neonatal rats with ischemic PVL. *Neuroscience* **2015**, 284, 444–458.
- Nuñez, J.L.; McCarthy, M.M. Estradiol Exacerbates Hippocampal Damage in a Model of Preterm Infant Brain Injury. Endocrinology 2003, 144, 2350–2359.
- 141. Ma, H.; Sinha, B.; Pandya, R.S.; Lin, N.; Popp, A.J.; Li, J.; Yao, J.; Wang, X. Therapeutic Hypothermia as a Neuroprotective Strategy in Neonatal Hypoxic-Ischemic Brain Injury and Traumatic Brain Injury. *Curr. Mol. Med.* **2012**, *12*, 1282–1296.
- 142. RCFN (Report of the Committee on Fetus & Newborn). Hypothermia & neonatal encephalopathy. *Pediatrics* **2014**, 133, 1146–1150.
- 143. Guillet, R.; Edwards, A.D.; Thoresen, M.; Ferriero, D.M.; Gluckman, P.D.; Whitelaw, A.; Gunn, A.J. Seven- to eight-year follow-up of the CoolCap trial of head cooling for neonatal encephalopathy. *Pediatr. Res.* **2011**, *71*, 205–209.
- 144. Shankaran, S.; Laptook, A.R.; Pappas, A.; McDonald, S.A.; Das, A.; Tyson, J.E.; Poindexter, B.B.; Schibler, K.; Bell, E.F.; Heyne, R.J.; et al. Effect of depth and duration of cooling on deaths in the NICU among neonates with hypoxic ischemic encephalopathy: A randomized clinical trial. *JAMA* **2014**, *312*, 2629–2639.
- 145. Shankaran, S. Outcomes of Hypoxic-Ischemic Encephalopathy in Neonates Treated with Hypothermia. *Clin. Perinatol.* **2014**, *41*, 149–159.
- 146. Laptook, A.R. Therapeutic Hypothermia for Preterm Infants with Hypoxic-Ischemic Encephalopathy: How Do We Move Forward? *J. Pediatr.* **2017**, *183*, 8–9.
- 147. Laura, F.; Mori, A.; Tataranno, M.L.; Muraca, M.C.; Rodriquez, D.C.; Giomi, S.; Coviello, C.; Buonocore, G. Therapeutic hypothermia in a late preterm infant. *J. Matern. Neonatal Med.* **2012**, 25, 125–127.
- 148. Shankaran, S. Hypoxic-ischemic Encephalopathy and Novel Strategies for Neuroprotection. Clin. Perinatol. 2012, 39, 919–929.
- 149. Potter, M.; Rosenkrantz, T.; Fitch, R.H. Behavioral and neuroanatomical outcomes in a rat model of preterm hypoxic-ischemic brain Injury: Effects of caffeine and hypothermia. *Int. J. Dev. Neurosci.* **2018**, 70, 46–55.
- 150. Juul, S.E.; Mayock, D.E.; Comstock, B.A.; Heagerty, P.J. Neuroprotective potential of erythropoietin in neonates; design of a randomized trial. *Matern. Health Neonatol. Perinatol.* **2015**, *1*, 27.
- 151. Atik, A.; Harding, R.; De Matteo, R.; Kondos-Devcic, D.; Cheong, J.; Doyle, L.W.; Tolcos, M. Caffeine for apnea of prematurity: Effects on the developing brain. *NeuroToxicology* **2017**, *58*, 94–102.
- 152. Dobson, N.R.; Patel, R.M.; Smith, P.B.; Kuehn, D.R.; Clark, J.; Vyas-Read, S.; Herring, A.; Laughon, M.M.; Carlton, D.; Hunt, C.E. Trends in Caffeine Use and Association between Clinical Outcomes and Timing of Therapy in Very Low Birth Weight Infants. *J. Pediatr.* **2014**, *164*, 992–998.
- 153. Kua, K.P.; Lee, S.W.H. Systematic review and meta-analysis of clinical outcomes of early caffeine therapy in preterm neonates. *Br. J. Clin. Pharmacol.* **2016**, *83*, 180–191.
- 154. Lodha, A.; Seshia, M.; McMillan, D.D.; Barrington, K.; Yang, J.; Lee, S.K.; Shah, P.S.; Canadian Neonatal Network. Association of Early Caffeine Administration and Neonatal Outcomes in Very Preterm Neonates. *JAMA Pediatr.* **2015**, *169*, 33–38.
- 155. Patel, R.M.; Leong, T.; Carlton, D.P.; Vyas-Read, S. Early caffeine therapy and clinical outcomes in extremely preterm infants. *J. Perinatol.* **2013**, *33*, 134–140.
- 156. Schmidt, B.; Roberts, R.S.; Davis, P.; Doyle, L.W.; Barrington, K.J.; Ohlsson, A.; Solimano, A.; Tin, W.; Caffeine for Apnea of Prematurity Trial Group. Long-Term Effects of Caffeine Therapy for Apnea of Prematurity. N. Engl. J. Med. 2007, 357, 1893–1902.

157. Schmidt, B.; Anderson, P.J.; Doyle, L.W.; Dewey, D.; Grunau, R.E.; Asztalos, E.V.; Davis, P.G.; Tin, W.; Moddemann, D.; Solimano, A.; et al. Survival without disability to age 5 years after neonatal caffeine therapy for apnea of prematurity. *JAMA* **2012**, 307, 275–282.

- 158. Juárez-Méndez, S.; Carretero, R.; Martínez-Tellez, R.; Silva-Gómez, A.B.; Flores, G. Neonatal caffeine administration causes a permanent increase in the dendritic length of prefrontal cortical neurons of rats. *Synapse* **2006**, *60*, 450–455.
- 159. Guillet, R.; Dunham, L. Neonatal Caffeine Exposure and Seizure Susceptibility in Adult Rats. Epilepsia 1995, 36, 743-749.
- 160. Back, S.A.; Craig, A.; Luo, N.L.; Ren, J.; Akundi, R.; Ribeiro, I.; Rivkees, S.A. Protective effects of caffeine on chronic hypoxia-induced perinatal white matter injury. *Ann. Neurol.* **2006**, *60*, 696–705.
- 161. Bona, E.; Adén, U.; Fredholm, B.B.; Hagberg, H. The effect of long term caffeine treatment on hypoxic-ischemic brain damage in the neonate. *Pediatr. Res.* **1995**, *38*, 312–318.
- 162. Winerdal, M.; Urmaliya, V.; Winerdal, M.E.; Fredholm, B.B.; Winqvist, U.A. Single dose caffeine protects the neonatal mouse brain against hypoxia-ischemia. *PLoS ONE* **2017**, *12*, e0170545.
- 163. Hagberg, H.; Gressens, P.; Mallard, C. Inflammation during fetal and neonatal life: Implications for neurologic and neuropsychiatric disease in children and adults. *Ann. Neurol.* **2011**, *71*, 444–457.
- 164. McLeod, R.; Rosenkrantz, T.; Fitch, R.H. Effects of caffeine on neonatal hypoxia ischemia outcomes in females. In Proceedings of the Society for Neuroscience, San Diego, CA, USA, 3–7 November 2018.
- 165. Mcleod, R.; Koski, R.; Rosenkrantz, T.; Fitch, H.R. Sex differences in the protective mechanism of action of caffeine on microglia in a rodent model of preterm hypoxic ischemic injury. In Proceedings of the Society for Neuroscience, Chicago, IL, USA, 8–11 November 2021.
- 166. Fan, X.; Kavelaars, A.; Heijnen, C.J.; Groenendaal, F.; Van Bel, F. Pharmacological Neuroprotection after Perinatal Hypoxic-Ischemic Brain Injury. *Curr. Neuropharmacol.* **2010**, *8*, 324–334.
- 167. Fan, X.; Van Bel, F.; Van Der Kooij, M.A.; Heijnen, C.J.; Groenendaal, F. Hypothermia and erythropoietin for neuroprotection after neonatal brain damage. *Pediatr. Res.* **2013**, *73*, 18–23.
- 168. Hill, C.A.; Alexander, M.L.; McCullough, L.D.; Fitch, R.H. Inhibition of X-Linked Inhibitor of Apoptosis with Embelin Differentially Affects Male versus Female Behavioral Outcome following Neonatal Hypoxia-Ischemia in Rats. *Dev. Neurosci.* **2011**, *33*, 494–504.
- 169. Golomb, M.R.; Fullerton, H.J.; Nowak-Gottl, U.; Deveber, G.; International Pediatric Stroke Study Group. Male predominance in childhood ischemic stroke: Findings from the international pediatric stroke study. *Stroke* **2009**, *40*, 52–57.
- 170. Golomb, M.R.; Zimmer, J.A.; Garg, B.P. Age-related variation in the presentation of childhood stroke varies with inclusion criteria. *Acta Paediatr.* **2009**, *99*, 6.
- 171. Lauterbach, M.D.; Raz, S.; Sander, C.J. Neonatal hypoxic risk in preterm birth infants: The influence of sex and severity of respiratory distress on cognitive recovery. *Neuropsychology* **2001**, *15*, 411–420.
- 172. Mayoral, S.R.; Omar, G.; Penn, A.A. Sex Differences in a Hypoxia Model of Preterm Brain Damage. *Pediatr. Res.* **2009**, *66*, 248–253.
- 173. Peacock, J.L.; Marston, L.; Marlow, N.; Calvert, S.A.; Greenough, A. Neonatal and infant outcome in boys and girls born very prematurely. *Pediatr. Res.* **2012**, *71*, 305–310.
- 174. Raz, S.; Debastos, A.K.; Newman, J.B.; Batton, D. Extreme prematurity and neuropsychological outcome in the preschool years. *J. Int. Neuropsychol. Soc.* **2009**, *16*, 169–179.
- 175. Raz, S.; Lauterbach, M.D.; Hopkins, T.L.; Glogowski, B.K.; Porter, C.L.; Riggs, W.; Sander, C.J. A female advantage in cognitive recovery from early cerebral insult. *Dev. Psychol.* **1995**, *31*, 958–966.
- 176. Benavides, A.; Metzger, A.; Tereshchenko, A.; Conrad, A.; Bell, E.F.; Spencer, J.; Ross-Sheehy, S.; Georgieff, M.; Magnotta, V.; Nopoulos, P.; et al. Sex specific outcomes in the preterm brain. *Pediatr. Res.* **2018**, *85*, 55–62.
- 177. Donders, J.; Hoffman, N.M. Gender differences in learning and memory after pediatric traumatic brain injury. *Neuropsychology* **2002**, *16*, 491–499.
- 178. Gualtieri, C.T.; Ondrusek, M.G.; Finley, C. Attention deficit disorders in adults. Clin. Neuropharmacol. 1985, 8, 343–356.
- 179. Rutter, M.; Caspi, A.; Moffitt, T. Using sex differences in psychopathology to study causal mechanisms: Unifying issues and research strategies. *J. Child Psychol. Psychiatry* **2003**, *44*, 1092–1115.
- 180. Charriaut-Marlangue, C.; Besson, V.C.; Baud, O. Sexually Dimorphic Outcomes after Neonatal Stroke and Hypoxia-Ischemia. *Int. J. Mol. Sci.* **2017**, *19*, 61.
- 181. Hindmarsh, G.J.; O'Callaghan, M.J.; Mohay, H.A.; Rogers, Y.M. Gender differences in cognitive abilities at 2 years in ELBW infants. *Early Hum. Dev.* **2000**, *60*, 115–122.
- 182. Kesler, S.R.; Reiss, A.L.; Vohr, B.; Watson, C.; Schneider, K.C.; Katz, K.H.; Maller-Kesselman, J.; Silbereis, J.; Constable, R.T.; Makuch, R.W.; et al. Brain Volume Reductions within Multiple Cognitive Systems in Male Preterm Children at Age Twelve. *J. Pediatr.* 2008, 152, 513–520.e1.
- 183. Sanches, E.F.; Arteni, N.; Nicola, F.; Aristimunha, D.; Netto, C.A. Sexual dimorphism and brain lateralization impact behavioral and histological outcomes following hypoxia–ischemia in P3 and P7 rats. *Neuroscience* **2015**, 290, 581–593.
- 184. Morsing, E.; Åsard, M.; Ley, D.; Stjernqvist, K.; Maršál, K. Cognitive Function After Intrauterine Growth Restriction and Very Preterm Birth. *Pediatrics* **2011**, 127, e874–e882.
- 185. Wallace, I.F.; Rose, S.A.; McCarton, C.M.; Kurtzberg, D.; Vaughan, H.G., Jr. Relations between infant neurobehavioral performance and cognitive outcome in very low birth weight preterm infants. *J. Dev. Behav.* **1995**, *16*, 309–317.

Life 2022, 12, 1514 16 of 16

186. Carwile, E.; Wagner, A.K.; Crago, E.; Alexander, S.A. Estrogen and stroke: A review of the current literature. *J. Neurosci. Nurs.* **2009**, *41*, 18–25.

- 187. McCarthy, M.M. Molecular aspects of sexual differentiation of the rodent brain. Psychoneuroendocrinology 1994, 19, 415–427.
- 188. McCarthy, M.M. Estradiol and the Developing Brain. Physiol. Rev. 2008, 88, 91-134.
- 189. McCarthy, M.M.; Arnold, A.P.; Ball, G.F.; Blaustein, J.D.; De Vries, G.J. Sex differences in the brain: The not so inconvenient truth. *J. Neurosci.* **2012**, 32, 2241–2247.
- 190. Liu, F.; Li, Z.; Li, J.; Siegel, C.; Yuan, R.; McCullough, L.D. Sex differences in caspase activation after experimental stroke. *Stroke* **2009**, *40*, 1842–1848.
- 191. Manwani, B.; McCullough, L.D. Sexual Dimorphism in Ischemic Stroke: Lessons from the Laboratory. *Women's Health* **2011**, 7, 319–339.
- 192. Nijboer, C.H.; Kavelaars, A.; van Bel, F.; Heijnen, C.J.; Groenendaal, F. Gender-Dependent Pathways of Hypoxia-Ischemia-Induced Cell Death and Neuroprotection in the Immature P3 Rat. *Dev. Neurosci.* **2007**, *29*, 385–392.
- 193. Renolleau, S.; Fau, S.; Charriaut-Marlangue, C. Gender-Related Differences in Apoptotic Pathways After Neonatal Cerebral Ischemia. *Neuroscientist* **2007**, *14*, 46–52. https://doi.org/10.1177/1073858407308889.
- 194. O'Driscoll, D.N.; Greene, C.; Molloy, E.J. Immune function? A missing link in the gender disparity in preterm neonatal outcomes. *Expert Rev. Clin. Immunol.* **2017**, *13*, 1061–1071.
- 195. Waddell, J.; Hanscom, M.; Edwards, N.S.; McKenna, M.C.; McCarthy, M.M. Sex differences in cell genesis, hippocampal volume and behavioral outcomes in a rat model of neonatal HI. *Exp. Neurol.* **2016**, *275*, 285–295.
- 196. Ment, L.R.; Oh, W.; Ehrenkranz, R.A.; Philip, A.G.; Vohr, B.; Allan, W.; Duncan, C.C.; Scott, D.T.; Taylor, K.J.; Katz, K.H. Lowdose indomethacin and prevention of intraventricular hemorrhage: A multicenter randomized trial. *Pediatrics* **1994**, *93*, 543–550.
- 197. Ment, L.R.; Vohr, B.R.; Makuch, R.W.; Westerveld, M.; Katz, K.H.; Schneider, K.C.; Duncan, C.C.; Ehrenkranz, R.; Oh, W.; Philip, A.G.; et al. Prevention of intraventricular hemorrhage by indomethacin in male preterm infants. *J. Pediatr.* **2004**, *145*, 832–834.
- 198. Feng, E.; Jiang, L. Peptidomic analysis of hippocampal tissue for explore leptin neuroprotective effect on the preterm ischemia-hypoxia brain damage model rats. *Neuropharmacology* **2019**, *162*, 107803.
- 199. Poupon-Bejuit, L.; Rocha-Ferreira, E.; Thornton, C.; Hagberg, H.; Rahim, A.A. Neuroprotective Effects of Diabetes Drugs for the Treatment of Neonatal Hypoxia-Ischemia Encephalopathy. *Front. Cell. Neurosci.* **2020**, 14, 112. https://doi.org/10.3389/fncel.2020.00112.
- 200. Rocha-Ferreira, E.; Poupon, L.; Zelco, A.; Leverin, A.-L.; Nair, S.; Jonsdotter, A.; Carlsson, Y.; Thornton, C.; Hagberg, H.; Rahim, A.A. Neuroprotective exendin-4 enhances hypothermia therapy in a model of hypoxic-ischaemic encephalopathy. *Brain* **2018**, 141, 2925–2942. https://doi.org/10.1093/brain/awy220.