

MNV-1 CW1: MS1 raw spectra (10 eV)

A) P monomer and dimer, B) + 250 μ M blood group B type-1 tetrasaccharide (HBGA)

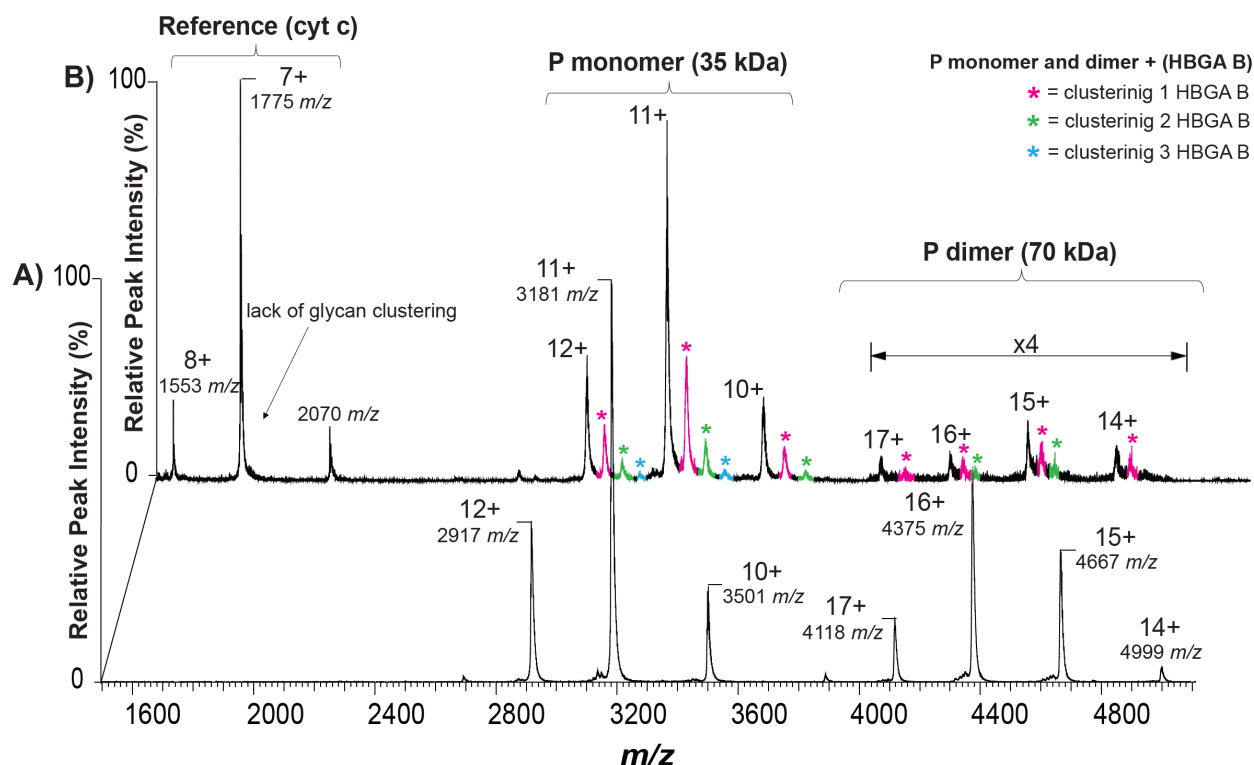


Figure S1 glycan clustering on murine norovirus MNV-1 (CW1) P domain. **A)** Native mass spectra of the P monomer and P dimer are shown, with a monomeric charge state distribution from 12+ to 10+ and a mass of 35 kDa and a dimeric charge state distribution of 17+ to 14+ with a mass of 70 kDa. **B)** Native mass spectra of cytochrome c (cyt c, 11 μ M) with the P monomer (4 μ M) and dimer (four times magnification) in presence of HBGA B clustering. HBGA B ligand concentration (250 μ M) in 250 mM ammonium acetate solution at pH 7.0).

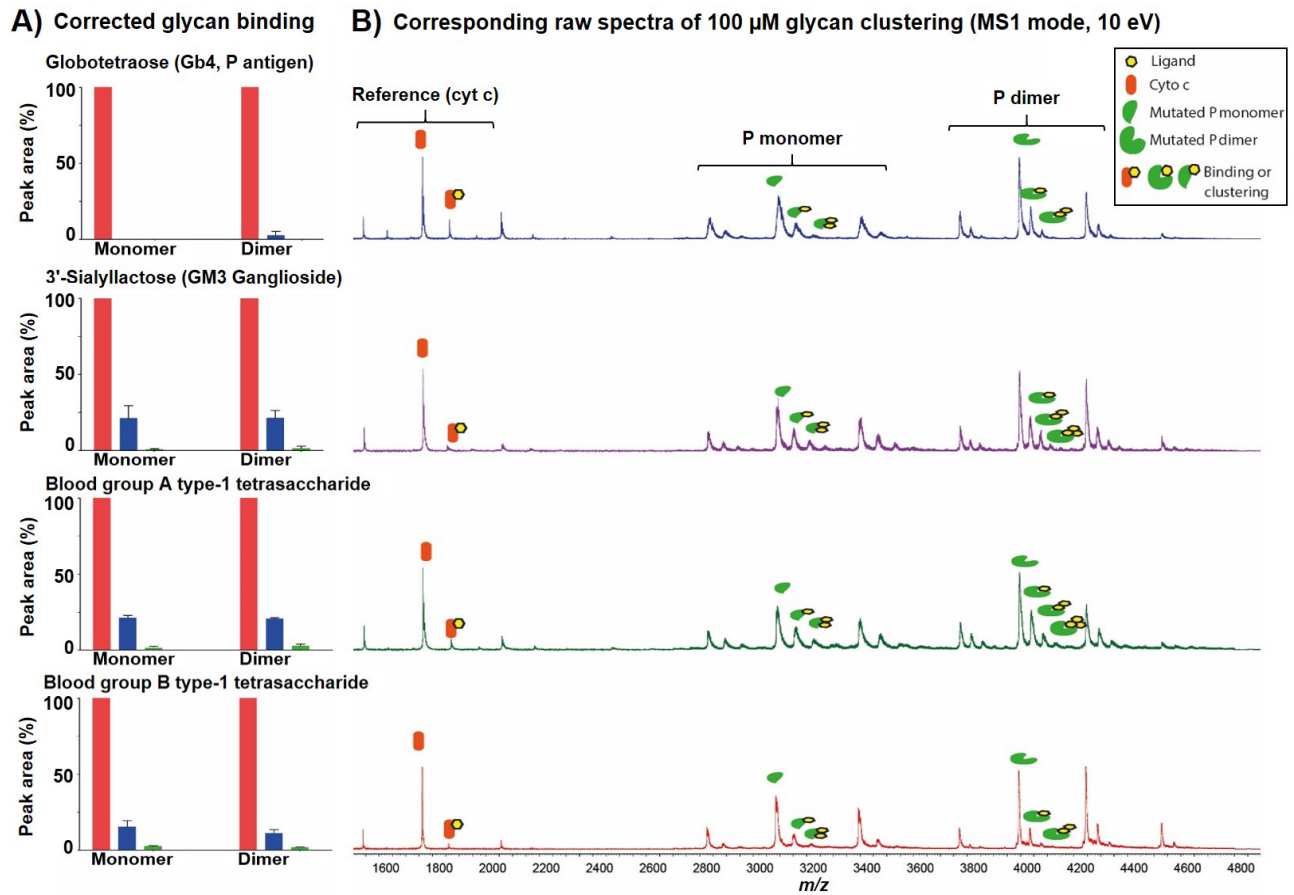
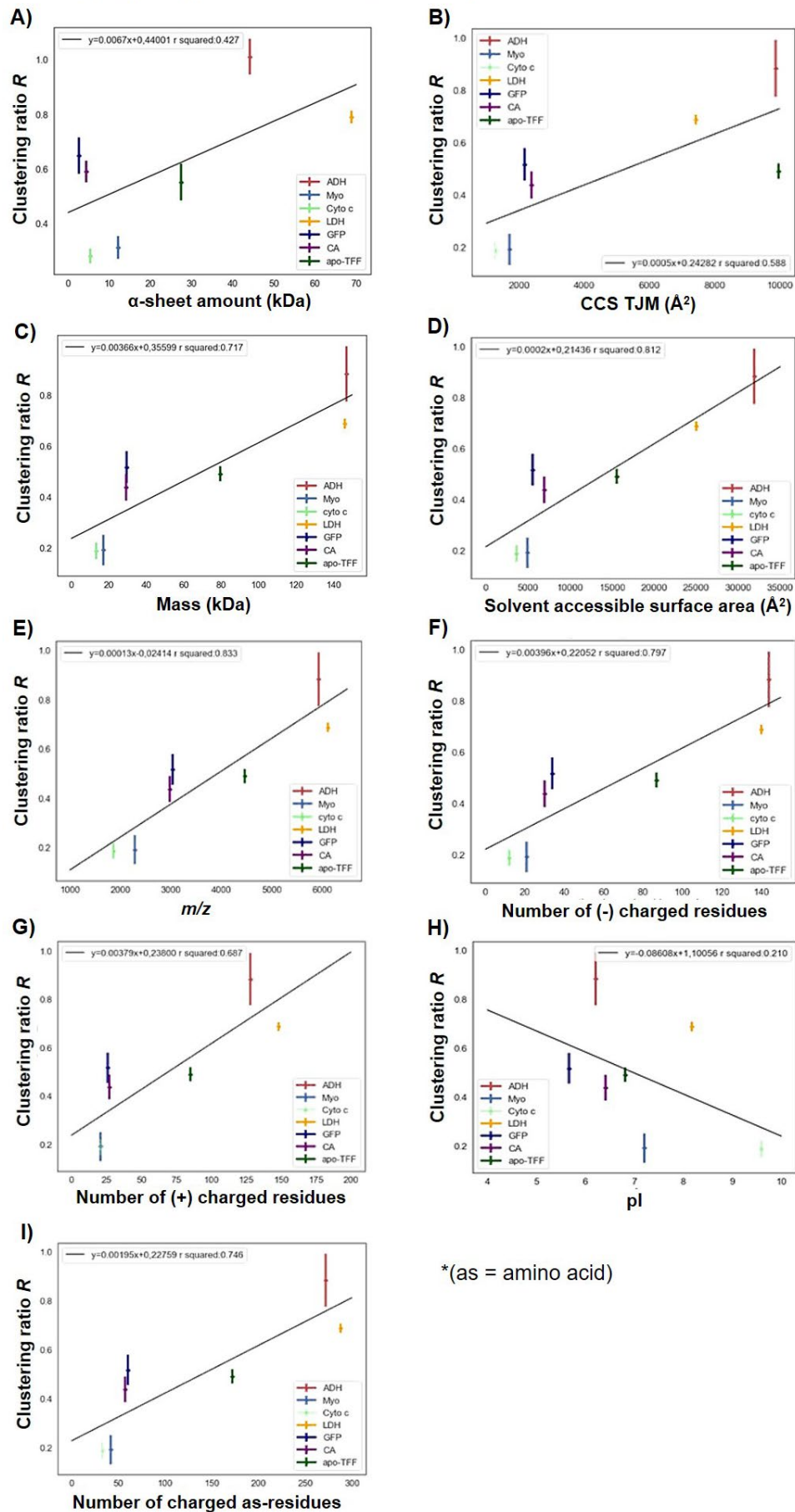


Figure S2 glycan clustering on mutated hNoV GII.4 MI001 strain P domain. **A)** The corrected binding of Gb4, GM3, blood group A and B type-1 tetrasaccharides (HBGA A, HBGA B) to mutated hNoV MI001 P dimer is shown, respectively in 100 μ M glycan concentration. **B)** Native mass spectra of cytochrome c (cyt c) with mutated hNoV MI001 P dimer in presence of different ligand Gb4 (dark blue), GM3 (purple), HBGA A (green), HBGA B (red) clustering (150 mM ammonium acetate solution at pH7). Signal intensity was normalized to the base peak in the spectra.

Blood group B type-1 tetrasaccharide 300 μ M



*(as = amino acid)

Figure S3. Correlation of clustering ratios to multiple protein property parameters. The correlation of different physiochemical parameters of protein (**A**) absolute share of α -helix in a protein, **B**) CCS TJM, **C**) protein mass (kDa), **D**) solvent accessible area, **E**) mass to charge ratio (m/z), **F**) number of negative charged residues, **G**) number of positive charged residues, **H**) pI, **I**) number of charged residues) to unspecific glycan clustering ratios R of seven reference proteins (ADH, CAII, cyt c, GFP, Myo. LDH, apo-TFF) at a blood group B type-1 tetrasaccharide (HBGA B) concentration of 300 μ M was analysed. Native MS experiments were performed with a fixed concentration of P dimer (1 μ M) and reference proteins (3 μ M) at 150 mM ammonium acetate, pH 7. The reference protein structures and protein sequence information is derived from corresponding PDB files (see Table 1). The black line represents the linear regression.

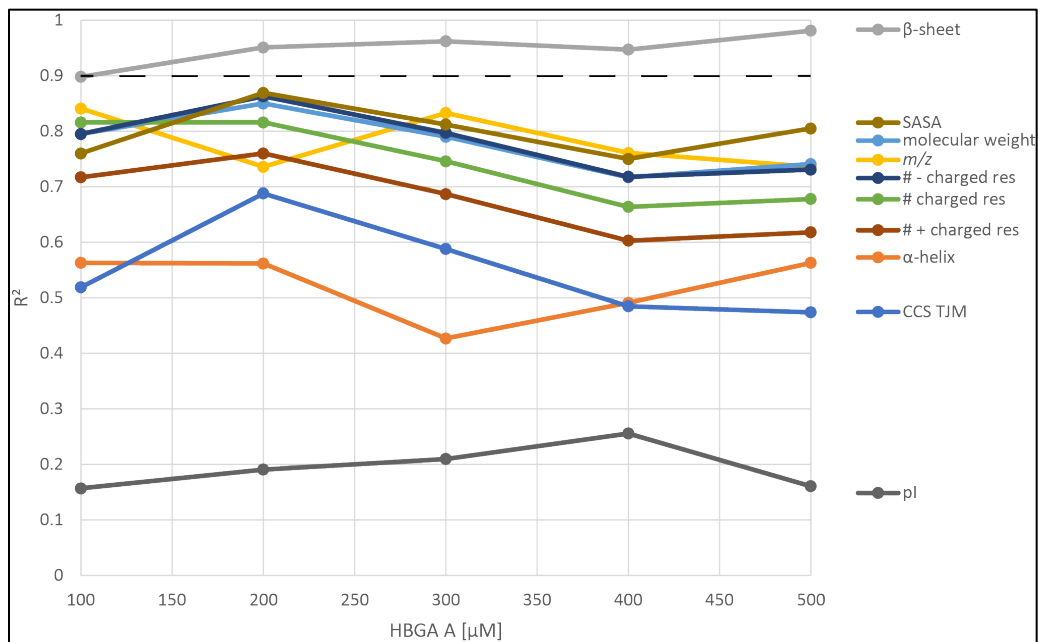


Figure S4. Correlation plotted as R^2 of clustering ratios R to different protein properties over employed HBGA A concentration. R^2 of the different parameters (β -sheet amount in kDa, of α -helix amount in kDa, charged residues, molecular weight of the protein, m/z) obtained for correlations at indicated HBGA A concentrations for the seven reference proteins. This is an alternative representation of the data displayed in Fig. 4A.

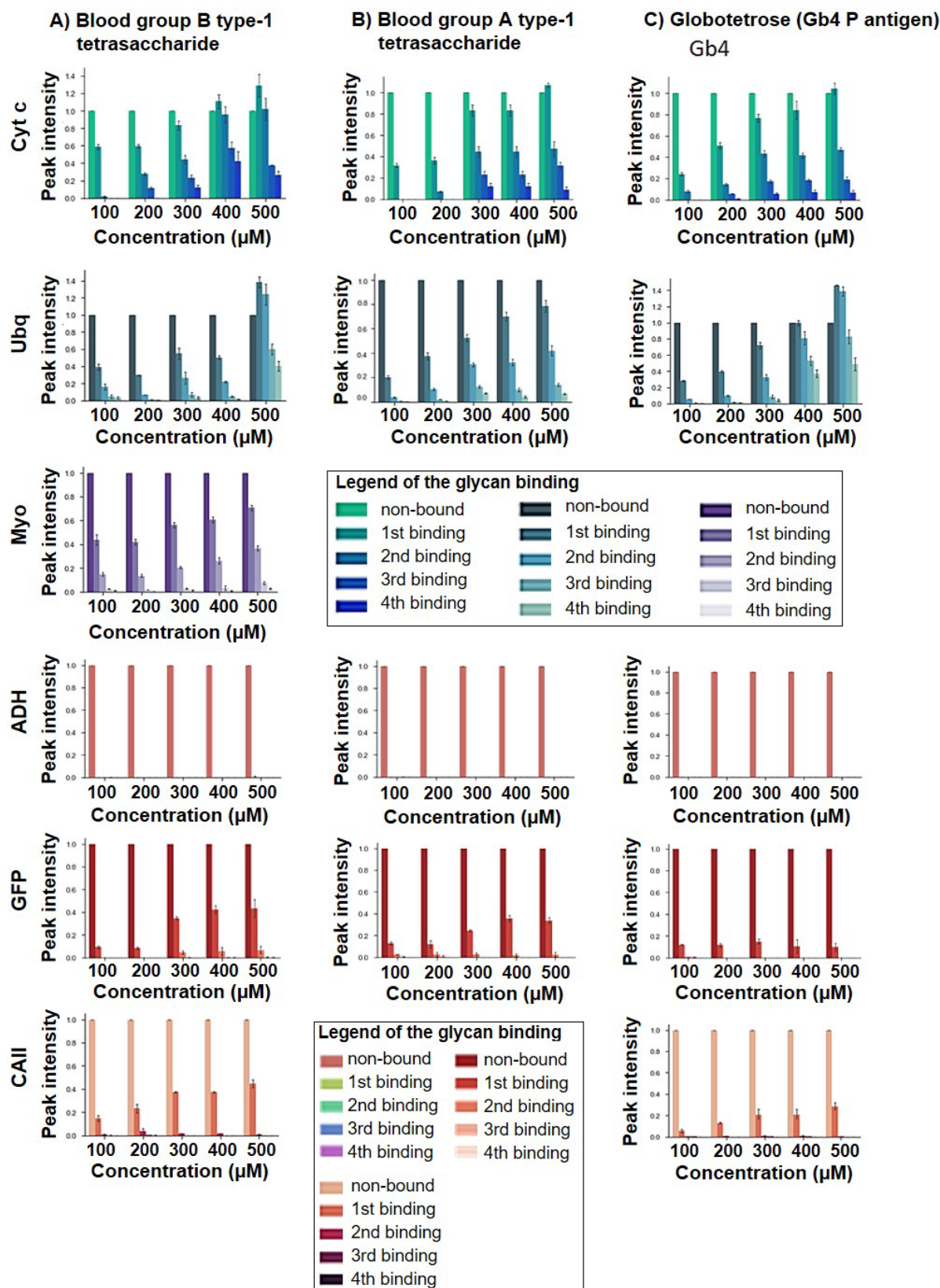
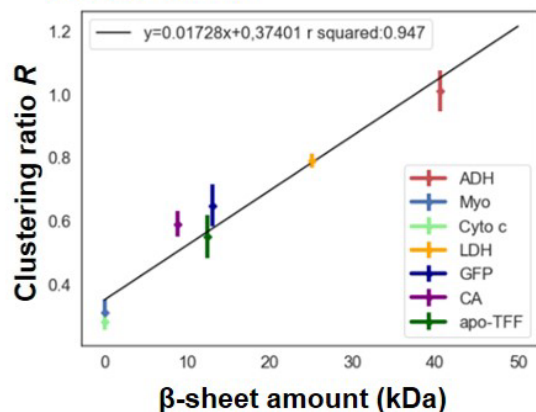


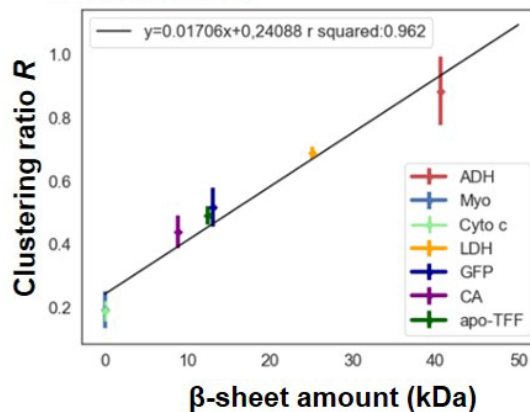
Figure S5. Specific binding of different carbohydrates on the P dimer after correction of the unspecific glycan clustering. Correction is based on the different reference proteins (glycans: blood group A and B

type-1 tetrasaccharide (HBGA), Globotetrose (Gb4, P antigen) at 100 - 500 μM concentration; ref. proteins: cyt c, Ubq, Myo, ADH, GFP, CAII).

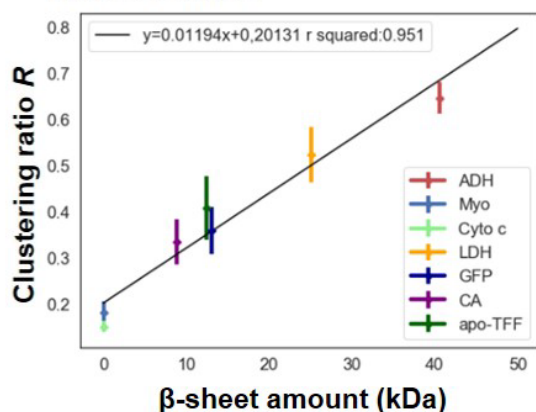
A) 400 μM blood group B type-1 tetrasaccharide



B) 300 μM blood group B type-1 tetrasaccharide



C) 200 μM blood group B type-1 tetrasaccharide



D) 100 μM blood group B type-1 tetrasaccharide

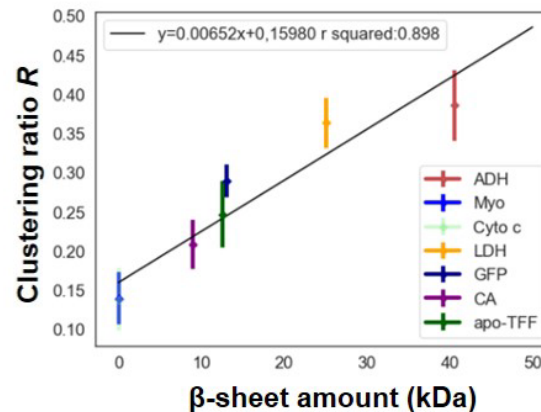


Figure S6. The relation between the amount of beta-sheet in the reference protein and the unspecific glycan clustering ratio R at different ligand concentrations (400 μM (A), 300 μM (B), 200 μM (C) and 100 μM (D)). The R value is calculated from the peak-area of the reference proteins (ADH, CA, cyto c, GFP, Myo, Ubq, LDH, apo-TFF). Native MS experiment were performed with a fixed concentration of P dimer (1 μM) and reference proteins (3 μM) at 150 mM ammonium acetate, pH7. The black line represents the linear regression between the unspecific clustering ratio of each reference protein and the corresponding mass of beta-sheets. The reference protein structures and protein sequence information is derived from each of corresponding defined PDB file.

Table S1. Glycan clustering ratio of HBGA B interaction with P dimer analysed with native MS. Data correction is based on the reference protein method.

Ref . Protein	glycan clustering ratio														
	100 µM			200 µM			300 µM			400 µM			500 µM		
	HBGA B	Gb4	HBGA A	HBGA B	Gb4	HBGA A	HBGA B	Gb4	HBGA A	HBGA B	Gb4	HBGA A	HBGA B	Gb4	HBGA A
Myo	0.14±0.03	0.14±0.02	0.13±0.03	0.18±0.02	0.17±0.03	0.16±0.03	0.19±0.06	0.20±0.03	0.22±0.03	0.31±0.04	0.30±0.01	0.29±0.04	0.36±0.05	0.37±0.06	0.34±0.03
Cyt c	0.14±0.01	0.10±0.01	0.18±0.03	0.15±0.01	0.15±0.00	0.16±0.02	0.19±0.03	0.19±0.03	0.19±0.03	0.29±0.02	0.29±0.04	0.27±0.02	0.35±0.03	0.45±0.02	0.41±0.09
Ubq	0.04±0.01	0.03±0.01	0.06±0.03	0.04±0.01	0.04±0.01	0.04±0.01	0.10±0.03	0.12±0.02	0.13±0.01	0.18±0.01	0.21±0.20	0.18±0.01	0.25±0.00	0.09±0.03	0.10±0.05
GFP	0.30±0.02	0.27±0.02	0.30±0.02	0.41±0.02	0.32±0.07	0.35±0.05	0.44±0.04	0.53±0.01	0.54±0.01	0.57±0.01	0.69±0.04	0.68±0.04	0.67±0.02	0.80±0.03	0.86±0.04
CA	0.22±0.03	0.21±0.01	0.20±0.05	0.32±0.05	0.32±0.07	0.37±0.01	0.40±0.04	0.50±0.00	0.42±0.02	0.60±0.05	0.60±0.05	0.57±0.02	0.75±0.07	0.75±0.03	0.74±0.07
ADH	0.42±0.03	0.34±0.04	0.39±0.02	0.64±0.06	0.66±0.03	0.65±0.02	0.97±0.08	0.81±0.11	0.88±0.10	1.02±0.08	1.03±0.08	0.98±0.05	1.45±0.11	1.26±0.03	1.32±0.10
apo-TFF	0.37±0.03	0.36±0.05	0.39±0.02	0.56±0.05	0.57±0.06	0.57±0.05	0.62±0.03	0.63±0.02	0.65±0.02	0.74±0.01	0.69±0.02	0.71±0.03	0.82±0.03	0.80±0.05	0.79±0.03
LDH	0.38±0.03	0.35±0.04	n.a.	0.54±0.05	0.51±0.08	n.a.	0.69±0.02	0.68±0.02	n.a.	0.77±0.05	0.80±0.01	n.a.	1.02±0.04	0.91±0.06	n.a.
SAGA P dimer	0.44±0.07	0.35±0.06	0.37±0.13	0.62±0.05	0.60±0.09	0.66±0.10	1.01±0.02	1.05±0.09	0.98±0.08	1.31±0.20	1.21±0.15	1.34±0.26	1.55±0.05	1.40±0.21	1.44±0.15

Table S2. Bound number of carbohydrates on P dimer analysed with native MS. Data correction is based on the reference protein method.

Ref. Protein	Bound number of HBGA B on Saga P dimer					Bound number of HBGA A on Saga P dimer					Bound number of Gb4 on Saga P dimer				
	100	200	300	400	500	100	200	300	400	500	100	200	300	400	500
Myo	2	3	3	3	4	2	3	3	3	4	2	3	3	3	4
Cyto c	2	3	4	4	4	2	3	3	4	4	1	2	3	3	4
Ubq	2	3	4	4	4	2	3	4	4	4	2	3	4	4	4
GFP	1	1	2	2	2	1	1	1	1	1	2	2	2	2	2
CA	1	2	2	2	2	1	1	1	1	1	1	1	2	2	2
ADH	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
apo-TFF	1	1	1	1	2	0	0	1	1	1	0	0	1	1	1
LDH	0	0	0	0	1	0	0	0	0	1	0	0	0	0	1

Table S3. Protein physiochemical properties used for plots. Data is obtained from Protein Data Bank in Europe (<https://www.ebi.ac.uk/pdbe-srv/pdbechem/>)

Ref. Protein	Analysed protein physiochemical feature									
	The share of β-sheet	The share of α-helix	mass	CCS TJM	positive charge residues number	negative charge residues number	pI	charge residues number	solvent accessible area	m/z
ADH	40.60	44.21	147.00	9876.81	128.00	144	6.21	272.00	31928.42	5933
apo-TFF	12.51	27.46	79.60	9959.74	85.00	87	6.81	172.00	15595.88	4469
CA	8.91	4.40	29.10	2378.38	27.00	30	6.41	57.00	6984.81	2985
cyto c	0.00	5.44	13.20	1286.00	21.00	12	9.59	33.00	3618.71	1864
GFP	13.08	2.61	29.60	2164.24	26.00	34	5.67	60.00	5608.96	3037
LDH	25.07	68.81	146.10	7421.70	148.00	140	8.17	288.00	25045.90	6120
Myo	0.00	12.14	17.00	1714.99	21.00	21	7.20	42.00	4980.50	2283