

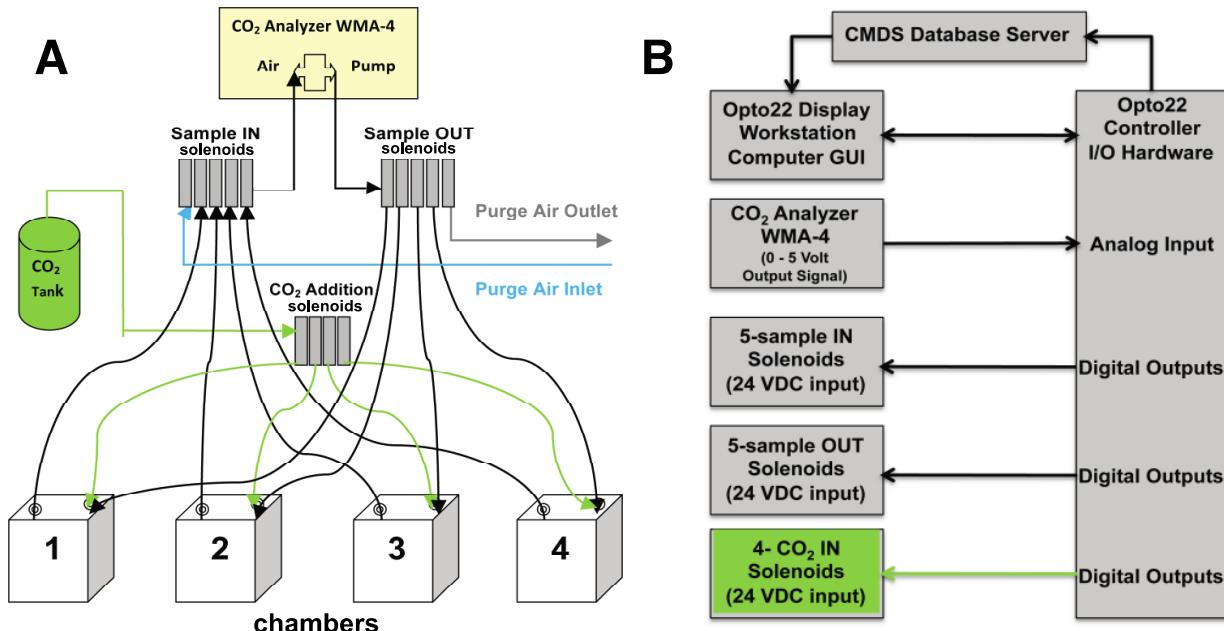
# Supplementary Information

**Table S1.** Pyrosequencing primers used in diversity analyses.

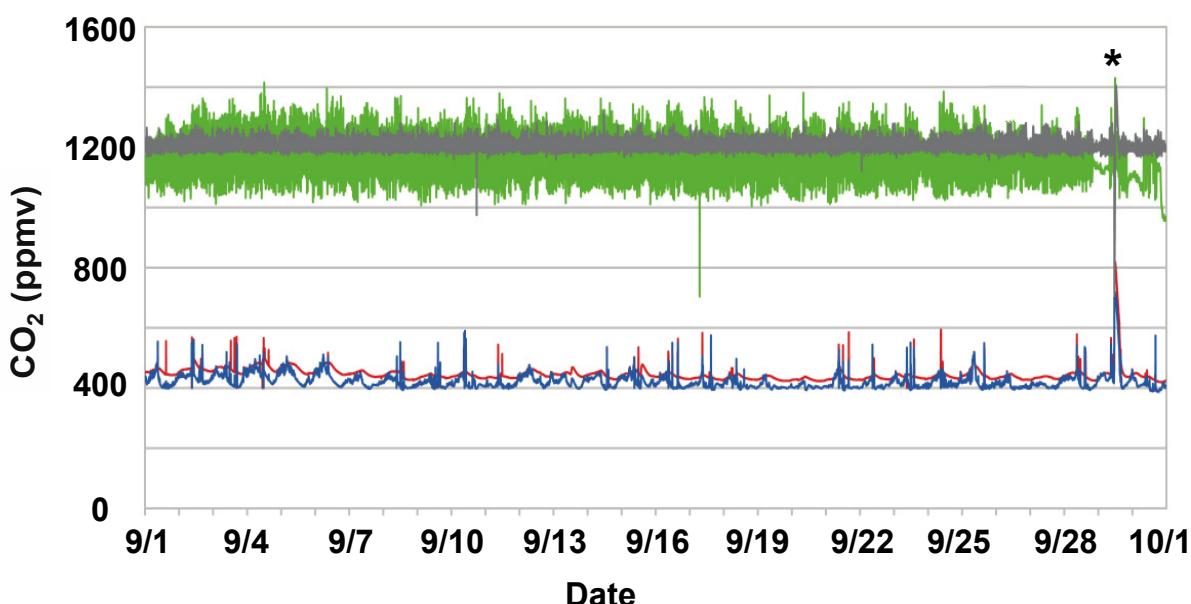
Name <sup>1</sup>	Primer Sequence <sup>2</sup>
454B-27F	5'- <u>GCCTTGCAGCCGCTCAG</u> <b>T</b> CAGAGTTGATCCTGGCTCAG-3'
454A-338R-C1-T0	5'- <u>GCCTCCCTCGGCCATCAGAACCAACC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C1-T2	5'- <u>GCCTCCCTCGGCCATCAGAACGAAGC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C1-T4	5'- <u>GCCTCCCTCGGCCATCAGCCCCAACCC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C1-T6	5'- <u>GCCTCCCTCGGCCATCAGCCGGCCTT</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C2-T0	5'- <u>GCCTCCCTCGGCCATCAGGAAATTGG</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C2-T2	5'- <u>GCCTCCCTCGGCCATCAGAACGCCCTT</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C2-T4	5'- <u>GCCTCCCTCGGCCATCAGCCTACCGC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C2-T6	5'- <u>GCCTCCCTCGGCCATCAGCCCGCCAT</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C3-T0	5'- <u>GCCTCCCTCGGCCATCAGCCAACCTT</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C3-T2	5'- <u>GCCTCCCTCGGCCATCAGTTCGAACGC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C3-T4	5'- <u>GCCTCCCTCGGCCATCAGAACATACCGC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C3-T6	5'- <u>GCCTCCCTCGGCCATCAGAACGCCAT</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C4-T0	5'- <u>GCCTCCCTCGGCCATCAGTTAAGGCC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C4-T2	5'- <u>GCCTCCCTCGGCCATCAGAACAGAACCC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C4-T4	5'- <u>GCCTCCCTCGGCCATCAGAACAGC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'
454A-338R-C4-T6	5'- <u>GCCTCCCTCGGCCATCAGCCTACGCC</u> <i>CAT</i> GCTGCCTCCGTAGGAGT-3'

Notes: <sup>1</sup> Name reflects the 454 Life Sciences Primer; the location within the 16S rRNA gene; the experimental chamber (C1, C2, C3, C4); and the time period the sample was collected (T0, T2, T4, T6 months). Note the forward primer 454B-27F was used for all samples; <sup>2</sup> Sequence reflects 454 Life Sciences Primer A/B (underlined); eight-base unique barcode; two-base linker (red italics); and the bacterial 16S rRNA gene primer (bold).

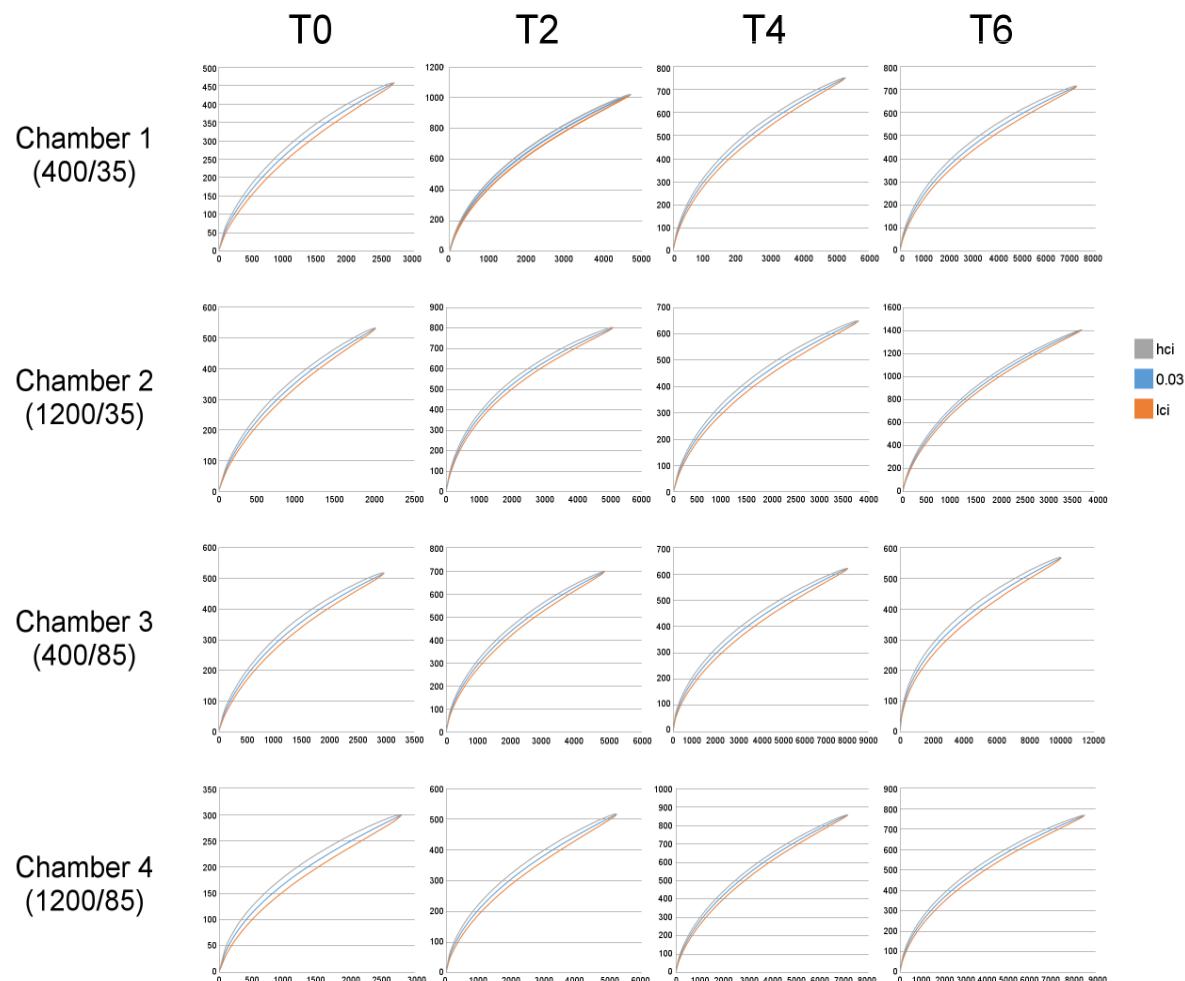
**Figure S1.** Diagrammatic representation of the environmental manipulation chambers. (A) Atmospheric gas flow control into the four environmental chambers. CO<sub>2</sub> flow is denoted in green; (B) Opto 22 hardware monitoring that controls CO<sub>2</sub> gas flow and records data to database server.



**Figure S2.** Representative month of CO<sub>2</sub> readings taken throughout the six-month microbialite in cubation. Readings taken every 8 min with CO<sub>2</sub> analyzer depict the tight regulation of the atmospheric headspace within each environmental chamber. Colors depict the four treatments: blue (Chamber 1, 400 ppmv CO<sub>2</sub>, 35‰ salinity); green (Chamber 2, 1200 ppmv CO<sub>2</sub>, 35‰ salinity); red (Chamber 3, 400 ppmv CO<sub>2</sub>, 85‰ salinity); and grey (Chamber 4, 1200 ppmv CO<sub>2</sub>, 85‰ salinity). Asterisk denotes a sampling event.



**Figure S3.** Rarefaction curves plotted 16S rRNA gene amplicon library from microbialites exposed to variable CO<sub>2</sub> and salinity levels. The blue curve represents the average OUTs observed at a 97% similarity threshold, while the grey and orange curves represent the upper and lower bounds, respectively, of the 95% confidence interval for that average. The X- and Y-axes represent the number of the sequences and OUTs, respectively.



© 2014 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).