



Article Start-Up Performance and Process Kinetics of a Two-Stage Partial Nitrification–Anaerobic Ammonium Oxidation Reactor

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Abstract: The study of two-stage partial nitrification-anaerobic ammonium oxidation (PN/A) reactors, which are advantageous in engineering applications, still lacks research on process kinetics. Therefore, in this study, the start-up performance and process kinetics of a two-stage PN/A reactor were evaluated by controlling the reaction conditions, for which the two reactors were inoculated with sludge, incubated separately, and then operated in tandem. Increasing the ammonia load of the reactor during the 60 d stabilization period resulted in a nitrogen accumulation rate of 96.93% and a $[NO_2^- - N]_{Eff}/[NH_4^+ - N]_{Eff}$ ratio of 1.33, which is close to the theoretical value of 1.32. Successful initiation of the A reactor was achieved after 55 d of operation by inoculating with anammox-activated sludge and granular activated carbon, and the PN and A reactors then successfully operated in combination for 20 d, with an average $NH_4^+ - N$ efficiency of 99.04% and the $NH_4^+ - N$ load of the A reactor showing an "S-shaped" curve. An analysis of the microbial growth kinetic models indicated that the removal of $NH_4^+ - N$ could be successfully described by the logistic, modified logistic, modified Gompertz, and modified Boltzmann models. A strong association between the model and the dependent variable was observed. The process kinetic analysis showed that the removal of $NH_4^+ - N$ from reactor A could be simulated under steady-state conditions, using the Grau second-order model. The parameters obtained from the model analysis are expected to help predict the denitrification performance of the reactor, facilitate operational management and control, and thus provide a promising research basis for the introduction of automated control systems.

Keywords: partial nitrification; anammox; kinetic model

1. Introduction

Nitrogenous pollutants, which reach water bodies via the discharge of nutrients, have become ubiquitous in the aquatic environment, leading to eutrophication, ecosystem damage, and toxicological risk to human life [1]. Biological methods are important for controlling water pollution due to their high efficiency, durability, and lack of secondary pollution [2]. The coupled biological denitrification process of partial nitrification/anammox (PN/A) has become a key research target for biological treatment due to its high denitrification efficiency, with 60% less aeration required and 90% excess activated sludge [3]. However, partial nitrification can eliminate ammonia-oxidizing bacteria (AOB) while retaining nitrite-oxidizing bacteria (NOB) by controlling factors such as temperature and hydraulic residence time. AOB dominates the conversion of NH₄⁺ – N to NO₂ – N [4], and anammox is a biological reaction in which anaerobic AOB (AnAOB) uses NH₄⁺ – N as an electron donor and NO₂⁻ – N as an electron acceptor to produce N₂ and a small amount of NO₃⁻ – N under anaerobic conditions [5,6]. The effectiveness of PN/A processes in



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). treating wastewater with high ammonia nitrogen content, such as waste leachate [7], farming wastewater [8], municipal wastewater [9], and sludge digestate [10], has been widely studied, with results suggesting certain economic advantages in using the method [11].

PN/A process applications are mainly one- or two-stage [12], with one-stage PN/A accounting for over 80% of PN/A processes worldwide [13]. Although research focus and applications tend to favor one-stage PN/A due to its smaller and more economical footprint [14], two-stage PN/A has greater potential for tighter control, a shorter start-up time, and reduced inhibition [15]. For influent wastewater with large variations in terms of water quality and quantity, the two-stage type is associated with greater advantages, such as the front-end PN section acting as a buffer and separate optimization of the PN/A process [16,17].

Studies investigating the start-up of two-stage PN/A have generally focused on the start-up effects and microbial characteristics [18], with few scholars focusing on the kinetics or attempting to simulate the startup and operation processes of the PN/A reactor, and even fewer focused on the narrowly applied two-stage PN/A reactor. The aim of this study was thus to analyze the start-up performance and system kinetics of a two-stage PN/A reactor, determine a kinetic model that is consistent with the observed results, and provide a reference for the start-up conditions and environmental control of the reactor in practical applications. The results are expected to facilitate better prediction, management, and optimization of the two-stage PN/A reactor.

2. Materials and Methods

2.1. Experimental Setup

The experimental setup was divided into two parts: partial nitrification (PN section) and anammox (A section), as shown in Figure 1. The PN section consisted of aerobic and sedimentation tanks, while the A section consisted of a pH adjustment tank and a UASB reactor. All reaction bodies were composed of organic glass. The volumes of the aerobic and sedimentation tanks were 9 and 8 L, respectively. Water was fed from the upper end of the aerobic tank, which was equipped with pH poles, temperature heating rods, and aeration heads to control the pH, temperature, and dissolved oxygen (DO), respectively. The volumes of the pH adjustment sedimentation and UASB tanks were 14.3 and 61 L, respectively. Water from the sedimentation tank was mixed with the temperature- and pH-adjusted water from the pH-adjustment tank and fed to the UASB tank from the bottom. The total effective volume of the reactor was 61 L, the sampling port was set on the right side, and the outer wall was insulated with thermal insulation material.



Figure 1. Diagram of a PN/A reactor: (**a**) the aerobic reactor, (**b**) the settling basin, (**c**) the pH-regulating tank, and (**d**) the UASB reactor.

2.2. Experimental Water and Inoculated Sludge

Cultivation was performed separately during the reactor start-up and load-raising phases, with water dispensed separately into the PN and A reactors until both reached the ideal operating load. The two reactors were then connected in series. This method reduces the difficulty that is associated with cultivation and improves the cultivation efficiency, while reducing the likelihood that the two reactors will interfere with each other during early cultivation, thus increasing the start-up time.

PN section reactor: Tap water was used in preparing the experimental water, thus avoiding interference from random bacteria, and the concentration of $[NH_4^+ - N]_{Inf}$ was 500 mg/L. Specific data are represented in Table 1.

Category	Pharmaceuticals	Concentration	Pharmaceuticals	Concentration
Experimental water	(NH ₄) ₂ SO ₄ KH ₂ PO ₄ NaHCO ₃	500–5500 mg/L Formulated by concentration Formulated by concentration	CaCl ₂ ·2H ₂ O MgSO ₄ ·7H ₂ O Trace elements 1 and 2	0.113 g/L 0.1 g/L 0.5 and 1 mL/L
Trace elements I	FeSO ₄ ·7H ₂ O MnCl ₂ ·4H ₂ O	10 0.352	$\begin{array}{c} C_{10}H_{14}N_2Na_2O_3\\ CoCl_2\cdot 6H_2O \end{array}$	5.6 0.096
Trace elements II	NiCl ₂ ·6H ₂ O ZnSO ₄ ·7H ₂ O NaMoO ₄ ·2H ₂ O	0.08 0.172 0.11	$\begin{array}{c} CuSO_4{\cdot}5H_2O\\ Na_2SeO_4{\cdot}10H_2O\\ C_{10}H_{14}N_2Na_2O_3 \end{array}$	0.1 0.105 5.0

Table 1. Simulated influent compositions.

A-section reactor: Similar to the PN reactor, $(NH_4)_2SO_4$ (Sinopharm Chemical Reagent Co. Ltd., Shanghai, China) and NaNO₂ (Sinopharm Chemical Reagent Co. Ltd., Shanghai, China) were prepared at the required concentrations. KH_2PO_4 (Sinopharm Chemical Reagent Co. Ltd., Shanghai, China) was used to adjust the pH of the feed water. $KHCO_3$ was added at 1500 mg/L, CaCl₂-7H₂O at 226.6 mg/L, MgSO₄-7H₂O at 200 mg/L, and Na₂S₂O₃ at 24.81 mg/L. Trace elements: The dosages and compositions of the trace elements were consistent with those in the PN section.

Inoculated sludge:

Activated sludge for use in the aerobic tank of the PN reactor was obtained from CASS tank No. 1 at the YanShan town wastewater treatment plant in the Guangxi Zhuang autonomous region. Impurities and debris were screened, and sediment was removed after resting and settling to improve the inoculated sludge concentration, thereby improving adaptation to the $[NH_4^+ - N]_{Inf}$ concentration.

Anaerobic ammonia oxidation-activated sludge from the research laboratory of Professor Furukawa at Kumamoto University, Japan, was used in the UASB tank of the A-section reactor. Treated granular BC, which was obtained by hammering bamboo charcoal to a particle size of approximately 1–2 mm, washing with distilled water, and soaking overnight before removing all liquid, was mixed with the sludge before cultivation for rapid enrichment with anaerobic ammonia-oxidizing bacteria. Color changes that are associated with anaerobic ammonia-oxidizing bacteria were observed in the resulting granular sludge.

A-section reactor's parameters: Pool temperature was controlled to within 35 ± 1 °C, using a ring heating rod; pH was controlled at 7.5–7.6 by adding acidic solution; and aeration of inlet water was obtained using 99% high-purity nitrogen. A small amount of anhydrous sodium sulfite was added to maintain the DO concentration of the inlet water below 0.1 mg/L.

2.3. Analysis Methods

2.3.1. Water Quality Analysis

The pH was determined using a pH meter (PHS-3C, INESA, Shanghai, China). The mixed liquor suspended solid concentration (MLSS) and mixed liquor volatile suspended solid concentration (MLVSS) were measured by weight. $NH_4^+ - N$, NO_2^-N , and $NO_3^- - N$ were measured using a continuous flow analyzer (Skalar Co. Ltd., Breda, Netherlands), and regular measurements of DO and temperature were obtained using a portable dissolved oxygen Meter (Hach Co. Ltd., CO, USA) and a thermometer, respectively.

2.3.2. Data Analysis

The free ammonia (FA), nitrogen accumulation rate (NAR), ammonia conversion efficiency (ACE), ammonia load (NLR), and settling velocity (SV) were calculated using the following equations (Equations (1)–(4)).

$$FA = \frac{17}{14} \frac{\left[NH_4^+ - N\right]_{Eff} \times 10^{pH}}{e^{6344/(273+T)} + 10^{pH}}$$
(1)

$$NAR = \frac{[NO_2^- - N]_{Eff}}{[NO_2^- - N]_{Eff} + [NO_3^- - N]_{Eff}} \times 100\%$$
(2)

$$ACE = \frac{[NH_4^+ - N]_{Inf} - [NH_4^+ - N]_{Eff}}{[NH_4^+ - N]_{Inf}} \times 100\%$$
(3)

$$NLR = \frac{([N H_4^+ - N]_{Inf} - [N H_4^+ - N]_{Eff}) \times Q}{HRT \times V}$$
(4)

where $[NH_4^+ - N]_{Eff}$ is the effluent concentration of $NH_4^+ - N$; $[NO_2^- - N]_{Eff}$ is the effluent concentration of $NO_2^- - N$; $[NO_3^- - N]_{Eff}$ is the effluent concentration of $NO_3^- - N$; $[NH_4^+ - N]_{Inf}$ is the influent concentration of $NH_4^+ - N$; Q is the intake volume; T is the temperature of the reactor; and H is the settlement height.

3. Discussion

3.1. PN Section Reactor Start-Up

The NLR was added during the operation of the PN section reactor to improve the cultivation of the activated sludge, improving its capacity for treating high-ammonianitrogen wastewater, while increasing the concentration of FA to screen AOB and eliminate NOB. Complete NOB and AOB activity inhibition has been reported at FA concentrations of >0.6 and 10 mg/L, respectively [19]; however, severe inhibition of AOB activity occurs only at FA concentrations > 40 mg/L. Because FA concentration inhibits AOB at greater concentration ranges than NOB, nitrite accumulation could be successfully screened and enriched if the FA concentration was controlled within a reasonable range. As shown in Figure 2, the PN section reactor was operated for 60 d and was able to meet the semi-nitrification and feed water requirements for subsequent anaerobic ammonia oxidation. Acclimation was performed for 1–3 d, load ramp-up for 4–40 d, and stabilization for 40–60 d.

Because 500 mg/L of $[NH_4^+ - N]_{Inf}$ that was used during the adaptation period was much higher than that in the domestic sewage that was treated with activated sludge, the removal of $NH_4^+ - N$ was not effective, and only 25–30% of the $NH_4^+ - N$ was converted to $NO_3^- - N$ and $NO_2^- - N$. $[NO_3^- - N]_{Eff}$ was greater than $[NO_2^- - N]_{Eff}$ because only HCO₃, an inorganic carbon source, was available in the artificially prepared water.

The ratio of nitrate to nitrite in the effluent reached 1, and the NAR reached 92.74% on day 4 of the load-boosting period, indicating that the conversion rate of ammonia nitrogen exceeded 50% and that the activated sludge in the plant had adapted to the 500 mg/L $[NH_4^+ - N]_{Inf}$ concentration. Therefore, from the fourth day onwards, $[NH_4^+ - N]_{Inf}$ was added every 2–5 d to load the influent until the reactor was upgraded to the highest $[NH_4^+ - N]_{Inf}$ of 5496.48 mg/L. Although fluctuations were observed in the $[NO_3^- - N]_{Eff}$ and $[NO_2^- - N]_{Eff}$ content during this period, a gradually increasing water nitrite-nitrogen concentration trend was still evident overall, while $[NO_3^- - N]_{Eff}$ was maintained at a low

level. An obvious inflection point for $[NO_3^- - N]_{Eff}$ and $[NH_4^+ - N]_{Eff}$ on day 31 was due to damage to the influent pump, with the sudden decrease in influent water leading to a corresponding decrease in the FA concentration and reduced AOB and NOB inhibition. However, a significant increase was observed only in the $[NO_2^- - N]_{Eff}$ levels, indicating a significant advantage for the AOB volume in the water column, along with a higher degree of nitrification.



Figure 2. Ammonia nitrogen in influent and effluent; nitrite nitrogen in the effluent; nitrate nitrogen in the effluent; and NLR, FA, NAR, and ACE during 60 d PN operation.

According to Li et al. [20], the concentration of HCO_3^- during short-course nitrification can improve the conversion of ammonia nitrogen and show a significantly increase $[NO_2^- - N]_{Eff}$ in the short term. Therefore, the short-range nitrification reaction was connected to the anammox reactor in the follow-up experiments, and the alkaline solution was changed to NaOH on the 55th day, allowing the ammonia load in the short-range nitrification reactor to reach approximately half that of the previous level, with the ammonia nitrogen concentration in the reactor effluent similar to that of the effluent nitrite nitrogen. The method provides sufficient amounts of the reaction substrate, $NH_4^+ - N$, for subsequent anaerobic ammonia oxidation. As shown in Figure 2, after the NAR reached 95.81% on day 4, it remained high as the ammonia load increased, albeit with certain fluctuations. A NAR of <90% was observed on only 14 of the 60 d that followed start-up, with the NAR reaching 96.93% and the $[NO_2^- - N]_{Eff}/[NH_4^+ - N]_{Eff}$ ratio being 1.33, which is close to the theoretical value of 1.32, during the stabilization period. This indicates high nitrite accumulation over some time, with the AOB completely eluting the NOB, indicating effective and successful nitrification.

3.2. A-Section Reactor Start-Up

The UASB process was selected for the A-section reactor. The feasibility of this process for treating high-concentration ammonia-nitrogen wastewater has been proven previously [21]. The addition of granular activated carbon accelerates the formation of granular sludge from AnAOB, which grows in granular form and is considered a suitable and promising method for anaerobic ammonia oxidation [22]. The laboratory scale-activated sludge AnAOB was used for 55 d, during which time the $[NH_4^+ - N]_{Inf}$ and $[NO_2^- - N]_{Inf}$ concentrations were gradually increased, with the highest influent concentrations reaching 745.8 mg/L and 597.7 mg/L, respectively. The NLR reached 4.3 kg-N/m³/d, and the con-

centrations of reactor $[NH_4^+ - N]_{Eff}$ and $[NO_2^- - N]_{Eff}$ concentrations is maintained at a low concentration throughout the 55 d-phase. In particular, maintaining the $[NO_2^- - N]_{Eff}$ concentration below 10 mg/L prevented the high $[NO_2^- - N]_{Eff}$ from inhibiting the activity of the bacterial strains. The reactor was observed to operate stably with only one load shock. Adaptation was performed over 1–3 d, load boosting over 4–45 d, and stabilization over 45–55 d.

The effluent that was present within the reactor on day 1 was not effective during the start-up period because the strictly anaerobic AAOB strains require a certain adaptation period following transferal and are exposed to oxygen during the process, rendering them inactive [23,24]. In addition, the 60 L reactor is six times larger than the previous laboratory-scale reactor in terms of volume. To ensure the same concentration, more activated carbon particles were added to the reactor; however, the lack of microbial attachment to the newly added activated carbon particles rendered culture necessary. The effluent effect was observed to improve over two consecutive days from the second day, with the $[NH_4^+ - N]_{Eff}$ decreasing to 17.3 mg/L and the $[NO_2^- - N]_{Eff}$ to 1 mg/L, indicating that the strain was fully suitable for the new reactor and the load could be increased. The content of Figure 3 shows the internal reactor changes during the startup phase of A-section reactor. The upper graph shows A-section reactor influent ammonia nitrogen, influent nitrite, effluent ammonia nitrogen, effluent nitrite, effluent ammonia nitrogen and ammonia load. The lower graph shows the ammonia conversion rate, nitrite conversion rate, influent ammonia/nitrogen/nitrite ratio, and nitrate-nitrogen conversion rate within the reactor during the startup phase of A-section reactor.



Figure 3. This figure shows the internal changes in reactor A at the start-up stage. The above diagram shows $InfNH_4^+$, $InfNO_2^-$, $EffNH_4^+$, $EffNO_2^-$, $EffNO_3^-$ and TolNLR of the reactor. In order to start reactor A, the ammonia load in the reactor was gradually increased during the start-up stage: that is, the daily influent ammonia nitrogen and influent nitrous oxide were gradually increased. The effluent ammonia nitrogen and nitrous oxide remained at a low level, and the effluent nitrate nitrogen

increased with the increase in the load. The following figure shows the $[NH_4^+]$ removal rate, $[NO_2^-]$ removal rate, $[InfNO_2^-/InfNH_4^+]$ of influent water and $[InfNO_3^-/InfNH_4^+]$ in reactor A at the start-up stage. As shown in the figure, the $[InfNO_3^-/InfNH_4^+]$ gradually increases with the start-up process, and $[InfNO_2^-/InfNH_4^+]$ gradually approaches the same level.

Stabilization period: Loading was steadily upgraded from day 4 to 45, at which point an inflection point appeared in the reactor when the ammonia nitrogen load reached the maximum value of $4.3 \text{ kg-N/m}^3/\text{d}$. The reactor began to show blockage from the activated carbon particles on day 46, with immediate deterioration observed in the effluent water quality. To avoid excessive high-water nitrite-affecting activity, the blockage problem was solved, and the load was slowly reduced to $3 \text{ kg-N/m}^3/\text{d}$ to extend the hydraulic residence time, followed by a slow restoration to $4.3 \text{ kg-N/m}^3/\text{d}$. This technique was observed to protect the strain activity to the maximum extent, with the reactor returning to its original performance and stable operation maintained after a week of buffering.

The anammox reaction equation is represented by the following equation [25]:

$1.32NO_{2}^{-} + 1NH_{4}^{+} + 0.13[H^{+}] + 0.066HCO_{3}^{-} \rightarrow 1.02505N_{2} + 0.26NO_{3}^{-} + 2.025H_{2}O + 0.066CH_{2}O_{0.5}N_{0.15}$ (5)

As seen in the formula, $[NO_3^- - N]_{Eff}$ increases as $[NH_4^+ - N]_{Inf}$ and $[NO_2^- - N]_{Inf}$ in the influent water increase. This is because, for every 1 mol/L of $NH_4^+ - N$ consumed in the anammox reaction equation, 1.32 mol/L of $NO_2^- - N$ and 0.26 mol/L of $NO_3^- - N$ are theoretically produced [26,27]. A certain reactor load can be used to appropriately reduce the influent ammonia nitrogen and nitrite nitrogen concentrations while shortening the hydraulic retention time to increase the load from the side, meaning that the effluent nitrate nitrogen concentration and total nitrogen content can be reduced with the premise of ensuring the load.

 $NH_4^+ - N$ removal rate: As $[NH_4^+ - N]_{Inf}$ increased, and $[NH_4^+ - N]_{Eff}$ decreased, indicating the good treatment effect of the reactor. The increase in $[NH_4^+ - N]_{Inf}$ from 68.8 mg/L at the beginning to a maximum of 597.7 mg/L was accompanied by a decrease in $[NH_4^+ - N]_{Eff}$, from 40.3 mg/L to about 5 mg/L during the stabilization period, at which point an $NH_4^+ - N$ removal rate of approximately 98% was observed, with a maximum of 99.3%.

 $NO_2^- - N$ removal rate: $NO_2^- - N$ removal was maintained at approximately 99%, except during the pre-startup and pipe-blockage periods. The $[NO_2^- - N]_{Inf}$ increased from 64.5 mg/L to 745.8 mg/L, with anammox treatment of the $NO_2^- - N$ wastewater leading to a $[NO_2^- - N]_{Eff}$ of only 1–2 mg/L, indicating particularly strong anammox treatment ability. These results indicate strong activity for the anaerobic ammonia-oxidizing bacteria, with outstanding treatment capacity.

As shown in Figure 3, the ratio of $[NO_2^- - N]_{inf}/[NH_4^+ - N]_{inf}$ in the reactor ranged from 1.22 to 1.35, while the ratio of $[NO_3^- - N]_{Eff}/[NH_4^+ - N]_{Eff}$ ranged from 0.17 to 0.26, remaining at approximately 0.25 during stabilization. The reaction ratio in this reactor was not stable but fluctuated, which may be because the conditions under actual operation differ from those in the laboratory. Many variables affect the actual reaction process, and the regulation of the reaction conditions cannot be realized instantaneously. Secondly, the microbial community in the reactor is not singular, and a small number of other microorganisms may be involved in substrate removal within the reactor. Chen [28] suggested that the slow proliferation of anaerobic anammox bacteria means that part of the reaction substrate is likely consumed by other miscellaneous bacteria in the anammox reactor, which is the same as the data analysis results obtained in this experiment. These factors can lead to an imbalance in the ammonia nitrogen and nitrite nitrogen removal ratios of the influent material, which fluctuates around the theoretical value of 1.32. However, in general, the proportion observed in the reactor is still in line with the proportion of anaerobic ammoxidation bacteria, indicating that anaerobic ammoxidation is the main reaction occurring within the reactor.

3.3. Joint Operation

Due to the strong sludge activity described in Section 2.1, the influent ammonia nitrogen concentration was reduced to 300 mg/L. The PN section effluent was then directly used as the A-section influent, and the two operated jointly to observe the effluent effect of the combined process.

3.3.1. Partial Nitrification Stage

The combined process was operated for 20 d, with no obvious inflection points observed. The $[NH_4^+ - N]_{Inf}$ of approximately 300 mg/L in the PN reactor was associated with a $[NH_4^+ - N]_{Eff}$ of 105–125 mg/L, $[NO_2^- - N]_{Eff}$ of 130–150 mg/L, and $[NO_3^- - N]_{Eff}$ of 5–20 mg/L, indicating that the nitrogen reached the effluent, which acted as influent for the anammox stage. Anammox requires a strict theoretical substrate concentration ratio of $NO_2^- - N:NH_4^+ - N = 1:1.32$, and the similarity of the obtained ratio to the theoretical value is an important indicator of whether the next step of anaerobic ammonia oxidation will proceed normally. The ratio of nitrite nitrogen to ammonia nitrogen in the effluent of approximately 1.2 for the PN reactor thus satisfied the partial nitrification effluent water, with an average NAR of 88.67% and the highest value of 94.70% indicating sufficient nitrite for the subsequent anammox reaction.

3.3.2. Anammox Stage

The partial-nitrification effluent entering the reactor was best treated by the mediumefficiency anammox reactor, in which $[NO_3^- - N]_{Eff}$ was maintained below 5 mg/L and $[NO_2^- - N]_{Eff}$ below 5 mg/L. The results indicated that the reactor operated well in the latter stage of the anammox process and that it could therefore be connected to the partial nitrification reactor in the former stage. The removal rates of $NH_4^+ - N$ and $NO_2^- - N$ in the later anammox stage were stable, with the removal rate of $NH_4^+ - N$ remaining at >96% and the removal rate of $NO_2^- - N$ stable at approximately 99% for a long period. The TN removal rate of the reactor was stable at above 80%, and compared to the artificial water distribution, the semi-nitrosated effluent of short-range nitrification contained approximately 20 mg/L of nitrogen in the form of nitrate. As seen in Figure 4, the increase in the nitrate nitrogen concentration did not affect treatment in the anammox stage. The ratio of nitrite nitrogen to ammonia nitrogen consumed was approximately 1.2, whereas the ratio of generated nitrate nitrogen to ammonia nitrogen consumed was approximately 0.22. This indicates that the high-efficiency anammox reaction achieved a good removal from the semi-nitrosated effluent during the partial nitrification stage.

3.4. Growth Kinetics Analysis

The start-up process of a microbial reactor involves the growth, reproduction, and expansion of the functional microorganisms, with microorganisms adapting to their environment during start-up and propagation and expansion occurring during loading. Correspondingly, an improvement in the treatment effect of the microbial reactor can mark an increase in the microbial population [29]. However, the growth of microorganisms is not unlimited and is affected by the living space, nutrient concentration, and other conditions. Therefore, the growth curve of microorganisms is bound to be exponential in the early stages and inhibited by restrictions on the living environment in the later stages [30]. In this experiment, the growth of the anammox bacteria in the A-section reactor was slow in the early stages, during which time microbes adapted to the new environment, adjusting their metabolic activities. However, as the substrate concentration and microbial population in the influent water increase, the anammox reaction proceeds more rapidly, with the total denitrification load tending towards constancy when the substrate concentration or available space stabilizes, after which the bacterial growth rate decreases, entering a stable period.



Figure 4. This graph shows the reaction data for 20 days of combined operation of the reactor. The above figure shows [InfNH₄⁺] of the reactor; [Eff NH₄⁺], [EffNO₂⁻], and [EffNO₃⁻] of the PN reactor; [EffNO₂⁻] and [EffNO₃⁻] of the A reactor; and [TolNLR] of the combined reactor. The following figure shows the NAR and [EffNO₂⁻/EffNH₄⁺] of the PN reactor and the [NH₄⁺] removal rate and [NO₂⁻] removal rate of reactor A.

Li et al. [20] used four kinetic models to simulate the growth of anammox microbial cells in the presence of a low-concentration substrate and successfully predicted the ideal microbial growth in a low-concentration UASB reactor. Therefore, cell growth in an anammox microbial reactor with a high substrate concentration was investigated in this study [31]. The process in the A-section reactor was analyzed using a microbial growth kinetic model.

Logistic model N_t =
$$\frac{K}{1 + \exp(a - rt)}$$
 (6)

Modified logistic model N_t =
$$\frac{K}{1 + \exp[4r(\lambda - t)/K + 2)]}$$
 (7)

Modified Gompertz model N_t = Kexp
$$\left[-\exp\left(\frac{er}{K}(\lambda - t) + 1\right)\right]$$
 (8)

Modified Boltzmann model N_t = K +
$$\frac{M - K}{1 + \exp\left(\frac{t - t_c}{t_d}\right)}$$
 (9)

 N_t is the NLR at time, t, which is proportional to the microbial growth rate (kg-N/m³-d); K is the maximum NLR removal load (kg-N/m³-d); a is a constant; r is the maximum instantaneous growth rate for total denitrification; λ is the denitrification delay time (d); M is the minimum total nitrogen removal load (g/L·d); t is the run time (d); t_c corresponds to K/2 (d); and td is a time constant (d).

Fitting of the experimental data to the NLR of reactor A resulted in high R^2 values of >0.95 for all four growth models (Table 2), indicating that all are suitable for describing the start-up performance of the A-section reactor in the PN/A reactor. The fitted curve

for the start-up period was in the shape of an "S" (Figure 5), indicating that the anammox proliferation process can be divided into an adaptation period, an active period, and a stabilization period. The fitted maximum ammonia nitrogen removal loads were consistent at 4.47 kg-N/ m^3 -d for both the logistic and the modified logistic models, whereas the maximum instantaneous growth rates (r) of 0.0903, 0.1009, and 0.0918 were obtained for ammonia nitrogen removal using the logistic model, Gompertz model, and Boltzmann model, respectively. Delay times (λ) of 8.609 and 6.152 d were obtained for ammonia removal using the modified logistic and modified Gompertz models, respectively, while fitting to the modified Boltzmann model resulted in a minimum $NH_4^+ - N$ removal load (M) of 0.113 kg-N/m³-d, corresponding to a t_c of 32.595 d and a t_d of 10.953 d. The significantly shorter delay times for ammonia nitrogen removal in this study as compared to the 17.28 and 13.49 d obtained by Li et al. [20] indicate a higher reaction rate under the high concentration conditions, indicating a better growth environment for anammox bacteria with better proliferation and enrichment. By analyzing these four models, it was possible to obtain the maximum NLR, instantaneous growth rate, and delay time for $NH_4^+ - N$ removal. These results help predict the ammonia nitrogen removal performance in practice and are thus expected to improve reactor operation management and control.

Table 2. Summary of growth kinetic parameters in different models.

Model					R ²
Logistic model	K 4.470	a 2.777	r 0.0903	-	0.9528
Modified logistic model	K 4.469	λ 8.609	r 0.1009		0.9528
Modified Gompertz model	K 5.536	λ 6.152	r 0.0918		0.9535
Modified Boltzmann model	K 4.610	M 0.113	t _c 32.595	t _d 10.953	0.9507

3.5. Analysis of Process Dynamics

Process kinetics modeling is an analytical method that is used for monitoring system performance, the fitted kinetic results used to estimate reactor treatment efficiency under the same operating conditions [32]. The Grau second-order model is the most widely used mathematical model for analyzing the performance of anaerobic reactors and can describe the dynamic behavior of anaerobic bioreactors [33,34]. The general equation of Grau's second-order model is shown in Equation (10):

$$-\frac{d_{s}}{d_{t}} = k_{1} X \left(\frac{S_{e}}{S_{i}}\right)^{2}$$
(10)

where k_1 is the kinetic constant (d⁻¹), X is the sludge concentration (g/L), S_i represents $[NH_4^+ - N]_{Inf}$ (mg/L), and S_e represents $[NH_4^+ - N]_{Eff}$ (mg/L). If the boundary condition ($S_e = S_i$) is used, t = HRT, and the following equation can be obtained by linearization:

$$\frac{S_i H}{S_i - S_e} = H + \frac{S_i}{k_1 X} \tag{11}$$

where H is the hydraulic retention time (HRT) listed in Table 3. If $S_i/(kX)$ is expressed as a constant b, and $S_i/(S_i-S_e)$ is expressed as E (a percentage of substrate removal), the above equation can be modified as follows:

$$\mathbf{E}\mathbf{H} = \mathbf{b} + \mathbf{a}\mathbf{H} \tag{12}$$



Time(day)

Figure 5. Growth kinetic model of an A-section reactor for ammonia nitrogen removal under (**a**) the logistic model, (**b**) the modified logistic model, (**c**) the modified Gompertz model, and (**d**) the modified Boltzmann model.

Time(day)

Time (d)	HRT (Hydraulic Retention Time) (h)	
1–10	4.5	
21–20	2.5	
21–30	2.2	
31–40	2	
41–55	2.3	
Grau second-order model		
R ²	0.9984	
a	1.0163	
b	0.0361	

 Table 3. Operating conditions and modeling parameters of the reactor for different periods of time.

The fit is shown in Figure 5 for EH placed on the *y*-axis and H on the *x*-axis. The equation for predicting the $[NH_4^+ - N]_{Eff}$ of reactor A is derived using Equation (13).

$$S_e = \left(1 - \frac{1}{1.063 + \frac{0.0361}{HRT}}\right) S_i \tag{13}$$

As shown in Figure 6, fitting this model resulted in $R^2 = 0.9984$, a = 1.0163, and b = 0.0361, allowing a better curve to be obtained. The equation for predicting the effluent ammonia nitrogen concentration also provides new concepts for an automated control system that can manage two-stage PN/A reactors.



Figure 6. Process dynamics model (H is the hydraulic retention time (HRT), and E is the reciprocal of the percentage of substrate removal).

4. Conclusions

The two-stage PN/A reactor, which is advantageous in engineering applications, has not been rigorously studied in terms of process kinetics. Therefore, in this study, the start-up performance and process kinetics of the two-stage PN/A reactor were evaluated. The results showed that increasing the NLR during the stabilization period 60 d after initiation resulted in a NAR of 96.93% and $[NO_2^- - N]_{Eff}/[NH_4^+ - N]_{Eff}$ of 1.33, which is close to the theoretical value of 1.32 in the PN reactor. The A-stage reactor was successfully started 55 d into operation by inoculation with anaerobic ammonia-oxidizing activated sludge and adding granular activated carbon. The ammonia load showed an S-shaped trend during the start-up of the A-stage reactor.

A logistic model, modified logistic model, modified Gompertz model, and modified Boltzmann model were used to plot the experimental data of ammonia nitrogen removal load. The results showed that the theoretical model correctly explained the kinetic curve of anaerobic ammonia oxidation in the experimental data. The curves in the image show a strong correlation between the model and the dependent variable. The maximum ammonia nitrogen removal load and the maximum instantaneous growth rate of nitrogen removal were obtained by using a growth kinetic model analysis. The maximum ammonia nitrogen removal load and the maximum instantaneous growth rate of nitrogen removal were obtained by using a growth kinetic model analysis. This helps to predict the performance of ammonia nitrogen removal in actual operation and improves the operational management and control of the reactor. Certain results were obtained during the study; however, the short time period over which the experiment was performed means that problems may not have been thoroughly elucidated, and further research and exploration are required to provide more reliable technical support for practical engineering use in the future. The recommendations are as follows:

- The results of this experiment were based on an artificial water distribution, and further in-depth research is required to determine the water quality in an actual complex.
- (2) This experiment has not yet been conducted from a microbial point of view, and followup experiments should be supplemented with other techniques such as scanning electron microscopy and DNA identification.

- (3) Increasing the load in an anaerobic ammonia oxidation reactor will result in pipe blockage, impacting the performance of the reactor, and this effect should be noted during any further experimental processes.
- (4) The combined processes of nitrosation and anaerobic ammonia oxidation produce nitrate nitrogen, and subsequent research could address the application of this reaction in further analysis and research.

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