

Supplementary information

**Ecotoxicity Assessment of the Water Extracts from
Metal-Contaminated and Hydrocarbon-Contaminated
Soils**

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Table S1 Ecotoxicological tests of the soils

Contact	Organism	Endpoint of the measurement	Standard bioassay	References
Direct contact (whole soil)	<i>Avena sativa</i> L.	Growth	ISO/DIS 15799, 1999	[1]
	<i>Brassica rapa</i> L.	Growth	ISO/DIS 15799, 1999	
	<i>Eisenia fetida</i>	Mortality	OECD 207, 1984	
	<i>Eisenia fetida</i>	Avoidance behavior	ISO/DIS 17512-1, 2008	
	Folsomia candida	Mortality and reproduction	OECD, 2009	[2]
		Chronic toxicity tests	ISO, 2014	[3]
	Radish (<i>Raphanus lativus</i> L.)	Germination	ASTM, 2014	[4]
	<i>Vibrio fischeri</i> (Microtox)	Bioluminescence inhibition	Environmental, 1998	[5]
	<i>Brassica juncea</i>	Seed germination and growth	OECD, 2006	
	<i>Escherichia coli</i>	SOS-Induction Factor	SOS-Chromotest kit	
	<i>Vibrio fischeri</i> (Microtox)	Bioluminescence inhibition	-	[6]
	Wheat (<i>Triticum aestivum</i> L.)	14-day root elongation	-	[7]
	Rape (<i>Brassica napus</i> L.)	Seed germination rate, seedling growth in 7-day seed germination tests.	-	[8]
	Cucumber (<i>Cucumis Sativus</i> L.)			
	Garden pea (<i>Pisum sativum</i>)	Root nodulation and leghaemoglobin assay	-	[9]
	<i>Eisenia fetida</i>	Acute, chronic and genotoxicity assays	OECD, 1984, 2004	
	Wheat (<i>Triticum aestivum</i>)	5 d-shoot and root elongation assays and 14 d-shoot biomass measurement	ISO 11269-1; 11269-2, 2012	[10]
	Sorghum (<i>Sorghum bicolor</i>)			
	Tomato (<i>Lycopersicon esculentum</i>)			
	Lettuce (<i>Lactuca sativa</i>)			
	Garden cress (<i>Lepidium sativum</i>)			
	White mustard (<i>Sinapis alba</i>)			
	<i>Lactuca sativa</i>	Germination and growth	ISO, 2005	[11]

		inhibition (14 d)		
	<i>Eisebua fetida</i>	Survival (14 d)	ISO, 1993	
	<i>Folsomia candida</i>	Avoidance (20 min)	Not standardized	
	<i>Folsomia candida</i>	Reproduction (40 d)	ISO, 1999	
Indirect contact (Soil-water extract)	<i>Avena sativa</i> L.	Germination index (GI)	Fuentes et al., 2004; Alvarenga et al., 2012	DIN 38414-S4 (1984).[1]
	<i>Brassica rapa</i> L.	GI (%)= [RSG×RRG]/100		
	<i>Lepidium sativum</i> L.			
	<i>Vibrio fischeri</i>	Bioluminescence inhibition	ISO 11348-2, 1998	
	<i>Daphnia magna</i>	Immobilization/mortality	ISO 6341, 1996	
	<i>Vibrio fischeri</i> (Microtox)	Bioluminescence inhibition	EN 12457-2 protocol, EC 2002.	Liquid-to-solid ratio of 100 g/L [2]
	<i>Vibrio fischeri</i> (Microtox)	Bioluminescence inhibition	-	The leaching solution comprised a 7.0 g soil sample mixed with 21 mL NaCl solution 2 % (w/v) heated to 50 °C and centrifuged at 160 rpm for 90 min. [5]
	<i>Vibrio fischeri</i> (Microtox)	Bioluminescence inhibition	Microbics Corporation, 1995.	Soil elutriates were obtained after mixing 15 g of soil and 30 g Milli Q water for 24 h. [6]
	<i>Luctuca sativa</i> L.	Germination, root and shoot elongation and fresh biomass	Filter paper test. OECD 208, 2006.	Solid-to-water ratio of 1:10 (m/V) according to DIN 38414-S4 (1984). [12]
	<i>Avena sativa</i> L.			
	Microalgae (<i>Pseudokirchneriella subcapitata</i>)	Growth inhibition	OECD 201, 1984.	
	Microcrustacean (<i>Daphnia magna</i>)	Immobilization	ISO 6341, 1996.	
	<i>Azomonas agilis</i>	Dehydrogenase activity	Hungarian standard (HS 21978-30)	For soil extraction 40 g of the soil sample was suspended in 80 ml deionised water. [13]
	Mustard seeds <i>Sinapis alba</i>	Length of germinated root and shoot	Hungarian standard (HS 21978-8)	

	<i>Panagrellus redivivus</i>	Reproduction inhibition	-	
	<i>Tetrahymena pyriformis</i>	Reproduction inhibition	-	
	Mustard seeds <i>Sinapis alba</i>	Root and shoot elongation	-	
	Chlorococcal algae <i>Desmodesmus subspicatus</i>	Growth inhibition	ISO 8692, 2004	Czech legislation for characterization of waste and contaminated soils. Solid-to-water ratio of 1:10 (m/V) according to DIN 38414-S4 (1984). [14]
	<i>Brassica chinensis</i>	Seed germination and root elongation	Wong et al., 2001	Sludge extracts were prepared by shaking 2 g fresh sewage sludge with 20 ml of milli-Q water. [15]
	<i>Vibrio fischeri</i>	Luminescence inhibition (30 min)	ISO 1998a, 1998b	[11]
	Micro-crustaceans <i>Daphnia magna</i>	Immobilization (48 h)	ISO, 1996	
	Algae <i>Pseudokirchneriella subcapitata</i>	Growth inhibition (72 h)	ISO, 2004	
	Rotifers <i>Brachionus calyciflorus</i>	Growth inhibition (48 h)	ISO 2008a, 2008b	
	Animals Mouse lymphoma cells	Chromosomic change (24 h)	Not standardized	
	Zebrafish larvae	Developmental neurotoxicity	-	The village soil leachate composed of 100 mL deionized water and 100 g village soil sample (1 g/mL) was shaken at 145 rpm on a thermostatic oscillator at 28°C (3d). [16]
	Zebrafish embryos	Developmental toxicity and thyroid hormone assay	-	100 g of village soil samples was added 100 mL of ultrapure water (1 g/mL). [17]
	Zebrafish embryos	Developmental toxicity	-	[18]

	Hepatocellular carcinoma HepG2 cell	Cancer cell metastasis	-	Following filtration with a 100-mesh sieve, a 100 g sample was added to a sealed beaker with 100 mL double-distilled water. [19]
	<i>Vibrio fischeri</i>	Bioluminescence inhibition	-	DIN 38414-S4 [20]
	<i>Lactuca sativa</i>	Germination; Root elongation	-	
	<i>Raphanus sativus</i>	Germination; Root elongation	-	
	<i>Dictyostelium discoideum</i>	Mortality; Endocytosis; Replication	-	

Reference

- Alvarenga, P.; Palma, P.; de Varennes, A.; Cunha-Queda, A.C. A contribution towards the risk assessment of soils from the São Domingos Mine (Portugal): Chemical, microbial and ecotoxicological indicators. *Environ. Pollut.* **2012**, *161*, 50-56.
- Kołtowski, M.; Charmsa, B.; Skubiszewska-Zięba, J.; Oleszczuk, P. Effect of biochar activation by different methods on toxicity of soil contaminated by industrial activity. *Ecotox. Environ. Safe.* **2017**, *136*, 119-125.
- Lin, X.L.; Sun, Z.J.; Zhao, L.; Ma, J.; Li, X.; He, F.; Hou, H. Toxicity of exogenous hexavalent chromium to soil-dwelling springtail *Folsomia candida* in relation to soil properties and aging time. *Chemosphere* **2019**, *224*, 734-742.
- Yavuz, B.; Januszewski, B.; Chen, T.F.; Delgado, A.G.; Westerhoff, P.; Rittmann, B. Using radish (*Raphanus lativus* L.) germination to establish a benchmark dose for the toxicity of ozonated-petroleum byproducts in soil. *Chemosphere* **2023**, *313*, 137382.
- Wang, S.; Cheng, F.L.; Shao, Z.G.; Wu, B.; Guo, S.H. Effects of thermal desorption on ecotoxicological characteristics of heavy petroleum-contaminated soil. *Sci. Total Environ.* **2023**, *857*, 159405.
- Domínguez, C.M.; Ventura, P.; Checa-Fernández, A.; Santos, A. Comprehensive study of acute toxicity using Microtox® bioassay in soils contaminated by lindane wastes. *Sci. Total Environ.* **2023**, *856*, 159146.
- Lin, L.; Zhu, B.J.; Qu, X.Z.; Gu, X.Y. Are Ni-Cd toxicity models derived from simple bioassay applicable to natural soils? A bioassay-MSMs coupling approach. *J. Hazard. Mater.* **2022**, *440*, 129830.
- Li, Y.Y.; Tian, X.Y.; Liang, J.L.; Chen, X.L.; Ye, J.Y.; Liu, Y.S.; Liu, Y.Y.; Wei, Y.M. Remediation of hexavalent chromium in contaminated soil using amorphous iron pyrite: Effect on leachability, bioaccessibility, phytotoxicity and long-term stability. *Environ. Pollut.* **2020**, *264*, 114804.
- Sivaram, A.K.; Logeshwaran, P.; Lockington, R.; Naidu, R.; Megharaj, M. Phytoremediation efficacy assessment of polycyclic aromatic hydrocarbons contaminated soils using garden pea (*Pisum sativum*) and earthworms (*Eisenia fetida*). *Chemosphere* **2019**, *229*, 227-235.
- Delerue, F.; Masfaraud, J.F.; Lascourrèges, J.F.; Atteia, O. A multi-site approach to investigate the role of toxicity and confounding factors on plant bioassay results. *Chemosphere* **2019**, *219*, 482-492.

11. Lors, C.; Ponge, J.F.; Aldaya, M.M.; Damidot, D. Comparison of solid and liquid-phase bioassays using ecoscores to assess contaminated soils. *Environ. Pollut.* **2011**, *159*, 2974–2981.
12. Santos, E.S.; Abreu, M.M.; de Varennes, A.; Macías, F.; Leitão, S.; Cerejeira, M.J. Evaluation of chemical parameters and ecotoxicity of a soil developed on gossan following application of polyacrylates and growth of *Spergularia purpurea*. *Sci. Total Environ.* **2013**, *461–462*, 360–370.
13. Leitgib, L.; Kálmán, J.; Gruiz, K. Comparison of bioassays by testing whole soil and their water extract from contaminated sites. *Chemosphere* **2007**, *66*, 428–434.
14. Kočí, V.; Mocová, K.; Kulovaná, M.; Vosáhllová, S. Phytotoxicity tests of solid wastes and contaminated soils in the Czech Republic. *Environ. Sci. Pollut. Res.* **2010**, *17*, 611–623.
15. Wong, J.W.C.; Li, K.; Fang, M.; Su, D.C. Toxicity evaluation of sewage sludges in Hong Kong. *Environ. Int.* **2001**, *27*, 373–380.
16. Yang, F.L.; Yun, Y.; Li, G.K.; Sang, N. Heavy metals in soil from gangue stacking areas increase children health risk and causes developmental neurotoxicity in zebrafish larvae. *Sci. Total Environ.* **2021**, *794*, 148629.
17. Yang, F.L.; Li, G.K.; Sang, N. Embryonic exposure to soil samples from a gangue stacking area induces thyroid hormone disruption in zebrafish. *Chemosphere* **2019**, *236*, 124337.
18. Legler, J.; van Velzen, M.; Ceniñ, P.H.; Houtman, C.J.; Lamoree, M.H.; Wegener, J.W. Effect-directed analysis of municipal landfill soil reveals novel developmental toxicants in the zebrafish *Danio rerio*. *Environ. Sci. Technol.* **2011**, *45*, 8552–8558.
19. Yun, Y.; Gao, R.; Yue, H.F.; Liu, X.F.; Li, G.K.; Sang, N. Polycyclic aromatic hydrocarbon (PAH)-containing soils from coal gangue stacking areas contribute to epithelial to mesenchymal transition (EMT) modulation on cancer cell metastasis. *Sci. Total Environ.* **2017**, *580*, 632–640.
20. Rodríguez-Ruiz, A.; Asensio, V.; Zaldibar, B.; Soto, M.; Marigómez, I. Toxicity assessment through multiple endpoint bioassays in soils posing environmental risk according to regulatory screening values. *Environ. Sci. Pollut. Res.* **2014**, *21*, 9689–9708.