

Comparison of the Efficiency of Deammonification under Different DO concentration in a Laboratory-Scale Sequencing Batch Reactor

Hussein Ezzi Al-Hazmi ^{1*}, Zhixuan Yin ², Dominika Grubba ¹, Joanna Majtacz ¹ and Jacek Makinia ¹

¹ Affiliation 1 Faculty of Civil and Environmental Engineering, Gdańsk University of Technology, Narutowicza Street 11/12, Gdańsk 80-233 Poland; hussienalhazmi@yahoo.com, dominika.grubba@pg.edu.pl, joamajta@pg.edu.pl, jmakinia@pg.edu.pl

² School of Environmental and Municipal Engineering, Qingdao University of Technology, 11 Fushun Road, Qingdao 266033, PR China (zhixuanyin@outlook.com)

* Correspondence: hussein.alhazmi66@gmail.com; Tel.: +48 730567469

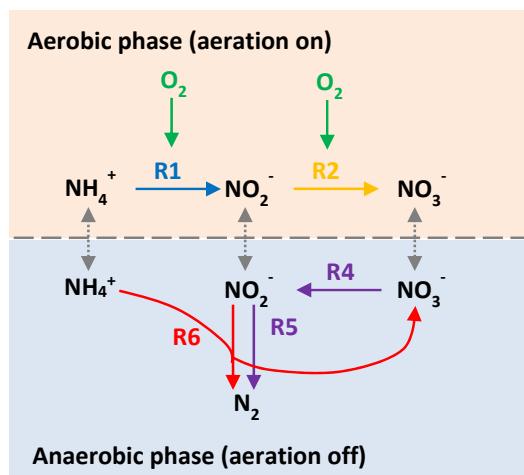


Figure S1. The conceptual model for nitrogen transformation under aeration on/off conditions. (R1: Aerobic growth of AOB; R2: Aerobic growth of NOB; R4-5: Anoxic growth of HDB; R6: Anaerobic growth of AnAOB).

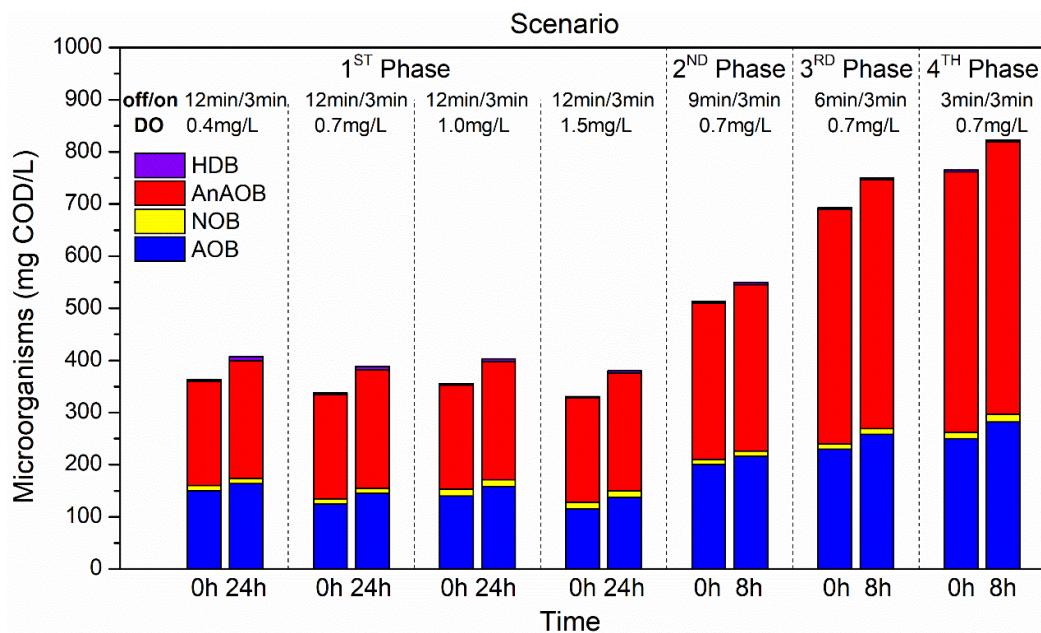


Figure S2. The initial microorganisms' composition and their predicted value after the batch test under different scenarios. (The initial composition was firstly estimated considering the results of our previous study (Al-Hazmi et al., 2020), metagenomic analysis and the measured MLVSS, and then the composition was slightly adjusted in during the preliminary model simulations).

Reference:

Al-Hazmi, H. E., Lu, X., Majtacz, J., et al. Optimization of the Aeration Strategies in a Deammonification Sequencing Batch Reactor for Efficient Nitrogen Removal and Mitigation of N₂O Production[J]. *Environmental Science & Technology*, **2020**, 55(2): 1218-1230.

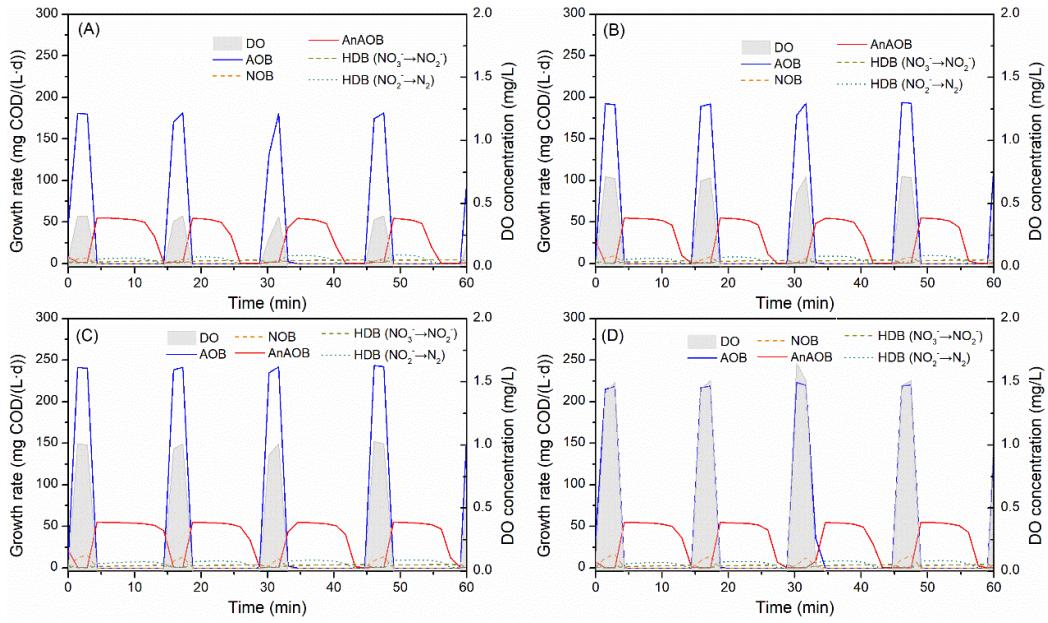


Figure S3. The variation in the simulated growth rates of different microorganisms with different DO set point of A) 0.4 mg O₂/L, B) 0.7 mg O₂/L, C) 1.0 mg O₂/L, D) 1.5 mg O₂/L at the same intermittent aeration mode off/on (12/3 min) conditions.

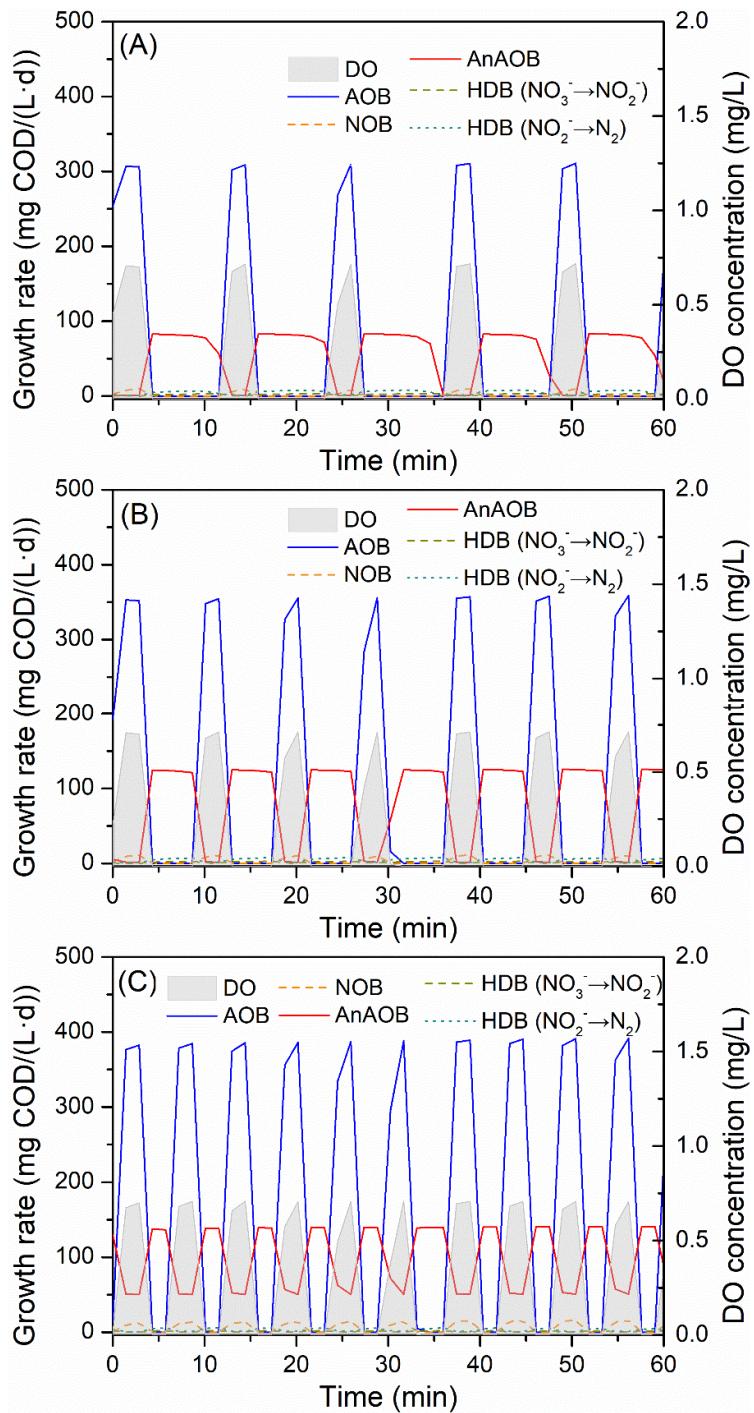


Figure S4. The variation in the simulated growth rates of different microorganisms with different intermittent aeration mode off/on A) 9min/3min, B) 6min/3min, C) 3min/3 min) at the same DO set point 0.7 mg O₂/L.

Table S1. Definition of the state variables of the proposed model.

No.	Symbol	State variable	Unit
1	S_I	Inert soluble matter	mg COD/L
2	S_S	Readily biodegradable substrate	mg COD/L
3	S_{O_2}	Dissolved oxygen	mg O ₂ /L
4	$S_{NO_3^-}$	Nitrate	mg N/L
5	$S_{NO_2^-}$	Nitrite	mg N/L
6	S_{N_2}	Nitrogen gas	mg N/L
7	$S_{NH_4^+}$	Ammonium and free ammonia	mg N/L
8	S_{ND}	Soluble biodegradable organic nitrogen	mg N/L
9	X_{AOB}	Aerobic ammonium-oxidizing biomass(AOB)	mg COD/L
10	X_{NOB}	Nitrite-oxidizing biomass(NOB)	mg COD/L
11	X_H	Aerobic heterotrophs biomass	mg COD/L
12	X_{HDB}	Anoxic heterotrophic denitrifying biomass (HDB)	mg COD/L
13	X_{AnAOB}	Anaerobic ammonium-oxidizing biomass (AnAOB)	mg COD/L
14	X_S	Slowly biodegradable substrate	mg COD/L
15	X_P	Particulate unbiodegradable matter	mg COD/L
16	X_{ND}	Particulate biodegradable organic nitrogen	mg N/L

Table S2. The matrix of the proposed model.

Component → i		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Process ↓ j		S_I	S_S	S_{O_2}	$S_{NO_3^-}$	$S_{NO_2^-}$	S_{N_2}	$S_{NH_4^+}$	S_{ND}	X_{AOB}	X_{NOB}	X_H	X_{HDB}	X_{AnAOB}	X_S	X_P	X_{ND}
R1	Aerobic growth of AOB, $NH_4^+ \rightarrow NO_2^-$			$-\frac{3.43 - Y_{AOB}}{Y_{AOB}}$		$\frac{1}{Y_{AOB}}$		$-i_{XB} - \frac{1}{Y_{AOB}}$		1							
R2	Aerobic growth of NOB, $NO_2^- \rightarrow NO_3^-$			$-\frac{1.14 - Y_{NOB}}{Y_{NOB}}$	$\frac{1}{Y_{NOB}}$	$-\frac{1}{Y_{NOB}}$		$-i_{XB}$		1							
R3	Aerobic growth of Heterotrophs		$-\frac{1}{Y_H}$	$-\frac{1 - Y_H}{Y_H}$				$-i_{XB}$			1						
R4	Anoxic growth of HDB, $NO_3^- \rightarrow NO_2^-$		$-\frac{1}{Y_{HDB}}$		$-\frac{1 - Y_{HDB}}{1.14Y_{HDB}}$	$\frac{1 - Y_{HDB}}{1.14Y_{HDB}}$		$-i_{XB}$				1					
R5	Anoxic growth of HDB, $NO_2^- \rightarrow N_2$		$-\frac{1}{Y_{HDB}}$			$-\frac{1 - Y_{HDB}}{1.71Y_{HDB}}$	$\frac{1 - Y_{HDB}}{1.71Y_{HDB}}$	$-i_{XB}$			1						
R6	Anaerobic growth of AnAOB, $NH_4^+ + NO_2^- \rightarrow N_2 + NO_3^-$				$\frac{1}{1.41}$	$-\frac{1}{Y_{AnAOB}} - 1.41$	$\frac{2}{Y_{AnAOB}}$	$-i_{XB} - \frac{1}{Y_{AnAOB}}$				1					
R7	Decay of AOB									-1				$1 - f_P$	f_P	$i_{XB} - f_P i_{XP}$	
R8	Decay of NOB									-1				$1 - f_P$	f_P	$i_{XB} - f_P i_{XP}$	
R9	Decay of Heterotrophs									-1				$1 - f_P$	f_P	$i_{XB} - f_P i_{XP}$	
R10	Decay of HDB										-1			$1 - f_P$	f_P	$i_{XB} - f_P i_{XP}$	
R11	Decay of AnAOB											-1		$1 - f_P$	f_P	$i_{XB} - f_P i_{XP}$	
R12	Hydrolysis of entrapped organics	f_I	$1 - f_I$					i_{XS}							-1		
R13	Hydrolysis of entrapped organic N								1							-1	
R14	Ammonification of soluble organic N								1	-1							

Table S3. The Kinetic equations of the proposed model.

Process ↓ j		Rate equation
R1	Aerobic growth of AOB, $\text{NH}_4^+ \rightarrow \text{NO}_2^-$	$\mu_{AOB}^{R1} \frac{S_{\text{NH}_4^+}}{K_{\text{NH}_4^+}^{R1} + S_{\text{NH}_4^+}} \frac{S_{O_2}}{K_{O_2}^{R1} + S_{O_2}} X_{AOB}$
R2	Aerobic growth of NOB, $\text{NO}_2^- \rightarrow \text{NO}_3^-$	$\mu_{NOB}^{R2} \frac{S_{\text{NO}_2^-}}{K_{\text{NO}_2^-}^{R2} + S_{\text{NO}_2^-}} \frac{S_{O_2}}{K_{O_2}^{R2} + S_{O_2}} X_{NOB}$
R3	Aerobic growth of Heterotrophs	$\mu_H^{R3} \frac{S_S}{K_S^{R3} + S_S} \frac{S_{O_2}}{K_{O_2,H} + S_{O_2}} X_H$
R4	Anoxic growth of HDB, $\text{NO}_3^- \rightarrow \text{NO}_2^-$	$\mu_{HDB}^{R4} \frac{S_S}{K_S^{R4} + S_S} \frac{S_{\text{NO}_3^-}}{K_{\text{NO}_3^-}^{R4} + S_{\text{NO}_3^-}} \frac{K_{i,O_2,HDB}}{K_{i,O_2,HDB} + S_{O_2}} \eta_g^{R4} X_{HDB}$
R5	Anoxic growth of HDB, $\text{NO}_2^- \rightarrow \text{N}_2$	$\mu_{HDB}^{R5} \frac{S_S}{K_S^{R5} + S_S} \frac{S_{\text{NO}_2^-}}{K_{\text{NO}_2^-}^{R5} + S_{\text{NO}_2^-}} \frac{K_{i,O_2,HDB}}{K_{i,O_2,HDB} + S_{O_2}} \eta_g^{R5} X_{HDB}$
R6	Anaerobic growth of AnAOB, $\text{NH}_4^+ + \text{NO}_2^- \rightarrow \text{N}_2 + \text{NO}_3^-$	$\mu_{AnAOB}^{R6} \frac{S_{\text{NH}_4^+}}{K_{\text{NH}_4^+}^{R6} + S_{\text{NH}_4^+}} \frac{S_{\text{NO}_2^-}}{K_{\text{NO}_2^-}^{R6} + S_{\text{NO}_2^-}} \frac{K_{i,O,AnAOB}}{K_{i,O,AnAOB} + S_{O_2}} X_{AnAOB}$
R7	Decay of AOB	$b_{AOB} X_{AOB}$
R8	Decay of NOB	$b_{NOB} X_{NOB}$

R9	Decay of Heterotrophs	$b_H X_H$
R10	Decay of HDB	$b_{HDB} X_{HDB}$
R11	Decay of AnAOB	$b_{AnAOB} X_{AnAOB}$
R12	Hydrolysis of entrapped organics	$k_h \frac{X_S/X_H}{K_X + (X_S/X_H)} \left[\left(\frac{S_{O_2}}{K_{O_2,H} + S_{O_2}} \right) + \eta_{hANOX} \left(\frac{K_{O_2,H}}{K_{O_2,H} + S_{O_2}} \right) \left(\frac{S_{NO_3^-}}{K_{NO_3^-} + S_{NO_3^-}} \right) \right] X_H$
R13	Hydrolysis of entrapped organic N	$r_{12}(X_{ND}/X_S)$
R14	Ammonification of soluble organic N	$K_a \cdot S_{ND} \cdot X_H$

Table S4. Stoichiometric parameters and conversion factors.

Symbol	Definition	Unit	Reference range	Default	Calibrated	References
Model stoichiometry						
Y_{AOB}	AOB yield	g COD/g N	0.15-0.24	0.18		1-13
Y_{NOB}	NOB yield	g COD/g N	0.041-0.06	0.06		1,3,5,6,7, 8,13,15
Y_H	heterotrophic yield	g COD/g COD	0.5-0.67	0.666		1,3,7,8,13, 14,16,17
Y_{HDB}	HDB yield	g COD/g COD	0.5-0.67	0.666		1,3,7,8,13, 14,16,17
Y_{AnAOB}	AMX yield	g COD/g N	0.114-0.17	0.17		13,15, 18-24
f_p	fraction of biomass leading to particulate products	g COD/g COD	0.08-0.2	0.08		1,3,7,13, 14,22
f_I	Production of soluble inerts in hydrolysis	g COD/g COD	0.02	0.02		14
Composite variable stoichiometry						
icv	XCOD/VSS	g COD/g VSS	1.42-1.48	1.48		14,25
fbod	BOD ₅ /BODultimate ratio	-	0.66	0.66		14
i_{XB}	N content of active biomass	g N/g COD	0.07-0.875	0.086		1-4,6-15, 18
i_{XP}	N content of endogenous/inert mass	g N/g COD	0.02-0.068	0.06		1,3,13,14
i_{XS}	nitrogen content of COD in slowly biodegradable substrate	g N/g COD	0.015	0.015		14

References: ¹(Hiatt and Grady, 2008); ²(Law et al., 2012); ³(Ni et al., 2011); ⁴(Mampaey et al., 2011); ⁵(Wiesmann et al., 1994); ⁶(Mampaey et al., 2013); ⁷(Kampschreur et al., 2007); ⁸(Ni et al., 2013a); ⁹(Ni et al., 2013b); ¹⁰(Peng et al., 2015a and 2015b); ¹¹(Pocquet et al., 2013); ¹²(Guo and Vanrolleghem, 2014); ¹³(von Schulthess and Gujer, 1996); ¹⁴(Henze et al., 2000); ¹⁵(Volcke et al., 2010); ¹⁶(Samie et al., 2011); ¹⁷(Pan et al., 2013); ¹⁸(Takács et al., 2007); ¹⁹(Koch et al., 2000); ²⁰(Strous et al., 1998); ²¹(Bi et al., 2015); ²²(Dapena-Mora et al., 2004); ²³(Hao et al., 2002); ²⁴(Ni et al., 2009); ²⁵(Comeau, 2008)

Table S5. Kinetic parameters and their values (*value at T = 20°C).

Symbol	Definition	Unit	Reference range	Default	Calibrated	References
AOB						
μ_{AOB}^{R1}	the maximum specific growth rate	1/d	0.39-5.76	1.2		1-17
$K_{NH_4^+}^{R1}$	ammonia half saturation coefficient	g N/m ³	0.02-27.5	1		1,4,5, 8-12, 18-22
$K_{O_2}^{R1}$	oxygen half saturation coefficient	g O ₂ /m ³	0.02-2.1	0.4		1,2,4-6, 8-15, 18-20, 22,23
b_{AOB}	AOB decay rate	1/d	0.071-0.13	0.096		2,5,8, 9,19
NOB						
μ_{NOB}^{R2}	the maximum specific growth rate	1/d	0.38-1.44	1.44		2,3,5, 6,8,9, 16,17,19,
$K_{NO_2^-}^{R2}$	nitrite half saturation coefficient	g N/m ³	0.16-5.5	1.2		5,8,9, 16,19,22, 24
$K_{O_2}^{R2}$	oxygen half saturation coefficient	g O ₂ /m ³	0.16-2.2	0.74		2,5,6, 8,9,16, 19,22-25
b_{NOB}	NOB decay rate	1/d	0.06-0.096	0.096		2,5,8, 9,19
Heterotrophs						
μ_H^{R3}	heterotrophic maximum specific growth rate	1/d	2-6.25	6		1-3,8,9
K_S^{R3}	readily biodegradable substrate half saturation coefficient	g COD/m ³	2-20	2		1-3,5, 9,26,27
$K_{O_2,H}$	oxygen half saturation coefficient	g O ₂ /m ³	0.1-2	0.2		1-3,5, 8,9,26
b_H	aerobic heterotrophic	1/d	0.2-0.62	0.62		1-3,5, 9

Symbol	Definition	Unit	Reference range	Default	Calibrated	References
	decay rate					
HDB						
μ_{HDB}^{R4}	heterotrophic maximum specific growth rate	1/d	2-6.25	6		1-3,8,9
μ_{HDB}^{R5}	heterotrophic maximum specific growth rate	1/d	2-6.25	6		1-3,8,9
K_S^{R4}	readily biodegradable substrate half saturation coefficient	g COD/m ³	2-20	20		1-3,5, 9,26,27
K_S^{R5}	readily biodegradable substrate half saturation coefficient	g COD/m ³	2-20	2		1-3,5, 9,26,27
$K_{NO_3^-}^{R4}$	nitrate half saturation coefficient	g N/m ³	0.2-0.5	0.5		1-3,5, 8,9,26, 28
$K_{NO_2^-}^{R5}$	nitrite half saturation coefficient	g N/m ³	0.06-8	0.2		2,3,5, 8,9,26, 28
$K_{i,O_2,HDB}$	oxygen inhibition coefficient	g O ₂ /m ³	0.1-2	0.2		1-3,5, 8,9,26
η_g^{R4}	anoxic growth factor 1, reducing nitrate to nitrite	-	0.029-0.8	0.2		1-3,9,29
η_g^{R5}	anoxic growth factor 2, reducing nitrite to dinitrogen	-	0.075-0.81	0.6		2,3,9, 29,30
b_{HDB}	anoxic heterotrophic decay rate	1/d	0.2-0.62	0.62		1-3,5, 9
AnAOB						

Symbol	Definition	Unit	Reference range	Default	Calibrated	References
μ_{AnAOB}^{R6}	the maximum specific growth rate	1/d	0.019-0.14	0.03	0.14	3,16-17, 27,31-36
$K_{NH_4^+}^{R6}$	ammonia half saturation coefficient	g N/m ³	0.03-21	0.07		16,31,32, 34,37-39
$K_{NO_2^-}^{R6}$	nitrite half saturation coefficient	g N/m ³	0.005-2	0.05		16,31-34, 38,39,40
$K_{i,O,AnAOB}$	oxygen inhibition coefficient	g O ₂ /m ³	0.01-0.4	0.01	0.4	31,37,38
b_{AnAOB}	anammox bacteria decay rate	1/d	0.0003-0.016	0.003		3,27, 31-34
Hydrolysis						
k_h	maximum specific hydrolysis rate	1/d	1.5-4.5	3		1,2,5 9,30
K_X	slowly biodegradable substrate half saturation coefficient	g COD/g COD	0.03-1	0.03		1,2,5 9
η_{hANOX}	anoxic hydrolysis factor	-	0.4-0.8	0.4		1,2
Ammonification						
K_a	ammonification rate	m ³ /g COD/d	0.08-0.1608	0.08		1,2

References: ¹(Henze et al., 2000); ²(Hiatt and Grady, 2008); ³(Samie et al., 2011); ⁴(Law et al., 2012); ⁵(Ni et al., 2011); ⁶(Mampaey et al., 2013); ⁷(Hellinga et al., 1998); ⁸(Kampschreur et al., 2007); ⁹(Ni et al., 2013a); ¹⁰(Ni et al., 2013b); ¹¹(Ni et al., 2014b); ¹²(Peng et al., 2015a and 2015b); ¹³(Pocquet et al., 2013); ¹⁴(Guo and Vanrolleghem, 2014); ¹⁵(Pocquet et al., 2016); ¹⁶(Van Hulle et al., 2012); ¹⁷(Ni et al., 2014a); ¹⁸(Mampaey et al., 2011); ¹⁹(Wiesmann et al., 1994); ²⁰(Van Hulle et al., 2007); ²¹(Pynnaert, 2003); ²²(Cao et al., 2017); ²³(Guisasola et al., 2005); ²⁴(Hydromantis, 2014); ²⁵(Isanta et al., 2015); ²⁶(von Schulthess and Gujer, 1996); ²⁷(Liu et al., 2016); ²⁸(Pan et al., 2013); ²⁹(Pan et al., 2015); ³⁰(Mannina et al., 2018); ³¹(Koch et al., 2000); ³²(Dapena-Mora et al., 2004); ³³(Hao et al., 2002); ³⁴(Ni et al., 2009); ³⁵(Lotti et al., 2015); ³⁶(Bae et al., 2010); ³⁷(Strous et al., 1998); ³⁸(Bi et al., 2015); ³⁹(Stewart et al., 2017); ⁴⁰(Lotti et al., 2014)

Table S6. Sensitivity coefficients calculated for the adjusted stoichiometric parameters.

	NH ₄ ⁺	NO ₂ ⁻	NO ₃ ⁻		NH ₄ ⁺	NO ₂ ⁻	NO ₃ ⁻
Y_{AOB}	1.1240	-3.7559	-0.0680	f_P	-0.0349	0.0053	0.0008
Y_{NOB}	-0.0024	0.5836	-0.2832	f_I	-0.0019	0.0552	0.0444
Y_H	0.0091	-0.5809	-0.0142	i_{XB}	0.2848	-0.0271	0.0001
Y_{HDB}	-0.0021	0.0040	0.0022	i_{XP}	0.0330	0.0044	-0.0011
Y_{AnAOB}	0.5607	2.7116	0.0459				

Table S7. Sensitivity coefficients calculated for the adjusted kinetic parameters.

	NH ₄ ⁺	NO ₂ ⁻	NO ₃ ⁻		NH ₄ ⁺	NO ₂ ⁻	NO ₃ ⁻
AOB							
μ_{AOB}^{R1}	-1.2539	3.9972	0.0734	$K_{O_2}^{R1}$	0.3525	-1.2334	-0.0066
$K_{NH_4^+}^{R1}$	0.0049	-0.0427	0.0009	b_{AOB}	0.0161	-0.0460	0.0003
NOB							
μ_{NOB}^{R2}	0.0016	-0.6168	0.3008	$K_{O_2}^{R2}$	-0.0009	0.3368	-0.1627
$K_{NO_2^-}^{R2}$	-0.0022	0.0505	-0.0224	b_{NOB}	0.0004	0.0073	-0.0049
Heterotrophs							
μ_H^{R3}	-0.0008	0.0005	0.0004	$K_{O_2,H}^{R3}$	-0.0014	0.0119	0.0009
K_S^{R3}	0.0005	0.0010	0.0004	b_H	0.2815	0.1174	0.0003
HDB							
μ_{HDB}^{R4}	0.0000	0.0013	-0.0013	$K_{NO_2^-}^{R5}$	0.0000	0.0000	0.0000
μ_{HDB}^{R5}	0.0000	-0.0032	0.0000	$K_{i,O_2,HDB}^{R5}$	-0.0014	0.0119	0.0009
K_S^{R4}	0.0005	0.0010	0.0004	η_g^{R4}	0.0000	0.0013	-0.0013
K_S^{R5}	0.0005	0.0010	0.0004	η_g^{R5}	0.0000	-0.0032	0.0000
$K_{NO_3^-}^{R4}$	0.0000	0.0000	0.0000	b_{HDB}	0.0004	0.0000	0.0000
AnAOB							
μ_{AnAOB}^{R6}	-0.5824	-3.1105	0.1033	$K_{i,O,AnAOB}^{R6}$	-0.0870	-0.4533	0.0152
$K_{NH_4^+}^{R6}$	0.0024	0.0030	-0.0012	b_{AnAOB}	0.0007	0.0004	0.0000
$K_{NO_2^-}^{R6}$	0.0021	0.0122	-0.0014				
Hydrolysis							
k_h	-0.0057	-0.0272	-0.0015	η_{hANOX}	-0.0055	-0.0080	-0.0003
K_X	0.0033	0.0188	0.0009				
Ammonification							
K_a	-0.8837	-0.0038	0.0000				

References

- Bae, H., Park, K.-S., Chung, Y.-C., Jung, J.-Y., 2010. Distribution of anammox bacteria in domestic WWTPs and their enrichments evaluated by real-time quantitative PCR. *Process Biochemistry* 45, 323–334. <https://doi.org/10.1016/j.procbio.2009.10.004>
- Bi, Z., Takekawa, M., Giri, P., Soda, S., Zhou, J., Qiao, S., Ike, M., Zhen, B., Takekawa, M., Park, G., Soda, S., Qiao, S., Ike, M., 2015. Effects of the C/N ratio and bacterial populations on nitrogen removal in the simultaneous anammox and heterotrophic denitrification process: Mathematic modeling and batch experiments. *Chemical Engineering Journal* 280, 606–613. <https://doi.org/10.1016/j.cej.2015.06.028>
- Cao, Y., van Loosdrecht, M.C.M., Daigger, G.T., 2017. Mainstream partial nitritation-anammox in municipal wastewater treatment: status, bottlenecks, and further studies. *Applied Microbiology and Biotechnology* 101, 1365–1383. <https://doi.org/10.1007/s00253-016-8058-7>
- Dapena-Mora, A., Van Hulle, S.W., Luis Campos, J., Méndez, R., Vanrolleghem, P.A., Jetten, M., 2004. Enrichment of Anammox biomass from municipal activated sludge: experimental and modelling results: Experimental and modelling results of Anammox enrichment. *Journal of Chemical Technology & Biotechnology* 79, 1421–1428. <https://doi.org/10.1002/jctb.1148>
- Guisasola, A., Jubany, I., Baeza, J.A., Carrera, J., Lafuente, J., 2005. Respirometric estimation of the oxygen affinity constants for biological ammonium and nitrite oxidation. *Journal of Chemical Technology and Biotechnology* 80, 388–396. <https://doi.org/10.1002/jctb.1202>
- Guo, L., Vanrolleghem, P.A., 2014. Calibration and validation of an activated sludge model for greenhouse gases no. 1 (ASMG1): Prediction of temperature-dependent N₂O emission dynamics. *Bioprocess and Biosystems Engineering* 37, 151–163. <https://doi.org/10.1007/s00449-013-0978-3>
- Hao, X., Heijnen, J.J., van Loosdrecht, M.C.M., 2002. Sensitivity analysis of a biofilm model describing a one-stage completely autotrophic nitrogen removal (CANON) process. *Biotechnology and Bioengineering* 77, 266–277. <https://doi.org/10.1002/bit.10105>
- Hellinga, C., Schellen, A.A.J.C., Mulder, J.W., Van Loosdrecht, M.C.M., Heijnen, J.J., 1998. The SHARON process: An innovative method for nitrogen removal from ammonium-rich waste water. *Water Science and Technology* 37, 135–142. [https://doi.org/10.1016/S0273-1223\(98\)00281-9](https://doi.org/10.1016/S0273-1223(98)00281-9)
- Henze, M., Gujer, W., Mino, T., van Loosdrecht, M., 2000. Activated Sludge Models ASM1, ASM2, ASM2d and ASM3. IWA Publishing, UK.
- Hiatt, W.C., Grady, C.P.L., 2008. An updated process model for carbon oxidation, nitrification, and denitrification. *Water environment research : a research publication of the Water Environment Federation* 80, 2145–2156. <https://doi.org/10.2175/106143008X304776>
- Hydromantis, 2014. GPS-X Technical Reference.

- Isanta, E., Reino, C., Carrera, J., Pérez, J., 2015. Stable partial nitritation for low-strength wastewater at low temperature in an aerobic granular reactor. *Water Research* 80, 149–158. <https://doi.org/10.1016/j.watres.2015.04.028>
- Kampschreur, M.J., Picioreanu, C., Tan, N., Kleerebezem, R., Jetten, M.S.M., van Loosdrecht, M.C.M., 2007. Unraveling the source of nitric oxide emission during nitrification. *Water Environ Res* 79, 2499–2509. <https://doi.org/10.2175/193864707787976470>
- Koch, G., Egli, K., Van der Meer, J.R., Siegrist, H., 2000. Mathematical modeling of autotrophic denitrification in a nitrifying biofilm of a rotating biological contactor. *Water Science and Technology* 41, 191–198.
- Law, Y., Ni, B.-J., Lant, P., Yuan, Z., 2012. N₂O production rate of an enriched ammonia-oxidising bacteria culture exponentially correlates to its ammonia oxidation rate. *Water Research* 46, 3409–3419. <https://doi.org/10.1016/j.watres.2012.03.043>
- Liu, Y., Sun, J., Peng, L., Wang, D., Dai, X., Ni, B.-J., 2016. Assessment of Heterotrophic Growth Supported by Soluble Microbial Products in Anammox Biofilm using Multidimensional Modeling. *Scientific Reports* 6, 1–11. <https://doi.org/10.1038/srep27576>
- Lotti, T., Kleerebezem, R., Abelleira-Pereira, J.M., Abbas, B., van Loosdrecht, M.C.M., 2015. Faster through training: The anammox case. *Water Research* 81, 261–268. <https://doi.org/10.1016/j.watres.2015.06.001>
- Lotti, T., Kleerebezem, R., Lubello, C., van Loosdrecht, M.C.M., 2014. Physiological and kinetic characterization of a suspended cell anammox culture. *Water Research* 60, 1–14. <https://doi.org/10.1016/j.watres.2014.04.017>
- Mampaey KE, Beuckels B, Kampschreur MJ, Kleerebezem R, van Loosdrecht MCM, Volcke EIP. 2011. Modelling nitrous and nitric oxide emissions by autotrophic ammonium oxidizing bacteria. In: Proceedings IWA/WEF Nutrient Recovery and Management Conference, 2011 Miami, FL, USA, January 9–12.
- Mampaey, K.E., Beuckels, B., Kampschreur, M.J., Kleerebezem, R., van Loosdrecht, M.C.M., Volcke, E.I.P., 2013. Modelling nitrous and nitric oxide emissions by autotrophic ammonia-oxidizing bacteria. *Environmental Technology* 34, 1555–1566. <https://doi.org/10.1080/09593330.2012.758666>
- Mannina, G., Cosenza, A., Ekama, G.A., 2018. A comprehensive integrated membrane bioreactor model for greenhouse gas emissions. *Chemical Engineering Journal* 334, 1563–1572. <https://doi.org/10.1016/j.cej.2017.11.061>
- Ni, B.-J., Chen, Y.-P., Liu, S.-Y., Fang, F., Xie, W.-M., Yu, H.-Q., 2009. Modeling a granule-based anaerobic ammonium oxidizing (ANAMMOX) process. *Biotechnology and Bioengineering* 103, 490–499. <https://doi.org/10.1002/bit.22279>
- Ni, B.-J., Joss, A., Yuan, Z., 2014a. Modeling nitrogen removal with partial nitritation and anammox in one floc-based sequencing batch reactor. *Water Research* 67, 321–329. <https://doi.org/10.1016/j.watres.2014.09.028>
- Ni, B.-J., Peng, L., Law, Y., Guo, J., Yuan, Z., Advanced, 2014b. Modeling of nitrous oxide production by autotrophic ammonia-oxidizing bacteria with multiple production pathways. *Environmental Science and Technology* 48, 3916–3924. <https://doi.org/10.1021/es405592h>

Ni, B.-J., Ruscalleda, M., Pellicer-Nacher, C., Smets, B.F., 2011. Modeling nitrous oxide production during biological nitrogen removal via nitrification and denitrification: Extensions to the general ASM models. *Environmental Science and Technology* 45, 7768–7776. <https://doi.org/10.1021/es201489n>

Ni, B.-J., Ye, L., Law, Y., Byers, C., Yuan, Z., 2013a. Mathematical modeling of nitrous oxide (N₂O) emissions from full-scale wastewater treatment plants. *Environmental science & technology* 47, 7795–803. <https://doi.org/10.1021/es4005398>

Ni, B.-J., Yuan, Z., Chandran, K., Vanrolleghem, P.A., Murthy, S., 2013b. Evaluating four mathematical models for nitrous oxide production by autotrophic ammonia-oxidizing bacteria. *Biotechnology and Bioengineering* 110, 153–163. <https://doi.org/10.1002/bit.24620>

Pan, Y., Ni, B.J., Lu, H., Chandran, K., Richardson, D., Yuan, Z., 2015. Evaluating two concepts for the modelling of intermediates accumulation during biological denitrification in wastewater treatment. *Water Research* 71, 21–31. <https://doi.org/10.1016/j.watres.2014.12.029>

Pan, Y., Ni, B.-J., Yuan, Z., 2013. Modeling Electron Competition among Nitrogen Oxides Reduction and N₂O Accumulation in Denitrification. *Environmental Science and Technology* 47, 11083–11091. <https://doi.org/10.1021/ef501790e>

Peng, L., Ni, B.-J., Ye, L., Yuan, Z., 2015a. The combined effect of dissolved oxygen and nitrite on N₂O production by ammonia oxidizing bacteria in an enriched nitrifying sludge. *Water Research* 73, 29–36. <https://doi.org/10.1016/j.watres.2015.01.021>

Peng, L., Ni, B.-J., Ye, L., Yuan, Z., 2015b. Selection of mathematical models for N₂O production by ammonia oxidizing bacteria under varying dissolved oxygen and nitrite concentrations. *Chemical Engineering Journal* 281, 661–668. <https://doi.org/10.1016/j.cej.2015.07.015>

Pocquet, M., Queinnec, I., Sperandio, M., 2013. Adaptation and identification of models for nitrous oxide (N₂O) production by autotrophic nitrite reduction. In: Proc. 11th IWA Conf. Instrum. Control Autom. ICA2013 Narbonne, France, September 18-20.

Pocquet, M., Wu, Z., Queinnec, I., Sperandio, M., 2016. A two pathway model for N₂O emissions by ammonium oxidizing bacteria supported by the NO/N₂O variation. *Water Research* 88, 948–959. <https://doi.org/10.1016/j.watres.2015.11.029>

Samie, G., Bernier, J., Rocher, V., Lessard, P., 2011. Modeling nitrogen removal for a denitrification biofilter. *Bioprocess and Biosystems Engineering* 34, 747–755. <https://doi.org/10.1007/s00449-011-0524-0>

Stewart, H.A., Al-Omari, A., Bott, C., De Clippeleir, H., Su, C., Takacs, I., Wett, B., Massoudieh, A., Murthy, S., 2017. Dual substrate limitation modeling and implications for mainstream deammonification. *Water Research* 116, 95–105. <https://doi.org/10.1016/j.watres.2017.03.021>

Strous, M., Heijnen, J.J., Kuenen, J.G., Jetten, M.S.M., 1998. The sequencing batch reactor as a powerful tool for the study of slowly growing anaerobic ammonium-oxidizing microorganisms. *Applied microbiology and biotechnology* 50,

589–596.

Takács, I., Vanrolleghem, P.A., Wett, B., Murthy, S., 2007. Elemental balance based methodology to establish reaction stoichiometry in environmental modeling. *Water Science and Technology* 56, 37–41. <https://doi.org/10.2166/wst.2007.606>

Van Hulle, S.W., Volcke, E.I., Teruel, J.L., Donckels, B., van Loosdrecht, M.C., Vanrolleghem, P.A., 2007. Influence of temperature and pH on the kinetics of the Sharon nitritation process. *Journal of Chemical Technology & Biotechnology* 82, 471–480. <https://doi.org/10.1002/jctb>

Van Hulle, S.W.H., Callens, J., Mampaey, K.E., van Loosdrecht, M.C.M., Volcke, E.I.P., 2012. N₂O and NO emissions during autotrophic nitrogen removal in a granular sludge reactor – a simulation study. *Environmental Technology* 33, 2281–2290. <https://doi.org/10.1080/09593330.2012.665492>

Volcke, E.I.P., Picioreanu, C., De Baets, B., van Loosdrecht, M.C.M., 2010. Effect of granule size on autotrophic nitrogen removal in a granular sludge reactor. *Environmental Technology* 31, 1271–1280. <https://doi.org/10.1080/09593330.2013.859711>

von Schulthess, R., Gujer, W., 1996. Release of nitrous oxide (N₂O) from denitrifying activated sludge: Verification and application of a mathematical model. *Water Research* 30, 521–530. [https://doi.org/10.1016/0043-1354\(95\)00204-9](https://doi.org/10.1016/0043-1354(95)00204-9)

Wiesmann, U., Choi, I.S., Dombrowski, E.-M., 1994. Fundamentals of Biological Wastewater Treatment. *Generations Journal Of The American Society On Aging* 6, 400. <https://doi.org/10.1002/9783527609604>