



Article Scaling Up from Leaf to Whole-Plant Level for Water Use Efficiency Estimates Based on Stomatal and Mesophyll Behaviour in *Platycladus orientalis*

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Abstract: Prediction of whole-plant short-term water use efficiency ($WUE_{s,P}$) is essential to indicate plant performance and facilitate comparison across different temporal and spatial scales. In this study, an isotope model was scaled up from the leaf to the whole-plant level, in order to simulate the variation in WUE_{S P} in response to different CO₂ concentrations (C_a ; 400, 600, and 800 µmol·mol⁻¹) and soil water content (SWC; 35–100% of field capacity). For WUE_{s,P} modelling, leaf gas exchange information, plant respiration, and "unproductive" water loss were taken into account. Specifically, in shaping the expression of the WUE_{s,P}, we emphasized the role of both stomatal (g_{sw}) and mesophyll conductance (g_m) . Simulations were compared with the measured results to check the model's applicability. The verification showed that estimates of g_{sw} from the coupled photosynthesis ($P_{n,L}$) g_{sw} model accounting for the effect of soil water stress slightly outperformed the model neglecting the soil water status effect. The established coupled $P_{n,L}$ - g_m model also proved more effective in estimating g_m than the previously proposed model. Introducing the two diffusion control functions into the whole-plant model, the developed model for WUE_{s,P} effectively captured its response pattern to different Ca and SWC conditions. Overall, this study confirmed that the accurate estimation of $WUE_{s,P}$ requires an improved predictive accuracy of g_{sw} and g_m . These results have important implications for predicting how plants respond to climate change.

Keywords: mesophyll conductance; stomatal conductance; stable isotope; soil water stress; water use efficiency; whole-plant level

1. Introduction

Water use efficiency (WUE), which refers to the ratio of carbon assimilation to water transpired by plants (i.e., water loss), is essential in optimizing plant water use [1]. The WUE can be defined at different temporal and spatial scales. At the leaf level, WUE describes the leaf net photosynthetic rate ($P_{n,L}$) relative to the leaf transpiration rate (E_L). Both processes are controlled by stomatal conductance (g_{sw}). The $P_{n,L}$ is also controlled by mesophyll conductance (g_m), in addition to g_{sw} , as recent studies demonstrated that mesophyll resistance is not negligible [2,3] and may be as important as stomatal conductance [4]. At the whole-plant level, all photosynthetic and non-photosynthetic parts contribute to respiration and water loss. However, the canopy accounts for the most significant part of carbon assimilation and transpiration water loss. Therefore, changes in g_{sw} (and or g_m) may decrease or increase the whole-plant WUE, especially at smaller temporal scales.



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Investigating whole-plant WUE at smaller temporal scales (hours or days) not only facilitates our understanding of whole-plant long-term (months, years, or decades) WUE and the underlying mechanism but also allows us to compare across different temporal and spatial scales. There have, however, been a few attempts to relate g_{sw} (and or g_m) to whole-plant WUE at smaller temporal scales, or models to predict the response pattern of whole-plant short-term WUE (WUE_{s,P}) to environmental changes. The estimation of WUE_{s,P} is frequently conducted on the assumption that leaf short-term WUE (WUE_{s,L}) is representative of WUE_{s,P} [5]. However, there may be a gap between the daily integrals of leaf and whole-plant WUE, as carbon and water loss from non-photosynthetic tissue can result in a decrease in WUE_{s,P} while not affecting WUE_{s,L}. Therefore, it is critical to obtain adequate predictions of whole-plant WUE at smaller temporal scales.

It has been suggested that the leaf WUE model can be scaled to the whole-plant level by taking into account "unproductive" water loss and carbon use by respiration, independent of photosynthesis [6–8]. Built upon this concept, the Farquhar et al. (1989) [7] model relates leaf gas exchange properties and carbon discrimination to whole-plant WUE, but it ignores the effect of mesophyll resistance (the inverse of $g_{\rm m}$) on carbon discrimination (Δ). Thus, the contribution of g_m to Δ needs to be considered [9], and that g_m should have been incorporated in the approach of Farquhar et al. (1989) to predict whole-plant WUE accurately. This hypothesis was supported by our previous findings [10], which found that the whole-plant model emphasizing the role of g_m outperformed the Farquhar et al. (1989) [7] model. Despite years of research, the three most widely used approaches for determining g_m , including the high number of gas exchange properties or measurements of gas exchange combined with chlorophyll fluorescence or carbon isotope discrimination [11], use complex parameters associated with complicated measurements, limiting the easy determination of g_m . In contrast, the soil water content and potential g_m (unstressed g_m , $g_{m,p}$)-dependent empirical model proposed by Keenan et al. (2010) [12], can easily be used. Unfortunately, the model is still flawed in reflecting the influence of other environmental factors and gas exchange properties on g_m . A practical and relatively simple representation of mesophyll behaviour may lie at the heart of a valid and useful prediction of $WUE_{s,P}$. Furthermore, the revised whole-plant model [10] for WUE_{s,P} included the presence of g_{sw} , in addition to g_m , thereby representing the linkage between WUE_{s,P} and g_{sw} . Although several models have been proposed to describe stomatal behaviour, including the simple coupled photosynthesis–stomatal conductance ($P_{n,L}$ - g_{sw}) model and its modified versions, it remains unclear which approach is the most useful. In general, the WUE model scaling from the leaf to the whole-plant level needs to be revised and improved based on wellmodelled stomatal and mesophyll behaviors.

The latest observations showed that globally-averaged atmosphere CO₂ concentration (C_a) reached a new high (413.2 ± 0.2 µmol·mol⁻¹) in 2020 [13]. If the upward trend of C_a continues, soil water stress may be intensified by climate change in many areas. Making it crucial to predict how WUE_{s,P} responds to the different C_a and soil water content (SWC). Therefore, we developed a model to estimate g_m based on the empirical relationship between g_m and $P_{n,L}$ (i.e., the coupled photosynthesis-mesophyll conductance model), and the revised g_m model and the previously established g_{sw} model were then incorporated into the whole-plant WUE model to estimate the variation in WUE_{s,P}. Measurements of whole-plant net CO₂ gas exchange (root systems have been excluded from measurements, i.e., aboveground measurements) and transpiration under different $C_a \times SWC$ conditions were conducted concurrently, allowing us to calculate the actual WUE_{s,P} and to compare the measured results with simulations obtained from the developed whole-plant WUE model. Our aim was, first, to establish a reliable model for g_m ; second, to check the applicability of the whole-plant WUE model scaled from the leaf level, based on estimations of stomatal and mesophyll behavior.

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2. Theoretical Background

2.1. Coupled g_{sw} - $P_{n,L}$ Model

Previous studies found that leaf stomatal conductance (g_{sw} , mol H₂O·m⁻²·s⁻¹) is highly correlated with photosynthesis ($P_{n,L}$, µmol·m⁻²·s⁻¹). Based on this, a series of models on the basis of the linear relationship between g_{sw} and $P_{n,L}$ have been proposed [14–16]. By incorporating the effect of leaf-to-air vapor pressure deficit (D), Leuning et al. (1995) [17] established an alternative coupled $P_{n,L}$ - g_{sw} model based on former studies

$$g_{\rm sw} = g_{0,\rm sw} + g_1 P_{\rm n,L} \frac{f(D)}{C_{\rm s} - \Gamma} \tag{1}$$

where $g_{0,sw}$ and g_1 are fitted parameters and $g_{0,sw}$ is considered to represent the residual stomatal conductance (mol H₂O·m⁻²·s⁻¹); C_s is the leaf surface CO₂ concentration (µmol·mol⁻¹); Γ is the CO₂ compensation point (µmol·mol⁻¹); f(D) is the vapor pressure deficit-dependent function. To describe the effect of D on stomatal behaviour, numerous expressions have been introduced [14,17–21]. Lloyd (1991) [19] and Yu et al. (2001) [21] consistently found that the precision of estimation was highest when imposing the function $f(D) = h_s$, with h_s referring to the relative humidity at leaf surface in %. Thus, we adopted the expression $f(D) = h_s$ in the Leuning et al. (1995) model [17].

The model introduced by Leuning et al. (1995) [17] has been widely used to predict gas exchange properties at the leaf scale [16,22], albeit without taking into account the response of water stress. To overcome this limitation, Egea et al. (2011) [23] proposed an improved model, which incorporated a soil water stress-dependent function ($f(\theta_s)$, calculated by Equation (5)), to describe the behaviour of gas exchange properties

$$g_{\rm sw} = g_{0,\rm sw} + g_1 P_{\rm n,L} \frac{f(\theta_{\rm s})f(D)}{C_{\rm s} - \Gamma}$$
⁽²⁾

2.2. Coupled g_m - P_n Model

Models which can easily represent mesophyll behaviour in response to environmental drivers are still scarce. Considering the restrictions of soil water stress on g_m (mol $CO_2 \cdot m^{-2} \cdot s^{-1}$), Keenan et al. (2010) [12] proposed a function to predict the linkage between g_m and soil water status

$$g_{\rm m} = f(\theta_{\rm m})g_{\rm m,p} \tag{3}$$

where $f(\theta_m)$ is the mesophyll conductance limitation function, which depends on soil water stress (calculated by Equation (5)); $g_{m,p}$ is the potential (unstressed) g_m . This model has been used to represent the feedback of g_m to soil water stress [1], but does not consider the response of mesophyll behaviour to other environmental drivers, such as C_a. In fact, g_m is affected by increases or decreases in C_a , and even changes more subtly with changes in C_a than in g_{sc} ($g_{sc} = g_{sw}/1.6$) [24]. Previous studies have observed that the $P_{n,L}$ increased linearly with g_m [25–27], which prompted us to establish a coupled $P_{n,L}$ - g_m function to model g_m by imposing similar limitation functions to mesophyll behavior as those imposed to stomatal behaviour. Based on the empirical relationship between $P_{n,L}$ and g_m , the proposed model is as follows

$$g_{\rm m} = g_{0,\rm m} + g_2 P_{\rm n,L} \frac{f(\theta_{\rm m})f(D)}{C_{\rm s} - \Gamma}$$
 (4)

where $g_{0,m}$ and g_2 are fitted parameters, and $g_{0,m}$ is considered to represent the residual mesophyll conductance (mol CO₂·m⁻²·s⁻¹).

The two soil water stress-dependent limitation functions, $f(\theta_s)$ and $f(\theta_s)$, were expressed as [12,28]

$$f(\theta_{i}) = \begin{cases} 1 & \theta \geq \theta_{c} \\ \left[\frac{\theta - \theta_{w}}{\theta_{c} - \theta_{w}}\right]^{q_{i}} & \theta_{w} \leq \theta \leq \theta_{c} \\ 0 & \theta \leq \theta_{w} \end{cases}$$
(5)

where θ is the soil volumetric water content (%); θ_c and θ_w are soil water content levels at field capacity (26.20%) and permanent wilting point (4.08%), respectively; parameter q_j is a measure of the nonlinearity of the effects of soil water stress on the limiting mechanisms; the subscript i = s and m represent stomatal and mesophyll limitations, respectively. In this study, the selected values for tunable parameters of q_s and q_m were 0.25, 0.50, 0.75, 1.00, 1.25, and 1.50, within the previously reported range [12,23].

2.3. Leaf and Whole-Plant WUE Model

The leaf instantaneous water use efficiency (WUE_{i,L}, mmol·mol⁻¹) is the ratio of leaf net photosynthetic rate ($P_{n,L}$, μ mol·m⁻²·s⁻¹) to transpiration rate (E_L , mmol·m⁻²·s⁻¹) [6]

$$WUE_{i,L} = \frac{P_{n,L}}{E_L} = \frac{P_{n,L}}{g_{sw}D}$$
(6)

Substituting the Egea et al. (2011) [23] model (Equation (2)) and the Leuning et al. (1995) [17] model (Equation (1)) into Equation (6), we obtain the following formulas, respectively

$$WUE_{i,L} = \frac{P_{n,L}}{D} \times \frac{C_s - \Gamma}{(C_s - \Gamma)g_{0,sw} + g_1 P_{n,L} f(\theta_s) f(D)}$$
(7)

$$WUE_{i,L} = \frac{P_{n,L}}{D} \times \frac{C_s - \Gamma}{(C_s - \Gamma)g_{0,sw} + g_1 P_{n,L} f(D)}$$
(8)

The WUE_{i,L} inferred from Equation (7) with well parameterized q_s ($q_s = 0.25$, see Section 4.1) is model configuration 1, and that inferred from Equation (8) is model configuration 2.

The whole-plant instantaneous water use efficiency (WUE_{i,P}, mmol·mol⁻¹) is the ratio of whole-plant net photosynthetic rate ($P_{n,p}$, μ mol·h⁻¹) to transpiration rate (E_p , mmol·h⁻¹) [7]. Considering respiration and water loss from the non-photosynthetic organs, the ratio of instantaneous net photosynthesis to transpiration can be scaled from the leaf to the whole-plant level

$$WUE_{i,P} = \frac{P_{n,P}}{E_p} = \frac{P_{n,L}}{E_L} \times \frac{(1 - \phi_{c,i})}{(1 + \phi_{w,i})} = \frac{P_{n,L}}{g_{sw}D} \times \frac{(1 - \phi_{c,i})}{(1 + \phi_{w,i})}$$
(9)

where $\phi_{c,i} = (3.6 P_{n,L} \times LA - P_{n,P})/(3.6 P_{n,L} \times LA)$, with *LA* referring to plant total leaf area in m²) is the proportion of respiration from non-photosynthetic parts (twigs and stem) during the daytime, and $\phi_{w,i} = (E_P - 3.6 E_L \times LA)/(3.6 E_L \times LA)$ is the proportion of water loss from non-photosynthetic parts during the daytime. Similarly, we substituted the simulated g_{sw} , calculated via the Egea et al. (2011) [23] model (Equation (2)) and the Leuning et al. (1995) [17] model (Equation (1)) into Equation (9), obtaining the following formulas, respectively

$$WUE_{i,P} = \frac{P_{n,L}}{D} \times \frac{C_s - \Gamma}{(C_s - \Gamma)g_{0,sw} + g_1 P_{n,L} f(\theta_s) f(D)} \times \frac{(1 - \phi_{c,i})}{(1 + \phi_{w,i})}$$
(10)

$$WUE_{i,P} = \frac{P_{n,L}}{D} \times \frac{C_{s} - \Gamma}{(C_{s} - \Gamma)g_{0,sw} + g_{1}P_{n,L}f(D)} \times \frac{(1 - \phi_{c,i})}{(1 + \phi_{w,i})}$$
(11)

The whole-plant short-term water use efficiency ($WUE_{s,P}$) is the ratio of whole-plant cumulative CO₂ assimilation to water loss. At the diel time scale, not only the role of

respiration and water loss from non-photosynthetic parts (twigs and stem) during the daytime need to be included, but also respiration and water loss from whole parts (leaf, twigs, and stem) during the nighttime contribute substantially to $WUE_{s,P}$. When all these processes are taken into account, the time-integrated $WUE_{s,P}$ is as follows

WUE_{s,P} =
$$\frac{\int P_{n,P}}{\int E_p} = \frac{\int P_{n,L}}{\int E_L} \times \frac{(1 - \phi_{c,s})}{(1 + \phi_{w,s})} = \frac{\int P_{n,L}}{\overline{g_{sw}}\overline{D}} \times \frac{(1 - \phi_{c,s})}{(1 + \phi_{w,s})}$$
 (12)

where $\phi_{c,s} = (3.6 P_{n,L} \times LA - P_{n,P} + R_P)/(3.6 P_{n,L} \times LA)$, with R_P referring to nighttime respiration in mmol·h⁻¹) is the proportion of respiration from non-photosynthetic parts (twigs and stem) during the whole time and from leaves during the nighttime; $\phi_{w,s} = (E_P - 3.6 E_L \times LA + E_d)/(3.6 E_L \times LA)$, with E_d referring to nighttime transpiration in mol·h⁻¹) is the proportion of water loss from non-photosynthetic parts (twigs and stem) during the whole time and from leaves during the nighttime. The above time integral is denoted as \int . According to Fick's law

$$\frac{P_{n,L}}{g_{sw}} = \frac{C_a}{1.6} \times (1 - \frac{C_i}{C_a})$$
(13)

where C_i is the leaf intercellular CO₂ concentration (µmol·mol⁻¹). The photosynthetic ¹³C discrimination (Δ , ‰) reflects the physiological properties over short time scales [29–31]. From the variant of the Farquhar et al. (1989) [7] classical model, including the effect of g_m on Δ , the short-term C_i/C_a ratio can be written as follows

$$\frac{C_{\rm i}}{C_{\rm a}} = \frac{\Delta_{\rm mea} - a + (b - a_{\rm m})\frac{8\rm sw}{1.6\rm g_{\rm m}}}{b - a + (b - a_{\rm m})\frac{8\rm sw}{1.6\rm g_{\rm m}}}$$
(14)

where *a* is the fractionation associated with the atmospheric CO₂ diffusion at the boundary layer (4.4‰); *a*_m is the fractionation of CO₂ diffusion and dissolution in the liquid phase (1.8‰); *b* is the fractionation during carboxylation (29‰); Δ_{mea} is measured photosynthetic ¹³C discrimination = $(\delta^{13}C_a - \delta^{13}C_l)/(1 + \delta^{13}C_l)$, with $\delta^{13}C_a$ and $\delta^{13}C_l$ referring to $\delta^{13}C$ of atmospheric CO₂ and water-soluble organic materials (WSOM, fast-turn-over carbohydrates) in leaves, respectively.

Substituting Equation (13) and Equation (14) into Equation (12), we obtain the following equation

$$WUE_{s,P} = \frac{C_{a}}{1.6D} \times \frac{b - \Delta}{b - a + (b - a_{m})\frac{g_{sw}}{g_{m}}} \times \frac{(1 - \phi_{c,s})}{(1 + \phi_{w,s})}$$
(15)

Similar to the simulation of $WUE_{i,L}$, two model configurations were applied in Equation (15), and we obtained the following equations

$$WUE_{s,P} = \frac{C_{a}}{1.6D} \times \frac{(1 - \phi_{c,s})}{(1 + \phi_{w,s})} \times \frac{b - \Delta}{b - a + (b - a_{m}) \times \frac{(C_{s} - \Gamma)g_{0,sw} + g_{1}P_{n,L}f(\theta_{s})f(D)}{(C_{s} - \Gamma)g_{0,m} + g_{2}P_{n,L}f(\theta_{m})f(D)}}$$
(16)

$$WUE_{s,P} = \frac{C_{a}}{1.6D} \times \frac{(1 - \phi_{c,s})}{(1 + \phi_{w,s})} \times \frac{b - \Delta}{b - a + (b - a_{m}) \times \frac{(C_{s} - \Gamma)g_{0,sw} + g_{1}P_{n,L}f(\theta_{s})f(D)}{(C_{s} - \Gamma)f(\theta_{m})g_{m,p}}}$$
(17)

3. Material and Methods

3.1. Experimental Design and Management

The experiment was carried out in April 2018 at the Chinese Forest Ecosystems Research Network (116°05′ E, 40°03′ N), situated at the Western Hill, Beijing, North China, using 7-year-old *Platycladus orientalis* saplings of the same genotype of a temperate origin. The plants were each transplanted into 15.51-L pots containing soil collected from a local *Platycladus orientalis* stand. The soil type is sandy loam, and the field capacity (θ_c , 26.2%)

and permanent wilting point (θ_w , 4.08%) of the soil and plants were determined by a pilot experiment. The θ_c was measured by soil water content (SWC) sensors (HOBO–U30, Onset, Cape Cod, Massachusetts, USA) after soil samples absorbed water for 24 h with no vertical underwater droplets. The $\theta_{\rm W}$ was determined by the same sensors when leaves produced wilting and could not be restored by supplemental water, that is, below the wilting point leaf water potential (measured by portable plant water potential meter (WP4C, Decagon, Pullman, WA, USA); data not shown) did not increase with the increase in SWC. Platycladus orientalis samplings with similar growth status and canopy structure (approximately 1.4 m high) were grown in a greenhouse. After acclimation in the greenhouse for two months, saplings were moved to growth chambers (FH-230, Taiwan Hipoint Corporation, Kaohsiung City, Taiwan) and subjected to a nested design with three CO_2 concentration (C_a) levels and five SWC regimes. The controlled environment (light, air temperature, and relative humidity) in the growth chambers was set to simulate natural growth conditions. From 07:00 to 19:00 (simulating daytime), all white LED lights were turned on, with 60% relative humidity and 25 °C. From 19:00 to 07:00 (simulating nighttime), all white LED lights were turned off, with 80% relative humidity and 18 °C. In North China, P. orientalis saplings are generally grown under the forest canopy, which receives a lower photosynthetic photon flux density (with an average of $230 \pm 37 \,\mu mol \cdot m^{-2} \cdot s^{-1}$) than full sunlight (with an average of $350 \pm 41 \,\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$) at daytime during the growing season. Thus, the low level of light intensity in the growth chamber ($220 \pm 20 \ \mu mol \cdot m^{-2} \cdot s^{-1}$) was considered to be approximately appropriate to simulate the growth of understory saplings.

To realize orthogonal treatments, two growth chambers were used. One growth chamber (Figure 1a) was connected to a CO_2 tank and ambient atmosphere with an intake pipe, which was used to maintain elevated C_a of 600 µmol·mol⁻¹ (C_{600}) or 800 µmol·mol⁻¹ (C_{800}) . Another growth chamber (Figure 1b) was only connected to ambient atmosphere with an intake pipe to maintain C_a of approximately 400 μ mol·mol⁻¹ (C_{400}). CO₂ sensors and control systems inside the growth chambers can continuously monitor and adjust C_a steady near the enactment value, with a standard deviation of 50 μ mol·mol⁻¹. Each C_a treatment was subjected to five SWC regimes: (1) 35-45% of field capacity, FC, (simulating severe drought), (2) 50–60% of FC (moderate drought), (3) 60–70% of FC (mild drought), (4) 70–80% of FC (well-watered), and (5) 95–100% of FC (excessively watered). The FC of the potting soil was 26.20%. For the sake of calculative simplicity, we assumed that the SWC gradient was: (1) 10.48%, (2) 14.41%, (3) 17.03%, (4) 19.65%, and (5) 26.20%, respectively. The SWC in the upper 10 to 15 cm was continuously measured by sensors (HOBO–U30, Onset, Cape Cod, Massachusetts, USA), and the water status of each potting soil was checked twice daily and irrigated manually to achieve the target SWC regimes. The surface of the potting soil was covered with an approximately 2-cm layer of perlite to reduce soil evaporation. Each treatment ($C_a \times SWC$) lasted for 30 days and had three pot-grown saplings as replicates. As one growth chamber was able to hold five pots, the experiment was performed progressively from June to November 2018, where treatments were maintained at $C_{400} \times \text{SWC}$ (in chamber b) and $C_{600} \times \text{SWC}$ (in chamber a) from June to August, and at $C_{400} \times SWC$ (in chamber b) from September to November. The pots were rearranged frequently to exclude position effects.



Figure 1. Schematic of growth chambers used in the experiments. One growth chamber (**a**) was used to maintain elevated CO_2 concentration of 600 µmol·mol⁻¹ or 800 µmol·mol⁻¹. Another growth chamber (**b**) was used to maintain CO_2 concentration of 400 µmol·mol⁻¹. There were five pots inside each chamber.

3.2. Measurements

3.2.1. Whole-Plant Carbon Balance and Measurement

After the saplings had been subjected to the 30-day $C_a \times SWC$ treatment, whole-plant carbon balance was measured inside the growth chambers using the static chamber as designed by Jasoni et al. (2005) [32]. The static chamber measured $50 \times 50 \times 150$ cm, and in its interior, a pocket weather meter was incorporated (Kestrel 5500, Nielsen-Kellerman, Boothwyn, PA, USA) to monitor air temperature (T_a, K) and pressure (P, P_a) . To avoid soil respiration, the substrate surface was tightly sealed with airtight plastic film as described by Escalona et al. (2013) [33]. Prior to each measurement, the sapling was enclosed in the static chamber, and the fan on its top turned on for 30 s to ensure that the flux was mixed well. The C_a in the static chamber was measured by an infrared gas analyzer (Li-8100, Li-Cor, Lincoln, NE, USA), starting after the flux was well mixed (initial C_a , i.e., C_0) and finishing after the measurement had lasted for 3 min (final C_a , i.e., C_1). Measurements for each sapling were repeated three times and conducted at 9:00–10:00, 13:00–14:00, and 17:00–18:00 during daytime and at 22:00–23:00, 2:00–3:00, and 6:00–7:00 during nighttime. During the 3 min, the C_a in the closed static chamber gradually decreased in the day but increased in darkness. The whole-plant daytime net photosynthetic rate $(P_{n,p})$ and the nighttime respiratory rate (R_p) were calculated as follows

$$P_{n,P} = \frac{V}{\Delta t} \times \frac{273.15}{T_a} \times \frac{P}{101,325} \times \frac{1}{22.41} \times (C_0 - C_1) \times \frac{60}{1000}$$
(18)

$$R_{n,P} = \frac{V}{\Delta t} \times \frac{273.15}{T_a} \times \frac{P}{101,325} \times \frac{1}{22.41} \times (C_1 - C_0) \times \frac{60}{1000}$$
(19)

where *V* is the chamber volume (L) and $\Delta t = 3$ min is the time duration. The *P*_{n,p} and R_p were calculated from values measured during daytime and nighttime, respectively.

3.2.2. Whole-Plant Transpiration Measurements

The whole-plant daytime transpiration rate (E_p) was measured from the beginning until the end of the experiment by a Flow 32-1K system (Dynamax, Houston, TX, USA). The Flow 32-1K system includes gauges installed at approximately 25 cm above the stem base and a CR1000 logger (Campbell Scientific, Logan, UT, USA), which continuously collected E_p data every 15 min. In this study, each treatment ($C_a \times$ SWC) lasted for 30 days. The E_p values remained relatively stable from the 21st day of orthogonal treatments, which were used for data analysis. The whole-plant nighttime transpiration rate (E_d) was measured by mass loss during the night. Total plant nighttime transpiration was obtained from the difference in pots weight at the onset (19:00) and end of night (7:00). During plant nighttime transpiration measurements, the substrate surface was tightly sealed with airtight plastic film as described by Escalona et al. (2013) [33] to avoid soil evaporation. Measurements were made every 3 days.

The measured WUE_{i,P} was the ratio between $P_{n,p}$ to E_p ($P_{n,p}/E_p$), while the modelled WUE_{i,P} was calculated by different model configurations. In model configuration 1 (Equation (10)), g_{sw} was calculated by Equation (2) with well parameterized q_s ($q_s = 0.25$, see Results 3.1). The model configuration 2 is Equation (11) with no additional parameterization associated with soil water stress.

The measured WUE_{s,P} was the ratio between accumulative carbon gain and cumulative water loss, that is, WUE_{s,P} = $(P_{n,P} - R_P)/(E_P + E_d)$. In contrast, the modelled WUE_{s,P} were calculated by different model configurations. In model configuration 1 (Equation (16)), g_m was calculated by Equation (4) with well parameterized q_m ($q_m = 0.25$, see Section 4.2), and g_{sw} was calculated by Equation (2) with well parameterized q_s ($q_s = 0.25$, see Section 4.1). In model configuration 2 (Equation (17)), g_m was calculated by Equation (3) with well parameterized q_m ($q_m = 0.50$, see Section 4.2), and g_{sw} was calculated by Equation (1).

3.2.3. Leaf Gas Exchange and Stable Isotope Analysis

On the day of whole-plant carbon balance measurements, leaf gas change properties ($P_{n,L}$, E_L , g_{sw} , and C_i), leaf temperature (T_L), and leaf surface relative humidity (RH) were measured inside the growth chambers on mature leaves, using a portable gas exchange system (Li-6400, Li-Cor, Lincoln, NE, USA) fitted with a needle leaf chamber. The measurements were conducted at different positions (upper, middle, and lower crown) and made on at least three different leaves in each canopy layer at 9:00, 13:00, and 17:00. No significant differences (p > 0.05) in these measurements among different canopy layers were observed. Almost all leaves were exposed to similar light intensities and, thus, the effect of internal leaves was not considered. In this study, we assumed that a period of 30 days was long enough for saplings to be subjected to the treatments, according to our pilot experiment as described by Zhang et al. (2019) [10]. Measured leaf instantaneous water use efficiency (WUE_{i,L}) was calculated as the ratio between $P_{n,L}$ and E_L ($P_{n,L/}E_L$).

The leaves used for gas exchange measurements were detached, immediately wrapped in tinfoil, and preserved in liquid nitrogen. Leaf water-soluble organic matter (WSOM) was extracted using the same method as described by Zhang et al. (2019) [10]. The obtained WSOM was dried and then combusted in an elemental analyzer (Flash EA 1112, Thermo Finnigan, California, USA) coupled to a continuous-flow stable isotope ratio mass spectrometer (DELTAplusXP, Thermo Finnigan, California, USA). The δ^{13} C of leaf WSOM ($\delta^{13}C_1$) was analyzed using the stable isotope ratio mass spectrometer with a precision of $\pm 0.1\%$. In addition, at the end of each treatment, atmosphere samples from the growth chamber were also collected (at least three replicates), and the δ^{13} C of the atmosphere ($\delta^{13}C_a$) was analyzed by the stable isotope ratio mass spectrometer. Measured g_m was obtained by carbon isotope discrimination combined with gas exchange measurements as previously described by Zhang et al. (2019) [10], i.e.,

$$g_{\rm m} = \frac{(b - a_{\rm i}) \times \frac{P_{\rm n,L}}{C_{\rm a}}}{(\Delta_{\rm lin} - \Delta_{\rm mea})}$$
(20)

where a_i is the fractionation of CO₂ diffusion and dissolution in the liquid phase (1.8‰), and Δ_{lin} is photosynthetic ¹³C discrimination (‰) calculated by the version of the Farquhar et al. (1982) [34] simple linear model, namely,

$$\Delta_{\text{lin}} = a + (b' - a) C_{\text{i}}: C_{\text{a}}$$
(21)

where b' is the fractionation relevant to the reactions of Rubisco and PEP carboxylase (27%) [6].

3.2.4. Whole-Plant Total Leaf Area Measurement

At the end of the experiment, saplings were harvested and separated into different parts. A portion of leaves with different widths and shapes were selected as subsamples. Leaf subsample fresh weight (FW_{sub}) was immediately determined using electronic balance with an accuracy of \pm 0.001 g, and the leaf area for subsample (LA_{sub}) was determined using image processing software for Photoshop. Subsequently, these leaves were dried at 80 °C for 48 h in an oven to obtain their dry weight (DW_{sub}). The dry weights of the remaining harvested leaves (DW_{rest}) were also determined. The whole-plant total leaf area (LA) of each sapling was calculated as follows

$$LA = R_{\rm D} \times DW = (LA_{\rm sub}/DW_{\rm sub}) \times (DW_{\rm sub} + DW_{\rm rest} + DW_{\rm iso})$$
(22)

In this equation, R_D is leaf area per dry weight (m²·g⁻¹), *DW* is whole-plant total dry weight (g), and $DW_{iso} = FW_{iso} \times DW_{sub}/FW_{sub}$, with FW_{iso} referring to fresh weight of leaves used for isotope analysis in g) is dry weight of leaves used for isotope analysis (g).

3.3. Data Analysis

All statistical analyses were conducted using SPSS 19.0. The influences of C_a and SWC on mean variables of g_{sw} , g_m , and WUE (including WUE_{i,L}, WUE_{i,P}, and WUE_{s,P}) were determined by two-way analysis of variance (ANOVA), and results were considered statistically significant at p < 0.05. Deviations of the modeled g_{sw} , g_m , and WUE from their measurements were absolute differences between the modeled and measured values. Relationships between the measured and modeled values in g_{sw} , g_m , and WUE were assessed using general linear regression analysis.

4. Results

4.1. Measured and Modelled Responses of g_{sw} to SWC and C_a

Changes in SWC and C_a significantly affected g_{sw} (p < 0.05), with a maximum of 0.0963 mmol H₂O·m⁻²·s⁻¹ at C₄₀₀ × 19.65% of SWC and a minimum of 0.0155 mol H₂O·m⁻²·s⁻¹ at C₈₀₀ × 10.48% of SWC (Figure 2). In all cases, g_{sw} decreased with elevated C_a. The g_{sw} increased sharply as water stress was alleviated irrespective of C_a, and this effect was less evident when SWC exceeded 17.03% and even decreased when g_{sw} peaked at 19.65% of SWC (Figure 2).



Figure 2. Response of measured leaf stomatal conductance (g_{sw} , mol $H_2O \cdot m^{-2} \cdot s^{-1}$) to three CO_2 concentrations (C_a) × five soil water contents (SWC). C_{400} , C_{600} , and C_{800} are C_a of 400, 600, and 800 µmol·mol⁻¹. Data represent mean values ± SD.

The g_{sw} simulated by the two coupled $P_{n,L}$ - g_{sw} model (Equations (1) and (2)) decreased in response to elevated C_a (Figure 3). In the absence of additional parameterization associated with soil water stress (Equation (1)), the g_{sw} increased as the soil water status improved and reached maximum values at 19.65% of SWC, with a slight decrease thereafter. In contrast, when the effect of soil water stress was incorporated in the coupled $P_{n,L}$ - g_{sw} model (Equation (2)), the simulated g_{sw} generally increased as SWC increased, regardless of the value imposed by q_s (Figure 3).



• Equation(2), $q_s=1.25$ • Equation (2), $q_s=1.50$ • Equation (1)

Figure 3. The estimated leaf stomatal conductance (g_{sw} , mol H₂O·m⁻²·s⁻¹) in *Platycladus orientalis* saplings under different soil water contents (SWC) and CO₂ concentrations (C_a) conditions, based on different models (Equations (1) and (2)). C₄₀₀, C₆₀₀, and C₈₀₀ are C_a of 400 (**a**), 600 (**b**), and 800 µmol·mol⁻¹ (**c**). Tunable parameter q_s is a measure of the nonlinearity of the effects of soil water stress on the stomatal limiting mechanisms. Data represent mean values ± SD.

The correlation between the measured and calculated g_{sw} is shown in Table 1. When applying Equation (2), we found a strong correlation between the calculated and the measured g_{sw} (p < 0.01), and the correlation coefficient R^2 decreased from 0.88 to 0.68 as q_s increased from 0.25 to 1.50. The calculated g_{sw} based on Equation (1) also significantly correlated with the measured g_{sw} (p < 0.01; $R^2 = 0.87$). However, when applying Equation (2), at $q_s = 0.25$, the calculated g_{sw} (higher R^2 and slope closer to (1) was closer to measured g_{sw} than when using Equation (1). Additionally, with Equation (2), there was less deviation (0.0084 ± 0.0053 mol H₂O·m⁻²·s⁻¹) between the measured and calculated g_{sw} than with Equation (1) (0.0086 ± 0.0062 mol H₂O·m⁻²·s⁻¹). This showed that the $P_{n,L}$ – g_{sw} model, which incorporates the soil water stress ($q_s = 0.25$, Equation (2)), better predicts g_{sw} than Equation (1).

Table 1. Correlation analysis between measured and modeled leaf stomatal conductance (g_{sw} , mol H₂O·m⁻²·s⁻¹) using different models (Equations (1) and (2)).

Model	Regression of Measured and Modelled Leaf g_{sw}			
	Linear Regression Equation	R^2	p	
Equation (2), $q_{\rm s} = 0.25$	y = 0.88x + 0.01	0.88	< 0.01	
Equation (2), $q_{\rm s} = 0.50$	y = 0.86x + 0.01	0.86	< 0.01	
Equation (2), $q_{\rm s} = 0.75$	y = 0.83x + 0.01	0.83	< 0.01	
Equation (2), $q_{\rm s} = 1.00$	y = 0.79x + 0.01	0.79	< 0.01	
Equation (2), $q_{\rm s} = 1.25$	y = 0.74x + 0.02	0.74	< 0.01	
Equation (2), $q_{\rm s} = 1.50$	y = 0.68x + 0.02	0.68	< 0.01	
Equation (1)	y = 0.87x + 0.01	0.87	< 0.01	

4.2. Measured and Modelled Responses of g_m to SWC and C_a

The g_m ranged between 0.0131 and 0.0571 mol CO₂·m⁻²·s⁻¹, significantly lower than g_{sw} (p < 0.05). Elevation of C_a produced significant changes in g_m . In all case, elevated C_a decreased g_m (Figure 4). Additionally, SWC significantly influenced the g_m (p < 0.05) in a similar pattern as g_{sw} . Under low soil moisture content, g_m increased rapidly with SWC. However, the rate of increase in g_m decreased when SWC exceeded 17.03% and even decreased at SWC between 19.65% and 25.55% (Figure 4).



Figure 4. Response of measured leaf mesophyll conductance (g_m , mol CO₂·m⁻²·s⁻¹) to three CO₂ concentrations (C_a) × five soil water contents (SWC). C_{400} , C_{600} , and C_{800} are C_a of 400, 600, and 800 µmol·mol⁻¹. Data represent mean values ± SD.

The simulated g_m , calculated by the SWC- and $g_{m,0}$ -dependent function (Equation (3)) and the coupled $P_{n,L}$ - g_m model (Equation (4)), is presented in Figure 5. Regardless of the model used, the calculated g_m decreased with C_a (Figure 5). Applying Equation (3), the simulated g_m increased almost linearly with an increase in SWC levels and tended to be higher with lower q_m , except under excess SWC (25.55% of SWC) (Figure 5a,c,e). In contrast, the simulated g_m calculated by Equation (4), using various q_m values, produced a more complicated tendency to SWC. (Figure 5b,d,f).

The relationships between measured and calculated g_m based on Equations (3) and (4) are shown in Table 2. Both model approaches produced significant relationships between simulated and measured results (p < 0.05). Setting the same q_m value, Equation (4) led to a higher R^2 (0.44 ~ 0.79) between the estimated and measured results than that of Equation (3) (0.34 ~ 0.52), and the former caused less deviation (0.0055 ± 0.0038 ~ 0.0097 ± 0.0046) from measurements than the latter (0.0090 ± 0.0058 ~ 0.0159 ± 0.0078). Therefore, the proposed coupled $P_{n,L}$ - g_m model with well parameterized q_m ($q_m = 0.25$, Equation (4)) effectively improved the predictive accuracy of g_m compared to the previously introduced $g_{m,p}$ - and SWC-dependent model (Equation (3)).

4.3. Measured and Modeled Instantaneous WUE at Leaf and Whole-Plant Level

At the leaf level, elevated C_a significantly enhanced the measured WUE_{i,L} (p < 0.05). Variations in SWC also significantly influenced the measured WUE_{i,L} (p < 0.05), which increased as the severe drought was alleviated (SWC increased from 10.48% to 14.41%), followed by a decline with increasing SWC levels and was almost constant when the SWC was above 19.65% (Figure 6a). In both model configurations, the response pattern of simulated WUE_{i,L} to SWC × C_a was similar to that of measured values, except that the simulated WUE_{i,L} increased as the SWC improved from 14.41 to 17.03% at C₄₀₀ and C₆₀₀, departing from the observed decreasing trend (Figure 6a,b).



Figure 5. The estimated leaf mesophyll conductance $(g_m, \text{ mol } \text{CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1})$ is based on different models (Equation (3), $(\mathbf{a}, \mathbf{c}, \mathbf{e})$; and Equation (4), $(\mathbf{b}, \mathbf{d}, \mathbf{f})$ under varying soil water contents (SWC) and CO₂ concentrations (C_a). C₄₀₀, C₆₀₀, and C₈₀₀ are C_a of 400 (**a**,**b**), 600 (**c**,**d**), and 800 µmol·mol⁻¹ (**e**,**f**). Tunable parameter q_m is a measure of the nonlinearity of the effects of soil water stress on the mesophyll limiting mechanisms. Data represent mean values \pm SD.

At the whole-plant level, it was observed that C_a and SWC significantly influenced (p < 0.05) the measured instantaneous WUE (WUE_{i,P}). In general, the measured WUE_{i,P} was higher at elevated C_a levels (Figure 6c). When the SWC increased from 10.48% to 14.41%, the percentage increase in the measured WUE_{i,P} was more pronounced at C_{800} than at C_{400} and C_{600} . In response to further increases in SWC, the measured WUE_{i,P} generally decreased sharply with further rises in SWC, but this trend was lesser when the soil water status was more than 19.65% of SWC. In both model configurations, the measured and simulated WUE_{i,P} values were similar in their response patterns to SWC × C_a , except when the SWC increased from 14.41% to 17.03% at C_{400} and C_{600} (Figure 6c,d).

	Regression of Measured and Modeled Leaf gave		
Model			Jean Ssw
	Linear Regression Equation	R^2	p
Equation (3), $q_{\rm m} = 0.25$	y = 0.30x + 0.03	0.50	< 0.01
Equation (3), $q_{\rm m} = 0.50$	y = 0.44x + 0.02	0.52	< 0.01
Equation (3), $q_{\rm m} = 0.75$	y = 0.53x + 0.02	0.48	< 0.05
Equation (3), $q_{\rm m} = 1.00$	y = 0.58x + 0.01	0.43	< 0.05
Equation (3), $q_{\rm m} = 1.25$	y = 0.61x + 0.01	0.38	< 0.05
Equation (3), $q_{\rm m} = 1.50$	y = 0.61x + 0.03	0.34	< 0.05
Equation (3), $q_{\rm m} = 0.25$	y = 0.79x + 0.01	0.79	< 0.01
Equation (3), $q_{\rm m} = 0.50$	y = 0.72x + 0.01	0.72	< 0.01
Equation (3), $q_{\rm m} = 0.75$	y = 0.65x + 0.01	0.65	< 0.01
Equation (3), $q_{\rm m} = 1.00$	y = 0.57x + 0.02	0.57	< 0.01
Equation (3), $q_{\rm m} = 1.25$	y = 0.51x + 0.02	0.51	< 0.01
Equation (3), $q_{\rm m} = 1.50$	y = 0.44x + 0.02	0.44	< 0.01

Table 2. Correlation analysis between measured and modeled leaf mesophyll conductance ($g_{m,}$ mol CO₂·m⁻²·s⁻¹) using different models (Equations (3) and (4)).



Figure 6. Measured (**a**) and simulated (**b**) leaf water use efficiency (WUE_{i-L}, mmol·mol⁻¹), and measured (**c**) and simulated (**d**) whole-plant level instantaneous water use efficiency (WUE_{i-P}, mmol·mol⁻¹) in different model configurations under five soil water contents (SWC) × three CO₂ concentrations (C_a) conditions. C_{400} , C_{600} , and C_{800} are C_a of 400, 600, and 800 µmol·mol⁻¹. Data represent mean values ± SD.

At the leaf and whole-plant level, both models revealed a strong correlation between the measured and calculated instantaneous WUE (p < 0.01). However, the relationship was stronger for model configuration 1 (C1), relative to model configuration 2 (C2) (Figure 7). In C1, the calculated WUE_{i,L} (WUE_{i,P}) deviated from measured WUE_{i,L} by 3.12 ± 2.44 (2.59 ± 1.86) mmol·mol⁻¹, which was slightly less than that realized with C2 (3.14 ± 2.52 (2.62 ± 1.90) mmol·mol⁻¹ (Figure 7). This indicates that C1 was more accurate than C2 in predicting WUE_{i,L} and WUE_{i,P}.



Figure 7. Correlation analysis between measured and modeled results of leaf instantaneous water use efficiency (WUE_{i-L} , $mmol \cdot mol^{-1}$) estimated by different model configurations (**a**), as well as between measured and modeled results of whole-plant instantaneous water use efficiency (WUE_{i-P} , $mmol \cdot mol^{-1}$) estimated by different model configurations (**b**).

4.4. Comparison of Measured and Modeled $WUE_{s,P}$ Values

The measured and simulated WUE_{s,P} values are shown in Figure 8. At severe drought (10.48% of SWC), the measured WUE_{s,P} peaked at C₆₀₀ and was lowest at C₈₀₀, whereas the simulated WUE_{s,P}, in both model configurations, reached its maximum at C₆₀₀ and was lowest at C₄₀₀ (Figure 8). At an improved soil water status (SWC at 14.41% ~ 25.55%), the measured and simulated WUE_{s,P} values significantly increased due to elevated C_a levels (p < 0.01). The measured WUE_{s,P} was also significantly influenced by SWC, generally responding in a similar manner as the measured WUE_{i,P} in response to SWC. When the saplings were subjected to SWC of 14.41% ~ 25.55%, in C1, the response pattern of simulated WUE_{s,P} to SWC was consistent with that of the measured values. In contrast, in C2, the simulated WUE_{s,P} increased as the SWC increased from 19.65 to 26.20% under any C_a, which differed from the response pattern of the measured values (Figure 8).

In both model configurations, there was a strong correlation between the measured and calculated WUE_{s,P} (p < 0.01). However, the correlation (R^2) was stronger for the C1 model, relative to the C2 model (Figure 9). In the C1 model, the calculated WUE_{s,P} deviated from measured WUE_{s,P} by 2.77 ± 2.23 mmol·mol⁻¹, compared with 2.91 ± 2.95 mmol·mol⁻¹ for the C2 model (Figure 9). Therefore, compared with C2, C1 better predicts the actual WUE_{s,P}.



Figure 8. Measured (**a**) and modelled (**b**) whole-plant short-term water use efficiency (WUE_{s,P}, mmol·mol⁻¹) under five soil water contents (SWC) × three CO₂ concentrations (C_a) conditions. C_{400} , C_{600} , and C_{800} are C_a of 400, 600, and 800 µmol·mol⁻¹. Data represent mean values ± SD.



Model Configuration 1 • Model Configuration 2

Figure 9. Correlation analysis between measured and modeled whole-plant short-term water use efficiency ($WUE_{s,P}$, mmol·mol⁻¹) estimated by different model configurations.

5. Discussion

5.1. Model Performance for Estimating g_{sw} and g_m

Soil water stress exclusion in the coupled $P_{n,L}$ - g_{sw} model (Equation (1)) for response patterns of g_{sw} performed reasonably well under non-limiting soil water conditions (Table 1), which is in agreement with previous studies conducted in almond trees [22] and in maize and soybean plants [21]. In response to contrasting soil water treatments, the combination of a soil moisture-dependent function with the coupled $P_{n,L}$ - g_{sw} model (Equation (2)) using

well parameterized q_s (0.25), was slightly more capable of representing the observed pattern of g_{sw} than Equation (1). These results align with a previous study, which highlights the importance of including the water stress function in the coupled $P_{n,L}$ - g_{sw} model [35]. However, adding the soil water stress-dependent function to the coupled $P_{n,L}$ - g_{sw} model contributed little to the improvement of model performance for g_{sw} . This can be ascribed to the fact that the measured $P_{n,L}$ incorporated the effect of the soil water status, resulting in the estimation of g_{sw} from the coupled A_n - g_{sw} model accounting for the effect of soil water.

The Keenan et al. (2010) [12] model (Equation (3)) for g_m was insufficient to take into account the impact of C_a and was therefore less suitable to simulate g_m (Table 2). In contrast, the predictive accuracy improved considerably when estimating g_m using the coupled $P_{n,L}$ - g_m model (Equation (4)). Therefore, the proposed coupled $P_{n,L}$ - g_m model is valid and promising for simulating g_m , despite its phenomenological nature and dependence on physiological hypotheses. Furthermore, imposing $q_m = 0.25$ in Equation (4) provided the best fit with the measured values (Figure 6), indicating that the limitation strength of g_m was similar to that of g_{sw} . This result conflicts with the general findings that stomatal behaviour imposed a higher limitation on photosynthesis than mesophyll behavior [22,36,37]. However, such a phenomenon may not always occur. For example, Pérez-Martín et al. (2009) [4] observed minor difference between stomatal and mesophyll limitations and reported that stiffer and more sclerophyllous leaves would provide greater mesophyll resistance during CO₂ diffusion.

In addition, this study found that, mostly, g_{sw} (and g_m) values varied with SWC, even if the influence of C_a on g_{sw} (and g_m) was significant. Centritto et al. (2002) [38] also found that g_{sw} was significantly lower in water-stressed seedlings than in well-watered seedlings, while elevated C_a did not significantly influence g_{sw} under either well-watered or water-stressed conditions. However, Flexas et al. (2007) [24] observed that both g_{sw} and g_m were much higher in 400 µmol·mol⁻¹ air than those in 1000 µmol·mol⁻¹ air. Thus, C_a effects on g_{sw} (or even g_m) may not be universal across species.

5.2. Different Model Configurations for Estimating $WUE_{s,P}$

In our proposed short-term WUE model (Equation (15)), scaling up from the leaf to the whole-plant level, there are diffusive limitation parameters. The C1 inferred from Equations (2) and (4) could more accurately represent the observed WUE_{s,P} than the C2 inferred from Equations (1) and (3) (Figure 9). This leads us to infer that the model scaling up from the leaf to whole-plant level, based on more accurate stomatal and mesophyll behaviour predictions, could be used to estimate WUE_{s,P} with a high level of precision. In addition, the developed model for estimating WUE_{s,P} also contains photosynthetic parameters by introducing the coupled $P_{n,L}$ - g_{sw} and $P_{n,L}$ - g_m models. Rather than estimating $P_{n,L}$ via the photosynthesis model [21,39], the estimates of WUE_{s,P} were calculated from measured $P_{n,L}$ values to exclude the situation that errors in the representation of g_{sw} and g_m might be compensated or overwhelmed by errors in simulated $P_{n,L}$. In such a situation, we can identify the influence of precision of stomatal and mesophyll modelling on the credibility and accuracy of the developed WUE_{s,P} model.

For WUE_{i,L} and WUE_{i,P} modelling, the C1 inferred from the more accurate g_{sw} model incorporating a soil water stress-dependent function (Equation (2)) slightly outperformed the C2 (Figure 7a,b). However, the R^2 between measured and modelled WUE_{s,P} were lower than those of WUE_{i,L} and WUE_{i-P} (Figures 7 and 9), most likely because the involvement of more parameters in the isotope-inferred WUE_{s,P} model could introduce more uncertainties and errors. For example, complications arising from post-photosynthetic carbon isotope fractionations are not considered as the process is still difficult to assess and largely unknown [40,41]. Furthermore, the effects of photorespiration and mitochondrial respiration on photosynthetic ¹³C discrimination are still the subject of debate [25] and were thus ignored in the current study.

5.3. Uncertainties of WUE_{*s*,*P*} Introduced by g_{sw} and g_m

Uncertainty analysis was conducted to further determine the uncertainties of WUE_{s,P} associated with stomatal and mesophyll behaviour simulations. Using the most effective approach to reproduce g_{sw} (Equation (2), with tunable parameter $q_s = 0.25$) and g_m (Equation (4), with tunable parameter $q_m = 0.25$), the average uncertainties (s.d.) in g_{sw} and g_m were 17.10% (14.14%) and 15.39% (10.98%), respectively. The WUE_{s,P} estimated from C1 caused average uncertainties (s.d.) of 24.09% (21.61%). The relatively small discrepancies between mean value and standard deviation in uncertainties of g_{sw} , g_m , and WUE_{s,P} indicate that these estimation methods were not stable, although model performance was improved. In addition, the WUE_{s,P} was more sensitive to g_{sw} than to g_m . That is, 10% error in g_{sw} introduced 6.17% error in WUE_{s,P}, while 10% error in g_m introduced a smaller error of 4.48% in WUE_{s,P}. Although the stomatal and mesophyll limitations were similar to those of the photosynthetic process in this study, the leaf transpiration is exclusively controlled by g_{sw} when the v is almost constant [9,10,42] (Seibt et al., 2008; Zhao et al., 2017; Zhang et al., 2019), which could result in the g_{sw} being a more influential factor for WUE_{s,P} than the g_m .

Overall, the explored whole-plant model, based on well-characterized coupled $P_{n,L}$ - g_{sw} (Equation (2)) and $P_{n,L}$ - g_m models (Equation (4)), is applicable for evaluating variation in WUE_{s,P} in response to C_a and SWC. However, we recognize that *Platycladus orientalis* is a very specific plant, and the results are hard to generalize for all other plants. It is therefore important to collect data from different plants to further examine the model. In addition, using only data of pot-grown saplings acclimated in growth chambers, with relatively similar canopy components (canopy structure, light interception), is not convincing enough for a general verification of the developed modelling approach. For field-grown plants with a complex canopy structure, water potential and gas exchange information for individual leaves cannot be consistent for the whole-plant level [43,44], leading to difficulties in generalizing the estimation of WUE_{s,P} from leaf properties. Moreover, root systems have been excluded from gas exchange measurements due to it being impossible to separate root and soil respiration for technical restriction. In conclusion, the ability of the whole-plant model to simulate WUE_{s,P} features should still be explored and improved.

6. Conclusions

In this study, the performances of coupled $P_{n,L}$ - g_{sw} and $P_{n,L}$ - g_m models were evaluated using leaf gas exchange measurements. We found the coupled $P_{n,L}$ - g_{sw} model incorporating the water stress-dependent function with well parameterized q_s (Equation (2)) agreed slightly better with the measured g_{sw} values than the model excluding the soil water stress effect (Equation (1)), and the established coupled $P_{n,L}$ - g_m model with well parameterized q_m (Equation (4)) allowed for a more reliable estimation of g_m than the previously introduced $g_{m,p}$ - and SWC-dependent model (Equation (3)). Based on the well-characterized models describing stomatal and mesophyll behavior, an isotopic model, scaling from the leaf to whole-plant level for estimating WUE_{s,P} (Equation (16)), was then established and validated. We found the developed model for WUE_{s,P} proved effective at capturing response patterns to C_a and SWC. Therefore, introducing the model performing well for g_{sw} and g_m into the Farquhar et al. (1989) model was applicable and represents a promising approach for describing whole-plant WUE at smaller temporal scales.

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Abbreviations

The follow	ing abbreviations are used in this manuscript:
SSWC	Soil water content
θ (SWC)	Actual soil water content
$\theta_{\rm c}$	Soil water content at field capacity (26.20%)
θ_{W}	Soil water content at permanent wilting point (4.08%)
Ca	Atmosphere CO ₂ concentration (μ mol·mol ⁻¹). The C ₆₀₀ and C ₈₀₀ are C _a levels of
	600 μ mol·mol ⁻¹ and 800 μ mol mol ⁻¹ , and the C ₄₀₀ is C _a level of 400 μ mol·mol ⁻¹ .
WUE	Water use efficiency (mmol·mol ⁻¹)
WUE _{i-L}	Leaf instantaneous water use efficiency (mmol mol^{-1})
WUE _{i-P}	Whole-plant instantaneous water use efficiency (mmol mol^{-1})
WUE _{s-P}	Whole-plant short-term water use efficiency (mmol·mol ⁻¹)
$P_{n,L}$	Leaf daytime net photosynthetic rate (μ mol·m ⁻² ·s ⁻¹)
$E_{\rm L}$	Leaf daytime transpiration rate (mmol·m ^{-2} ·s ^{-1})
$P_{n,P}$	Whole-plant daytime net photosynthetic rate (mmol h^{-1})
$\int P_{n,P}$	Whole-plant cumulative net carbon sequestration over a day-night cycle $(mmol^{-1})$
Ē _P	Whole-plant daytime transpiration rate (mol· h^{-1})
Ed	Whole-plant nighttime transpiration rate (mol·h $^{-1}$)
$\int E_{\mathbf{P}}$	Whole-plant cumulative transpiration over a day-night cycle (mol^{-1})
[°] R _P	Whole-plant nighttime respiration rate (mmol h^{-1})
$C_{\rm s}$	Leaf surface CO_2 concentration (µmol·mol ⁻¹)
C_{i}	Leaf intercellular CO ₂ concentration (μ mol·mol $^{-1}$)
8b	Leaf boundary layer conductance (mol $CO_2 \cdot m^{-2} \cdot s^{-1}$)
8sw	Leaf stomatal conductance (mol $H_2O \cdot m^{-2} \cdot s^{-1}$)
8sc	Leaf stomatal conductance for CO_2 (mol $CO_2 \cdot m^{-2} \cdot s^{-1}$)
g 1	Fitted parameter associated with the photosynthesis-stomatal conductance model
<i>8</i> 0,sw	Fitted parameter, and $g_{0,sw}$ is considered to represent the residual stomatal conductance (mol H ₂ O·m ⁻² ·s ⁻¹)
$f(\theta_s)$	Stomatal conductance limitation function that depends on soil water stress
g _s	The exponents involved in the stomatal conductance limitation function
gm	Leaf mesophyll conductance (mmol $CO_2 \cdot m^{-2} \cdot s^{-1}$)
gm.p	Potential (unstressed) $g_{\rm m}$ (mmol CO ₂ ·m ⁻² ·s ⁻¹)
82	Fitted parameter associated with the photosynthesis–mesophyll conductance model
Sm 0	Fitted parameter, and $g_{0 \text{ m}}$ is considered to represent the residual mesophyll conductance
0,0	$(\text{mol } \text{CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1})$
$f(\theta_{\rm m})$	Mesophyll conductance limitation function that depends on soil water stress
$q_{\rm m}$	The exponents involved in the mesophyll conductance limitation function
$\Delta_{\rm mea}$	Measured short-term photosynthetic ¹³ C discrimination (‰)
Δ_{lin}	The ¹³ C discrimination calculated by the linear model (‰)
$\delta^{13}C_a$	The δ^{13} C of atmosphere CO ₂ (‰)
$\delta^{13}C_1$	The δ^{13} C of leaf water-soluble organic materials (WSOM) (‰)
а	Fractionation associated with the CO_2 diffusion in air (4.4‰)
b'	Fractionation relevant to the reactions of Rubisco and PEP carboxylase (27‰)
a _m	Fractionation of CO_2 diffusion and dissolution in the liquid phase (1.8%)
a _i	Fractionation of CO_2 diffusion and dissolution in the liquid phase (1.8%)
b	Fractionation during carboxylation (29‰)
е	Discrimination value for the mitochondrial respiration (dark respiration)

f Discrimination value for photorespiration

- Γ CO₂ compensation point with dark respiration
- *k* Carboxylation efficiency
- *D* Water vapor pressure difference between the intercellular spaces of the leaf and the leaf external air (mbar)
- $\phi_{w,i}$ Instantaneous proportion of "unproductive" water loss, that is, water lost by transpiration from twigs and stems during the day
- $\phi_{c,i}$ Instantaneous proportion of carbon fixed during photosynthesis, that is, subsequently lost by respiration from twigs and stems during the day
- $\phi_{w,s}$ Proportion of "unproductive" water loss at short time scale (over a day–night cycle), that is, water lost by transpiration from twigs and stems during the day, and from twigs, stems, and leaves at night
- $\phi_{c,s}$ Proportion of carbon fixed during photosynthesis at short time scale (over a day–night cycle), that is, subsequently lost by respiration from twigs and stems over the whole period, and from leaves during the night
- *LA* Total leaf area (m^2)
- DW Dry weight (g)

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