

Supplementary Information

Assessment of Glucocorticoid Removal by UVA/Chlorination and Ozonation: Performance Comparison in Kinetics, Degradation Pathway, and Toxicity

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Table S1. Available data on measured concentrations of FA, TA, and CP in various environmental samples.

Glucocorticoid	Type of samples	Country	Concentration (ng/L)	Ref.
Triamcinolone acetonide	Hospital wastewater	Netherlands	14–41	[4]
Fluocinolone acetonide	Surface water	USA	0.19 ± 0.15	[19]
Triamcinolone acetonide	Surface water	USA	9.00±0.18	[19]
Clobetasol propionate	Surface water	USA	0.64 ± 0.16	[19]
Fluocinolone acetonide	Surface water	Germany	<0.05–1.0	[20]
Triamcinolone acetonide	Surface water	Germany	0.05–12	[20]
Clobetasol propionate	Surface water	Germany	<0.02–3.4	[20]
Fluocinolone acetonide acetate	Surface water	Italian	0.3–1.2	[21]
Fluocinolone acetonide acetate	Urban wastewater	Italian	2.3	[21]
Triamcinolone acetonide	Surface water	Italian	0.9–2.6	[21]
Triamcinolone acetonide	Urban wastewater	Italian	6.4	[21]
Triamcinolone acetonide	Hospital wastewater	Switzerland	14	[22]
Triamcinolone acetonide	Surface water	Switzerland	<1	[22]
Clobetasol propionate	Hospital wastewater	Switzerland	7	[22]
Clobetasol propionate	Surface water	Switzerland	<1	[22]
Triamcinolone acetonide	Surface water	Hungary	<0.63	[23]

Table S2-1. Pseudo-first-order kinetic models and corresponding R^2 of GCs under different light intensities during UVA/chlorination.

parameter	Rate constants (k; h ⁻¹)	coefficients of determination (R^2)
FA	180 mW/cm ²	0.99763
	220 mW/cm ²	0.99558
	250 mW/cm ²	0.99306
	280 mW/cm ²	0.98601
TA	180 mW/cm ²	0.99467
	220 mW/cm ²	0.99775

CP	250 mW/cm ²	0.25434	0.99441
	280 mW/cm ²	0.33204	0.99085
	180 mW/cm ²	0.16623	0.98153
	220 mW/cm ²	0.22465	0.87598
	250 mW/cm ²	0.20964	0.88711
	280 mW/cm ²	0.23123	0.88929

Table S2-2. Pseudo-first-order kinetic models and corresponding R^2 of GCs under different chlorine dosages during UVA/chlorination.

parameter	Rate constants (k; h ⁻¹)	coefficients of determination (R^2)
FA	0 mg/L	0.99763
	2.5 mg/L	0.99426
	5 mg/L	0.99827
	10 mg/L	0.99824
	20 mg/L	0.99747
	0 mg/L	0.96715
	2.5 mg/L	0.98740
TA	5 mg/L	0.98554
	10 mg/L	0.89792
	20 mg/L	0.98884
	0 mg/L	0.99467
	2.5 mg/L	0.99584
	5 mg/L	0.99516
	10 mg/L	0.99913
CP	20 mg/L	0.99629

Table S3-1. Pseudo-first-order kinetic models and corresponding R^2 of GCs under different O₃ dosages during ozonation.

parameter	Rate constants (k; min ⁻¹)	coefficients of determination (R^2)
FA	19 mg/L	0.99703
	31 mg/L	0.99142
	38 mg/L	0.92409
	59 mg/L	0.94070
	67 mg/L	0.85893
	19 mg/L	0.99206
	31 mg/L	0.97076
TA	31 mg/L	0.97076
	38 mg/L	0.89608
	59 mg/L	0.87522
	67 mg/L	0.80473
	19 mg/L	0.98912
	31 mg/L	0.98839
	38 mg/L	0.92646
CP	59 mg/L	0.89267
	67 mg/L	0.81886

Table S3-2. Pseudo-first-order kinetic models and corresponding R^2 of GCs under different initial pH values during ozonation.

parameter		Rate constants (k; min ⁻¹)	coefficients of determination (R^2)
FA	4	0.11914	0.98616
	5	0.01835	0.86718
	6.2	0.00913	0.88802
	7.7	0.01240	0.97747
TA	4	0.21968	0.89827
	5	0.02187	0.96061
	6.2	0.01416	0.88335
	7.7	0.01571	0.97318
CP	4	0.27527	0.94872
	5	0.02909	0.93142
	6.2	0.01489	0.89591
	7.7	0.02321	0.97886

Table S4-1. General information of identified transformation products of FA during UVA/chlorination.

Product	Species	m/z	Molecular formula	Relative molecular weight	Diff (ppm)
FA-P ₃₉₄	(M-H)-	393.1507	C ₂₁ H ₂₇ FO ₆	394.1592	1.62
FA-P _{432a}	(M-H)-	431.1866	C ₂₁ H ₃₀ F ₂ O ₇	432.196	2.58
FA-P _{432b}	(M-H)-	431.1869	C ₂₄ H ₂₉ FO ₆	432.1948	1.85
FA-P _{448a}	(M-H)-	447.2019	C ₂₁ H ₃₃ FO ₉	448.2097	2.83
FA-P _{448b}	(M-H)-	447.2019	C ₂₄ H ₃₂ O ₈	448.2097	1.28
FA-P _{450a}	(M-H)-	449.1974	C ₂₁ H ₃₂ F ₂ O ₈	450.2065	2.99
FA-P _{450b}	(M-H)-	449.1985	C ₂₄ H ₃₁ FO ₇	450.2054	-0.57
FA-P ₄₇₀	(M-H)-	469.2034	C ₂₄ H ₃₂ F ₂ O ₇	470.2116	2.26
FA-P ₄₈₆	(M-H)-	485.1763	C ₂₄ H ₃₂ ClFO ₇	486.1821	1.9
FA-P ₅₀₂	(M-H)-	501.1697	C ₂₄ H ₃₂ ClFO ₈	502.177	-0.09
FA-P ₅₀₄	(M-H)-	503.1854	C ₂₄ H ₃₄ ClFO ₈	504.1926	0.82

Table S4-2. General information of identified transformation products of TA during UVA/chlorination.

Product	Species	m/z	Molecular formula	Relative molecular weight	Diff (ppm)
TA-P ₃₅₈	(M-H)-	357.1499	C ₂₁ H ₂₃ FO ₄	358.158	1.97
TA-P ₃₆₅	(M-H)-	355.1551	C ₂₁ H ₂₄ O ₅	365.1624	0.04
TA-P ₄₆₅	(M-H)-	455.2026	C ₂₃ H ₃₃ FO ₈	465.2159	2.95
TA-P ₄₇₆	(M-H)-	475.2014	C ₂₃ H ₃₇ ClO ₈	476.2177	0
TA-P ₅₀₆	(M-H)-	505.1848	C ₂₃ H ₃₅ ClO ₁₀	506.1919	-0.36

Table S4-3. General information of identified transformation products of CP during UVA/ chlorination.

Product	Species	m/z	Molecular formula	Relative molecular weight	Diff (ppm)
CP-P ₄₆₄	(M-H)-	463.1881	C ₂₂ H ₃₄ ClFO ₇	464.1977	2.37
CP-P ₄₆₈	(M-H)-	467.1981	C ₂₅ H ₃₄ ClFO ₅	468.2079	2.8
CP-P ₄₇₀	(M-H)-	469.2243	C ₂₄ H ₃₅ FO ₈	470.2316	-0.06
CP-P _{482a}	(M-H)-	481.1772	C ₂₂ H ₃₆ Cl ₂ O ₇	482.1838	-2.84
CP-P _{482b}	(M-H)-	481.1781	C ₂₅ H ₃₂ ClFO ₆	482.1871	1.65
CP-P ₄₈₄	(M-H)-	484.1948	C ₂₅ H ₃₄ ClFO ₆	484.2028	2.98
CP-P ₅₀₂	(M-H)-	501.1429	C ₂₄ H ₃₂ Cl ₂ O ₇	502.1525	2.29
CP-P ₅₁₀	(M-H)-	509.1740	C ₂₃ H ₃₆ Cl ₂ O ₈	510.1787	-2.93
CP-P ₅₁₂	(M-H)-	511.1893	C ₂₃ H ₃₈ Cl ₂ O ₈	512.1944	-2.76

Table S5-1. General information of identified transformation products of FA during ozonation.

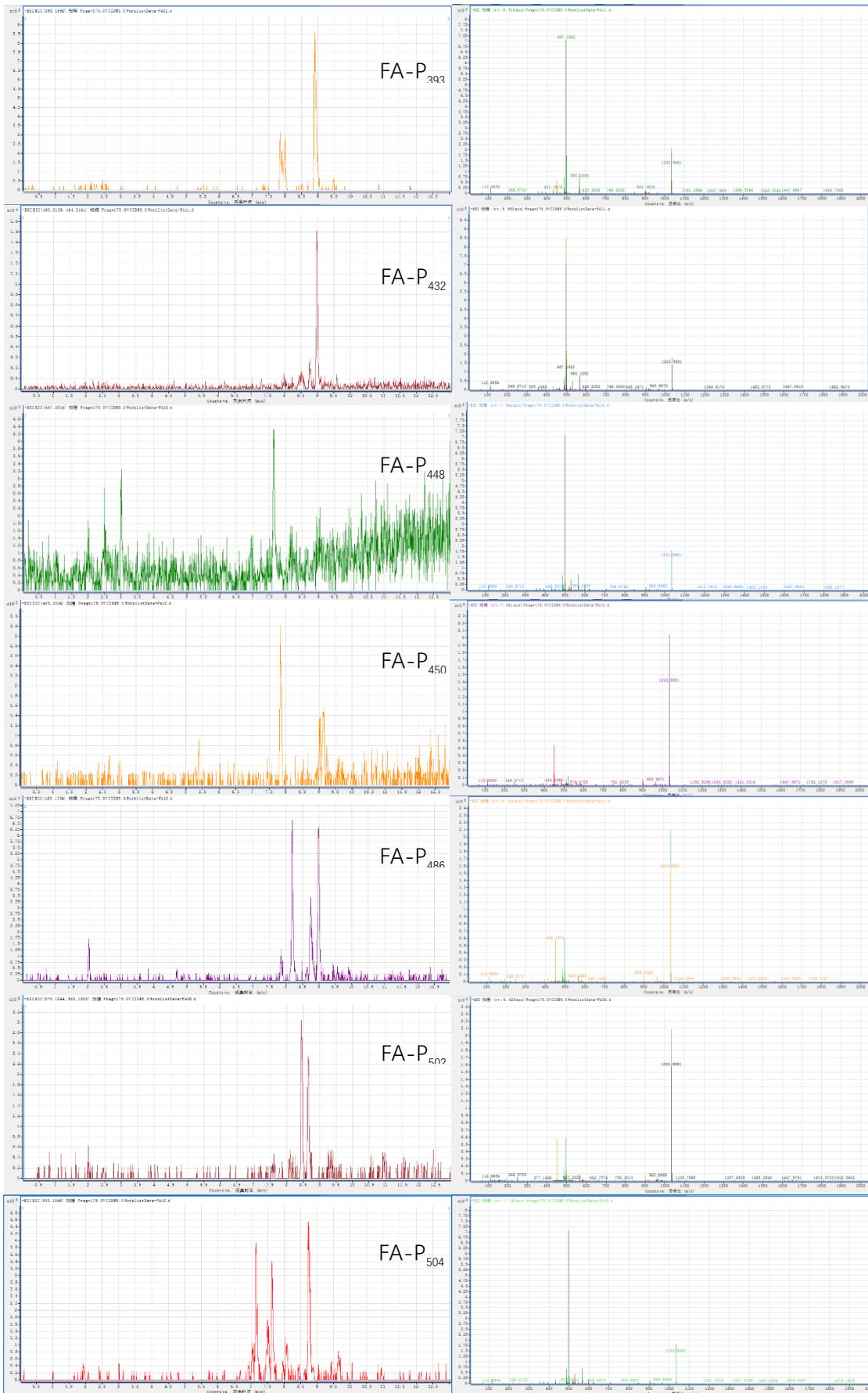
Product	Species	m/z	Molecular formula	Relative molecular weight	Diff (ppm)
FA-P _{382a}	(M-H)-	381.1709	C ₂₀ H ₂₇ FO ₆	382.1792	2.11
FA-P _{382b}	(M-H)-	381.1707	C ₂₃ H ₂₆ O ₅	382.178	0.42
FA-P ₄₀₀	(M-H)-	399.1819	C ₂₀ H ₂₉ FO ₇	400.1897	1.14
FA-P ₄₀₆	(M-H)-	405.1709	C ₂₂ H ₂₇ FO ₆	406.1792	1.39
FA-P ₄₁₀	(M-H)-	409.1667	C ₂₁ H ₂₇ FO ₇	410.1741	0.58
FA-P ₄₃₂	(M-H)-	431.1867	C ₂₁ H ₃₀ F ₂ O ₇	432.196	2.45
FA-P ₄₄₂	(M-H)-	441.1737	C ₂₂ H ₂₈ F ₂ O ₇	442.1803	-1.5
FA-P ₄₆₄	(M-H)-	463.1778	C ₂₁ H ₃₀ F ₂ O ₉	464.1858	1.12
FA-P ₄₁₄	(M-H)-	413.1602	C ₂₃ H ₂₆ O ₇	414.1679	1.71
FA-P ₄₁₆	(M-H)-	415.1757	C ₂₃ H ₂₈ O ₇	416.1835	1.36
FA-P ₄₂₄	(M-H)-	423.1816	C ₂₂ H ₂₉ FO ₇	424.1897	2.28
FA-P ₄₃₄	(M-H)-	433.1671	C ₂₀ H ₂₈ F ₂ O ₈	434.1752	2.17
FA-P ₄₃₆	(M-H)-	435.1757	C ₂₃ H ₂₉ FO ₇	436.1898	0.62
FA-P ₄₃₈	(M-H)-	437.1766	C ₂₃ H ₂₈ F ₂ O ₆	438.1854	2.9
FA-P ₄₇₂	(M-H)-	471.1836	C ₂₃ H ₃₀ F ₂ O ₈	472.1909	0.07

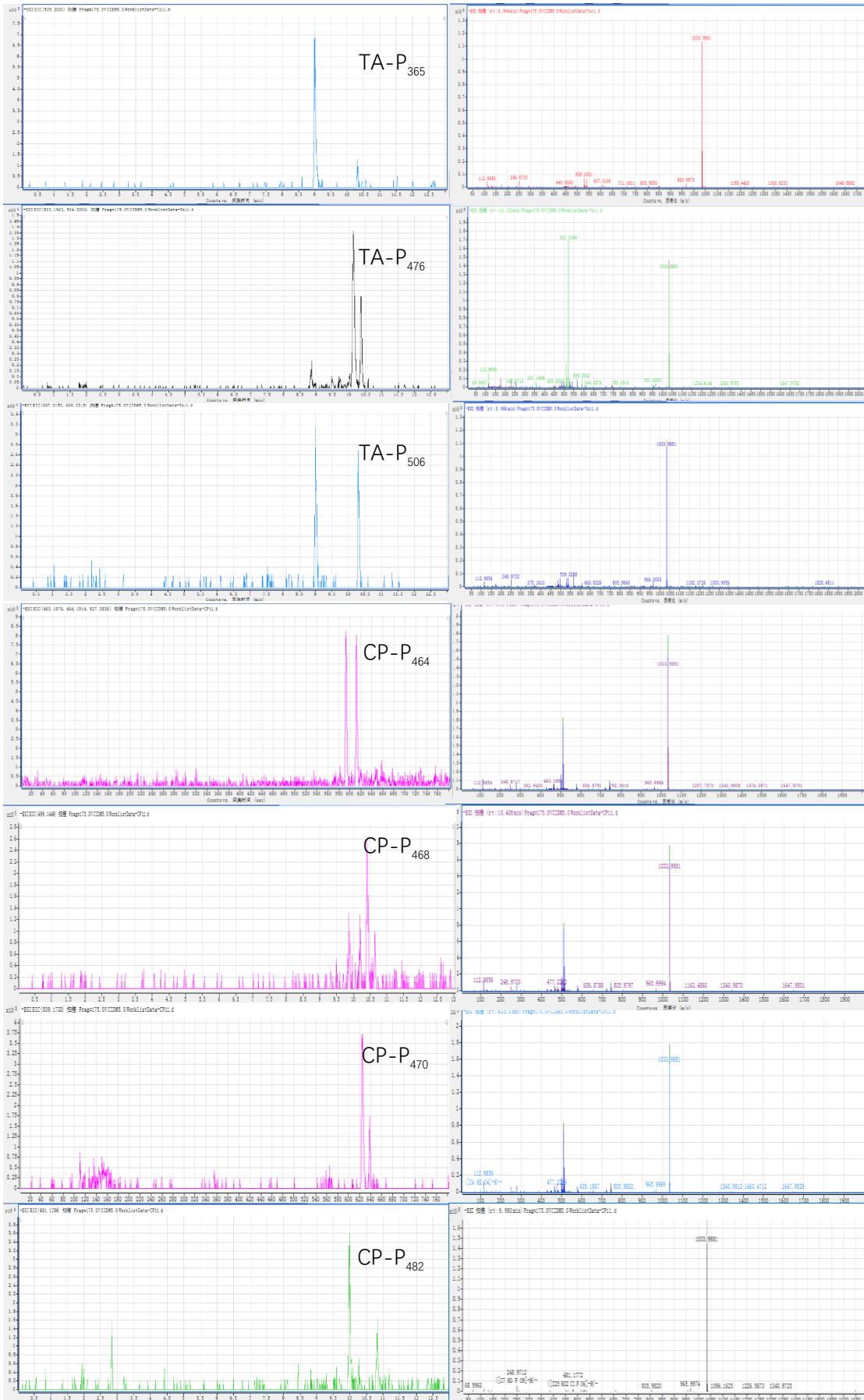
Table S5-2. General information of identified transformation products of TA during ozonation.

Product	Species	m/z	Molecular formula	Relative molecular weight	Diff (ppm)
TA-P ₃₉₆	(M-H)-	395.1508	C ₂₀ H ₂₅ FO ₇	396.1584	0.88
TA-P ₄₂₂	(M-H)-	421.1865	C ₂₂ H ₃₀ O ₈	422.1941	0.58
TA-P ₄₂₄	(M-H)-	423.2015	C ₂₂ H ₃₂ O ₈	424.2097	1.9
TA-P ₄₃₀	(M-H)-	429.1901	C ₂₄ H ₃₀ O ₇	430.1992	2.45
TA-P _{432a}	(M-H)-	431.2074	C ₂₁ H ₃₃ FO ₈	432.2159	2.73
TA-P _{432b}	(M-H)-	431.2087	C ₂₄ H ₃₂ O ₇	432.2148	-1.54
TA-P ₄₄₂	(M-H)-	441.1930	C ₂₂ H ₃₁ FO ₈	442.2003	0.09
TA-P _{448a}	(M-H)-	447.2020	C ₂₁ H ₃₃ FO ₉	448.2109	2.89
TA-P _{448b}	(M-H)-	447.2022	C ₂₄ H ₃₂ O ₈	448.2097	0.54
TA-P ₄₅₈	(M-H)-	457.1865	C ₂₂ H ₃₁ FO ₉	458.1952	2.99
TA-P ₄₇₆	(M-H)-	475.1971	C ₂₂ H ₃₃ FO ₁₀	476.2058	1.45
TA-P ₄₆₀	(M-H)-	459.2020	C ₂₂ H ₃₃ FO ₉	460.2190	2.82
TA-P ₄₈₄	(M-H)-	483.2032	C ₂₄ H ₃₃ FO ₉	484.2109	0.38
TA-P ₄₈₈	(M-H)-	487.1982	C ₂₃ H ₃₃ FO ₁₀	488.2058	0.9

Table S5-3. General information of identified transformation products of CP during ozonation.

Product	Species	m/z	Molecular formula	Relative molecular weight	Diff (ppm)
CP-P ₄₁₂	(M-H)-	411.1576	C ₂₁ H ₂₉ ClO ₆	412.1653	1.54
CP-P ₄₂₄	(M-H)-	423.1570	C ₂₂ H ₂₉ ClO ₆	424.1653	0.67
CP-P ₄₂₈	(M-H)-	427.1531	C ₂₁ H ₂₉ ClO ₇	428.1602	0.64
CP-P ₄₃₂	(M-H)-	431.1623	C ₂₄ H ₂₉ ClO ₅	432.1704	-0.25
CP-P ₄₄₀	(M-H)-	439.1519	C ₂₂ H ₂₉ ClO ₇	440.1602	4.46
CP-P ₄₄₄	(M-H)-	443.1621	C ₂₂ H ₃₀ ClFO ₆	444.1715	2.73
CP-P ₄₆₆	(M-H)-	465.1675	C ₂₄ H ₃₁ ClO ₇	466.1758	3.62
CP-P ₄₈₆	(M-H)-	485.1743	C ₂₄ H ₃₂ ClFO ₇	486.1821	1.32
CP-P ₄₉₀	(M-H)-	489.1684	C ₂₃ H ₃₂ ClFO ₈	490.177	3.05
CP-P ₅₁₅	(M-H)-	514.1534	C ₂₅ H ₃₂ ClFO ₈	515.1832	3.46
CP-P ₄₉₈	(M-H)-	497.1742	C ₂₅ H ₃₂ ClFO ₇	498.1821	0.96
CP-P ₅₀₂	(M-H)-	501.1691	C ₂₄ H ₃₂ ClFO ₈	502.177	2.44





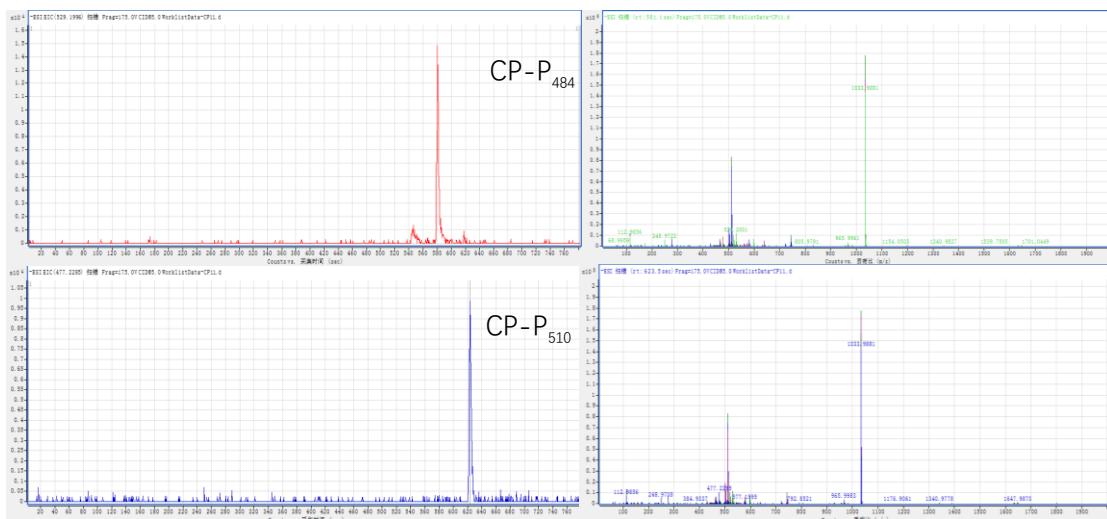
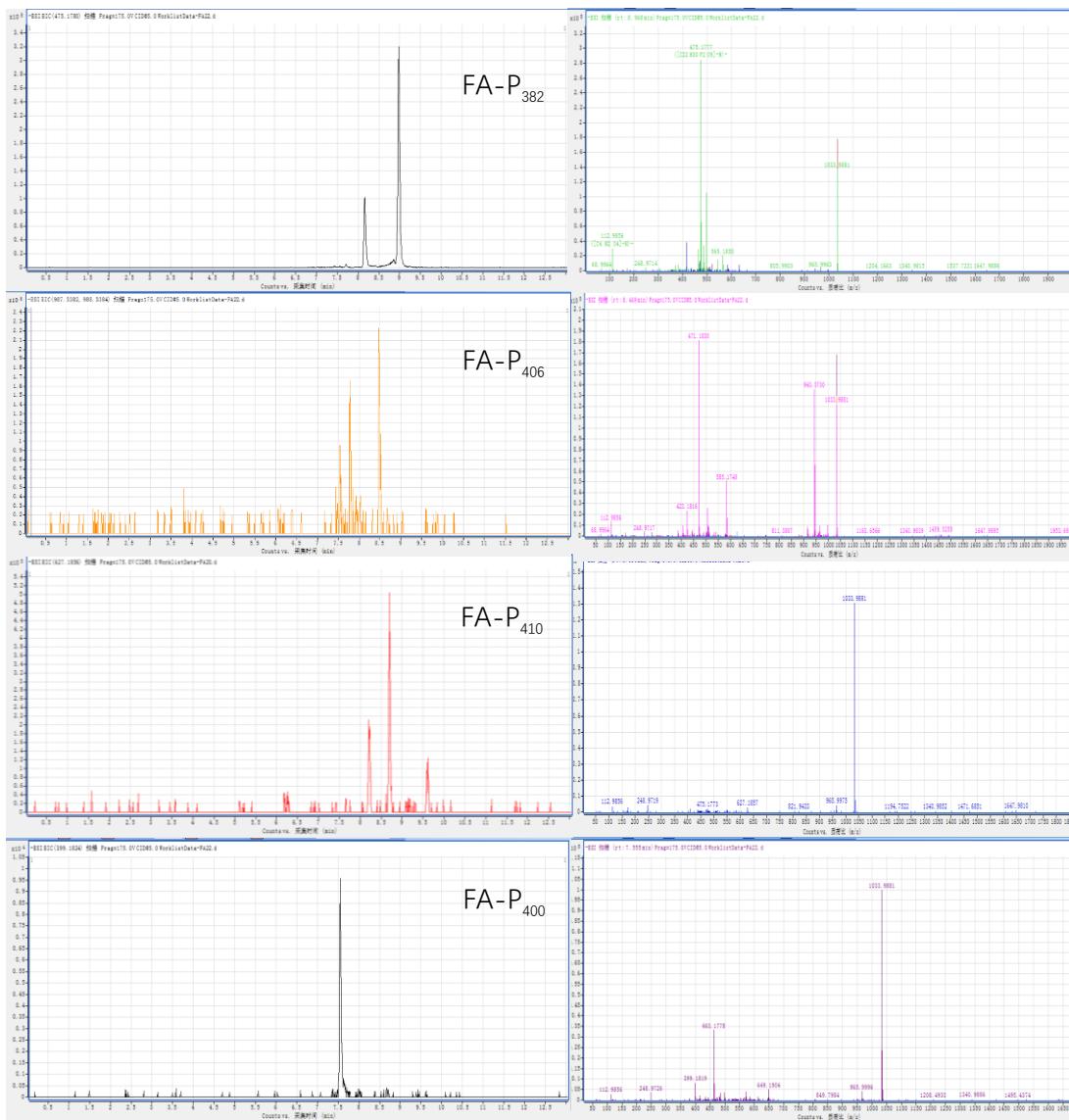
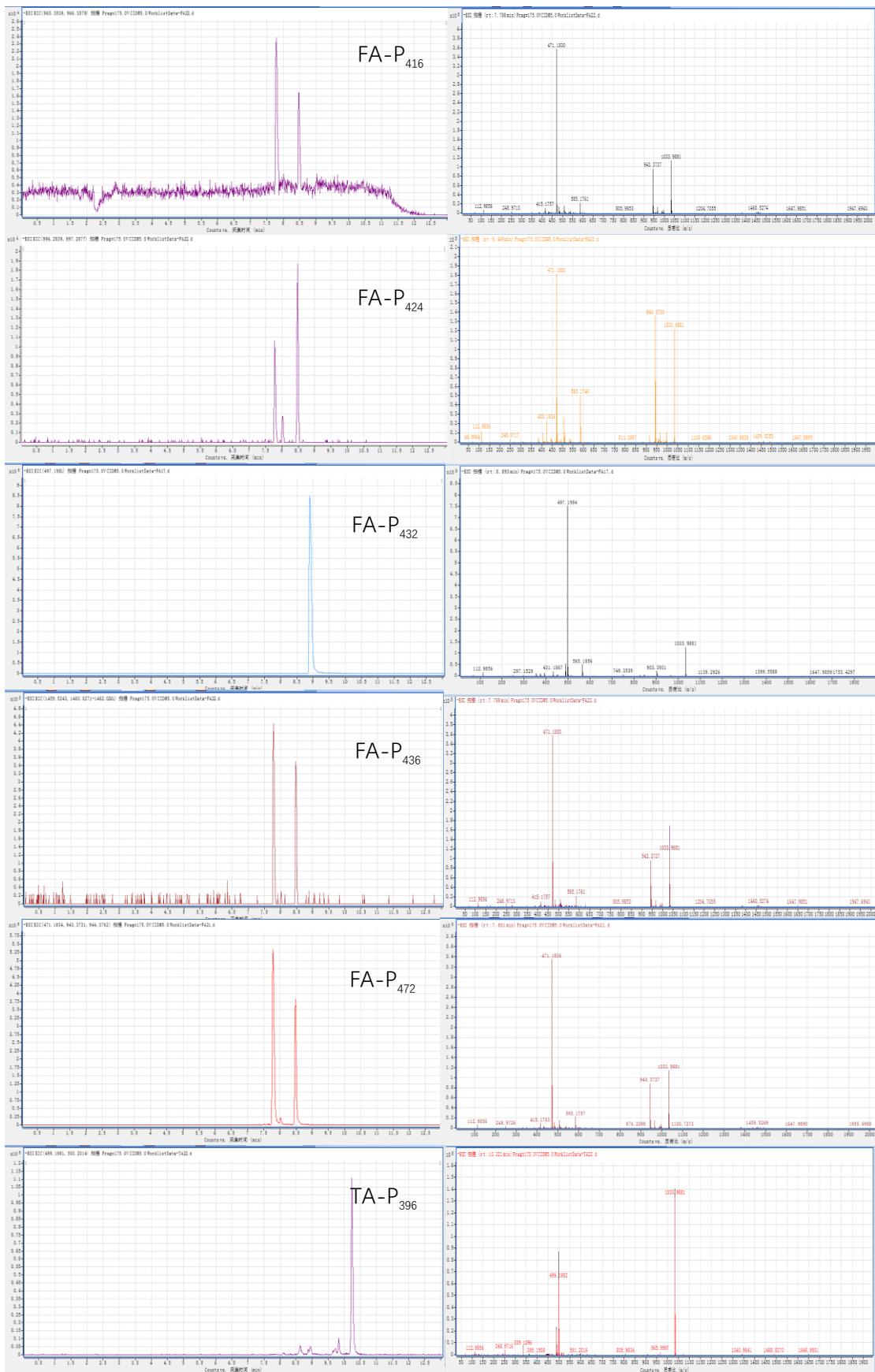
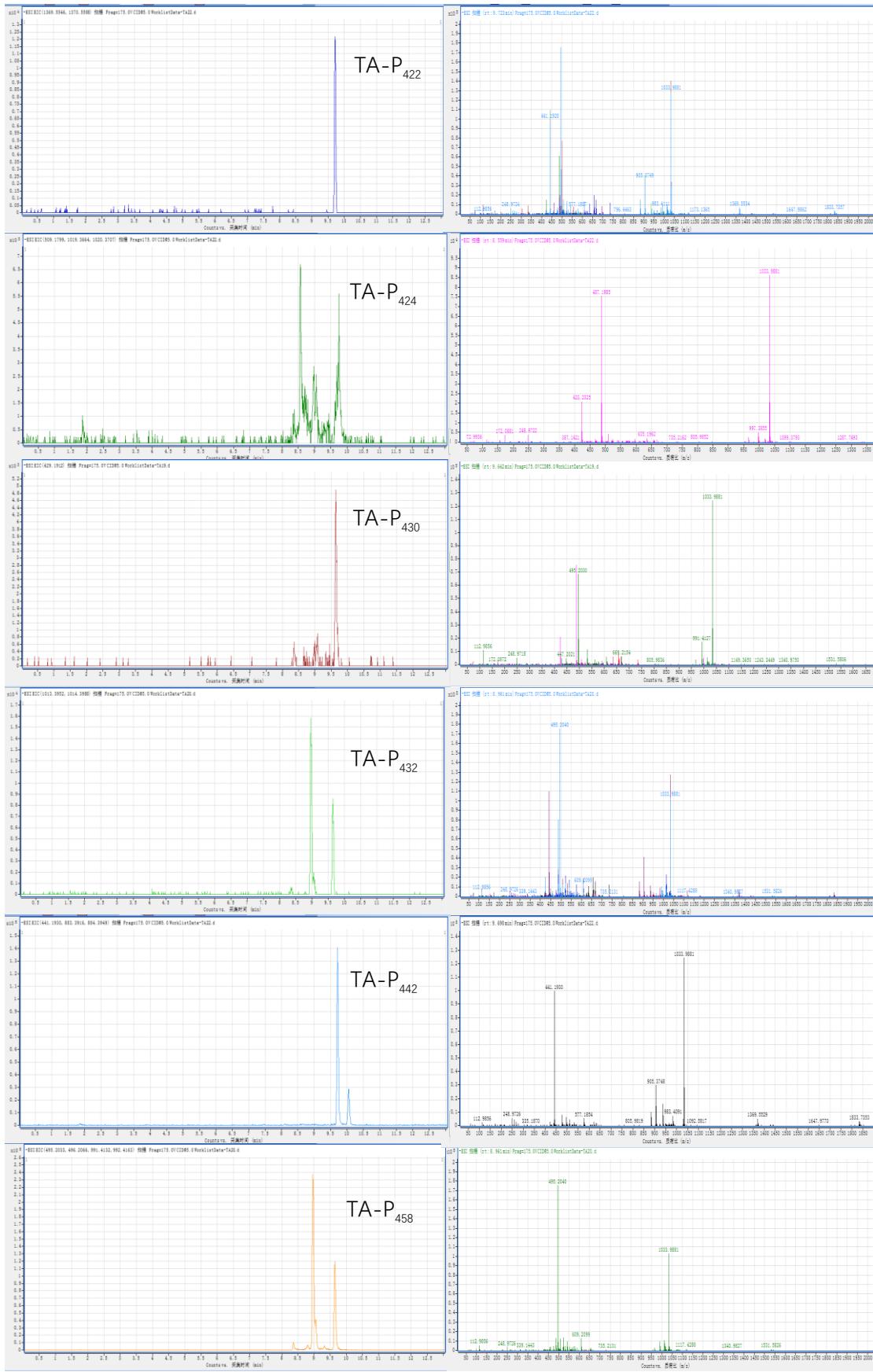
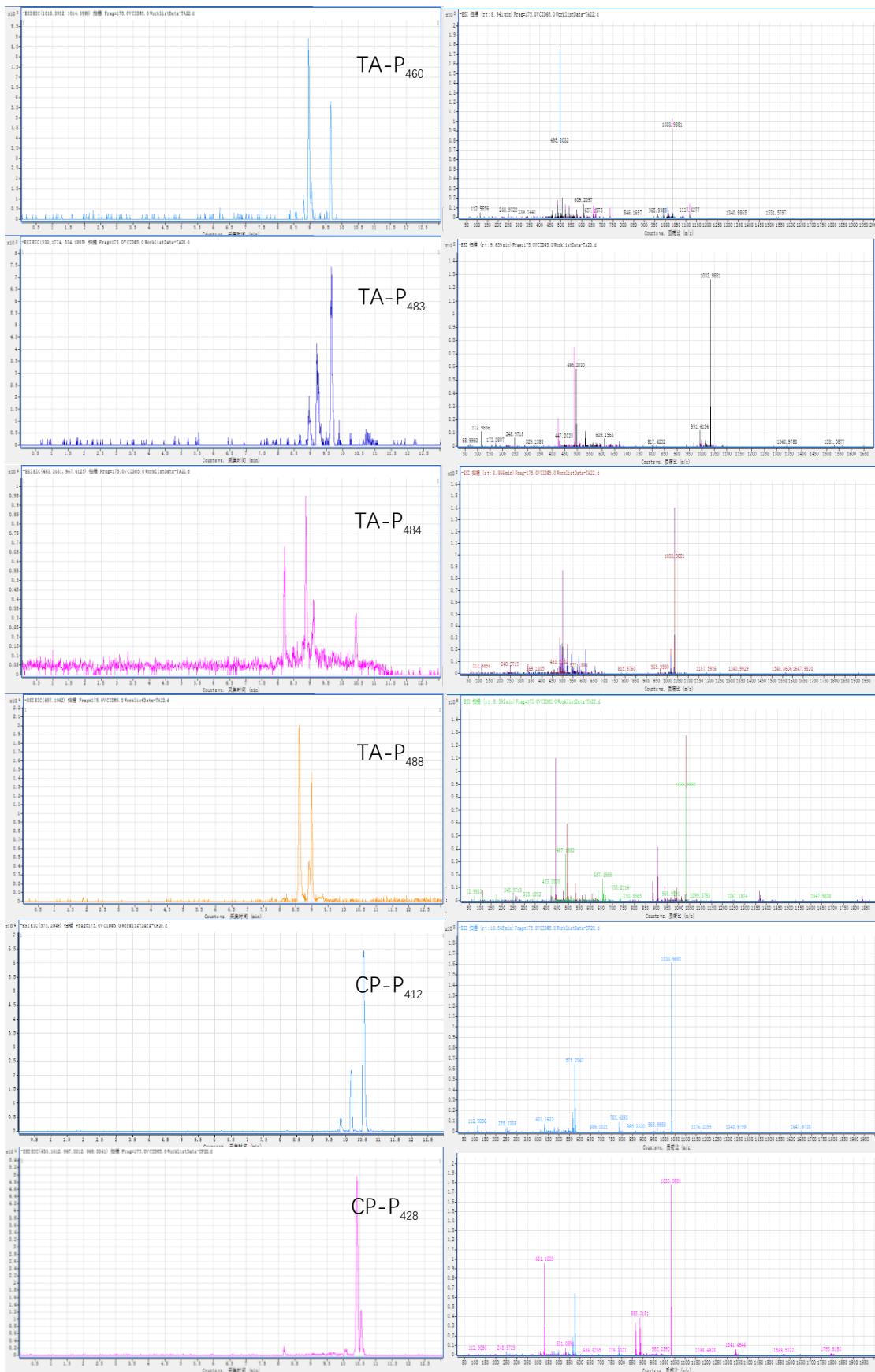


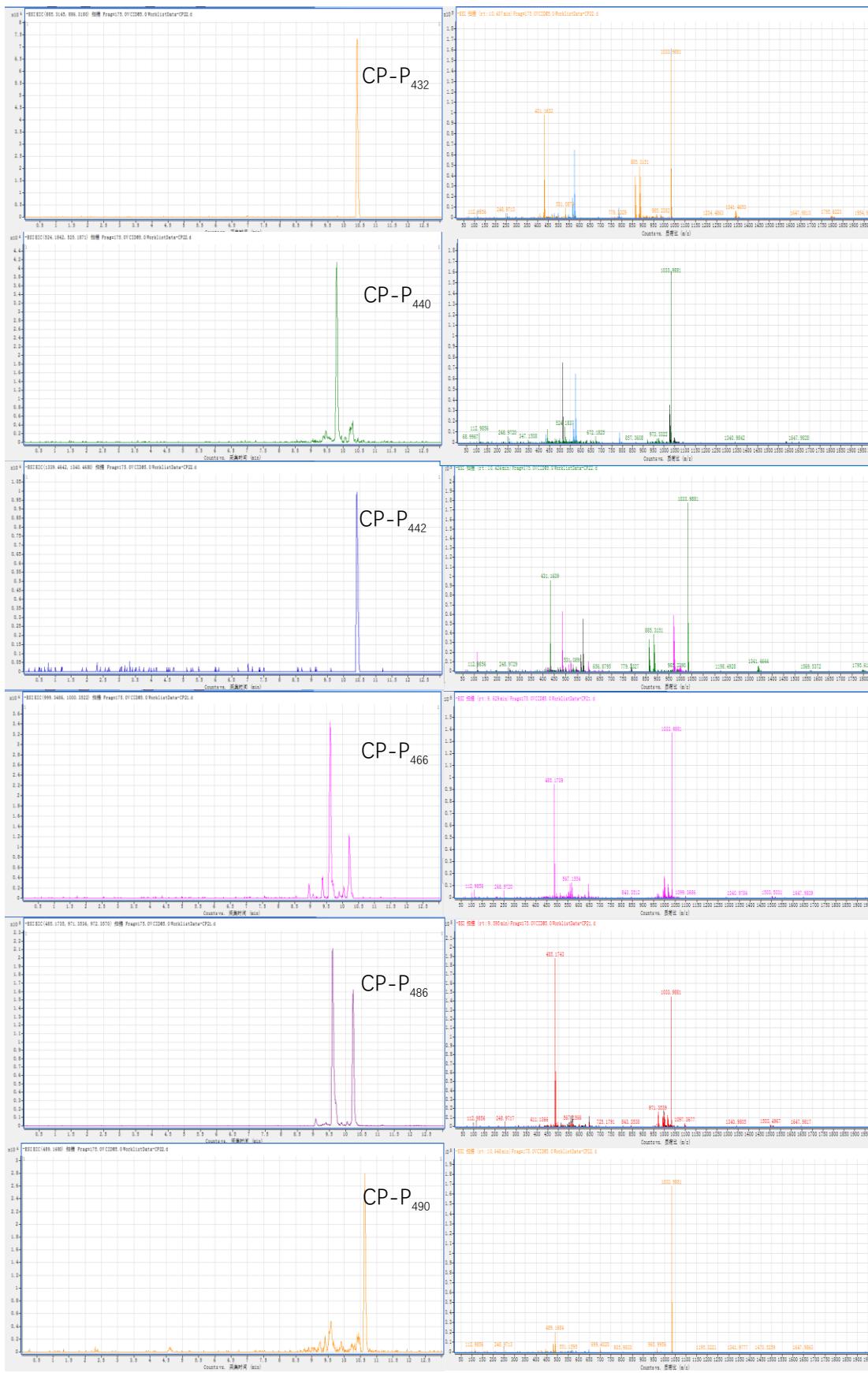
Figure S1-1. Extracted ion chromatographs and mass spectrometry fragments for intermediates of GCs during UVA/chlorination.











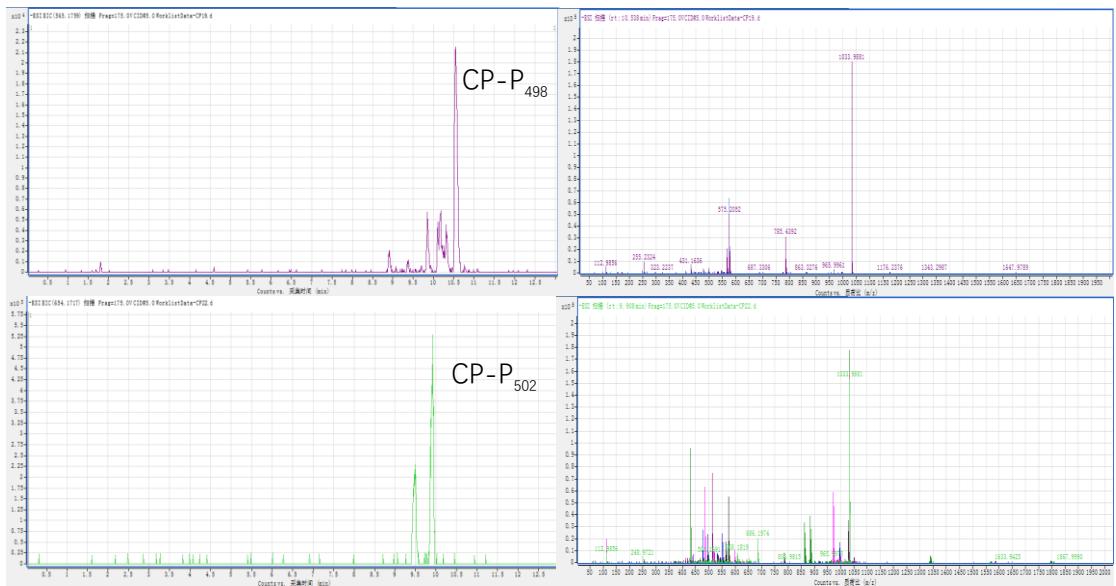


Figure S1.2 Extracted ion chromatographs and mass spectrometry fragments for intermediates of GCs during ozonation.

References

1. Schriks, M.; van Leerdam, J.A.; van der Linden, S.C.; van der Burg, B.; van Wezel, A.P.; de Voogt, P. High-Resolution Mass Spectrometric Identification and Quantification of Glucocorticoid Compounds in Various Wastewaters in The Netherlands. *Environ. Sci. Technol.* **2010**, *44*, 4766–4774. <https://doi.org/10.1021/es100013x>.
2. Wu, S.; Jia, A.; Daniels, K.D.; Park, M.; Snyder, S.A. Trace analysis of corticosteroids (CSs) in environmental waters by liquid chromatography-tandem mass spectrometry. *Talanta* **2019**, *195*, 830–840. <https://doi.org/10.1016/j.talanta.2018.11.113>.
3. Weizel, A.; Schluesener, M.P.; Dierkes, G.; Ternes, T.A. Occurrence of Glucocorticoids, Mineralocorticoids, and Progestogens in Various Treated Wastewater, Rivers, and Streams. *Environ. Sci. Technol.* **2018**, *52*, 5296–5307. <https://doi.org/10.1021/acs.est.7b06147>.
4. Speltini, A.; Merlo, F.; Maraschi, F.; Sturini, M.; Contini, M.; Calisi, N.; Profumo, A. Thermally condensed humic acids onto silica as SPE for effective enrichment of glucocorticoids from environmental waters followed by HPLC-HESI-MS/MS. *J. Chromatogr. A* **2018**, *1540*, 38–46. <https://doi.org/10.1016/j.chroma.2018.02.004>.
5. Ammann, A.A.; Macikova, P.; Groh, K.J.; Schirmer, K.; Suter, M.J.F. LC-MS/MS determination of potential endocrine disruptors of cortico signalling in rivers and wastewaters (vol 406, pg 7653, 2014). *Anal. Bioanal. Chem.* **2016**, *408*, 4821–4822. <https://doi.org/10.1007/s00216-016-9581-1>.
6. Tolgyesi, A.; Verebey, Z.; Sharma, V.K.; Kovacsics, L.; Fekete, J. Simultaneous determination of corticosteroids, androgens, and progesterone in river water by liquid chromatography-tandem mass spectrometry. *Chemosphere* **2010**, *78*, 972–979. <https://doi.org/10.1016/j.chemosphere.2009.12.025>.