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Phosphate Removal from Nursery Runoff Water Using an Iron-Based Remediation System

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Abstract: Phosphorous (P) losses from containerized plant production nurseries can be significant due to the low nutrient retention capacities of the media components. As environmental regulators establish, refine, and enforce nutrient criteria, effective methods are needed to reduce amounts of P in runoff and drainage water. This study investigated the use of a small scale flow-through ferrous iron (Fe(II))-based remediation system for chemically precipitating P. This system consisted of four inter-connected tanks, with the first two maintained under anaerobic conditions and the last two maintained under aerobic conditions. FeSO₄ was introduced into the first of the aerobic tanks at different rates to achieve Fe:P ratios of 0, 9.0, 16.3, and 21.2. Water samples were collected from the systems, and P removal was monitored by ion chromatography. Phosphorus removal efficiencies of 78, 95, and 99% were observed for each respective treatment, indicating great potential for this conceptual system at Fe:P dosing ratios ≥ 16.3 and phosphorus concentrations between 3 and 5 mg/L. This type of system may especially be useful for nurseries with space limitations.

Keywords: nutrient; P; mitigation

1. Introduction

The ornamental horticulture industry is economically significant in the U.S., with wholesale and retail sales sectors generating a total output of \$21.08 billion, 232,648 jobs, \$8.75 billion in employee earnings, and \$13.17 billion in value-added benefits in 2015 [1]. Fertilizer applications are essential for producing high-quality ornamental plants in most commercial operations, especially in situations where the plants are grown in soilless media. Soilless media may be composed of combinations of many different types of raw materials and by-products, including aged pine bark, coir, peat, and sand. Many soilless media components are lightweight and much more porous relative to mineral soils, which reduces handling and shipping costs. One consequence of applying fertilizers to soilless media is that they may leach from the pots where they are applied [2–8]. Leaching of nutrients into nursery drainage ditches and retention ponds may result in increased clogging of filters (due to excessive algae growth) and increased drainageway vegetation management due to excessive growth of aquatic and marginal plants. Likewise, off-site discharges of nutrient-enriched drainage water may result in unwanted shifts of aquatic ecosystems to more algal-dominated systems [2,7–9]. In fact, nutrient pollution from nitrogen and phosphorus is the primary cause of water quality impairments within the state of Florida [10]. Many states, including Florida, are developing or have developed more stringent rules for limiting nutrient discharges into public waterbodies, including numeric water quality criteria [11] and total maximum daily loads (TMDLs) [12] to improve water quality. As a result, plant production nursery managers must consider how to optimize their operations to prevent off-site

discharges of nutrients, and in some cases remove nutrients from drainage water to improve water quality for reuse on the nursery.

Nutrient removal from drainage water using constructed wetlands is one option for nurseries having available land area that can be dedicated to non-plant-production purposes [9,13]. However, in space-limited nurseries, enough land may not be available for building constructed wetlands. In these cases, smaller-scale, high-throughput remediation systems would be particularly useful. Wilson and Albano [14] described such a system for removing nitrate from nursery drainage water. This proof-of-concept anaerobic bioreactor system consisted of four inter-connected tanks filled with Kaldness media. Molasses (injected into the first tanks) was used as a source of carbon for denitrifying microflora. As tested, the system consistently removed 80 to 100% of the nitrate-nitrogen added into the system, but water discharged had a very low redox potential (~-300 to -400 mV). One disadvantage of this system is that the water discharged from the bioreactors is anoxic and has a very low redox potential due to microbial metabolism within the bioreactors. Discharge of this water into waterbodies with living organisms could cause deleterious effects due to the low dissolved oxygen content. Given that the discharge water must be aerated before discharge, it is possible that an aeration module could also be optimized to precipitate phosphorus out of solution at the same time. Ferrous iron (Fe(II)) is stable in aqueous solutions under reducing conditions. Ferrous iron is also present in the groundwater throughout much of Florida. Iron is one of the controlling factors regulating phosphorous (P) release/uptake by sediments under anaerobic and aerobic conditions [15–17]. Under aerobic conditions, Fe(II) is quickly oxidized to ferric iron (Fe(III)), which is not very soluble. In the presence of phosphate (PO_4^{3-}), the change from Fe(II) to Fe(III) can co-precipitate PO_4^{3-} as iron phosphate (FePO₄).

Several studies [18–21] have evaluated the use of iron salts to precipitate PO_4^{3-} from wastewaters. Previous researchers reported that molar ratios of Fe^{2+} :P, as well as pH (circum-neutral), and redox potential (aerobic conditions) are important factors affecting the efficiency for PO_4^{3-} removal from wastewater [18,19,21]. Working with crude sewage at the lab-scale, Thistleton et al. [19] reported that Fe^{2+} :P molar ratios of 1, 1.5, 2, and 3.4, resulted in P removal efficiencies of 52, 60, 72, and 85%, respectively, under aerobic, neutral pH conditions. They suggested that a 3:1 Fe^{2+} :P ratio was optimal for removal of phosphorus from sewage. In an earlier study, Svanks [18] reported P removal efficiencies of 97.4, 98.4, and 100% using $FeSO_4$ -Fe treatment ratios of 1.5, 2.9, and 3.9, respectively. Both studies were conducted in jars at the bench scale. Other researchers evaluated more complex systems for removing P from sewage-impacted water. Zang et al. [21] reported 99% reduction of P concentrations (10 mg P/L initially) in wastewater treated with $FeSO_4$ at an Fe^{2+} :P molar ratio of 2, and under aerobic, neutral pH conditions. Working with a similar system, Wang et al. [20] reported a 97% reduction in P concentrations in sewage water treated with $FeSO_4$ at a Fe^{2+} :P molar ratio of 2.99, with initial and final P concentrations of 6.41 and 0.16 mg P/L, respectively.

Phosphate precipitation using Fe salts is a relatively common practice in municipal wastewater treatment facilities. However, this type of treatment practice has not been adapted for use at smaller scales, such as space-limited ornamental plant nurseries. Development of a phosphate removal module that could work in sequence with the previously described nitrate removal modules could provide a valuable tool for addressing nutrient-related water quality issues. The objectives of the current proof-of-concept study were to evaluate the potential for adding an aerated module with Fe(II) injection to the system described by Wilson and Albano [14] and to evaluate the influence of iron injection rates on the removal efficiency of PO_4^{3-} .

2. Materials and Methods

2.1. System Description

This study was conducted at a 42.9 ha containerized plant nursey (Grandiflora Nursery), located in Alachua, Florida, FL, USA. Based on the system described by Wilson and Albano [14], duplicate

small-scale flow-through bioreactor systems were constructed using inter-connected polyethylene tanks (Figure 1). These systems were set up next to a retention pond located at the lowest elevation of an approximately 14 ha production area on the nursery. In this case, water was pumped from the retention pond into a 242 L polyethylene distribution tank (Figure 1) using a Tsurumi 0.497 kW model LB-480 submersible water pump (Tsurumi Pump, Glendale Heights, IL, USA). The flow rates through the systems were maintained at 7.6 ± 0.2 L/min. Given that the bottom sediments in the pond were very light and easily suspended, the pump was suspended approximately 0.5 m from the water surface using an inflated rubber inner tube for automotive tires. The inner tube was secured to the top of an 18.9 L polyethylene container with 6.3 mm holes drilled throughout the entire surface area (approximately 2.5 cm apart). A layer of nylon window screening was placed within the suspended container, and the pump was then inserted. The window screening was then closed around the pump to keep fish and debris out of the pump. The pump was connected to the distribution tank using approximately 22 m of 1.9 cm i.d. (inner diameter) hose. The flow rate of water into the distribution tank was controlled using a valve.



Figure 1. Diagram of bioremediation treatment-train system. (**A**) Side view showing tank profiles, positions, Kaldness media positions, and connections between tanks in each position. (**B**) Top view of replicate systems showing water inflow and outflow points and points for FeSO₄, air, and nutrient injections. T = tank; M= molasses tank; Fe(II) FeSO₄ injection.

Duplicate bioreactor systems were connected to the distribution tank using 5.08 cm i.d. polyvinyl chloride (PVC) fittings and pipes. Each bioreactor system consisted of four 242 L inter-connected

polyethylene tanks (height: 0.79 m, width at top: 0.79 m). Each tank was filled with 151 L of Kaldness media to provide surface area for microbes for denitrification. The Kaldness media had a measured porosity of 18.5%. Water from the distribution tank entered the top of the first tank in each series. Water from the first tank discharged into the second tank through a 5.08 cm i.d. PVC connector between the bottoms of the tanks. Water flow through the remaining tanks was staggered similarly. For this study, only the first two tanks in each bioreactor series were used for nitrate removal as described in Wilson and Albano [14]. Molasses (Alachua Feed and Seed, Gainesville, FL) was provided as a carbon source for denitrification in the first two tanks (i.e., T1, T2, T5, T6) in each bioreactor series (Figure 1). It was pumped into these tanks at the point-of-entry for water entering from the distribution tank at a rate of 0.7 mL/min using an electronic metering pump (Pulsafeeder, Punta Gorda, FL, USA). The last two tanks in each bioreactor series (T3, T4, T7, T8; Figure 1B) were used to evaluate PO_4^{3-} removal. Five air stones $(30.4 \times 3.8 \times 3.8 \text{ cm})$ connected to a 1.3 kW regenerative blower (model SST30, Pentair, Apopka, FL, USA) capable of producing air flows of 1.7 m³/min at an elevation of 1.2 m. The stones were then added to these tanks to reestablish aerobic conditions conducive to PO₄³⁻ precipitation. In addition, 20% iron sulfate (FeSO₄; Southern Ag., Orlando, FL, USA) was pumped into the first of these two aerobic tanks (T3 and T7) in each series using another electronic metering pump (Pulsafeeder, Punta Gorda, FL, USA) at variable rates depending on the trial.

To evaluate the effects of FeSO₄-dosing rates on the removal of PO₄^{3–} from the water, known amounts of PO₄^{3–} were pumped into the distribution tank. This arrangement was necessary due to the variable and sometimes low concentrations (below method detection limits) of PO₄^{3–} present in the retention pond water. In this case, a concentrated nutrient stock solution was mixed in a separate tank using Peter's Professional 20N-10P-20K General Purpose fertilizer (Alachua Feed and Seed, Gainesville, FL, USA). The Peters fertilizer was mixed in batches at a rate of 11.3 kg/208 L of water from the retention pond. This nutrient stock solution was injected into the distribution tank at a rate of 8 mL/min using an electronic metering pump (Pulsameter, Punta Gorda, FL, USA), with a target concentration in the distribution tank being $1.79 \pm 0.72 \text{ mg/L PO4}^{3-}$ -P. This concentration is within the range of PO4^{3–}-P concentrations reported in nursery runoff water in previous studies [4,9]. After flow-through acclimation to the pond water and nutrient solution, the effects of FeSO₄ injection rates were evaluated. A stock solution of FeSO₄ (65 g Fe²⁺/L) was injected at rates of 0, 2.0, 3.6, and 4.7 mL/min to achieve Fe:P molar ratios of 0, 9.0, 16.3, and 21.2, respectively.

2.2. Sample Collection and Analysis

Physicochemical properties were measured on each sampling day. Eh measurements were taken using an Accumet pH/volt meter (Fisher Scientific, Waltham, MA, USA) equipped with a flat chlorinated polyvinyl chloride (CPVC) redox electrode (Cole-Parmer, Vernon Hills, IL, USA). Water pH was measured using a Yellow Springs Instruments (YSI) data sonde equipped with a 650 Multiparameter Display System (Yellow Spring Instruments Inc., Yellow Springs, OH, USA).

Water samples were hand-collected from each tank at 1-hr intervals for 2 to 3 h on each day of evaluation, depending on weather conditions. Immediately after collection, samples were filtered through a 0.2 μ m syringe filter (Thermo Scientific, Rockwood, TN, USA) into 50 mL polypropylene centrifuge tubes (Fisher Scientific, Waltham, MA, USA) and stored in a cooler at 4 °C for transport to the lab in Gainesville, FL where they were frozen, then subsequently shipped to and analyzed at the USDA-ARS Horticultural Research Laboratory, Ft. Pierce, FL, USA. Dissolved PO₄³⁻ concentrations were quantified in samples by ion chromatography following the guidance of U.S. Environmental Protection Agency [22] (1997) and Nesterenko [23]. Briefly, analyses were conducted using an ICS 1000 (Dionex Corp., Sunnyvale, CA, USA) fitted with an IonPac AS14 column (Dionex Corp., Sunnyvale, CA, USA), operating at 1800 to 1900 psi with sodium bicarbonate as the eluent, and with the detector and suppressor set at 17 mS and 24 milliamps, respectively. The system was fitted with a 75 μ L sample loop. Known-concentration standards and distilled-deionized water method blanks were used to ensure accurate and precise quantification.

2.3. Data Analysis

Phosphate concentrations at each position within the treatment-train of the duplicate systems (i.e., 1st, 2nd, 3rd, and 4th tank positions in each series) were combined for all samples collected during each FeSO₄-dosing scenario for analysis. Data were subjected to analysis of variance (ANOVA; P = 0.05) with means comparisons using Student's *t*-test (JMP Pro 14, SAS Institute, Inc., Cary, NC, USA) to identify statistically significant changes in PO₄^{3–} concentrations relative to FeSO₄-dosing rates and positions within the treatment-train. In an effort to confirm precipitation of phosphorus and predict its identity in precipitated form, the equilibrium speciation model Visual MINTEQ 3.1 program (Jon Petter Gustafsson, KTH, Sweden; https://vminteq.lwr.kth.se/) was populated with physical and chemical parameters associated with Position 3 in the treatment system. While equilibrium was not reached in the flow-through system, this model has been used successfully to predict precipitation in non-static swine waste management operations [24]. Input data are provided in Tables S1 and S2. Scenarios for each Fe²⁺ dosing ratio and a mean PO₄^{3–} concentration of 5.5 mg/L were evaluated.

3. Results

Variable FeSO₄-dosing rates were chosen to compare the effective PO_4^{3-} removal efficiency associated with increasing Fe²⁺:P molar ratios. A summary of pH and Eh values associated with these treatments is shown in Table 1. pH ranged from 7.2 to 8.4 in the controls, and from 7.5 to 9.6 during the Fe²⁺ dosing events. pH generally increased as the season progressed, which may have been associated with an algal bloom that occurred in the retention pond serving as the water source. pH generally decreased to minimal values in the last position for nitrate removal due to microbial respiration and release of organic acids associated with the molasses [14]. Likewise, Eh values were generally highest in the incoming water, decreasing to minimal levels in position 2 tanks as would be expected due to microbial metabolism [14]. Eh was not restored to control levels in position 4 tanks, indicating that additional aeration would be needed for complete restoration of the water to initial conditions.

Fe:P Ratio	Position				
	Distribution Tank	1	2	3	4
		pН			
0	7.9 ± 0.4	7.5 ± 0.3	7.16 ± 0.2	7.9 ± 0.2	8.4 ± 0.2
9.0	9.0 ± 0.9	8.0 ± 0.3	7.5 ± 0.2	7.7 ± 0.6	8.1 ± 0.6
16.3	9.4 ± 0.3	8.7 ± 0.3	7.9 ± 0.3	8.6 ± 0.1	9.4 ± 0.1
21.2	9.6 ± 0.4	9.0 ± 0.2	8.4 ± 0.1	8.2 ± 0.1	8.8 ± 0.1
		Eh			
0	114 ± 16	14 ± 15	-16 ± 20	114 ± 14	120 ± 11
9.0	29 ± 26	-154 ± 24	-233 ± 27	-146 ± 25	-72 ± 19
16.3	74 ± 43	-127 ± 22	-204 ± 27	-143 ± 12	-41 ± 18
21.2	136 ± 32	-90 ± 18	-178 ± 26	-112 ± 14	-61 ± 9

Table 1. Summary of pH and Eh (mV) measurements in tanks at each position of the treatment-train.

Phosphate-P concentrations at each position of the treatment-train as affected by FeSO₄-dosing rates are shown in Figure 2. As expected, no significant changes in P occurred in any of the positions when no FeSO₄ was injected into the systems, although there was a nominal decrease of 20 to 27% in the third and fourth positions (Figure 2A). Precipitation of calcium phosphates have been shown to be more favorable in conditions with higher pH (>8.5) and Ca/P ratios >1.67 [25], which could partially explain the reduction observed. Injection of FeSO₄ at a rate of 2 mL/min (9.0 Fe²⁺:P ratio) reduced PO₄^{3–}-P concentrations by 77 and 80% in tank positions 3 and 4, respectively (Figure 2B); while injection at rates of 3.6 (16.3 Fe²⁺:P ratio) and 4.7 mL/min (21.2 Fe²⁺:P ratio) reduced PO₄^{3–}-P concentrations by 94 to 96% (Figure 2C) and 97 to 100% (Figure 2D) in the respective positions.



Figure 2. Phosphate concentrations relative to position within the treatment-train and FeSO₄ dosing rate. Different letters indicate differences amongst different Fe²⁺:P dosing ratios relative to positions as indicated by ANOVA with LSD Means calculations (P = 0.05).

The effects of FeSO₄ dosing rates of PO_4^{3-} -P concentrations at each separate position within the treatment-train are compared in Figure 3. Concentrations within position 1 tanks were statistically similar at all FeSO₄ dosing rates as would be expected since these tanks are farthest removed from the point of FeSO₄ injection (Figure 3A). Phosphate-P concentrations in tanks at position 2 were significantly lower than the zero-FeSO₄-dosing rate trials during the 16.3 Fe²⁺:P-dosing trials (even though FeSO₄ should not have been present), but not during the 9.0 and 21.2 Fe²⁺:P-dosing events (Figure 3B). One possible explanation for these differences may be due to the FeSO₄ solution being injected into the position 3 tanks which, along with position 4 tanks, were forcefully aerated to rapidly mix the Kaldness media in the tanks. This forceful agitation may have resulted in backflow of FeSO₄-dosed water through the connection between the position 2 and 3 tanks (Figure 1A) resulting in the lower PO₄³⁻-P concentrations detected. Given that the currents within tanks changed as the air stone positions were changed with routine maintenance, it is impossible to predict the extent of

possible backflow. Phosphate concentrations dramatically decreased in positions 3 and 4 where the FeSO₄ was injected into the system (Figure 3C,D). Phosphate-P concentrations in position 3 decreased by 74, 96, and 100% at the respective Fe²⁺:P-dosing ratios of 9.0, 16.3, and 21.2, relative to the control. Likewise, $PO_4^{3^-}$ -P concentrations in position 4 tanks were reduced by 75, 94, and 97% at the same respective Fe²⁺:P-dosing rates. Based on these results, a minimal FeSO₄ addition rate of 3.6 mL/min (Fe²⁺:P ratio = 16.3) was enough to consistently remove the PO₄³⁻ from solution. The equilibrium speciation model Visual MINTEQ predicted that nearly all of the PO₄³⁻ (5.497–5.498 mg/L; Tables S3 and S4) should have precipitated under all of the FeSO₄-dosing scenarios evaluated, with vivianite (Fe₃(PO₄)₂·8H₂O) being the dominant precipitated mineral.



Figure 3. Phosphate concentrations (mg/L) relative to FeSO₄ dosing rates in tanks at positions 1, 2, 3, and 4 in the bioremediation treatment-train. Different letters indicate differences amongst positions relative to Fe²⁺:P dosing ratios as indicated by ANOVA with LSD Means calculations (P = 0.05).

4. Discussion

In the current study, greater than 94% removal only occurred at Fe^{2+} :P molar dosing ratios of 16.3 and higher, which is in contrast with results previously reported by Zang et al. [21], and Wang et al. [20], Thistleton et al. [19], and Svanks et al. [18]. A likely explanation for this difference is the fact that Zang et al. [21] and Wang et al. [20] were working with much smaller scale (30 L) membrane bioreactors with hydraulic retention times of 10 h as opposed to approximately 0.2 h per tank in the current systems. Additionally, their systems partially recycled the water being treated, as opposed to treating the water in a single pass. Likewise, the earlier jar studies (Thistleton et al. [19]; Svanks et al. [18]) were not conducted under flow-through conditions. As a result, there was likely more time and opportunity for the ions to comingle and precipitate in the non-flow-through systems. Thus, the need for higher molar ratios of Fe^{2+} :P in the current study was likely due to inefficiencies associated with mixing within the single-pass, flow-through systems.

One issue that was encountered is the release of the precipitated Fe(III) iron from the remediation systems. Given the forcefulness of the aeration system, precipitated iron did not settle out from the water column within tanks at positions 3 and 4, but instead exited the systems with the outflow. This resulted in orange staining of sediments in the lake into which the systems discharged. To counteract this occurrence, a 75 L polyethylene container was added after the water exited the systems to allow for settling. This tank was filled with nylon window screening to baffle and slow the flow rate, and further facilitate settling and recovery of the precipitated iron phosphate. According to Wilfert et al. [26], vivianite is the dominant (70–90%) iron phosphate mineral released in chemically treated sewage sludge. In the current study, analysis with Visual MINTEQ predicted similar results. Vivianite may be separated magnetically from sludge [26–28]. Approximately 60% of PO₄^{3–} within this material may be recovered [28]. This recovered vivianite can be used as fertilizer which may serve to offset some of the system operating costs. Dissolving vivianite with potassium hydroxide may also precipitate the iron fraction for reuse in PO₄^{3–} removal.

The current system is a small-scale design to determine proof-of-concept. At full scale, a subterranean system is envisioned as the ideal design to provide dual use of space in production areas and roadways. Nutrient-enriched surface water would drain through the system by gravity, rather than by the utilization of pumps. Site-specific engineering and design are needed to accommodate typical flow volumes and nutrient loadings. Implementation of a pilot-scale study would be beneficial to gain insight into how the system would perform under site-specific conditions. Consumable expenses for the operation of the system include the purchase of iron salts and molasses. In the current study, a 22.6 Kg (50 lb) bag of FeSO₄ and a 18.9 L (5 gal.) container of molasses were purchased for \$17.11 and \$24.25, respectively. Treating approximately 460 L/h of pond water according to the 16.3 molar dosing ratio would require an estimated 216 mL/h of FeSO₄ stock solution at a concentration of 176.8 g FeSO₄ distributed at a flow rate of 3.6 mL/min and an estimated 42 mL/h of molasses distributed at a flow rate of 0.7 mL/min. At these rates, costs for FeSO₄ and molasses would be an estimated \$22.46 and \$9.05 per week, respectively.

5. Conclusions

Results from this proof-of-concept study indicate that it is possible to add a PO_4^{3-} -precipitation module to the bioremediation system described by Wilson and Albano [14]. In this case, FeSO₄ quantities needed to effectively remove PO_4^{3-} from the water is much higher than previously reported studies due to the flow-through nature of the system. While the system shows promise, an analysis of the economics associated with the blowers used for aeration is needed to determine feasibility and scales for use.

Supplementary Materials: The following are available online at http://www.mdpi.com/2073-4441/11/4/795/s1, Table S1: Ion concentrations (mg/L) for Visual MINTEQ 3.1 analysis, Table S2: Chemical and physical properties for Visual MINTEQ 3.1 analysis, Table S3: Possible mineral precipitates included in Visual MINTEQ 3.1 analysis, Table S4: Soluble and insoluble fractions of Fe²⁺ and PO₄³⁻ predicted by Visual MINTEQ 3.1.

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