



Article

Nitrogen and Phosphorus Uptake Dynamics in Tropical Cerrado Woodland Streams

Nícolas Reinaldo Finkler ¹, Flavia Tromboni ²,*, Iola Gonçalves Boëchat ³, Björn Gücker ³ and Davi Gasparini Fernandes Cunha ¹

- Department of Hydraulic and Sanitation Engineering, São Carlos School of Engineering, University of São Paulo, São Carlos, São Paulo CEP 13560-590, Brazil; nicolas.finkler@gmail.com (N.R.F.); davig@sc.usp.br (D.G.F.C.)
- Global Water Center and Department of Biology, University of Nevada, Reno, NV 89557, USA
- Department of Geosciences, Federal University of São João del-Rei, São João del-Rei, Minas Gerais CEP 36301-360, Brazil; iboechat@ufsj.edu.br (I.G.B.); guecker@ufsj.edu.br (B.G.)
- * Correspondence: flavia.tromboni@gmail.com; Tel.: +1-775-420-7335

Received: 15 July 2018; Accepted: 31 July 2018; Published: 14 August 2018



Abstract: Pollution abatement through phosphorus and nitrogen retention is a key ecosystem service provided by streams. Human activities have been changing in-stream nutrient concentrations, thereby altering lotic ecosystem functioning, especially in developing countries. We estimated nutrient uptake metrics (ambient uptake length, areal uptake rate, and uptake velocity) for nitrate (NO₃-N), ammonium (NH₄-N), and soluble reactive phosphorus (SRP) in four tropical Cerrado headwater streams during 2017, through whole-stream nutrient addition experiments. According to multiple regression models, ambient SRP concentration was an important explanatory variable of nutrient uptake. Further, best models included ambient NO₃-N and water velocity (for NO₃-N uptake metrics), dissolved oxygen (DO) and canopy cover (for NH₄-N); and DO, discharge, water velocity, and temperature (for SRP). The best kinetic models describing nutrient uptake were efficiency-loss (R² from 0.47–0.88) and first-order models (R² from 0.60–0.85). NO₃–N, NH₄–N, and SRP uptake in these streams seemed coupled as a result of complex interactions of biotic P limitation, abiotic P cycling processes, and the preferential uptake of NH₄-N among N-forms. Global change effects on these tropical streams, such as temperature increase and nutrient enrichment due to urban and agricultural expansion, may have adverse and partially unpredictable impacts on whole-stream nutrient processing.

Keywords: low-order streams; nutrient retention; self-purification capacity; Tracer Additions for Spiraling Curve Characterization; tropical water bodies

1. Introduction

Streams are biogeochemical active systems that alter the amount and chemical form of nutrients and organic matter transported from their catchments to downstream systems [1,2]. Stream nitrogen (N) and phosphorus (P) retention is considered an important ecosystem service as it controls the export of these nutrients to downstream rivers, lakes, estuaries, and ultimately oceans, mitigating the negative consequences of cultural eutrophication [3,4]. Human activities have considerably altered nutrient availability, mobility, and distribution in freshwater systems [5–7], affecting ecosystem-wide nutrient and organic matter dynamics [8], and thus freshwater ecosystem functioning.

Variability in N and P retention rates and efficiency has been associated with primary production [9–11], hydrology and stream geomorphology [12,13], carbon availability [14–16], and nutrient limitation and availability [11,17–19]. Several studies have assessed how environmental

Water 2018, 10, 1080 2 of 17

concentration influences the uptake of a particular nutrient [20–23]. Some empirical studies have considered multi-elemental effects and dynamics [24,25], but they are rare in tropical regions (but see [19]). In addition, some authors have explored correlations between N and P uptake in streams including measurements of either nitrate–nitrogen (NO₃–N) [26,27] or ammonium–nitrogen (NH₄–N) [28,29], but rarely both.

Few studies have measured the uptake of soluble reactive phosphorus (SRP), NO_3-N , and NH_4-N together (e.g., [30]), and those suggested that colimitation by N and P can occur in streams [19]. There is increasing evidence of the predominance of colimitation and interactions among multiple elements in both aquatic and terrestrial ecosystems (e.g., [8,31–33]). The assessment of nutrient limitation is important because the dynamics and downstream transport of a limiting nutrient differ from those of a non-limiting one [19]. Understanding the implications of elemental interactions in nutrient retention studies is essential for the management of water quality in streams and downstream ecosystems.

Studies of nutrient dynamics in the tropics are rare, but especially important in developing countries, which experience high rates of population growth and rural-urban migration [34]. In these countries, 90–95% of all sewage, and 70% of industrial waste are still not treated [35]. The lack of sewage treatment and increasing urbanization in catchments can significantly alter in-stream nutrient concentrations [11,36]. Other impacts occurring in tropical regions are related to deforestation of pristine vegetation for pastoral and agricultural uses. These are associated with decreases in stream nutrient concentrations in some tropical catchments, due to the depletion of soil organic matter stocks, but with increases in other catchments, probably due to fertilizer use (see literature review in [37]). Some studies have reported strong correlations between uptake rates and ambient nutrient availability [38–40], while others found no influence of background nutrient concentrations on retention [41], highlighting the need for further research.

Nutrient uptake can be affected by nutrient enrichment in three different ways. First, the biotic community can track nutrient availability, resulting in constant uptake velocity and linear increases in areal uptake rate with increasing nutrient concentration [2,17]. Second, biological processes can be less efficient or saturated under higher ambient concentrations. Saturation or loss of efficiency causes uptake velocity to decline and areal uptake rate to increase in a curvilinear fashion with increasing concentration [11,42,43]. Third, the uptake of a particular nutrient can covary with the relative availability of another nutrient [18,44]. For example, P enrichment can lead to N limitation and stimulate N uptake, and vice-versa.

The aforementioned studies usually examined nutrient enrichment associated with land use gradients (e.g., [18,22,42,45]). Land use can also influence other variables, such as water discharge, light availability, sediment inputs, and ecosystem metabolism, all of which directly or indirectly influence nutrient uptake [46,47]. Here, we investigated a gradient in nutrient concentration across forested, tropical headwater streams. We studied whole-stream NO₃-N, NH₄-N, and SRP uptake across a gradient of ambient stream nutrient concentrations to expand our knowledge about interactions between N and P cycling and how these interactions are affected by nutrient availability in tropical streams. We studied four streams located in the Brazilian Cerrado savanna biome to address the following research questions: (1) How does nutrient uptake vary across a gradient of ambient nutrient availability? (2) How do the absolute and relative availability of one nutrient influence the uptake of the other nutrient? (3) Are there positive or negative relationships between NO₃-N, NH₄-N, and SRP uptake rates? (4) Are nutrient uptake rates higher or lower than those reported for temperate streams in the literature? In general, we hypothesized a higher demand for NH₄-N than for SRP and NO₃–N, because NH₄–N is the N-form preferentially assimilated by biota [45]. More enriched streams were expected to be less retentive than less enriched ones, showing lower areal uptake rates and uptake velocities. Finally, we hypothesized that none of the studied streams would exhibit saturation conditions (i.e., significant fits to the Michaelis-Menten kinetic model) because of the absence of chronical nutrient inputs.

Water 2018, 10, 1080 3 of 17

2. Methods

We carried this study out in four low-order Cerrado streams (discharge less than $20 \text{ L} \cdot \text{s}^{-1}$) located in São Carlos and Brotas (São Paulo, Southeastern Brazil; Table 1; Figure 1), and in each of them, we selected a representative ~100-m long stream reach. The Cerrado biome (Brazilian woodland savannah) is the second-largest South American biome, rich in springs and low-order lotic networks, which contribute to 8 of the 12 large Brazilian river basins [48]. The sampling sites Espraiado (ESP) and Broa (BRO) were located in relatively preserved basins with extensive and dense riparian vegetation, natural substrate, and restricted access. Canchim (CAN) was located in the EMBRAPA Pecuária Sudeste experimental farm, presenting a preserved riparian zone and natural substrata. Mineirinho tributary (TBM) had an urban drainage basin with fragmented riparian vegetation and advanced erosive features; however, it did not receive domestic or industrial effluents and its headwaters were preserved. The studied streams differed in water depth (from 0.04 to 0.48 m), and were relatively narrow with wetted widths ranging from 0.50 to 1.20 m.

Table 1. Characterization of the streams used in this study with geographic coordinate, drainage area, and general reach characterization.

Stream (Site Code)	Geographic Coordinates	Drainage Area (km²)	General Characteristics
Broa (BRO)	22°11′40.93″ S 47°53′55.78″ W	2.68	Natural vegetation, forested and dense riparian zone, natural substrate, located upstream from a natural wetland
Canchim (CAN)	21°57′54.69″ S 47°50′38.02″ W	1.17	Natural vegetation, forested and dense riparian zone, natural substrate, located next to the spring
Espraiado (ESP)	21°58′46.75″ S 47°52′23.11″ W	2.49	Natural vegetation, forested and dense riparian zone, meandric channel, natural substrate
Mineirinho tributary (TBM)	22°00′12.78″ S 47°55′40.82″ W	0.82	Relatively degraded vegetation, natural substrate, presence of erosive features and pluvial runoff

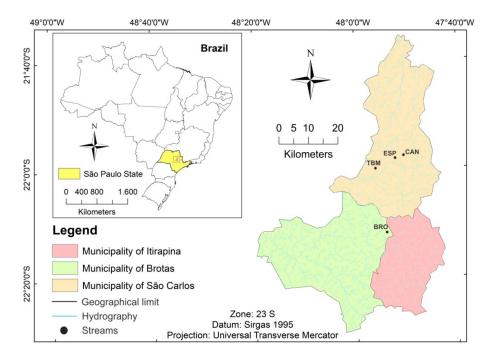


Figure 1. Sampling locations BRO (Broa), CAN (Canchim), ESP (Espraiado), and TBM (Mineirinho River tributary) in southeast Brazil.

Water 2018, 10, 1080 4 of 17

We ran four sets of nutrient additions over the year 2017 (January, April, July, and October), encompassing wet and dry seasons. All addition experiments were carried out under base-flow conditions. We used the tracer additions for spiraling curve characterization (TASCC) approach described in the literature [49] to estimate uptake metrics from pulsed nutrient additions. This method allows determining total ($_{tot\text{-}dyn}$), added ($_{add\text{-}dyn}$), and ambient ($_{amb}$) uptake metrics. We calculated the uptake metrics uptake length ($_{Sw}$), areal uptake rate (U), and uptake velocity ($_{f}$) for different N and P forms according to the nutrient spiraling concept [2].

To all stream reaches, we simultaneously added NO₃-N as NaNO₃, NH₄-N as NH₄Cl, and SRP as K₂HPO₄ as bioavailable reactive tracers to characterize nutrient dynamics, and chloride (Cl⁻) as NaCl as a conservative tracer to characterize stream hydrodynamics. The added mass of conservative tracer was calculated prior to each experiment to increase in-stream electrical conductivity (EC) at measurable, but moderate levels (i.e., 5–10-fold of background EC), while the added mass of nutrients was calculated to raise instream concentrations to up to 2-5-fold of background concentrations [49]. The ratio of NH₄-N to NO₃-N in nutrient additions ranged from 0.25 to 0.62. We dissolved all salts in a 5 L bucket with stream water and then poured the solution carefully into the stream at the top of the experimental reach over one minute (i.e., as a slug). Electric conductivity was measured at 10 s intervals over the experiment using a multiparameter probe at the downstream end of the reach (Model HI 9829, HANNA Instruments, Woonsocket, RI, USA). At this station, we took water samples over the full pulse, with sampling frequency ranging from 15 s to 5 min as a function of EC rate of rise or decline, resulting in 21-26 samples per experiment, in order to obtain a well-characterized breakthrough curve. Immediately before the additions, we collected three water samples to determine nutrient background (i.e., ambient) concentrations. Water samples were filtered immediately upon collection (GF/C Glass Microfiber Membranes, 0.45 µm, Whatman International, Kent, UK) and frozen at −18 °C until analysis. All water samples were analyzed within a maximum of two weeks after collection.

Stream water variables (i.e., pH, EC, temperature, and dissolved oxygen) were measured using the multiparameter probe (HANNA HI 9829, HANNA Instruments, Woonsocket, RI, US). All nutrient concentrations were determined via colorimetry using a Hach DR 4000V spectrophotometer (Hach Environmental, Loveland, CO, USA). The method used for NH₄–N was based on the literature [50], modified for a 7 mL sample volume, and the ones used for SRP and NO₃–N followed the literature [51]. Dissolved organic carbon (DOC) was analyzed using a Shimadzu SSM 5000 TOC analyzer (Shimadzu Corp., Norcross, GA, USA) the combustion method.

We also estimated the canopy cover percentage in each stream reach using a concave densitometer (Forestry Suppliers Inc., Jackson, MS, USA) following the literature [52,53]. Dilution gauging was used to measure stream discharge at the downstream ends of each experimental reach [54]. Air temperature and total precipitation were obtained from a nearby meteorological station belonging to the Brazilian National Institute of Meteorology (INMET, www.inmet.gov.br), located in São Carlos (code: 83726, latitude $-21'96^{\circ}$, longitude $-47'86^{\circ}$, altitude 856 m).

Differences among ambient nutrient concentrations across streams were tested using a Kruskal–Wallis test due to the non-normality of data. Spearman rank correlation was initially used to explore general correlations between uptake metrics and in-stream variables (i.e., physical, chemical, and hydraulic variables). We performed general regression analyses using uptake metrics as dependent variables, and as independent variables, those selected as important uptake drivers based on literature research (e.g., [17,30,55–57]). Among the selected variables, we excluded those that were autocorrelated (i.e., through Spearman rank tests). The remaining set of variables was then submitted to backward stepwise selection to obtain the multiple linear regression models that best described relationships between uptake metrics and environmental variables. We considered data from all streams and samplings together in regression models; therefore, the generated models incorporated typical spatial and seasonal variability of the study region, which we considered to be representative of tropical

Water 2018, 10, 1080 5 of 17

Cerrado woodlands. All data were previously ln(x + 1) transformed, in order to achieve normally distribution, as a prerequisite of linear regression models.

We also adjusted our experimental data to three kinetic models to evaluate the uptake dynamic as function of nutrient concentration. Here, we used the TASCC total dynamics metrics (i.e., total areal uptake rate as $U_{tot\text{-}dyn}$, and total dynamic nutrient concentration as $C_{tot\text{-}dyn}$ [49]). The Michaelis–Menten kinetic model (M–M) represents saturation in uptake when nutrient availability greatly exceeds uptake via biological processes [18]. The efficiency-loss (ELS) kinetic model suggests that nutrient uptake is less efficient at high concentrations, even if saturation is not reached [20]. The first-order kinetic model (FTO) assumes that there is no saturation of uptake rate, and that the relationship between uptake rate and nutrient concentration is positive and linear. This model is often assumed to represent nutrient uptake in pristine, nutrient-limited settings.

Linear regression (p < 0.05) was used to determine the regression fit with the ELS and FTO models. Dependent and independent variables were $\ln(x+1)$ transformed to satisfy the normality of residuals assumption of this method and reduce the effects of extreme values. Fits to the M–M model were processed using untransformed data [20]. Saturation was considered if there was a significant adjustment to the M–M model and the calculated K_m (i.e., the half-saturation constant) was within the range of experimental nutrient concentrations. To fit the data to all three models, we used least squares regressions with the Levenberg–Marquardt algorithm. We conducted statistical analyses using Statistica 10 (Statsoft, Tulsa, OK, USA) and fits to kinetics models and graphical representation were performed with Origin 2017 (OriginLab Corp., Northampton, MA, USA).

3. Results

3.1. General Characterization of the Streams

Total annual precipitation between December 2016 and November 2017 was 1042 mm. Monthly precipitation varied between 0 and 178 mm, with 72% of total rainfall concentrated between December 2016 and May 2017. Stream discharge varied between 2.0 and 19.2 $\rm L\cdot s^{-1}$ considering all sites and sampling dates (Table 2). Water velocities ranged from 0.04 to 0.60 m·s⁻¹. Ranges for EC were 4–32 $\rm \mu S\cdot cm^{-1}$, for water temperature 15–23 °C, for pH 4.4–6.5, and for dissolved oxygen 5.4–8.6 mg·L⁻¹. The riparian zones were dense in all sites with canopy cover varying between 77% and 98%, with ESP as the most forested site (92–98%).

Table 2. Ambient physical and chemical characteristics of studied streams. DOC (dissolved organic carbon), DO (dissolved oxygen), EC (electric conductivity), T (water temperature), Q (stream discharge), v (water velocity), H (depth), w (wetted width), and CC (percentage of canopy cover).

Site	DOC (mg·L ⁻¹)	DO (mg·L ⁻¹)	pН	EC (μS·cm ⁻¹)	T (°C)	Q (L·s ⁻¹)	v (m·s ⁻¹)	H (cm)	w (m)	CC (%)
BRO	1.3-3.8	6.9-7.9	4.4-5.3	4–6	16.5-21.6	7.5-11.2	0.10-0.30	4–16	0.6 - 1.0	90–98
ESP	1.4 - 3.3	5.5-7.4	4.9 - 5.6	11-19	15-20.6	5.0-19.2	0.06 - 0.26	5-48	0.5 - 0.6	93–98
TBM	1.2 - 2.4	7.1 - 8.3	5.8 - 6.1	19-28	17.5-22.8	3.4-6.9	0.10 - 0.60	1–5	1.0-1.2	77–91
CAN	1.6-2.7	7.5–8.5	5.9-6.5	21–32	15-20.5	2.0-4.0	0.04-0.25	1–8	0.9 - 1.1	92–97

Background dissolved organic carbon concentration did not differ significantly among streams (p=0.94) and was always lower than $4.0~{\rm mg\cdot L^{-1}}$ (Table 2). Concentrations of NO₃–N and NH₄–N (Table 3) also did not differ significantly among streams (p=0.47 and p=0.57, respectively), with annual medians of the four studied streams ranging from 37–457 ${\rm \mu g\cdot L^{-1}}$ for NO₃–N, and 5–36 ${\rm \mu g\cdot L^{-1}}$ for NH₄–N. However, there were significant differences in SRP concentrations among streams (p<0.005), with annual medians ranging between 1 and 23 ${\rm \mu g\cdot L^{-1}}$. Accordingly, we classified our streams using background SRP concentrations to facilitate the visualization of SRP limitation effects on nutrient uptake. Therefore, all graphs and tables are presented in the following order of SRP concentration (Table 3): BRO < ESP < TBM < CAN.

Table 3. Ambient uptake length $(S_{w-amb}; m)$, areal uptake rate (U_{amb}) , uptake velocity $(V_{f-amb}; mm \cdot min^{-1})$, and nutrient concentration $(C_{amb}; \mu g \cdot L^{-1})$ during nutrient additions in four tropical streams for each nutrient form expressed as annual medians (minimum-maximum values). SRP—soluble reactive phosphorus.

Site	NH ₄ -N			NO ₃ -N				SRP				
	S _{w-amb} (m)	U_{amb} $(\mu g \cdot m^{-2} \cdot min^{-1})$	V _{f-amb} (mm·min ⁻¹)	C _{amb} (μg·L ⁻¹)	S _{w-amb} (m)	$U_{amb} \ (\mu g \cdot m^{-2} \cdot min^{-1})$	V _{f-amb} (mm·min ⁻¹)	C _{amb} (μg·L ⁻¹)	S _{w-amb} (m)	U_{amb} $(\mu g \cdot m^{-2} \cdot min^{-1})$	V _{f-amb} (mm·min ⁻¹)	$C_{amb} (\mu g \cdot L^{-1})$
All sites	70	75	7.5	6	150	593	2.4	315	144	35	4.3	13
	(32–253)	(5–593)	(0.8–68)	(5–36)	(7–418)	(56–46,373)	(0.5–123)	(37–457)	(55–559)	(9–149)	(0.5–17)	(1–23)
BRO	44	251	24.3	8	63	8829	32.0	357	64	53	16.0	3
	(32–59)	(121–593)	(16–68)	(5–24)	(8–418)	(714–46,373)	(2–123)	(177–382)	(55–105)	(10–79)	(7–17)	(1–5)
ESP	90	112	9	8	249	1131	4.0	322	138	87	8.0	11
	(57–152)	(41–274)	(7–25)	(5–36)	(139–313)	(667–2038)	(3.6–5.6)	(183–407)	(42–154)	(60–149)	(7–12)	(8–13)
TBM	116	13	3	6	194	264	1.3	142	283	12	1.0	13
	(62–172)	(10–20)	(1.6–3)	(5–6.7)	(95–351)	(56–519)	(1-3)	(37–392)	(156–560)	(9–15)	(0.6–1)	(8–15)
CAN	193	8	0.9	10	132	313	1.0	316	201	24	1.0	21
	(35–253)	(76–96)	(0.7–6)	(5–12)	(131–373)	(155–328)	(0.5–1.6)	(202–457)	(77–391)	(10–36)	(0.5–1.5)	(18–23)

Water 2018, 10, 1080 7 of 17

3.2. Nutrient Uptake Metrics and Kinetics

Among streams, annual median uptake lengths ranged from 44–193 m for NH₄–N, 63–249 m for NO₃–N, and 64–283 m for SRP (Table 3). Annual median areal uptake rates ranged from 8–251 $\mu g \cdot m^{-2} \cdot min^{-1}$ among streams for NH₄–N, 264–8829 $\mu g \cdot m^{-2} \cdot min^{-1}$ for NO₃–N, and 12–87 $\mu g \cdot m^{-2} \cdot min^{-1}$ for SRP. Annual median uptake velocities ranged from 0.9–24.3, 1.0–32.0, and 1.0–16.0 mm·min⁻¹ for NH₄–N, NO₃–N, and SRP, respectively (Table 3).

Spearman rank correlations used for parameter selection are available as Supplementary Material (Table S1). For multiple regression models, we used U_{amb} and V_{f-amb} as the dependent variables rather than S_{w-amb} . Uptake length is sensitive to stream discharge, that is, hydrologic transport, and accordingly, we found only weak relationships between S_{w-amb} and other environmental variables. Therefore, we considered U_{amb} and V_{f-amb} to be more representative of biogeochemical nutrient demand. All multiple linear regression models were significant (p < 0.05, adjusted R^2 0.48 to 0.89; Table 4). In these multiple regressions, there was a negative influence of SRP concentrations on N uptake rates and velocity. NO₃–N concentration had a positive effect on NO₃–N uptake rate. Furthermore, SRP concentrations had a negative influence on SRP uptake velocity, while NH₄–N concentrations did not affect any nutrient uptake metrics. More forested streams appeared to be more NH₄–N retentive than more open-canopy ones, as we found a strong positive relationship between the percentage of canopy cover and NH₄–N uptake rate. DO concentrations had a negative effect on NH₄–N uptake, while DO, water temperature, and velocity had negative effects on SRP retention.

Table 4. Multiple linear regressions of ambient uptake metrics (U_{amb} and V_{f-amb}) as function of environmental variables. All data used were ln-transformed [ln(x + 1)].

Dependent Variable	Independent Variable	В	SE	<i>p</i> -Value	Adjusted R ²
	Intercept	2.20	3.50	0.54	
$U_{amb} NO_3-N (\mu g \cdot m^{-2} \cdot min^{-1})$	SRP _{amb}	-1.33	0.52	2×10^2	0.482
_	NO_{3amb}	1.14	0.57	3×10^2	
	Intercept	-12.25	13.15	0.37	
$U_{amb} NH_4-N (\mu g \cdot m^{-2} \cdot min^{-1})$	SRP_{amb}	-1.64	0.24	3×10^5	0.824
O _{amb} 1114–11 (μg·III ·IIIII)	DO	-4.68	1.52	1×10^2	0.024
	%CC	6.61	2.65	3×10^2	
	Intercept	24.28	4.20	1×10^4	
	DO	-4.60	1.13	2×10^3	
U_{amb} SRP ($\mu g \cdot m^{-2} \cdot min^{-1}$)	Q	0.47	0.21	4×10^2	0.816
	V	-3.08	0.99	1×10^2	
	T	-3.85	0.92	1×10^3	
	Intercept	5.31	0.97	2×10^4	
V _{f-amb} NO ₃ −N (mm·min ⁻¹)	SRP _{amb}	-1.12	0.36	1×10^2	0.508
	v	-5.31	2.20	3×10^2	
	Intercept	13.03	3.00	1×10^3	
V _{f-amb} NH ₄ −N (mm·min ⁻¹)	SRP _{amb}	-1.35	0.22	$8 imes 10^4$	0.731
	DO	-3.74	1.39	2×10^2	
	Intercept	21.34	3.10	3×10^{5}	
	SRP _{amb}	-1.05	0.12	2×10^6	
V _{f-amb} SRP (mm·min ⁻¹)		-3.71	0.84	1×10^3	0.889
,	T	-2.97	0.73	2×10^3	
	v	-2.77	0.79	4×10^3	

For all streams, there was a better fit of total dynamic uptake rate ($U_{tot\text{-}dyn}$) with total dynamic nutrient concentration ($C_{tot\text{-}dyn}$) than with total dynamic uptake velocity ($V_{f\text{-}tot\text{-}dyn}$) (Figure 2, Supplementary Tables S2–S4). For uptake rate (U) kinetics (Table 5), there was no significant fit to the M–M model, suggesting no saturation of uptake rates. In general, the relationship between U and nutrient concentration did not appear to follow a specific model across streams. For U, site BRO

Water 2018, 10, 1080 8 of 17

had a better fit to the ELS model for all nutrients (R^2 = 0.12, 0.47, and 0.79 for NO₃–N, NH₄–N, and SRP, respectively) than to M–M and FTO models. Site ESP showed a better fit to the ELS model for NO₃–N U (R^2 = 0.70) and SRP U (R^2 = 0.88), while NH₄–N U followed the FTO model (R^2 = 0.80). Site TBM followed the FTO model for NO₃–N U and SRP U (R^2 = 0.60 and R^2 = 0.83, respectively), while for NH₄–N U, the ELS model showed the best fit (R^2 = 0.68). The FTO model showed the best fit in site CAN for NH₄–N and SRP U (R^2 = 0.70 and R^2 = 0.85, respectively), while there was a better fit to the ELS model for NO₃–N U (R^2 = 0.73) in CAN.

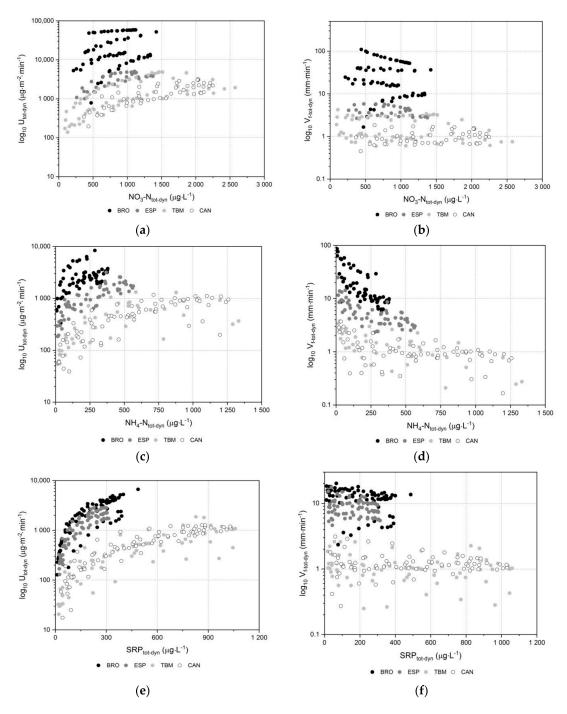


Figure 2. Relationships between $U_{tot\text{-}dyn}$, $V_{f\text{-}ot\text{-}dyn}$, and $C_{tot\text{-}dyn}$ for NO_3 –N, NH_4 –N, and SRP during the period from January to October 2017. (a) $U_{tot\text{-}dyn}$ versus $C_{tot\text{-}dyn}$ for NO_3 –N; (b) $V_{f\text{-}ot\text{-}dyn}$ versus $C_{tot\text{-}dyn}$ for NO_3 –N; (c) $U_{tot\text{-}dyn}$ versus $C_{tot\text{-}dyn}$ for NH_4 –N; (d) $V_{f\text{-}ot\text{-}dyn}$ versus $C_{tot\text{-}dyn}$ for NH_4 –N; (e) $U_{tot\text{-}dyn}$ versus $C_{tot\text{-}dyn}$ for SRP; (f) $V_{f\text{-}ot\text{-}dyn}$ versus $C_{tot\text{-}dyn}$ for SRP.

Water 2018, 10, 1080 9 of 17

Table 5. Kinetic model adjustments between $V_{\text{ft-tot-dyn}}$ versus $C_{\text{tot-dyn}}$ and $U_{\text{tot-dyn}}$ versus $C_{\text{tot-dyn}}$ for each stream considering all data collected from January to October 2017. In any case, R^2 is represented. ELS: efficiency-loss; FTO: first-order; M-M: Michaelis-Menten.

Stream	Best Kinetic Model (R2) for $V_{\text{ft-tot-dyn}}$ versus $N_{\text{tot-dyn}}$						
Stream	NO ₃ -N	NH ₄ -N	SRP				
BRO	-	M-M (0.653)	-				
ESP	FTO (0.649)	ELS (0.658)	-				
TBM	-	FTO (0.453)	-				
CAN	_	ELS (0.364)	-				
Stream	Best Kinetic Model (R ²) for U _{tot-dyn} versus N _{tot-dyn}						
Stream	NO ₃ -N	NH ₄ -N	SRP				
BRO	-	ELS (0.465)	ELS (0.787)				
ESP	ELS (0.697)	FTO (0.795)	ELS (0.881)				
TBM	FTO (0.596)	ELS (0.679)	FTO (0.831)				
CAN	ELS (0.733)	FTO (0.702)	FTO (0.854)				

 $-R^2 < 0.200$; $-R^2 < 0.200$; and slope > 1.

4. Discussion

Forested heterotrophic headwater streams, such as the streams we studied [58], typically have a considerable nutrient retention capacity (i.e., short S_{w-amb} and high U_{amb} and V_{f-amb} [10,14]). Several studies have also reported substantial NO₃-N, NH₄-N, and SRP retention in pristine tropical headwater streams [40,41,57,59]. The authors of [41] attributed high rates of NH₄–N uptake in tropical headwater streams to microbial activity, which is probably energy-limited because decomposition of terrestrial leaf litter is rapid, dissolved organic C is strongly retained by mineral soils, and light availability limits primary production in these streams. Further, the authors of [57] highlighted the importance of structural complexity, which increases residence time of solutes and available instream surface area, both important variables for aquatic microbial biofilms and thus for ammonium uptake in tropical Cerrado savanna streams. High nutrient uptake in tropical streams appears to be related to these processes rather than to water column nutrient concentrations, or just high temperature increasing microbial metabolism. In general, the tropical Cerrado woodland stream sites studied here corresponded well with this pattern; nutrient uptake lengths were short and uptake velocities and rates for all nutrient forms were high compared with literature data from temperate streams, especially for N forms. However, the metrics reported here were similar to those reported from other tropical streams of similar size (Table 6; representing a literature review of peer-reviewed articles on nutrient spiraling in tropical streams published prior to July 2018 and available at Web of Science and Science Direct). For a comparison with temperate systems, we used data from a literature review [14], which presented uptake metrics from 52 published studies, mostly of them carried out in temperate zones.

Table 6. Compilation of NO_3 –N, NH_4 –N, and SRP uptake results from natural streams sites located in different regions. The values are shown as range of metrics (min–max) when available; otherwise, the mean values are represented. Data from Ensign and Doyle [14] are uptake metrics from first-order streams.

Nutrient	Stream	Location	S _{w-amb} (m)	U_{amb} $(\mu g \cdot m^{-2} \cdot min^{-1})$	V _{f-amb} (mm·min ⁻¹)	Reference
	_	Temperate	101–478	5.8–19.1	0.8-4.2	[14]
	Barra Pequena		467–499	94	-	[19]
NO ₃ -N	Q Bisley –		1192 315–8480	16 -	1.92 0.13–0.94	[59] [40]
	BRO ESP TBM CAN	Tropical	6 (8–418) 249 (139–313) 194 (95–351) 132 (131–373)	8829 (714–46,373) 1131 (667–2038) 264 (56–519) 313 (155–328)	32 (2–123) 4 (3.6–5.6) 1.3 (1–3) 1 (0.5–1.6)	This study
	_	Temperate	23–275	5.3-52.6	2.5-17.0	[14]
NH ₄ -N	Barra Pequena Q. Bisley R. Cipó –	Tropical	138–501 15–26 –	5.45–14.2 33–42 1000	- 87.3 - 0.3–8.5	[19] [59] [13] [41]
	BRO ESP TBM CAN	порка	44 (32–59) 90 (57–152) 116 (62–172) 193 (35–253)	251 (121–593) 112 (41–274) 13 (10–20) 8 (76–96)	24.3 (16–68) 9 (7–25) 3 (1.6–3) 0.9 (0.7–6)	This study
	_	Temperate	24–161	3.4–15.7	1.5-6.6	[14]
SRP	Barra Pequena Carapa		461–1065 –	6.5–63 950–1150	_ _	[19] [60]
SKI	BRO ESP TBM CAN	Tropical	64 (55–105) 138 (42–154) 283 (156–560) 201 (77–391)	53 (10–79) 86 (60–149) 12 (9–15) 24 (10–36)	16 (7–17) 8 (7–12) 1 (0.6–1) 1 (0.5–1.5)	This study

Low SRP concentrations and predominantly high ambient molar DIN:SRP ratios (annual median of 24 to 279 across streams) would predict short uptake lengths and high uptake velocities for SRP compared with DIN forms. The authors of [61] suggested P limitation in streams at molar DIN:SRP ratios higher than 20, and consequently, a stoichiometric dominance of P uptake over N uptake. However, NH₄–N presented the shortest S_{w-amb} and the greatest V_{f-amb} in our study, indicating high retention and demand for NH₄–N. NO₃–N had the longest S_{w-amb} among all nutrient forms, indicating lowest retention relative to hydrologic transport of this nutrient form, and SRP had the lowest V_{f-amb} , suggesting lowest demand.

Above a molar Redfield ratio of N:P of 16:1 [62], P is expected to limit algal growth (N is in excess), and below this ratio there is an N deficit (P is in excess). The authors of [58] presented molar TN:TP ratios of 44 to 195 in the stream water for the same reaches we studied here, which would suggest P limitation. The lack of a relationship between SRP uptake metrics and DIN concentrations further supports P limitation as the increased availability of N should not facilitate SRP uptake if phosphorus is the primary limiting nutrient. Surprisingly, the slope of the relationship of N uptake rates and velocities, as well as $U_{\text{DIN}:}U_{\text{SRP}}$ ratios, with SRP concentrations was persistently negative in our study. We hypothesize that this negative relationship may have been caused by the rather special P geochemistry of the studied region (see discussion in subsequent paragraphs), that is, anaerobic sediment conditions stimulating P release, but at the same time negatively affecting aerobic assimilatory N uptake. In conclusion, the rather complex relationships between NO_3-N , NH_4-N , and SRP uptake in our streams appear to be due to the interplay of biotic P limitation, abiotic P cycling processes, and the preferential uptake of NH_4-N among N-forms.

While inorganic N uptake appeared to be driven mostly by biotic process in our study, because it was dominated by NO₃-uptake (i.e., higher areal NO₃–N than NH₄–N uptake rates), that does not present relevant abiotic uptake processes, SRP uptake might be related to abiotic processes. The relative importance of biotic and abiotic mechanisms for P retention can vary depending on P loading rates and in-stream conditions. At low levels of P loading, biotic uptake exceeds sorption [63], while under elevated P inputs, sorption can become a predominant mechanism depending on stream geochemistry [64]. Experimental whole-stream SRP additions and laboratory assays based on sediment sorption isotherms have shown that abiotic factors can dominate P retention in streams [60,65,66].

Abiotic P uptake occurs through sorption, which includes both adsorption to surfaces of cationic minerals and precipitation with electrolytes [67,68], and could be high in small streams, in which the sediment surface area to water volume ratio is high and contact time of stream water with sediments is long [69]. Fe and Al complexed with organic matter can be responsible for P sorption [68], however, we did not find relationships between SRP uptakes and dissolved organic C. In sediments dominated by Fe minerals, such as those present in Cerrado streams [70], reduction of soluble ferrous oxyhydroxide compounds results in the formation of P sorption sites. This reduction is the result of facultative organisms using ferric iron as an electron acceptor during their metabolic process in absence of oxygen. On the other hand, a reduced sediment surface layer can allow for considerable PO_4^{3-} release from Fe(III) oxide, whereas an oxidized sediment surface represents an efficient geochemical barrier for sediment P release [71].

Phosphorus uptake can be primarily governed by temperature, which can be attributed to biological mechanisms [72]. Therefore, if biotic uptake was relevant in our streams, the relationship with temperature should have been positive, because increases in thermal energy stimulate biotic P uptake [73]. However, we found a negative effect of temperature on P uptake, suggesting lower P uptake at high temperatures. We also found a negative slope in the correlations between SRP U_{amb} (B = -4.60), as well as V_{f-amb} (B = -3.71), and DO. The negative effects of both higher temperature and DO concentration on P uptake seem contradictory, as low DO saturation occurs at higher temperature. As both higher streamwater DO and temperature should stimulate aerobic biotic P uptake in the advective zone, these negative relationships may also point to the importance of sediment processes, that is, geochemical SRP sorption and SRP release processes [71]. In conclusion, our data suggested that abiotic P sorption may be a relevant process in our streams.

Among inorganic N-forms, the demand for NH_4-N was high, while the mass removal of NO_3-N was greater; NH_4-N V_{f-amb} was higher than NO_3-N V_{f-amb} , but NO_3-N U_{amb} was much greater than NH_4-N U_{amb} . The persistently high NH_4-N V_{f-amb} suggested preferential assimilation of NH_4-N by aquatic biota (bacteria, fungi, and algae) over NO_3-N , which is consistent with studies carried out in the literature [74–76]. In general, these authors observed that consumers track water column NH_4-N more closely than water column NO_3-N , as a result of the lower energy-demand associated with the assimilation of NH_4-N than with that of NO_3-N .

Despite the lower demand for NO₃–N relative to supply, as represented by its uptake velocities, the utilization of NO₃–N was considerable. Areal uptake rates (U) for NO₃–N were up to 80 times greater than U_{amb} for NH₄–N. This high U_{amb} showed that NO₃–N is an important source of N in these systems. Some studies [23,42] have related this condition to the generally greater background concentrations of NO₃–N than those of NH₄–N. For instance, the authors of [15] observed that NO₃–N concentrations 10- to 1000-fold greater than NH₄–N reduced NH₄–N retention to down to three times in streams in New Hampshire, USA. Similarly, the N demand of microorganisms was primarily satisfied by NO₃–N as a result of its greater availability, that is, 95% of the total DIN concentrations across our streams.

The relationships between stream nutrient uptake and its concentrations have received much attention in recent studies that also used pulse nutrient additions for kinetic analysis [16,27,47,77]. These authors highlighted that the amount of data provided by the TASCC method is useful for kinetic modelling. However, pulsed additions do not represent ambient uptake from a whole-stream

perspective, because they do not reflect stable-state transport conditions, but rather transient experimental conditions, with rising and falling limbs of breakthrough curves representing different transport processes [2].

We did not observe saturation conditions, that is, no significant fit to the M-M model was found across streams, which was expected because there was no evidence of chronical nutrient enrichment. In some cases, the lack of saturation suggests the existence of a mass transfer component, high-saturation sorption kinetics, or even the occurrence of dissimilatory processes such as nitrification and denitrification that may only saturate at very high concentrations [17]. Our study did not address the specific processes that dominate nutrient uptake, but demonstrated the dominance of biotic nitrate uptake and suggested the importance of abiotic SRP uptake. First-order responses to experimental nutrient enrichment or seasonal or spatial variation in nutrient concentration are commonly observed in streams with low to moderate nutrient concentrations [43], but efficiency loss and especially saturation are the typical responses of streams with chronic nutrient inputs. High streamwater nutrient concentrations due to agricultural and urban land use have been reported for tropical regions [37]. Across 35 highly urbanized tropical watersheds, NO₃-N and NH₄-N concentrations as high as 0.41 and $4.42 \text{ mg} \cdot \text{L}^{-1}$ have been reported [36]. Similar studies in other tropical streams with chronic nutrient loading would advance our understanding about the potential occurrence of nutrient saturation in tropical streams. Further, more nutrient uptake data is required to understand uptake as function of concentration across a wide variety of tropical streams and rivers, including assessments of abiotic versus biotic uptake, and limitation by mass transfer.

In our study, the uptake of different nutrients and nutrient forms appeared to be tightly associated. All nutrient uptake velocities were positively and strongly correlated with each other, which could suggest the occurrence of co-limitation in our streams [30]. The role of co-limitation in nutrient utilization by microorganisms has been widely investigated in recent studies [19,28,29,78], but had been rarely investigated in tropical stream ecology (but see [19]). As nutrient limitation is an important driver of nutrient uptake [79–81], future investigations should aim at more detailed assessments on how anthropogenic impacts on tropical streams are related to their nutrient limitation and co-limitation conditions. Finally, the relatively high uptake rates observed in the studied tropical Cerrado woodland streams highlight the importance of preservation of these headwater streams for the management of ecosystem functioning and services of Cerrado, and potentially other tropical catchments.

5. Conclusions

Here, we evaluated the variability in whole-stream uptake of three important nutrient forms, which is, SRP, NH₄–N, and NO₃–N, in relatively undisturbed, tropical woodland streams, and found relatively high and coupled uptake rates of these nutrients. Relationships between NO₃–N, NH₄–N, and SRP uptake in these streams seemed to be due to rather complex interactions of biotic P limitation, abiotic P cycling processes, and the preferential uptake of NH₄–N among N-forms. Drivers of nutrient uptake were partially nutrient form-specific and included unexpected effects, such as negative temperature dependencies of SRP uptake and negative relationships between the uptake of all nutrient forms and SRP concentration. Thus, global change effects on these tropical streams, such as temperature increases due to climate warming and decreased shading as a result of riparian clear-cutting, as well as nutrient enrichment due to urban and agricultural expansion, may have adverse and partially unpredictable impacts on whole-stream nutrient processing, and thus catchment biogeochemistry. Future studies should investigate the mechanisms of nutrient spiraling in tropical streams, such as abiotic versus biotic uptake processes, mass transfer limitation, and nutrient limitation and co-limitation, and how human activities influence them.

Supplementary Materials: The following are available online at http://www.mdpi.com/2073-4441/10/8/1080/s1, Table S1: Spearman's correlation coefficients between ambient uptake metrics (S_{w-amb} , S_{w-amb} , untrient concentrations and ratios, and in-stream physical and chemical variables. Significant correlations are highlighted in bold (p < 0.05), Table S2: Statistical parameters of Michaelis–Menten models used to evaluate the

relationship between total dynamic uptake rate ($U_{tot-dyn}$) or total dynamic uptake velocity ($V_{f-tot-dyn}$), and total dynamic nutrient concentration ($C_{tot-dyn}$) for both streams. For the Michaelis–Menten model, the maximum uptake rate (U_{max} ; $\mu g \cdot m^{-2} \cdot min^{-1}$) and the half saturation constant (K_m ; $\mu g \cdot L^{-1}$) are shown. The adjusted R^2 and p-value of fits are reported in brackets, Table S3: Statistical parameters of the efficiency-loss model ($U = aN^b$) used to evaluate the fit between the relationship between total dynamic uptake rate ($U_{tot-dyn}$), total dynamic uptake velocity ($V_{f-tot-dyn}$), and total dynamic nutrient concentration ($C_{tot-dyn}$) for both streams. For the efficiency-loss model, a represents the intercept and b is the exponent. The adjusted R^2 and b-value of the fits are also reported in brackets, Table S4: Statistical parameters of the first-order model (U = a + bN) used to evaluate the fit between the relationship between total dynamic uptake rate ($U_{tot-dyn}$), total dynamic uptake velocity ($V_{f-tot-dyn}$), and total dynamic nutrient concentration ($C_{tot-dyn}$) for both streams. For the efficiency-loss model, a represents the intercept and b is the slope. The adjusted R^2 and b-value of the fits are also reported in brackets.

Author Contributions: N.R.F and D.G.F.C. designed the framework of the study, conducted fieldwork and analyzed data. N.R.F. coordinated analysis of dissolved nutrients of the samples, and also the modelling of nutrient uptake metrics. I.G.B. and B.G. revised statistical analysis. All authors discussed the results and commented on the manuscript.

Funding: This study was supported by FAPESP (São Paulo Research Foundation, Grant 2016/14176-1) and CNPq Productivity Grant 300899/2016-5). NSF Macrosystems Grant 1442595 supported F.T.

Acknowledgments: N.R.F. thanks CNPq (National Council for Scientific and Technological Development) for scholarship. Special thanks to M.S. Ferreira (University of São Paulo, Brazil), who provided help with mapping and A. Schechner (Kansas State University, USA) for editing the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Newbold, J.D.; O'Neill, R.V.; Elwood, J.W.; Van Winkle, W. Nutrient Spiraling in Streams: Implications for Nutrient Limitation and Invertebrate Activity. *Am. Nat.* **1982**, *120*, 628–652. [CrossRef]
- 2. Stream Solute Workshop. Concepts and Methods for Assessing Solute Dynamics in Stream Ecosystems. *J. N. Am. Benthol. Soc.* **1990**, *9*, 95–119. [CrossRef]
- 3. Rabalais, N.N. Nitrogen in Aquatic Ecosystems. AMBIO J. Hum. Environ. 2002, 31, 102–112. [CrossRef]
- 4. Vanni, M.J.; Renwick, W.H.; Bowling, A.M.; Horgan, M.J.; Christian, A.D. Nutrient stoichiometry of linked catchment-lake systems along a gradient of land use. *Freshw. Biol.* **2011**, *56*, 791–811. [CrossRef]
- 5. Matson, P.A.; McDowell, W.H.; Townsend, A.R.; Vitousek, P.M. The globalization of N deposition: Ecosystem consequences in tropical environments. *Biogeochemistry* **1999**, *46*, 67–83. [CrossRef]
- 6. Bennett, E.M.; Carpenter, S.R.; Caraco, N.F. Human Impact on Erodable Phosphorus and Eutrophication: A Global Perspective. *AIBS Bull.* **2001**, *51*, 227–231. [CrossRef]
- 7. Galloway, J.N.; Townsend, A.R.; Erisman, J.W.; Bekunda, M.; Cai, Z.; Freney, J.R.; Martinelli, L.A.; Seitzinger, S.P.; Sutton, M.A. Transformation of the Nitrogen Cycle: Recent Trends, Questions, and Potential Solutions. *Science* 2008, 320, 889–892. [CrossRef] [PubMed]
- 8. Elser, J.J.; Bracken, M.E.S.; Cleland, E.E.; Gruner, D.S.; Harpole, W.S.; Hillebrand, H.; Ngai, J.T.; Seabloom, E.W.; Shurin, J.B.; Smith, J.E. Global analysis of nitrogen and phosphorus limitation of primary producers in Freshwater, marine and terrestrial ecosystems. *Ecol. Lett.* **2007**, *10*, 1135–1142. [CrossRef] [PubMed]
- 9. Hall, R.J.O.; Tank, J.L. Ecosystem metabolism controls nitrogen uptake in streams in Grand Teton National Park, Wyoming. *Limnol. Oceanogr.* **2003**, *48*, 1120–1128. [CrossRef]
- 10. Fellows, C.S.; Valett, H.M.; Dahm, C.N.; Mulholland, P.J.; Thomas, S.A. Coupling nutrient uptake and energy flow in headwater streams. *Ecosystems* **2006**, *9*, 788–804. [CrossRef]
- 11. Gücker, B.; Pusch, M.T. Regulation of nutrient uptake in eutrophic lowland streams. *Limnol. Oceanogr.* **2006**, *51*, 1443–1453. [CrossRef]
- 12. Valett, H.M.; Crenshaw, C.L.; Wagner, P.F.; Oct, N. Stream Nutrient Uptake, Forest Succession and Biogeochemical Theory. *Ecology* **2002**, *83*, 2888–2901. [CrossRef]
- 13. Gücker, B.; Boëchat, I. Stream morphology controls ammonium retention. *Ecology* **2004**, *85*, 2818–2827. [CrossRef]
- 14. Ensign, S.H.; Doyle, M.W. Nutrient spiraling in streams and river networks. *J. Geophys. Res. Biogeosci.* **2006**, 111, 1–13. [CrossRef]

Water 2018, 10, 1080 14 of 17

15. Bernhardt, E.S.; Hall, R.O.; Likens, G.E. Whole-system estimates of nitrification and nitrate uptake in streams of the Hubbard Brook Experimental Forest. *Ecosystems* **2002**, *5*, 419–430. [CrossRef]

- 16. Rodríguez-Cardona, B.; Wymore, A.S.; McDowell, W.H. DOC:NO₃⁻ ratios and NO₃⁻ uptake in forested headwater streams. *J. Geophys. Res. Biogeosci.* **2016**, *121*, 205–217. [CrossRef]
- 17. Dodds, W.K.; López, A.J.; Bowden, W.B.; Gregory, S.; Grimm, N.B.; Hamilton, S.K.; Hershey, A.E.; Martí, E.; McDowell, W.H.; Meyer, J.L.; et al. N uptake as a function of concentration in streams. *J. N. Am. Benthol. Soc.* **2002**, *21*, 206–220. [CrossRef]
- 18. Earl, S.R.; Valett, H.M.; Webster, J.R. Nitrogen saturation in stream ecosystems. *Ecology* **2006**, *87*, 3140–3151. [CrossRef]
- 19. Tromboni, F.; Thomas, S.A.; Gücker, B.; Neres-Lima, V.; Lourenço-Amorim, C.; Moulton, T.P.; Silva-Junior, E.F.; Feijó-Lima, R.; Boëchat, I.G.; Zandonà, E. Nutrient limitation and the stoichiometry of nutrient uptake in a tropical rainforest stream. *J. Geophys. Res. Biogeosci.* 2018. [CrossRef]
- 20. O'Brien, J.M.; Dodds, W.K.; Wilson, K.C.; Murdock, J.N.; Eichmiller, J. The saturation of N cycling in Central Plains streams: 15N experiments across a broad gradient of nitrate concentrations. *Biogeochemistry* **2007**, *84*, 31–49. [CrossRef]
- 21. Hall, R.O.; Bernhardt, E.S.; Likens, G.E. Relating Nutrient Uptake with Transient Storage in Forested Mountain Streams. *Limnol. Oceanogr.* **2008**, 47, 255–265. [CrossRef]
- 22. Martí, E.; Fonollà, P.; Von Schiller, D.; Sabater, F.; Argerich, A.; Ribot, M.; Riera, J.L. Variation in stream C, N and P uptake along an altitudinal gradient: A space-for-time analogue to assess potential impacts of climate change. *Hydrol. Res.* **2009**, *40*, 123. [CrossRef]
- 23. Ribot, M.; Von Schiller, D.; Peipoch, M.; Sabater, F.; Grimm, N.B.; Martí, E. Influence of nitrate and ammonium availability on uptake kinetics of stream biofilms. *Freshw. Sci.* **2013**, *32*, 1155–1167. [CrossRef]
- 24. Hoellein, T.J.; Tank, J.L.; Rosi-Marshall, E.J.; Entrekin, S.A.; Lamberti, G.A. Controls of nutrient variation on spatial and temporal variation of uptake in three Michigan headwater streams. *Limnology* **2007**, *52*, 1964–1977. [CrossRef]
- 25. Von Schiller, D.; Martí, E.; Riera, J.L.; Ribot, M.; Argerich, A.; Fonolla, P.; Sabater, F. Inter-annual, Annual, and Seasonal Variation of P and N Retention in a Perennial and an Intermittent Stream. *Ecosystems* **2008**, 11, 670–687. [CrossRef]
- 26. Bechtold, H.A.; Marcarelli, A.M.; Baxter, C.V.; Inouye, R.S. Effects of N, P, and organic carbon on stream biofilm nutrient limitation and uptake in a semi-arid watershed. *Limnol. Oceanogr.* **2012**, *57*, 1544–1554. [CrossRef]
- 27. Piper, L.R.; Cross, W.F.; McGlynn, B.L. Colimitation and the coupling of N and P uptake kinetics in oligotrophic mountain streams. *Biogeochemistry* **2017**, *132*, 165–184. [CrossRef]
- 28. Hill, B.H.; Mccormick, F.H.; Harvey, B.C.; Johnson, S.L.; Warren, M.L.; Elonen, C.M. Microbial enzyme activity, nutrient uptake and nutrient limitation in forested streams. *Freshw. Biol.* **2010**, *55*, 1005–1019. [CrossRef]
- 29. Schade, J.D.; Macneill, K.; Thomas, S.A.; Camille Mcneely, F.; Welter, J.R.; Hood, J.; Goodrich, M.; Power, M.E.; Finlay, J.C. The stoichiometry of nitrogen and phosphorus spiralling in heterotrophic and autotrophic streams. *Freshw. Biol.* **2011**, *56*, 424–436. [CrossRef]
- 30. Gibson, C.A.; Reilly, C.M.O.; Conine, A.L.; Lipshutz, S.M. Nutient uptake across a gradient of nutrient concentration and ratios at the landscape scale. *J. Geophys. Res. Biogeosci.* **2015**, *120*, 326–340. [CrossRef]
- 31. Wymore, A.S.; Coble, A.A.; Rodríguez-Cardona, B.; McDowell, W.H. Nitrate uptake across biomes and the influence of elemental stoichiometry: A new look at LINX II. *Glob. Biogeochem. Cycles* **2016**, *30*, 1183–1191. [CrossRef]
- 32. Schlesinger, W.H.; Cole, J.J.; Finzi, A.C.; Holland, E.A. Introduction to coupled biogeochemical cycles. *Front. Ecol. Environ.* **2011**, *9*, 5–8. [CrossRef]
- 33. Marklein, A.R.; Houlton, B.Z. Nitrogen inputs accelerate phosphorus cycling rates across a wide variety of terrestrial ecosystems. *New Phytol.* **2012**, *193*, 696–704. [CrossRef] [PubMed]
- 34. UNFPA. The State of World Population. 2017. Available online: https://www.unfpa.org/sites/default/files/sowp/downloads/UNFPA_PUB_2017_EN_SWOP.pdf (accessed on 1 August 2018).
- 35. MEA. Millennium Ecosystem Assessment. Ecosystems and Human Well Being: Synthesis; Island Press: Washington, DC, USA, 2005.

Water 2018, 10, 1080 15 of 17

36. Tromboni, F.; Dodds, W.K. Relationships between Land Use and Stream Nutrient Concentrations in a Highly Urbanized Tropical Region of Brazil: Thresholds and Riparian Zones. *Environ. Manag.* **2017**, *60*, 30–40. [CrossRef] [PubMed]

- 37. Gücker, B.; Silva, R.C.S.; Graeber, D.; Monteiro, J.A.F.; Brookshire, E.N.J.; Chaves, R.C.; Boëchat, I.G. Dissolved nutrient exports from natural and human-impacted Neotropical catchments. *Glob. Ecol. Biogeogr.* **2016**, 25, 378–390. [CrossRef]
- 38. Mulholland, P.J.; Tank, J.L.; Webster, J.R.; Bowden, W.B.; Dodds, W.K.; Gregory, S.V.; Grimm, N.B.; Hamilton, S.K.; Johnson, S.L.; Mart, E.; et al. Can uptake length in streams be determined by nutrient addition experiments? Results from an interbiome comparison study. *J. N. Am. Benthol. Soc.* **2002**, *21*, 544–560. [CrossRef]
- 39. Solomon, C.T.; Hotchkiss, E.R.; Moslemi, J.M.; Ulseth, A.J.; Stanley, E.H.; Hall, R.O., Jr.; Flecker, A.S. Sediment size and nutrients regulate denitrification in a tropical stream. *J. N. Am. Benthol. Soc.* **2009**, 29, 480–490. [CrossRef]
- 40. Potter, J.D.; McDowell, W.H.; Merriam, J.L.; Peterson, B.J.; Thomas, S.M. Denitrification and total nitrate uptake in streams of a tropical landscape. *Ecol. Appl.* **2010**, 20, 2104–2115. [CrossRef] [PubMed]
- 41. Koenig, L.E.; Song, C.; Wollheim, W.M.; Rüegg, J.; Mcdowell, W.H. Nitrification increases nitrogen export from a tropical river network. *Freshw. Sci.* **2017**, *36*, 698–712. [CrossRef]
- 42. Newbold, J.D.; Bott, T.L.; Kaplan, L.A.; Dow, C.L.; Jackson, J.K.; Aufdenkampe, A.K.; Martin, L.A.; Van Horn, D.J.; de Long, A.A. Uptake of nutrients and organic C in streams in New York City drinking-water-supply watersheds. *J. N. Am. Benthol. Soc.* **2006**, 25, 998–1017. [CrossRef]
- 43. Mulholland, P.J.; Helton, A.M.; Poole, G.C.; Hall, R.O.; Hamilton, S.K.; Peterson, B.J.; Tank, J.L.; Ashkenas, L.R.; Cooper, L.W.; Dahm, C.N.; et al. Stream denitrification across biomes and its response to anthropogenic nitrate loading. *Nature* 2008, 452, 202–205. [CrossRef] [PubMed]
- 44. Gibson, C.A.; O'Reilly, C.M. Organic matter stoichiometry influences nitrogen and phosphorus uptake in a headwater stream. *Freshw. Sci.* **2012**, *31*, 395–407. [CrossRef]
- 45. Bernot, M.J.; Tank, J.L.; Royer, T.V.; David, M.B. Nutrient uptake in streams draining agricultural catchments of the midwestern United States. *Freshw. Biol.* **2006**, *51*, 499–509. [CrossRef]
- 46. Niyogi, D.K.; Simon, K.S.; Townsend, C.R. Land use and stream ecosystem functioning: Nutrient uptake in streams that contrast in agricultural development. *Arch. Hydrobiol.* **2004**, *160*, 471–486. [CrossRef]
- 47. Covino, T.; McGlynn, B.; McNamara, R. Land use/land cover and scale influences on in-stream nitrogen uptake kinetics. *J. Geophys. Res. Biogeosci.* **2012**, *117*, 1–13. [CrossRef]
- 48. Fonseca, B.M.; De Mendonça-Galvão, L.; Padovesi-Fonseca, C.; De Abreu, L.M.; Fernandes, A.C.M. Nutrient baselines of Cerrado low-order streams: Comparing natural and impacted sites in Central Brazil. *Environ. Monit. Assess.* **2014**, *186*, 19–33. [CrossRef] [PubMed]
- 49. Covino, T.P.; McGlynn, B.L.; McNamara, R.A. Tracer Additions for Spiraling Curve Characterization (TASCC): Quantifying stream nutrient uptake kinetics from ambient to saturation. *Limnol. Oceanogr. Methods* **2010**, *8*, 484–498. [CrossRef]
- 50. Solórzano, L. Determination of ammonia in natural waters by phenol hypochlorite method. *Limnol. Oceanogr.* **1969**, *14*, 799–801. [CrossRef]
- 51. APHA. Standard Methods for the Examination of Water and Wastewater, 22nd ed.; American Water Works Assn: Washington, DC, USA, 2012.
- 52. Lemmon, P.E. A New Instrument for Measuring Forest Overstory Density. J. For. 1957, 55, 667–668.
- 53. Lemmon, P.E. A Spherical Densiometer for Estimating Forest Overstory Density. For. Sci. 1956, 2, 314–320.
- 54. Webster, J.R.; Valett, H.M. *Solute Dynamics. Methods in Stream Ecology;* Hauer, F.R., Lamberti, G., Eds.; Elsevier Inc.: New York, NY, USA, 2006; pp. 169–185, ISBN 9780123329080.
- 55. Alexander, R.B.; Smith, R.A.; Schwarz, G.E. Effect of stream channel size on the delivery of nitrogen to the Gulf of Mexico. *Nature* **2000**, 403, 758–762. [CrossRef] [PubMed]
- 56. Webster, J.R.; Mulholland, P.J.; Tank, J.L.; Valett, H.M.; Dodds, W.K.; Peterson, B.J.; Bowden, W.B.; Dahm, C.N.; Findlay, S.; Gregory, S.V. Others Factors affecting ammonium uptake in streams—an inter-biome perspective. *Freshw. Biol.* **2003**, *48*, 1329–1352. [CrossRef]
- 57. Rosa, R.D.S.; Aguiar, A.C.F.; Boëchat, I.G.; Gücker, B. Impacts of fish farm pollution on ecosystem structure and function of tropical headwater streams. *Environ. Pollut.* **2013**, 174, 204–213. [CrossRef] [PubMed]

Water 2018, 10, 1080 16 of 17

58. Saltarelli, W.A.; Dodds, W.K.; Tromboni, F.; Calijuri, M.C.; Neres-lima, V.; Jordão, C.E.; Cunha, D.G.F. Variation of stream metabolism along a tropical environmental gradient. *J. Limnol.* **2018**. [CrossRef]

- 59. Merriam, J.L.; McDowell, W.H.; Tank, J.L.; Wollheim, W.M.; Crenshaw, C.L.; Johnson, S.L. Characterizing nitrogen dynamics, retention and transport in a tropical rainforest stream using an in situ ¹⁵N addition. *Freshw. Biol.* **2002**, *47*, 143–160. [CrossRef]
- 60. Small, G.E.; Ardón, M.; Duff, J.H.; Jackman, A.P.; Ramírez, A.; Triska, F.J.; Pringle, C.M. Phosphorus retention in a lowland Neotropical stream following an eight-year enrichment experiment. *Freshw. Sci.* **2016**, *35*, 1–11. [CrossRef]
- 61. Schanz, F.; Juon, H. Two different methods of evaluating nutrient limitations of periphyton bioassays, using water from the River Rhine and eight of its tributaries. *Hydrobiologia* **1983**, *102*, 187–195. [CrossRef]
- 62. Dodds, W.; Whiles, M. Nitrogen, sulfur, phosphorus, and other nutrients. In *Freshwater Ecology*; Dodds, W.K., Whiles, M., Eds.; Elsevier Inc.: Amsterdam, The Netherlands, 2010; ISBN 978-0-12-374724-2.
- 63. Newbold, J.D.; Elwood, J.W.; O'Neill, R.V.; Sheldon, A.L. Phosphorus Dynamics in a Woodland Stream Ecosystem: A Study of Nutrient Spiralling. *Ecology* **1983**, *64*, 1249–1265. [CrossRef]
- 64. Meyer, J.L. The Role of Sediments and Bryophytes in Phosphorus Dynamics in a Head Water Stream Ecosystem. *Limnol. Oceanogr.* **1979**, 24, 365–375. [CrossRef]
- 65. Demars, B.O.L. Whole-stream phosphorus cycling: Testing methods to assess the effect of saturation of sorption capacity on nutrient uptake length measurements. *Water Res.* **2008**, 42, 2507–2516. [CrossRef] [PubMed]
- 66. Powers, S.M.; Stanley, E.H.; Lottig, N.R. Quantifying phosphorus uptake using pulse and steady-state approaches in streams. *Limnol. Oceanogr. Methods* **2009**, *7*, 498–508. [CrossRef]
- 67. House, W.A.; Warwick, M.S. Interactions of phosphorus with sediments in the River Swale, Yorkshire, UK. *Hydrol. Process.* **1999**, *13*, 1103–1115. [CrossRef]
- 68. Reddy, K.R.; Kadlec, R.H.; Flaig, E.; Gale, P.M. Phosphorus retention in streams and wetlands: A review. *Crit. Rev. Environ. Sci. Technol.* **1999**, 29, 83–146. [CrossRef]
- 69. House, W.H.; Denison, F.H.; Armitage, P. Comparison of the uptake of inorganic P to a suspended and stream bed-sediment. *Water Res.* **1995**, *29*, 767–779. [CrossRef]
- 70. Silva, J.S.O.; da Bustamante, M.M.C.; Markewitz, D.; Krusche, A.V.; Ferreira, L.G. Effects of land cover on chemical characteristics of streams in the Cerrado region of Brazil. *Biogeochemistry* **2011**, *105*, 75–88. [CrossRef]
- 71. Mortimer, C.H. Chemical exchanges between sediments and water in the Great Lakes—Speculations on probable regulatory mechanisms. *Limnol. Oceanogr.* **1971**, *16*, 387–404. [CrossRef]
- 72. Angelo, A.D.J.D.; Webster, J.R.; Benfield, E.F. Mechanisms of Stream Phosphorus Retention: An Experimental Study. J. N. Am. Benthol. Soc. 1991, 10, 225–237. [CrossRef]
- 73. Valett, H.M.; Thomas, S.A.; Mulholland, P.J.; Webster, J.R.; Dahm, C.N.; Fellows, C.S.; Crenshaw, C.L.; Peterson, C.G. Endogenous and exogenous control of ecosystem function: N cycling in headwater streams. *Ecology* **2008**, *89*, 3515–3527. [CrossRef] [PubMed]
- 74. Dodds, W.K.; Priscu, J.C.; Ellis, B.K. Seasonal uptake and regeneration of inorganic nitrogen and phosphorus in a large oligotrophic lake: Size-fractionation and antibiotic treatment. *J. Plankton Res.* **1991**, *13*, 1339–1358. [CrossRef]
- 75. Pastor, A.; Peipoch, M.; Cañas, L.; Chappuis, E.; Ribot, M.; Gacia, E.; Riera, J.L.; Martí, E.; Sabater, F. Nitrogen stable isotopes in primary uptake compartments across streams differing in nutrient availability. *Environ. Sci. Technol.* **2013**, 47, 10155–10162. [CrossRef] [PubMed]
- 76. Oviedo-Vargas, D.; Royer, T.V.; Johnson, L.T. Dissolved organic carbon manipulation reveals coupled cycling of carbon, nitrogen, and phosphorus in a nitrogen-rich stream. *Limnol. Oceanogr.* **2013**, *58*, 1196–1206. [CrossRef]
- 77. Diemer, L.A.; McDowell, W.H.; Wymore, A.S.; Prokushkin, A.S. Nutrient uptake along a fire gradient in boreal streams of Central Siberia. *Freshw. Sci.* **2015**, *34*, 1443–1456. [CrossRef]
- 78. Appling, A.P.; Heffernan, J.B. Nutrient Limitation and Physiology Mediate the Fine-Scale (De)coupling of Biogeochemical Cycles. *Am. Nat.* **2014**, *184*, 384–406. [CrossRef] [PubMed]
- 79. Tank, J.L.; Reisinger, A.J. Nutrient Limitation and Uptake Rates in Streams and Rivers of the Greater Yellowstone Ecosystem. *Univ. Wyoming Natl. Park Serv. Res. Cent. Annu. Rep.* **2013**, *36*, 153–159.

80. King, S.A.; Heffernan, J.B.; Cohen, M.J. Nutrient flux, uptake, and autotrophic limitation in streams and rivers. *Freshw. Sci.* **2014**, *33*, 85–98. [CrossRef]

81. Jarvie, H.P.; Smith, D.R.; Norton, L.R.; Edwards, F.K.; Bowes, M.J.; King, S.M.; Scarlett, P.; Davies, S.; Dils, R.M.; Bachiller-Jareno, N. Phosphorus and nitrogen limitation and impairment of headwater streams relative to rivers in Great Britain: A national perspective on eutrophication. *Sci. Total Environ.* **2018**, *621*, 849–862. [CrossRef] [PubMed]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).