



# Article Exploring Adiponectin in Autosomal Dominant Kidney Disease: Insight and Implications

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Abstract: Autosomal Dominant Polycystic Kidney Disease (ADPKD) is a common monogenic disorder characterized by renal cysts and progressive renal failure. In kidney diseases, adipose tissue undergoes functional changes that have been associated with increased inflammation and insulin resistance mediated by release of adipokines. Adiponectin is involved in various cellular processes, such as energy and inflammatory and oxidative processes. However, it remains to be determined whether adiponectin is involved in the concomitant metabolic dysfunctions present in PKD. In this scenario, we aimed to analyze: (a) PPARy, ADIPOQ, ADIPOR1 and ADIPOR2 gene variations in 92 ADPKD patients through PCR-Sanger sequencing; and (b) adiponectin levels and its oligomerization state by ELISA and Western Blot. Our results indicated that: (a) 14 patients carried the PPARy SNP, 29 patients carried the ADIPOQ SNP rs1501299, and 25 patients carried the analyzed ADIPOR1 SNPs. Finally, 82 patients carried ADIPOR2 SNPs; and (b) Adiponectin is statistically lower in ADPKD patients compared to controls, and further statistically lower in ESRD than in non-ESRD patients. An inverse relationship between adiponectin and albumin and between adiponectin and creatinine and a direct relationship between adiponectin and eGFR were found. Interestingly, significantly lower levels of adiponectin were found in patients bearing the ADIPOQ rs1501299 SNP and associated with low levels of eGFR. In conclusion, adiponectin levels and the presence of ADIPOQ rs1501299 genotype are significantly associated with a worse ADPKD phenotype, indicating that both could potentially provide important insights into the disease. Further studies are warranted to understand the pathophysiological role of adiponectin in ADPKD patients.

Keywords: adiponectin; ADPKD1-2; SNPs; ADIPOQ; PPARy

## 1. Introduction

Autosomal Dominant Polycystic Kidney Disease (ADPKD) is a common monogenic disorder with an increasing prevalence in the last decade [1,2]. The patients affected by ADPKD are characterized by progressive formation of cysts in the kidneys with progressive renal failure; in addition, more than half suffer from an increased predisposition to atherosclerosis, chronic inflammation, and show a higher risk of cardiovascular disease [3]. Although a fully reliable disease marker is still missing, several biochemical markers that can be assayed in patients' blood and urine such as albumin, creatinine and total proteins [4].



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**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). In the majority of patients, the cause of disease is a mutation in the PKD1 ( $\pm 85\%$ ) or PKD2 ( $\pm 15\%$ ) genes that encode the polycystin-1 or polycystin-2 proteins, respectively [5,6].

The molecular mechanisms involved are not yet well understood, but obesity represents a condition that contribute to the risk of developing chronic renal failure, even in the absence of any other metabolic dysregulation [7]. The metabolic changes that adipose tissue undergoes in kidney disease are associated with increased inflammation and insulin resistance mediated by adipokine release, activation of the renin-angiotensin aldosterone system (RAAS), and oxidative stress [8].

Among adipokines, adiponectin is a cytokine of 30 kDa, accounting for approximately 0.01% of total serum proteins [9]. Adiponectin assembles to form three major oligomeric forms: the trimers (LMW), the hexamers (MMW), and the high-molecularweight (HMW) [10]. Initially studied for its metabolic functions (insulin sensitization and enhancement of fatty acid oxidation), adiponectin has attracted attention for additional functions such as the inhibition of inflammatory and oxidative processes [11]. Consequently, serum adiponectin levels have been found modulated in both metabolic (obesity and related disorders) and immune and inflammatory disorders [12–14].

In obesity, reduced adiponectin levels have been associated with insulin resistance, cardiovascular diseases and obesity related kidney diseases. On the other hand, patients with chronic kidney disease have dysregulated levels of adiponectin which correlate with disease progression [15–18]. More importantly, the adiponectin involvement in renal alterations has been demonstrated independently of the presence of obesity, suggesting a role for this adipokine in kidney patho-physiology [19]. Furthermore, adiponectin is inversely related to renal function, and represents a predictor of end-stage renal disease with a strong association with disease severity and mortality [20]. Moreover, serum adiponectin levels are elevated in patients with albuminuria and are positively correlated with urine albumin/creatinine ratio [18].

Adiponectin is encoded by the ADIPOQ gene located on chromosome 3q27 [21]. Genetic variations in the ADIPOQ gene have been found associated with adiponectin expression in several studies and in various pathologies [22–24]. Currently, only one study has examined the distribution of the ADIPOQ rs1501299, which significantly differs between CKD and controls [25]. Adiponectin exerts its biological effects mainly by ADIPOR1 and ADIPOR2 receptors, which are expressed in the kidneys and whose activation could prevent or improve diabetic nephropathy [26]. Furthermore, both ADIPORs have been linked with a high risk of CVD. In addition, Peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ), which regulates transcription of ADIPOQ gene, is associated with increased adiponectin levels and a reduced risk of type 2 diabetes [27].

In this scenario, our study was aimed to investigate the ADIPOQ, ADIPOR1 and ADI-POR2, PPAR $\gamma$  genetic variations in 92 ADPKD patients. In addition, the adiponectin serum levels were tested and compared to a sex-, BMI- and age-matched group of 104 healthy controls. Finally, the oligomerization state of adiponectin was investigated.

To our knowledge, while the amount literature about the role of adiponectin in several renal disorders is increasing, no data are available about the regulation of adiponectin levels in PKD patients. Finally, the most relevant SNPs in the genes encoding for ADIPOQ, Adiponectin receptors 1, 2 (ADIPOR1, ADIPOR2) and PPARγ were investigated.

#### 2. Materials and Methods

#### 2.1. Patients Selection

This is a secondary analysis of the previous published paper [28]; here, 92 patients (mean age  $42.84 \pm 14.7$ ) diagnosed with ADPKD were recruited from Nephrology Unit, Department of Public Health, "Federico II" University, Naples, and Hospital outpatient clinic, "Federico II", Naples. The study inclusion criteria and the genetic characterization of ADPKD patients has been previously performed [28]. Briefly, the inclusion criteria were either with a positive family history of PKD and met the unified criteria for ultrasonographic diagnosis of PKD; or were without PKD family history, but had confirmed renal cysts

through imaging as per the unified criteria for ultrasonographic diagnosis of PKD [28]. Supplementary Table S1 shows the pathogenic variants found in the selected population. Blood samples were collected after a 12 h overnight fasting period and centrifuged to collect serum. Serum aliquots were immediately frozen in liquid nitrogen and stored at -80 °C. The following clinical and biochemical values were recorded (see Table 1): BMI, total cholesterol, triglycerides, glycemia, AST, ALT, WBC, neutrophils %, lymphocytes%, NLR, RBC, HGB, EGFR.

Parameter		$\begin{array}{l} \text{ADPKD } n = 92 \\ \text{Mean (SD)} \end{array}$	Controls $n = 104$ Mean (SD)	<i>p</i> -Value
Age (years)		42.84 (14.7)	45.96 (16.6)	0.169
Sex	F M	49 43	48 56	0.86
BMI $(kg/m^2)$		25.81 (4.8)	24.66 (2.5)	0.065
Systolic pressure (mmHg)		132.89 (16.7)	123.66 (3.35)	0.006
Diastolic pressure (mmHg)		82.35 (10.3)	83.66 (5.23)	0.50
Total cholesterol (mg/dL)		178.48 (35.9)	185.84 (33.1)	0.06
Triglycerides (mg/dL)		101.34 (67.3)	94.06 (50.51)	0.39
Glycemia (mg/dL)		81.47 (17.3)	83.96 (15.0)	0.29
AST (U/L)		20.66 (6.4)	20.36 (6.5)	0.74
ALT (U/L)		20.04 (11.4)	22.15 (15.9)	0.08
WBC ( $\times 10^3 / \mu L$ )		6.91 (2.0)	6.57 (1.5)	0.30
Neutrophils %		60.20 (10.2)	56.91 (9.6)	0.02
Lymphocytes %,		28.15 (7.8)	32.63 (8.2)	0.00
NLR		2.46 (1.3)	1.95 (0.9)	0.004
RBC ( $\times 10^6/\mu$ L)		4.66 (0.7)	4.92 (0.9)	0.06
HGB (g/dL)		15.27 (1.3)	14.09 (1.3)	0.40
Albumin (mg/dL)		4.55 (0.3)	4.70 (0.3)	0.01
Creatinine $(mg/dL)$		1.29 (1.1)	0.91 (0.2)	0.001
Uric Acid (mg/dL)		5.45 (1.6)	4.79 (1.2)	0.005
Urea (mg/dL)		49.89 (30.2)	36.20 (9.5)	0.000
EGFR (mL/min/1.73 m <sup>2</sup> )		78.99 (33.3)	90 (6.7)	

Table 1. Comparison of the biochemical and clinical features of the study participants.

A total of 104 healthy volunteers, aged  $45.91 \pm 16.6$  years, constituted the control group. The research protocol was conducted in accordance with the principles of the Helsinki II Declaration; according to the current legislation in Italy, informed consent was obtained from each subject. The protocol of this study was approved by the Federico II Ethical Committee (protocol code 2018-000477-77, date of approval: 16 November 2022).

## 2.2. ELISA

Total serum adiponectin concentration was measured by enzyme-linked immunosorbent assay (ELISA) utilizing a polyclonal antibody, in house produced, versus a human adiponectin sequence region (H2N-ETTTQGPGVLLPLPKG-COOH), as previously reported [29]. Each sample was tested three times in duplicate.

## 2.3. Western Blotting

Serum samples were quantified for total proteins by Bradford's method (Bio-Rad, Hercules, CA, USA); 5  $\mu$ g of total proteins were treated with 1× Laemmli buffer, heated at 95 °C for 5 min and loaded under non-reducing conditions on 10%SDS-PAGE gel and transferred as previously described [30]. The blots were scanned by using ChemiDoc MP imaging system (Bio-Rad, Hercules, CA, USA) and analyzed by densitometry with ImageJ software (version 1.53) (http://rsbweb.nih.gov.ij/), (accessed on 1 October 2023). A representative sample of 30 patients and 30 healthy controls was analyzed and tested two times in duplicate.

#### 2.4. SNPs Analysis by Sanger Sequencing

Genomic DNA was amplified by singleplex PCR using the FailSafe PCR System (Epicentre, Madison, WI, USA). The following SNPs have been analyzed in the entire population of ADPKD patients through Sanger sequencing: ADIPOQ c.45T>C; ADIPOQ c.268G>A; ADIPOQ c.214+62G>T PPAR $\gamma$  c.34C>G; AdipoR1 c.94-12A>G; AdipoR1 c. 94-8T>G; AdipoR2 c.650+20G>A; AdipoR2 c.\*1642C>T; AdipoR2 c.\*1718C>T; AdipoR2 c.795G>A; AdipoR2 171+48A>T; (Supplementary Table S2). Universally tagged sequencing primers were designed using the following software: Primer3 version 1.1.4 (http://www.sourceforge.net; accessed on 1 October 2023). Primers are available on request. Thermal cycling was performed with 15 cycles [30 s at 98 °C; 30 s at 62 °C (-0.5 °C each cycle); 60 s at 72 °C], followed by 15 cycles (30 s at 98 °C; 30 s at 55 °C; 60 s at 72 °C).

## 2.5. Statistical Analysis

The data were analyzed with SPSS software. (Version 27.0, IBM Corp, Armonk, NY, USA). Results are presented as mean  $\pm$  standard deviation (SD), and *p* < 0.05 was considered statistically significant. To compare groups (ADPKD vs. Control), we used unpaired *t*-tests. Furthermore, the adiponectin levels were adjusted for weight, age, BMI, systolic and diastolic pressure, and then compared between groups (ADPKD and Controls). Pearson correlation (r) was used to evaluate the linear correlation between different variables.

## 3. Results

#### 3.1. Biochemical and Clinical Parameters of ADPKD Patients Compared to Controls

The baseline characteristics of the study participants are shown in Table 1. There were no statistically significant differences in age, sex, BMI, diastolic pressure, glycemia, total cholesterol, triglycerides between the two groups. Neutrophils, NLR and systolic pressure values were statistically higher in ADPKD patients compared to controls (p value < 0.001), indicating an inflammatory state in patients. Albumin and lymphocytes values were lower in patients than in controls (p value = 0.01). As expected, creatinine, uric acid, and urea were higher in patients compared to controls (p value < 0.001).

## 3.2. Adiponectin Evaluation and Oligomerization State

Adiponectin levels were statistically lower in ADPKD patients compared to healthy controls (14.13  $\pm$  4.2 vs. 16.73  $\pm$  3.8 µg/mL; *p* value < 0.0001) (Figure 1A). Furthermore, adiponectin levels were adjusted for weight, age, BMI, systolic and diastolic pressure and then compared between the groups (ADPKD and controls); adiponectin levels were still lower in ADPKD patients than in controls (Table 2).



**Figure 1.** Total serum adiponectin is decreased in ADPKD patients compared to controls. (A) Adiponectin levels were statistically reduced in patients with ADPKD compared to controls  $(14.13 \pm 4.2 \text{ vs. } 16.73 \pm 3.8 \text{ } \mu\text{g/mL}; p \text{ value } < 0.0001)$  (B) Adiponectin levels were statistically lower in a subgroup of ADPKD patients at ESRD vs. age-matched ADPKD patients not in ESRD  $(13.74 \pm 4.40 \text{ vs. } 15.65 \pm 3.58 \text{ } \mu\text{g/mL}; p \text{ value } = 0.02)$ . \* p < 0.05. \*\*\*\* p < 0.0001.

A directed		Adiponectin						
Aujusteu			Controls		ADPKD			- P
		Mean		ES	Mean	±	ES	
Age	years	16.7	±	0.402	14.1	±	0.425	0.00001
Body Weight	kg	16.6	$\pm$	0.414	14.3	±	0.431	0.0003
BMI	kg/m <sup>2</sup>	16.7	±	0.404	14.2	±	0.428	0.00004
Systolic pressure	mmHG	17.0	±	0.717	14.3	±	0.445	0.002
Diastolic Pressure	mmHG	17.1	$\pm$	0.700	14.3	±	0.441	0.0009

**Table 2.** Adiponectin comparison between ADPKD patients and controls after adjustment for age,body weight, BMI, systolic pressure, and diastolic pressure.

Interestingly, adiponectin was further statistically lower in ESRD than in non-ESRD patients (11.62  $\pm$  2.5 vs. 14.54  $\pm$  4.4  $\mu$ g/mL; *p* value = 0.04) (Figure 1B).

Additionally, we analyzed the oligomeric distribution of adiponectin in ADPKD patients compared to healthy controls by Western blotting (Figure 2). Three bands corresponding to HMW ( $\geq$ 250 kDa), MMW ( $\sim$ 180 kDa), and LMW ( $\sim$ 70 kDa) oligomers were evident for both controls and ADPKD patients (Figure 2A). As shown in Figure 2B, the densitometric analysis of oligomeric distribution showed that ADPKD patients had a lower expression of HMW oligomers compared to controls.



**Figure 2.** Oligomeric distribution of adiponectin in ADPKD patients and healthy controls. (**A**) Representative Western blot image for adiponectin oligomers, HMW, MMW, and LMW in the serum of controls and patients diagnosed with ADPKD. (**B**) Graphical representation of pixel quantization of analyzed controls and ADPKD patients. \*\*\*\* p < 0.0001.

Subsequently, within patients, we analyzed adiponectin correlations with several clinical parameters to evaluate its relationship with disease prognosis. We found an inverse relationship between adiponectin and albumin (p = 0.014) (Figure 3A) and an inverse relationship between adiponectin and creatinine (p = 0.021) (Figure 3B). A direct relationship between adiponectin and eGFR was found (p = 0.04) (Figure 3C). Altogether, these correlations suggest that adiponectin is inversely related to disease severity and poor prognosis.

Successively, we divided patients into three groups, defining three classes of severity, according to the presence of the following parameters: BMI > 25, creatinine > 1.1 mg/dL, systolic pressure > 120, diastolic pressure > 80, urea > 43 mg/dL, uric acid > 6 mg/dL, NLR > 3, eGFR < 90. According to each parameter, the three groups were divided as follows: 1 mild (0–1 point), 2 moderate (2–4 points), 3 severe (>4 points). Adiponectin distribution within the three groups supported the evidence that it correlates with disease seriousness: indeed, group 1 had higher levels of adiponectin (14.88  $\pm$  5.1) compared to group 2 (13.60  $\pm$  3.2) and group 3 (13.94  $\pm$  4.2).



**Figure 3.** Correlation analysis between adiponectin and clinical parameters in ADPKD patients and controls. Dashed lines indicate control subjects while dark lines refer to patients. Adiponectin inversely correlated with albumin (**A**), and creatinine (**B**) in both ADPKD patients and controls. Adiponectin positively correlated with eGFR (**C**) in ADPKD patients.

## 3.3. SNPs Analysis in PPAR $\gamma$ , ADIPOQ, ADIPOR1 and ADIPOR2 Genes

The SNP analysis, performed by PCR and Sanger sequencing, revealed 14 patients bearing the PPAR $\gamma$  rs1801282 SNP (Table 3). Regarding ADIPOQ, we found in 26, 4 and 29 patients with the following SNPs: rs2241766, rs62625753, rs1501299. Further, 23 and 25 patients bore the analyzed ADIPOR1 SNPs, rs2275737 and rs2275738, respectively. Finally, 18, 23, and 16 patients carried the following ADIPOR2 SNPs: rs16928751, rs1044471, rs12342 (see Table 3).

**Table 3.** PPARγ, ADIPOQ, ADIPOR1, ADIPOR2 gene polymorphisms distribution in ADPKD population.

Gene	SNP (rs Number)	n Patients	Het/Homo	Age	Adiponectin (µg/mL) (st.dev)	eGFR (mL/min/1.73 mq) (st.dev)	Systolic Blood Pressure (mm/Hg) (st.dev)	Diastolic Blood Pressure (mm/Hg) (st.dev)
PPARγ	c.34 C>G (rs1801282)	14	14/0	46 (15.35)	15.92 (3.27)	65.21 (36.87)	133.30 (22.65)	83.52 (18.18)
ADIPOQ	c.45C>T (rs2241766)	26	20/6	43 (14.84)	14.11 (4.39)	66.18 (31.85)	136.09 18.17)	82.97 (9.49)
	c.268G>A (rs62625753)	4	4/0	58 (18.6)	14.25 (5.00)	50.25 (37.73)	136.67 (5.77)	88.33 (7.63)
	c.214+62G>T (rs1501299)	29	24/5	43.7(16.28)	14.5 (4.42)	78.3 (35.15)	134.0 (18.11)	84.4 (10.21)
ADIPOR1	c. 94-8T>G (rs2275737)	23	17/6	44 (12.92)	13.25 (3.86)	64.34 (31.36)	134.14 (13.36)	82.14 (8.16)
	c. 94-12>G (rs2275738)	25	18/7	44 (12.92)	13.25 (3.86)	64.34 (31.36)	134.14 (13.36)	82.14 (8.16)

Gene	SNP (rs Number)	<i>n</i> Patients	Het/Homo	Age	Adiponectin (µg/mL) (st.dev)	eGFR (mL/min/1.73 mq) (st.dev)	Systolic Blood Pressure (mm/Hg) (st.dev)	Diastolic Blood Pressure (mm/Hg) (st.dev)
	c.795G>A (rs16928751)	18	16/2	44 (13.60)	14.11 (3.97)	67.80 (38.35)	133.53 (13.89)	84.12 (10.64)
ADIPOR2	c.1718C>T (rs1044471)	23	17/6	41 (12.65)	12.98 (3.56)	68.99 (33.83)	132.32 (13.50)	80.89 (8.39)
	c.1642C>T (rs12342)	16	13/3	45 (14.47)	14.57 (4.39)	68.86 (34.50)	134.25 (15.83)	82.97 (9.53)

Table 3. Cont.

When we analyzed adiponectin levels and the main clinical parameters in those patients with SNPs, we found that patients bearing the PPAR $\gamma$  rs1801282 SNP had significantly higher adiponectin levels than the non-mutated patients (15.92 ± 3.27 vs. 13.81 ± 3.76 µg/mL; *p* value = 0.04) (Figure 4A). Interestingly, when we considered ADIPOQ mutated patients, no significant modulation of adiponectin levels was found in the SNPs patients groups.



**Figure 4.** Serum total adiponectin is expressed differently among ADPKD patients with SNPs in PPAR $\gamma$  and ADIPOQ genes. (**A**) Patients bearing the PPAR $\gamma$  rs1801282 SNP had higher adiponectin levels compared to the non-mutated ones (15.92 ± 3.27 vs. 13.81 ± 3.76 µg/mL; *p* value = 0.04). (**B**) The oligomeric state of adiponectin was also analyzed. (**C**) Patients bearing the PPAR $\gamma$  rs1801282 SNP showed an oligomeric profile comparable to that of control subjects and non-mutated patients. \* *p* < 0.05. \*\*\*\* *p* < 0.0001.

Subsequently, we analyzed the oligomeric profile of those patients bearing the PPAR $\gamma$  rs1801282 SNP with healthy controls (Figure 4B,C). As shown in panel B, the oligomeric profile was comparable among the groups, with evidence for a modulation of adiponectin total levels in the three groups (both PPAR $\gamma$  and ADIPOQ had lower levels than controls while PPAR $\gamma$  higher than ADIPOQ).

## 4. Discussion

In this study, conducted comparing an ADPKD Italian cohort to a healthy volunteer group, we analyzed adiponectin levels and its oligomerization state in relation to biochemical, genetic, and clinical parameters.

To date, numerous studies have been published providing details on the association between adiponectin and kidney function [15–18,31]. In detail, serum adiponectin concentrations are higher in patients with renal insufficiency [32]. Much of the previous work on the association between adiponectin and CKD has produced conflicting results, and has originated primarily from studies with small sample sizes and/or unadjusted comparisons between CKD patients and those without CKD. In addition, most of them considered elderly subjects and/or CKD patients, where significantly increased levels of serum adiponectin were found [15–18]. Furthermore, some studies have proposed that elevated serum adiponectin levels may be considered a prognostic marker in the progression of CKD due to the negative correlation with glomerular filtration [17]. In addition, adiponectin is significantly positively associated with severity of CKD measured by eGFR and urinary albumin [33]. However, to our knowledge, there are no studies that have considered the regulation of adiponectin levels in ADPKD patients.

Here, we found that, contrary to what is reported in CKD, adiponectin is statistically lower in ADPKD patients compared to healthy controls. Importantly, adiponectin levels are directly correlated with EGFR in our cohort of ADPKD patients and are further lower in ESRD patients compared to non-ESRD patients, indicating a relationship between adiponectin and renal health and an inverse relation with ADPKD severity. The literature data that report adiponectin up-regulation in renal diseases, but no data are available for a difference in ADPKD. Functionally, adiponectin role in kidney disorders have been related to multiple factors, i.e., the regulation of inflammatory processes in nephrons, involvement in vascular calcifications, and in the control of the nutritional status [34]. This study fails to clarify the functional role of adiponectin in ADPKD, but the data obtained, such as the different modulation of adiponectin in ESRD patients, and the direct correlation with eGFR, together with the inverse relationship with creatinine and albumin, suggest that the expression of this adipokine may be related to renal functioning. It is noticeable that, to our knowledge, there is no evidence of a correlation between adiponectin and serum albumin, but serum adiponectin levels are reported elevated in patients with albuminuria and positively correlated with the urine albumin/creatinine ratio [18].

On the other hand, in type I diabetic and diabetic nephropathy patients, it has been demonstrated that adiponectin predicts all-cause mortality and ESRD [17].

From this perspective, adiponectin may represent a central messenger in establishing a cross-talk between adipose tissue and kidney; furthermore, based on our data and those present in the literature, it could be hypothesized that adiponectin acts as an antiinflammatory cytokine and, therefore, participates in the progression of the disease.

Several genetic polymorphisms have been described to have a role in the regulation of adiponectin levels [35]; consequently, we analyzed the most relevant SNPs in ADIPOQ and PPAR $\gamma$  genes, finding 14 patients bearing the PPAR $\gamma$  rs1801282 SNP and 29 patients bearing the ADIPOQ SNP (rs1501299). The PPAR SNP rs1801282, as reported in the study by Zusi et al., has a distribution in the Italian population of 12% in heterozygosity, comparable to that found in the present paper (15%) in ADPKD Italian patients [36]. Interestingly, PPAR $\gamma$  mutated patients had higher adiponectin levels compared to negative patients; such data is in accordance with previous studies that described higher levels of adiponectin in relation to the PPAR $\gamma$  SNP in several diseases, such as diabetes [37,38]. However, it is important to notice that, for the first time, we described this association in a renal disorder. Adiponectin expression is clearly related to PPAR $\gamma$ , since administration of the synthetic PPAR $\gamma$  ligands, the thiazolidinediones (TZDs), increases adiponectin concentrations [39]. Interestingly, its insulin-sensitizing effects are partly mediated by the increased expression of adiponectin [40]. Considering ADIPOQ mutated patients, Pileggi et al. described a

frequency of 12% and 41% of homozygous and heterozygous patients, respectively, in Italian subjects [41]; here, we report a 5.4% of frequency for homozygous and 26% for heterozygous patients. The rs1501299 SNP was previously analyzed in several populations, all describing the mutated genotype as associated with lower adiponectin levels [42]. Indeed, after a hypocaloric diet, the mutated genotype of an ADIPOQ gene variant (rs1501299) was related to a worse improvement in adiponectin levels within a cohort of obese patients [43]. Similar results were reported in healthy school-aged children of Greek origin under a fiber diet [44]. In addition, the distribution of the ADIPOQ rs1501299 significantly differs between CKD and controls. However, although several studies considered metabolic diseases, to our knowledge, this is the first study considering a population characterized by renal disease. Recently, a family harboring a 10-nucleotide deletion mutation in ADIPOQ was reported to co-segregate with diabetes and end-stage renal disease. This mutation introduces a frameshift in exon 3, resulting in a premature termination codon that disrupts translation of adiponectin's globular domain [45].

Adiponectin exerts its effects via interaction with two specific receptors, AdipoR1 and AdipoR2 [46]. Previously, a significant association between ADIPOR2 SNPs and eGFR have been identified suggesting a potential effect of ADIPORs gene variants on kidney function [46]. Therefore, here we investigated the principal SNPs in ADIPOR1-2 genes in our ADPKD population in relation to biochemical and clinical parameters, but no relevant associations were found with biochemical parameters. Kobayashi et al. described several AdipoR2 SNPs associated with kidney function, suggesting that the effects of this polymorphism on adiponectin receptor may affect kidney function in the elderly Japanese population [47]. However, in our population, these SNPs were not present. Overall, our results indicated that determining genetic background could benefit ADPKD patient management and expand the number of genetic susceptibility biomarkers.

It is important to consider that our study does have several limitations: first, our study missed follow-up information; secondly, genetic background is missing in the control subjects, and this might weaken the discriminatory statistical power to identify true associations of genetic variants and clinical characteristics.

## 5. Conclusions

In conclusion, this report demonstrated that serum adiponectin levels could influence the clinical phenotype of ADPKD patients, and that to analyze the association between polymorphisms and ADPKD risk could potentially provide important insights into ameliorating kidney outcome. Our results suggest that additional longitudinal studies and clinical trials should be conducted to investigate if adipocytokines play a role in the development and progression of ADPKD. Further studies that focus on the adiponectin pathway and gene–environment interactions ADPKD patients will aid us in obtaining a deeper understanding of adiponectin role in the establishment and prognosis of PKDs. Studies with larger sample sizes are required to improve the precision of point estimates.

**Supplementary Materials:** The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/genes15040484/s1, Table S1: Genetic characterization of ADPKD patients analyzed in the study. PKD1 and PKD2 gene mutations were shown. Table S2: SNPs in PPARg, ADIPOQ, ADIPOR1, ADIPOR2 genes.

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