

Editorial

Special Issue “Cardiovascular Genetics”

Andreas Brodehl ^{1,*},, Hendrik Milting ¹ and Brenda Gerull ²,

¹ Erich and Hanna Klessmann Institute, Heart and Diabetes Center NRW, University Hospital of the Ruhr-University Bochum, Georgstrasse 11, 32545 Bad Oeynhausen, Germany; hmlting@hdz-nrw.de

² Comprehensive Heart Failure Center (CHFC), Department of Internal Medicine I, University Hospital Würzburg, 97080 Würzburg, Germany; Gerull_B@ukw.de

* Correspondence: abrodehl@hdz-nrw.de; Tel.: +49-(0)5731-973530

Since the beginnings of cardiovascular genetics, it became evident in thousands of clinical cases that many cardiomyopathies, channelopathies, aortopathies as well as complex multifactorial diseases such as coronary artery disease, atherosclerosis or atrial fibrillation (AF) have a genetic etiology.

In this Special Issue, Ho et al. summarized the genetic relationship between genetic risk factors for cardiovascular diseases and dementia [1]. Vesa et al. showed that single nucleotide polymorphisms (SNPs) in *CYP4F2* (leukotriene-B(4) omega-hydroxylase 1) and *VKORC1* (vitamin K epoxide reductase complex 1) might be risk factors for plaque formation in atherosclerosis [2]. As non-alcoholic fatty liver disease (NAFLD) shares molecular metabolic pathways with atherosclerosis-related cardiovascular diseases (CVDs), Castaldo et al. investigated if NAFLD-associated SNPs might also be related with sub-clinical atherosclerosis [3]. However, the authors excluded an association of specific SNPs involved in NAFLD with atherosclerosis [2]. Peripheral arterial disease (PAD) is frequently caused by the atherosclerosis of leg arteries. In addition to environmental factors such as, e.g., smoking, it is estimated that about 20% of cases with PAD might be influenced by heritable factors [4]. In this context, Renner et al. showed that a specific SNP in the promoter region of the *EPO* gene is associated with an early onset of PAD and could in consequence serve as a potential biomarker [5]. Familial hypercholesterolemia (FH) can be caused by mutations in *LDLR* (low density lipoprotein receptor), *APOB* (apolipoprotein B) and *PCSK9* (proprotein convertase subtilisin/kexin type 9). Meshkov et al. genotyped a large cohort of Russian patients with FH and identified over 200 variants in these genes [6]. The authors reported that over 35% of them are novel and specific for the Russian population [6]. Aortic aneurysms can lead to aortic dissections and contribute to sudden death. Creamer et al. summarized in this Special Issue the genetic and molecular knowledge about hereditary aortopathies [7]. Interestingly, the transcriptomic analysis of human ascending aortic tissue performed by Zhou et al. revealed detailed molecular insights into pathways involved in aortic dissection [8]. In addition to inflammation, the cell death and degeneration of smooth muscle cells, the authors focused on autophagy pathways [6]. The work of Greene et al. defined the transcriptomes of normal aortic valves and those with aortic stenosis (AS) and aortic insufficiency (AI) [9]. Of note, valves with AS and AI have a unique specific gene expression pattern [7].

In clinical practice, cardiomyopathies are classified according to their structural and functional features into dilated (DCM), hypertrophic (HCM), arrhythmogenic (ACM), restrictive (RCM) and left-ventricular non-compaction (LVNC) cardiomyopathies. All cardiomyopathies can be caused by genetic and non-genetic factors including, e.g., myocarditis. In 1990, the first HCM-associated mutation was identified in *MYH7*, encoding myosin heavy chain β [10]. In recent decades, several other HCM-associated genes have been described. The majority of these genes encode sarcomere or sarcomere-associated proteins such as, e.g., *MYBPC3* encoding myosin binding protein C3 [11]. Due to the amount and size of HCM-associated genes, next generation sequencing (NGS) techniques



Citation: Brodehl, A.; Milting, H.; Gerull, B. Special Issue “Cardiovascular Genetics”. *Genes* **2021**, *12*, 479. <https://doi.org/10.3390/genes12040479>

Received: 10 March 2021

Accepted: 23 March 2021

Published: 26 March 2021

Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

are frequently used in genetic diagnostics. In this context, the work of Fernlund et al. showed the impact of broad exome sequencing in combination with a virtually defined cardiomyopathy gene panel covering the 60 most likely genes by reanalyzing cases with pediatric cardiomyopathy. The authors revealed pathogenic or likely pathogenic mutations and in 30% additional genetic variants of unknown significance in 50% of the cases [12]. HCM can be also associated with syndromic diseases such as Potocki–Schaffer [13] or Noonan syndrome [14]. Caiazza et al. described an interesting case, where the index patient with Noonan syndrome and HCM carries a likely pathogenic variant in *MYBPC3* and in addition a known pathogenic *PTPN11* (protein tyrosine phosphatase non-receptor type 11) mutation [15]. This report underlines the high relevance of broad genetic testing, which is important for the genetic counseling of the patients and their family members even if syndromic features are present. Peripartum cardiomyopathy (PPCM) is a cardiac disease associated with heart failure and systolic dysfunction during pregnancy or in the first months after delivery. In the majority of cases, PPCM is idiopathic. However, a significant amount of cases with PPCM have a genetic etiology overlapping with other cardiomyopathies. Spracklen et al. presented a review in this Special Issue about the genetic factors involved in PPCM [16]. In most cases, the specific molecular and cellular changes of specific cardiomyopathy-associated mutations are unknown or can only be modelled in cell culture or in animal models because of the lack of explanted human myocardial tissue from mutation carriers. Of note, Sielemann et al. used RNA-sequencing to perform detailed transcriptome analyses revealing differentially expressed genes and regulated pathways in explanted human myocardial tissue samples from cardiomyopathy patients with mutations in *TTN* (titin), *LMNA* (lamin A/C), *RBM20* (RNA binding motif protein 20) and *PKP2* (plakophilin-2) [17]. *TTN*, *LMNA* and *RBM20* are the major DCM genes [18–20] and mutations in *PKP2* are common in patients with ACM [21]. Interestingly, the gene expression signatures differ significantly in these four genetic cardiomyopathies indicating a complex and specific remodeling process in the diseased human hearts depending on the specific genotype [17]. However, the clinical cardiac phenotypes associated with mutations in specific genes can be broad. For example, *RBM20* mutations can cause DCM [22], ACM [23] or LVNC [24]. In this context, Vakhrushev et al. report a *RBM20* variant identified in a patient with ventricular arrhythmia but without further structural abnormalities, broadening the clinical spectrum of *RBM20*-associated cardiomyopathies [25]. *RBM20* is a cardiac splicing factor and is involved in the splicing of several different cardiac genes such as *TTN* and *RYR2* (ryanodine receptor 2) [26,27]. Another example of a gene causing broad cardiac presentations is *DES*, encoding the muscle specific intermediate filament protein desmin. Desmin filaments connect different multiple-protein complexes and cell organelles such as the desmosomes, Z-bands, mitochondria and nuclei [28]. *DES* mutations were found in patients with skeletal myopathies [29] or with different cardiomyopathies such as DCM [30], ACM [31,32] and RCM [33,34]. Kulikova et al. reported in this Special Issue an LVNC family carrying the *DES* missense mutation p.A337P, leading to a severe filament assembly defect in transfected cells [35]. This genetic finding is in good agreement with further reports of other groups identifying *DES* as a novel LVNC gene [36–38]. In contrast to monogenic inherited cardiomyopathies, AF is a complex heart rhythm disorder caused by environmental and multi-genic risk factors [39] and significantly increases the risk for stroke and heart failure. Apixaban is an anticoagulant drug used to reduce the risk for stroke in patients with non-valvular AF [40]. The work of Rosian et al. showed that there was no significant association between peak apixaban plasma concentrations in patients with specific SNPs in the *ABCB1* gene, encoding an ATP-binding cassette (ABC) transporter [41].

In summary, in this Special Issue, different genetic and genomic studies are summarized, describing the impact of genetic factors for different cardiovascular diseases. In addition, three transcriptome analyses of human aortic valves, the ascending aortic tissue and myocardial tissue of mutation carriers with specified genetic cardiomyopathies revealed interesting molecular gene expression patterns in health and disease.

Funding: This research received no external funding.

Acknowledgments: We thank Greta Marie Pohl for proof reading.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Ho, W.M.; Wu, Y.Y.; Chen, Y.C. Genetic Variants behind Cardiovascular Diseases and Dementia. *Genes* **2020**, *11*, 1514. [[CrossRef](#)]
2. Vesa, S.C.; Vlaicu, S.I.; Vacaras, V.; Crisan, S.; Sabin, O.; Pasca, S.; Trifa, A.P.; Rusz-Fogarasi, T.; Sava, M.; Buzoianu, A.D. CYP4F2 and VKORC1 Polymorphisms Amplify the Risk of Carotid Plaque Formation. *Genes* **2020**, *11*, 822. [[CrossRef](#)]
3. Castaldo, L.; Laguzzi, F.; Strawbridge, R.J.; Baldassarre, D.; Veglia, F.; Vigo, L.; Tremoli, E.; de Faire, U.; Eriksson, P.; Smit, A.J.; et al. Genetic Variants Associated with Non-Alcoholic Fatty Liver Disease Do Not Associate with Measures of Sub-Clinical Atherosclerosis: Results from the IMPROVE Study. *Genes* **2020**, *11*, 1243. [[CrossRef](#)]
4. Kullo, I.J.; Leeper, N.J. The genetic basis of peripheral arterial disease: Current knowledge, challenges, and future directions. *Circ. Res.* **2015**, *116*, 1551–1560. [[CrossRef](#)]
5. Renner, W.; Kaiser, M.; Khuen, S.; Trummer, O.; Mangge, H.; Langsenlehner, T. The Erythropoietin rs1617640 Gene Polymorphism Associates with Hemoglobin Levels, Hematocrit and Red Blood Cell Count in Patients with Peripheral Arterial Disease. *Genes* **2020**, *11*, 1305. [[CrossRef](#)] [[PubMed](#)]
6. Meshkov, A.; Ershova, A.; Kiseleva, A.; Zotova, E.; Sotnikova, E.; Petukhova, A.; Zharikova, A.; Malyshev, P.; Rozhkova, T.; Blokhina, A.; et al. The LDLR, APOB, and PCSK9 Variants of Index Patients with Familial Hypercholesterolemia in Russia. *Genes* **2021**, *12*, 66. [[CrossRef](#)]
7. Creamer, T.J.; Bramel, E.E.; MacFarlane, E.G. Insights on the Pathogenesis of Aneurysm through the Study of Hereditary Aortopathies. *Genes* **2021**, *12*, 183. [[CrossRef](#)]
8. Zhou, Z.; Liu, Y.; Zhu, X.; Tang, X.; Wang, Y.; Wang, J.; Xu, C.; Wang, D.; Du, J.; Zhou, Q. Exaggerated Autophagy in Stanford Type A Aortic Dissection: A Transcriptome Pilot Analysis of Human Ascending Aortic Tissues. *Genes* **2020**, *11*, 1187. [[CrossRef](#)] [[PubMed](#)]
9. Greene, C.L.; Jaatinen, K.J.; Wang, H.; Koyano, T.K.; Bilbao, M.S.; Woo, Y.J. Transcriptional Profiling of Normal, Stenotic, and Regurgitant Human Aortic Valves. *Genes* **2020**, *11*, 789. [[CrossRef](#)]
10. Geisterfer-Lowrance, A.A.; Kass, S.; Tanigawa, G.; Vosberg, H.P.; McKenna, W.; Seidman, C.E.; Seidman, J.G. A molecular basis for familial hypertrophic cardiomyopathy: A beta cardiac myosin heavy chain gene missense mutation. *Cell* **1990**, *62*, 999–1006. [[CrossRef](#)]
11. Gerull, B.; Klaassen, S.; Brodehl, A. The genetic landscape of cardiomyopathies. In *Genetic Causes of Cardiac Disease*; Springer: Berlin/Heidelberg, Germany, 2019; pp. 45–91.
12. Fernlund, E.; Kissopoulou, A.; Green, H.; Karlsson, J.E.; Ellegard, R.; Arstrand, H.K.; Jonasson, J.; Gunnarsson, C. Hereditary Hypertrophic Cardiomyopathy in Children and Young Adults-The Value of Reevaluating and Expanding Gene Panel Analyses. *Genes* **2020**, *11*, 1472. [[CrossRef](#)]
13. Romeike, B.F.; Shen, Y.; Nishimoto, H.K.; Morton, C.C.; Layman, L.C.; Kim, H.G. Spectrum of genes involved in a unique case of Potocki Schaffer syndrome with a large chromosome 11 deletion. *Clin. Neuropathol.* **2014**, *33*, 238–244. [[CrossRef](#)] [[PubMed](#)]
14. El Bouchikhi, I.; Belhassan, K.; Moufid, F.Z.; Iraqui Houssaini, M.; Bouguenouch, L.; Samri, I.; Atmani, S.; Ouldlim, K. Noonan syndrome-causing genes: Molecular update and an assessment of the mutation rate. *Int. J. Pediatr. Adolesc. Med.* **2016**, *3*, 133–142. [[CrossRef](#)]
15. Caiazza, M.; Rubino, M.; Monda, E.; Passariello, A.; Fusco, A.; Cirillo, A.; Esposito, A.; Pierro, A.; De Fazio, F.; Pacileo, R.; et al. Combined PTPN11 and MYBPC3 Gene Mutations in an Adult Patient with Noonan Syndrome and Hypertrophic Cardiomyopathy. *Genes* **2020**, *11*, 947. [[CrossRef](#)] [[PubMed](#)]
16. Spracklen, T.F.; Chakafana, G.; Schwartz, P.J.; Kotta, M.C.; Shaboodien, G.; Ntusi, N.A.B.; Sliwa, K. Genetics of Peripartum Cardiomyopathy: Current Knowledge, Future Directions and Clinical Implications. *Genes* **2021**, *12*, 103. [[CrossRef](#)]
17. Sielemann, K.; Elbeck, Z.; Gartner, A.; Brodehl, A.; Stanasiuk, C.; Fox, H.; Palusziewicz, L.; Tiesmeier, J.; Wlost, S.; Gummert, J.; et al. Distinct Myocardial Transcriptomic Profiles of Cardiomyopathies Stratified by the Mutant Genes. *Genes* **2020**, *11*, 1430. [[CrossRef](#)]
18. Gerull, B.; Gramlich, M.; Atherton, J.; McNabb, M.; Trombitas, K.; Sasse-Klaassen, S.; Seidman, J.G.; Seidman, C.; Granzier, H.; Labeit, S.; et al. Mutations of TTN, encoding the giant muscle filament titin, cause familial dilated cardiomyopathy. *Nat. Genet.* **2002**, *30*, 201–204. [[CrossRef](#)]
19. Brodsky, G.L.; Muntoni, F.; Miocic, S.; Sinagra, G.; Sewry, C.; Mestroni, L. Lamin A/C gene mutation associated with dilated cardiomyopathy with variable skeletal muscle involvement. *Circulation* **2000**, *101*, 473–476. [[CrossRef](#)]
20. Gaertner, A.; Klauke, B.; Felski, E.; Kassner, A.; Brodehl, A.; Gerdes, D.; Stanasiuk, C.; Ebbinghaus, H.; Schulz, U.; Dubowy, K.O.; et al. Cardiomyopathy-associated mutations in the RS domain affect nuclear localization of RBM20. *Hum. Mutat.* **2020**, *41*, 1931–1943. [[CrossRef](#)] [[PubMed](#)]
21. Gerull, B.; Heuser, A.; Wichter, T.; Paul, M.; Basson, C.T.; McDermott, D.A.; Lerman, B.B.; Markowitz, S.M.; Ellinor, P.T.; MacRae, C.A.; et al. Mutations in the desmosomal protein plakophilin-2 are common in arrhythmogenic right ventricular cardiomyopathy. *Nat. Genet.* **2004**, *36*, 1162–1164. [[CrossRef](#)]

22. Brauch, K.M.; Karst, M.L.; Herron, K.J.; de Andrade, M.; Pellikka, P.A.; Rodeheffer, R.J.; Michels, V.V.; Olson, T.M. Mutations in ribonucleic acid binding protein gene cause familial dilated cardiomyopathy. *J. Am. Coll. Cardiol.* **2009**, *54*, 930–941. [[CrossRef](#)]
23. Parikh, V.N.; Caleshu, C.; Reuter, C.; Lazzaroni, L.C.; Ingles, J.; Garcia, J.; McCaleb, K.; Adesiyun, T.; Sedaghat-Hamedani, F.; Kumar, S.; et al. Regional Variation in RBM20 Causes a Highly Penetrant Arrhythmogenic Cardiomyopathy. *Circ. Heart Fail.* **2019**, *12*, e005371. [[CrossRef](#)]
24. Sedaghat-Hamedani, F.; Haas, J.; Zhu, F.; Geier, C.; Kayvanpour, E.; Liss, M.; Lai, A.; Frese, K.; Pribe-Wolferts, R.; Amr, A.; et al. Clinical genetics and outcome of left ventricular non-compaction cardiomyopathy. *Eur. Heart J.* **2017**, *38*, 3449–3460. [[CrossRef](#)] [[PubMed](#)]
25. Vakhrushev, Y.; Kozyreva, A.; Semenov, A.; Sokolnikova, P.; Lubimtseva, T.; Lebedev, D.; Smolina, N.; Zhuk, S.; Mitrofanova, L.; Vasichkina, E.; et al. RBM20-Associated Ventricular Arrhythmias in a Patient with Structurally Normal Heart. *Genes* **2021**, *12*, 94. [[CrossRef](#)] [[PubMed](#)]
26. Fochi, S.; Lorenzi, P.; Galasso, M.; Stefani, C.; Trabetti, E.; Zipeto, D.; Romanelli, M.G. The Emerging Role of the RBM20 and PTBP1 Ribonucleoproteins in Heart Development and Cardiovascular Diseases. *Genes* **2020**, *11*, 402. [[CrossRef](#)] [[PubMed](#)]
27. Guo, W.; Schafer, S.; Greaser, M.L.; Radke, M.H.; Liss, M.; Govindarajan, T.; Maatz, H.; Schulz, H.; Li, S.; Parrish, A.M.; et al. RBM20, a gene for hereditary cardiomyopathy, regulates titin splicing. *Nat. Med.* **2012**, *18*, 766–773. [[CrossRef](#)] [[PubMed](#)]
28. Brodehl, A.; Gaertner-Rommel, A.; Milting, H. Molecular insights into cardiomyopathies associated with desmin (DES) mutations. *Biophys Rev.* **2018**, *10*, 983–1006. [[CrossRef](#)]
29. Winter, L.; Wittig, I.; Peeva, V.; Eggers, B.; Heidler, J.; Chevessier, F.; Kley, R.A.; Barkovits, K.; Strecker, V.; Berwanger, C.; et al. Mutant desmin substantially perturbs mitochondrial morphology, function and maintenance in skeletal muscle tissue. *Acta Neuropathol.* **2016**, *132*, 453–473. [[CrossRef](#)]
30. Taylor, M.R.; Slavov, D.; Ku, L.; Di Lenarda, A.; Sinagra, G.; Carniel, E.; Haubold, K.; Boucek, M.M.; Ferguson, D.; Graw, S.L.; et al. Prevalence of desmin mutations in dilated cardiomyopathy. *Circulation* **2007**, *115*, 1244–1251. [[CrossRef](#)]
31. Bermudez-Jimenez, F.J.; Carriel, V.; Brodehl, A.; Alaminos, M.; Campos, A.; Schirmer, I.; Milting, H.; Abril, B.A.; Alvarez, M.; Lopez-Fernandez, S.; et al. Novel Desmin Mutation p.Glu401Asp Impairs Filament Formation, Disrupts Cell Membrane Integrity, and Causes Severe Arrhythmogenic Left Ventricular Cardiomyopathy/Dysplasia. *Circulation* **2018**, *137*, 1595–1610. [[CrossRef](#)]
32. Protonotarios, A.; Brodehl, A.; Asimaki, A.; Jager, J.; Quinn, E.; Stanasiuk, C.; Ratnavadivel, S.; Futema, M.; Akhtar, M.M.; Gossios, T.D.; et al. The novel desmin variant p.Leu115Ile is associated with a unique form of biventricular Arrhythmogenic Cardiomyopathy. *Can. J. Cardiol.* **2020**. [[CrossRef](#)]
33. Brodehl, A.; Pour Hakimi, S.A.; Stanasiuk, C.; Ratnavadivel, S.; Hendig, D.; Gaertner, A.; Gerull, B.; Gummert, J.; Paluszewicz, L.; Milting, H. Restrictive Cardiomyopathy is Caused by a Novel Homozygous Desmin (DES) Mutation p.Y122H Leading to a Severe Filament Assembly Defect. *Genes* **2019**, *10*, 918. [[CrossRef](#)]
34. Herrmann, H.; Cabet, E.; Chevalier, N.R.; Moosmann, J.; Schultheis, D.; Haas, J.; Schowalter, M.; Berwanger, C.; Weyerer, V.; Agaimy, A.; et al. Dual Functional States of R406W-Desmin Assembly Complexes Cause Cardiomyopathy With Severe Intercalated Disc Derangement in Humans and in Knock-In Mice. *Circulation* **2020**, *142*, 2155–2171. [[CrossRef](#)]
35. Kulikova, O.; Brodehl, A.; Kiseleva, A.; Myasnikov, R.; Meshkov, A.; Stanasiuk, C.; Gartner, A.; Divashuk, M.; Sotnikova, E.; Koretskiy, S.; et al. The Desmin (DES) Mutation p.A337P Is Associated with Left-Ventricular Non-Compaction Cardiomyopathy. *Genes* **2021**, *12*, 121. [[CrossRef](#)]
36. Kubanek, M.; Schimerova, T.; Piherova, L.; Brodehl, A.; Krebssova, A.; Ratnavadivel, S.; Stanasiuk, C.; Hansikova, H.; Zeman, J.; Palecek, T.; et al. Desminopathy: Novel Desmin Variants, a New Cardiac Phenotype, and Further Evidence for Secondary Mitochondrial Dysfunction. *J. Clin. Med.* **2020**, *9*, 937. [[CrossRef](#)]
37. Tamiya, R.; Saito, Y.; Fukamachi, D.; Nagashima, K.; Aizawa, Y.; Ohkubo, K.; Hatta, T.; Sezai, A.; Tanaka, M.; Ishikawa, T.; et al. Desmin-related myopathy characterized by non-compaction cardiomyopathy, cardiac conduction defect, and coronary artery dissection. *ESC Heart Fail.* **2020**, *7*, 1338–1343. [[CrossRef](#)]
38. Marakhonov, A.V.; Brodehl, A.; Myasnikov, R.P.; Sparber, P.A.; Kiseleva, A.V.; Kulikova, O.V.; Meshkov, A.N.; Zharikova, A.A.; Koretsky, S.N.; Kharlap, M.S.; et al. Noncompaction cardiomyopathy is caused by a novel in-frame desmin (DES) deletion mutation within the 1A coiled-coil rod segment leading to a severe filament assembly defect. *Hum. Mutat.* **2019**, *40*, 734–741. [[CrossRef](#)]
39. Roselli, C.; Rienstra, M.; Ellinor, P.T. Genetics of Atrial Fibrillation in 2020: GWAS, Genome Sequencing, Polygenic Risk, and Beyond. *Circ. Res.* **2020**, *127*, 21–33. [[CrossRef](#)] [[PubMed](#)]
40. Ioannou, A.; Tsappa, I.; Metaxa, S.; Missouris, C.G. Non-valvular atrial fibrillation: Impact of apixaban on patient outcomes. *Patient Relat. Outcome Meas.* **2017**, *8*, 121–131. [[CrossRef](#)] [[PubMed](#)]
41. Rosian, A.N.; Rosian, S.H.; Kiss, B.; Stefan, M.G.; Trifa, A.P.; Ober, C.D.; Anchidin, O.; Buzoianu, A.D. Interindividual Variability of Apixaban Plasma Concentrations: Influence of Clinical and Genetic Factors in a Real-Life Cohort of Atrial Fibrillation Patients. *Genes* **2020**, *11*, 438. [[CrossRef](#)] [[PubMed](#)]