



**1** Supplementary information

# Theranostic aza-BODIPY as vector for enhanced Boron Neutron Capture Therapy applications

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## 8 Materials and methods for chemical synthesis and characterization

9 Reactions were carried out in analytical reagent grade solvents from Carlo Erba under normal 10 atmosphere. Dry solvents, purchased from Carlo Erba, were non-stabilized and dried using a MB-11 SPS-800 (MBraun) or PureSolv-MD-5 (Inert®). All reagents purchased from Sigma Aldrich<sup>TM</sup>, 12 Thermo Fisher Scientific<sup>™</sup> or ACROS Organics<sup>™</sup> were used as received without further purification. 13 Sodium mercaptododecaborate (10B) was purchased from Katchem<sup>TM</sup>. Reactions were monitored by 14 thin-layer chromatography and RP-HPLC-MS. Analytical thin-layer chromatography was performed 15 with Merck 60 F254 silica gel (precoated sheets, 0.2 mm thick). Column chromatography was carried 16 out using silica gel (Sigma Aldrich; 40-63 µm 230-400 mesh 60Å). Ion exchange was executed using 17 an Amberlite<sup>™</sup> IRA410Cl ion-exchange resin.

(<sup>1</sup>H, <sup>11</sup>B, <sup>13</sup>C, <sup>19</sup>F)-NMR spectra were recorded at 298 or 343 K on Bruker 500 Avance III or 600
Avance III spectrometers. Chemical shifts are given relative to TMS (<sup>1</sup>H, <sup>13</sup>C), BF<sub>3</sub>\*Et<sub>2</sub>O (<sup>11</sup>B, <sup>10</sup>B), CFCl<sub>3</sub>
(<sup>19</sup>F), and were referenced to the residual solvent signal. High resolution mass spectra (HR-MS) were
recorded on a Thermo LTQ Orbitrap XL ESIMS spectrometer. NMR and mass-analyses were
performed at the "Plateforme d'Analyse Chimique et de Synthèse Moléculaire de l'Université de
Bourgogne" (PACSMUB).

HPLC-MS analyses were obtained from a Thermo-Dionex Ultimate 3000 instrument (pump + autosampler at 20 °C + column oven at 25 °C) equipped with a diode array detector (Thermo-Dionex DAD 3000-RS) and a MSQ Plus single quadrupole mass spectrometer equipped with Phenomenex Kinetex® column (2.6 μm C18 100 Å, LC Column 50 x 2.1 mm).

- 28 The employed gradient for analyses was as follows:
- 29

Time [min]	% H2O + 0.1% formic acid	% ACN + 0.1% formic acid	Flow [mL/min]
0	95	5	0.5
5	0	100	0.5
6.5	0	100	0.5
6.6	95	5	0.5
8.5	95	5	0.5
8.51	95	5	0.05

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Table S1: HPLC analytical gradient.

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Semi-preparative separations were executed on a HPLC-system, from Shimadzu, that was
equipped with 2 LC-20AT pumps, a SPD-20A UV/Vis detector, a FRC-10A fraction collector, a SIL10AP sampler and a CBM-20A control unit. The column was a Shim-Pack GIST 5 µm C18 10x250 mm
column obtained from Shimadzu too. The gradient using a mixture of ACN and water with 0.1% TFA
and a flow rate of 5 mL/min was as follows:

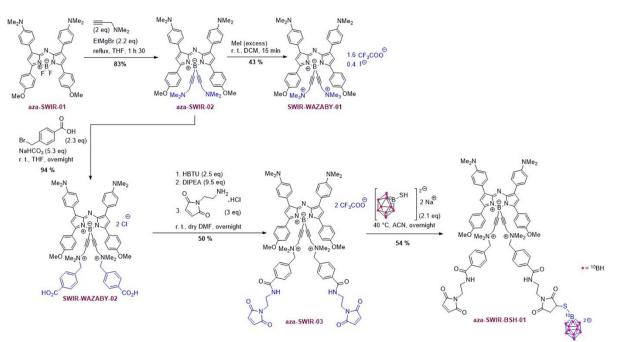
n				
9	Time	% H2O +	% ACN +	Flow
0				
	[min]	0.1 %TFA	0.1 %TFA	[mL/min]
<u>,</u>	0	75	25	5
1	5	75	25	5
	25	0	100	5
	28	0	100	5
	30	75	25	5
	50	75	20	5



Table S2: Detailed of gradients.

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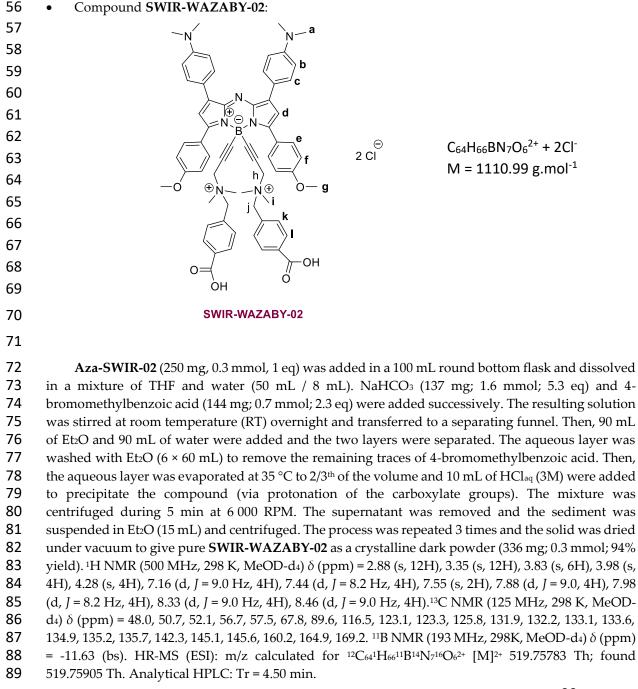
### 50 Synthesis and characterization

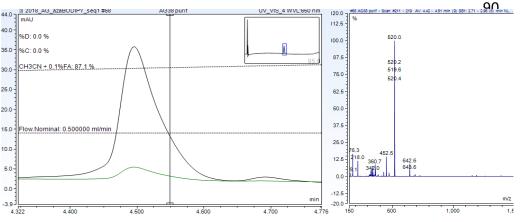


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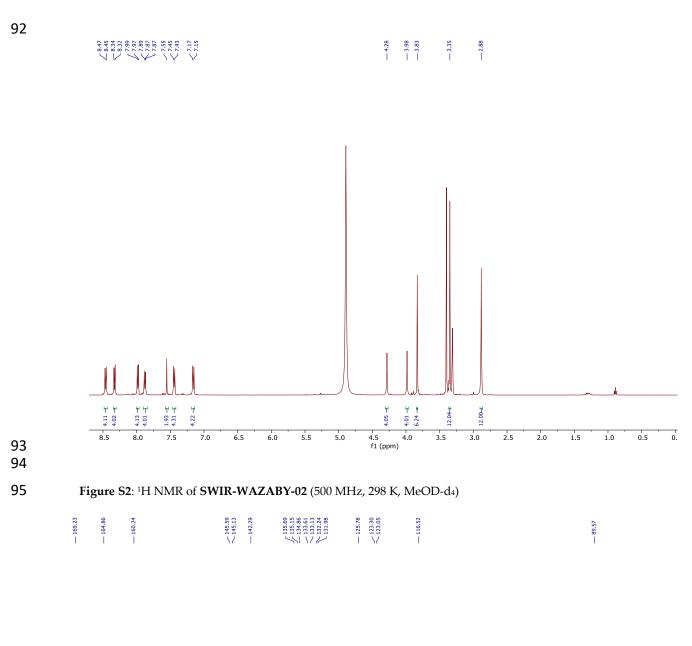
Scheme S1: Synthetic pathways of aza-SWIR-BSH-01.

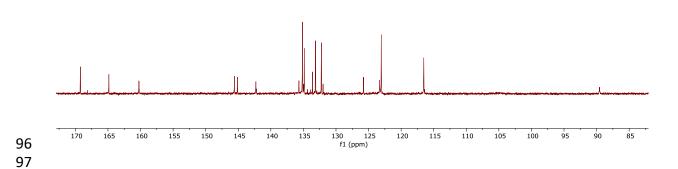
- 53 Aza-SWIR-01, aza-SWIR-02, and SWIR-WAZABY-01 were synthesized according to procedures,
- 54 we previously reported [1].
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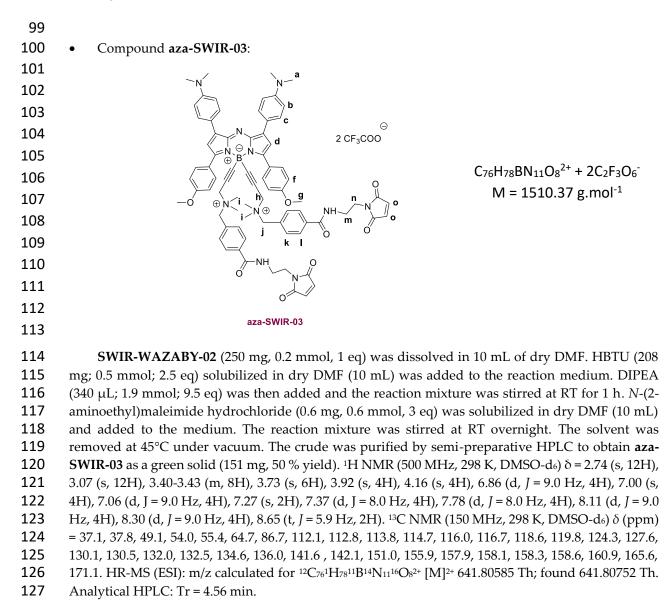
91 Figure S1: Analytical HPLC of SWIR-WAZABY-02

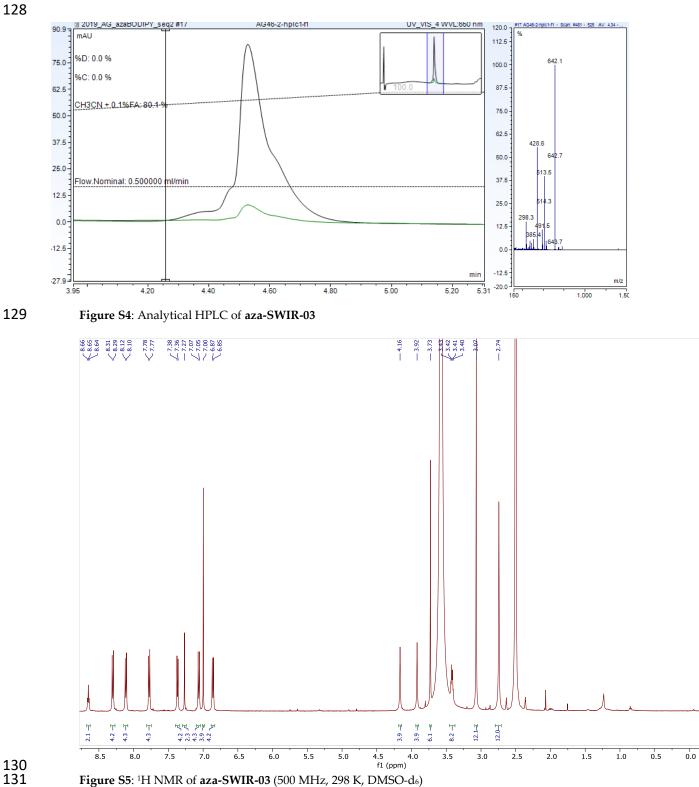


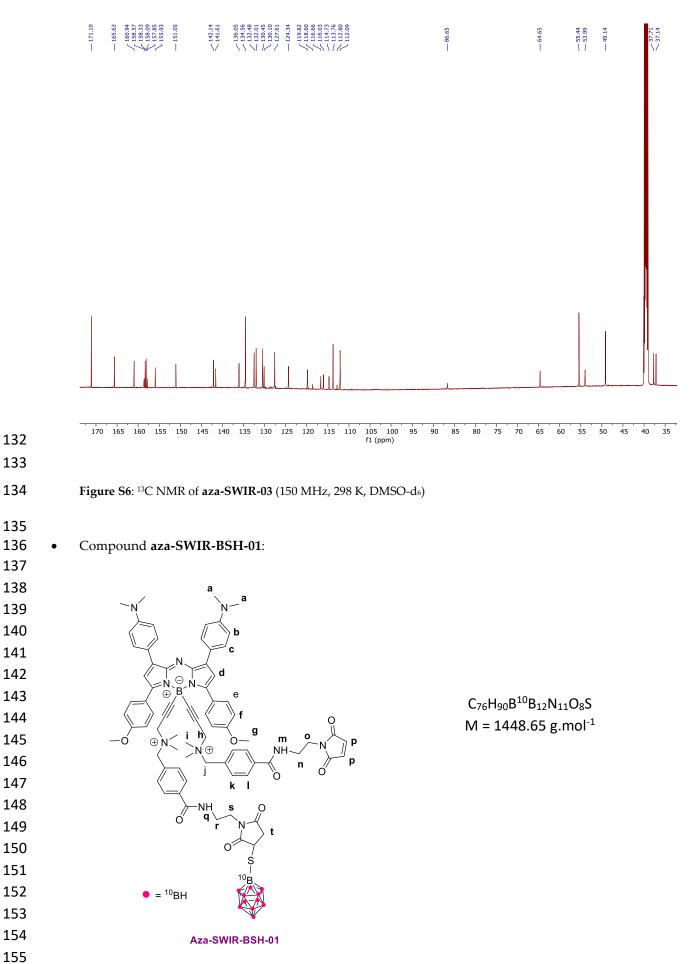


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#### Figure S3: <sup>13</sup>C NMR of SWIR-WAZABY-02 (125 MHz, 298 K, MeOD-d4)

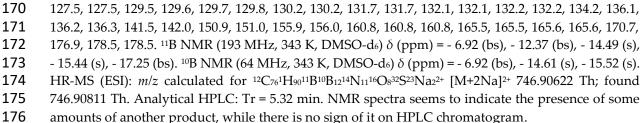


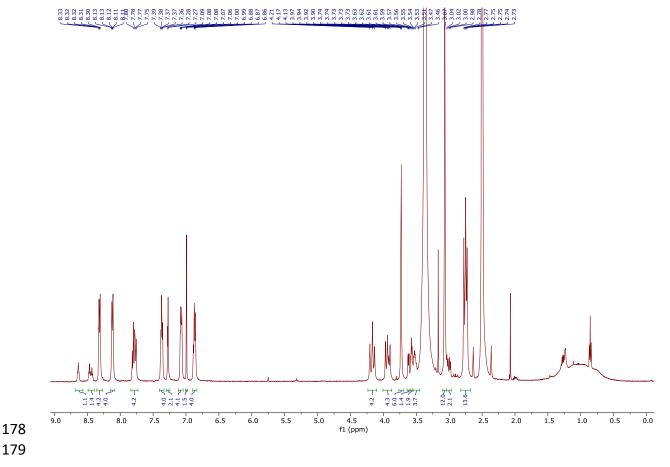




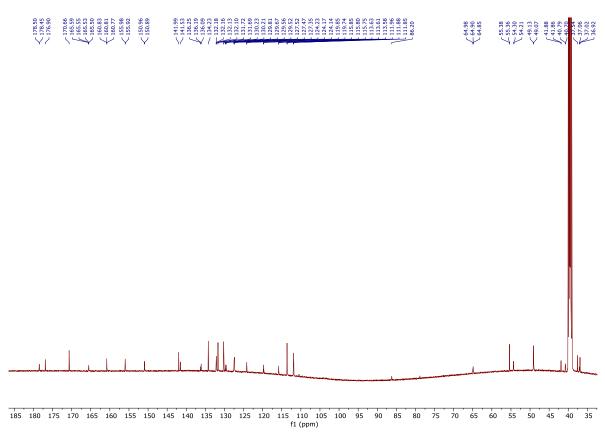
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**Aza-SWIR-03** (120 mg, 79 μmol, 1 eq) was dissolved in 4 mL of ACN in a 10 mL round-bottom flask. Sodium mercaptoundecahydrododecaborate (<sup>10</sup>B) (42 mg; 166 μmol; 2.1 eq) was added and the reaction mixture was stirred overnight at 40 °C, leading to the formation of a precipitate. The mixture was centrifuged during 5 min at 6 000 RPM. The supernatant was removed and the sediment was suspended in ACN (15 mL), the process was repeated three times with ACN, once with DCM (15 mL), three time with EtzO (15 mL) and twice with pentane (15 mL) to obtain **aza-SWIR-BSH-01** as a green solid (62 mg, 54%). <sup>1</sup>H NMR\* (500 MHz, 298 K, DMSO-d<sub>6</sub>) δ = 1.03 (bs, 11H), 2.73–2.78 (m, 14H), 3.01 (dd, *J* = 8.3; 18.4 Hz, 2H), 3.07 (s, 12H), 3.46–3.56 (m, 4H), 3.58 (t, *J* = 5.8 Hz, 2H), 3.62 (dd, *J* = 8.3; 3.1 Hz, 1H), 3.73–3.74 (m, 6H), 3.90–3.97 (m, 2H), 4.17 (t, *J* = 17.9 Hz, 4H), 6.85–6.89 (m, 4H), 6.99–7.00 (m, 2H), 7.07–7.09 (m, 4H), 7.26–7.28 (m, 2H), 7.36–7.39 (m, 4H), 7.76–7.82 (m, 4H), 8.10–8.13 (m, 4H), 8.30–8.33 (m, 4H), 8.43 (t, *J* = 5.8 Hz, 1H), 8.64 (t, *J* = 5.8 Hz, 1H). <sup>13</sup>C NMR (150 MHz, 343 K, DMSO-d<sub>6</sub>) δ (ppm) = 36.9, 37.0, 37.1, 37.5, 40.7, 40.8, 41.9, 41.9, 49.1, 49.1, 54.2, 54.3, 55.4, 55.4, 64.9, 64.9, 65.0, 86.2, 111.9, 111.9, 112.0, 113.6, 113.6, 113.6, 115.8, 115.8, 115.8, 119.7, 119.9, 124.1, 124.2, 124.2, 127.4, 127.5, 127.5, 129.5, 129.5, 129.6, 120.7, 120.9, 120.2,







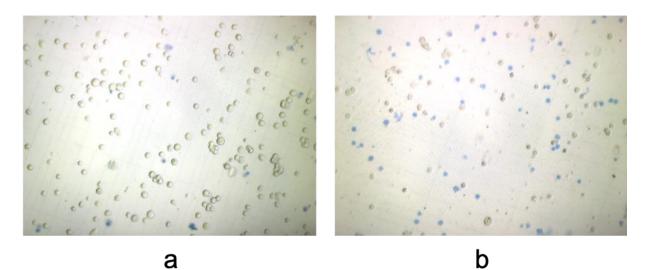


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**183** Figure S8: <sup>13</sup>C NMR of aza-SWIR-BSH-01 (150 MHz, 343 K, DMSO-d<sub>6</sub>.

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Figure S9: U-251 cells incubated with aza-SWIR-BSH-01 before (a) and after (b) 10 min of neutron
 exposure. The clichés were taken from microscopy observation. Cells were diluted with Trypan blue
 to count the viable cells and exclude the dead cells (in blue) on Malassez slides. The presence of huge
 number of blue cells in (b) evidences the strong and immediate impact of the neutron exposure on
 cells incubated with aza-SWIR-BSH-01.

Cells 2020, 9, 1953

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194	1.	Godard, A.; Kalot, G.; Pliquett, J.; Busser, B.; Le Guevel, X.; Wegner, K.D.; Resch-Genger, U.;
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196		Organic Dyes for High Contrast In Vivo NIR-II Imaging. Bioconjug. Chem. 2020, 31,
197		1088-1092, doi:10.1021/acs.bioconjchem.0c00175.
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