

Review

Alzheimer's Disease, a Lipid Story: Involvement of Peroxisome Proliferator-Activated Receptor α

Francisco Sáez-Orellana ^{1,2,†}, Jean-Noël Octave ^{1,2} and Nathalie Pierrot ^{1,2,†,*}

¹ Université Catholique de Louvain, Alzheimer Dementia, Avenue Mounier 53, SSS/IONS/CEMO-Bte B1.53.03, B-1200 Brussels, Belgium; francisco.saez@uclouvain.be (F.S.-O.); jean-noel.octave@uclouvain.be (J.-N.O.)

² Institute of Neuroscience, Alzheimer Dementia, Avenue Mounier 53, SSS/IONS/CEMO-Bte B1.53.03, B-1200 Brussels, Belgium

* Correspondence: nathalie.pierrot@uclouvain.be; Tel.: +32-2-764-93-34

† These authors contributed equally to this work.

Received: 17 April 2020; Accepted: 12 May 2020; Published: 14 May 2020

Abstract: Alzheimer's disease (AD) is the leading cause of dementia in the elderly. Mutations in genes encoding proteins involved in amyloid- β peptide ($A\beta$) production are responsible for inherited AD cases. The amyloid cascade hypothesis was proposed to explain the pathogeny. Despite the fact that $A\beta$ is considered as the main culprit of the pathology, most clinical trials focusing on $A\beta$ failed and suggested that earlier interventions are needed to influence the course of AD. Therefore, identifying risk factors that predispose to AD is crucial. Among them, the epsilon 4 allele of the *apolipoprotein E* gene that encodes the major brain lipid carrier and metabolic disorders such as obesity and type 2 diabetes were identified as AD risk factors, suggesting that abnormal lipid metabolism could influence the progression of the disease. Among lipids, fatty acids (FAs) play a fundamental role in proper brain function, including memory. Peroxisome proliferator-activated receptor α (PPAR α) is a master metabolic regulator that regulates the catabolism of FA. Several studies report an essential role of PPAR α in neuronal function governing synaptic plasticity and cognition. In this review, we explore the implication of lipid metabolism in AD, with a special focus on PPAR α and its potential role in AD therapy.

Keywords: Alzheimer's; risk factors; PPARs; PPAR α , lipids; fatty acids; modulators; cognition; sex; therapy

1. Alzheimer Disease: A Dementing Illness

With a prevalence doubling every 5 years beyond 65, Alzheimer's disease (AD) is a devastating neurodegenerative disorder, which is the most common cause of dementia in the elderly. In 2019, Alzheimer's Disease International estimated that there are over 50 million people living with dementia globally, a figure set to increase to 152 million by 2050 [1].

Memory loss is one of the main clinical features of AD onset, and numerous neuropsychological tests allow for the assessment of the cognitive functions of Alzheimer's patients [2,3]. Along with a decline in cognitive performance, AD is characterized by the coexistence in the brain of two main neuropathological lesions: intraneuronal neurofibrillary tangles composed of hyperphosphorylated microtubule-associated protein tau and extracellular senile plaques containing the amyloid- β ($A\beta$) peptide generated from the sequential proteolytic processing of its precursor, the amyloid precursor protein (APP). Although the definitive diagnosis of the disease was previously achieved by the postmortem neuropathological brain examination, the detection of specific AD biomarkers in the cerebrospinal fluid, including $A\beta$ and tau, constitutes an early examination and a reliable diagnosis

[4]. Moreover, recent non-invasive imaging techniques using A β - and tau-PET tracers have led to the preclinical diagnosis of AD, allowing its evolution during the patient's lifetime to be tracked [5–7]. As positron emission tomography (PET) imaging studies have shown that A β accumulation occurs long before the onset of clinical AD and given that mutations in the *APP* gene can act as fully penetrant in rare inherited early-onset AD cases (EOAD, about 1% of the cases, (Figure 1)), the amyloid cascade hypothesis was proposed to explain the pathogeny. According to this hypothesis, a gradual accumulation and aggregation of A β initiate a neurodegenerative cascade resulting in neurofibrillary tangles formation, cell loss, vascular damage and dementia [8,9]. Although this hypothesis was strengthened by the discovery that mutations in presenilins (Figure 1), the catalytic subunits of the γ -secretase complex, lead to an increase in A β production [10], several studies have challenged the amyloid hypothesis over the past ten years [11,12]. Mounting evidence reports that mutations in the *Presenilin 1* (*PSEN1*) gene associated with EOAD have heterogenic manifestations. It was indeed recently shown that the most common mutations found in *PSEN1* decrease the activity of the γ -secretase [13,14] or lead to a loss of its function [15], indicating that in some cases, *PSEN1* mutations either hyper-activate or reduce the activity of the γ -secretase complex. Moreover, γ -secretase activity assessed in brain samples from EOAD and non-demented controls was similar, while it displayed some dysfunctions in a few brain samples from late-onset AD cases (LOAD), which represent the vast majority of AD cases [14]. This suggests that γ -secretase may also play a role in some LOAD cases, in which brain A β production levels are similar to those observed in unaffected controls [16].

Moreover, several studies have shown that humans with Down syndrome, who harbor three copies of the *APP* gene that leads to the overexpression of APP protein, have an age-dependent increased risk for developing AD and develop clinical features and neuropathological changes similar to those observed in AD (for review, see [17]). This aforementioned study suggests that AD could be a combination of different pathologies with diverse etiologies [18] leading to dementia. This is supported by recent findings showing that some pathologies have similar clinical markers and manifestations to those observed in AD, as reported in limbic-predominant, age-related TDP-43 encephalopathy (LATE), in which senile plaques and neurofibrillary tangles that define AD have been brought out in the brain [19].

Since A β that builds up in plaques also deposits during normal brain aging, amyloid deposition occurring in the hippocampus and cerebral cortex of AD patients potentially explains deficits in memory and cognitive function observed. Despite the various isoforms of A β produced, A β toxicity rate is dependent on its state of assembly. Among the three assemblies state of A β (monomers, soluble oligomers and insoluble fibrils) (for review, see [20]), soluble A β oligomers are organized into different structures ranging from dimers, trimers, tetramers, pentamers, decamers and dodecamers, among others [21–23]. Toxic soluble oligomers have been identified in AD brains [24–26]. However, in some cases, higher aggregates, such as fibrils, showed protective effects in AD models [27], suggesting that there is an inverse correlation between the size of A β assembly and its toxicity. Since A β dimers form a more stable structure, these dimeric units are described to be the building blocks for toxic aggregates [28]. This supports the idea that disassembling plaques or fibrillar structures could be detrimental if not accompanied by strategies to remove oligomeric aggregates of A β . However, it remains difficult to influence the course of AD by removing amyloid deposits. Indeed, some approaches were developed to inhibit secretases activities involved in the release of A β or to remove amyloid deposits from the brain using active or passive immunotherapy [29]. While first attempts completely failed in Phase III clinical trials due to the widespread function of secretases and the development of encephalitis in some patients [30], recent attempts have shown that although senile plaques can be effectively removed from the AD brain, cognitive performance is not improved in these patients [31]. Despite the fact that recent results with an immunotherapy clinical trial using aducanumab targeting aggregated forms of A β have been encouraging and could prove efficacious [32], this clearly suggests that when structural modifications are found in the brain due to accumulation of abnormal proteins, the proposed treatments arrive too late and are inefficient [33]. Consequently, we must focus on modifications in physiological functions, which could occur long

before abnormal protein deposition. Among them, abnormal lipid metabolism could be an important early event in the pathogenesis of AD.

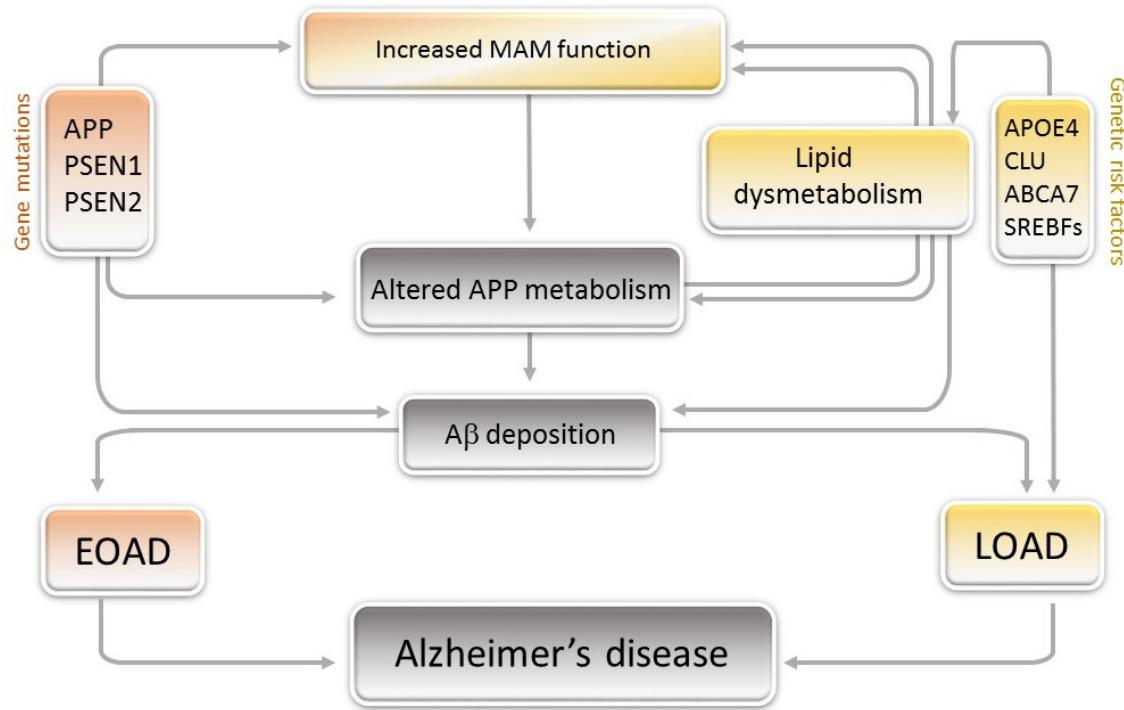


Figure 1. Gene mutations and genetic risk factors linked to lipid dysmetabolism and the progression of Alzheimer's disease (AD). Gene mutations responsible for inherited early-onset AD cases (EOAD, gene mutations) and genetic risk factors for late-onset AD cases (LOAD, genetic risk factors) lead to altered amyloid precursor protein (APP) processing and brain amyloid- β (A β) deposition. Disruption of lipid homeostasis induces abnormal lipid composition in rafts and increased mitochondria-associated endoplasmic reticulum membrane (MAM) function in which targeted APP is proteolytically processed into A β by presenilins (PSEN). Conversely, cleavage of APP directly affects cellular lipid composition by altering the synthesis of several lipids that are enriched in lipid rafts. Abbreviations: APOE4 (Apolipoprotein E4); CLU (Clusterin); ABCA7 (ATP-binding cassette subfamily A member 7); SREBFs (Sterol regulatory element-binding genes).

2. Linking Lipids to Alzheimer's Disease

While in EOAD, A β accumulation in the brain is caused by gene mutations, the vast majority of AD cases are late-onset AD cases (LOAD, 99% of cases), in which the source of A β accumulation in the brain is still unknown. Nevertheless, it is well established that the epsilon 4 allele of the Apolipoprotein E (APOE) gene, encoding the main lipid carrier in the brain, is a genetic risk factor for AD. People who are homozygous for this allele have ten times greater risk to develop AD [34]. Therefore, a relationship between AD and lipid metabolism has been established. Furthermore, large-scale genome-wide association studies on AD first confirmed that APOE is a major risk factor and provided evidence that at least 20 genetic susceptibility loci in addition to APOE genotype are associated with AD [35,36] (Figure 2). Among them are genes encoding Clusterin and ABCA7, two proteins involved in lipid metabolism [37–40] (Figures 1 and 2). Therefore, the identification of these susceptibility loci supports the hypothesis that perturbation of lipid metabolism favors the progression of AD [41]. This hypothesis is sustained by recent reports showing that genetic polymorphisms in SREBF genes encoding sterol regulatory element-binding proteins (SREBPs), transcription factors activating lipid metabolism-related genes involved in cholesterol and fatty acids biosynthesis [42,43], were associated with an increased risk of schizophrenia and LOAD [44–46]. Disturbances in the signaling and expression of SREBPs were indeed reported in LOAD cases and, in a rare case of EOAD, harboring a microduplication in the locus of APP gene [47,48].

Additional support comes from metabolomic studies that have shown changes in the lipid content of plasma, cerebrospinal fluid and in brain tissue from AD [49–52]. Moreover, perturbations in brain fatty acids profiles observed in brain regions vulnerable to AD pathology [49,53] could influence AD pathogenesis by promoting A β accumulation and tau pathology [54–56].

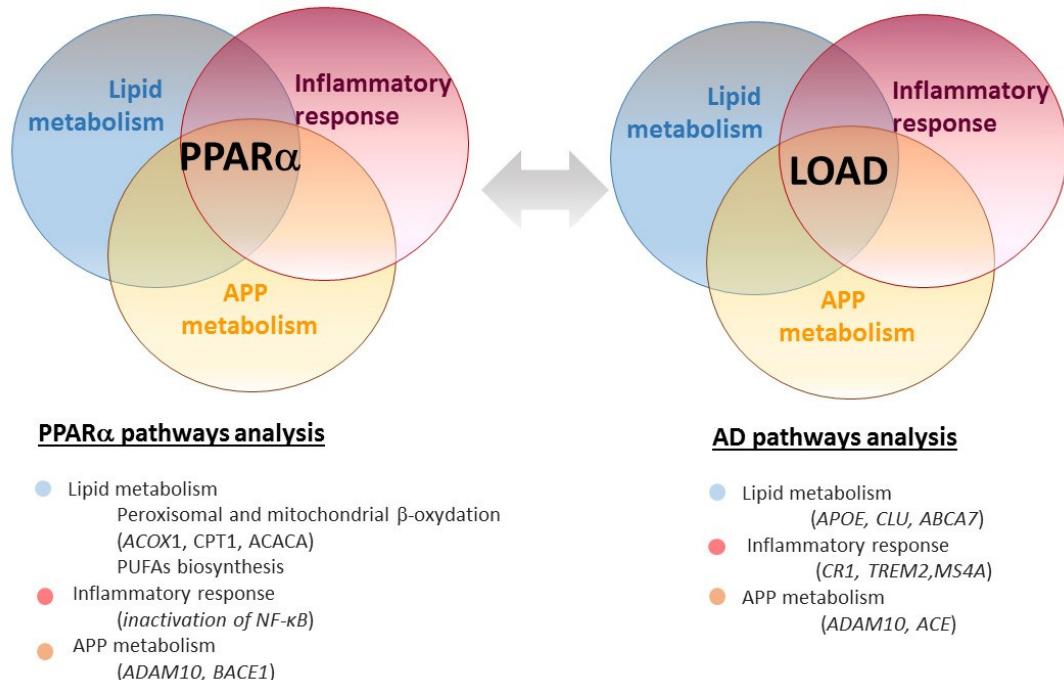


Figure 2. Common pathways regulated by proliferator-activated receptor α (PPAR α) and involved in the etiology of AD. Peroxisome proliferator-activated receptor α (PPAR α) is a transcription factor that governs pathways involved in the metabolism of lipids, inflammatory response and the metabolism of the amyloid precursor protein (APP), which have been implicated in the etiology of late-onset Alzheimer's disease (LOAD). Genome-wide association studies identified several genetic risk factors for LOAD, which are involved in pathways that are governed by PPAR α . Abbreviations: ABCA7 (ATP-binding cassette subfamily A member 7), ACE (Angiotensin-converting enzyme gene), APOE (Apolipoprotein E), APP (Amyloid precursor protein), ACACA (Acetyl-CoA carboxylase), ACOX1 (Acyl-CoA oxidase), ADAM10 (ADAM Metallopeptidase Domain 10), BACE1 (β -Site amyloid precursor protein-cleaving enzyme 1), CLU (Clusterin), CPT1 (Carnitine palmitoyl transferase), CR1 (Complement receptor 1), NF- κ B (Nuclear factor κ B), MS4A (Membrane-spanning 4A), PUFAs (Polyunsaturated fatty acids), TREM2 (Triggering receptor expressed on myeloid cells 2).

3. Lipids in Alzheimer Disease: Involvement of Fatty Acids in Cognitive Function

The human brain contains the second-highest concentration of lipids (50–60% of its dry weight) after adipose tissue [57]. Due to their structural diversity and their involvement in a wide range of biological processes, lipids play a fundamental role in maintaining brain physiological functions. Phospholipids, sterols, sphingolipids, fatty acids and triacylglycerols are the five main brain lipid classes, which are involved in neuronal differentiation, synaptogenesis, and brain development (for reviews, see [58,59]). At the subcellular level, lipids are basic structural components of cell membranes and are enriched in the myelin sheath surrounding nerve cell axons, which regulates the ability of a neuron to trigger action potentials encoding information [60]. Moreover, lipids and their derivatives modulate membrane fluidity and permeability, which regulate trafficking, localization and function of ion pumps, channels, receptors and transporters at the plasma membrane [61,62]. In particular, any change in lipid homeostasis affects the lipid composition of membrane lipid rafts, which are cholesterol and sphingolipid-enriched microdomains [63] where most of the synaptic-related proteins involved in synaptic transmission and plasticity are embedded [64]. Since lipid rafts

from AD brains displayed important changes in their composition [65,66], disturbance in the function of lipid-rafts-associated synaptic proteins could contribute to the development of neuropathological events that favor amyloidogenesis and proteins aggregation [67,68].

One of the main pathological characteristics involved in the pathogenesis of AD implies the APP protein, which is hydrolyzed by β - and γ -secretases, leading to the deposition of A β in the brain. APP is a single-pass transmembrane protein with a large extracellular region that contains several domains involved in APP dimerization, protein–protein or metal interactions (e.g., heparin- and copper-binding domains). The APP trans-membrane-helix domain in which the A β sequence is inserted and the APP intracellular domain contain cholesterol-binding [69] and YENPTY motifs that regulate the subcellular location, trafficking, and proteolytic processing of APP, respectively [70] (for more details, see [71,72]). These domains and motifs engage APP and its cleavage products in a plethora of physiological functions ranging from synaptic transmission, plasticity, development, neuroprotection, trophic function, cell adhesion, apoptosis, calcium and lipid homeostasis, among others [73–75].

The cholesterol-binding motif found in APP plays an essential role in the interaction of APP with proteins involved in cholesterol metabolism (e.g., SREBP1) [48] and in its location in lipid rafts present in synaptic vesicles and mitochondria-associated endoplasmic reticulum (ER) membranes (MAMs) [76] (Figure 1). MAMs are enriched in cholesterol and sphingomyelin and are points of physical contact between the outer mitochondrial membrane and ER. While they play an essential role in the metabolism of glucose, phospholipids, cholesterol and calcium homeostasis [77,78], MAMs regulate APP processing. Indeed, presenilins and γ -secretase activity, previously localized at the ER, are enriched in MAMs, in which β -cleaved fragment generating A β accumulates [79,80]. Interestingly, the activity/function and expression of MAM-associated proteins increase in human and mouse AD brains long before A β deposition [81], suggesting a potential role of MAMs in the pathophysiology of AD [81,82]. From these data, the concept of the MAM hypothesis in AD emerged (reviewed in more details in [83],[84]).

While rafts are described as noncaveolar lipid microdomains, caveolae are cholesterol-enriched membrane invaginations found in the Golgi network, exocytotic vesicles, ER and plasma membrane in which surface protein markers caveolin are embedded [85,86]. Caveolae are involved in cellular cholesterol transport and are docking sites for signaling proteins and receptors and are therefore considered as hotspots for cell–cell communication [87]. Although caveolae-dependent cell signaling is not yet fully understood, several studies have reported the involvement of caveolin proteins in the pathogenesis of AD [88,89]. Caveolin expression levels are upregulated in the hippocampus and the frontal cortex of AD brain compared to control, suggesting a link between the expression of caveolin and dysregulation of cholesterol homeostasis observed in AD [89–91]. Moreover, increased expression in caveolin promotes oxidative stress and APP processing into A β [89,92] that could favor the progression of AD. Although cholesterol and sphingolipid-enriched membrane microdomains could take part in AD physiopathology, fatty acids seem to also contribute to its occurrence.

Fatty acids (FAs) are the major essential monomeric constituents of all lipids [93] and therefore are key components of cellular membranes [94]. They can be unesterified (free) or esterified to plasma membrane phospholipids and are classified based on the length of their carbon chain. FAs are either saturated, monounsaturated, or polyunsaturated (PUFAs) (for review, see [95]). While the brain can produce saturated and monosaturated FAs by de novo lipogenesis, essential PUFAs cannot be synthesized in sufficient quantities [96] and therefore are provided by the diet [94]. Within the brain, palmitic acid and stearic acid are the main saturated FAs, and oleic acid is the main monounsaturated one. Linoleic and α -linolenic essential FAs are transformed into arachidonic and docosahexaenoic acids, the major brain ω -6 and ω -3 PUFAs, respectively [94,95].

PUFAs play a critical role in neurogenesis, synaptic function, inflammation, glucose homeostasis, mood and cognition [94]. As they play a critical role in brain development and functioning [97], high concentrations of dietary saturated long-chain FAs and a decrease in dietary

consumption of ω -3 PUFAs have been associated with neurological dysfunction and neuropsychiatric disorders, including neurodegenerative diseases such as AD [98–100]. In addition, diets in the western population are rich in saturated FAs and low in PUFAs [101], which are not only associated with the development of obesity but also to cognitive dysfunction.

Levels of docosahexaenoic acid (DHA), the major brain ω -3 PUFAs, have been reported to be decreased in plasma and post-mortem brains from AD patients [49–51]. Although DHA dietary supplements did not improve memory, cognition or mood [50,102,103], higher dietary intake of DHA is associated with decreased risk of neurological disorders [104] and dementia in elderly individuals [105]. Interestingly, ω -3 PUFAs supplementation in individuals with mild cognitive impairment and in AD patients without the *APOE4* allele has shown benefits [102,106]. While low brain DHA levels were shown to impair behavior in AD mouse models [107,108], the dietary supplementation of ω -3 PUFAs in rodents facilitated hippocampal synaptic plasticity and improved cognitive deficits of aged mice and in several animal models of AD [50,102,109–112]. Moreover, in non-pathological conditions, maternal intake of ω -3 PUFAs increases hippocampal plasticity and cognition in healthy pups rodents [113].

While cellular and molecular mechanisms underlying such effects are poorly understood, more and more studies put forward the involvement of nuclear receptors.

4. RXRs, LXR α s and PPARs Nuclear Receptors in AD

4.1. Nuclear Receptors

The nuclear receptors superfamily of ligand-dependent transcription factors regulates energy balance, inflammation, and lipid and glucose metabolism [114]. They control target genes expression through their binding with sequence-specific elements located in gene promoter regions [114]. Structurally, they contain an amino-terminal activation domain needed for the recruitment of coactivators, a carboxyl-terminal ligand- and a DNA-binding domain. Among these receptors, retinoid X receptors (RXRs), liver X receptors (LXR α s) and peroxisome proliferator-activated receptors (PPARs) act as master regulators of lipid metabolism by *trans*-activating genes encoding enzymes involved in lipid and fat metabolism. Therefore, they are abundantly expressed in metabolically active tissues, including the brains of rodents and humans [115]. Due to their anti-inflammatory and potential neuroprotective effects, RXRs, LXR α s and PPARs activation with specific agonists emerged as promising approaches for treating brain pathologies in several mouse models of Parkinson's, Huntington and Alzheimer's diseases, multiple and amyotrophic lateral sclerosis, stroke and even in a mouse model with physiological brain aging-dependent cognitive decline (reviewed in [116,117]).

4.2. RXRs

Among the three RXR isotypes identified (RXR α , β and γ), RXR α is mainly expressed in the liver, lungs, muscles, kidneys, epidermis and intestine. While RXR β is expressed ubiquitously, RXR γ is enriched in the brain, heart and muscles. RXRs can be activated by 9-*cis* retinoic acid, linoleic, linolenic and DHA acids, natural RXR ligands [118,119]. As a strong agonist of the RXRs, the retinoid bexarotene synthetic agonist [120], which the U.S. FDA approved for the treatment of cutaneous T-cell lymphoma [121], was described to improve cognitive deficits in AD mouse models [122–127] mainly by inducing the transcription and lipidation of APOE and reducing microglial expression of pro-inflammatory genes among others [128,129]. Although we previously reported that bexarotene improved cognition in a patient with mild AD [130], its efficacy in clinical trials for treating AD pathology has been disappointing [131,132].

4.3. LXR α s

As an obligate binding partner of LXR α s, RXRs form permissive heterodimers with two LXR isoforms, LXR α and β [114]. LXR α is abundantly expressed in the liver, intestine, kidney, spleen and adipose tissue, whereas LXR β is ubiquitously expressed at a lower level but more widely in the brain and mainly in the hippocampus. LXR α s are activated by oxysterols, most prominently hydroxylated

forms of cholesterol [133,134]. They play therefore a critical role in the control of whole-body cholesterol homeostasis and exert potent anti-inflammatory actions [135]. Once activated, they control the transcription of target genes involved in lipid transport and biosynthesis, such as APOE and SREBP, respectively [136,137]. The expression of the SREBP1 isoform is mediated by LXR_s to ensure FAs synthesis needed for the esterification of free cholesterol for protecting cells from a detrimental cholesterol overload. Moreover, unesterified PUFAs exert feedback inhibition on the expression of SREBP1 by antagonizing the oxysterol LXR receptor [138,139].

4.4. PPARs

PPARs were first described for their ability to induce peroxisomal proliferation in the liver in response to xenobiotics [140]. Afterward, they were considered as master metabolic regulators involved in energy homeostasis [141]. They act principally as lipid sensors and regulate whole-body metabolism in response to dietary lipid intake and control their subsequent metabolism and storage by inducing or repressing the expression of genes involved in the metabolism of lipid and glucose [142]. The three PPARs isoforms identified (PPAR α , β/δ and γ) have partially overlapping functions and tissue distribution in mammals. PPAR α is highly expressed in the liver, heart and kidney but has low levels in the brain [143,144] and more particularly in the hippocampus of rodents and primates [144–149]. PPAR α plays an important role in the regulation of FAs catabolism [150] by controlling the expression of genes encoding acyl-CoA oxidase, carnitine palmitoyl transferase and acetyl-CoA carboxylase, enzymes that tightly regulate FAs peroxisomal and mitochondrial β -oxidation, respectively (for reviews, see [151,152]) (Figure 1). Consistent with the first central role of PPAR α in FAs catabolism [150], PPAR α null mice exhibit greater lipid accumulation [153].

While PPAR γ isoform is mainly expressed in white and brown adipose tissue, the large intestine and spleen, in which it regulates adipogenesis, energy balance, lipid biosynthesis and inflammation [154], PPAR β/δ is expressed ubiquitously in all tissues and is the most abundant isoform found in liver, kidney, adipose tissue and skeletal muscle, where it plays mainly a role in FAs oxidation [155].

Although PPARs expression is ubiquitous in the human and mouse brain, PPAR α and γ are expressed in both neurons and astrocytes, while PPAR β/δ isoform is exclusively neuronal [115].

Although PPARs were first classified as orphan receptors, many natural and synthetic agonists of PPARs are used in the treatment of glucose and lipid disorders. Several endogenous ligands from dietary lipids and their metabolites were identified, among them the essential FA DHA and eicosanoids [156,157]. Recently, hexadecanamide, 9-octadecanamide and 3-hydroxy-(2,2)-dimethyl butyrate have been identified as endogenous PPAR α ligands in mouse brain hippocampus [158]. Moreover, several synthetic ligands are widely used in clinical practices, among them fibrates and thiazolidinediones, PPAR α and γ agonists, used in the treatment of hypertriglyceridemia and diabetes mellitus, respectively (for review, see [159]). In addition, PPARs ligands [160] decrease A β burden, tau phosphorylation and inflammation and improve behavior in AD mouse models [161,161].

Due to their overlapping expression in all brain cell types from mouse and human and given that they share similarities in their ligand-binding domains [162,163], a tight interconnection between PPARs isoforms was described a couple of years ago [164,165]. The mutual interactions observed between PPAR α , β/δ and γ lead to the concept of a “PPAR triad” in the brain (reviewed in [166]). This concept emerged from data reporting that the activation of a PPAR isoform affects the expression of other PPARs due to the low isotype specificity of endogenous PPAR ligands [166,167]. Indeed, the simultaneous activation of different PPARs isoforms was first shown in C6 glioma and lipopolysaccharide (LPS)-stimulated astrocytes, in which the activation of PPAR β/δ increases the expression of PPAR γ and to some extent that of PPAR α in a positive feedback loop [164,165]. Moreover, such crosstalk between PPARs was also reported in primary cortical neurons and in ischemic rat brain, where PPAR γ activation stimulates the interleukin-1 receptor antagonist production through the activation of the PPAR β/δ [168]. Conversely, PPAR α agonist reduces the expression of PPAR β/δ in LPS-stimulated primary cultures of astrocytes in a negative feedback loop, leading to the downregulation of the cyclooxygenase (COX)-2 enzyme involved in the synthesis of

the endogenous PPAR agonist prostaglandin [164],[166]. While the activation of PPAR α represses the expression of COX-2, PPAR β/δ activation upregulates the expression of both COX-2 and cytosolic phospholipase A2, producing PUFAs [164],[166] (Figure 2). Therefore, the cross-talk between PPAR isoforms highlights a “PPAR network”, in which the activation of each PPAR participates in the fine-tuning of genes expression. These interconnections between PPARs should be considered to design appropriate therapeutic strategies for neurodegenerative disorders, including AD.

5. PPARs in Alzheimer’s Disease Therapy: The Promising Role of PPAR α

5.1. PPAR γ and PPAR β/δ in AD

Considering that AD and metabolic diseases such as obesity and type 2 diabetes have overlapping metabolic dysfunctions (e.g., dyslipidemia, glucose metabolism impairment and insulin resistance) and given that PPARs metabolic regulators are expressed in the brain, it is not surprising that changes in PPARs signaling might lead to dementia [169–172].

Since PPAR γ and PPAR β/δ regulate both lipid and carbohydrate metabolism and insulin sensitivity, these receptors represent an attractive therapeutic target for AD. The reduction in glucose metabolic rates observed in the AD brain occurs decades before onset of clinical symptoms and supports the idea that metabolic deficits are upstream events, which may influence the course of AD [173]. As a defining feature of AD, brain glucose hypometabolism leads to a decrease in the O-GlcNAcylation (O-GlcNAc) of proteins, including both tau and APP. While an increase in brain O-GlcNAc protects against tau and A β peptide toxicity, a decrease in O-GlcNAc promotes neurodegeneration [174].

Moreover, brain insulin resistance promotes AD pathophysiology by disrupting energy homeostasis and insulin signaling pathways [175,176]. Impairment in insulin signaling favors A β -mediated oxidative stress, A β secretion, brain amyloid deposition and tau pathology (reviewed in [177–179]). Therefore, targeting PPAR β/δ and PPAR γ with specific drugs represents an effective strategy to preserve carbohydrate metabolism, insulin-sensitizing pathways and cognitive performance. By far, PPAR γ was first considered as a promising target for the treatment of AD. While the thiazolidinedione class of PPAR γ agonists has shown improvement in cognitive behavior in murine models of AD [161,180,181], human clinical trials using PPAR γ agonists are less encouraging [182,183]. Although the chronic treatment of diabetic patients with the PPAR γ agonist pioglitazone reduces dementia risk by 47% [184], Takeda and U.S. partner Zinfandel Pharmaceuticals decided to give up and stop testing a 20-year-old diabetes medicine that fails once more in AD therapy, a lack of success attributed to the low penetrance of glitazones in the brain.

In contrast to PPAR γ , PPAR β/δ is highly expressed throughout the brain and therefore represents a new therapeutic target of interest in AD [185]. Indeed, treatments using PPAR β/δ agonists have been reported to decrease brain neuroinflammation, neurodegeneration, amyloid burden and improve cognitive function in several AD mouse and rat models [186–189]. Recently, a Phase IIa clinical study of the dual PPAR δ and γ agonist T3D-959 reports plasma metabolome profile changes on lipid, glucose and insulin-related metabolism and improvements of cognitive function (presumably associated with APOE genotype) in a small cohort of patients with mild to moderate AD [190].

5.2. PPAR α in AD

Although the function of PPAR α in the brain remained elusive for a long time, more and more studies indicate that PPAR α is involved in physiological and pathological brain functions (e.g., in the sleep-wake cycle [191,192], depression [193–196], epilepsy [197–199], stroke [200–203] and schizophrenia [204]). PPAR α modulators (e.g., oleoylethanolamide, a natural PPAR α ligand; Wy14643 and fibrates, two synthetic PPAR α agonists) regulate dopamine and hippocampal brain-derived neurotrophic factor (BDNF) signaling pathways to rescue depression-related behaviors [193,195,196] and nicotinic acetylcholine receptors and endocannabinoid signaling to alleviate epilepsy and schizophrenia-like effects in mice [197,204].

5.2.1. PPAR α Function in Brain Energy Supply

In addition to their anti-inflammatory and potential neuroprotective effects [117,172,205–207], PPARs, in particular PPAR α , are master metabolic regulators of energy homeostasis [141]. Several studies report that PPAR α plays an essential role in maintaining brain energy supply. Ketone bodies, which are derived from FAs oxidation, are mainly produced in the liver during prolonged fasting or starvation and represent a significant alternative source of fuel to compensate for a lack of glucose in the brain [208–210]. The ketogenic diet has been therefore used in the treatment of several neurological diseases, including Parkinson’s and Alzheimer’s diseases, traumatic brain injury and epilepsy (reviewed in [211]). More and more evidence indicates that the ketogenic diet shows benefits in both in vitro and in vivo AD models. Treatment with the ketone body d- β -hydroxybutyrate protects hippocampal neurons from A β toxicity [212] and ketogenic diet decreases brain amyloid pathology in a mouse model of AD [213]. Moreover, the oral administration of the ketogenic compound AC-1202 reduces oxidative stress and inflammation and improves cognitive function in mild to moderate AD patients [214].

5.2.2. PPAR α and Cognitive Function

More recently, an essential role of PPAR α in cognition has emerged. By using a passive-avoidance task, Mazzola et al. reported that memory acquisition is enhanced in rats treated with the PPAR α agonist Wy14643 [215]. Moreover, treatment of mice with the Wy14643 attenuates cognitive impairments induced by scopolamine, a muscarinic acetylcholine receptor antagonist [216]. Consistent with the potential role of PPAR α in cognition and memory, PPAR α -deficient mice have spatial learning and long-term memory deficits [149], indicating that PPAR α is required for normal cognitive function [217]. Roy et al. have shown that PPAR α , and not PPAR γ and PPAR β/δ isoforms, regulates the expression of a set of synaptic-related proteins involved in excitatory neurotransmission, including BDNF, GluN2A and GluN2B subunits containing N-methyl-D-aspartate receptors (NMDARs) and GluA1 subunit containing alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPARs) [149]. In agreement with this, we have recently reported that the absence of PPAR α severely impairs hippocampal long-term potentiation (LTP), which is defined as an activity-dependent enhancement of synaptic strength involved in memory processing [218], and GluA1 expression in male mice [125].

Moreover, Roy et al. identified a PPAR-responsive element in the promoter of genes encoding the cAMP response element-binding (CREB) protein and therefore identified it as a PPAR α target [149]. Interestingly, recent data indicate that RXR activation induces neuronal CREB signaling and increases dendritic complexity and branching of neurons promoting their differentiation and development [219,220]. In addition, activation of RXRs upregulates the expression of synaptic markers and improves cognition in a mouse model of AD [124]. Altogether, these data indicate that effects mediated by RXR activation on the expression of synaptic-related proteins and cognition could be PPAR α -mediated.

It is also interesting to note that Chikahisa et al. recently reported that PPAR α -null mice exhibit an enhancement of fear learning [221]. This enhancement results from an increase in levels of dopamine and its metabolites in the amygdala [221], suggesting that PPAR α is likewise involved in the regulation of emotional memory via the dopamine pathway in the amygdala.

5.2.3. Potential Link between PPAR α and AD

The relevance of a potential beneficial effect of PPAR α for dementia is supported by some studies showing that polymorphisms in *PPARA* gene encoding PPAR α were associated with an increased risk of AD. In 2003, Brune et al. were the first to report an association of the *PPARA* L162V polymorphism with the AD risk [222]. They indicate that this risk is even higher in carriers harboring a polymorphism in *INS* gene encoding insulin [222]. The interaction of *INS* and *PPARA* genes in AD was thereafter investigated by Kölsch and colleagues [223]. In their study, they report an interaction on AD risk between *PPARA* L162V and *INS* in Northern Europeans, in whom A β 42 and pro-

inflammatory cytokines levels were increased in the cerebrospinal fluid (CSF) [223,224]. However, Sjölander et al. later reported a lack of replication of these studies [225]. They did not find significant differences in genotype or allele distributions between AD patients and controls and found no influence of PPAR α variants on CSF markers [225]. Although these conflicting results question the promising role of PPAR α in AD therapy, previous results indicate that expression levels of PPAR α and β/δ are significantly reduced, whereas PPAR γ expression is selectively increased in AD brains [226], indicating that PPARs function is impaired in AD and therefore may contribute to the progression of the disease.

5.2.4. PPAR α Ligands and AD

More and more studies report the beneficial effects of several PPAR α synthetic agonists on cognitive behavior in several AD mouse models. Among them, fibrates (e.g., fenofibrate, bezafibrate, ciprofibrate and gemfibrozil) are a class of lipid-lowering drugs used in the treatment of metabolic syndromes, including hypertriglyceridemia, obesity and type 2 diabetes, which prevents the progression of atherosclerotic lesions, cardiovascular events and non-alcoholic fatty liver disease (reviewed in [227,228]). Among fibrates, fenofibrate has been widely used, but its relatively low efficiency as PPAR α agonist [229,230] leads to the development of pemaflibrate, a more potent and selective agonist for PPAR α [231–233]. Recent results from two Japanese Phase III clinical studies indicate that pemaflibrate improves lipid profiles in patients with type 2 diabetes and hypertriglyceridemia [234] and was useful for dyslipidemia, with a much higher efficacy than fenofibrate [235].

The salutary effects of fibrates on memory have been reported in several preclinical AD models. It was recently demonstrated that administration of the PPAR α activator gemfibrozil decreases amyloid plaque burden, microgliosis and astrogliosis in the hippocampus and cortex of 5XFAD mice [236], a well-characterized transgenic mouse model of AD, in which age-dependent synaptic and cognitive deficits occur [237]. Although a decrease in the expression of PPAR α was observed in the brain of 5XFAD mice [125,236], oral administration of gemfibrozil or pemaflibrate improves spatial learning, memory and hippocampal LTP, respectively, in these mice [125,236].

More recently, Luo et al. reported that amyloid pathology, memory deficits and anxiety were reversed in the APP-PSEN1 Δ E9 mouse model of AD treated with either gemfibrozil or Wy14643 [238]. The effects observed were mediated by a PPAR α -dependent enhancement of autophagosome biogenesis [238].

In addition, the activation of PPAR α with non-conventional ligands such as statins or aspirin, cholesterol-lowering and nonsteroidal anti-inflammatory drugs, improves [239] hippocampal plasticity and memory in 5XFAD but not in 5XFAD/*Ppara*-null mice by mediating the transcriptional activation of BDNF and CREB, respectively [239,240]. Moreover, oral administration of low-dose aspirin decreased amyloid plaque pathology in 5xFAD mice by stimulating PPAR α -mediated lysosomal biogenesis [241].

Overall, these results support the potential for using PPAR α ligands as a promising strategy for the treatment of AD.

Among PPAR α ligands, gemfibrozil has recently been assessed as a possible treatment for AD. Although gemfibrozil has first been tested in a Phase I clinical trial (NCT00966966 [242]) in healthy volunteers to evaluate its safety and absorption (unpublished data), a second early Phase I trial (NCT02045056 [243]) is testing its efficiency to prevent AD by evaluating its ability to increase microRNA107 (mir-107) levels in participants with either no or mild cognitive impairment. It was previously shown that expression of mir-107, a small noncoding RNA involved in the regulation of gene expression [244], is reduced in AD and may accelerate disease progression through the regulation of β -Site amyloid precursor protein-cleaving enzyme 1 (BACE1) [245,246], an endopeptidase that cleaves APP to generate A β [247,248]. Moreover, gemfibrozil-mediated activation of PPAR α has been reported to promote the non-amyloidogenic APP processing [249]. In 2015, Corbett et al. identified PPAR-responsive elements in the promoter of the gene encoding the α -secretase “a disintegrin and metalloproteinase domain-containing protein 10” (ADAM10), a new

PPAR α target [249] (Figure 2). APP cleavage by ADAM10 precludes A β generation and results in the release of a soluble APP α (sAPP α) fragment which exerts neurotrophic and neuroprotective properties involved in the maintenance of dendritic integrity in the hippocampus [250]. Treatment of wild-type mouse hippocampal neurons with gemfibrozil increases sAPP α and decreases A β production [249]. Moreover, the production of brain endogenous A β is increased in PPAR α -deficient mice and exacerbated in 5XFAD/Ppara-null when compared to wild-type and 5xFAD respective littermates [249].

Although PPAR α is indubitably involved in the non-amyloidogenic processing of APP, PPAR γ was previously demonstrated to also regulate A β production by controlling the expression of *Bace1* gene. PPAR γ activation with specific agonists (e.g., thiazolidinediones and non-steroidal anti-inflammatory drugs including ibuprofen) decreases the expression of BACE1 [251], whereas a lack of PPAR γ has an opposite effect in cultured mouse embryonic fibroblasts [252], suggesting that PPAR γ is a repressor of *Bace1*.

5.2.5. PPAR α and Sex

While sexual dimorphisms of PPAR γ agonist rosiglitazone were previously reported on insulin sensitization and glucose in mice [253], most *in vivo* studies have analyzed the potential effects of PPAR α modulators on cognition mainly in male and not in female rodents. We have previously reported a sex-regulated gene dosage effect of PPAR α on synaptic plasticity [125]. PPAR α activation improves synaptic plasticity only in male but not in female 5XFAD mice [125]. These observations were concomitant with a higher expression of PPAR α in brains of males as compared to females [125]. Such differences in PPAR α expression between male and female rodents were previously reported in liver [254], lymphocytes [255] and ischemic brain [202]. Moreover, a sexual dimorphism was also observed in hippocampus-dependent behaviors. Numerous studies have previously reported that the magnitude and maintenance of LTP were larger in males than in females, not only at CA3-CA1 synapses but also in the dentate gyrus-CA3 and temporoammonic-CA1 synapses of the hippocampus [256–261].

The most obvious difference between males and females is sexual hormones. Hormones are known to influence the expression of PPAR α in a sex-specific manner since gonadectomy of male rats decreases PPAR α expression [254]. Estrogens, such as estradiol, are known to improve synaptic plasticity [262,263], and behavior is affected in ovariectomized female rats [264–266]. In humans, cognitive impairments in older women have long been attributed to the decrease in circulating estradiol levels after menopause. Exogenous restitution of this hormone during the perimenopausal period ameliorates such impairments [267,268]. In addition, estrogen replacement therapy in women in a specific time window is associated with reduced incidence of AD (reviewed in [269]). Although no differences in PPAR α expression were reported between men and women in skeletal muscles [270], several studies indicate that human circulating mononuclear and T cells exhibit sex differences driven by the expression of PPAR α and PPAR γ [271,272]. In their study, Zhang et al. showed that the treatment of T cells with androgens increases PPAR α and decreases PPAR γ expression [272].

It is known that women are at a higher risk for AD (two-thirds of those with AD are women). This results partly from differences between men and women in life-expectation and biology (e.g., sex-specific differences in gene expression, hormone levels, brain structure and function and in inflammatory response) (reviewed in [273,274]). Such differences are not exclusively related to AD but are also observed in cardiovascular diseases, metabolic syndromes and diabetes, where postmenopausal women or women with endocrine disorders (e.g., in Polycystic Ovary Syndrome or Primary Ovarian Insufficiency, in which levels of androgens or estrogens are increased or decreased, respectively), are at higher risk to develop these pathologies when compared to non-affected women (reviewed in [275]). This suggests a potential role for estrogens in metabolic function and in particular in brain metabolism. Indeed, impairment in estrogenic regulation of brain glucose metabolism was previously reported during perimenopause [276,277], and brain hypometabolism reported in women in menopausal transition is associated with cognitive dysfunctions [278,279]. This could result from a decrease in the activation of estrogen receptors (members of the superfamily class of nuclear

receptors) in brain areas involved in learning and memory processes, including the prefrontal cortex and hippocampus [280]. Moreover, impairments in the estrogenic regulation of mitochondrial bioenergetics [281] could lead to subsequent oxidative stress, promoting A β accumulation and neuronal dysfunction [282]. Furthermore, sex differences in PPARs expression and function reported in rodent and human brain could result from changes in sex hormones levels and cross-talks between estrogen receptors and PPARs network (reviewed in [283]), suggesting a role for these receptors in sexual dimorphisms observed in metabolism, inflammatory response and brain function. Moreover, sex differences of the plasma and brain lipidome have been mentioned in humans [284–287], supporting a potential role of PPAR α in this aspect.

6. Conclusions

AD is a multifactorial neurodegenerative disorder in which cognitive deficits occurred. AD is influenced by genotype and environmental factors. Among risk factors identified, genomic loci encoding proteins involved in lipid metabolism and altered lipidome of AD brain suggest that brain lipid metabolism is impaired in AD. Moreover, obesity and type 2 diabetes metabolic disorders, identified as AD risk factors, support the essential role of lipid homeostasis in the etiology of AD. Interestingly, PPARs are nuclear transcription factors that govern pathways implicated in the etiology of AD, including lipid metabolism and inflammatory response (Figure 2). Among them, PPAR α involved in the catabolism of FAs plays a crucial role in cognitive brain function. While disease-modifying treatments for AD are seeking to interfere with the pathogenic steps responsible for clinical symptoms, PPARs modulators are a promising target in AD therapy. Among PPARs, PPAR α has a particular interest since it is the only PPAR isoform described to have neuronal functions involved in memory processes.

As the expression of PPARs is modified in the AD brain, the characterization of new synthetic molecules able to activate several PPARs isoforms could be needed for an efficient treatment for AD. Alternative strategies could be therefore to design novel pan-agonists that can simultaneously activate PPAR α , PPAR β and PPAR γ . Their efficiencies were previously demonstrated in preclinical mouse model of AD and therefore deserve further investigation.

Since a sexual dimorphism of PPAR α agonist was observed in mechanisms underlying memory processes *in vivo*, sex differences should be considered in therapy aiming to use PPAR α modulators. In humans, the influence of sex on the incidence, manifestation and treatment of numerous neurological and psychiatric diseases is well documented. Therefore, it is of crucial importance to decipher sex differences in AD, in which complex cognitive and neuropsychiatric symptoms occur, in order to define novel PPAR α sex-specific therapeutic strategies.

Author Contributions: F.S.-O. and N.P. designed and wrote the manuscript. J.-N.O. critically evaluated and approved the review. All authors have read and agreed to the published version of the manuscript.

Funding: This review received no external funding.

Acknowledgments: Research in the area addressed in this review was supported by the Fondation Louvain, the Belgian Fonds pour la Recherche Scientifique, Interuniversity Attraction Poles Program-Belgian State-Belgian Science Policy, The Belgian Fonds de la Recherche Scientifique Médicale, the Queen Elisabeth Medical Foundation and the Fondation pour la Recherche sur la Maladie d’Alzheimer, CONICYT Becas Chile Program.

Conflicts of Interest: The authors declare no conflict of interest related to the material presented in this review.

References

1. Alzheimer’s Disease International. *World Alzheimer Report 2019, Attitudes to Dementia*; Alzheimer’s Disease International: London, UK, 2019.
2. Feldman, H.H.; Jacova, C.; Robillard, A.; Garcia, A.; Chow, T.; Borrie, M.; Schipper, H.M.; Blair, M.; Kertesz, A.; Chertkow, H. Diagnosis and treatment of dementia: 2. Diagnosis. *Cmaj: Can. Med. Assoc. J.* **2008**, *178*, 825–836, doi:10.1503/cmaj.070798.

3. Kuslansky, G.; Katz, M.; Verghese, J.; Hall, C.B.; Lapuerta, P.; LaRuffa, G.; Lipton, R.B. Detecting dementia with the Hopkins Verbal Learning Test and the Mini-Mental State Examination. *Arch. Clin. Neuropsychol.* **2004**, *19*, 89–104, doi:10.1016/S0887-6177(02)00217-2.
4. Parnetti, L.; Lanari, A.; Silvestrelli, G.; Saggese, E.; Rebaldi, P. Diagnosing prodromal Alzheimer's disease: Role of CSF biochemical markers. *Mech. Ageing Dev.* **2006**, *127*, 129–132, doi:10.1016/j.mad.2005.09.022.
5. Marcus, C.; Mena, E.; Subramaniam, R.M. Brain PET in the diagnosis of Alzheimer's disease. *Clin Nucl Med* **2014**, *39*, e413–e426, doi:10.1097/RNU.0000000000000547.
6. Iaccarino, L.; Sala, A.; Caminiti, S.; Perani, D. The emerging role of PET imaging in dementia [version 1; peer review: 3 approved]. *F1000 Res.* **2017**, *6*, doi:10.12688/f1000research.11603.1.
7. Villemagne, V.L.; Doré, V.; Burnham, S.C.; Masters, C.L.; Rowe, C.C. Imaging tau and amyloid- β proteinopathies in Alzheimer disease and other conditions. *Nat. Rev. Neurol.* **2018**, *14*, 225–236, doi:10.1038/nrneurol.2018.9.
8. Hardy, J.; Allsop, D. Amyloid deposition as the central event in the aetiology of Alzheimer's disease. *Trends Pharmacol. Sci.* **1991**, *12*, 383–388, doi:10.1016/0165-6147(91)90609-V.
9. Selkoe, D.J.; Hardy, J. The amyloid hypothesis of Alzheimer's disease at 25 years. *Embo Mol. Med.* **2016**, *8*, 595–608, doi:10.15252/emmm.201606210.
10. Bergmans, B.A.; De Strooper, B. γ -secretases: From cell biology to therapeutic strategies. *Lancet Neurol.* **2010**, *9*, 215–226, doi:10.1016/S1474-4422(09)70332-1.
11. Hillen, H. The Beta Amyloid Dysfunction (BAD) Hypothesis for Alzheimer's Disease. *Front. Neurosci.* **2019**, *13*, doi:10.3389/fnins.2019.01154.
12. Castellani, R.J.; Plascencia-Villa, G.; Perry, G. The amyloid cascade and Alzheimer's disease therapeutics: Theory versus observation. *Lab. Investig.* **2019**, *99*, 958–970, doi:10.1038/s41374-019-0231-z.
13. Sun, L.; Zhou, R.; Yang, G.; Shi, Y. Analysis of 138 pathogenic mutations in presenilin-1 on the in vitro production of A β 42 and A β 40 peptides by γ -secretase. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E476–E485, doi:10.1073/pnas.1618657114.
14. Ben-Gedalya, T.; Moll, L.; Bejerano-Sagie, M.; Frere, S.; Cabral, W.A.; Friedmann-Morvinski, D.; Slutsky, I.; Burstyn-Cohen, T.; Marini, J.C.; Cohen, E. Alzheimer's disease-causing proline substitutions lead to presenilin 1 aggregation and malfunction. *Embo J.* **2015**, *34*, 2820–2839, doi:10.15252/embj.201592042.
15. Xia, D.; Watanabe, H.; Wu, B.; Lee, Sang H.; Li, Y.; Tsvetkov, E.; Bolshakov, Vadim Y.; Shen, J.; Kelleher, Raymond J. Presenilin-1 Knockin Mice Reveal Loss-of-Function Mechanism for Familial Alzheimer's Disease. *Neuron* **2015**, *85*, 967–981, doi:10.1016/j.neuron.2015.02.010.
16. Szaruga, M.; Veugelen, S.; Benurwar, M.; Lismont, S.; Sepulveda-Falla, D.; Lleo, A.; Ryan, N.S.; Lashley, T.; Fox, N.C.; Murayama, S.; et al. Qualitative changes in human γ -secretase underlie familial Alzheimer's disease. *J Exp Med* **2015**, *212*, 2003–2013, doi:10.1084/jem.20150892.
17. Lott, I.T.; Head, E. Dementia in Down syndrome: Unique insights for Alzheimer disease research. *Nat. Rev. Neurol.* **2019**, *15*, 135–147, doi:10.1038/s41582-018-0132-6.
18. Gong, C.-X.; Liu, F.; Iqbal, K. Multifactorial Hypothesis and Multi-Targets for Alzheimer's Disease. *J. Alzheimer's Dis.* **2018**, *64*, S107–S117, doi:10.3233/JAD-179921.
19. Nelson, P.T.; Dickson, D.W.; Trojanowski, J.Q.; Jack, C.R.; Boyle, P.A.; Arfanakis, K.; Rademakers, R.; Alafuzoff, I.; Attems, J.; Brayne, C.; et al. Limbic-predominant age-related TDP-43 encephalopathy (LATE): Consensus working group report. *Brain* **2019**, *142*, 1503–1527, doi:10.1093/brain/awz099.
20. Goure, W.F.; Krafft, G.A.; Jerecic, J.; Hefti, F. Targeting the proper amyloid-beta neuronal toxins: A path forward for Alzheimer's disease immunotherapy. *Alzheimer's Res. Ther.* **2014**, *6*, 42, doi:10.1186/alzrt272.
21. Walsh, D.M.; Tseng, B.P.; Rydel, R.E.; Podlisny, M.B.; Selkoe, D.J. The Oligomerization of Amyloid β -Protein Begins Intracellularly in Cells Derived from Human Brain. *Biochemistry* **2000**, *39*, 10831–10839, doi:10.1021/bi001048s.
22. Chen, Y.-R.; Glabe, C.G. Distinct Early Folding and Aggregation Properties of Alzheimer Amyloid- β Peptides A β 40 and A β 42: STABLE TRIMER OR TETRAMER FORMATION BY A β 42. *J. Biol. Chem.* **2006**, *281*, 24414–24422, doi:10.1074/jbc.M602363200.
23. Ahmed, M.; Davis, J.; Aucoin, D.; Sato, T.; Ahuja, S.; Aimoto, S.; Elliott, J.I.; Van Nostrand, W.E.; Smith, S.O. Structural conversion of neurotoxic amyloid- β 1–42 oligomers to fibrils. *Nat. Struct. Mol. Biol.* **2010**, *17*, 561–567, doi:10.1038/nsmb.1799.

24. Shankar, G.M.; Li, S.; Mehta, T.H.; Garcia-Munoz, A.; Shepardson, N.E.; Smith, I.; Brett, F.M.; Farrell, M.A.; Rowan, M.J.; Lemere, C.A.; et al. Amyloid- β protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat. Med.* **2008**, *14*, 837–842, doi:10.1038/nm1782.
25. Lesné, S.E.; Sherman, M.A.; Grant, M.; Kuskowski, M.; Schneider, J.A.; Bennett, D.A.; Ashe, K.H. Brain amyloid- β oligomers in ageing and Alzheimer's disease. *Brain* **2013**, *136*, 1383–1398, doi:10.1093/brain/awt062.
26. Kayed, R.; Head, E.; Thompson, J.L.; McIntire, T.M.; Milton, S.C.; Cotman, C.W.; Glabe, C.G. Common Structure of Soluble Amyloid Oligomers Implies Common Mechanism of Pathogenesis. *Science* **2003**, *300*, 486–489, doi:10.1126/science.1079469.
27. Cohen, E.; Paulsson, J.F.; Blinder, P.; Burstyn-Cohen, T.; Du, D.; Estepa, G.; Adame, A.; Pham, H.M.; Holzenberger, M.; Kelly, J.W.; et al. Reduced IGF-1 Signaling Delays Age-Associated Proteotoxicity in Mice. *Cell* **2009**, *139*, 1157–1169, doi:10.1016/j.cell.2009.11.014.
28. O'Nuallain, B.; Freir, D.B.; Nicoll, A.J.; Risse, E.; Ferguson, N.; Herron, C.E.; Collinge, J.; Walsh, D.M. Amyloid β -Protein Dimers Rapidly Form Stable Synaptotoxic Protofibrils. *J. Neurosci.* **2010**, *30*, 14411–14419, doi:10.1523/jneurosci.3537-10.2010.
29. Canter, R.G.; Penney, J.; Tsai, L.-H. The road to restoring neural circuits for the treatment of Alzheimer's disease. *Nature* **2016**, *539*, 187–196, doi:10.1038/nature20412.
30. Eisai. Eisai And Biogen To Discontinue Phase III Clinical Studies Of BACE Inhibitor Elenbecestat In Early Alzheimer's Disease. Available online: <http://eisai.mediaroom.com/2019-09-13-Eisai-And-Biogen-To-Discontinue-Phase-III-Clinical-Studies-Of-BACE-Inhibitor-Elenbecestat-In-Early-Alzheimers-Disease> (accessed on 01/04/2020).
31. Liu, P.-P.; Xie, Y.; Meng, X.-Y.; Kang, J.-S. History and progress of hypotheses and clinical trials for Alzheimer's disease. *Signal Transduct Target* **2019**, *4*, 29–29, doi:10.1038/s41392-019-0063-8.
32. Schneider, L. A resurrection of aducanumab for Alzheimer's disease. *Lancet Neurol.* **2020**, *19*, 111–112, doi:10.1016/S1474-4422(19)30480-6.
33. Huang, L.-K.; Chao, S.-P.; Hu, C.-J. Clinical trials of new drugs for Alzheimer disease. *J. Biomed. Sci.* **2020**, *27*, 18, doi:10.1186/s12929-019-0609-7.
34. Yu, J.-T.; Tan, L.; Hardy, J. Apolipoprotein E in Alzheimer's Disease: An Update. *Annu. Rev. Neurosci.* **2014**, *37*, 79–100, doi:10.1146/annurev-neuro-071013-014300.
35. Kunkle, B.W.; Grenier-Boley, B.; Sims, R.; Bis, J.C.; Damotte, V.; Naj, A.C.; Boland, A.; Vronskaya, M.; van der Lee, S.J.; Amlie-Wolf, A.; et al. Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates $\text{A}\beta$, tau, immunity and lipid processing. *Nat. Genet.* **2019**, *51*, 414–430, doi:10.1038/s41588-019-0358-2.
36. Cuyvers, E.; Sleegers, K. Genetic variations underlying Alzheimer's disease: Evidence from genome-wide association studies and beyond. *Lancet Neurol.* **2016**, *15*, 857–868, doi:10.1016/S1474-4422(16)00127-7.
37. De Roeck, A.; Van Broeckhoven, C.; Sleegers, K. The role of ABCA7 in Alzheimer's disease: Evidence from genomics, transcriptomics and methylomics. *Acta Neuropathol.* **2019**, *138*, 201–220, doi:10.1007/s00401-019-01994-1.
38. Harold, D.; Abraham, R.; Hollingworth, P.; Sims, R.; Gerrish, A.; Hamshere, M.L.; Pahwa, J.S.; Moskvina, V.; Dowzell, K.; Williams, A.; et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat. Genet.* **2009**, *41*, 1088–1093, doi:10.1038/ng.440.
39. Hollingworth, P.; Harold, D.; Sims, R.; Gerrish, A.; Lambert, J.-C.; Carrasquillo, M.M.; Abraham, R.; Hamshere, M.L.; Pahwa, J.S.; Moskvina, V.; et al. Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat. Genet.* **2011**, *43*, 429–435, doi:10.1038/ng.803.
40. Lambert, J.-C.; Heath, S.; Even, G.; Campion, D.; Sleegers, K.; Hiltunen, M.; Combarros, O.; Zelenika, D.; Bullido, M.J.; Tavernier, B.; et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat. Genet.* **2009**, *41*, 1094–1099, doi:10.1038/ng.439.
41. Di Paolo, G.; Kim, T.-W. Linking lipids to Alzheimer's disease: Cholesterol and beyond. *Nat. Rev. Neurosci.* **2011**, *12*, 284–296, doi:10.1038/nrn3012.
42. Shimano, H.; Shimomura, I.; Hammer, R.E.; Herz, J.; Goldstein, J.L.; Brown, M.S.; Horton, J.D. Elevated levels of SREBP-2 and cholesterol synthesis in livers of mice homozygous for a targeted disruption of the SREBP-1 gene. *J. Clin. Investig.* **1997**, *100*, 2115–2124, doi:10.1172/JCI119746.

43. Horton, J.D. Sterol regulatory element-binding proteins: Transcriptional activators of lipid synthesis. *Biochem. Soc. Trans.* **2002**, *30*, 1091–1095, doi:10.1042/bst0301091.
44. Chen, Y.; Bang, S.; McMullen, M.F.; Kazi, H.; Talbot, K.; Ho, M.-X.; Carlson, G.; Arnold, S.E.; Ong, W.-Y.; Kim, S.F. Neuronal Activity-Induced Sterol Regulatory Element Binding Protein-1 (SREBP1) is Disrupted in Dysbindin-Null Mice—Potential Link to Cognitive Impairment in Schizophrenia. *Mol. Neurobiol.* **2017**, *54*, 1699–1709, doi:10.1007/s12035-016-9773-x.
45. Picard, C.; Julien, C.; Frappier, J.; Miron, J.; Théroux, L.; Dea, D.; Breitner, J.C.S.; Poirier, J. Alterations in cholesterol metabolism-related genes in sporadic Alzheimer's disease. *Neurobiol. Aging* **2018**, *66*, e181–e189, doi:10.1016/j.neurobiolaging.2018.01.018.
46. Steen, V.M.; Skrede, S.; Polushina, T.; López, M.; Andreassen, O.A.; Fernø, J.; Hellard, S.L. Genetic evidence for a role of the SREBP transcription system and lipid biosynthesis in schizophrenia and antipsychotic treatment. *Eur. Neuropsychopharmacol.* **2017**, *27*, 589–598, doi:10.1016/j.euroneuro.2016.07.011.
47. Wang, C.; Zhao, F.; Shen, K.; Wang, W.; Siedlak, S.L.; Lee, H.-g.; Phelix, C.F.; Perry, G.; Shen, L.; Tang, B.; et al. The sterol regulatory element-binding protein 2 is dysregulated by tau alterations in Alzheimer disease. *Brain Pathol.* **2019**, *29*, 530–543, doi:10.1111/bpa.12691.
48. Pierrot, N.; Tyteca, D.; D'auria, L.; Dewachter, I.; Gailly, P.; Hendrickx, A.; Tasiaux, B.; Haylani, L.E.; Muls, N.; N'Kuli, F.; et al. Amyloid precursor protein controls cholesterol turnover needed for neuronal activity. *Embo Mol. Med.* **2013**, *5*, 608–625, doi:10.1002/emmm.201202215.
49. Cunnane, S.C.; Schneider, J.A.; Tangney, C.; Tremblay-Mercier, J.; Fortier, M.; Bennett, D.A.; Morris, M.C. Plasma and Brain Fatty Acid Profiles in Mild Cognitive Impairment and Alzheimer's Disease. *J. Alzheimer's Dis.* **2012**, *29*, 691–697, doi:10.3233/JAD-2012-110629.
50. Joffre, C.; Nadjar, A.; Lebbadi, M.; Calon, F.; Laye, S. n-3 LCPUFA improves cognition: The young, the old and the sick. *Prostaglandins Leukot. Essent. Fat. Acids* **2014**, *91*, 1–20, doi:10.1016/j.plefa.2014.05.001.
51. Lukiw, W.J.; Pappolla, M.; Pelaez, R.P.; Bazan, N.G. Alzheimer's Disease—A Dysfunction in Cholesterol and Lipid Metabolism. *Cell. Mol. Neurobiol.* **2005**, *25*, 475–483, doi:10.1007/s10571-005-4010-6.
52. Proitsi, P.; Kim, M.; Whiley, L.; Pritchard, M.; Leung, R.; Soininen, H.; Kloszewska, I.; Mecocci, P.; Tsolaki, M.; Vellas, B.; et al. Plasma lipidomics analysis finds long chain cholesteryl esters to be associated with Alzheimer's disease. *Transl. Psychiatry* **2015**, *5*, e494–e494, doi:10.1038/tp.2014.127.
53. Snowden, S.G.; Ebshiana, A.A.; Hye, A.; An, Y.; Pletnikova, O.; O'Brien, R.; Troncoso, J.; Legido-Quigley, C.; Thambisetty, M. Association between fatty acid metabolism in the brain and Alzheimer disease neuropathology and cognitive performance: A nontargeted metabolomic study. *Plos Med.* **2017**, *14*, e1002266, doi:10.1371/journal.pmed.1002266.
54. Amtul, Z.; Uhrig, M.; Wang, L.; Rozmahel, R.F.; Beyreuther, K. Detrimental effects of arachidonic acid and its metabolites in cellular and mouse models of Alzheimer's disease: Structural insight. *Neurobiol. Aging* **2012**, *33*, e821–e831, doi:10.1016/j.neurobiolaging.2011.07.014.
55. Barbero-Camps, E.; Fernández, A.; Martínez, L.; Fernández-Checa, J.C.; Colell, A. APP/PS1 mice overexpressing SREBP-2 exhibit combined A β accumulation and tau pathology underlying Alzheimer's disease. *Hum. Mol. Genet.* **2013**, *22*, 3460–3476, doi:10.1093/hmg/ddt201.
56. Grimm, M.O.W.; Kuchenbecker, J.; Grösgen, S.; Burg, V.K.; Hundsdörfer, B.; Rothhaar, T.L.; Friess, P.; de Wilde, M.C.; Broersen, L.M.; Penke, B.; et al. Docosahexaenoic Acid Reduces Amyloid β Production via Multiple Pleiotropic Mechanisms. *J. Biol. Chem.* **2011**, *286*, 14028–14039, doi:10.1074/jbc.M110.182329.
57. Hamilton, J.A.; Hillard, C.J.; Spector, A.A.; Watkins, P.A. Brain Uptake and Utilization of Fatty Acids, Lipids and Lipoproteins: Application to Neurological Disorders. *J. Mol. Neurosci.* **2007**, *33*, 2–11, doi:10.1007/s12031-007-0060-1.
58. Grimm, M.O.W.; Michaelson, D.M.; Hartmann, T. Omega-3 fatty acids, lipids, and apoE lipidation in Alzheimer's disease: A rationale for multi-nutrient dementia prevention. *J. Lipid Res.* **2017**, *58*, 2083–2101, doi:10.1194/jlr.R076331.
59. Hussain, G.; Wang, J.; Rasul, A.; Anwar, H.; Imran, A.; Qasim, M.; Zafar, S.; Kamran, S.K.S.; Razzaq, A.; Aziz, N.; et al. Role of cholesterol and sphingolipids in brain development and neurological diseases. *Lipids Health Dis.* **2019**, *18*, 26, doi:10.1186/s12944-019-0965-z.
60. Fields, R.D. A new mechanism of nervous system plasticity: Activity-dependent myelination. *Nat. Rev. Neurosci.* **2015**, *16*, 756–767, doi:10.1038/nrn4023.

61. Duncan, A.L.; Song, W.; Sansom, M.S.P. Lipid-Dependent Regulation of Ion Channels and G Protein-Coupled Receptors: Insights from Structures and Simulations. *Annu. Rev. Pharmacol. Toxicol.* **2020**, *60*, 31–50, doi:10.1146/annurev-pharmtox-010919-023411.
62. Cordero-Morales, J.F.; Vásquez, V. How lipids contribute to ion channel function, a fat perspective on direct and indirect interactions. *Curr. Opin. Struct. Biol.* **2018**, *51*, 92–98, doi:10.1016/j.sbi.2018.03.015.
63. Munro, S. Lipid Rafts: Elusive or Illusive? *Cell* **2003**, *115*, 377–388, doi:10.1016/S0092-8674(03)00882-1.
64. Sebastião, A.M.; Colino-Oliveira, M.; Assaife-Lopes, N.; Dias, R.B.; Ribeiro, J.A. Lipid rafts, synaptic transmission and plasticity: Impact in age-related neurodegenerative diseases. *Neuropharmacology* **2013**, *64*, 97–107, doi:10.1016/j.neuropharm.2012.06.053.
65. Martín, V.; Fabelo, N.; Santpere, G.; Puig, B.; Marín, R.; Ferrer, I.; Díaz, M. Lipid Alterations in Lipid Rafts from Alzheimer’s Disease Human Brain Cortex. *J. Alzheimer’s Dis.* **2010**, *19*, 489–502, doi:10.3233/JAD-2010-1242.
66. Raquel, M.; Noemí, F.; Cecilia, F.-E.; Ana, C.-A.; Deiene, R.-B.; David, Q.-A.; Fátima, M.-H.; Mario, D. Lipid Raft Alterations in Aged-Associated Neuropathologies. *Curr. Alzheimer Res.* **2016**, *13*, 973–984, doi:<http://dx.doi.org/10.2174/1567205013666160314150017>.
67. Hicks, D.; Nalivaeva, N.; Turner, A. Lipid Rafts and Alzheimer’s Disease: Protein-Lipid Interactions and Perturbation of Signaling. *Front. Physiol.* **2012**, *3*, doi:10.3389/fphys.2012.00189.
68. Dart, C. SYMPOSIUM REVIEW: Lipid microdomains and the regulation of ion channel function. *J. Physiol.* **2010**, *588*, 3169–3178, doi:10.1113/jphysiol.2010.191585.
69. Barrett, P.J.; Song, Y.; Van Horn, W.D.; Hustedt, E.J.; Schafer, J.M.; Hadziselimovic, A.; Beel, A.J.; Sanders, C.R. The Amyloid Precursor Protein Has a Flexible Transmembrane Domain and Binds Cholesterol. *Science* **2012**, *336*, 1168–1171, doi:10.1126/science.1219988.
70. Coburger, I.; Hoefgen, S.; Than, M.E. The structural biology of the amyloid precursor protein APP – a complex puzzle reveals its multi-domain architecture. **2014**, *395*, 485, doi:10.1515/hsz-2013-0280.
71. van der Kant, R.; Goldstein, Lawrence S.B. Cellular Functions of the Amyloid Precursor Protein from Development to Dementia. *Dev. Cell* **2015**, *32*, 502–515, doi:10.1016/j.devcel.2015.01.022.
72. Müller, U.C.; Deller, T.; Korte, M. Not just amyloid: Physiological functions of the amyloid precursor protein family. *Nat. Rev. Neurosci.* **2017**, *18*, 281–298, doi:10.1038/nrn.2017.29.
73. Ludewig, S.; Korte, M. Novel Insights into the Physiological Function of the APP (Gene) Family and Its Proteolytic Fragments in Synaptic Plasticity. *Front. Mol. Neurosci.* **2017**, *9*, doi:10.3389/fnmol.2016.00161.
74. Grimm, M.O.W.; Mett, J.; Grimm, H.S.; Hartmann, T. APP Function and Lipids: A Bidirectional Link. *Front. Mol. Neurosci.* **2017**, *10*, doi:10.3389/fnmol.2017.00063.
75. Hefter, D.; Ludewig, S.; Draguhn, A.; Korte, M. Amyloid, APP, and Electrical Activity of the Brain. *Neuroscientist* **2020**, *26*, 231–251, doi:10.1177/1073858419882619.
76. DelBove, C.E.; Strothman, C.E.; Lazarenko, R.M.; Huang, H.; Sanders, C.R.; Zhang, Q. Reciprocal modulation between amyloid precursor protein and synaptic membrane cholesterol revealed by live cell imaging. *Neurobiol. Dis.* **2019**, *127*, 449–461, doi:10.1016/j.nbd.2019.03.009.
77. Area-Gomez, E.; Schon, E.A. Mitochondria-associated ER membranes and Alzheimer disease. *Curr. Opin. Genet. Dev.* **2016**, *38*, 90–96, doi:10.1016/j.gde.2016.04.006.
78. Janikiewicz, J.; Szymański, J.; Malinska, D.; Patalas-Krawczyk, P.; Michalska, B.; Duszyński, J.; Giorgi, C.; Bonora, M.; Dobrzyn, A.; Wieckowski, M.R. Mitochondria-associated membranes in aging and senescence: Structure, function, and dynamics. *Cell Death Dis.* **2018**, *9*, 332, doi:10.1038/s41419-017-0105-5.
79. Area-Gomez, E.; de Groof, A.J.C.; Boldogh, I.; Bird, T.D.; Gibson, G.E.; Koehler, C.M.; Yu, W.H.; Duff, K.E.; Yaffe, M.P.; Pon, L.A.; et al. Presenilins Are Enriched in Endoplasmic Reticulum Membranes Associated with Mitochondria. *Am. J. Pathol.* **2009**, *175*, 1810–1816, doi:10.2353/ajpath.2009.090219.
80. Pera, M.; Larrea, D.; Guardia-Laguarda, C.; Montesinos, J.; Velasco, K.R.; Agrawal, R.R.; Xu, Y.; Chan, R.B.; Di Paolo, G.; Mehler, M.F.; et al. Increased localization of APP-C99 in mitochondria-associated ER membranes causes mitochondrial dysfunction in Alzheimer disease. *Embo J.* **2017**, *36*, 3356–3371, doi:10.1525/embj.201796797.
81. Hedskog, L.; Pinho, C.M.; Filadi, R.; Rönnbäck, A.; Hertwig, L.; Wiegner, B.; Larssen, P.; Gellhaar, S.; Sandebring, A.; Westerlund, M.; et al. Modulation of the endoplasmic reticulum–mitochondria interface in Alzheimer’s disease and related models. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 7916–7921, doi:10.1073/pnas.1300677110.

82. Area-Gomez, E.; del Carmen Lara Castillo, M.; Tambini, M.D.; Guardia-Laguarta, C.; de Groof, A.J.C.; Madra, M.; Ikenouchi, J.; Umeda, M.; Bird, T.D.; Sturley, S.L.; et al. Upregulated function of mitochondria-associated ER membranes in Alzheimer disease. *Embo J.* **2012**, *31*, 4106–4123, doi:10.1038/emboj.2012.202.
83. Area-Gomez, E.; de Groof, A.; Bonilla, E.; Montesinos, J.; Tanji, K.; Boldogh, I.; Pon, L.; Schon, E.A. A key role for MAM in mediating mitochondrial dysfunction in Alzheimer disease. *Cell Death Dis.* **2018**, *9*, 335, doi:10.1038/s41419-017-0215-0.
84. Area-Gomez, E.; Schon, E.A. On the Pathogenesis of Alzheimer's Disease: The MAM Hypothesis. *Faseb J.* **2017**, *31*, 864–867, doi:10.1096/fj.201601309.
85. McMahon, H.T.; Boucrot, E. Membrane curvature at a glance. *J. Cell Sci.* **2015**, *128*, 1065–1070, doi:10.1242/jcs.114454.
86. Simons, K.; Toomre, D. Lipid rafts and signal transduction. *Nat. Rev. Mol. Cell Biol.* **2000**, *1*, 31–39, doi:10.1038/35036052.
87. Srivastav, R.K.; Ansari, T.M.; Prasad, M.; Vishwakarma, V.K.; Bagga, P.; Ahsan, F. Caveolins; An Assailant or An Ally of Various Cellular Disorders. *Drug Res (Stuttg)* **2019**, *69*, 419–427, doi:10.1055/a-0850-6518.
88. Chauhan, N.B. Membrane dynamics, cholesterol homeostasis, and Alzheimer's disease. *J. Lipid Res.* **2003**, *44*, 2019–2029, doi:10.1194/jlr.R300010-JLR200.
89. Gaudreault, S.B.; Dea, D.; Poirier, J. Increased caveolin-1 expression in Alzheimer's disease brain. *Neurobiol. Aging* **2004**, *25*, 753–759, doi:10.1016/j.neurobiolaging.2003.07.004.
90. Alsaqati, M.; Thomas, R.S.; Kidd, E.J. Proteins Involved in Endocytosis Are Upregulated by Ageing in the Normal Human Brain: Implications for the Development of Alzheimer's Disease. *J. Gerontol. : Ser. A* **2017**, *73*, 289–298, doi:10.1093/gerona/glx135.
91. Cameron, P.L.; Ruffin, J.W.; Bollag, R.; Rasmussen, H.; Cameron, R.S. Identification of Caveolin and Caveolin-Related Proteins in the Brain. *J. Neurosci.* **1997**, *17*, 9520–9535, doi:10.1523/jneurosci.17-24-09520.1997.
92. Head, B.P.; Peart, J.N.; Panneerselvam, M.; Yokoyama, T.; Pearn, M.L.; Niesman, I.R.; Bonds, J.A.; Schilling, J.M.; Miyano, A.; Headrick, J.; et al. Loss of Caveolin-1 Accelerates Neurodegeneration and Aging. *PLoS ONE* **2010**, *5*, e15697, doi:10.1371/journal.pone.0015697.
93. Chang, C.-Y.; Ke, D.-S.; Chen, J.-Y. Essential Fatty Acids and Human Brain. *Acta Neurol. Taiwanica* **2009**, *18*, 231–241.
94. Bazinet, R.P.; Layé, S. Polyunsaturated fatty acids and their metabolites in brain function and disease. *Nat. Rev. Neurosci.* **2014**, *15*, 771–785, doi:10.1038/nrn3820.
95. Tracey, T.J.; Steyn, F.J.; Wolvetang, E.J.; Ngo, S.T. Neuronal Lipid Metabolism: Multiple Pathways Driving Functional Outcomes in Health and Disease. *Front. Mol. Neurosci.* **2018**, *11*, doi:10.3389/fnmol.2018.00010.
96. Moore, S.A. Polyunsaturated fatty acid synthesis and release by brain-derived cells in vitro. *J. Mol. Neurosci.* **2001**, *16*, 195–200, doi:10.1385/JMN:16:2-3:195.
97. Luchtman, D.W.; Song, C. Cognitive enhancement by omega-3 fatty acids from child-hood to old age: Findings from animal and clinical studies. *Neuropharmacology* **2013**, *64*, 550–565, doi:10.1016/j.neuropharm.2012.07.019.
98. Hussain, G.; Schmitt, F.; Loeffler, J.-P.; Gonzalez De Aguilar, J. Fattening the brain: A brief of recent research. *Front. Cell. Neurosci.* **2013**, *7*, doi:10.3389/fncel.2013.00144.
99. Conquer, J.A.; Tierney, M.C.; Zecevic, J.; Bettger, W.J.; Fisher, R.H. Fatty acid analysis of blood plasma of patients with alzheimer's disease, other types of dementia, and cognitive impairment. *Lipids* **2000**, *35*, 1305–1312, doi:10.1007/s11745-000-0646-3.
100. Schaefer, E.J.; Bongard, V.; Beiser, A.S.; Lamon-Fava, S.; Robins, S.J.; Au, R.; Tucker, K.L.; Kyle, D.J.; Wilson, P.W.F.; Wolf, P.A. Plasma Phosphatidylcholine Docosahexaenoic Acid Content and Risk of Dementia and Alzheimer Disease: The Framingham Heart Study. *Arch. Neurol.* **2006**, *63*, 1545–1550, doi:10.1001/archneur.63.11.1545.
101. Odermatt, A. The Western-style diet: A major risk factor for impaired kidney function and chronic kidney disease. *Am. J. Physiol. -Ren. Physiol.* **2011**, *301*, F919–F931, doi:10.1152/ajpregnol.00068.2011.
102. Quinn, J.F.; Raman, R.; Thomas, R.G.; Yurko-Mauro, K.; Nelson, E.B.; Van Dyck, C.; Galvin, J.E.; Emond, J.; Jack, C.R.; Weiner, M.; et al. Docosahexaenoic Acid Supplementation and Cognitive Decline in Alzheimer Disease: A Randomized Trial. *JAMA* **2010**, *304*, 1903–1911, doi:10.1001/jama.2010.1510.

103. Mazereeuw, G.; Lanctôt, K.L.; Chau, S.A.; Swardfager, W.; Herrmann, N. Effects of omega-3 fatty acids on cognitive performance: A meta-analysis. *Neurobiol. Aging* **2012**, *33*, e1417–e1429, doi:10.1016/j.neurobiolaging.2011.12.014.
104. Layé, S. Polyunsaturated fatty acids, neuroinflammation and well being. *Prostaglandins Leukot. Essent. Fat. Acids (Plefa)* **2010**, *82*, 295–303, doi:10.1016/j.plefa.2010.02.006.
105. Féart, C.; Peuchant, E.; Letenneur, L.; Samieri, C.; Montagnier, D.; Fourrier-Reglat, A.; Barberger-Gateau, P. Plasma eicosapentaenoic acid is inversely associated with severity of depressive symptomatology in the elderly: Data from the Bordeaux sample of the Three-City Study. *Am. J. Clin. Nutr.* **2008**, *87*, 1156–1162, doi:10.1093/ajcn/87.5.1156.
106. Freund-Levi, Y.; Eriksdotter-Jönsson, M.; Cederholm, T.; Basun, H.; Faxén-Irving, G.; Garlind, A.; Vedin, I.; Vessby, B.; Wahlund, L.-O.; Palmblad, J. ω-3 Fatty Acid Treatment in 174 Patients With Mild to Moderate Alzheimer Disease: OmegAD Study: A Randomized Double-blind Trial. *Arch. Neurol.* **2006**, *63*, 1402–1408, doi:10.1001/archneur.63.10.1402.
107. Calon, F.; Lim, G.P.; Yang, F.; Morihara, T.; Teter, B.; Ubeda, O.; Rostaing, P.; Triller, A.; Salem, N.; Ashe, K.H.; et al. Docosahexaenoic Acid Protects from Dendritic Pathology in an Alzheimer's Disease Mouse Model. *Neuron* **2004**, *43*, 633–645, doi:10.1016/j.neuron.2004.08.013.
108. Calon, F.; Cole, G. Neuroprotective action of omega-3 polyunsaturated fatty acids against neurodegenerative diseases: Evidence from animal studies. *Prostaglandins Leukot. Essent. Fat. Acids* **2007**, *77*, 287–293, doi:10.1016/j.plefa.2007.10.019.
109. Connor, S.; Tenorio, G.; Clandinin, M.T.; Sauvé, Y. DHA supplementation enhances high-frequency, stimulation-induced synaptic transmission in mouse hippocampus. *Appl. Physiol. Nutr. Metab.* **2012**, *37*, 880–887, doi:10.1139/h2012-062.
110. Labrousse, V.F.; Nadjar, A.; Joffre, C.; Costes, L.; Aubert, A.; Grégoire, S.; Bretillon, L.; Layé, S. Short-Term Long Chain Omega3 Diet Protects from Neuroinflammatory Processes and Memory Impairment in Aged Mice. *PLoS ONE* **2012**, *7*, e36861, doi:10.1371/journal.pone.0036861.
111. Hashimoto, M.; Hossain, S.; Shimada, T.; Sugioka, K.; Yamasaki, H.; Fujii, Y.; Ishibashi, Y.; Oka, J.-I.; Shido, O. Docosahexaenoic acid provides protection from impairment of learning ability in Alzheimer's disease model rats. *J. Neurochem.* **2002**, *81*, 1084–1091, doi:10.1046/j.1471-4159.2002.00905.x.
112. Cole, G.M.; Frautschy, S.A. Docosahexaenoic Acid Protects from Amyloid and Dendritic Pathology in an Alzheimer's Disease Mouse Model. *Nutr. Health* **2006**, *18*, 249–259, doi:10.1177/026010600601800307.
113. Kavraal, S.; Oncu, S.K.; Bitikta, S.; Artis, A.S.; Dolu, N.; Gunes, T.; Suer, C. Maternal intake of Omega-3 essential fatty acids improves long term potentiation in the dentate gyrus and Morris water maze performance in rats. *Brain Res.* **2012**, *1482*, 32–39, doi:10.1016/j.brainres.2012.09.002.
114. Evans, Ronald M.; Mangelsdorf, David J. Nuclear Receptors, RXR, and the Big Bang. *Cell* **2014**, *157*, 255–266, doi:10.1016/j.cell.2014.03.012.
115. Warden, A.; Truitt, J.; Merriman, M.; Ponomareva, O.; Jameson, K.; Ferguson, L.B.; Mayfield, R.D.; Harris, R.A. Localization of PPAR isotypes in the adult mouse and human brain. *Sci. Rep.* **2016**, *6*, 27618, doi:10.1038/srep27618.
116. Moutinho, M.; Landreth, G.E. Therapeutic potential of nuclear receptor agonists in Alzheimer's disease. *J. Lipid Res.* **2017**, *58*, 1937–1949, doi:10.1194/jlr.R075556.
117. Zolezzi, J.M.; Santos, M.J.; Bastías-Candia, S.; Pinto, C.; Godoy, J.A.; Inestrosa, N.C. PPARs in the central nervous system: Roles in neurodegeneration and neuroinflammation. *Biol. Rev.* **2017**, *92*, 2046–2069, doi:10.1111/brv.12320.
118. Wolf, G. Is 9-Cis-Retinoic Acid the Endogenous Ligand for the Retinoic Acid-X Receptor? *Nutr. Rev.* **2006**, *64*, 532–538, doi:10.1111/j.1753-4887.2006.tb00186.x.
119. Heyman, R.A.; Mangelsdorf, D.J.; Dyck, J.A.; Stein, R.B.; Eichele, G.; Evans, R.M.; Thaller, C. 9-cis retinoic acid is a high affinity ligand for the retinoid X receptor. *Cell* **1992**, *68*, 397–406, doi:10.1016/0092-8674(92)90479-V.
120. Chitranshi, N.; Dheer, Y.; Kumar, S.; Graham, S.L.; Gupta, V. Molecular docking, dynamics, and pharmacology studies on bexarotene as an agonist of ligand-activated transcription factors, retinoid X receptors. *J. Cell. Biochem.* **2019**, *120*, 11745–11760, doi:10.1002/jcb.28455.
121. Zhang, C.; Duvic, M. Treatment of cutaneous T-cell lymphoma with retinoids. *Dermatol. Ther.* **2006**, *19*, 264–271, doi:10.1111/j.1529-8019.2006.00083.x.

122. Jiang, Q.; Lee, C.Y.D.; Mandrekar, S.; Wilkinson, B.; Cramer, P.; Zelcer, N.; Mann, K.; Lamb, B.; Willson, T.M.; Collins, J.L.; et al. ApoE Promotes the Proteolytic Degradation of A β . *Neuron* **2008**, *58*, 681–693, doi:10.1016/j.neuron.2008.04.010.
123. Cramer, P.E.; Cirrito, J.R.; Wesson, D.W.; Lee, C.Y.D.; Karlo, J.C.; Zinn, A.E.; Casali, B.T.; Restivo, J.L.; Goebel, W.D.; James, M.J.; et al. ApoE-Directed Therapeutics Rapidly Clear β -Amyloid and Reverse Deficits in AD Mouse Models. *Science* **2012**, *335*, 1503–1506, doi:10.1126/science.1217697.
124. Mariani, M.M.; Malm, T.; Lamb, R.; Jay, T.R.; Neilson, L.; Casali, B.; Medarametla, L.; Landreth, G.E. Neuronally-directed effects of RXR activation in a mouse model of Alzheimer’s disease. *Sci. Rep.* **2017**, *7*, 42270, doi:10.1038/srep42270.
125. Pierrot, N.; Ris, L.; Stancu, I.-C.; Doshina, A.; Ribeiro, F.; Tyteca, D.; Baugé, E.; Laloyer, F.; Malong, L.; Schakman, O.; et al. Sex-regulated gene dosage effect of PPAR α on synaptic plasticity. *Life Sci. Alliance* **2019**, *2*, e201800262, doi:10.26508/lسا.201800262.
126. Savage, J.C.; Jay, T.; Goduni, E.; Quigley, C.; Mariani, M.M.; Malm, T.; Ransohoff, R.M.; Lamb, B.T.; Landreth, G.E. Nuclear Receptors License Phagocytosis by Trem2⁺ Myeloid Cells in Mouse Models of Alzheimer’s Disease. *J. Neurosci.* **2015**, *35*, 6532–6543, doi:10.1523/jneurosci.4586-14.2015.
127. Yamanaka, M.; Ishikawa, T.; Griep, A.; Axt, D.; Kummer, M.P.; Heneka, M.T. PPAR γ /RXR α -Induced and CD36-Mediated Microglial Amyloid- β Phagocytosis Results in Cognitive Improvement in Amyloid Precursor Protein/Presenilin 1 Mice. *J. Neurosci.* **2012**, *32*, 17321–17331, doi:10.1523/jneurosci.1569-12.2012.
128. Liang, Y.; Lin, S.; Beyer, T.P.; Zhang, Y.; Wu, X.; Bales, K.R.; DeMattos, R.B.; May, P.C.; Li, S.D.; Jiang, X.-C.; et al. A liver X receptor and retinoid X receptor heterodimer mediates apolipoprotein E expression, secretion and cholesterol homeostasis in astrocytes. *J. Neurochem.* **2004**, *88*, 623–634, doi:10.1111/j.1471-4159.2004.02183.x.
129. Zhao, J.; Fu, Y.; Liu, C.-C.; Shinohara, M.; Nielsen, H.M.; Dong, Q.; Kanekiyo, T.; Bu, G. Retinoic Acid Isomers Facilitate Apolipoprotein E Production and Lipidation in Astrocytes through the Retinoid X Receptor/Retinoic Acid Receptor Pathway. *J. Biol. Chem.* **2014**, *289*, 11282–11292, doi:10.1074/jbc.M113.526095.
130. Pierrot, N.; Lhommel, R.; Quenon, L.; Hanseeuw, B.; Dricot, L.; Sindic, C.; Maloteaux, J.-M.; Octavea, J.-N.; Ivanoiu, A. Targretin Improves Cognitive and Biological Markers in a Patient with Alzheimer’s Disease. *J. Alzheimer’s Dis.* **2016**, *49*, 271–276, doi:10.3233/JAD-150405.
131. Ghosal, K.; Haag, M.; Verghese, P.B.; West, T.; Veenstra, T.; Braunstein, J.B.; Bateman, R.J.; Holtzman, D.M.; Landreth, G.E. A randomized controlled study to evaluate the effect of bexarotene on amyloid- β and apolipoprotein E metabolism in healthy subjects. *Alzheimers Dement (N Y)* **2016**, *2*, 110–120, doi:10.1016/j.trci.2016.06.001.
132. Cummings, J.L.; Zhong, K.; Kinney, J.W.; Heaney, C.; Moll-Tudla, J.; Joshi, A.; Pontecorvo, M.; Devous, M.; Tang, A.; Bena, J. Double-blind, placebo-controlled, proof-of-concept trial of bexarotene in moderate Alzheimer’s disease. *Alzheimer’s Res. Ther.* **2016**, *8*, 4, doi:10.1186/s13195-016-0173-2.
133. George J. Schroepfer, J. Oxysterols: Modulators of Cholesterol Metabolism and Other Processes. *Physiol. Rev.* **2000**, *80*, 361–554, doi:10.1152/physrev.2000.80.1.361.
134. Ma, L.; Nelson, E.R. Oxysterols and nuclear receptors. *Mol. Cell. Endocrinol.* **2019**, *484*, 42–51, doi:10.1016/j.mce.2019.01.016.
135. Bensinger, S.J.; Tontonoz, P. Integration of metabolism and inflammation by lipid-activated nuclear receptors. *Nature* **2008**, *454*, 470–477, doi:10.1038/nature07202.
136. Gan, X.; Kaplan, R.; Menke, J.G.; MacNaul, K.; Chen, Y.; Sparrow, C.P.; Zhou, G.; Wright, S.D.; Cai, T.-Q. Dual Mechanisms of ABCA1 Regulation by Geranylgeranyl Pyrophosphate. *J. Biol. Chem.* **2001**, *276*, 48702–48708, doi:10.1074/jbc.M109402200.
137. Langmann, T.; Liebisch, G.; Moehle, C.; Schifferer, R.; Dayoub, R.; Heiduczek, S.; Grandl, M.; Dada, A.; Schmitz, G. Gene expression profiling identifies retinoids as potent inducers of macrophage lipid efflux. *Biochim. Et Biophys. Acta (Bba) - Mol. Basis Dis.* **2005**, *1740*, 155–161, doi:10.1016/j.bbadiis.2004.11.016.
138. Ou, J.; Tu, H.; Shan, B.; Luk, A.; DeBose-Boyd, R.A.; Bashmakov, Y.; Goldstein, J.L.; Brown, M.S. Unsaturated fatty acids inhibit transcription of the sterol regulatory element-binding protein-1c (SREBP-1c) gene by antagonizing ligand-dependent activation of the LXR. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 6027–6032, doi:10.1073/pnas.111138698.
139. Yoshikawa, T.; Shimano, H.; Yahagi, N.; Ide, T.; Amemiya-Kudo, M.; Matsuzaka, T.; Nakakuki, M.; Tomita, S.; Okazaki, H.; Tamura, Y.; et al. Polyunsaturated Fatty Acids Suppress Sterol Regulatory Element-binding

- Protein 1c Promoter Activity by Inhibition of Liver X Receptor (LXR) Binding to LXR Response Elements. *J. Biol. Chem.* **2002**, *277*, 1705–1711, doi:10.1074/jbc.M105711200.
140. Reddy, J.K.; Rao, M.S. Xenobiotic-Induced Peroxisome Proliferation: Role of Tissue Specificity and Species Differences in Response in the Evaluation of the Implications for Human Health. Berlin, Heidelberg; pp. 43–53.
141. Wang, Y.-X. PPARs: Diverse regulators in energy metabolism and metabolic diseases. *Cell Res.* **2010**, *20*, 124–137, doi:10.1038/cr.2010.13.
142. Michalik, L.; Auwerx, J.; Berger, J.P.; Chatterjee, V.K.; Glass, C.K.; Gonzalez, F.J.; Grimaldi, P.A.; Kadlakova, T.; Lazar, M.A.; O’Rahilly, S.; et al. International Union of Pharmacology. LXI. Peroxisome Proliferator-Activated Receptors. *Pharmacol. Rev.* **2006**, *58*, 726–741, doi:10.1124/pr.58.4.5.
143. Auboeuf, D.; Rieusset, J.; Fajas, L.; Vallier, P.; Frerring, V.; Riou, J.P.; Staels, B.; Auwerx, J.; Laville, M.; Vidal, H. Tissue Distribution and Quantification of the Expression of mRNAs of Peroxisome Proliferator-Activated Receptors and Liver X Receptor- α in Humans: No Alteration in Adipose Tissue of Obese and NIDDM Patients. *Diabetes* **1997**, *46*, 1319–1327, doi:10.2337/diab.46.8.1319.
144. Braissant, O.; Foufelle, F.; Scotto, C.; Dauça, M.; Wahli, W. Differential expression of peroxisome proliferator-activated receptors (PPARs): Tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat. *Endocrinology* **1996**, *137*, 354–366, doi:10.1210/endo.137.1.8536636.
145. Rivera, P.; Arrabal, S.; Vargas, A.; Blanco Calvo, E.; Serrano, A.; Pavon, F.J.; Rodriguez de Fonseca, F.; Suarez, J. Localization of peroxisome proliferator-activated receptor alpha (PPAR α) and N-acyl phosphatidylethanolamine phospholipase D (NAPE-PLD) in cells expressing the Ca $^{2+}$ -binding proteins calbindin, calretinin, and parvalbumin in the adult rat hippocampus. *Front. Neuroanat.* **2014**, *8*, doi:10.3389/fnana.2014.00012.
146. Moreno, S.; Farioli-Vecchioli, S.; Cerù, M.P. Immunolocalization of peroxisome proliferator-activated receptors and retinoid \times receptors in the adult rat CNS. *Neuroscience* **2004**, *123*, 131–145, doi:10.1016/j.neuroscience.2003.08.064.
147. Gofflot, F.; Chartoire, N.; Vasseur, L.; Heikkinen, S.; Dembele, D.; Le Merrer, J.; Auwerx, J. Systematic Gene Expression Mapping Clusters Nuclear Receptors According to Their Function in the Brain. *Cell* **2007**, *131*, 405–418, doi:10.1016/j.cell.2007.09.012.
148. Cullingford, T.E.; Bhakoo, K.; Peuchen, S.; Dolphin, C.T.; Patel, R.; Clark, J.B. Distribution of mRNAs Encoding the Peroxisome Proliferator-Activated Receptor α , β , and γ and the Retinoid X Receptor α , β , and γ in Rat Central Nervous System. *J. Neurochem.* **1998**, *70*, 1366–1375, doi:10.1046/j.1471-4159.1998.70041366.x.
149. Roy, A.; Jana, M.; Corbett, Grant T.; Ramaswamy, S.; Kordower, Jeffrey H.; Gonzalez, Frank J.; Pahan, K. Regulation of Cyclic AMP Response Element Binding and Hippocampal Plasticity-Related Genes by Peroxisome Proliferator-Activated Receptor α . *Cell Rep.* **2013**, *4*, 724–737, doi:10.1016/j.celrep.2013.07.028.
150. Staels, B.; Dallongeville, J.; Auwerx, J.; Schoonjans, K.; Leitersdorf, E.; Fruchart, J.-C. Mechanism of Action of Fibrate on Lipid and Lipoprotein Metabolism. *Circulation* **1998**, *98*, 2088–2093, doi:doi:10.1161/01.CIR.98.19.2088.
151. and, J.K.R.; Hashimoto, T. PEROXISOMAL β -OXIDATION AND PEROXISOME PROLIFERATOR-ACTIVATED RECEPTOR α : An Adaptive Metabolic System. *Annu. Rev. Nutr.* **2001**, *21*, 193–230, doi:10.1146/annurev.nutr.21.1.193.
152. Schreurs, M.; Kuipers, F.; Van Der Leij, F.R. Regulatory enzymes of mitochondrial β -oxidation as targets for treatment of the metabolic syndrome. *Obes. Rev.* **2010**, *11*, 380–388, doi:10.1111/j.1467-789X.2009.00642.x.
153. Chung, K.W.; Lee, E.K.; Lee, M.K.; Oh, G.T.; Yu, B.P.; Chung, H.Y. Impairment of PPAR α and the Fatty Acid Oxidation Pathway Aggravates Renal Fibrosis during Aging. *J. Am. Soc. Nephrol.* **2018**, *29*, 1223–1237, doi:10.1681/asn.2017070802.
154. Semple, R.K.; Chatterjee, V.K.K.; O’Rahilly, S. PPAR γ and human metabolic disease. *J. Clin. Investig.* **2006**, *116*, 581–589, doi:10.1172/JCI28003.
155. Barish, G.D.; Narkar, V.A.; Evans, R.M. PPAR δ : A dagger in the heart of the metabolic syndrome. *J. Clin. Investig.* **2006**, *116*, 590–597, doi:10.1172/JCI27955.
156. Kliewer, S.A.; Sundseth, S.S.; Jones, S.A.; Brown, P.J.; Wisely, G.B.; Koble, C.S.; Devchand, P.; Wahli, W.; Willson, T.M.; Lenhard, J.M.; et al. Fatty acids and eicosanoids regulate gene expression through direct interactions with peroxisome proliferator-activated receptors α and γ . *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 4318–4323, doi:10.1073/pnas.94.9.4318.

157. Desvergne, B. PPARs special issue: Anchoring the present to explore the future. *Biochim. Et Biophys. Acta (Bba) Mol. Cell Biol. Lipids* **2007**, *1771*, 913–914, doi:10.1016/j.bbalip.2007.07.005.
158. Roy, A.; Kundu, M.; Jana, M.; Mishra, R.K.; Yung, Y.; Luan, C.-H.; Gonzalez, F.J.; Pahan, K. Identification and characterization of PPAR α ligands in the hippocampus. *Nat. Chem. Biol.* **2016**, *12*, 1075–1083, doi:10.1038/nchembio.2204.
159. Cheng, H.S.; Tan, W.R.; Low, Z.S.; Marvalim, C.; Lee, J.Y.H.; Tan, N.S. Exploration and Development of PPAR Modulators in Health and Disease: An Update of Clinical Evidence. *Int. J. Mol. Sci.* **2019**, *20*, 5055.
160. Burris, T.P. PPAR α ligands make memories. *Nat. Chem. Biol.* **2016**, *12*, 993–994, doi:10.1038/nchembio.2241.
161. Heneka, M.T.; Reyes-Irisarri, E.; Hüll, M.; Kummer, M.P. Impact and Therapeutic Potential of PPARs in Alzheimer's Disease. *Curr Neuropharmacol* **2011**, *9*, 643–650, doi:10.2174/157015911798376325.
162. Korbecki, J.; Bobiński, R.; Dutka, M. Self-regulation of the inflammatory response by peroxisome proliferator-activated receptors. *Inflamm. Res.* **2019**, *68*, 443–458, doi:10.1007/s00011-019-01231-1.
163. Marco, F.; Francesca, F.; Maria Paola, C.; Sandra, M. Neuroprotective Properties of Peroxisome Proliferator-Activated Receptor Alpha (PPAR α) and its Lipid Ligands. *Curr. Med. Chem.* **2014**, *21*, 2803–2821, doi:<http://dx.doi.org/10.2174/0929867321666140303143455>.
164. Aleshin, S.; Grabeklis, S.; Hanck, T.; Sergeeva, M.; Reiser, G. Peroxisome Proliferator-Activated Receptor (PPAR)- γ Positively Controls and PPAR α Negatively Controls Cyclooxygenase-2 Expression in Rat Brain Astrocytes through a Convergence on PPAR β/δ via Mutual Control of PPAR Expression Levels. *Mol. Pharmacol.* **2009**, *76*, 414–424, doi:10.1124/mol.109.056010.
165. Leisewitz, A.V.; Urrutia, C.R.; Martinez, G.R.; Loyola, G.; Bronfman, M. A PPARs cross-talk concertedly commits C6 glioma cells to oligodendrocytes and induces enzymes involved in myelin synthesis. *J. Cell. Physiol.* **2008**, *217*, 367–376, doi:10.1002/jcp.21509.
166. Aleshin, S.; Reiser, G. Role of the peroxisome proliferator-activated receptors (PPAR)- α , β/δ and γ triad in regulation of reactive oxygen species signaling in brain. *Biol. Chem.* **2013**, *394*, 1553, doi:10.1515/hsz-2013-0215.
167. Aleshin, S.; Strokin, M.; Sergeeva, M.; Reiser, G. Peroxisome proliferator-activated receptor (PPAR) β/δ , a possible nexus of PPAR α - and PPAR γ -dependent molecular pathways in neurodegenerative diseases: Review and novel hypotheses. *Neurochem. Int.* **2013**, *63*, 322–330, doi:10.1016/j.neuint.2013.06.012.
168. Glatz, T.; Stöck, I.; Nguyen-Ngoc, M.; Gohlke, P.; Herdegen, T.; Culman, J.; Zhao, Y. Peroxisome-proliferator-activated receptors γ and peroxisome-proliferator-activated receptors β/δ and the regulation of interleukin 1 receptor antagonist expression by pioglitazone in ischaemic brain. *J. Hypertens.* **2010**, *28*, 1488–1497, doi:10.1097/HJH.0b013e3283396e4e.
169. Arnold, S.E.; Arvanitakis, Z.; Macauley-Rambach, S.L.; Koenig, A.M.; Wang, H.-Y.; Ahima, R.S.; Craft, S.; Gandy, S.; Buettner, C.; Stoeckel, L.E.; et al. Brain insulin resistance in type 2 diabetes and Alzheimer disease: Concepts and conundrums. *Nat. Rev. Neurol.* **2018**, *14*, 168–181, doi:10.1038/nrneurol.2017.185.
170. Mazon, J.N.; de Mello, A.H.; Ferreira, G.K.; Rezin, G.T. The impact of obesity on neurodegenerative diseases. *Life Sci.* **2017**, *182*, 22–28, doi:10.1016/j.lfs.2017.06.002.
171. Procaccini, C.; Santopaoolo, M.; Faicchia, D.; Colamatteo, A.; Formisano, L.; de Candia, P.; Galgani, M.; De Rosa, V.; Matarese, G. Role of metabolism in neurodegenerative disorders. *Metabolism* **2016**, *65*, 1376–1390, doi:10.1016/j.metabol.2016.05.018.
172. Wójtowicz, S.; Strosznajder, A.K.; Jeżyna, M.; Strosznajder, J.B. The Novel Role of PPAR Alpha in the Brain: Promising Target in Therapy of Alzheimer's Disease and Other Neurodegenerative Disorders. *Neurochem. Res.* **2020**, *45*, 972–988, doi:10.1007/s11064-020-02993-5.
173. Mosconi, L. Glucose metabolism in normal aging and Alzheimer's disease: Methodological and physiological considerations for PET studies. *Clin Transl Imaging* **2013**, *1*, 10.1007/s40336-40013-40026-y, doi:10.1007/s40336-013-0026-y.
174. Gong, C.-X.; Liu, F.; Iqbal, K. O-GlcNAcylation: A regulator of tau pathology and neurodegeneration. *Alzheimer's Dement.* **2016**, *12*, 1078–1089, doi:10.1016/j.jalz.2016.02.011.
175. Kandimalla, R.; Thirumala, V.; Reddy, P.H. Is Alzheimer's disease a Type 3 Diabetes? A critical appraisal. *Biochim. Et Biophys. Acta (Bba) - Mol. Basis Dis.* **2017**, *1863*, 1078–1089, doi:10.1016/j.bbadiis.2016.08.018.
176. de la Monte, S.M.; Wands, J.R. Review of insulin and insulin-like growth factor expression, signaling, and malfunction in the central nervous system: Relevance to Alzheimer's disease. *J. Alzheimer's Dis.* **2005**, *7*, 45–61, doi:10.3233/JAD-2005-7106.

177. De Felice, F.G. Alzheimer's disease and insulin resistance: Translating basic science into clinical applications. *J. Clin. Investig.* **2013**, *123*, 531–539, doi:10.1172/JCI64595.
178. Mullins, R.J.; Diehl, T.C.; Chia, C.W.; Kapogiannis, D. Insulin Resistance as a Link between Amyloid-Beta and Tau Pathologies in Alzheimer's Disease. *Front. Aging Neurosci.* **2017**, *9*, doi:10.3389/fnagi.2017.00118.
179. Gonçalves, R.A.; Wijesekara, N.; Fraser, P.E.; De Felice, F.G. The Link Between Tau and Insulin Signaling: Implications for Alzheimer's Disease and Other Tauopathies. *Front. Cell. Neurosci.* **2019**, *13*, doi:10.3389/fncel.2019.00017.
180. Skerrett, R.; Pellegrino, M.P.; Casali, B.T.; Taraboanta, L.; Landreth, G.E. Combined Liver X Receptor/Peroxisome Proliferator-activated Receptor γ Agonist Treatment Reduces Amyloid β Levels and Improves Behavior in Amyloid Precursor Protein/Presenilin 1 Mice. *J. Biol. Chem.* **2015**, *290*, 21591–21602, doi:10.1074/jbc.M115.652008.
181. Escribano, L.; Simón, A.-M.; Gimeno, E.; Cuadrado-Tejedor, M.; López de Maturana, R.; García-Osta, A.; Ricobaraza, A.; Pérez-Mediavilla, A.; Del Río, J.; Frechilla, D. Rosiglitazone Rescues Memory Impairment in Alzheimer's Transgenic Mice: Mechanisms Involving a Reduced Amyloid and Tau Pathology. *Neuropsychopharmacology* **2010**, *35*, 1593–1604, doi:10.1038/npp.2010.32.
182. Gold, M.; Gold, M.; Alderton, C.; Zvartau-Hind, M.; Egginton, S.; Saunders, A.M.; Irizarry, M.; Craft, S.; Landreth, G.; Linnamägi, Ü.; et al. Rosiglitazone Monotherapy in Mild-to-Moderate Alzheimer's Disease: Results from a Randomized, Double-Blind, Placebo-Controlled Phase III Study. *Dement. Geriatr. Cogn. Disord.* **2010**, *30*, 131–146, doi:10.1159/000318845.
183. Galimberti, D.; Scarpini, E. Pioglitazone for the treatment of Alzheimer's disease. *Expert Opin. Investig. Drugs* **2017**, *26*, 97–101, doi:10.1080/13543784.2017.1265504.
184. Heneka, M.T.; Fink, A.; Doblhammer, G. Effect of pioglitazone medication on the incidence of dementia. *Ann. Neurol.* **2015**, *78*, 284–294, doi:10.1002/ana.24439.
185. Woods, J.W.; Tanen, M.; Figueroa, D.J.; Biswas, C.; Zycband, E.; Moller, D.E.; Austin, C.P.; Berger, J.P. Localization of PPAR δ in murine central nervous system: Expression in oligodendrocytes and neurons. *Brain Res.* **2003**, *975*, 10–21, doi:10.1016/S0006-8993(03)02515-0.
186. Sergey, K.; Jill, C.R.; Douglas, L.F. A PPARdelta Agonist Reduces Amyloid Burden and Brain Inflammation in a Transgenic Mouse Model of Alzheimers Disease. *Curr. Alzheimer Res.* **2009**, *6*, 431–437, doi:<http://dx.doi.org/10.2174/156720509789207949>.
187. Tong, M.; Deochand, C.; Didsbury, J.; de la Monte, S.M. T3D-959: A Multi-Faceted Disease Remedial Drug Candidate for the Treatment of Alzheimer's Disease. *J. Alzheimer's Dis.* **2016**, *51*, 123–138, doi:10.3233/JAD-151013.
188. Reich, D.; Gallucci, G.; Tong, M.; de la Monte, S.M. Therapeutic Advantages of Dual Targeting of PPAR- δ and PPAR- γ in an Experimental Model of Sporadic Alzheimer's Disease. *J Parkinsons Dis Alzheimers Dis* **2018**, *5*, doi:10.13188/2376-922X.1000025.
189. de la Monte, S.M.; Tong, M.; Schiano, I.; Didsbury, J. Improved Brain Insulin/IGF Signaling and Reduced Neuroinflammation with T3D-959 in an Experimental Model of Sporadic Alzheimer's Disease. *J. Alzheimer's Dis.* **2017**, *55*, 849–864, doi:10.3233/JAD-160656.
190. Chamberlain, S.; Gabriel, H.; Strittmatter, W.; Didsbury, J. An Exploratory Phase IIa Study of the PPAR delta/gamma Agonist T3D-959 Assessing Metabolic and Cognitive Function in Subjects with Mild to Moderate Alzheimer's Disease. *J. Alzheimer's Dis.* **2020**, *73*, 1085–1103, doi:10.3233/JAD-190864.
191. Murillo-Rodríguez, E.; Guzmán, K.; Arankowsky-Sandoval, G.; Salas-Crisóstomo, M.; Jiménez-Moreno, R.; Arias-Carrión, O. Evidence that activation of nuclear peroxisome proliferator-activated receptor alpha (PPAR α) modulates sleep homeostasis in rats. *Brain Res. Bull.* **2016**, *127*, 156–163, doi:10.1016/j.brainresbull.2016.09.007.
192. Mijangos-Moreno, S.; Poot-Aké, A.; Guzmán, K.; Arankowsky-Sandoval, G.; Arias-Carrión, O.; Zaldívar-Rae, J.; Sarro-Ramírez, A.; Murillo-Rodríguez, E. Sleep and neurochemical modulation by the nuclear peroxisome proliferator-activated receptor α (PPAR- α) in rat. *Neurosci. Res.* **2016**, *105*, 65–69, doi:10.1016/j.neures.2015.09.005.
193. Jiang, B.; Wang, Y.-J.; Wang, H.; Song, L.; Huang, C.; Zhu, Q.; Wu, F.; Zhang, W. Antidepressant-like effects of fenofibrate in mice via the hippocampal brain-derived neurotrophic factor signalling pathway. *Br. J. Pharmacol.* **2017**, *174*, 177–194, doi:10.1111/bph.13668.

194. Li, M.; Wang, D.; Bi, W.; Jiang, Z.-e.; Piao, R.; Yu, H. Palmitoylethanolamide Exerts Antidepressant-Like Effects in Rats: Involvement of PPAR Pathway in the Hippocampus. *J. Pharmacol. Exp. Ther.* **2019**, *369*, 163–172, doi:10.1124/jpet.118.254524.
195. Scheggi, S.; Melis, M.; De Felice, M.; Aroni, S.; Muntoni, A.L.; Pelliccia, T.; Gambarana, C.; De Montis, M.G.; Pistis, M. PPAR α modulation of mesolimbic dopamine transmission rescues depression-related behaviors. *Neuropharmacology* **2016**, *110*, 251–259, doi:10.1016/j.neuropharm.2016.07.024.
196. Jiang, B.; Huang, C.; Zhu, Q.; Tong, L.-J.; Zhang, W. WY14643 produces anti-depressant-like effects in mice via the BDNF signaling pathway. *Psychopharmacology* **2015**, *232*, 1629–1642, doi:10.1007/s00213-014-3802-0.
197. Puligheddu, M.; Pillolla, G.; Melis, M.; Lecca, S.; Marrosu, F.; De Montis, M.G.; Scheggi, S.; Carta, G.; Murru, E.; Aroni, S.; et al. PPAR-Alpha Agonists as Novel Antiepileptic Drugs: Preclinical Findings. *PLoS ONE* **2013**, *8*, e64541, doi:10.1371/journal.pone.0064541.
198. Citraro, R.; Russo, E.; Leo, A.; Russo, R.; Avagliano, C.; Navarra, M.; Calignano, A.; De Sarro, G. Pharmacokinetic-pharmacodynamic influence of N-palmitoylethanamine, arachidonyl-2'-chloroethylamide and WIN 55,212-2 on the anticonvulsant activity of antiepileptic drugs against audiogenic seizures in DBA/2 mice. *Eur. J. Pharmacol.* **2016**, *791*, 523–534, doi:10.1016/j.ejphar.2016.09.029.
199. Porta, N.; Vallée, L.; Lecointe, C.; Bouchaert, E.; Staels, B.; Bordet, R.; Auvin, S. Fenofibrate, a peroxisome proliferator-activated receptor- α agonist, exerts anticonvulsive properties. *Epilepsia* **2009**, *50*, 943–948, doi:10.1111/j.1528-1167.2008.01901.x.
200. Zhou, Y.; Yang, L.; Ma, A.; Zhang, X.; Li, W.; Yang, W.; Chen, C.; Jin, X. Orally administered oleoylethanolamide protects mice from focal cerebral ischemic injury by activating peroxisome proliferator-activated receptor- α . *Neuropharmacology* **2012**, *63*, 242–249, doi:10.1016/j.neuropharm.2012.03.008.
201. Sun, Y.; Alexander, S.P.H.; Garle, M.J.; Gibson, C.L.; Hewitt, K.; Murphy, S.P.; Kendall, D.A.; Bennett, A.J. Cannabinoid activation of PPAR alpha; a novel neuroprotective mechanism. *Br. J. Pharmacol.* **2007**, *152*, 734–743, doi:10.1038/sj.bjp.0707478.
202. Dotson, A.L.; Wang, J.; Chen, Y.; Manning, D.; Nguyen, H.; Saugstad, J.A.; Offner, H. Sex differences and the role of PPAR alpha in experimental stroke. *Metab. Brain Dis.* **2016**, *31*, 539–547, doi:10.1007/s11011-015-9766-x.
203. Dotson, A.L.; Wang, J.; Liang, J.; Nguyen, H.; Manning, D.; Saugstad, J.A.; Offner, H. Loss of PPAR α perpetuates sex differences in stroke reflected by peripheral immune mechanisms. *Metab. Brain Dis.* **2016**, *31*, 683–692, doi:10.1007/s11011-016-9805-2.
204. Costa, M.; Squassina, A.; Congiu, D.; Chillotti, C.; Niola, P.; Galderisi, S.; Pistis, M.; Del Zompo, M. Investigation of endocannabinoid system genes suggests association between peroxisome proliferator activator receptor- α gene (PPARA) and schizophrenia. *Eur. Neuropsychopharmacol.* **2013**, *23*, 749–759, doi:10.1016/j.euroneuro.2012.07.007.
205. D'Agostino, G.; La Rana, G.; Russo, R.; Sasso, O.; Iacono, A.; Esposito, E.; Raso, G.M.; Cuzzocrea, S.; Lo Verme, J.; Piomelli, D.; et al. Acute Intracerebroventricular Administration of Palmitoylethanolamide, an Endogenous Peroxisome Proliferator-Activated Receptor- α Agonist, Modulates Carrageenan-Induced Paw Edema in Mice. *J. Pharmacol. Exp. Ther.* **2007**, *322*, 1137–1143, doi:10.1124/jpet.107.123265.
206. Cuzzocrea, S.; Mazzon, E.; Di Paola, R.; Peli, A.; Bonato, A.; Britti, D.; Genovese, T.; Muià, C.; Crisafulli, C.; Caputi, A.P. The role of the peroxisome proliferator-activated receptor- α (PPAR- α) in the regulation of acute inflammation. *J. Leukoc. Biol.* **2006**, *79*, 999–1010, doi:10.1189/jlb.0605341.
207. B., D.O.; A., F.; MP., C.; S., M. Targeting PPARalpha in Alzheimer's Disease. *Curr. Alzheimer Res.* **2018**, *15*, 345–354, doi:<http://dx.doi.org/10.2174/1567205014666170505094549>.
208. Kersten, S.; Stienstra, R. The role and regulation of the peroxisome proliferator activated receptor alpha in human liver. *Biochimie* **2017**, *136*, 75–84, doi:10.1016/j.biochi.2016.12.019.
209. Bougarne, N.; Weyers, B.; Desmet, S.J.; Deckers, J.; Ray, D.W.; Staels, B.; De Bosscher, K. Molecular Actions of PPAR α in Lipid Metabolism and Inflammation. *Endocr. Rev.* **2018**, *39*, 760–802, doi:10.1210/er.2018-00064.
210. Cunnane, S.C.; Courchesne-Loyer, A.; Vandenberghe, C.; St-Pierre, V.; Fortier, M.; Hennebelle, M.; Croteau, E.; Bocti, C.; Fulop, T.; Castellano, C.-A. Can Ketones Help Rescue Brain Fuel Supply in Later Life? Implications for Cognitive Health during Aging and the Treatment of Alzheimer's Disease. *Front. Mol. Neurosci.* **2016**, *9*, doi:10.3389/fnmol.2016.00053.
211. Yang, H.; Shan, W.; Zhu, F.; Wu, J.; Wang, Q. Ketone Bodies in Neurological Diseases: Focus on Neuroprotection and Underlying Mechanisms. *Front. Neurol.* **2019**, *10*, doi:10.3389/fneur.2019.00585.

212. Kashiwaya, Y.; Takeshima, T.; Mori, N.; Nakashima, K.; Clarke, K.; Veech, R.L. D- β -Hydroxybutyrate protects neurons in models of Alzheimer's and Parkinson's disease. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 5440–5444, doi:10.1073/pnas.97.10.5440.
213. Yao, J.; Chen, S.; Mao, Z.; Cadenas, E.; Brinton, R.D. 2-Deoxy-D-Glucose Treatment Induces Ketogenesis, Sustains Mitochondrial Function, and Reduces Pathology in Female Mouse Model of Alzheimer's Disease. *PLoS ONE* **2011**, *6*, e21788, doi:10.1371/journal.pone.0021788.
214. Henderson, S.T.; Vogel, J.L.; Barr, L.J.; Garvin, F.; Jones, J.J.; Costantini, L.C. Study of the ketogenic agent AC-1202 in mild to moderate Alzheimer's disease: A randomized, double-blind, placebo-controlled, multicenter trial. *Nutr. Metab.* **2009**, *6*, 31, doi:10.1186/1743-7075-6-31.
215. Mazzola, C.; Medalie, J.; Scherma, M.; Panlilio, L.V.; Solinas, M.; Tanda, G.; Drago, F.; Cadet, J.L.; Goldberg, S.R.; Yasar, S. Fatty acid amide hydrolase (FAAH) inhibition enhances memory acquisition through activation of PPAR-alpha nuclear receptors. *Learn Mem* **2009**, *16*, 332–337, doi:10.1101/lm.1145209.
216. Xu, H.; You, Z.; Wu, Z.; Zhou, L.; Shen, J.; Gu, Z. WY14643 Attenuates the Scopolamine-Induced Memory Impairments in Mice. *Neurochem. Res.* **2016**, *41*, 2868–2879, doi:10.1007/s11064-016-2002-1.
217. D'Agostino, G.; Cristiano, C.; Lyons, D.J.; Citraro, R.; Russo, E.; Avagliano, C.; Russo, R.; Raso, G.M.; Meli, R.; De Sarro, G.; et al. Peroxisome proliferator-activated receptor alpha plays a crucial role in behavioral repetition and cognitive flexibility in mice. *Mol. Metab.* **2015**, *4*, 528–536, doi:10.1016/j.molmet.2015.04.005.
218. Bliss, T.V.P.; Collingridge, G.L. A synaptic model of memory: Long-term potentiation in the hippocampus. *Nature* **1993**, *361*, 31–39, doi:10.1038/361031a0.
219. Mounier, A.; Georgiev, D.; Nam, K.N.; Fitz, N.F.; Castranio, E.L.; Wolfe, C.M.; Cronican, A.A.; Schug, J.; Lefterov, I.; Koldamova, R. Bexarotene-Activated Retinoid X Receptors Regulate Neuronal Differentiation and Dendritic Complexity. *J. Neurosci.* **2015**, *35*, 11862–11876, doi:10.1523/jneurosci.1001-15.2015.
220. Nam, K.N.; Mounier, A.; Fitz, N.F.; Wolfe, C.; Schug, J.; Lefterov, I.; Koldamova, R. RXR controlled regulatory networks identified in mouse brain counteract deleterious effects of A β oligomers. *Sci. Rep.* **2016**, *6*, 24048, doi:10.1038/srep24048.
221. Chikahisa, S.; Chida, D.; Shiuchi, T.; Harada, S.; Shimizu, N.; Otsuka, A.; Tanioka, D.; Séi, H. Enhancement of fear learning in PPAR α knockout mice. *Behav. Brain Res.* **2019**, *359*, 664–670, doi:10.1016/j.bbr.2018.09.020.
222. Brune, S.; Kölsch, H.; Ptak, U.; Majores, M.; Schulz, A.; Schlosser, R.; Rao, M.L.; Maier, W.; Heun, R. Polymorphism in the peroxisome proliferator-activated receptor α gene influences the risk for Alzheimer's disease. *J. Neural Transm.* **2003**, *110*, 1041–1050, doi:10.1007/s00702-003-0018-6.
223. Kölsch, H.; Lehmann, D.J.; Ibrahim-Verbaas, C.A.; Combarros, O.; van Duijn, C.M.; Hammond, N.; Belbin, O.; Cortina-Borja, M.; Lehmann, M.G.; Aulchenko, Y.S.; et al. Interaction of insulin and PPAR- α genes in Alzheimer's disease: The Epistasis Project. *J. Neural Transm.* **2012**, *119*, 473–479, doi:10.1007/s00702-011-0732-4.
224. Heun, R.; Kölsch, H.; Ibrahim-Verbaas, C.A.; Combarros, O.; Aulchenko, Y.S.; Breteler, M.; Schuur, M.; van Duijn, C.M.; Hammond, N.; Belbin, O.; et al. Interactions between PPAR- α and inflammation-related cytokine genes on the development of Alzheimer's disease, observed by the Epistasis Project. *Int J Mol Epidemiol Genet* **2012**, *3*, 39–47.
225. Sjölander, A.; Minthon, L.; Bogdanovic, N.; Wallin, A.; Zetterberg, H.; Blennow, K. The PPAR- α gene in Alzheimer's disease: Lack of replication of earlier association. *Neurobiol. Aging* **2009**, *30*, 666–668, doi:10.1016/j.neurobiolaging.2007.07.018.
226. de la Monte, S.M.; Wands, J.R. Molecular indices of oxidative stress and mitochondrial dysfunction occur early and often progress with severity of Alzheimer's disease. *J. Alzheimer's Dis.* **2006**, *9*, 167–181, doi:10.3233/JAD-2006-9209.
227. Staels, B.; Maes, M.; Zambon, A. Fibrates and future PPAR α agonists in the treatment of cardiovascular disease. *Nat. Clin. Pract. Cardiovasc. Med.* **2008**, *5*, 542–553, doi:10.1038/ncpcardio1278.
228. Gross, B.; Pawlak, M.; Lefebvre, P.; Staels, B. PPARs in obesity-induced T2DM, dyslipidaemia and NAFLD. *Nat. Rev. Endocrinol.* **2017**, *13*, 36–49, doi:10.1038/nrendo.2016.135.
229. Brown, W.V.; Dujovne, C.A.; Farquhar, J.W.; Feldman, E.B.; Grundy, S.M.; Knopp, R.H.; Lasser, N.L.; Mellies, M.J.; Palmer, R.H.; Samuel, P. Effects of fenofibrate on plasma lipids. Double-blind, multicenter study in patients with type IIA or IIB hyperlipidemia. *Arterioscler. : Off. J. Am. Heart Assoc. Inc.* **1986**, *6*, 670–678, doi:doi:10.1161/01.ATV.6.6.670.
230. Brown, W.V. Focus on Fenofibrate. *Hosp. Pract.* **1988**, *23*, 31–40, doi:10.1080/21548331.1988.11703636.

231. Hennuyer, N.; Duplan, I.; Paquet, C.; Vanhoutte, J.; Woitain, E.; Touche, V.; Colin, S.; Vallez, E.; Lestavel, S.; Lefebvre, P.; et al. The novel selective PPAR α modulator (SPPARM α) pemaflibrate improves dyslipidemia, enhances reverse cholesterol transport and decreases inflammation and atherosclerosis. *Atherosclerosis* **2016**, *249*, 200–208, doi:10.1016/j.atherosclerosis.2016.03.003.
232. Yamazaki, Y.; Abe, K.; Toma, T.; Nishikawa, M.; Ozawa, H.; Okuda, A.; Araki, T.; Oda, S.; Inoue, K.; Shibuya, K.; et al. Design and synthesis of highly potent and selective human peroxisome proliferator-activated receptor α agonists. *Bioorganic Med. Chem. Lett.* **2007**, *17*, 4689–4693, doi:10.1016/j.bmcl.2007.05.066.
233. Fruchart, J.-C. Selective peroxisome proliferator-activated receptor α modulators (SPPARM α): The next generation of peroxisome proliferator-activated receptor α -agonists. *Cardiovasc. Diabetol.* **2013**, *12*, 82, doi:10.1186/1475-2840-12-82.
234. Araki, E.; Yamashita, S.; Arai, H.; Yokote, K.; Satoh, J.; Inoguchi, T.; Nakamura, J.; Maegawa, H.; Yoshioka, N.; Tanizawa, Y.; et al. Effects of Pemaflibrate, a Novel Selective PPAR α Modulator, on Lipid and Glucose Metabolism in Patients With Type 2 Diabetes and Hypertriglyceridemia: A Randomized, Double-Blind, Placebo-Controlled, Phase 3 Trial. *Diabetes Care* **2018**, *41*, 538–546, doi:10.2337/dc17-1589.
235. Ishibashi, S.; Arai, H.; Yokote, K.; Araki, E.; Suganami, H.; Yamashita, S. Efficacy and safety of pemaflibrate (K-877), a selective peroxisome proliferator-activated receptor α modulator, in patients with dyslipidemia: Results from a 24-week, randomized, double blind, active-controlled, phase 3 trial. *J. Clin. Lipidol.* **2018**, *12*, 173–184, doi:10.1016/j.jacl.2017.10.006.
236. Chandra, S.; Roy, A.; Jana, M.; Pahan, K. Cinnamic acid activates PPAR α to stimulate Lysosomal biogenesis and lower Amyloid plaque pathology in an Alzheimer's disease mouse model. *Neurobiol. Dis.* **2019**, *124*, 379–395, doi:10.1016/j.nbd.2018.12.007.
237. Oakley, H.; Cole, S.L.; Logan, S.; Maus, E.; Shao, P.; Craft, J.; Guillozet-Bongaarts, A.; Ohno, M.; Disterhoft, J.; Van Eldik, L.; et al. Intraneuronal β -Amyloid Aggregates, Neurodegeneration, and Neuron Loss in Transgenic Mice with Five Familial Alzheimer's Disease Mutations: Potential Factors in Amyloid Plaque Formation. *J. Neurosci.* **2006**, *26*, 10129–10140, doi:10.1523/jneurosci.1202-06.2006.
238. Luo, R.; Su, L.-Y.; Li, G.; Yang, J.; Liu, Q.; Yang, L.-X.; Zhang, D.-F.; Zhou, H.; Xu, M.; Fan, Y.; et al. Activation of PPARA-mediated autophagy reduces Alzheimer disease-like pathology and cognitive decline in a murine model. *Autophagy* **2020**, *16*, 52–69, doi:10.1080/15548627.2019.1596488.
239. Roy, A.; Jana, M.; Kundu, M.; Corbett, Grant T.; Rangaswamy, Suresh B.; Mishra, Rama K.; Luan, C.-H.; Gonzalez, Frank J.; Pahan, K. HMG-CoA Reductase Inhibitors Bind to PPAR α to Upregulate Neurotrophin Expression in the Brain and Improve Memory in Mice. *Cell Metab.* **2015**, *22*, 253–265, doi:10.1016/j.cmet.2015.05.022.
240. Patel, D.; Roy, A.; Kundu, M.; Jana, M.; Luan, C.-H.; Gonzalez, F.J.; Pahan, K. Aspirin binds to PPAR α to stimulate hippocampal plasticity and protect memory. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E7408–E7417, doi:10.1073/pnas.1802021115.
241. Chandra, S.; Jana, M.; Pahan, K. Aspirin Induces Lysosomal Biogenesis and Attenuates Amyloid Plaque Pathology in a Mouse Model of Alzheimer's Disease via PPAR α . *J. Neurosci.* **2018**, *38*, 6682–6699, doi:10.1523/jneurosci.0054-18.2018.
242. Medicine, N.L.O. Study Evaluating Potential Interaction Between SAM-531 And Gemfibrozil When Co-Administered. Available online: <https://clinicaltrials.gov/ct2/show/NCT00966966> (accessed on 10 April 2020).
243. Medicine, N.L.O. Modulation of Micro-RNA Pathways by Gemfibrozil in Predementia Alzheimer Disease. Available online: <https://clinicaltrials.gov/ct2/show/NCT02045056> (accessed on 10 April 2020).
244. Krol, J.; Loedige, I.; Filipowicz, W. The widespread regulation of microRNA biogenesis, function and decay. *Nat. Rev. Genet.* **2010**, *11*, 597–610, doi:10.1038/nrg2843.
245. Wang, W.-X.; Rajeev, B.W.; Stromberg, A.J.; Ren, N.; Tang, G.; Huang, Q.; Rigoutsos, I.; Nelson, P.T. The Expression of MicroRNA miR-107 Decreases Early in Alzheimer's Disease and May Accelerate Disease Progression through Regulation of β -Site Amyloid Precursor Protein-Cleaving Enzyme 1. *J. Neurosci.* **2008**, *28*, 1213–1223, doi:10.1523/jneurosci.5065-07.2008.
246. Nelson, P.T.; Wang, W.-X. MiR-107 is Reduced in Alzheimer's Disease Brain Neocortex: Validation Study. *J. Alzheimer's Dis.* **2010**, *21*, 75–79, doi:10.3233/JAD-2010-091603.

247. Vassar, R.; Bennett, B.D.; Babu-Khan, S.; Kahn, S.; Mendiaz, E.A.; Denis, P.; Teplow, D.B.; Ross, S.; Amarante, P.; Loeloff, R.; et al. β -Secretase Cleavage of Alzheimer's Amyloid Precursor Protein by the Transmembrane Aspartic Protease BACE. *Science* **1999**, *286*, 735–741, doi:10.1126/science.286.5440.735.
248. Haniu, M.; Denis, P.; Young, Y.; Mendiaz, E.A.; Fuller, J.; Hui, J.O.; Bennett, B.D.; Kahn, S.; Ross, S.; Burgess, T.; et al. Characterization of Alzheimer's β -Secretase Protein BACE: A PEPSIN FAMILY MEMBER WITH UNUSUAL PROPERTIES. *J. Biol. Chem.* **2000**, *275*, 21099–21106, doi:10.1074/jbc.M002095200.
249. Corbett, G.T.; Gonzalez, F.J.; Pahan, K. Activation of peroxisome proliferator-activated receptor α stimulates ADAM10-mediated proteolysis of APP. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 8445–8450, doi:10.1073/pnas.1504890112.
250. Yuan, X.-Z.; Sun, S.; Tan, C.-C.; Yu, J.-T.; Tan, L. The Role of ADAM10 in Alzheimer's Disease. *J. Alzheimer's Dis.* **2017**, *58*, 303–322, doi:10.3233/JAD-170061.
251. Sastre, M.; Dewachter, I.; Landreth, G.E.; Willson, T.M.; Klockgether, T.; van Leuven, F.; Heneka, M.T. Nonsteroidal Anti-Inflammatory Drugs and Peroxisome Proliferator-Activated Receptor- γ Agonists Modulate Immunostimulated Processing of Amyloid Precursor Protein through Regulation of β -Secretase. *J. Neurosci.* **2003**, *23*, 9796–9804, doi:10.1523/jneurosci.23-30-09796.2003.
252. Sastre, M.; Dewachter, I.; Rossner, S.; Bogdanovic, N.; Rosen, E.; Borghgraef, P.; Evert, B.O.; Dumitrescu-Ozimek, L.; Thal, D.R.; Landreth, G.; et al. Nonsteroidal anti-inflammatory drugs repress β -secretase gene promoter activity by the activation of PPAR γ . *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 443–448, doi:10.1073/pnas.0503839103.
253. Duan, S.Z.; Usher, M.G.; Foley, E.L.; Milstone, D.S.; Brosius, F.C.; Mortensen, R.M. Sex dimorphic actions of rosiglitazone in generalised peroxisome proliferator-activated receptor- γ (PPAR- γ)-deficient mice. *Diabetologia* **2010**, *53*, 1493–1505, doi:10.1007/s00125-010-1748-2.
254. Jalouli, M.; Carlsson, L.; Améen, C.; Lindén, D.; Ljungberg, A.; Michalik, L.; Edén, S.; Wahli, W.; Oscarsson, J. Sex Difference in Hepatic Peroxisome Proliferator-Activated Receptor α Expression: Influence of Pituitary and Gonadal Hormones. *Endocrinology* **2003**, *144*, 101–109, doi:10.1210/en.2002-220630.
255. Dunn, S.E.; Ousman, S.S.; Sobel, R.A.; Zuniga, L.; Baranzini, S.E.; Youssef, S.; Crowell, A.; Loh, J.; Oksenberg, J.; Steinman, L. Peroxisome proliferator-activated receptor (PPAR) α expression in T cells mediates gender differences in development of T cell-mediated autoimmunity. *J. Exp. Med.* **2007**, *204*, 321–330, doi:10.1084/jem.20061839.
256. Maren, S.; De Oca, B.; Fanselow, M.S. Sex differences in hippocampal long-term potentiation (LTP) and Pavlovian fear conditioning in rats: Positive correlation between LTP and contextual learning. *Brain Res.* **1994**, *661*, 25–34, doi:10.1016/0006-8993(94)91176-2.
257. Qi, X.; Zhang, K.; Xu, T.; Yamaki, V.N.; Wei, Z.; Huang, M.; Rose, G.M.; Cai, X. Sex Differences in Long-Term Potentiation at Temporoammonic-CA1 Synapses: Potential Implications for Memory Consolidation. *PLoS ONE* **2016**, *11*, e0165891, doi:10.1371/journal.pone.0165891.
258. Monfort, P.; Gomez-Gimenez, B.; Llansola, M.; Felipo, V. Gender Differences in Spatial Learning, Synaptic Activity, and Long-Term Potentiation in the Hippocampus in Rats: Molecular Mechanisms. *Acad. Chem. Neurosci.* **2015**, *6*, 1420–1427, doi:10.1021/acschemneuro.5b00096.
259. Yang, D.-W.; Pan, B.; Han, T.-Z.; Xie, W. Sexual dimorphism in the induction of LTP: Critical role of tetanizing stimulation. *Life Sci.* **2004**, *75*, 119–127, doi:10.1016/j.lfs.2003.12.004.
260. Ris, L.; Angelo, M.; Plattner, F.; Capron, B.; Errington, M.L.; Bliss, T.V.P.; Godaux, E.; Giese, K.P. Sexual dimorphisms in the effect of low-level p25 expression on synaptic plasticity and memory. *Eur. J. Neurosci.* **2005**, *21*, 3023–3033, doi:10.1111/j.1460-9568.2005.04137.x.
261. Harte-Hargrove, L.C.; Varga-Wesson, A.; Duffy, A.M.; Milner, T.A.; Scharfman, H.E. Opioid Receptor-Dependent Sex Differences in Synaptic Plasticity in the Hippocampal Mossy Fiber Pathway of the Adult Rat. *J. Neurosci.* **2015**, *35*, 1723–1738, doi:10.1523/jneurosci.0820-14.2015.
262. Córdoba Montoya, D.A.; Carrer, H.F. Estrogen facilitates induction of long term potentiation in the hippocampus of awake rats1First published on the World Wide Web on 4 November 1997.1. *Brain Res.* **1997**, *778*, 430–438, doi:10.1016/S0006-8993(97)01206-7.
263. Good, M.; Day, M.; Muir, J.L. Cyclical changes in endogenous levels of oestrogen modulate the induction of LTD and LTP in the hippocampal CA1 region. *Eur. J. Neurosci.* **1999**, *11*, 4476–4480, doi:10.1046/j.1460-9568.1999.00920.x.
264. Daniel, J.M.; Lee, C.D. Estrogen replacement in ovariectomized rats affects strategy selection in the Morris water maze. *Neurobiol. Learn. Mem.* **2004**, *82*, 142–149, doi:10.1016/j.nlm.2004.06.001.

265. Korol, D.L.; Malin, E.L.; Borden, K.A.; Busby, R.A.; Couper-Leo, J. Shifts in preferred learning strategy across the estrous cycle in female rats. *Horm. Behav.* **2004**, *45*, 330–338, doi:10.1016/j.yhbeh.2004.01.005.
266. Gupta, R.R.; Sen, S.; Diepenhorst, L.L.; Rudick, C.N.; Maren, S. Estrogen modulates sexually dimorphic contextual fear conditioning and hippocampal long-term potentiation (LTP) in rats¹¹Published on the World Wide Web on 1 December 2000. *Brain Res.* **2001**, *888*, 356–365, doi:10.1016/S0006-8993(00)03116-4.
267. Joffe, H.; Hall, J.E.; Gruber, S.; Sarmiento, I.A.; Cohen, L.S.; Yurgelun-Todd, D.; Martin, K.A. Estrogen therapy selectively enhances prefrontal cognitive processes: A randomized, double-blind, placebo-controlled study with functional magnetic resonance imaging in perimenopausal and recently postmenopausal women. *Menopause* **2006**, *13*, 411–422, doi:10.1097/01.gme.0000189618.48774.7b.
268. Scott, E.; Zhang, Q.-g.; Wang, R.; Vadlamudi, R.; Brann, D. Estrogen neuroprotection and the critical period hypothesis. *Front. Neuroendocrinol.* **2012**, *33*, 85–104, doi:10.1016/j.yfrne.2011.10.001.
269. Merlo, S.; Spampinato, S.F.; Sortino, M.A. Estrogen and Alzheimer's disease: Still an attractive topic despite disappointment from early clinical results. *Eur. J. Pharmacol.* **2017**, *817*, 51–58, doi:10.1016/j.ejphar.2017.05.059.
270. Maher, A.C.; Akhtar, M.; Vockley, J.; Tarnopolsky, M.A. Women Have Higher Protein Content of β -Oxidation Enzymes in Skeletal Muscle than Men. *PLoS ONE* **2010**, *5*, e12025, doi:10.1371/journal.pone.0012025.
271. Wege, N.; Schutkowski, A.; Boenn, M.; Bialek, J.; Schlitt, A.; Noack, F.; Grosse, I.; Stangl, G.I. Men and women differ in their diurnal expression of monocyte peroxisome proliferator-activated receptor- α in the fed but not in the fasted state. *Faseb J.* **2015**, *29*, 2905–2911, doi:10.1096/fj.14-267575.
272. Zhang, M.A.; Rego, D.; Moshkova, M.; Kebir, H.; Chruscinski, A.; Nguyen, H.; Akkermann, R.; Stanczyk, F.Z.; Prat, A.; Steinman, L.; et al. Peroxisome proliferator-activated receptor (PPAR) α and - γ regulate IFN γ and IL-17A production by human T cells in a sex-specific way. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 9505–9510, doi:10.1073/pnas.1118458109.
273. Fisher, D.W.; Bennett, D.A.; Dong, H. Sexual dimorphism in predisposition to Alzheimer's disease. *Neurobiol. Aging* **2018**, *70*, 308–324, doi:10.1016/j.neurobiolaging.2018.04.004.
274. Riedel, B.C.; Thompson, P.M.; Brinton, R.D. Age, APOE and sex: Triad of risk of Alzheimer's disease. *J. Steroid Biochem. Mol. Biol.* **2016**, *160*, 134–147, doi:10.1016/j.jsbmb.2016.03.012.
275. Appelman, Y.; van Rijn, B.B.; ten Haaf, M.E.; Boersma, E.; Peters, S.A.E. Sex differences in cardiovascular risk factors and disease prevention. *Atherosclerosis* **2015**, *241*, 211–218, doi:10.1016/j.atherosclerosis.2015.01.027.
276. Brinton, R.D. Estrogen-induced plasticity from cells to circuits: Predictions for cognitive function. *Trends Pharmacol. Sci.* **2009**, *30*, 212–222, doi:10.1016/j.tips.2008.12.006.
277. Liu, F.; Day, M.; Muñiz, L.C.; Bitran, D.; Arias, R.; Revilla-Sánchez, R.; Grauer, S.; Zhang, G.; Kelley, C.; Pulito, V.; et al. Activation of estrogen receptor- β regulates hippocampal synaptic plasticity and improves memory. *Nat. Neurosci.* **2008**, *11*, 334–343, doi:10.1038/nn2057.
278. Scheyer, O.; Rahman, A.; Hristov, H.; Berkowitz, C.; Isaacson, R.S.; Diaz Brinton, R.; Mosconi, L. Female Sex and Alzheimer's Risk: The Menopause Connection. *J Prev Alzheimers Dis* **2018**, *5*, 225–230, doi:10.14283/jpad.2018.34.
279. Rahman, A.; Jackson, H.; Hristov, H.; Isaacson, R.S.; Saif, N.; Shetty, T.; Etingin, O.; Henchcliffe, C.; Brinton, R.D.; Mosconi, L. Sex and Gender Driven Modifiers of Alzheimer's: The Role for Estrogenic Control Across Age, Race, Medical, and Lifestyle Risks. *Front. Aging Neurosci.* **2019**, *11*, doi:10.3389/fnagi.2019.00315.
280. McEwen, B.S.; Akama, K.T.; Spencer-Segal, J.L.; Milner, T.A.; Waters, E.M. Estrogen effects on the brain: Actions beyond the hypothalamus via novel mechanisms. *Behav. Neurosci.* **2012**, *126*, 4–16, doi:10.1037/a0026708.
281. Yao, J.; Brinton, R.D. Estrogen Regulation of Mitochondrial Bioenergetics: Implications for Prevention of Alzheimer's Disease. In *Advances in Pharmacology*; Michaelis, E.K., Michaelis, M.L., Eds.; Academic Press: 2012; Vol. 64, pp. 327–371.
282. Mattson, M.P.; Magnus, T. Ageing and neuronal vulnerability. *Nat. Rev. Neurosci.* **2006**, *7*, 278–294, doi:10.1038/nrn1886.
283. Benz, V.; Kintscher, U.; Foryst-Ludwig, A. Sex-Specific Differences in Type 2 Diabetes Mellitus and Dyslipidemia Therapy: PPAR Agonists. In *Sex and Gender Differences in Pharmacology*; Regitz-Zagrosek, V., Ed.; Springer Berlin Heidelberg: Berlin, Heidelberg, 2012; pp. 387–410.

284. Wong, M.W.K.; Braidy, N.; Crawford, J.; Pickford, R.; Song, F.; Mather, K.A.; Attia, J.; Brodaty, H.; Sachdev, P.; Poljak, A. APOE Genotype Differentially Modulates Plasma Lipids in Healthy Older Individuals, with Relevance to Brain Health. *J. Alzheimer's Dis.* **2019**, *72*, 703–716, doi:10.3233/JAD-190524.
285. Díaz, M.; Fabelo, N.; Ferrer, I.; Marín, R. “Lipid raft aging” in the human frontal cortex during nonpathological aging: Gender influences and potential implications in Alzheimer’s disease. *Neurobiol. Aging* **2018**, *67*, 42–52, doi:10.1016/j.neurobiolaging.2018.02.022.
286. Mielke, M.M.; Haughey, N.J.; Han, D.; An, Y.; Bandaru, V.V.R.; Lyketsos, C.G.; Ferrucci, L.; Resnick, S.M. The Association Between Plasma Ceramides and Sphingomyelins and Risk of Alzheimer’s Disease Differs by Sex and APOE in the Baltimore Longitudinal Study of Aging. *J. Alzheimer's Dis.* **2017**, *60*, 819–828, doi:10.3233/JAD-160925.
287. Couttas, T.A.; Kain, N.; Tran, C.; Chatterton, Z.; Kwok, J.B.; Don, A.S. Age-Dependent Changes to Sphingolipid Balance in the Human Hippocampus are Gender-Specific and May Sensitize to Neurodegeneration. *J. Alzheimer's Dis.* **2018**, *63*, 503–514, doi:10.3233/JAD-171054.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).