

Table S1. NCBI GEO accession numbers and phenotypes of samples used in correlation studies

Study GEO Accession	Phenotype	Sample
<b>GSE130548</b>	Epithelial	GSM3742048.CEL
<b>GSE130548</b>	Epithelial	GSM3742049.CEL
<b>GSE130548</b>	Epithelial	GSM3742050.CEL
<b>GSE130548</b>	Mesenchymal	GSM3742051.CEL
<b>GSE130548</b>	Mesenchymal	GSM3742052.CEL
<b>GSE130548</b>	Mesenchymal	GSM3742053.CEL
<b>GSE55072</b>	Epithelial	GSM1329077.CEL
<b>GSE55072</b>	Epithelial	GSM1329078.CEL
<b>GSE55072</b>	Mesenchymal	GSM1329081.CEL
<b>GSE55072</b>	Mesenchymal	GSM1329082.CEL
<b>GSE55711</b>	Epithelial	GSM1342162.CEL
<b>GSE55711</b>	Epithelial	GSM1342163.CEL
<b>GSE55711</b>	Mesenchymal	GSM1342164.CEL
<b>GSE55711</b>	Mesenchymal	GSM1342165.CEL
<b>GSE77551</b>	Epithelial	GSM2054383.CEL
<b>GSE77551</b>	Epithelial	GSM2054384.CEL
<b>GSE77551</b>	Epithelial	GSM2054385.CEL
<b>GSE77551</b>	Mesenchymal	GSM2054377.CEL
<b>GSE77551</b>	Mesenchymal	GSM2054378.CEL
<b>GSE77551</b>	Mesenchymal	GSM2054379.CEL

Table S2. Sequence information of siRNAs used.

siRNA	Sequences	Vendor
siElf3_2	TTGAACCAACTTGTTGATAA	Qiagen
siElf3_10	GGUAAUACUACAAACGGGA	Dharmacon
siElf3_12	GGUUGGAGAGAGAGUCGGAU	Dharmacon
siCntrl	Silencer™ Negative Control No. 1 siRNA	Dharmacon
siEhf_1	AACAATTTATGTTTAATGAAA	Qiagen
siEhf_2	ACCCTTGATCTATTTAATCAA	Qiagen
siEhf_3	AAGGAACACTACAGTTGATAA	Qiagen
siEhf_4	CAGCAAATGGATTCTGATCAA	Qiagen

Table S3. Primers and corresponding UPL probes used in qPCR experiments.

Primers	Forward	Reverse	Probe
<b>Ahr</b>	TGCACAAGGAGTGGACGA	AGGAAGCTGGTCTGGGGTAT	27
<b>Cdh1</b>	ATCCTCGCCCTGCTGATT	ACCACCGTTCTCCTCCGTA	18
<b>Cdh2</b>	TCCCTGAGATACAGCGTCACT	ATAATGAAGATGCCCCGTTGG	17
<b>Cebpa</b>	AAACAACGCAACGTGGAGA	GCGGTCATTGTCACTGGTC	67
<b>Ehf</b>	TCATTGTCAAGACTGAACAAACC	GTCCAACAGATCTACTGTGCTACC	33
<b>Elf3</b>	ACCGAACCCTGACACACCT	AGCTGTACATGGCGTTGAAGT	46
<b>Exosc9</b>	CAAGGTGCCCCCTATAGTGCT	GGTCTGAGCTCTTATTTTCTTTGG	106
<b>Fn1</b>	AGGTGGACCCCGCTAAAC	TGCCGCAACTACTGTGATTC	31
<b>Gapdh</b>	AGCTTGTCAATCAACGGGAAG	TTTGATGTTAGTGGGGTCTCG	9
<b>Grhl3</b>	AAGGAAGATGTGAATGAACCTG	TCGTCCTCATTACTGTAGGGAAA	100
<b>Ovol2</b>	GTGAGGATTGCGGCTACAC	TGGTCACTGTTACATGCAG	79
<b>Snai1</b>	CTTGCTCCACAAGCACCA	GAGGATGGGGAGGTAGCAG	71
<b>Snai2</b>	ATCCTTGGGGCGTGTAAGT	TGAACCACTGTGATCCTTGG	6
<b>Zeb1</b>	TGGAGTTCAAAGGTTGTCGTT	TTGCCACATCAACACTGGTC	109
<b>Zeb2</b>	TTGCTCCAGGATGTGTGG	CACACACTTGTTTGTGTGCATATC	64
<b>Cas21</b>	CCACCTTTGACCCAGGAA	AGGCTCCTGCTTCACCTG	66
<b>Sox9</b>	GTACCCGCATCTGCACAAC	CTCCTCCACGAAGGGTCTCT	75
<b>Sp6</b>	TTCGGCCTAGGTCTCTTTCA	CCCACACACACACCTCATCT	32
<b>Tfcp2l1</b>	GGGGACTACTCGGAGCATCT	TTCCGATCAGCTCCCTTG	53
<b>Zfp750</b>	GAGCCAGCGTGAGAACAGA	CAGGAGAGTTCCTTCCGTCA	68
<b>Irf6</b>	GCTTGCTGCTCCTAACCTGA	CTTTCTGGTGGGCAATGAG	34
<b>Vdr</b>	CACCTGGCTGATCTTGTCAGT	CTGGTCATCAGAGGTGAGGTC	89
<b>Elf1</b>	CCAGAGGAAGCAACCATAGC	AACCTGGGTTGAAGCCTGTA	38

Table S4. Sequences of oligonucleotides used for the cloning of the Grhl3 promoter.

	Forward	Reverse
Grhl3 1kb Promoter	GCTAGCCTCGAGTGCCTAGCACAGAGCACTTAC	GGATCCAAGCTTGGTGCCGACTGCAGCTAGAC

Table S5. qPCR primers and UPL probes used in ChIP experiments

Primers	Forward	Reverse	Probe
<b>Grhl3_Prom_52</b>	TCTAGTTCTCCCGTTCCTTCC	TTCTTAGCTGAGGGGGTGAG	52
<b>Grhl3_Prom_108</b>	GGAGAAAAGAGAGGGGACTCA	TCACCTTCTCTACCCGGAAA	108

Table S6. Primers used in RT-PCR experiments

Primers	Forward	Reverse
<b>Casz1_RT</b>	TCTACTACCACGGCTGCCAC	TCGTTGCTGGATTCCTCGTG
<b>Zfp750_RT</b>	CGACTCCAGCAAGCTGAGCA	GGCGTTGCTTGCATACATGG
<b>Elf3_RT</b>	GTCTGGAGGGCAAGAAGAGC	CCAACCTCTTCTTCCTTCCA
<b>Tbp_RT</b>	AGCCTCAGTACAGCAATCAAC	GAACTTCACATCACAGCTCC