



SUPPLEMENTARY FIGURES





Cellular growth assessment after treatment of MCF7, MDA-MB-231, MDA-MB-468, T47D, SKBR3, MCF-10A and BJ-hTERT cell lines with different concentrations of thioalbamide (0.01 to 1 μ M) or Doxorubicin (0.01 to 5 μ M) for 72 h. Results, quantified by the MTT assay, are expressed as percentage of growth vs control cells treated with vehicle alone (DMSO). Values represent mean ± SD of three independent experiments, each performed with triplicate samples. *Results reported in Frattaruolo et al. 2017.





MDA-MB-468



Figure S2. Thioalbamide-induced DNA fragmentation in MDA-MB-468 cells. TdT-mediated dUTP nick-end-labeling (TUNEL) assay in MDA-MB-468 cells treated for 72 h with vehicle (Ctrl) or 50 nM thioalbamide. DAPI was used for DNA staining. Images at 20X magnification were taken with an Olympus BX41 microscope with CSV1.14 software, using a CAMXC-30 for image acquisition (scale bar: 25 μm)







Figure S3. Thioalbamide-induced DNA fragmentation in SKBR3 cells. TdT-mediated dUTP nick-end-labeling (TUNEL) assay in SKBR3 cells treated for 72 h with vehicle (Ctrl) or 100 nM thioalbamide. DAPI was used for DNA staining. Images at 20X magnification were taken with an Olympus BX41 microscope with CSV1.14 software, using a CAMXC-30 for image acquisition (scale bar: $25 \mu m$)









Figure S4. Thioalbamide-induced DNA fragmentation in T47D cells. TdT-mediated dUTP nick-end-labeling (TUNEL) assay in T47D cells treated for 72 h with vehicle (Ctrl) or 50 nM thioalbamide. DAPI was used for DNA staining. Images at 20X magnification were taken with an Olympus BX41 microscope with CSV1.14 software, using a CAMXC-30 for image acquisition (scale bar: $25 \mu m$)





MDA-MB-231



Figure S5. Thioalbamide-induced DNA fragmentation in MDA-MB-231 cells. TdT-mediated dUTP nick-end-labeling (TUNEL) assay in MDA-MB-231 cells treated for 72 h with vehicle (Ctrl) or 100 nM thioalbamide. DAPI was used for DNA staining. Images at 20X magnification were taken with an Olympus BX41 microscope with CSV1.14 software, using a CAMXC-30 for image acquisition (scale bar: $25 \mu m$)



Figure S6. Thioalbamide induces apoptosis in breast cancer cell lines. Quantification of positive cells in TUNEL assay performed on MCF7, MDA-MB-468, MDA-MB-231, SKBR3 and T47D. Values represent mean \pm SD of three independent experiments. ****P value <0.0001



Figure S7. Oxidative stress underlies thioalbamide cytotoxicity in breast cancer cell lines. Cell viability assessment of MDA-MB231 (a), MDA-MB-468 (b), T47D (c) and SKBR3 (d) cells after treatment for 72 h with 50 nM thioalbamide, alone or in association with vitamin E (Vit E), as indicated. **P value <0.01; ***P value <0.001; ***P value <0.001





Figure S8. Metabolic profile of breast cancer cells treated with thioalbamide. Metabolic profile (OCR and ECAR) of T47D and MDA-MB-468 cells treated with 50/100 nM thioalbamide for 48 h.Values represent mean \pm SEM of three indipendent experiments



Figure S9. Thioalbamide induces loss of mitochondrial membrane potential in breast cancer cells. Mitochondrial membrane potential was assessed after 72 h of treatment with thioalbamide, using MitoTracker Orange CM-H2TMRos probe. 50.000 events were acquired using SONY SH800 flow cytometer. Values represent mean \pm SD of three independent experiments. Treatment conditions: DMSO (-), 50 nM thioalbamide (+), 100 nM thioalbamide (++). 2µM FCCP was used as positive control. *P value <0.05; ****P value <0.0001







Figure S10. Thioalbamide reduces CD44 expression in breast cancer cells. CD44 antibody staining was assessed, after 72h of treatment with thioalbamide, using anti-human CD44 (BD Bioscience). 50.000 events were acquired using SONY SH800 flow cytometer. Values represent mean \pm SD of three independent experiments. Treatment conditions: DMSO (-), 50 nM thioalbamide (+). *P value <0.05; **P value <0.01