



# **MicroRNAs in Cancer and Cardiovascular Disease**

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**Abstract:** Although cardiac tumor formation is rare, accumulating evidence suggests that the two leading causes of deaths, cancers, and cardiovascular diseases are similar in terms of pathogenesis, including angiogenesis, immune responses, and fibrosis. These similarities have led to the creation of new exciting field of study called cardio-oncology. Here, we review the similarities between cancer and cardiovascular disease from the perspective of microRNAs (miRNAs). As miRNAs are well-known regulators of translation by binding to the 3'-untranslated regions (UTRs) of messenger RNAs (mRNAs), we carefully dissect how a specific set of miRNAs are both oncomiRs (miRNAs in cancer) and myomiRs (muscle-related miRNAs). Furthermore, from the standpoint of similar pathogenesis, miRNAs categories related to the similar pathogenesis are discussed; namely, angiomiRs, ImmunemiRs, and fibromiRs.

Keywords: cancer; cardiovascular disease; miRNA

## 1. Introduction

Cancer and cardiovascular disease are the leading causes of death across the globe accounting for one in six deaths [1] and 32% of all deaths worldwide [2], respectively, according to World Health Organization (WHO). Both cancer and cardiovascular disease are the umbrella terms commonly used to describe several disease etiologies. Each etiology of cancer and cardiovascular disease (e.g., lung cancer and ischemic heart disease, respectively) has its own distinct cause and progression pattern. However, recent research suggests that many aspects of cancer and cardiovascular disease are similar in terms of pathogenesis [3–5], leading to the development of specific field of study called cardio-oncology [6,7]. For example, both diseases involve dysregulated functionalities in vasculature, where abnormal vasculature (called, tumor vasculature [8]) occurs in cancer, while coronary artery disease is a type of cardiovascular disease caused by the narrowing or blockage of coronary arteries [9]. Another example is the involvement of immune responses, where prolonged or chronic inflammation is a hallmark of cancer [10–12] as well as cardiovascular disease [13–15]. The activation of immune responses often leads to the deposition of excessive extracellular matrices [16,17], which are another hallmark of cancer [17] and cardiac fibrosis as the end-stage of heart failure [18].

MicroRNAs (miRNAs) are evolutionary-conserved, regulatory short [~22 nucleotides (nt)] non-protein-coding RNAs that function as translational inhibitors by binding to the 3'-untranslated regions (3'-UTRs) of messenger RNAs (mRNAs) [19,20]. As one miRNA is predicted to bind hundreds of mRNAs due to its very short seed sequence (~6 nt) [21–23], it is speculated and experimentally shown for some miRNAs to regulate cascades of signaling pathways and their downstream targets. Due to their versatilities, dysregulation in miRNAs is linked to a variety of diseases, including cancer [24,25] and cardiovascular disease [26–28]. As the regulatory importance of miRNAs is experimentally proven, the therapeutic silencing of miRNAs is being explored [29–34]. However, due to their biodistributions (e.g., including their presence in the circulation [35–37]) and the presence of many target mRNAs for one miRNA, the precise mechanistic elucidation of each miRNA



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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). is urgently needed to advance into clinics. Since a specific miRNA is highly dependent on which target mRNAs are present in a specific biological context, it must be taken into consideration that the same miRNA can yield different biological outcomes depending on the specific cell or tissue [38]. This is especially important when considering miRNAs as potential therapeutic targets.

As cancer and cardiovascular disease share several aspects of disease causes and progressions, it is no surprise that many miRNAs are shown to be involved in pathogeneses of both cancer and cardiovascular disease. Because the heart is the least likely organ to harbor tumor growth [39], the communication between researchers working in miRNAs for either cancer or cardiovascular biology is scarce, although many miRNAs are found to be dysregulated in both diseases. To fill this gap in knowledge, here, we summarize the current status of miRNA research from the perspective of shared disease progression mechanisms in cancer and cardiovascular disease.

#### 2. OncomiRs vs. MyomiRs

According to the latest annotation provided by the GENCODE consortium (Release 41; https://www.gencodegenes.org/human/stats\_41.html; accessed on 3 October 2022), there are 1879 human miRNAs. Due to the intensive miRNA research in the last three decades [40], many (but not all) miRNAs have been studied functionally and some mechanistically. To date, miRNAs have been categorized based on their functionalities. These categories include oncomiRs and (cardiac) myomiRs to describe cancer- and striated muscle-related miRNAs, respectively. Although the heart consists of cell types other than cardiac muscle (cardiomyocytes), for simplicity, here, we will compare oncomiRs and myomiRs to understand the possible overlaps of the functional miRNAs in both cancer and cardiovas-cular disease.

As there are many different types of cancer, the list of oncomiRs is growing rapidly due to the availability of next generation sequencing (i.e., small RNA sequencing) to identify miRNAs overexpressed in tumor samples. As such, there are several dedicated databases for oncomiRs available, including miRCancer [41], OncomiR [42–44], and the OncoMir Cancer Database (OMCD) [45]. Compared to oncomiRs, the list of myomiRs is small, including *miR-1*, *miR-133a/b*, *miR-206*, *miR-208a/b*, *miR-302*, *miR-367*, *miR-486*, and *miR-499* [46,47]. Not surprisingly, all myomiRs are involved in tumorigenesis.

One of the most abundant miRNAs in the heart [48], *miR-133*, is a regulator of cardiac hypertrophy [49] and its down-regulation was observed in patients with myocardial infarction [50] (Figure 1). In gastric cancer, *miR-133* is down-regulated in gastric cancer patients and negatively associated with tumor size, invasion depth, and peripheral organ metastasis [51]. Mechanistically, *miR-133* targets 3'-UTR of the cell division cycle 42 (*CDC42*) gene to regulate the downstream effectors of CDC42, P21-activated kinases (PAKs). Similarly, an overexpression of *miR-133a* in the lung cancer cell lines, A549 and NCI-H1299, results in the suppression of cell proliferation, migration, and invasion by targeting matrix metallopeptidase 14 (*MMP14*) [52]. Another study shows that the overexpression of *miR-133b* in the lung cancer cell line, A549, re-sensitized the radioresistant A549 cells by targeting pyruvate kinase M1/2 (*PKM*, also known as *PKM2*) to regulate glycolysis [53]. Besides gastric and lung cancers, functions of *miR-133* are also reported in glioblastoma [54], oral cancer [55], and prostate cancer [56]. All other miRNAs are also shown to be functionally important for tumorigenesis, suggesting the importance of examining miRNAs in onco-cardiology.



**Figure 1.** OncomiRs and myomiRs. MyomiRs have a dual function and are involved in tumorigenesis. *miR-133* regulates the cardiac hypertrophy and improves the myocardial function after infarction, while it is associated with multiple tumors. *miR-133* targets 3'-UTR of *CDC42* and regulates PAKs, thus preventing the growth and metastasis of gastric cancer. Similarly, *miR-133* prevents the proliferation, migration, and invasion of lung cancer by targeting *MMP14* and *PKM2*. Furthermore, *miR-133* is involved also in the pathogenesis of glioblastoma and oral and prostate cancer. Figure created with BioRender.com, accessed on 24 October 2022.

## 3. Angiogenesis: AngiomiRs

Angiogenesis is the process of new blood vessel formation through the migration, growth, and differentiation of endothelial cells [57,58]. In cancer, angiogenesis allows for a tumor to grow as new vessels provide nutrients and oxygen to malignant cells [59,60]. In cardiovascular disease, therapeutic angiogenesis aims to provide the blood flow to the ischemic heart tissue [61,62]. Thus, in both diseases, angiogenesis is an important therapeutic target, although the opposite effects are observed. During angiogenesis, several miRNAs are functionally involved, which has created a specific term to describe these angiogenesis-related miRNAs called, angiomiRs (Figure 2). AngiomiRs include *miR-15/16*, *miR-17~92* cluster, *miR-18a*, *miR-19*, *miR-21*, *miR-23b*, *miR-27a/b*, *miR-29b*, *miR-30*, *miR-34a*, *miR-125b*, *miR-126*, *miR-128*, *miR-143*, *miR-145*, *miR-155*, *miR-192*, *miR-194*, *miR-199a*, *miR-200* family, *miR-204*, *miR-210*, *miR-217*, *miR-296*, *miR-378*, *miR-484*, *miR-494*, *miR-497*, *miR-542-3p*, *miR-573*, *miR-642*, and *let-7b* [63,64], which some are discussed below.

The *miR*-17~92 cluster was first reported in tumorigenesis [65] and is one of the most well-studied miRNA clusters [66,67]. By crossing *miR*-17~92 floxed mice with an inducible vascular endothelial cell specific Cre driver (Cdh5-cre/ERT2), Chamorro-Jorganes et al. demonstrated that retinal angiogenesis was reduced during the development of these mice [68]. Furthermore, the vascular endothelial growth factor (VEGF)-induced ear and tumor angiogenesis were reduced, suggesting that VEGF regulates *miR*-17~92 cluster expression leading to the regulation of angiogenesis. The involvement of the *miR*-17~92 cluster is well documented in various diseases, including cardiovascular disease [69,70]. Since the *miR*-17~92 cluster consists of *miR*-17, *miR*-18a, *miR*-19a, *miR*-20a, *miR*-19b-1, and *miR*-92a-1, each miRNA in this cluster is also shown to be important for angiogenesis, including tumorigenesis and cardiovascular disease. For example, *miR*-92a is dysregulated

in many forms of cancer, suggesting it is a potential diagnostic biomarker as well as a therapeutic target [71]. In the cardiovascular system, Bonauer et al. demonstrated that overexpression of *miR-92a* in endothelial cells inhibited angiogenesis in murine models of limb ischemia and myocardial infarction, while the silencing of *miR-92a* via antagomiR resulted in enhanced angiogensis and the functional recovery of the damaged tissues in murine disease models, suggesting *miR-92a* as a potential therapeutic target for ischemia diseases [72].

The *miR*-200 family is another well studied miRNA family that includes *miR*-141, *miR*-200a, *miR*-200b, *miR*-200c, and *miR*-429 [73]. In cancer, the *miR*-200 family is shown to play functional roles in cell malignant transformation and preventing tumor initiation [74]. By profiling epicardial adipose tissue from coronary artery disease (CAD) patients and non-CAD atherosclerotic patients, Zhang et al. demonstrated that the expressions of *miR*-141-3*p*, *miR*-200b, *miR*-200c-3*p*, and *miR*-429 are up-regulated in CAD patients compared to non-CAD patients [75]. By performing a series of experiments in vitro, the authors demonstrated that the overexpression of *miR*-200b-3*p* in human umbilical vein endothelial cells (HUVECs) resulted in increased apoptosis under oxidative stress. Mechanistically, *miR*-300b-3*p* targets histone deacetylase 4 (HDAC4) as the overexpression of HDAC4 reduced the increased apoptosis induced by inhibiting *miR*-200b-3*p*, suggesting that *miR*-200b-3*p* is a potential therapeutic target for atherosclerosis.



**Figure 2.** The dual role of angiomiRs in cancer and cardiac pathophysiology. The *miR*-17~92 cluster is involved in tumorigenesis and tumor vascularization. This cluster is also involved in retinal angiogenesis and the progression of cardiovascular disease. The members of the *miR*-200 family prevent the tumor initiation and malignant transformation, although they are upregulated in coronary artery disease. *MiR*-34a is a tumor suppressor involved in the development of thyroid cancer, head and neck squamous cell carcinoma, and cancer stem cells division. The overexpression of *miR*-34a suppress the proliferation and induces senescence in cardiomyocytes, fibroblasts, smooth muscle, and endothelial cells, by inhibiting sirtuin 1 (*SIRT1*). Figure created with BioRender.com, accessed on 24 October 2022.

*MiR-34a* is a tumor suppressor and considered as a diagnostic and prognostic biomarker as well as a therapeutic target in various cancers, including head and neck squamous cell carcinoma, thyroid cancer, and cancer stem cells [76,77]. Interestingly, the expression of *miR-34a* is increased in senescent HUVECs and in the heart and spleen of older mice [78]. When overexpressed, *miR-34a* suppressed cell cycle and proliferation by inhibiting sirtuin 1 (*SIRT1*). Because ageing is a hot topic to be investigated, subsequent research shows the functional importance of *miR-34a* in cell types other than endothelial cells in the heart, including in cardiomyocytes [79,80], fibroblasts [81], and smooth muscle cells [82,83]. This is not an isolated case as many other angiomiRs (and other miRNAs) are expressed rather ubiquitously, suggesting that examining miRNAs as a common mechanism of action for cardio-oncology is not a big surprise.

#### 4. Immune Responses: Immuno-miRs

Prolonged inflammation is a hallmark of cancer [10] that immune systems can have both positive and negative effects on regarding the development of tumors and prognostics of cancer patients [84]. Indeed, immunotherapy is a type of treatment using one's own immune system to fight cancer, but the success rates of immunotherapy drugs vary between 15–30% in most tumor types, while 50–80% in melanoma [85]. As the immune system is a complex system involving many different cell types (e.g., basophils, eosinophils, lymphocytes, macrophages, monocytes, and neutrophils) to fight against infection [86,87], understanding the immune system is also important in cardiovascular disease [88–90]. For example, myocardial infarction leads to the loss of cardiomyocytes, which are replaced by non-contracting scar tissue [91,92]. The immune system is a double-edged sword in the remodeling process of the infarcted heart as macrophages are necessary for repair in the acute phase as their systemic depletion results in impaired scar formation and the rupture of the left ventricle of the heart. However, the accumulation of macrophages in non-infarcted regions of the left ventricle leads to progressive myocyte attrition, collagen deposition, and loss of the pump function of the heart in a chronic phase of remodeling of the infarcted heart. As miRNAs are expressed in many immune cells and finetune the important signaling pathways, the list of immuno-miRs is growing rapidly [93,94]. As such, specialized databases for immune-miRs are available, including IRNdb [95], RNA2Immune [96], and RNAimmuno [97]. In the following, examples of immune-miRs are explained in cancer and cardiovascular disease.

Monocytes are a type of white blood cells (leukocytes) that can differentiate into macrophages and dendritic cells [98]. Furthermore, monocyte-derived macrophages can be polarized into inflammatory subtype, M1, and anti-inflammatory subtype, M2 [99]. The cascade of differentiation and polarizations are controlled by the coordinate actions of cytokines, which can be regulated at the transcriptional and post-transcriptional levels, where miRNAs can regulate the translation of transcription factors responsible for cytokine gene expressions. These microRNAs include *miR*-125a-3p and *miR*-26a-2 in M1 macrophages, while miR-27a, miR-29b-1, miR-132, miR-193b, and miR-222 constitute the M2 macrophages [100] (Figure 3A). For example, *miR*-222 targets ADAM metallopeptidase domain 17 (ADAM17) to modulate multidrug resistance in colorectal carcinoma [101] (Figure 3B). In breast cancer, the overexpression of *miR*-222 inhibits the chemotaxis of tumorassociated macrophages by targeting C-X-C motif chemokine ligand 12 (CXCL12) [102]. In the serum, the level of *miR*-222 is independently associated with atrial fibrillation (irregular heart rhythm) in patients with degenerative valvular heart disease [103]. In addition, the level of *miR*-222 is elevated in acute viral myocarditis caused by Coxsackievirus B3 [104]. Although the functions of *miR*-222 are mainly reported in cardiomyocytes [104–106] and cardiac fibroblasts [107,108], it is clear that immune-miRs in monocytes and macrophages are important regulators of immune responses in cancer and cardiovascular disease.



**Figure 3.** Immuno-miRs. (**A**) MiRNAs responsible for regulation of cytokine gene expressions leading to the differentiation of two types of monocytes-derived macrophages—inflammatory subtype M1 and anti-inflammatory subtype M2. (**B**) The role of *miR-222* in tumorigenesis and cardiovascular disease. *MiR-222* targets *ADAM17* to prevent multidrug resistant colorectal carcinoma. The inhibitory effect on the chemotaxis of tumor associated macrophages in breast cancer is mediated by targeting *CXCL12*. The overexpression of *miR-222* is associated with atrial fibrillation and Coxsackie virus caused myocarditis. (**C**) The overexpression of *miR-155* is associated lymph node metastasis in breast cancer and advance of esophageal, liver, and lung cancer. *Mir-155* regulates angiogenesis by controlling the expression of *AGTR1* in endothelial cells and *SOCS1* in monocytes/macrophages. (**D**) The extracellular RNA, *miR-146a-5p*, is highly presented in hepatocellular carcinoma derived exosomes and regulates the polarization of macrophages into M2 tumor-associated macrophages. On the contrary, the cardiomyocytes-derived *miR-146a-5p* inhibits the M2 macrophage polarization by targeting *TRAF6* while promoting M1 macrophage polarization. Figure created with BioRender.com, accessed on 24 October 2022.

Enriched in immune cells, *miR-155* is a master regulator of immune responses [109] (Figure 3C). In breast cancer, the increased expression of *miR-155* is associated with high tumor grade, advanced stage, and lymph node metastasis [110]. Similarly, *miR-155* is over-expressed in other forms of cancer, including esophageal cancer [111], liver cancer [112], and lung cancer [113], which calls for *miR-155* as a diagnostic and prognostic cancer biomarker [114] as well as therapeutic target [115]. As shown in the previous section, *miR-155* is an angiomiR so its function is well known in the endothelial cells and atherosclerosis [116,117]. Besides endothelial cells, *miR-155* is highly expressed in monocytes and macrophages, which Pankratz et al. used in knockout mice to elegantly demonstrate that *miR-155* regulates angiogenesis and arteriogenesis by controlling their target genes, angiotensin II receptor type 1 (*AGTR1*) and suppressor of cytokine signaling 1 (*SOCS1*) in endothelial and monocyte/macrophages, respectively [118].

Extracellular RNAs (exRNAs) are a type of cell–cell communication that are produced by a donor cell and are released into the extracellular environment (e.g., body fluid, circulation) [119]. They are contained in the lipid particles, such as extracellular vehicles (EVs), including exosomes. ExRNAs include proteins and RNAs, including miRNAs. For example, *miR*-146*a*-5*p* is enriched in the hepatocellular carcinoma-derived exosomes [120] (Figure 3D). The transcription factor, spalt like transcription factor 4 (SALL4), binds to the promoter of *miR-146a-5p* to directly control its expression in exosomes, thereby regulates the polarization of macrophages into M2 tumor-associated macrophages. In contrast, cardiomyocyte-derived exosomal *miR-146a-5p* promotes M1 macrophage polarization while inhibiting M2 macrophage polarization by targeting TNF receptor associated factor 6 (*TRAF6*) [121]. This is just of many miRNAs contained in exosomes.

The studying of immunology has intensified in recent years due to the rise of coronavirus disease 2019 (COVID-19) [122–125]. As there is a substantial risk of heart problems associated with COVID-19 and mRNA vaccines [126–129], it is likely that more and more miRNAs will be identified in the heart, which may have been studied in cancer previously to expand the list of immune-miRs in the cardio-oncology field.

#### 5. Fibrosis: fibromiRs

Fibrosis is a process in which fibroblasts and other mesenchymal cells are activated to become myofibroblasts to secrete an excess number of extracellular matrices (ECM; e.g., collagens, glycosaminoglycans, and glycoproteins) [130] (Figure 4A). It is the end stage of many diseases, including cardiovascular disease [18]. In cancer, cancer-associated fibroblasts (CAFs) promote tumorigenic features, including ECM deposition, epithelial-to-mesenchymal transition (EMT), and metastasis [131]. To understand fibrosis, many screening studies have been performed to identify differentially expressed genes and miRNAs, which are collectively called fibromiRs [132–135]. For example, *miR-21* is the most studied fibromiR [136–139] (Figure 4B). Not only is it highly expressed in many forms of cancer and suggested as potential diagnostic biomarkers of cancer types (breast, pancreatic, colorectal, and prostate cancer) [140], *miR-21* stimulates MAP kinase signaling in cardiac fibroblasts, thereby contributing to myocardial disease [141]. Furthermore, *miR-21* targets matrix metallopeptidase 2 (*Mmp2*) in cardiac fibroblasts of the infarcted heart via phosphatase and the tensin homolog (PTEN) pathway [142], suggesting the important signaling roles of *miR-21* in both cancer and cardiovascular disease.

Multiple reports show that another oncomiR, *miR*-22, is highly involved in tumor progression in multiple tumors, including breast cancer [143], acute myeloid leukemia (AML) [144], and hepatocellular carcinoma (HCC) [145,146]. The effect of *miR*-22 on HCC seems to be related to the early effect of *miR*-22 on liver fibrosis through its regulation of bone morphogenic protein 7 (*BMP7*) [147] (Figure 4C), which starts from a degenerative process and ultimately leads to HCC developing. Interestingly, *miR*-22 is reported to have the same effect on cardiac fibrosis [148] via the regulation of *Sirt1* and *HDAC4*. As the role of *miR*-22 in fibrosis is conserved in multiple diseases and tissues, this miRNA could serve as a potential therapeutic target in liver [149] and cardiac fibrosis [150].

Although fibroblasts can be found throughout the human body, they are heterogeneous populations of cells without any single cell surface marker that is specific for fibroblasts as many markers are expressed in other cell types, including epithelial and immune cells [151,152]. In this regard, microRNAs are involved in activating fibroblasts, which contribute to the heterogeneity of fibroblasts [153]. For example, the members of the miR-200 family, miR-141 and miR-200a, target C-X-C motif chemokine ligand 12 (CXCL12; also known as  $CXCL12\beta$ ) to regulate the immunosuppressive activity of a subtype of carcinoma-associated fibroblasts in ovarian cancer [154]. Besides the miR-200 family being angiomiRs as written in above subsection, *miR-200b* is negatively regulated by the epigenetic factor, DNA methyltransferase 3 alpha (Dnmt3a), to control autophagy in rat cardiac fibroblasts [155]. In addition, small RNA-seq experiment using rat cardiac fibroblasts induced with transforming growth factor-β1 (TGF-β1) showed 3 up- (miR-325-3p, miR-325-5p, and miR-210-5p) and 21 down-regulated miRNAs (e.g., miR-19a-3p, miR-19b-3p, miR-144-3p, and miR-200b-3p), potentially targeting genes involved in calcium and glutamatergic synapse signaling pathways [156]. Taken together, there are many shared fibromiRs between CAFs and cardiac fibroblasts.



**Figure 4.** FibromiRs. (**A**) The core mechanisms of fibrosis and carcinogenesis. Multiple cell types (e.g., fibroblasts, myofibroblasts, epithelial cells, and macrophages) are involved. The pathophysiological mechanisms include inflammation, epithelial to mesenchymal transition, extracellular matrix accumulation, and metastasis. (**B**) *MiR-21* is a diagnostic biomarker for multiple cancers, including breast, pancreatic, colorectal, and prostate. *MiR-21* stimulates cardiac fibroblasts by targeting *Mmp2* and the PTEN pathway, leading to the progression of myocardial disease. (**C**) *MiR-22* induces the liver fibrosis through *BMP7* leading to progression into hepatocellular carcinoma. In the heart, *miR-22* promotes cardiac fibrosis by targeting *Sirt1* and *HDAC4*. Figure created with BioRender.com, accessed on 24 October 2022.

#### 6. Conclusions

To maintain the homeostasis of the tissues and remodeling of the tissues upon damages, angiogenesis, immune responses, and fibrosis are interconnected. As such, miRNAs are identified to be involved in each cellular activity as angiomiRs, immuno-miRs, and fibromiRs, respectively. Not surprisingly, some miRNAs (e.g., *miR-17~92* cluster, *miR-34a*, and *miR-200* family) are involved in all three cellular activities, which some overlapping miRNAs are responsible for such cellular activities and responses. This is particularly interesting as cancer is considered as a complex adaptive ecosystem [157], in which cancer cells and the stromal cells transform, cooperate, and even co-evolve with each other over time and space [158]. Thus, it will be interesting to further investigate miRNAs from the perspective of the ecosystem in cancer and possibly in cardiovascular disease.

As cancer and cardiovascular disease are two of the leading causes of death worldwide, it will be exciting to find a common disease mechanism. As reviewed above, miRNAs are shared between these life-threatening diseases. Given that miRNAs are investigated as potential therapeutic targets, increased communication between researchers working with cancer and cardiovascular disease is necessary to find a potential cure for these diseases. To this end, the rise of the cardio-oncology field should facilitate a further understanding of the pathogeneses of these two diseases, possibly through miRNAs. **Author Contributions:** M.I., R.P. and S.U. wrote the manuscript, generated figures, and approved the final version of this manuscript. All authors have read and agreed to the published version of the manuscript.

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### References

- 1. Available online: https://www.who.int/news-room/fact-sheets/detail/cancer (accessed on 3 October 2022).
- 2. Available online: https://www.who.int/health-topics/cardiovascular-diseases#tab=tab\_1 (accessed on 3 October 2022).
- 3. Seton-Rogers, S. Cardiovascular disease and cancer communicate. Nat. Rev. Cancer 2020, 20, 552. [CrossRef]
- 4. Knisely, J.P.S.; Henry, S.A.; Saba, S.G.; Puckett, L.L. Cancer and cardiovascular disease. Lancet 2020, 395, 1904. [CrossRef]
- 5. De Boer, R.A.; Meijers, W.C.; van der Meer, P.; van Veldhuisen, D.J. Cancer and heart disease: Associations and relations. *Eur. J. Heart Fail.* **2019**, *21*, 1515–1525. [CrossRef]
- 6. Wang, Y.; Wang, Y.; Han, X.; Sun, J.; Li, C.; Adhikari, B.K.; Zhang, J.; Miao, X.; Chen, Z. Cardio-Oncology: A Myriad of Relationships Between Cardiovascular Disease and Cancer. *Front. Cardiovasc. Med.* **2022**, *9*, 727487. [CrossRef]
- Koutsoukis, A.; Ntalianis, A.; Repasos, E.; Kastritis, E.; Dimopoulos, M.A.; Paraskevaidis, I. Cardio-oncology: A Focus on Cardiotoxicity. *Eur. Cardiol. Rev.* 2018, 13, 64–69. [CrossRef]
- 8. Ruoslahti, E. Specialization of tumour vasculature. Nat. Rev. Cancer 2002, 2, 83–90. [CrossRef]
- 9. Malakar, A.K.; Choudhury, D.; Halder, B.; Paul, P.; Uddin, A.; Chakraborty, S. A review on coronary artery disease, its risk factors, and therapeutics. *J. Cell. Physiol.* 2019, 234, 16812–16823. [CrossRef]
- 10. Hiam-Galvez, K.J.; Allen, B.M.; Spitzer, M.H. Systemic immunity in cancer. Nat. Rev. Cancer 2021, 21, 345–359. [CrossRef]
- Gordon-Weeks, A.; Yuzhalin, A.E. Cancer Extracellular Matrix Proteins Regulate Tumour Immunity. *Cancers* 2020, 12, 3331. [CrossRef]
- 12. Gonzalez, H.; Hagerling, C.; Werb, Z. Roles of the immune system in cancer: From tumor initiation to metastatic progression. *Genes Dev.* **2018**, *32*, 1267–1284. [CrossRef]
- 13. Sorriento, D.; Iaccarino, G. Inflammation and Cardiovascular Diseases: The Most Recent Findings. *Int. J. Mol. Sci.* 2019, 20, 3879. [CrossRef]
- 14. Lopez-Candales, A.; Hernandez Burgos, P.M.; Hernandez-Suarez, D.F.; Harris, D. Linking Chronic Inflammation with Cardiovascular Disease: From Normal Aging to the Metabolic Syndrome. *J. Nat. Sci.* **2017**, *3*, e341.
- 15. Mason, J.C.; Libby, P. Cardiovascular disease in patients with chronic inflammation: Mechanisms underlying premature cardiovascular events in rheumatologic conditions. *Eur. Heart J.* **2015**, *36*, 482–489. [CrossRef]
- 16. Boyd, D.F.; Thomas, P.G. Towards integrating extracellular matrix and immunological pathways. *Cytokine* **2017**, *98*, 79–86. [CrossRef]
- 17. Pickup, M.W.; Mouw, J.K.; Weaver, V.M. The extracellular matrix modulates the hallmarks of cancer. *EMBO Rep.* 2014, 15, 1243–1253. [CrossRef]
- Ichiki, T.; Schirger, J.A.; Huntley, B.K.; Brozovich, F.V.; Maleszewski, J.J.; Sandberg, S.M.; Sangaralingham, S.J.; Park, S.J.; Burnett, J.C., Jr. Cardiac fibrosis in end-stage human heart failure and the cardiac natriuretic peptide guanylyl cyclase system: Regulation and therapeutic implications. J. Mol. Cell. Cardiol. 2014, 75, 199–205. [CrossRef]
- 19. Bartel, D.P. MicroRNAs: Genomics, biogenesis, mechanism, and function. Cell 2004, 116, 281–297. [CrossRef]
- 20. Gebert, L.F.R.; MacRae, I.J. Regulation of microRNA function in animals. Nat. Rev. Mol. Cell Biol. 2019, 20, 21–37. [CrossRef]
- 21. Chen, Y.; Wang, X. miRDB: An online database for prediction of functional microRNA targets. *Nucleic Acids Res.* **2020**, *48*, D127–D131. [CrossRef]
- 22. Agarwal, V.; Bell, G.W.; Nam, J.W.; Bartel, D.P. Predicting effective microRNA target sites in mammalian mRNAs. *Elife* 2015, *4*, e05005. [CrossRef]
- Paraskevopoulou, M.D.; Georgakilas, G.; Kostoulas, N.; Vlachos, I.S.; Vergoulis, T.; Reczko, M.; Filippidis, C.; Dalamagas, T.; Hatzigeorgiou, A.G. DIANA-microT web server v5.0: Service integration into miRNA functional analysis workflows. *Nucleic* Acids Res. 2013, 41, W169–W173. [CrossRef] [PubMed]
- Chen, S.; Wang, Y.; Li, D.; Wang, H.; Zhao, X.; Yang, J.; Chen, L.; Guo, M.; Zhao, J.; Chen, C.; et al. Mechanisms Controlling MicroRNA Expression in Tumor. *Cells* 2022, 11, 2852. [CrossRef] [PubMed]
- 25. Ding, L.; Gu, H.; Xiong, X.; Ao, H.; Cao, J.; Lin, W.; Yu, M.; Lin, J.; Cui, Q. MicroRNAs Involved in Carcinogenesis, Prognosis, Therapeutic Resistance and Applications in Human Triple-Negative Breast Cancer. *Cells* **2019**, *8*, 1492. [CrossRef]
- 26. Kansakar, U.; Varzideh, F.; Mone, P.; Jankauskas, S.S.; Santulli, G. Functional Role of microRNAs in Regulating Cardiomyocyte Death. *Cells* **2022**, *11*, 983. [CrossRef] [PubMed]
- 27. Song, R.; Hu, X.Q.; Zhang, L. Mitochondrial MiRNA in Cardiovascular Function and Disease. Cells 2019, 8, 1475. [CrossRef]
- 28. Mirna, M.; Paar, V.; Rezar, R.; Topf, A.; Eber, M.; Hoppe, U.C.; Lichtenauer, M.; Jung, C. MicroRNAs in Inflammatory Heart Diseases and Sepsis-Induced Cardiac Dysfunction: A Potential Scope for the Future? *Cells* **2019**, *8*, 1352. [CrossRef]
- Forterre, A.; Komuro, H.; Aminova, S.; Harada, M. A Comprehensive Review of Cancer MicroRNA Therapeutic Delivery Strategies. *Cancers* 2020, 12, 1852. [CrossRef]

- 30. Szczepanek, J.; Skorupa, M.; Tretyn, A. MicroRNA as a Potential Therapeutic Molecule in Cancer. Cells 2022, 11, 1008. [CrossRef]
- Momin, M.Y.; Gaddam, R.R.; Kravitz, M.; Gupta, A.; Vikram, A. The Challenges and Opportunities in the Development of MicroRNA Therapeutics: A Multidisciplinary Viewpoint. *Cells* 2021, 10, 3097. [CrossRef]
- O'Neill, C.P.; Dwyer, R.M. Nanoparticle-Based Delivery of Tumor Suppressor microRNA for Cancer Therapy. *Cells* 2020, 9, 521. [CrossRef]
- Seo, H.A.; Moeng, S.; Sim, S.; Kuh, H.J.; Choi, S.Y.; Park, J.K. MicroRNA-Based Combinatorial Cancer Therapy: Effects of MicroRNAs on the Efficacy of Anti-Cancer Therapies. *Cells* 2019, *9*, 29. [CrossRef] [PubMed]
- Quemener, A.M.; Centomo, M.L.; Sax, S.L.; Panella, R. Small Drugs, Huge Impact: The Extraordinary Impact of Antisense Oligonucleotides in Research and Drug Development. *Molecules* 2022, 27, 536. [CrossRef] [PubMed]
- 35. Cui, M.; Wang, H.; Yao, X.; Zhang, D.; Xie, Y.; Cui, R.; Zhang, X. Circulating MicroRNAs in Cancer: Potential and Challenge. *Front. Genet.* **2019**, *10*, 626. [CrossRef] [PubMed]
- 36. Wu, Y.; Li, Q.; Zhang, R.; Dai, X.; Chen, W.; Xing, D. Circulating microRNAs: Biomarkers of disease. *Clin. Chim. Acta* 2021, 516, 46–54. [CrossRef]
- Creemers, E.E.; Tijsen, A.J.; Pinto, Y.M. Circulating microRNAs: Novel biomarkers and extracellular communicators in cardiovascular disease? *Circ. Res.* 2012, 110, 483–495. [CrossRef]
- Murakami, Y.; Yasuda, T.; Saigo, K.; Urashima, T.; Toyoda, H.; Okanoue, T.; Shimotohno, K. Comprehensive analysis of microRNA expression patterns in hepatocellular carcinoma and non-tumorous tissues. *Oncogene* 2006, 25, 2537–2545. [CrossRef]
- Tyebally, S.; Chen, D.; Bhattacharyya, S.; Mughrabi, A.; Hussain, Z.; Manisty, C.; Westwood, M.; Ghosh, A.K.; Guha, A. Cardiac Tumors: JACC CardioOncology State-of-the-Art Review. *Cardio Oncol.* 2020, 2, 293–311. [CrossRef]
- 40. Lee, R.C.; Feinbaum, R.L.; Ambros, V. The *C. elegans* heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. *Cell* **1993**, *75*, 843–854. [CrossRef]
- 41. Xie, B.; Ding, Q.; Han, H.; Wu, D. miRCancer: A microRNA-cancer association database constructed by text mining on literature. *Bioinformatics* **2013**, *29*, 638–644. [CrossRef]
- 42. Sarver, A.L.; Subramanian, S. Competing endogenous RNA database. Bioinformation 2012, 8, 731–733. [CrossRef]
- Sarver, A.L.; Phalak, R.; Thayanithy, V.; Subramanian, S. S-MED: Sarcoma microRNA expression database. *Lab. Investig.* 2010, 90, 753–761. [CrossRef] [PubMed]
- 44. Sarver, A.L.; French, A.J.; Borralho, P.M.; Thayanithy, V.; Oberg, A.L.; Silverstein, K.A.; Morlan, B.W.; Riska, S.M.; Boardman, L.A.; Cunningham, J.M.; et al. Human colon cancer profiles show differential microRNA expression depending on mismatch repair status and are characteristic of undifferentiated proliferative states. *BMC Cancer* **2009**, *9*, 401. [CrossRef]
- 45. Sarver, A.L.; Sarver, A.E.; Yuan, C.; Subramanian, S. OMCD: OncomiR Cancer Database. *BMC Cancer* 2018, *18*, 1223. [CrossRef] [PubMed]
- Zilahi, E.; Adamecz, Z.; Bodoki, L.; Griger, Z.; Poliska, S.; Nagy-Vincze, M.; Danko, K. Dysregulated expression profile of myomiRs in the skeletal muscle of patients with polymyositis. *EJIFCC* 2019, 30, 237–245. [PubMed]
- McCarthy, J.J. MicroRNA-206: The skeletal muscle-specific myomiR. *Biochim. Biophys. Acta* 2008, 1779, 682–691. [CrossRef] [PubMed]
- 48. Leptidis, S.; El Azzouzi, H.; Lok, S.I.; de Weger, R.; Olieslagers, S.; Kisters, N.; Silva, G.J.; Heymans, S.; Cuppen, E.; Berezikov, E.; et al. A deep sequencing approach to uncover the miRNOME in the human heart. *PLoS ONE* **2013**, *8*, e57800. [CrossRef]
- Care, A.; Catalucci, D.; Felicetti, F.; Bonci, D.; Addario, A.; Gallo, P.; Bang, M.L.; Segnalini, P.; Gu, Y.; Dalton, N.D.; et al. MicroRNA-133 controls cardiac hypertrophy. *Nat. Med.* 2007, 13, 613–618. [CrossRef]
- 50. Bostjancic, E.; Brandner, T.; Zidar, N.; Glavac, D.; Stajer, D. Down-regulation of miR-133a/b in patients with myocardial infarction correlates with the presence of ventricular fibrillation. *Biomed. Pharmacother.* **2018**, *99*, 65–71. [CrossRef]
- 51. Cheng, Z.; Liu, F.; Wang, G.; Li, Y.; Zhang, H.; Li, F. miR-133 is a key negative regulator of CDC42-PAK pathway in gastric cancer. *Cell. Signal.* **2014**, *26*, 2667–2673. [CrossRef]
- 52. Xu, M.; Wang, Y.Z. miR133a suppresses cell proliferation, migration and invasion in human lung cancer by targeting MMP14. Oncol. Rep. 2013, 30, 1398–1404. [CrossRef]
- Liu, G.; Li, Y.I.; Gao, X. Overexpression of microRNA-133b sensitizes non-small cell lung cancer cells to irradiation through the inhibition of glycolysis. Oncol. Lett. 2016, 11, 2903–2908. [CrossRef] [PubMed]
- Xu, F.; Li, F.; Zhang, W.; Jia, P. Growth of glioblastoma is inhibited by miR-133-mediated EGFR suppression. *Tumour Biol.* 2015, 36, 9553–9558. [CrossRef] [PubMed]
- 55. Jung, J.E.; Lee, J.Y.; Park, H.R.; Kang, J.W.; Kim, Y.H.; Lee, J.H. MicroRNA-133 Targets Phosphodiesterase 1C in Drosophila and Human Oral Cancer Cells to Regulate Epithelial-Mesenchymal Transition. *J. Cancer* **2021**, *12*, 5296–5309. [CrossRef] [PubMed]
- 56. Tao, J.; Wu, D.; Xu, B.; Qian, W.; Li, P.; Lu, Q.; Yin, C.; Zhang, W. microRNA-133 inhibits cell proliferation, migration and invasion in prostate cancer cells by targeting the epidermal growth factor receptor. *Oncol. Rep.* **2012**, 27, 1967–1975. [CrossRef]
- 57. Potente, M.; Carmeliet, P. The Link Between Angiogenesis and Endothelial Metabolism. *Annu. Rev. Physiol.* **2017**, *79*, 43–66. [CrossRef] [PubMed]
- 58. Chung, A.S.; Ferrara, N. Developmental and pathological angiogenesis. Annu. Rev. Cell. Dev. Biol. 2011, 27, 563–584. [CrossRef]
- 59. Lugano, R.; Ramachandran, M.; Dimberg, A. Tumor angiogenesis: Causes, consequences, challenges and opportunities. *Cell. Mol. Life Sci.* 2020, 77, 1745–1770. [CrossRef]

- 60. Eelen, G.; Treps, L.; Li, X.; Carmeliet, P. Basic and Therapeutic Aspects of Angiogenesis Updated. *Circ. Res.* 2020, 127, 310–329. [CrossRef]
- 61. Yla-Herttuala, S.; Bridges, C.; Katz, M.G.; Korpisalo, P. Angiogenic gene therapy in cardiovascular diseases: Dream or vision? *Eur. Heart J.* **2017**, *38*, 1365–1371. [CrossRef] [PubMed]
- 62. Korpela, H.; Jarvelainen, N.; Siimes, S.; Lampela, J.; Airaksinen, J.; Valli, K.; Turunen, M.; Pajula, J.; Nurro, J.; Yla-Herttuala, S. Gene therapy for ischaemic heart disease and heart failure. *J. Intern. Med.* **2021**, 290, 567–582. [CrossRef]
- Salinas-Vera, Y.M.; Marchat, L.A.; Gallardo-Rincon, D.; Ruiz-Garcia, E.; Astudillo-De La Vega, H.; Echavarria-Zepeda, R.; Lopez-Camarillo, C. AngiomiRs: MicroRNAs driving angiogenesis in cancer (Review). *Int. J. Mol. Med.* 2019, 43, 657–670. [CrossRef] [PubMed]
- 64. Wang, S.; Olson, E.N. AngiomiRs-key regulators of angiogenesis. *Curr. Opin. Genet. Dev.* 2009, *19*, 205–211. [CrossRef] [PubMed]
  65. Ota, A.; Tagawa, H.; Karnan, S.; Tsuzuki, S.; Karpas, A.; Kira, S.; Yoshida, Y.; Seto, M. Identification and characterization of a
- novel gene, C13orf25, as a target for 13q31-q32 amplification in malignant lymphoma. *Cancer Res.* 2004, *64*, 3087–3095. [CrossRef]
  66. Mogilyansky, E.; Rigoutsos, I. The miR-17/92 cluster: A comprehensive update on its genomics, genetics, functions and increasingly important and numerous roles in health and disease. *Cell Death Differ.* 2013, *20*, 1603–1614. [CrossRef] [PubMed]
- 67. Mendell, J.T. miRiad roles for the miR-17-92 cluster in development and disease. *Cell* **2008**, 133, 217–222. [CrossRef] [PubMed]
- 68. Chamorro-Jorganes, A.; Lee, M.Y.; Araldi, E.; Landskroner-Eiger, S.; Fernandez-Fuertes, M.; Sahraei, M.; Quiles Del Rey, M.; van Solingen, C.; Yu, J.; Fernandez-Hernando, C.; et al. VEGF-Induced Expression of miR-17-92 Cluster in Endothelial Cells Is Mediated by ERK/ELK1 Activation and Regulates Angiogenesis. *Circ. Res.* 2016, 118, 38–47. [CrossRef]
- 69. Gu, H.; Liu, Z.; Zhou, L. Roles of miR-17-92 Cluster in Cardiovascular Development and Common Diseases. *Biomed. Res. Int.* 2017, 2017, 9102909. [CrossRef]
- Danielson, L.S.; Park, D.S.; Rotllan, N.; Chamorro-Jorganes, A.; Guijarro, M.V.; Fernandez-Hernando, C.; Fishman, G.I.; Phoon, C.K.; Hernando, E. Cardiovascular dysregulation of miR-17-92 causes a lethal hypertrophic cardiomyopathy and arrhythmogenesis. *FASEB J.* 2013, 27, 1460–1467. [CrossRef]
- Li, M.; Guan, X.; Sun, Y.; Mi, J.; Shu, X.; Liu, F.; Li, C. miR-92a family and their target genes in tumorigenesis and metastasis. *Exp. Cell Res.* 2014, 323, 1–6. [CrossRef]
- 72. Bonauer, A.; Carmona, G.; Iwasaki, M.; Mione, M.; Koyanagi, M.; Fischer, A.; Burchfield, J.; Fox, H.; Doebele, C.; Ohtani, K.; et al. MicroRNA-92a controls angiogenesis and functional recovery of ischemic tissues in mice. *Science* **2009**, *324*, 1710–1713. [CrossRef]
- Cavallari, I.; Ciccarese, F.; Sharova, E.; Urso, L.; Raimondi, V.; Silic-Benussi, M.; D'Agostino, D.M.; Ciminale, V. The miR-200 Family of microRNAs: Fine Tuners of Epithelial-Mesenchymal Transition and Circulating Cancer Biomarkers. *Cancers* 2021, 13, 5874. [CrossRef] [PubMed]
- 74. Humphries, B.; Yang, C. The microRNA-200 family: Small molecules with novel roles in cancer development, progression and therapy. *Oncotarget* 2015, *6*, 6472–6498. [CrossRef] [PubMed]
- 75. Zhang, F.; Cheng, N.; Du, J.; Zhang, H.; Zhang, C. MicroRNA-200b-3p promotes endothelial cell apoptosis by targeting HDAC4 in atherosclerosis. *BMC Cardiovasc. Disord.* **2021**, *21*, 172. [CrossRef] [PubMed]
- Li, W.J.; Wang, Y.; Liu, R.; Kasinski, A.L.; Shen, H.; Slack, F.J.; Tang, D.G. MicroRNA-34a: Potent Tumor Suppressor, Cancer Stem Cell Inhibitor, and Potential Anticancer Therapeutic. *Front. Cell Dev. Biol.* 2021, 9, 640587. [CrossRef] [PubMed]
- Kalfert, D.; Ludvikova, M.; Pesta, M.; Ludvik, J.; Dostalova, L.; Kholova, I. Multifunctional Roles of miR-34a in Cancer: A Review with the Emphasis on Head and Neck Squamous Cell Carcinoma and Thyroid Cancer with Clinical Implications. *Diagnostics* 2020, 10, 563. [CrossRef] [PubMed]
- Ito, T.; Yagi, S.; Yamakuchi, M. MicroRNA-34a regulation of endothelial senescence. *Biochem. Biophys. Res. Commun.* 2010, 398, 735–740. [CrossRef]
- 79. Yang, Y.; Cheng, H.W.; Qiu, Y.; Dupee, D.; Noonan, M.; Lin, Y.D.; Fisch, S.; Unno, K.; Sereti, K.I.; Liao, R. MicroRNA-34a Plays a Key Role in Cardiac Repair and Regeneration Following Myocardial Infarction. *Circ. Res.* **2015**, *117*, 450–459. [CrossRef]
- Boon, R.A.; Iekushi, K.; Lechner, S.; Seeger, T.; Fischer, A.; Heydt, S.; Kaluza, D.; Treguer, K.; Carmona, G.; Bonauer, A.; et al. MicroRNA-34a regulates cardiac ageing and function. *Nature* 2013, 495, 107–110. [CrossRef]
- Huang, Y.; Qi, Y.; Du, J.Q.; Zhang, D.F. MicroRNA-34a regulates cardiac fibrosis after myocardial infarction by targeting Smad4. Expert Opin. Ther. Targets 2014, 18, 1355–1365. [CrossRef]
- 82. Chen, Q.; Yang, F.; Guo, M.; Wen, G.; Zhang, C.; Zhu, J.; Xiao, Q.; Zhang, L. miRNA-34a reduces neointima formation through inhibiting smooth muscle cell proliferation and migration. *J. Mol. Cell. Cardiol.* **2015**, *89*, 75–86. [CrossRef]
- Badi, I.; Mancinelli, L.; Polizzotto, A.; Ferri, D.; Zeni, F.; Burba, I.; Milano, G.; Brambilla, F.; Saccu, C.; Bianchi, M.E.; et al. miR-34a Promotes Vascular Smooth Muscle Cell Calcification by Downregulating SIRT1 (Sirtuin 1) and Axl (AXL Receptor Tyrosine Kinase). Arterioscler. Thromb. Vasc. Biol. 2018, 38, 2079–2090. [CrossRef] [PubMed]
- Janssen, L.M.E.; Ramsay, E.E.; Logsdon, C.D.; Overwijk, W.W. The immune system in cancer metastasis: Friend or foe? J. Immunother. Cancer 2017, 5, 79. [CrossRef] [PubMed]
- 85. Esfahani, K.; Roudaia, L.; Buhlaiga, N.; Del Rincon, S.V.; Papneja, N.; Miller, W.H., Jr. A review of cancer immunotherapy: From the past, to the present, to the future. *Curr. Oncol.* **2020**, *27*, S87–S97. [CrossRef] [PubMed]
- Marshall, J.S.; Warrington, R.; Watson, W.; Kim, H.L. An introduction to immunology and immunopathology. *Allergy Asthma Clin. Immunol.* 2018, 14, 49. [CrossRef]
- 87. Nicholson, L.B. The immune system. Essays Biochem. 2016, 60, 275–301. [CrossRef]

- 88. Fernandez-Ruiz, I. Immune system and cardiovascular disease. Nat. Rev. Cardiol. 2016, 13, 503. [CrossRef] [PubMed]
- 89. Frostegard, J. Immunity, atherosclerosis and cardiovascular disease. BMC Med. 2013, 11, 117. [CrossRef]
- 90. Lafuse, W.P.; Wozniak, D.J.; Rajaram, M.V.S. Role of Cardiac Macrophages on Cardiac Inflammation, Fibrosis and Tissue Repair. *Cells* **2020**, *10*, 51. [CrossRef]
- Duncan, S.E.; Gao, S.; Sarhene, M.; Coffie, J.W.; Linhua, D.; Bao, X.; Jing, Z.; Li, S.; Guo, R.; Su, J.; et al. Macrophage Activities in Myocardial Infarction and Heart Failure. *Cardiol. Res. Pract.* 2020, 2020, 4375127. [CrossRef]
- 92. Lavine, K.J.; Pinto, A.R.; Epelman, S.; Kopecky, B.J.; Clemente-Casares, X.; Godwin, J.; Rosenthal, N.; Kovacic, J.C. The Macrophage in Cardiac Homeostasis and Disease: JACC Macrophage in CVD Series (Part 4). J. Am. Coll. Cardiol. 2018, 72, 2213–2230. [CrossRef]
- 93. Kroesen, B.J.; Teteloshvili, N.; Smigielska-Czepiel, K.; Brouwer, E.; Boots, A.M.; van den Berg, A.; Kluiver, J. Immuno-miRs: Critical regulators of T-cell development, function and ageing. *Immunology* **2015**, *144*, 1–10. [CrossRef] [PubMed]
- 94. Davidson-Moncada, J.; Papavasiliou, F.N.; Tam, W. MicroRNAs of the immune system: Roles in inflammation and cancer. *Ann. N. Y. Acad. Sci.* **2010**, *1183*, 183–194. [CrossRef] [PubMed]
- Denisenko, E.; Ho, D.; Tamgue, O.; Ozturk, M.; Suzuki, H.; Brombacher, F.; Guler, R.; Schmeier, S. IRNdb: The database of immunologically relevant non-coding RNAs. *Database* 2016, 2016, baw138. [CrossRef] [PubMed]
- Wang, J.; Li, S.; Wang, T.; Xu, S.; Wang, X.; Kong, X.; Lu, X.; Zhang, H.; Li, L.; Feng, M.; et al. RNA2Immune: A database of experimentally supported data linking non-coding RNA regulation to the immune system. *Genom. Proteom. Bioinform.* 2022. [CrossRef]
- Olejniczak, M.; Galka-Marciniak, P.; Polak, K.; Fligier, A.; Krzyzosiak, W.J. RNAimmuno: A database of the nonspecific immunological effects of RNA interference and microRNA reagents. RNA 2012, 18, 930–935. [CrossRef]
- Italiani, P.; Boraschi, D. From Monocytes to M1/M2 Macrophages: Phenotypical vs. Functional Differentiation. *Front. Immunol.* 2014, 5, 514. [CrossRef]
- 99. Yunna, C.; Mengru, H.; Lei, W.; Weidong, C. Macrophage M1/M2 polarization. Eur. J. Pharmacol. 2020, 877, 173090. [CrossRef]
- 100. Gombozhapova, A.; Rogovskaya, Y.; Shurupov, V.; Rebenkova, M.; Kzhyshkowska, J.; Popov, S.V.; Karpov, R.S.; Ryabov, V. Macrophage activation and polarization in post-infarction cardiac remodeling. *J. Biomed. Sci.* **2017**, *24*, 13. [CrossRef]
- Xu, K.; Liang, X.; Shen, K.; Sun, L.; Cui, D.; Zhao, Y.; Tian, J.; Ni, L.; Liu, J. MiR-222 modulates multidrug resistance in human colorectal carcinoma by down-regulating ADAM-17. *Exp. Cell Res.* 2012, *318*, 2168–2177. [CrossRef]
- 102. Li, Y.; Zhao, L.; Shi, B.; Ma, S.; Xu, Z.; Ge, Y.; Liu, Y.; Zheng, D.; Shi, J. Functions of miR-146a and miR-222 in Tumor-associated Macrophages in Breast Cancer. *Sci. Rep.* **2015**, *5*, 18648. [CrossRef]
- 103. Zhou, H.; Lin, S.; Li, X.; Guo, D.; Wang, Y.; Hu, Y. Serum miR-222 is independently associated with atrial fibrillation in patients with degenerative valvular heart disease. *BMC Cardiovasc. Disord.* **2021**, *21*, 98. [CrossRef] [PubMed]
- 104. Corsten, M.F.; Heggermont, W.; Papageorgiou, A.P.; Deckx, S.; Tijsma, A.; Verhesen, W.; van Leeuwen, R.; Carai, P.; Thibaut, H.J.; Custers, K.; et al. The microRNA-221/-222 cluster balances the antiviral and inflammatory response in viral myocarditis. *Eur. Heart J.* 2015, 36, 2909–2919. [CrossRef] [PubMed]
- 105. Liu, X.; Xiao, J.; Zhu, H.; Wei, X.; Platt, C.; Damilano, F.; Xiao, C.; Bezzerides, V.; Bostrom, P.; Che, L.; et al. miR-222 is necessary for exercise-induced cardiac growth and protects against pathological cardiac remodeling. *Cell Metab.* 2015, 21, 584–595. [CrossRef] [PubMed]
- Knyrim, M.; Rabe, S.; Grossmann, C.; Gekle, M.; Schreier, B. Influence of miR-221/222 on cardiomyocyte calcium handling and function. *Cell Biosci.* 2021, 11, 160. [CrossRef]
- 107. Verjans, R.; Peters, T.; Beaumont, F.J.; van Leeuwen, R.; van Herwaarden, T.; Verhesen, W.; Munts, C.; Bijnen, M.; Henkens, M.; Diez, J.; et al. MicroRNA-221/222 Family Counteracts Myocardial Fibrosis in Pressure Overload-Induced Heart Failure. *Hypertension* 2018, 71, 280–288. [CrossRef]
- 108. Wang, Z.; Wang, Z.; Gao, L.; Xiao, L.; Yao, R.; Du, B.; Li, Y.; Wu, L.; Liang, C.; Huang, Z.; et al. miR-222 inhibits cardiac fibrosis in diabetic mice heart via regulating Wnt/beta-catenin-mediated endothelium to mesenchymal transition. *J. Cell. Physiol.* 2020, 235, 2149–2160. [CrossRef] [PubMed]
- Alivernini, S.; Gremese, E.; McSharry, C.; Tolusso, B.; Ferraccioli, G.; McInnes, I.B.; Kurowska-Stolarska, M. MicroRNA-155-at the Critical Interface of Innate and Adaptive Immunity in Arthritis. *Front. Immunol.* 2017, *8*, 1932. [CrossRef]
- 110. Mattiske, S.; Suetani, R.J.; Neilsen, P.M.; Callen, D.F. The oncogenic role of miR-155 in breast cancer. *Cancer Epidemiol. Biomark. Prev.* **2012**, *21*, 1236–1243. [CrossRef]
- 111. Nariman-Saleh-Fam, Z.; Saadatian, Z.; Daraei, A.; Mansoori, Y.; Bastami, M.; Tavakkoli-Bazzaz, J. The intricate role of miR-155 in carcinogenesis: Potential implications for esophageal cancer research. *Biomark. Med.* **2019**, *13*, 147–159. [CrossRef]
- 112. Xin, X.; Lu, Y.; Xie, S.; Chen, Y.; Jiang, X.; Song, S.; Wang, L.; Pu, H.; Gui, X.; Li, T.; et al. miR-155 Accelerates the Growth of Human Liver Cancer Cells by Activating CDK2 via Targeting H3F3A. *Mol. Ther. Oncolytics.* **2020**, *17*, 471–483. [CrossRef]
- 113. Shao, C.; Yang, F.; Qin, Z.; Jing, X.; Shu, Y.; Shen, H. The value of miR-155 as a biomarker for the diagnosis and prognosis of lung cancer: A systematic review with meta-analysis. *BMC Cancer* **2019**, *19*, 1103. [CrossRef] [PubMed]
- Hosseini Mojahed, F.; Aalami, A.H.; Pouresmaeil, V.; Amirabadi, A.; Qasemi Rad, M.; Sahebkar, A. Clinical Evaluation of the Diagnostic Role of MicroRNA-155 in Breast Cancer. *Int. J. Genom.* 2020, 2020, 9514831. [CrossRef] [PubMed]
- 115. Bayraktar, R.; Van Roosbroeck, K. miR-155 in cancer drug resistance and as target for miRNA-based therapeutics. *Cancer Metastasis Rev.* 2018, 37, 33–44. [CrossRef]

- 116. Bruen, R.; Fitzsimons, S.; Belton, O. miR-155 in the Resolution of Atherosclerosis. *Front. Pharmacol.* **2019**, *10*, 463. [CrossRef] [PubMed]
- 117. Wei, Y.; Nazari-Jahantigh, M.; Neth, P.; Weber, C.; Schober, A. MicroRNA-126, -145, and -155: A therapeutic triad in atherosclerosis? *Arterioscler. Thromb. Vasc. Biol.* 2013, 33, 449–454. [CrossRef] [PubMed]
- 118. Pankratz, F.; Bemtgen, X.; Zeiser, R.; Leonhardt, F.; Kreuzaler, S.; Hilgendorf, I.; Smolka, C.; Helbing, T.; Hoefer, I.; Esser, J.S.; et al. MicroRNA-155 Exerts Cell-Specific Antiangiogenic but Proarteriogenic Effects During Adaptive Neovascularization. *Circulation* 2015, 131, 1575–1589. [CrossRef]
- 119. Gruner, H.N.; McManus, M.T. Examining the evidence for extracellular RNA function in mammals. *Nat. Rev. Genet.* **2021**, 22, 448–458. [CrossRef]
- 120. Yin, C.; Han, Q.; Xu, D.; Zheng, B.; Zhao, X.; Zhang, J. SALL4-mediated upregulation of exosomal miR-146a-5p drives T-cell exhaustion by M2 tumor-associated macrophages in HCC. *Oncoimmunology* **2019**, *8*, 1601479. [CrossRef]
- 121. Shimada, B.K.; Yang, Y.; Zhu, J.; Wang, S.; Suen, A.; Kronstadt, S.M.; Jeyaram, A.; Jay, S.M.; Zou, L.; Chao, W. Extracellular miR-146a-5p Induces Cardiac Innate Immune Response and Cardiomyocyte Dysfunction. *Immunohorizons* 2020, 4, 561–572. [CrossRef]
- 122. Merad, M.; Blish, C.A.; Sallusto, F.; Iwasaki, A. The immunology and immunopathology of COVID-19. *Science* 2022, 375, 1122–1127. [CrossRef]
- Diamond, M.S.; Kanneganti, T.D. Innate immunity: The first line of defense against SARS-CoV-2. Nat. Immunol. 2022, 23, 165–176.
   [CrossRef] [PubMed]
- 124. Bhardwaj, A.; Sapra, L.; Saini, C.; Azam, Z.; Mishra, P.K.; Verma, B.; Mishra, G.C.; Srivastava, R.K. COVID-19: Immunology, Immunopathogenesis and Potential Therapies. *Int. Rev. Immunol.* **2022**, *41*, 171–206. [CrossRef] [PubMed]
- 125. Mortaz, E.; Tabarsi, P.; Varahram, M.; Folkerts, G.; Adcock, I.M. The Immune Response and Immunopathology of COVID-19. *Front. Immunol.* **2020**, *11*, 2037. [CrossRef] [PubMed]
- 126. Urban, S.; Fulek, M.; Blaziak, M.; Iwanek, G.; Jura, M.; Fulek, K.; Guzik, M.; Garus, M.; Gajewski, P.; Lewandowski, L.; et al. COVID-19 Related Myocarditis in Adults: A Systematic Review of Case Reports. J. Clin. Med. 2022, 11, 5519. [CrossRef] [PubMed]
- 127. Kuehn, B.M. Cardiac Complications More Common After COVID-19 Than Vaccination. JAMA 2022, 327, 1951. [CrossRef]
- 128. Abbasi, J. The COVID Heart-One Year After SARS-CoV-2 Infection, Patients Have an Array of Increased Cardiovascular Risks. *JAMA* 2022, 327, 1113–1114. [CrossRef]
- 129. Verma, A.K.; Lavine, K.J.; Lin, C.Y. Myocarditis after COVID-19 mRNA Vaccination. N. Engl. J. Med. 2021, 385, 1332–1334. [CrossRef]
- 130. Plikus, M.V.; Wang, X.; Sinha, S.; Forte, E.; Thompson, S.M.; Herzog, E.L.; Driskell, R.R.; Rosenthal, N.; Biernaskie, J.; Horsley, V. Fibroblasts: Origins, definitions, and functions in health and disease. *Cell* **2021**, *184*, 3852–3872. [CrossRef]
- 131. Asif, P.J.; Longobardi, C.; Hahne, M.; Medema, J.P. The Role of Cancer-Associated Fibroblasts in Cancer Invasion and Metastasis. *Cancers* **2021**, *13*, 4720. [CrossRef]
- 132. Smolarz, B.; Durczynski, A.; Romanowicz, H.; Szyllo, K.; Hogendorf, P. miRNAs in Cancer (Review of Literature). *Int. J. Mol. Sci.* **2022**, *23*, 2805. [CrossRef]
- 133. Wang, S.; Lv, T.; Chen, Q.; Yang, Y.; Xu, L.; Zhang, X.; Wang, E.; Hu, X.; Liu, Y. Transcriptome sequencing and lncRNA-miRNAmRNA network construction in cardiac fibrosis and heart failure. *Bioengineered* 2022, *13*, 7118–7133. [CrossRef] [PubMed]
- Banerjee, M.; Ferragut Cardoso, A.; Al-Eryani, L.; Pan, J.; Kalbfleisch, T.S.; Srivastava, S.; Rai, S.N.; States, J.C. Dynamic alteration in miRNA and mRNA expression profiles at different stages of chronic arsenic exposure-induced carcinogenesis in a human cell culture model of skin cancer. *Arch. Toxicol.* 2021, *95*, 2351–2365. [CrossRef] [PubMed]
- 135. Zhang, Y.; Wang, D.; Peng, M.; Tang, L.; Ouyang, J.; Xiong, F.; Guo, C.; Tang, Y.; Zhou, Y.; Liao, Q.; et al. Single-cell RNA sequencing in cancer research. *J. Exp. Clin. Cancer Res.* **2021**, *40*, 81. [CrossRef] [PubMed]
- Surina, S.; Fontanella, R.A.; Scisciola, L.; Marfella, R.; Paolisso, G.; Barbieri, M. miR-21 in Human Cardiomyopathies. *Front. Cardiovasc. Med.* 2021, *8*, 767064. [CrossRef] [PubMed]
- 137. Jenike, A.E.; Halushka, M.K. miR-21: A non-specific biomarker of all maladies. Biomark. Res. 2021, 9, 18. [CrossRef]
- 138. Feng, Y.H.; Tsao, C.J. Emerging role of microRNA-21 in cancer. Biomed. Rep. 2016, 5, 395–402. [CrossRef] [PubMed]
- 139. Cheng, Y.; Zhang, C. MicroRNA-21 in cardiovascular disease. J. Cardiovasc. Transl. Res. 2010, 3, 251–255. [CrossRef]
- 140. Bautista-Sanchez, D.; Arriaga-Canon, C.; Pedroza-Torres, A.; De La Rosa-Velazquez, I.A.; Gonzalez-Barrios, R.; Contreras-Espinosa, L.; Montiel-Manriquez, R.; Castro-Hernandez, C.; Fragoso-Ontiveros, V.; Alvarez-Gomez, R.M.; et al. The Promising Role of miR-21 as a Cancer Biomarker and Its Importance in RNA-Based Therapeutics. *Mol. Ther. Nucleic Acids* 2020, 20, 409–420. [CrossRef]
- 141. Thum, T.; Gross, C.; Fiedler, J.; Fischer, T.; Kissler, S.; Bussen, M.; Galuppo, P.; Just, S.; Rottbauer, W.; Frantz, S.; et al. MicroRNA-21 contributes to myocardial disease by stimulating MAP kinase signalling in fibroblasts. *Nature* 2008, 456, 980–984. [CrossRef]
- 142. Roy, S.; Khanna, S.; Hussain, S.R.; Biswas, S.; Azad, A.; Rink, C.; Gnyawali, S.; Shilo, S.; Nuovo, G.J.; Sen, C.K. MicroRNA expression in response to murine myocardial infarction: MiR-21 regulates fibroblast metalloprotease-2 via phosphatase and tensin homologue. *Cardiovasc. Res.* 2009, *82*, 21–29. [CrossRef]
- 143. Song, S.J.; Poliseno, L.; Song, M.S.; Ala, U.; Webster, K.; Ng, C.; Beringer, G.; Brikbak, N.J.; Yuan, X.; Cantley, L.C.; et al. MicroRNA-antagonism regulates breast cancer stemness and metastasis via TET-family-dependent chromatin remodeling. *Cell* 2013, 154, 311–324. [CrossRef]

- 144. Song, S.J.; Ito, K.; Ala, U.; Kats, L.; Webster, K.; Sun, S.M.; Jongen-Lavrencic, M.; Manova-Todorova, K.; Teruya-Feldstein, J.; Avigan, D.E.; et al. The oncogenic microRNA miR-22 targets the TET2 tumor suppressor to promote hematopoietic stem cell self-renewal and transformation. *Cell Stem Cell* 2013, 13, 87–101. [CrossRef] [PubMed]
- 145. Jiang, R.; Deng, L.; Zhao, L.; Li, X.; Zhang, F.; Xia, Y.; Gao, Y.; Wang, X.; Sun, B. miR-22 promotes HBV-related hepatocellular carcinoma development in males. *Clin. Cancer Res.* 2011, *17*, 5593–5603. [CrossRef] [PubMed]
- 146. Zhang, L.; Yang, P.; Wang, J.; Liu, Q.; Wang, T.; Wang, Y.; Lin, F. MiR-22 regulated T cell differentiation and hepatocellular carcinoma growth by directly targeting Jarid2. *Am. J. Cancer Res.* **2021**, *11*, 2159–2173. [PubMed]
- 147. Ji, D.; Li, B.; Shao, Q.; Li, F.; Li, Z.; Chen, G. MiR-22 Suppresses BMP7 in the Development of Cirrhosis. *Cell. Physiol. Biochem.* 2015, *36*, 1026–1036. [CrossRef]
- 148. Huang, Z.P.; Wang, D.Z. miR-22 in cardiac remodeling and disease. Trends Cardiovasc. Med. 2014, 24, 267–272. [CrossRef]
- 149. Hu, Y.; Liu, H.X.; Jena, P.K.; Sheng, L.; Ali, M.R.; Wan, Y.Y. miR-22 inhibition reduces hepatic steatosis via FGF21 and FGFR1 induction. *JHEP Rep.* **2020**, *2*, 100093. [CrossRef]
- Wang, R.; Xu, Y.; Zhang, W.; Fang, Y.; Yang, T.; Zeng, D.; Wei, T.; Liu, J.; Zhou, H.; Li, Y.; et al. Inhibiting miR-22 Alleviates Cardiac Dysfunction by Regulating Sirt1 in Septic Cardiomyopathy. *Front. Cell Dev. Biol.* 2021, 9, 650666. [CrossRef]
- 151. Chang, H.Y.; Chi, J.T.; Dudoit, S.; Bondre, C.; van de Rijn, M.; Botstein, D.; Brown, P.O. Diversity, topographic differentiation, and positional memory in human fibroblasts. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 12877–12882. [CrossRef]
- 152. Muhl, L.; Genove, G.; Leptidis, S.; Liu, J.; He, L.; Mocci, G.; Sun, Y.; Gustafsson, S.; Buyandelger, B.; Chivukula, I.V.; et al. Single-cell analysis uncovers fibroblast heterogeneity and criteria for fibroblast and mural cell identification and discrimination. *Nat. Commun.* **2020**, *11*, 3953. [CrossRef]
- 153. LeBleu, V.S.; Neilson, E.G. Origin and functional heterogeneity of fibroblasts. FASEB J. 2020, 34, 3519–3536. [CrossRef] [PubMed]
- 154. Givel, A.M.; Kieffer, Y.; Scholer-Dahirel, A.; Sirven, P.; Cardon, M.; Pelon, F.; Magagna, I.; Gentric, G.; Costa, A.; Bonneau, C.; et al. miR200-regulated CXCL12beta promotes fibroblast heterogeneity and immunosuppression in ovarian cancers. *Nat. Commun.* 2018, 9, 1056. [CrossRef] [PubMed]
- 155. Zhao, X.D.; Qin, R.H.; Yang, J.J.; Xu, S.S.; Tao, H.; Ding, X.S.; Shi, K.H. DNMT3A controls miR-200b in cardiac fibroblast autophagy and cardiac fibrosis. *Inflamm. Res.* 2018, 67, 681–690. [CrossRef] [PubMed]
- 156. Liu, S.; Ke, W.; Liu, Y.; Zhao, Z.; An, L.; You, X.; Yang, F.; Yang, X.; Wang, G.; Zhao, X. Function analysis of differentially expressed microRNAs in TGF-beta1-induced cardiac fibroblasts differentiation. *BioSci. Rep.* 2019, 39, BSR20182048. [CrossRef] [PubMed]
- 157. Willis, A.J. The Ecosystem: An Evolving Concept Viewed Historically. Funct. Ecol. 1997, 11, 268–271. [CrossRef]
- 158. Luo, W. Nasopharyngeal Carcinoma Ecology Theory: Cancer as Multidimensional Spatiotemporal "Unity of Ecology and Evolution" Pathological Ecosystem. *Preprints* 2022, 2022100226. [CrossRef]