

SUPPLEMENTARY DATA

Emerging Role of miR-21-5p in Neuron–Glia Dysregulation and Exosome Transfer Using Multiple Models of Alzheimer’s Disease

Cells

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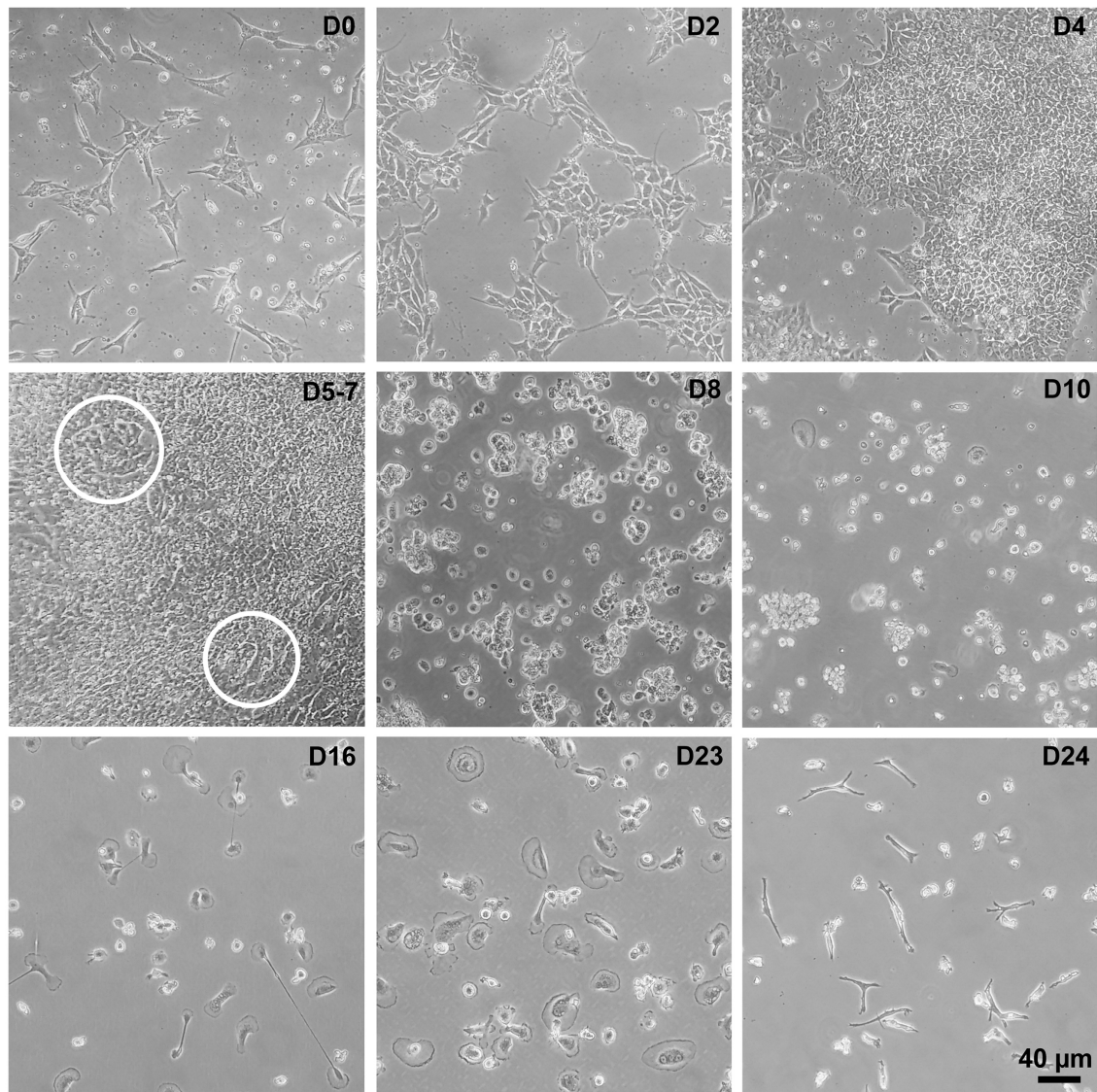
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Supplementary Table S1. List of primers used for mRNA/miRNA analysis by RT-qPCR.

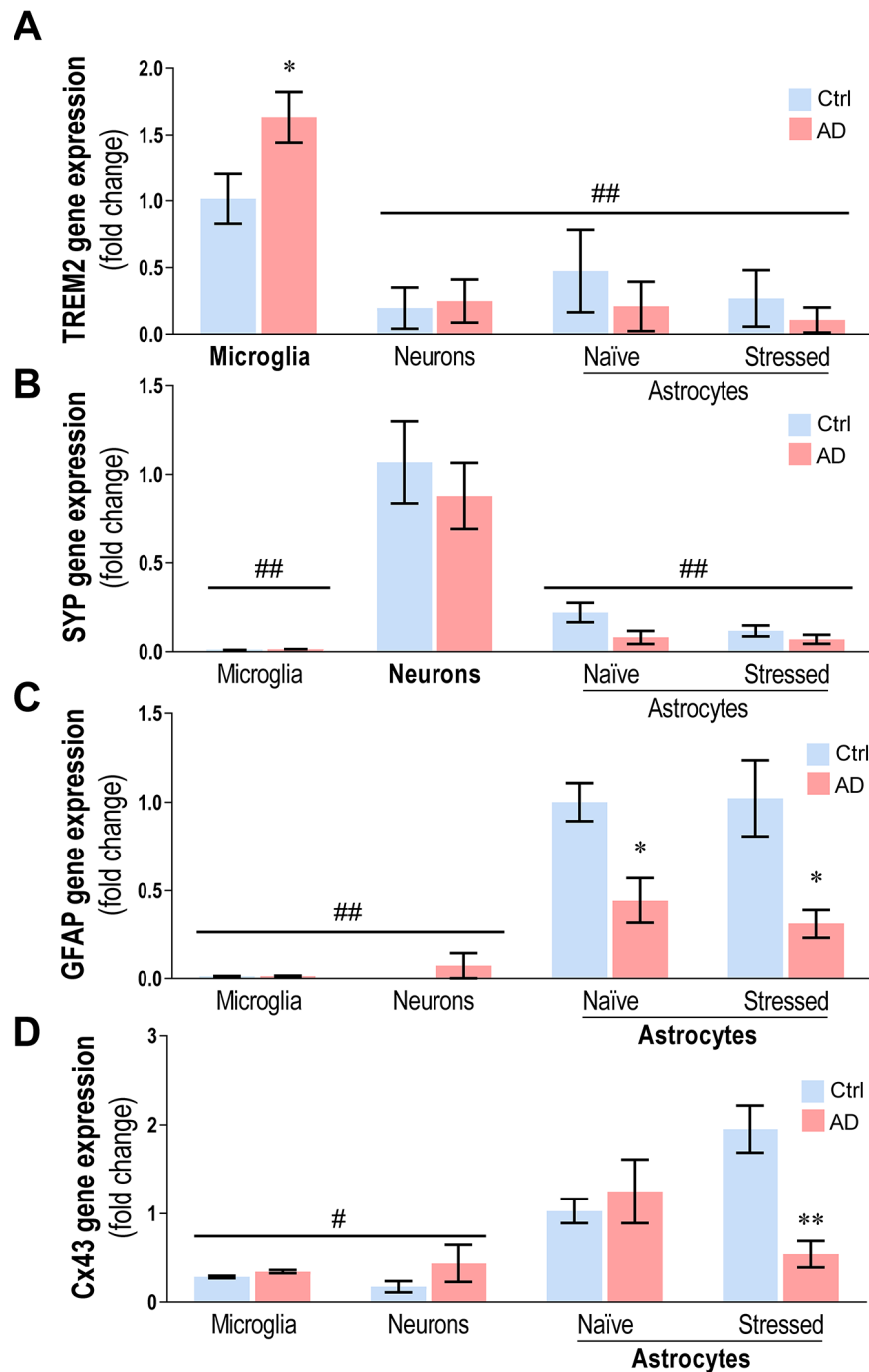
miRNAs	Target sequence (5' to 3')
hsa-miR-124-3p	UAAGGCACGCGGUGAAUGCC
hsa-miR-125b-5p	UCCCUGAGACCCUAACUUGUGA
hsa-miR-21-5p	UAGCUUAUCAGACUGAUGUUGA
hsa-miR-146a-5p	UGAGAACUGAAUCCAUGGGUU
hsa-miR-155-5p	UUAAUGCUAUCGUGAUAGGGGU
hsa-miR-23a-3p	AUCACAUUGCCAGGGAUUUCC
hsa-miR-29a-5p	ACUGAUUUUCUUUGGUGUUCAG
hsa-miR-122-5p	UGGAGUGUGACAAUGGUGUUUG
hsa-miR-219a-5p	UGAUUGUCCAAACGCAAUUCU
hsa-miR-338-5p	AACAAUAUCCUGGUGCUGAGUG
UniSp6 (spike-in)	(Reference sequence)
U6	(Reference sequence)
mRNAs <small>(human except indicated)</small>	Primer sequence (5' to 3')
<i>HMGB1</i>	FWD: CATCTCAGGGCCAAACCGATA REV: AGCAGACATGGTCTTCCACC
<i>MHC-II</i>	FWD: AGGGATTGCGCAAAAGCA REV: TCACCTCCATGTGCCTTACAGA
<i>CX3CR1</i>	FWD: GTGGTGTGCTGACAAAGCTTGAA REV: TCATGGGTGCCATCGTAAGAA
<i>CD68</i>	FWD: TCAGCTTTGGATTCATGCAG REV: AGGTGGACAGCTGGTGAAAG
<i>TNF-α</i>	FWD: AACCTCCTCTCTGCCATC REV: ATGTTCTGCTCCTCACA
<i>S100B</i>	FWD: TGTAGACCCTAACCCGGAGG REV: TGCATGGATGAGGAACGCAT
<i>iNOS</i>	FWD: TCCGAGGCAAACAGCACATTCA REV: GGGTTGGGGGTGTGGTGATGT
<i>TREM2</i>	FWD: ATGATGCGGGTCTCTACCAGTG REV: GCATCCTCGAAGCTCTCAGACT
<i>IL-6</i>	FWD: ATGAACTCCTTCTCCACAAGC REV: GTTTTCTGCCAGTGCCTCTTTG
<i>IL-8</i>	FWD: CTGCGCCAACACAGAAATTATTGTA REV: TTCACTGGCATCTTCACTGATTCTT
<i>IL-10</i>	FWD: CCTGGAGGAGGTGATGCCCCA REV: CCTGCTCCACGGCCTTGCTC
<i>Arg-1</i>	FWD: TGGAACTTGCATGGACA REV: AAGTCCGAAACAAGCCAA
<i>TGF-β</i>	FWD: TGCGCTTGAGATCTTCAA REV: GGGCTAGTCGCACAGAACT
<i>C/EBPα</i>	FWD: CAAAGCCAAGAAGTCGGTGGACAA REV: TCATTGTGACTGGTCAACTCCAGC

<i>PPARα</i>	FWD: CTCGAGGACACCGGAGAGG REV: CACGGAGCTGATCCCAAAGT
<i>PPARα (mouse)</i>	FWD: AACTGACGTTTGTGGCTGGTC REV: CACCATGTTGGATGGATGTGG
<i>PTEN (mouse)</i>	FWD: AATTCCCAGTCAGAGGCGCTATGT REV: GATTGCAAGTTCCGCCACTGAACA
<i>SYP</i>	FWD: AGTGCGCTAGAGCATTCTGG REV: CCACCATTCTGCCTCGCTTA
<i>GFAP</i>	FWD: GAGGTTGAGAGGGACAATCT REV: GCTTCATCTGCTTCCTGTCT
<i>Cx43</i>	FWD: GTTCAATCACTTGCGCTGAC REV: AGTTGAGTAGGCTTGAAC
<i>Ki-67</i>	FWD: TCCTTTGGTGGGCACCTAAGACCTG REV: TGATGGTTGAGGCTGTTCTTGATG
<i>GAPDH</i>	FWD: CGCTCTCTGCTCCTCCTGTT REV: CCATGGTGTCTGAGCGATGT
<i>β-actin</i>	FWD: CAGAGCCTCGCCTTTGCCGA REV: ATCCATGGTGAGCTGGCGGC

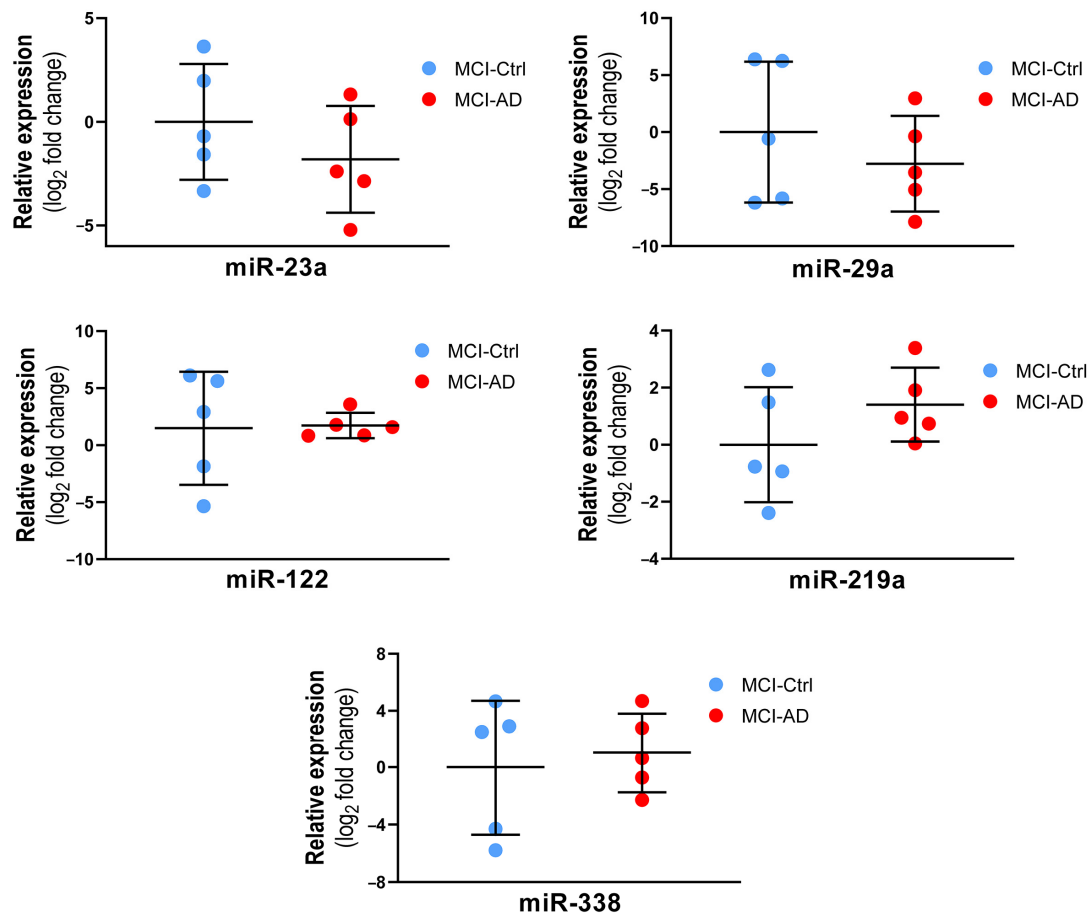
hsa, Homo sapiens; miRNA(miR), microRNA; mRNA, messenger RNA; RT-qPCR, real-time quantitative polymerase chain reaction; FWD, forward; REV, reverse; HMGB1, high mobility group box protein 1 coding gene; MHC-II, major histocompatibility complex class II coding gene; CX3CR1, CX3C chemokine receptor 1 gene; CD68, cluster of differentiation 68 gene; S100B, S100 calcium-binding protein B coding gene; TNFα, tumor necrosis factor alpha coding gene; iNOS, nitric oxide synthase coding gene; TREM2, triggering receptor expressed on myeloid cells 2 coding gene; IL-6, interleukin 6 coding gene; IL-8, interleukin 8 coding gene; IL-10, interleukin 10 coding gene; Arg-1, arginase-1 coding gene; TGF-β, transforming growth factor beta coding gene; C/EBPα, CCAAT enhancer binding protein alpha coding gene; PPARα, peroxisome proliferator-activated receptor alpha gene; PTEN, phosphatase and tensin homolog; SYP, synaptophysin gene; GFAP, glial fibrillary acidic protein; Cx43 (GJA1), connexin 43 (gap junction protein alpha 1); Ki-67 (MKI67), marker of proliferation Ki-67 gene; GAPDH, glyceraldehyde-3-phosphate dehydrogenase coding gene; β-actin coding gene.



Supplementary Figure S1. Representative brightfield images of the sequential differentiation stages associated to the differentiation protocol of microglia, in Figure 4 of the manuscript, and as described in Material and Methods. First, we induced early mesodermal differentiation from day 0 (D0) to D2, in low oxygen conditions. Then, at D2, we induced hematovascular mesodermal differentiation until D4, when we cultured cells back on normoxia to start the hemogenic endothelial differentiation. From D5 to D7, Hematopoietic progenitor cells (HPCs, highlighted in white circles) were expanded. At D8, myeloid progenitors were suspended, cultured in ultra-low attachment (ULA) plates, and expanded for 2 days. At D10, we started the primitive microglia differentiation until D16 to D23. Then at D24, primitive microglia were detached and plated into poly-D-lysine-coated plates and submitted to experiments. All images are at the same magnification and scale bar corresponds to 40 μ m. D, days in culture.



Supplementary Figure S2. Characteristic markers of microglia, neurons and astrocytes differentiated from induced pluripotent stem cells (iPSCs) support that each cell differentiation was successively achieved. (A) Gene expression of TREM2 (microglial marker) in iPSC-derived cells *vs.* Ctrl microglia; (B) Gene expression of SYP (neuronal marker) in iPSC-derived cells *vs.* Ctrl neurons; (C) Gene expression of GFAP, and (D) gene expression of Cx43 (astrocyte markers) in iPSC-derived cells *vs.* Ctrl naïve astrocytes. Results are mean \pm SEM from four independent experiments, presented in fold change. Two-tailed student's *t*-test: * $p < 0.05$; ** $p < 0.01$ *vs.* respective Ctrl paired cell type; One-way ANOVA # $p < 0.05$; ## $p < 0.05$ *vs.* each marker-specific cell type. TREM2, triggering receptor expressed on myeloid cells 2 coding gene; SYP, synaptophysin coding gene; GFAP, glial fibrillary acidic protein coding gene; Cx43 (or GJA1), connexin 43 (or gap junction protein alpha 1) coding gene; Ctrl, control; Naïve, non-immunostimulated astrocytes; stressed, immunostimulated with a cocktail composed by complement component C1q + interleukin-1 alpha + tumor necrosis factor alpha.



Supplementary Figure S3. Additional set of AD-associated miRNAs quantified in the CSF from MCI patients who fulfilled criteria for MCI due to AD (MCI-AD) *vs.* MCI patients with no biomarker criteria for AD (MCI-Ctrl). miR-23a, miR-29a, miR-122, miR-219a and miR-338 profile in the CSF samples from MCI-AD (red dots) side-by-side with miRNA profile in the CSF from MCI-Ctrl (blue dots). Results are mean \pm SEM from five different subjects of each group, presented in binary logarithm of fold change. Mann-Whitney U test with Bonferroni post-hoc correction: ** $p < 0.01$ MCI-AD *vs.* MCI-Ctrl. AD, Alzheimer's disease; Ctrl, control; MCI, mild cognitive impairment; CSF, cerebrospinal fluid.