

## **Supplementary material**

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## Supplementary text

**No transfer  $\text{NH}_4^+$  equation.** In the main text, we mentioned that when  $\text{NH}_4^+$  uptake equals the consumption of diatom, there is no transfer. We name the value of  $\text{NH}_4^+$  concentration in this condition as the *no transfer  $\text{NH}_4^+$  concentration*. Here is the derivation of no transfer  $\text{NH}_4^+$  concentration equation.

Firstly, we make the consumption of the diatom ( $\mu \times Q_N^D$ , only include N usage in growth) equal to  $\text{NH}_4^+$  uptake ( $V_{\text{NH}_4^+}$ ) and get Equation S1:

$$\mu \times Q_N^D = V_{\text{NH}_4^+} \quad \text{Equation S1}$$

Then use Equation S2 to represent  $V_{\text{NH}_4^+}$ :

$$V_{\text{NH}_4^+} = V_{\text{NH}_4^+}^{\max} \frac{[\text{NH}_4^+]}{[\text{NH}_4^+] + K_m} \quad \text{Equation S2}$$

And then come to a new Equation S3:

$$\mu \times Q_N^D = V_{\text{NH}_4^+}^{\max} \frac{[\text{NH}_4^+]}{[\text{NH}_4^+] + K_m} \quad \text{Equation S3}$$

Finally, we only keep  $[\text{NH}_4^+]$  on the left side and have Equation S4, which shows that *no transfer  $\text{NH}_4^+$*  can be only influenced by growth rate ( $\mu$ ), since all other parameters in the right side are constant.

$$[\text{NH}_4^+] = \frac{\mu \times Q_N^D \times V_{\text{NH}_4^+}^{\max} / K_m}{1 - \mu \times Q_N^D / K_m} \quad \text{Equation S4}$$

**Derivation of the key equations.** To help readers to better understand the key equations in the main text, in this section we describe the derivation of the key equations. The derivation used here is similar with a previous DDA modeling paper [1].

Figure S1 shows a schematic of the DDA model with flux notations. Here  $Q_i^j$  (pmol cell<sup>-1</sup>) means the cellular quota of element  $i$  in  $j$ , and  $J_i^{jk}$  (pmol cell<sup>-1</sup> d<sup>-1</sup>) represents transport of  $i$  from  $j$  to  $k$ , where  $i = \text{C or N}$  and  $j, k = \text{H (Heterocysts: Het), V (Vegetative cells: Veg) or D (Diatoms: Dia)}$ . In this derivation, we start with the dynamics of C and N quotas in three different cells.

Equation S5 and Equation S6 describe how C and N change in Het. Cellular C can be accumulated from Veg ( $J_C^{VH}$ ) and can be consumed by  $N_2$  fixation ( $F_C^{N_2fix}$ ). Cellular N can be consumed by transferring to Veg ( $-J_N^{HV}$ ) and can be obtained from  $N_2$  fixation ( $F_N^{N_2fix}$ ).

$$\frac{dQ_C^H}{dt} = J_C^{VH} - F_C^{N_2fix} \quad \text{Equation S5}$$

$$\frac{dQ_N^H}{dt} = -J_N^{HV} + F_N^{N_2fix} \quad \text{Equation S6}$$

Equation S7 and Equation S8 indicate element quotas dynamics ( $\frac{dQ_C^V}{dt}$  and  $\frac{dQ_N^V}{dt}$ ) in Veg. C can be obtained from photosynthesis in Veg ( $F_{pho}^V$ ) and transferred from diatom ( $J_C^{DV}$ ). And C can be consumed by being transferred to Het ( $J_C^{VH}$ ) and biosynthesis (including growth and respiration) of Veg and Het ( $\mu(Q_C^V + Q_C^H)(1 + E)$ ). As for N, it can be obtained from Het and can be transferred to Dia. And it can also be consumed by biosynthesis of Veg and Het ( $\mu(Q_N^V + Q_N^H)$ ).

$$\frac{dQ_C^V}{dt} = F_{pho}^V + J_C^{DV} - J_C^{VH} - \mu(Q_C^V + Q_C^H)(1 + E) \quad \text{Equation S7}$$

$$\frac{dQ_N^V}{dt} = J_N^{HV} - J_N^{VD} - \mu(Q_N^V + Q_N^H) \quad \text{Equation S8}$$

Equation S9 and Equation S10 are cellular C and N dynamics ( $\frac{dQ_C^D}{dt}$  and  $\frac{dQ_N^D}{dt}$ ) in Dia. C can be obtained from photosynthesis in Dia ( $F_{pho}^D$ ). And it can be consumed by transfer to Veg ( $J_C^{DV}$ ) and biosynthesis (including growth and respiration) of Dia ( $\mu Q_C^D(1 + E)$ ). As for N, it can be obtained from Veg ( $J_N^{VD}$ ) and from environmental  $NH_4^+$  uptake ( $V_{NH_4^+}$ ). And it can also be consumed by biosynthesis of Dia ( $\mu Q_N^D$ ).

$$\frac{dQ_C^D}{dt} = F_{pho}^D - J_C^{DV} - \mu Q_C^D(1 + E) \quad \text{Equation S9}$$

$$\frac{dQ_N^D}{dt} = J_N^{VD} - \mu Q_N^D + V_{NH_4^+} \quad \text{Equation S10}$$

After we get all of the above equations, we assume a steady state, which means that all of the element dynamic terms ( $\frac{dy}{dt}$ ) are zero. Then we sum up Equation S5, Equation S7 and Equation S9, and we get Equation S11, which is the C balance equation that we used in main text equation (1). And we also sum up Equation S6, Equation S8 and Equation S10, and get Equation S12, which is the N balance equation that we used in main text equation (3).

$$F_{pho}^D + F_{pho}^V = \mu(Q_C^V + Q_C^H + Q_C^D)(1 + E) + F_C^{N_2fix} \quad \text{Equation S11: the same as Equation (1) in the main text}$$

$$F_N^{N_2fix} + V_{NH_4^+} = \mu(Q_N^V + Q_N^H + Q_N^D) \quad \text{Equation S12: the same as Equation (3) in the main text}$$

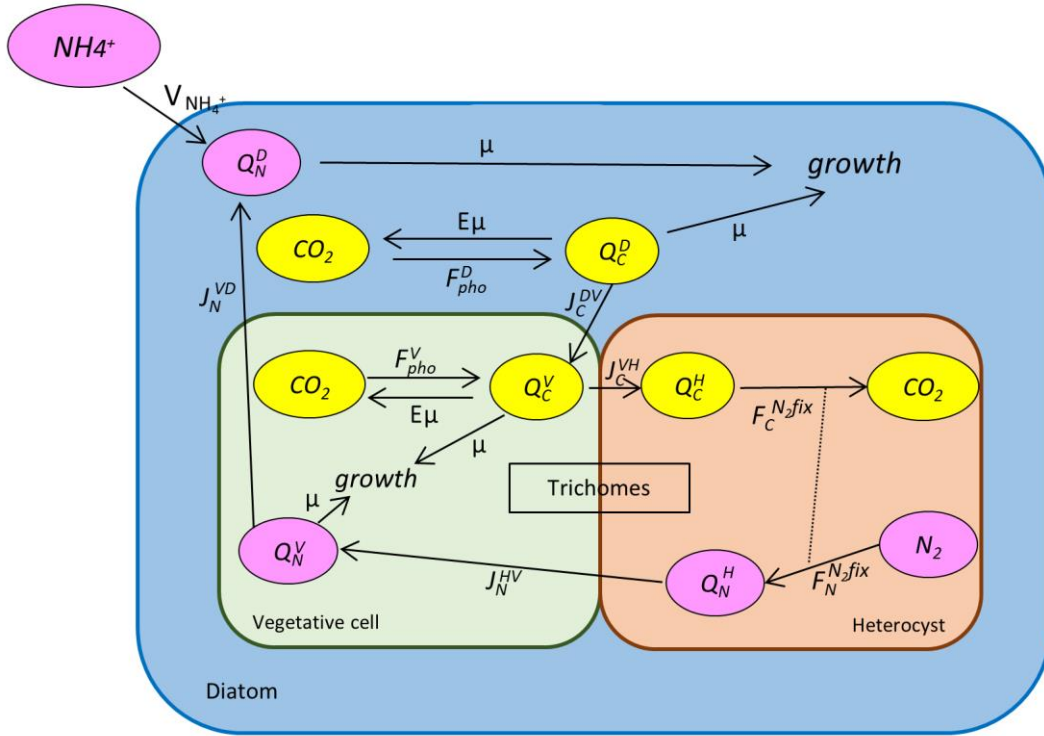


Figure S1. A schematic of the DDA model with flux notations. Blue frame and space: the host diatom. Green space and green frame: Vegetative cell in trichomes. Brown frame and light brown space: Heterocyst in trichomes. Yellow ovals: C pools. Pink ovals: N pools. Black arrows: The direction of element flux when the  $NH_4^+$  supply is not

enough for N consumption in the host diatom cell. Red arrows: Direction change when  $\text{NH}_4^+$  supply is more than the diatom consumption. Dash line: these are coupling between the processes. See Supplementary Methods for flux notations.  $E\mu$  represents  $E \times \mu$ , growth-dependent respiration.

**Light intensity.** Here we describe the equation (Equation S13) using light intensity as a variable in the calculation of photosynthesis. We assumed that total photosynthesis rate ( $F_{pho}^D + F_{pho}^V$ ) can increase with light intensity ( $I$ ) but becomes saturated when it reaches a maximum ( $F_{cfix}^{max}$ , this is the maximum photosynthesis rate for the whole DDA). Here,  $A_i$  represents the light saturation coefficient. We got  $F_{cfix}^{max}$  and  $A_i$  values from a previous modeling paper [2]. Given DDAs are often observed near the surface, we assumed strong enough light to allow saturation for both diatom and vegetative cells. And we also tested 50% maximum photosynthesis for vegetative cells to see if the difference of light-harvesting systems between diatom and cyanobacteria can change the results.

$$F_{pho}^D + F_{pho}^V = F_{cfix}^{max} (1 - e^{-A_i I}) \quad \text{Equation S13}$$

The simulation results of light effect are shown in Figure S1. Here, we found that light intensity can only influence C pathways (Figure S2). With increasing light intensity, photosynthesis rate and C transfer increase (Figure S2 a, c, e, f). However, N pathways are not influenced by light intensity (Figure S2 b, d). And the light effect is nearly identical when we tested 50% maximum photosynthesis for vegetative cells (Figure S3).

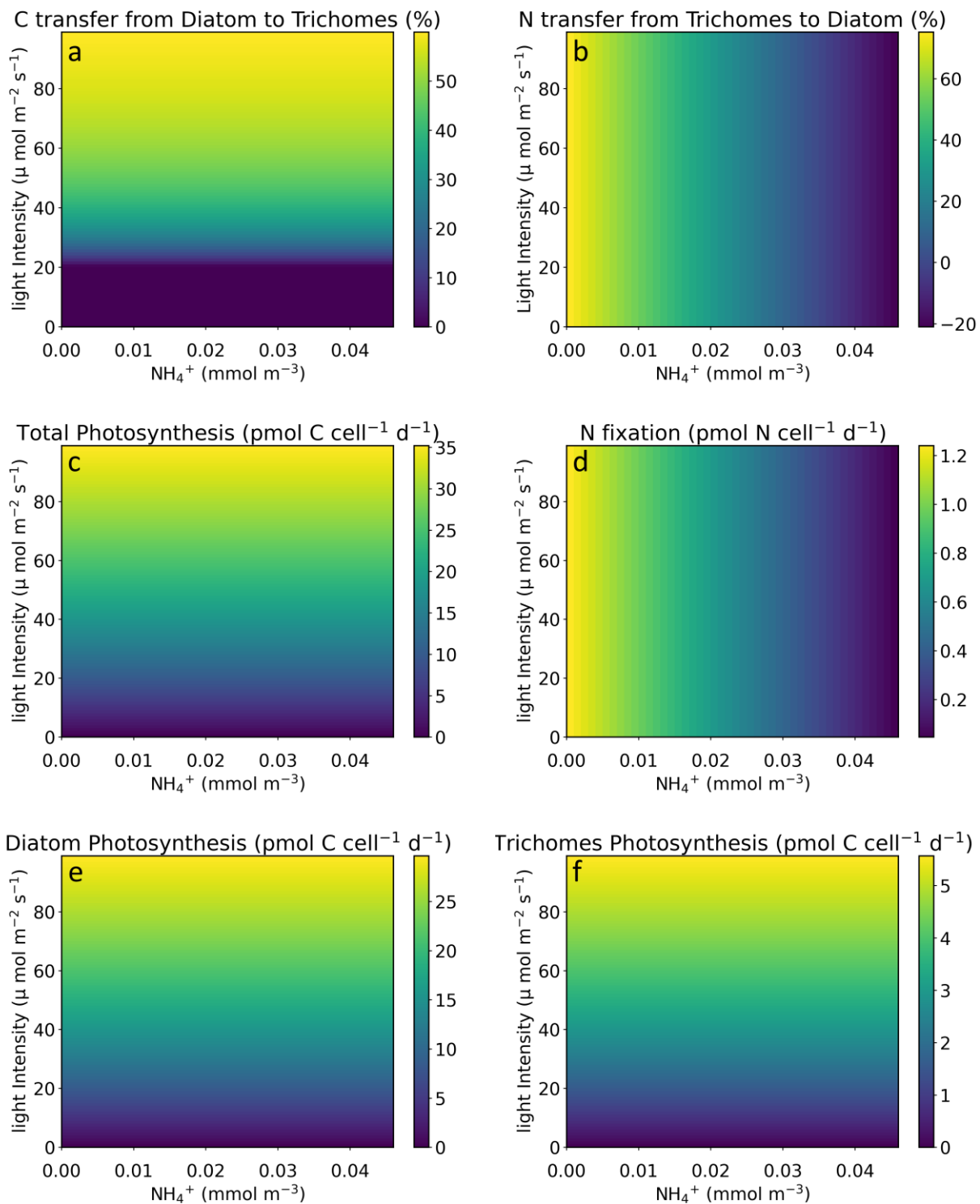


Figure S2. Simulated influence of the  $\text{NH}_4^+$  concentration and light intensity on element transfer and supply. (a) Effect of  $\text{NH}_4^+$  and light intensity on C transfer. (b) Effect of  $\text{NH}_4^+$  and light intensity on N transfer. (c), (e), (f)

Effect of  $\text{NH}_4^+$  and light intensity on photosynthesis, including total DDA, diatom and trichomes. (d) Effect of  $\text{NH}_4^+$  and light intensity on  $\text{N}_2$  fixation. For (a) and (b), the unit percentage means how much C and N transfer account for the total C and N consumption (or supply).

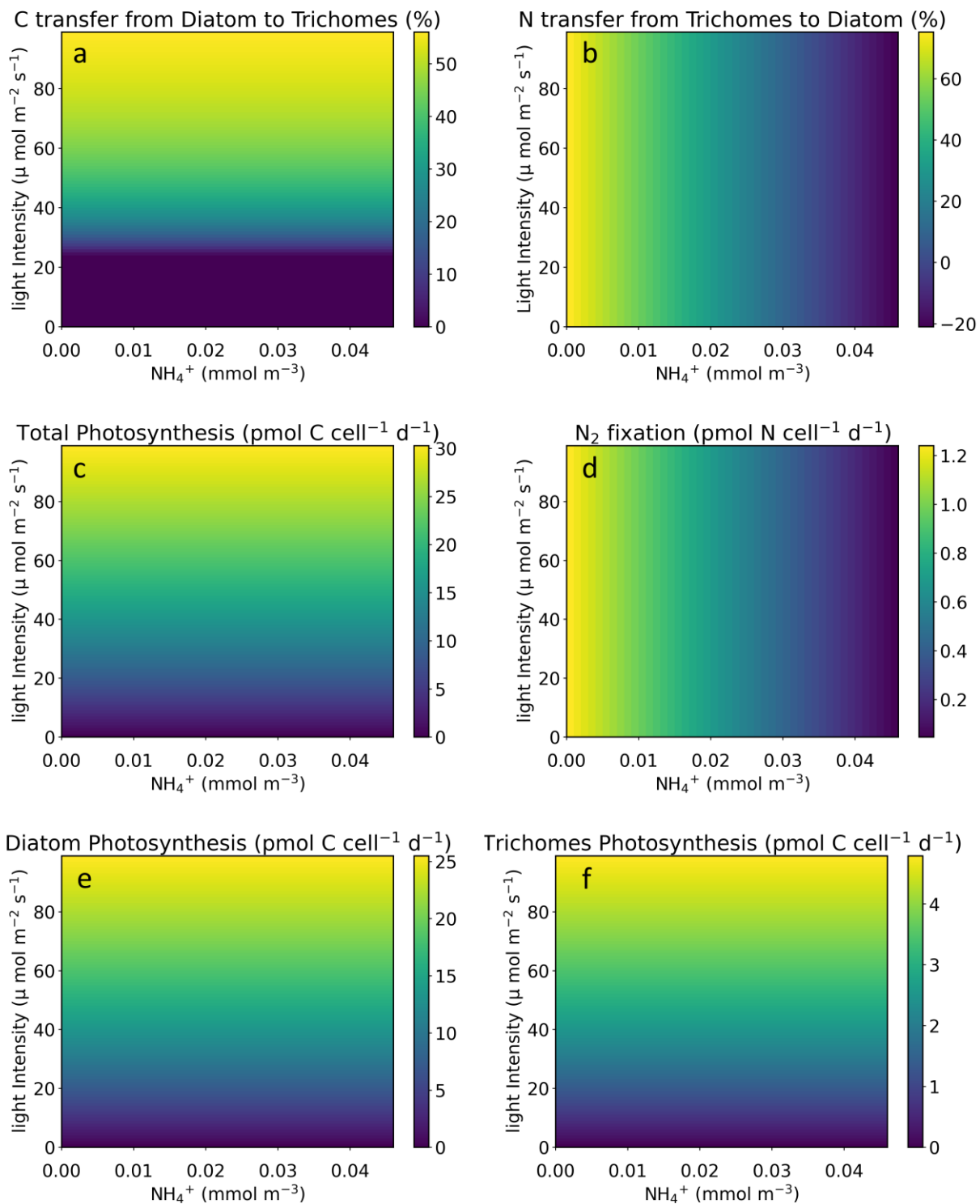


Figure S3. Sensitivity test: Simulated influence of the  $\text{NH}_4^+$  concentration and light intensity on element transfer and supply (50% maximum photosynthesis for vegetative cells). (a) Effect of  $\text{NH}_4^+$  and light intensity on C transfer. (b) Effect of  $\text{NH}_4^+$  and light intensity on N transfer. (c), (e), (f) Effect of  $\text{NH}_4^+$  and light intensity on photosynthesis,



including total DDA, diatom and trichomes. (d) Effect of  $\text{NH}_4^+$  and light intensity on  $\text{N}_2$  fixation. For (a) and (b), the unit percentage means how much C and N transfer account for the total C and N consumption (or supply).

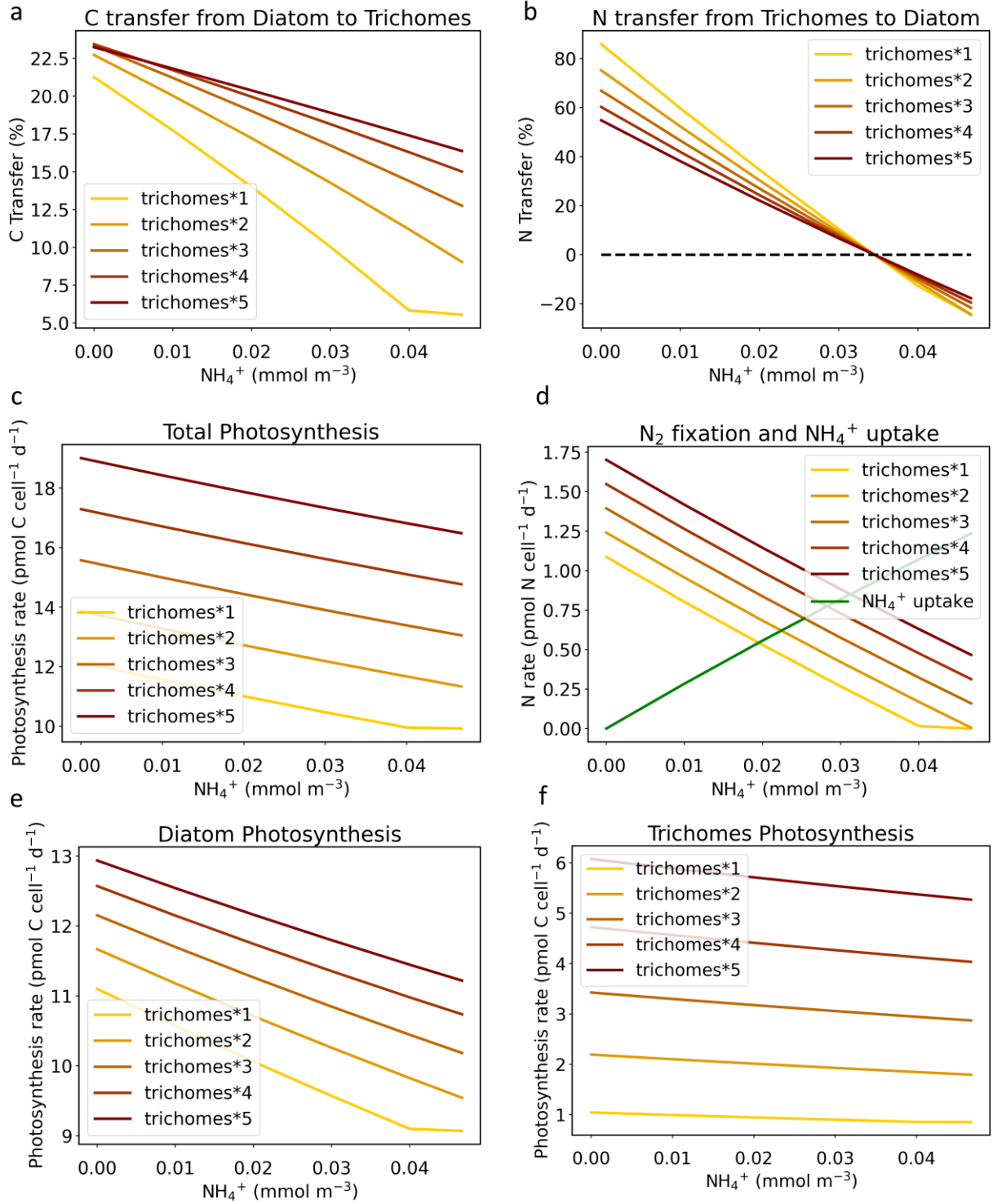


Figure S4. Simulated influences of the  $\text{NH}_4^+$  concentration and trichomes numbers on element transfer and supply.

(a) Effect of  $\text{NH}_4^+$  and trichomes number on C transfer. (b) Effect of  $\text{NH}_4^+$  and trichomes numbers on N transfer, the dash line is the no transfer  $\text{NH}_4^+$  concentration. (c), (e), (f) Effect of  $\text{NH}_4^+$  and trichomes numbers on photosynthesis, including total DDA, diatom and trichomes. (d) Effect of  $\text{NH}_4^+$  and trichomes number on  $\text{NH}_4^+$  uptake and  $\text{N}_2$  fixation. The different color lines mean different number of trichomes. For (a) and (b), the unit percentage means how much C and N transfer account for the total C and N consumption (or supply).

**Trichomes numbers.** Here, we showed the effect of the number of trichomes on C and N transfer, photosynthesis and  $\text{N}_2$  fixation.

**Growth rate.**

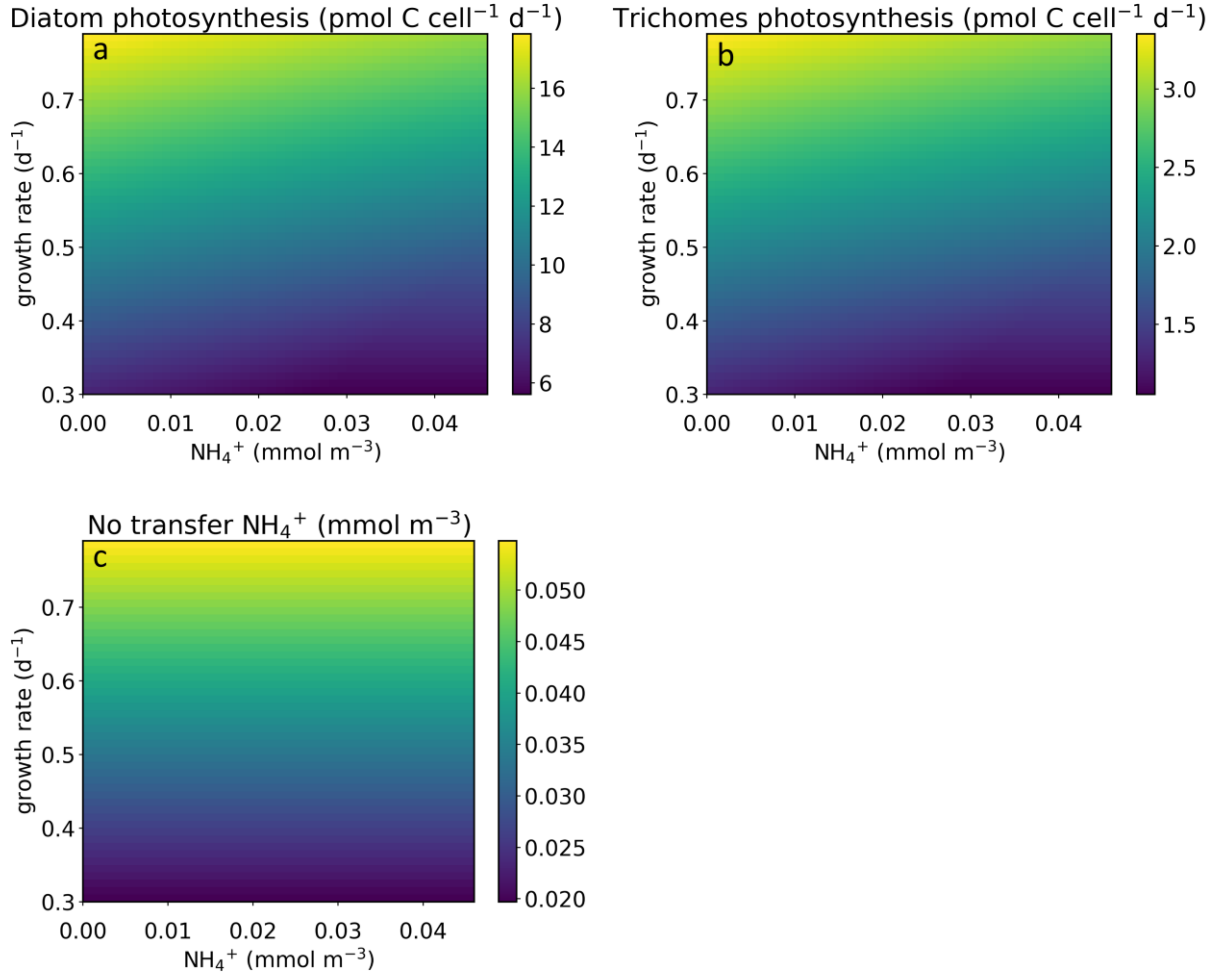


Figure S5. Simulated influences of the  $\text{NH}_4^+$  concentration and growth rate on photosynthesis and the no transfer  $\text{NH}_4^+$ . These are supplementary figures for the main text growth rate figures (main text Figure 3).

## Parameter table

Table S1. Parameters, units, and definitions for all equations

Parameter	Unit	Definition
$F_{pho}^D$	$\text{pmol C d}^{-1} \text{ cell}^{-1}$	Diatom photosynthesis rate

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$F_{pho}^V$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	Vegetative cells photosynthesis rate
$\mu$	d <sup>-1</sup>	DDA growth rate
$Q_C^V$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	cellular C quotas of vegetative cells per DDA
$Q_C^H$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	cellular C quotas of heterocysts per DDA
$Q_C^D$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	cellular C quotas of diatom per DDA
E	dimensionless unit	The ratio of respiration to biosynthesis
$F_C^{N_2fix}$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	C usage in N <sub>2</sub> fixation
$F_N^{N_2fix}$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	N usage in N <sub>2</sub> fixation
$F_{C:N}^{N_2fix}$	pmol C pmol N <sup>-1</sup>	C to N cost ratio in N <sub>2</sub> fixation
$V_{NH_4^+}$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	Diatom NH <sub>4</sub> <sup>+</sup> uptake rate
$Q_N^V$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	cellular N quotas of vegetative cells per DDA
$Q_N^H$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	cellular N quotas of heterocysts per DDA
$Q_N^D$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	cellular N quotas of diatom per DDA
$[NH_4^+]$	mmol m <sup>-3</sup>	NH <sub>4</sub> <sup>+</sup> concentration

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$V_{NH_4^+}^{max}$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	maximum NH <sub>4</sub> <sup>+</sup> uptake rate
$K_m$	mmol m <sup>-3</sup>	Half saturation concentration
$F_{Cfix}^{max}$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	Maximum photosynthesis rate
$A_i$	μ mol <sup>-1</sup> m <sup>2</sup> s	Light saturation coefficient
$I$	μ mol m <sup>-2</sup> s <sup>-1</sup>	Light intensity
$\frac{dQ_C^H}{dt}$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	Cellular C dynamics in heterocysts
$\frac{dQ_N^H}{dt}$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	Cellular N dynamics in heterocysts
$\frac{dQ_C^V}{dt}$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	Cellular C dynamics in vegetative cells
$\frac{dQ_N^V}{dt}$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	Cellular N dynamics in vegetative cells
$\frac{dQ_C^D}{dt}$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	Cellular C dynamics in diatoms
$\frac{dQ_N^D}{dt}$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	Cellular N dynamics in diatoms
$J_C^{VH}$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	C transfer from vegetative cells to heterocysts
$J_N^{HV}$	pmol N d <sup>-1</sup> cell <sup>-1</sup>	N transfer from heterocysts to vegetative cells
$J_C^{DV}$	pmol C d <sup>-1</sup> cell <sup>-1</sup>	C transfer from diatoms to vegetative cells

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 $J_N^{VD}$

pmol N d<sup>-1</sup> cell<sup>-1</sup>N transfer from vegetative cells to diatoms

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**Sensitivity test for changing the respiration to biosynthesis value**

In our model, we assume that the ratio of respiration to biosynthesis (E) is constant for diatoms, heterocysts, and vegetative cells. To make our conclusions more convincing, we also did a sensitivity test to explore whether our results would be different if we used different values for the three types of cells. We used the original E value (0.38) for diatoms and heterocysts, and doubled the E value for vegetative cells. The results are in Figure S6, Figure S7, Figure S8, Figure S9. Comparing the results in these figures: Figure S6 and main text Figure 2, Figure S7 and main text Figure 3, Figure S8 and Figure S4, we believe that the results are almost the same. And compared to main text Figure 5, Figure S9 shows higher C transfers, but the conclusion is still the same. In conclusion, this sensitivity test shows that using different E values does not influence the overall conclusion from the simulation results. Therefore, using the same value is fine.

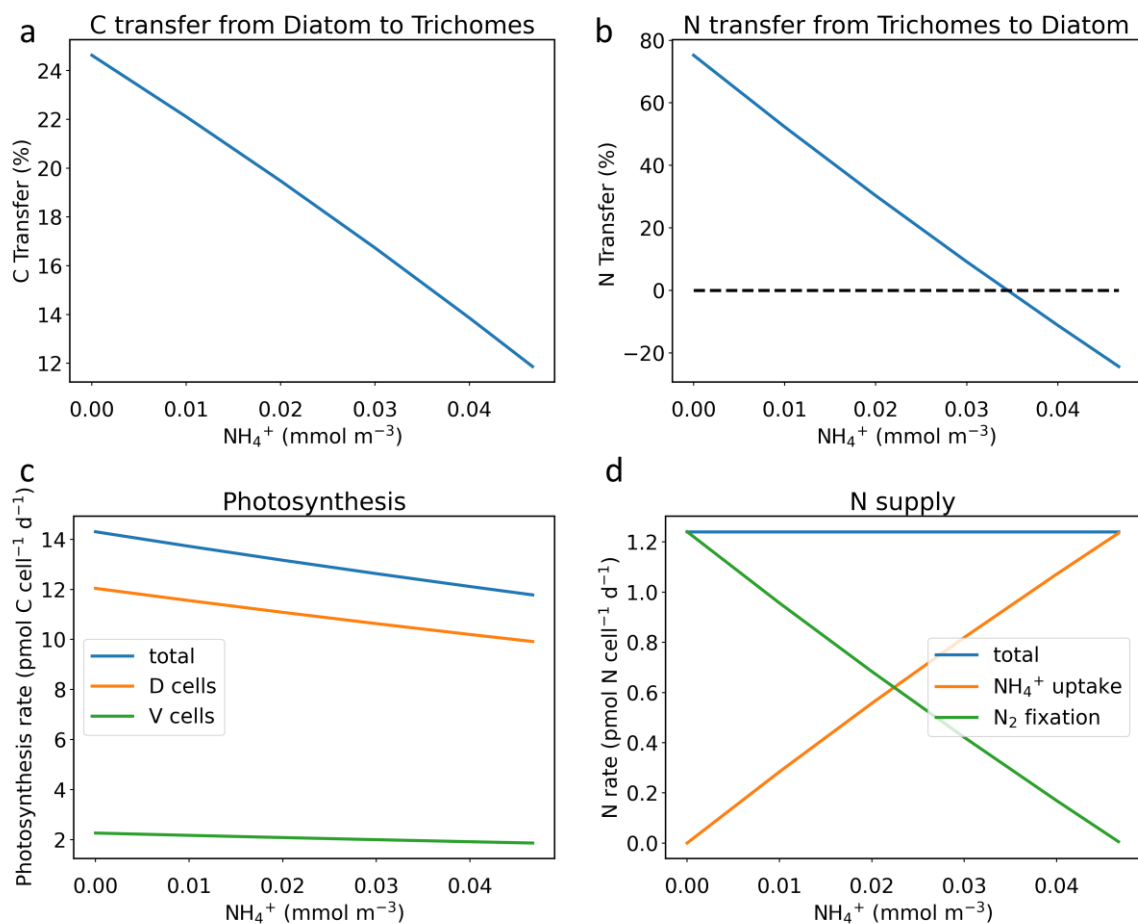


Figure S6. Sensitivity test: Simulated influences of the  $\text{NH}_4^+$  concentration on element supply and transfer. (a) Effect of  $\text{NH}_4^+$  on C transfer. (b) Effect of  $\text{NH}_4^+$  on N transfer, the dash line is *no transfer  $\text{NH}_4^+$  concentration*. (c) Effect of  $\text{NH}_4^+$  on photosynthesis, blue line is photosynthesis change in DDA, orange line is photosynthesis change in the host diatom, and green line is photosynthesis change in vegetative cell. (d) Effect of  $\text{NH}_4^+$  on N supply, blue line is total N supply change, orange line is  $\text{NH}_4^+$  uptake change, and green line is  $\text{N}_2$  fixation change. For (a) and (b), the unit percentage means how much C and N transfer account for the total C and N consumption (or supply).

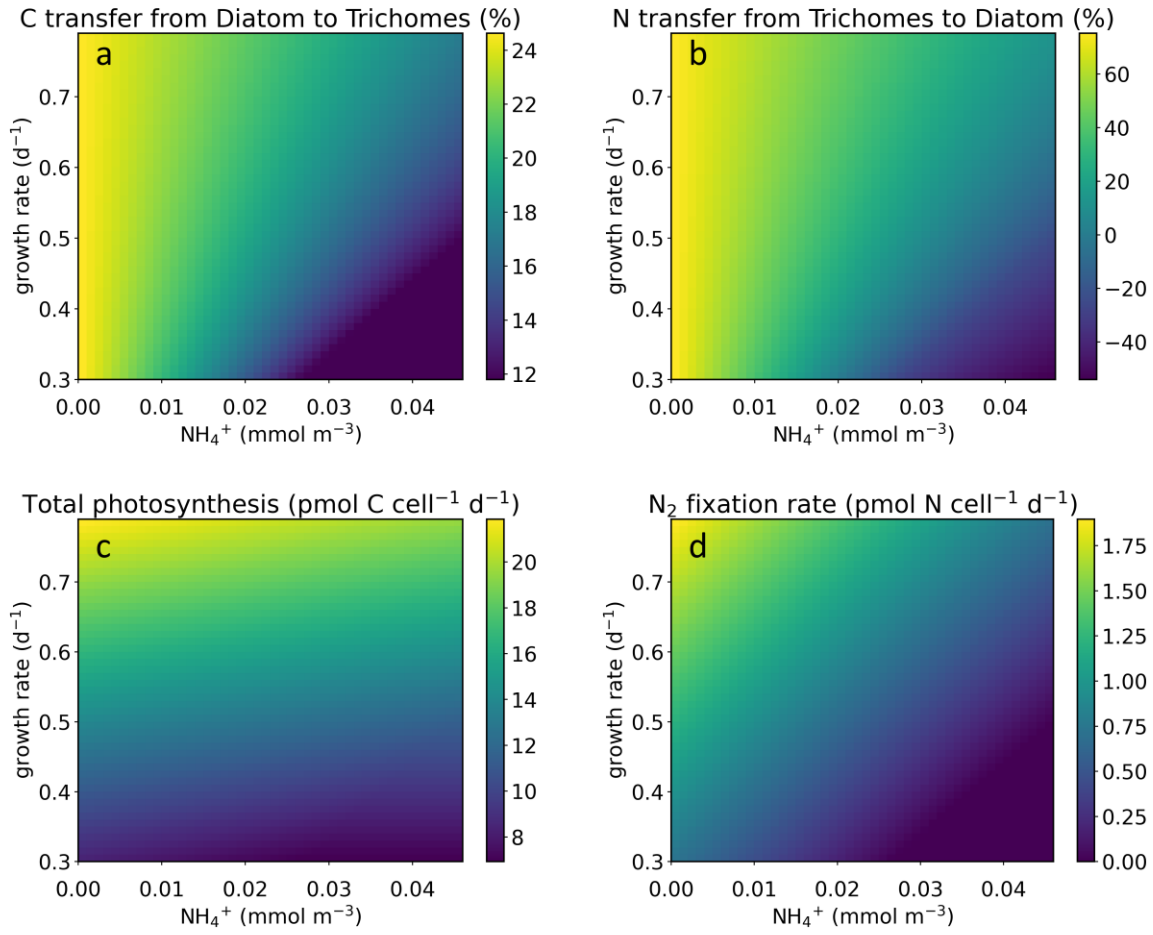


Figure S7. Sensitivity test: Simulated influence of the  $\text{NH}_4^+$  concentration and growth rate on element transfer and supply. (a) Effect of  $\text{NH}_4^+$  and growth rate on C transfer. (b) Effect of  $\text{NH}_4^+$  and growth rate on N transfer. (c) Effect of  $\text{NH}_4^+$  and growth rate on photosynthesis. (d) of  $\text{NH}_4^+$  and growth rate on  $\text{N}_2$  fixation. For (a) and (b), the unit percentage means how much C and N transfer account for the total C and N consumption (or supply).



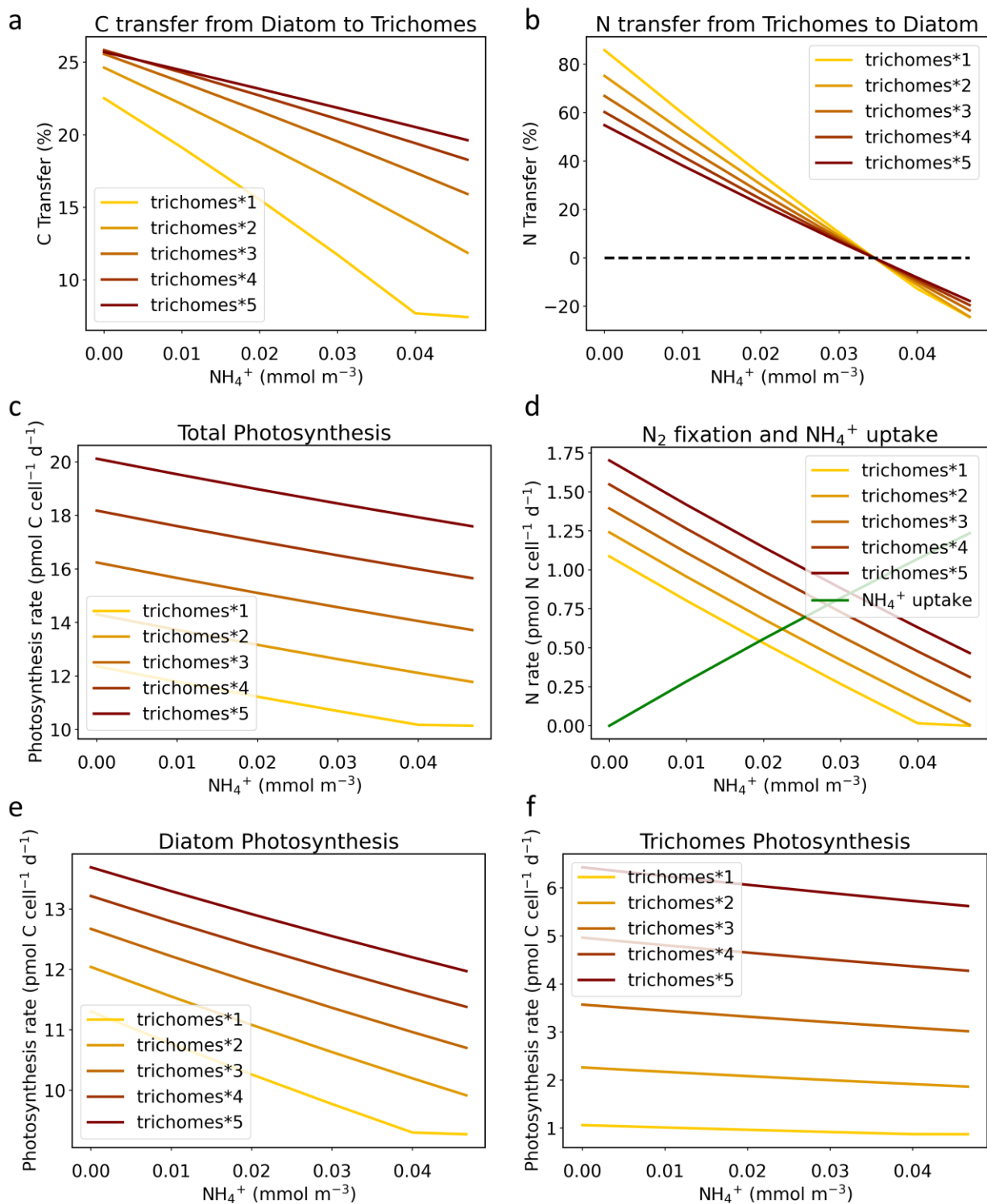


Figure S8. Sensitivity test: Simulated influences of the  $\text{NH}_4^+$  concentration and trichomes numbers on element transfer and supply. (a) Effect of  $\text{NH}_4^+$  and trichomes number on C transfer. (b) Effect of  $\text{NH}_4^+$  and trichomes numbers on N transfer, the dash line is no transfer  $\text{NH}_4^+$  concentration. (c), (e), (f) Effect of  $\text{NH}_4^+$  and trichomes numbers on photosynthesis, including total DDA, diatom and trichomes. (d) Effect of  $\text{NH}_4^+$  and trichomes number

on  $\text{NH}_4^+$  uptake and  $\text{N}_2$  fixation. The different color lines mean different number of trichomes. For (a) and (b), the unit percentage means how much C and N transfer account for the total C and N consumption (or supply).

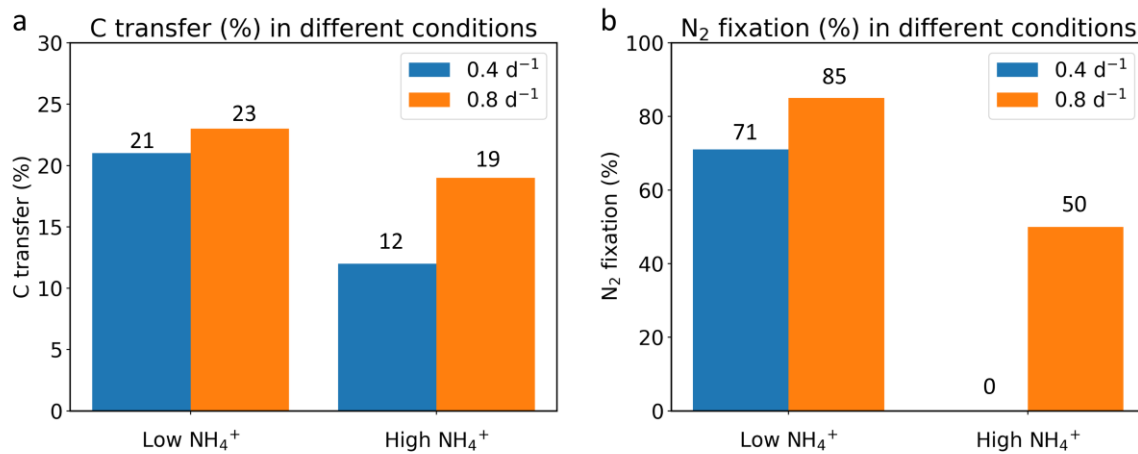


Figure S9. Sensitivity test: C transfer and  $\text{N}_2$  fixation in different conditions: low  $\text{NH}_4^+$ , high  $\text{NH}_4^+$ , low growth rate (blue bars), high growth rate (orange bars). (a) C transfer. (b)  $\text{N}_2$  fixation.

## References

1. Inomura, K.; Follett, C.L.; Masuda, T.; Eichner, M.; Prášil, O.; Deutsch, C. Carbon Transfer from the Host Diatom Enables Fast Growth and High Rate of  $\text{N}_2$  Fixation by Symbiotic Heterocystous Cyanobacteria. *Plants* **2020**, *9*, 8–16, doi:10.3390/plants9020192.
2. Inomura, K.; Wilson, S.T.; Deutsch, C. Mechanistic Model for the Coexistence of Nitrogen Fixation and Photosynthesis in Marine Trichodesmium. *mSystems* **2019**, *4*, 1–13.