

Review



Novel Balance Mechanism Participates in Stem Cell Therapy to Alleviate Neuropathology and Cognitive Impairment in Animal Models with Alzheimer's Disease

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Abstract: Stem cell therapy improves memory loss and cognitive deficits in animal models with Alzheimer's disease. The underlying mechanism remains to be determined, but it may involve the interaction of stem cells with hippocampal cells. The transplantation of stem cells alters the pathological state and establishes a novel balance based on multiple signaling pathways. The new balance mechanism is regulated by various autocrine and paracrine cytokines, including signal molecules that target (a) cell growth and death. Stem cell treatment stimulates neurogenesis and inhibits apoptosis, which is regulated by the crosstalk between apoptosis and autophagy—(b) A β and tau pathology. Aberrant A β plaques and neurofibrillary tau tangles are mitigated subsequent to stem cell intervention—(c) inflammation. Neuroinflammation in the lesion is relieved, which may be related to the microglial M1/M2 polarization—(d) immunoregulation. The transplanted stem cells modulate immune cells and shape the pathophysiological roles of immune-related genes such as TREM2, CR1, and CD33—(e) synaptogenesis. The functional reconstruction of synaptic connections can be promoted by stem cell therapy through multi-level signaling, such as autophagy, microglial activity, and remyelination. The regulation of new balance mechanism provides perspective and challenge for the treatment of Alzheimer's disease.

Keywords: Alzheimer's disease; stem cell therapy; neurogenesis; synaptogenesis; astrocyte; microglia; autophagy; apoptosis; immunoregulation; neuroinflammation

1. Alzheimer's Disease and Stem Cell Therapy

Alzheimer's disease (AD) is a neurodegenerative disorder, characterized by memory decline and cognitive impairment. In pathology, AD is manifested with A β peptide plaques, neurofibrillary tau tangles, neuronal death, synaptic alterations, and cerebral atrophy [1,2]. The etiology of AD is complicated by the diversity of risk factors, such as heredity, aging, infection, immunity, medicines, environmental pollutants, and sociopsychological factors [3–5]. Certain diseases have been considered as predisposing factors for AD, such as hypothyroidism, immune-related disease, virus infection, epilepsy, depression, and schizophrenia. The early onset AD locus is located on chromosomes 21, 14, and 1, while the late-onset AD locus is on chromosome 19 [6]. The expression of typical genes such as APP, S182, STM-2 and APOE is linked with the pathogenesis of AD [7,8]. Most sporadic AD may be the result of the interaction between genetic susceptibility and environmental factors. The development of AD is associated with the comprehensive effects of various mechanisms such as oxidative stress, apoptosis, autophagy, immunity, inflammation, cholesterol metabolism, and angiogenesis [9]. Aberrant A β deposits and



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). neurofibrillary tau aggregates induce neuronal death and synaptic loss. Some genes such as APOE4, ABCA7 and SLC24A4 are related to cholesterol metabolism that is implicated with P-tau trafficking [10,11]. Hence, statin drugs can decrease neurofibrillary tangle burden by competitively inhibiting HMG-CoA reductase [12]. The disturbance of neurotransmitters is connected with the clinical manifestations of AD, including acetylcholine system, monoamine system, and neuropeptides [13,14]. Immunoregulation plays an important role in the neuronal loss of patients with AD. The allelic variants of microglial TREM2 cannot control the balance between the formation and phagocytosis of A β proteins in the brain, increasing the risk of AD by nearly three times [15,16]. Free radical generation and oxidative stress cause neuronal apoptosis, which are related to the onset of AD as well [17].

The medical treatment of AD remains a challenge. Presently, only a few medicines have certain effects, including (a) acetylcholinesterase inhibitors such as donepezil, galantamine, rivastigmine, and tacrine [18]. They can compensate for the cholinergic decline by inhibiting acetylcholine turnover, (b) NMDA receptor antagonist memantine, and (c) $A\beta$ -directed monoclonal antibody aducanumab. They target $A\beta$ peptides to reduce their accumulation in the brain [19]. Other compounds that can reduce amyloid plaques, neurofibrillary tangles and neuroinflammation have been evaluated in clinical trials as well [20,21]. So far, no medications have been demonstrated surely to stop or delay the progression of AD. Stem cell therapy as a novel strategy has also been explored in animal models with AD (Figure 1). Research results prove that the transplantation of stem cells can improve memory and learning abilities, which can function in the AD-like animal models as reflected by extended effectiveness or longer life expectancy [9,22]. Despite the encouraging progress, therapeutic effect is expected to continue for the remaining life. Therefore, significant improvements are needed to enhance efficiency. The transplanted stem cells can proliferate and transdifferentiate, which compensate for neuronal loss and restore synaptic connection. The therapeutic mechanisms are essentially associated with neurogenesis and synaptogenesis. The source of stem cells may be autologous, allogenic, or iPS-derived [9,23–26]. Autologous stem cells can be isolated and purified from brain, fat, dental pulp, or bone marrow. In contrast, allogenic stem cells may be prepared from placenta, umbilical cord, or embryonic tissue. Additionally, the delivery methods affect the therapeutic effect of stem cells [9]. Different approaches have been compared based on feasibility and accessibility, but their therapeutic efficiency remains under investigation. It is confirmed that the transplanted stem cells can repair cognitive impairment and improve behavioral performance in AD-like animal models as demonstrated by Morris water maze test, Y-maze alternation test, plus-maze discriminative avoidance task, social recognition test, and open-field evaluation [24,27,28].



Figure 1. Stem cell therapy for animal models with Alzheimer's disease. The transplantation of stem cells can stimulate the secretion of autocrine and paracrine cytokinesis, which alters microenvironment and promotes neurogenesis as well as synaptogenesis. As a result, stem cell therapy alleviates neuropathology and improves behavioral performance in animal models with Alzheimer's disease.

2. Participant Cell Types of New Balance Mechanism

In the human brain, there are approximately 86 billion neurons and about the same number of non-neuronal glia cells [29]. The ratios of neurons to glia vary from one region to another. The transplantation of stem cells affects a variety of cell types such as neurons, oligodendrocytes, astrocytes, and microglia in the hippocampus. Intercellular interactions are regulated by different signal pathways, which bring about a series of pathophysiological changes and develop a novel balance.

- (1). Physical pressure. The local pressure of cerebral tissue can be increased after the stem cells are delivered through the intrahippocampal injection, but the same phenomenon is not found via the peripheral delivery. Nonetheless, the local physical pressure caused by mechanical force is almost negligible, since a similar therapeutic effects can be obtained through tail vein delivery as well [9,30,31].
- (2). Signaling molecules. The transportation of stem cells alters the microenvironment of cerebral tissue and stimulates the secretion of autocrine and paracrine cytokines, such as chemokines, leucocyte chemoattractant factors, transcription factors, inflammatory cytokines, fibrogenic cytokines, and growth factors (Table 1). Some factors are general products that can be secreted by all types of stem cells, whereas other cytokines are only produced by specific stem cells [32]. Those pragmatic cytokines participate in the

establishment of new balance mechanisms. The secretion of autocrine and paracrine cytokines plays important roles in neurogenesis and synaptogenesis.

- (3). Changes in various cell types (Figure 2, Table 2).
 - (a) Functional neurons play a central role in the brain. There are roughly 20 billion neurons in the human cortex. Each neuron has an average 7000 synaptic connections [33]. The number of synapses is relatively stabilized in adulthood. Neuronal synapses may decrease with aging, but they can also increase due to brain plasticity. The transplanted stem cells can stimulate neurogenesis and synapse formation. Newborn neurons may be from (i) the transdifferentiation of stem cells; and (ii) the activation of specialized multipotent stem cells in the brain. At present, stem cell therapy has overcome the concerns of uncertainty and safety, and its effectiveness has been validated as well.
 - (b) Oligodendrocyte is a specific subtype of neuroglia. In the central nervous system, their branch structures wrap around the neuronal axons to form an insulating myelin sheath. The physiological function of oligodendrocytes is to maintain neuronal insulation during the excitement of nerve signals. The complete structure of the myelin sheath provides a safety measure for signal transmission among neuronal synapses. Stem cell therapy restores neuronal networks by way of synaptogenesis that is protected by the myelin sheaths from oligodendrocytes [34,35].
 - (c) Astrocytes, also called astroglia, have projections covering local neurons. Astrocytes are the support system in the cerebral tissue to hold neurons in the position. Additionally, they can produce cytokines and interact with other cell types. For example, astrocytes participate in microglia-mediated inflammatory and immune processes [36]. Astrocytes are responsible for substance exchange. In the CNS, astrocytes contact both capillaries and neurons to transport nutrients. Moreover, the phagocytosis of astrocytes is implicated in the amyloid load of Alzheimer disease [37]. In the process of stem cell therapy, the precise roles of astrocytes are still unclear. After exposure to MSC-conditioned medium, the expression of pro-inflammatory factors such as IL-1 β , TNF- α and IL-6 was attenuated in cultured astrocytes [38]. The transplanted stem cells acted on astrocytes to modify neuroimmune and relieve neuroinflammation in vivo [39].
 - (d) Microglia are resident immune cells in the brain, equivalent to macrophages. Functional microglia take part in the neuroinflammation, immunomodulation, the elimination of $A\beta$ proteins, and tau pathology. As the first line of the neuroimmune system, microglia remove cerebral debris and protect neurons from harmful invasion. In contrast, the inflammatory factors released by microglia can cause receptor-induced neuronal apoptosis [40]. Fortunately, microglial activity can be modulated by the transplanted stem cells. So, stem cell therapy suppresses neuroinflammation and controls neuroimmune overreaction. Furthermore, microglia can detect neuronal injury and play a critical role in the maintenance of neuronal health. As immune cells, microglia have duality in the pathogenesis of AD. They can not only protect neurons by engulfing detrimental $A\beta$ proteins, but also damage neurons by secreting inflammatory cytokines [41–43]. The consequence may be beneficial or pernicious, which is determined by the comprehensive effect of multi-level signaling crosstalk.

Types	Examples	Function	References
Inflammatory cytokines	TNFα, IL-1, IL-2, IL-6, IL-10	To regulate inflammatory and immune responses, to participate in the regulation of cell growth and apoptosis, etc.	J. Clin. Endocrinol. Metab. 1998 Jun;83(6):2043–51; Immunotherapy. 2018 Sep;10(12):1053–1064;
Fibrogenic cytokines	FGF, TIMP-1	Proliferation of fibroblasts, collagen synthesis and extracellular fibrosis, immune mediators.	PLoS ONE. 2019 Apr 22;14(4):e0215678; Brain Res. 2004 Apr 16;1005(1–2):21–8.
Chemokines	CCL5, CXCL-10, CXCL-12,	Chemo-attractants, to guide the migration of cells, to regulate immunity, inflammation, angiogenesis, etc.	Stem Cells. 2012 Jul;30(7):1544–55; Cancer Res. 2011 Jun 1;71(11):3831–40; J. Cell Physiol. 2019 Aug;234(10):18707– 18719
Leucocyte chemoattractant factors	CINC-1, G-CSF, SCF, GM-CSF	To participate in im- mune/inflammatory cascade.	PLoS ONE. 2019 Apr 22;14(4):e0215678; Blood. 2000 Nov 15;96(10):3422–30.
Transcription factors	GATA-4, Nkx2.5, MEF2C	Response to intercellular and extracellular signals, transcriptional regulation in development, cell cycle, and pathogenesis.	Mol. Med. Rep. 2015 Aug;12(2):2607–21; Tissue Eng. Part A. 2011 Jan;17(1–2):45–58.
Growth factors	HGF, IGF-1	Signaling molecules promote cell differentiation and maturation.	Stem Cells Dev. 2010 Jul;19(7):1035–42; Int. J. Stem Cells. 2009 May;2(1):59–68.
Vascular endothelial growth factor	VEGF	To stimulate the formation of blood vessels.	Int. J. Stem Cells. 2009 May;2(1):59–68; Brain Res. 2004 Apr 16;1005(1–2):21–8.
Other	MCP-1, OPG	Selectively recruiting monocytes, to regulate bone metabolism.	Int. J. Stem Cells. 2009 May;2(1):59–68; J. Interferon Cytokine Res. 2009 Jun;29(6):313–26; Cell. 1997 Apr 18;89(2):309–19.

 Table 1. Autocrine and paracrine cytokines secreted by stem cells.

Mechanisms	Cell Types	Signaling Pathways	References
Immunoregulation	Neurons, Microglia, Astrocytes, Oligodendrocytes	To facilitate microglial M1/M2 polarization; to regulate the crosstalk between T cells and microglia; to mediate synaptic plasticity.	Neuroscience. 2019 Dec 1;422:99–118; Proc. Natl. Acad. Sci. USA. 2006 Mar 28;103(13): 5048–5053; Front. Synaptic Neurosci. 2018 Jun 13;10:14.
Inflammation	Neurons, Microglia, Astrocytes, Oligodendrocytes	To decrease the level of NF- κ B in astrocytes; to reduce the levels of TNF- α , IL-6, and MCP-1; to regulate cell growth and apoptosis.	Neuropathol. Appl. Neurobiol. 2017 Jun;43(4):299–314; Sci. Rep. 2020 Jul 1;10(1):10772; DOI:10.1186/s13024- 015-0035-6.
Neurogenesis	Neurons, Microglia, Astrocytes, Oligodendrocytes	To increase IGF-1 expression in the hippocampus; to increase N-acetylaspartate and Glutamate; to induce the expression of synaptophysin.	Exp. Ther. Med. 2017 Nov; 14(5): 4312–4320; Transl. Neurodegener. 2020 May 27;9(1):20; Hippocampus. 2017 Dec;27(12):1250–1263
Autophagy	Neurons, Microglia, Astrocytes, Oligodendrocytes	To increase cellular viability and LC3-II expression; to upregulate BECN1/Beclin 1 expression; to enhance mitophagy.	Autophagy. 2014 Jan;10(1):32–44; Mol. Neurobiol. 2019 Dec;56(12):8220–8236; Autophagy. 2021 Jan 19;1–20.
Apoptosis	Neurons, Microglia, Astrocytes, Oligodendrocytes	To regulate expression of hippocampal SIRT1, PCNA, p53, ac-p53, p21, and p16; to target caspase pathway; Ca ²⁺ signaling.	Behav. Brain Res. 2018 Feb 26;339:297–304; Front. Neurosci. 2018 May 22;12:333; Curr. Alzheimer Res. 2010 Sep;7(6):540–8; Sci. Rep. 2016 Aug 12;6:31450.
Angiogenesis	Neurons, Microglia, Astrocytes, Oligodendrocytes	BMSCs secrete VEGF, BDNF, NT-3, IGF-1, bFGF, GDNF and TGF. VEGF is the most important mitogen in the process of angiogenesis.	Brain Res. 2011 Jan 7; 1367:103–113; Int. J. Mol. Med. 2013 May;31(5):1087–96; Neuroreport. 2015 May 6;26(7):399–404.
Synaptogenesis	Neurons, Microglia, Astrocytes, Oligodendrocytes	To stimulate the production of BDNF and NGF for remyelination; peptide FG loop (FGL) amplifies remyelination and modulates neuroinflammation.	Cell Biol. Int. 2021 Feb;45(2):432–446; J. Neuroimmune. Pharmacol. 2016 Dec;11(4):708–720; Front. Cell Dev. Biol. 2021 Jul 2;9:680301.

 Table 2. New balance mechanism in the hippocampus involves multiple signaling pathways.



Figure 2. Participant cell types of a new balance mechanism. The transplantation of stem cells alters the pathological state and establishes a novel balance in the brain, which involve multiple signaling pathways such as neurogenesis, autophagy, apoptosis, inflammation, immunoregulation, the removal of aberrant proteins, neuroglial interaction, and angiogenesis. All cell types in the hippocampus participate in the establishment of the new balance mechanism. The therapeutic benefit of stem cells depends on the comprehensive effect of multi-level signaling crosstalk.

3. Representative Signaling Pathways of New Balance Mechanism

The transplantation of stem cells alters the pathological state and stimulates the secretion of autocrine and paracrine cytokines in the hippocampus. The remodeling process establishes a novel balance related to multiple signaling pathways. The new balance is an essential mechanism to improve the neuropathology and recognitive deficits of Alzheimer's disease, which has been validated by regulating representative pathways.

3.1. The Transplantation of Stem Cells Mediates Cell Growth and Death

The transplanted stem cells can survive in the hippocampus and further transdifferentiate into neurons as demonstrated in APP/PS1 transgenic mice [24,26]. Meanwhile, some newborn neurons may be derived from endogenous progenitors, which have been detected in C57BL/6 mice as well as in the tissue culture of a patient's cortex [44,45]. More details of in vivo conditions still need to be verified on the patient. Further, the beneficial cytokines produced by MSCs can stimulate proliferation through the indirect regulation of neurotrophic factors such as NGF, FGF2 and BDNF [46]. The comprehensive effect of transplanted stem cells is to promote neuronal growth or neurogenesis. Generally, the development of DA is presented with long-term and gradual characteristics, accompanying neuronal apoptosis/necroptosis/necrosis. Apoptosis is an important way of neuronal death, especially at the early stage of AD. Apoptosis is initiated in a controlled environment. Apoptotic body may be promptly removed via phagocytosis. Thereupon, histopathological changes are slight or insignificant. Tissue biopsy may be the only way to confirm the apoptosis in most cases. Perhaps, this is the reason that the low rate of apoptosis is observed in some stages. Apoptotic cell death rarely exhibits acute features such as inflammatory necrosis caused by microbial infection or thrombosis. The transplanted stem cells stimulate neurogenesis and inhibit apoptosis-related neuron death [9,47]. Besides, stem cell therapy decreases the generation of ROS and alleviates ROS-induced neuronal damage. Interestingly, short-term ROS exposure promotes the proliferation of neural progenitors whereas persistent ROS stimulation aggravates oxidative stress and neuronal apoptosis [48]. The transplanted stem cells may control the dynamic equilibrium between ROS generation and elimination, thereby regulating neurogenesis. In clinical, the oxidative damage in the advanced AD is very severe, leading to neuronal loss and cognitive decline [49]. The transplanted stem cells activate autophagy in AD-like animal models. The activation of autophagy is reflected by the upregulation of BECN1/Beclin 1 and the increased number of LC3-II-positive autophagosomes in the hippocampus, which boosts the clearance of AB peptides and the relief of oxidative stress [17]. Autophagy is a key mechanism to promote neurogenesis as demonstrated by the expression levels of signal molecules such as Beclin-1, atg5, LC3-II, and mTOR. The proliferation of neural progenitor cells in adult hippocampus is regulated by the PI3K/AKT/mTOR and ERK1/2 signaling pathways [50–52]. There is a crosstalk between autophagy and apoptosis (Figure 3). The induction of autophagy is begun while Beclin-1 is dissociated at the BH3-only domain of Bcl-2 proteins subsequent to the phosphorylation of Bcl-2. Activated autophagy alleviates neuronal apoptosis by altering the levels of IAPs, Bcl-2, caspase-8 and so forth. The autophagic response can be balanced by caspase activation. Activated caspase-8 cleaves Beclin-1 into C-terminal and Nterminal fragments to trigger apoptosis [53]. The cell fate is modified by the interaction of diverse BH3 proteins with Beclin-1 and caspase-8 [54]. The beneficial effect of transplanted stem cells may be through the upregulation of BECN1/Beclin-1, the modulation of Bcl-2 family, and the inhibition of caspase activity [22,55]. The crosstalk between autophagy and apoptosis modulates the therapeutic effect of transplanted stem cells. A synergistic effect may be acquired when the transplanted stem cells is combined with autophagic and/or apoptotic mediators.



Figure 3. Crosstalk between autophagy and apoptosis. Cell fate is regulated by the interaction between autophagy and apoptosis. There is a crosstalk between apoptosis and autophagy by sharing common regulators, such as p53, Atg5, caspase-8, Beclin-1/Bcl-2, and IAPs. Cellular FLIP inhibits caspase 8 and autophagosome formation that is mediated by LC3 conjugation. Autophagosome promotes the activation of caspase 8 through the platform consisting of ATG5, LC3 and p62. Bcl-2 family involves both autophagy and apoptosis by regulating signal molecules such as Beclin1 and BAX/BAK dimer. The activation of autophagy can degrade IAPs to facilitate apoptosis. Activated caspase-3 causes apoptosis but suppresses autophagy. The red line represents the inhibitory effect.

3.2. The Transplanted Stem Cells Regulate the Production and Removal of Aberrant Proteins

There are aberrant A β proteins and tau aggregates in the brain. Both A β plaques and tau tangles are increased with advanced age and/or genetic factors. The buildup of two proteins is associated with the pathogenesis of Alzheimer's disease, although the causal connection remains to be determined. Owing to the hindrance of $A\beta$ metabolic pathway with aging, the production of A β proteins, especially insoluble A β proteins, is more than their degradation. The $A\beta$ peptides form plaques to deposit in the brain of patients with AD. Genetic modification demonstrated that the down-regulation of Becn-1 increased extracellular A β deposition, whereas the high expression of Beclin-1 decreased the A β pathology in APP transgenic mice [54,56,57]. The reversal relationship provides the mechanistic link between autophagic Beclin-1 expression and cytotoxic Aß deposits. Aß proteins is derived from γ -secretase-hydrolyzed APP [58,59]. Simultaneously, γ -secretase also activates Notch receptors for A β metabolism [60,61]. Aging weakens the activation of the Notch signaling pathway and leads to the accumulation of hydrolyzed APP, which is closely related to the pathogenesis of AD. In addition, there is evidence that aberrant A β proteins can inhibit the PI3K/Akt signaling pathway and autophagic activity [62,63]. Cytotoxic Aß proteins can induce the apoptosis of primary cultured neurons. Furthermore, the injection of A β proteins into the hippocampus produces AD-like manifestations in animal models, showing similar changes to AD patients [64]. The accumulated A β proteins launch apoptotic, necroptotic, and necrotic mechanisms. Aβ-mediated cytotoxicity causes irreversible damage during cell maturation, which impairs neurogenesis by decreasing the survival rate of newborn neurons [65,66]. As a consequence, the integration of newly generated neurons into the hippocampal circuitry is decreased, resulting in the decline in learning and memory capabilities. Immunotherapy with antibodies targeting Aß proteins have been explored in clinical trials [67]. Obviously, aberrant A β deposits and weak neurogenesis are related to the pathogenesis of AD. Aging as a risk factor complicates the metabolism of AD-associated A β proteins [66]. Meta-analysis revealed that the transplantation of stem cells could decrease A β plaques in the hippocampus of APP/PS1 mice, which promoted the functional improvement of AD-like animals [9]. Sometimes, stem cell therapy cannot significantly decrease Aβ plaques in certain studies. Furthermore, certain drugs diminish A β protein load but may not ameliorate memory loss and cognitive deficits. Thus, the theory of $A\beta$ pathology is controversial. Neurofibrillary aggregates are formed by the hyperphosphorylation of microtubule-associated protein tau. Tau tangles are composed of tubular filaments, paired helical filaments, and hyperphosphorylated tau protein, which are associated with the decreased autophagy [68–70]. The intracellular accumulation of tau tangles can cause ER stress-induced apoptosis, but tau hyperphosphorylation may also induce apoptotic escape and initiates neurodegeneration [48,68,71]. The expression of JNK in the hippocampus and cortex of AD patients was exceedingly increased [72,73]. In rapidly aging mice with AD, the JNK cascade was dramatically higher than that in normal mice [74]. JNK may involve the regulation of tau protein via oxidative stress. The inhibition of JNK phosphorylation can decrease the level of phospho-tau proteins. AD-like tau pathology and cognitive impairment are exacerbated by reducing insulin/GSK-3β signaling activity [75]. Tau hyperphosphorylation and the CaM-CaMKIV signal pathway participate in the recovery of memory ability in AD-like rats [76]. The transplantation of stem cells can lower tau aggregates and inhibit neuronal apoptosis. Moreover, reduced tau tangles are beneficial to both young and aged AD-like animals [9,77,78]. The improvements of the aforementioned neuropathology are related to the enhancement of autophagy [79,80]. Clearly, stem cell therapy not only facilitates the elimination of aberrant proteins, but also prevents their formation. These are two different aspects that transplanted stem cells can deal with.

3.3. The Transplanted Stem Cells Can Produce Pro- and Anti-Inflammatory Cytokines

Inflammation is a response to a variety of stimuli such as infection, toxic metabolites, and autoimmunity. The initiation of neuroinflammation may be a protective action, but the actual consequence leads to harmful tissue damage. The triggers of neuroinflammation can be cytokines, metabolites, or aberrant A β proteins. A lot of evidence supports that neuroinflammation is an independent factor affecting the different stages of AD. Inflammatory cytokines, small molecular proteins secreted by glial cells in the brain, are key factors by binding to corresponding receptors on the cell surface. It was found that thirteen pro-inflammatory cytokines in patients with AD, including IL-1 β , IL-6, IL-18, TNF- α and so on, were significantly higher than those in the normal control [81,82]. Conversely, some anti-inflammatory cytokines play a protective effect in the pathogenesis of AD. For instance, IL-10 is the primary product of active monocytes. Its functions include phagocytosis, the expression of Th1 cytokines, the regulation of costimulatory molecules, and MHC class II antigen presentation [83]. IL-10 can inhibit inflammation by blocking the cytotoxicity of pro-inflammatory cytokines. The IL-10/STAT3 signal pathway can be regulated to rebalance the natural immunity in the brain, which may bring about beneficial effects on neuroinflammation [84]. The signal components in the classic IL-10 pathway are up-regulated in the hippocampus of AD patients. Besides, the cytokines IL-2 and IL-4 have anti-inflammatory effects similar to IL-10 [40,85]. Distinctly, inflammatory cytokines have protective and harmful effects. The NF- κ B signal pathway is related to inflammation, oxidative stress, and apoptosis in the brain [86]. The cerebral levels of BACE1 and NF- κ B p65 are markedly enhanced in patients with AD. Anti-inflammatory drugs or stem cell therapy can block the transcription of BACE1 as well as the production of A β , suggesting that the inhibition of NF- κ B-mediated BACE1 expression is the plausible target of AD treatment [87]. There are complicated interactions among hippocampal cells such as astrocytes, neurons, and microglia (Figure 4). Neurons are functional carriers in the brain, implicated in the inflammatory response by producing $A\beta$ deposits and tau tangles [88]. Meanwhile, neurons are also targets that need to be protected during neuroinflammation. Astrocytes provide support, protection, and nutrient supply to neurons under physiological conditions. Active astrocytes can secrete inflammatory cytokines, such as RANTES, MIP-1 α , MCP-1 and complement, to participate in the neuroinflammation [36,89]. The transplanted stem cells may suppress inflammation caused by astrocytes [90]. Pro-inflammatory factors such as IL-1 β , TNF- α , and IL-6 was decreased in cultured astrocytes following exposure to MSC-conditioned medium [38]. Microglia are the innate immune cells of the central nervous system. There are fine-tuning mechanisms for microglia to protect cerebral neurons. They can remove aberrant $A\beta$ protein plaques. Additionally, microglia are able to maintain neuronal connections as well as modulate the electrical activity. Microglial activity regulates neuronal function and vice versa. However, the above-mentioned relationship is interrupted in the pathogenesis of AD. The dysfunction of neuronal conduction is a prominent feature, leading to cognitive deficits. The pathogenic role of microglia in development of AD is demonstrated by genetic mutations [91]. The abnormal interaction between neuronal and microglial activities is engaged in the active cycle that deteriorates cognitive impairment. Microglial activation has a duality in the pathogenesis of AD. They protect neurons by engulfing detrimental substances and attack neurons by secreting inflammatory cytokines. The dual role of microglia may be due to the polarization of M1/M2 phenotype [92,93]. The classic M1 state can be activated by A β deposits to produce pro-inflammatory factors such as TNF- α , IL-1 β , IFN- γ , thereby exacerbating inflammatory cell death [42,43]. The M2 phenotype may generate anti-inflammatory factors such as IL-2, IL-4 or IL-10, facilitating cell repair and neuroprotection [94–96]. In the APP/PS1 transgenic models, the profiles of gene expression are overlapped between microglial M1 and M2 types. Accordingly, the exact role of microglia has not yet been determined. Both the beneficial and detrimental effects of microglia can be fulfilled in the pathogenesis of AD. Available data demonstrate that the transplanted stem cells take part in the regulation of immune and inflammatory processes. After the administration of stem cells, microglial activation stimulates the removal of $A\beta$ deposits and neuroinflammation is thereupon alleviated. Consequently, stem cell therapy can suppress inflammation. Furthermore, stem cells can recruit peripheral monocytes across blood-brain barrier into the lesion. The activation of the newly recruited monocytes can further accelerate the clearance of $A\beta$ peptides as well as apoptotic bodies. This phenomenon seems contradictory, but it does happen. Still, many intermediate details need to be clarified through future research. Nevertheless, the comprehensive effect of stem cell therapy is conducive to the improvement of neuropathology as well as cognitive impairment in Alzheimer's disease.



Figure 4. Regulation of inflammatory and neuroimmune responses. The transplanted stem cells inhibit neuroinflammation and participate in immunoregulation. Moreover, peripheral monocytes can be recruited to accelerate the removal of aberrant proteins. The signaling pathways of new balance mechanism form a complex network, but inflammatory/immune processes are key regulators to determine neurogenesis and synaptogenesis, which play a critical role in the pathogenesis of Alzheimer's disease.

3.4. Immunoregulation Is Modulated by the Transplanted Stem Cells

The CNS is immunologically privileged, since peripheral immune cells are usually blocked by the blood-brain barrier composed of astrocytes and endothelial cells. Pathological studies have revealed that viral, bacterial, and fungal infections are related to the pathogenesis of Alzheimer's disease. For example, HSV-1 DNA was found within amyloid plaques [97]. Borrelia burgdorferi bacterium caused Lyme neuroborreliosis and dementia [98]. The diffuse mycosis was related to the development of Alzheimer's disease. Further studies proved that fungal infections could occur in different brain regions of patients with AD, but are absent in the control individuals [99]. The pathogenesis of Alzheimer's disease may be partly explained by the microbial infection of CNS due to immunodeficiency, but this pathogen hypothesis needs more evidence to confirm the causality. It is well known that APOE4 and TREM2 variants associated with the development of AD may be susceptible to HSV-1 infection [100,101]. Another possibility is that both gene variants and HSV-1 infection are related to the pathogenesis of AD. In addition, the immune system decreases its protective capacity with aging. Advanced age (e.g., over 59 years old) significantly increased the mortality in patients with Alzheimer's disease after SARS-CoV-2 infection [102,103]. Therefore, aging is a predominant risk factor related to AD [104]. The dysfunction of the immune system in the brain is demonstrated by the partial mutations of TREM2 and CD33 genes [15,105]. In patients with AD, aberrant A β proteins activate T cells, perpetuating the cycle of immune-mediated cell injury and repair [106]. The neuroimmune

and immunoregulation are the basic targets of understanding the pathogenesis of AD. The activation of microglia participates in immunoregulation in the pathogenesis of AD. As innate immune cells in the brain, microglia have functions similar to macrophages and can be activated in response to microbial infections and toxic metabolites. Their effects on immunoregulation had been verified using the CRISPR knockout method [107]. Microglia protect the brain and maintain neuronal health by removing aberrant A β plaques as well as apoptotic bodies. Microglia contain the M1/M2 phenotype, playing a dual role in the pathogenesis of AD. Therefore, only immunosuppression or immunoenhancement cannot acquire beneficial effects on the development of AD. Perhaps, the damaged neuroimmune in the AD brain needs to be rebalanced. The IL-10/JAK1/STAT3 signaling pathway can regulate the establishment of immunobalance in the brain [84,108]. When bone marrow stem cells were transplanted into immunodeficient mice with AD, the transfused stem cells could restore missing immune cells for the elimination of A β plaques [109,110]. The transplanted stem cells can (a) inhibit microglial activation and neuroinflammation and (b) recruit peripheral monocytes across the blood-brain barrier into the lesion. These monocytes may switch the microglial M1/M2 phenotype to accelerate the removal of A β plaques in the AD brain [111-113] and (c) secret cytokines. Certain cytokines released by MSCs can facilitate cell survival and proliferation through the regulation of NGF, FGF2 and BDNF [46]. The transplanted stem cells promote neurogenesis and inhibit neurodegenerative cell death. Of note, a variety of autocrine and paracrine factors produce distinct functions. Some cytokines take part in relevant pathways to relieve neuropathology, but other factors are competitors or bystanders. Therefore, the pathophysiological roles of autocrine and paracrine factors should be scrutinized in future studies. Moreover, the expression of immune-related genes is modulated by transplanted stem cells, including TREM2, CR1, HLA-DRB5, CD33, MS4A, INPP5D, EPHA1, and CLU (Table 3). These genes influence the different stages of AD and play a crucial role in the pathogenesis of AD. The dysfunction of immune-related genes can be corrected by stem cell therapy, which has been demonstrated in AD-like models [9,109,110]. Microarray analysis and high-throughput gene sequencing have confirmed the gene profiles. Evidently, immune factors do participate in the pathogenesis of Alzheimer's disease. The immunoregulation can effectively alleviate neuropathology and improve cognitive function. Noticeably, the transplanted stem cells are neither immunosuppressant nor immunostimulant, but they function as a regulator or controller that balances the neuroimmune response to maintain neuronal health.

3.5. The Transplanted Stem Cells Participate in Synaptic Plasticity

The change of neuronal synapses is pivotal pathway to the new balance mechanism. Patients with AD show a decrease in the number of synapses. After stem cell treatment, the favorable improvement is verified by increasing the number of synapses [23,26,114]. Moreover, the synthesis of neurotransmitters is also enhanced, which is consistent with the effect of neurotransmitter drugs. The enhancement of the quantity and quality of neuron synapses may explain why stem cell therapy can improve the cognitive symptoms of AD-like animal models. The formation of synapses (synaptogenesis) in the nervous system covers the lifespan of healthy individual. This process is an essential requirement for maintaining the normal function of nerve activity. There is a certain degree of synaptic pruning between neurons and synapses through competition for neural growth factors. Therefore, synaptogenesis is regulated by autocrine and paracrine cytokines. The secretion of cytokines establishes a precise relationship between synaptogenesis and microglial activity. Microglia play an important role in protecting neuronal connections and maintaining the integrity of neural circuits. Microglia have a direct role in modulating the electrical activity of neurons. The presence of aberrant proteins and/or toxic factors can damage microglial function. The protective effect of microglia may thus be impaired. At this moment, dysfunctional microglia can hurt synaptic connections. The dysfunction of synaptic networks incur cognitive deficits in Alzheimer's disease. Canonical Wnt signal transduction

involves the early neurodevelopment in the brain and the maturation of the blood–brain barrier. Wnt/ β -catenin signaling regulates synaptic plasticity and the development of acetylcholine receptors, which may be related to the pathophysiology of AD [115,116]. Meanwhile, A β proteins can activate GSK3, thereby promoting the phosphorylation of tau protein as well as reducing the activity of Wnt [117–119]. Previous studies demonstrate that WASP-1 may significantly improve memory and synaptic transmission. The transplantation of stem cells can decrease aberrant A β peptides and tau aggregates to facilitate synaptogenesis. As proved in the iPS cells of AD patient, synaptogenesis is associated with lysosomal vacuolar-type H-ATPase and intracellular Ca²⁺ concentration. The impairment of autophagy inhibits synaptogenesis and neurogenesis [120]. The transplantation of neural stem cells stimulates cellular changes and improves behavioral performance, which may be attributed to the recovery of synaptic connectivity through neurotrophin release (i.e., GAP-43, BDNF) [121]. In addition, endogenous neural regeneration is enhanced by mobilizing the NCAM-derived peptide FG loop to amplify remyelination as well as modulate neuroinflammation [122,123].

Names	Function	References	
TREM2	Transmembrane glycoprotein. To mediate immune and inflammatory responses as microglial receptor.	Neurobiol. Dis. 2020 Nov;145:105072; Neurobiol. Dis. 2019 Jul;127:432–448.	
CR1	To regulate complement cascade and mediate immune adherence as well as phagocytosis.	Stem Cell Res. 2016 Nov;17(3):560–563.	
HLA-DRB5	To encode major histocompatibility complex class II protein involved in immune responses.	Neurol. Genet. 2018 Jan 18;4(1):e211; JAMA Neurol. 2015 Jan;72(1):15–24.	
CD33	Microglial receptor converged on immune-inflammatory pathways.	Neurobiol. Dis. 2019 Jul;127:432–448; Gerontology. 2019;65(4):323–331	
MS4A	Belonging to a class of four-transmembrane spanning proteins.	Aging Cell. 2019 Aug;18(4):e12964.	
INPP5D	At the plasma membrane, the protein hydrolyzes the 5' phosphate and regulates multiple signaling pathways.	EMBO Mol. Med. 2020 Mar 6;12(3):e10606.	
EPHA1	To regulate the developmental of nervous system.	Int. J. Comput. Biol. Drug Des. 2020;13(1):58–70; J. Immunol. 2020 Sep 1;205(5):1318–1322.	
CLU	Diverse functions such as protein chaperoning, apoptosis, complement activation, etc.	Mol Neurodegener. 2015 Jul 16;10:30; Turk J Med Sci. 2015;45(5):1082–6.	

Table 3. Immune-related genes are implicated in the pathogenesis of Alzheimer's disease.

4. Perspective

4.1. Therapeutic Efficiency and Synergistic Effect

The synergistic effect between neurotrophic cytokines and stem cells may increase therapeutic efficiency. The transplantation of stem cells can enhance neurotrophic factors such as BDNF and NGF [34,124–126]. Neurotrophic BDNF is related to the canonical nerve growth in the brain. NGF is a prototypical growth factor, involved in numerous biological processes such as the survival of target neurons, and the regulation of proliferation and neuroimmune. Therefore, the application of therapeutic stem cells may be pretreated with neurotrophic factors to produce synergistic effects.

4.2. Stem Cell Viability

There is a low survival of transplanted stem cells in the recipients, which is a real problem in therapeutic practice. Anyway, it can be improved by activating autophagy in stem cells. Novel strategy may consider that the transplanted stem cells are combined with the nanoparticles of autophagy-enhancing agents and/or apoptosis regulators, especially for the treatment of advanced AD.

4.3. The Improvement of Delivery Methods

Clinically, mannitol infusion is often applied to reduce intracranial pressure. The intraarterial infusion of mannitol can transiently open the blood–brain barrier by loosening tight junctions. This technique can be utilized for stem cell delivery. After the blood– brain barrier is opened, stem cells may be transfused through the peripheral vein instead of intracranial injection. In addition, the intranasal delivery of stem cells can acquire functional improvement in the APP/PS1 models of AD [127].

4.4. Exosomes

In the process of stem cell therapy, the details of exosomes produced by stem cells are unknown [128–130]. It is possible that the exosomes of stem cells stimulate the secretion of autocrine or paracrine cytokines to achieve therapeutic effects. Accordingly, the role of exosomes needs to be clarified through future analysis.

5. Challenges

- The selection of surveillance biomarkers. Currently, monitoring markers (i.e., Aβ42, T-tau and P-tau, or exosomes in cerebrospinal fluid and/or peripheral bloodstream) need to be optimized for the evaluation of therapeutic effects.
- (2). The timeline of the new balance mechanism. Following the transplantation of stem cells, the pathological state is altered and then a new balance is developed. However, it is unsure how long the dynamic reconstruction can be maintained. Perhaps, it is necessary to repeatedly transplant stem cells to obtain reliable therapeutic effects. At this time, it is important to optimize the relevant parameters of stem cell transplantation, including cell concentration, time interval, inoculation position, and delivery method.
- (3). Uncertainty and perplexity. The therapeutic effect of transplanted stem cells involves multiple mechanisms, such as immunomodulation, inflammation, apoptosis, neurogenesis, autophagy, and angiogenesis. The integration of various mechanisms establishes a new balance and brings about beneficial improvements. Nowadays, most of the above-mentioned mechanisms have been investigated and their roles have been elucidated. Nevertheless, the details of relevant mechanisms still need to be explored, such as autophagy and immunomodulation, the interaction between astrocytes and microglia, microglial activation and synaptic remodeling, etc.

In summary, stem cell therapy is beneficial to the improvement of animal models with AD, which is demonstrated by the alleviation of neuropathology and the amelioration of cognitive impairment. The transplantation of stem cells alters regional microenvironment by stimulating the secretion of autocrine and paracrine cytokines, which promotes neurogenesis as well as synaptogenesis. Potential mechanisms are associated with autophagy, apoptosis, the elimination of aberrant proteins, the interaction of different neuroglia, inflammation, and immunoregulation. Those functional activities alter the pathological state and establish a novel balance by integrating multiple signal pathways. The new balance mechanism is the comprehensive effect of multi-level signaling crosstalk in the brain, which not only lays a theoretical foundation for stem cell therapy but also provides perspectives and challenges for the treatment of Alzheimer's disease.

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Abbreviations

AD, Alzheimer's disease; A β , amyloid beta or β -amyloid; APP, amyloid-beta precursor protein; iPS cells, induced pluripotent stem cells; BACE1, β -site APP cleaving enzyme 1; HSV-1, herpes simplex virus type 1; WASP-1, Wnt activating small protein; BDNF, brain-derived neurotrophic factor; NGF, Nerve growth factor; FGF2, fibroblast growth factor 2; TREM2, triggering receptor expressed on myeloid cells 2; CR1, complement receptor type 1; MS4A, membrane-spanning 4A; INPP5D, inositol polyphosphate-5-phosphatase D; EPHA1, ephrin type-A receptor 1; CLU, clusterin; HLA-DRB5, HLA class II histocompatibility antigen, DR beta 5; IL-1 β , interleukin 1 β ; IL-10, interleukin-10; TNF- α , tumor necrosis factor α ; IFN- γ , interferon γ ; GAP-43, growth-associated protein 43; NCAM, neural cell adhesion molecule; MSC, mesenchymal stem cell; CNS, central nervous system; ROS, reactive oxygen species; Bcl-2, B-cell lymphoma 2; LC3, microtubule-associated proteins 1A/1B light chain 3B; LC3-II, lipid modified form of LC3; IGF-1, insulin-like growth factor 1; IAPs, inhibitor of apoptosis proteins.

References

- 1. Boche, D.; Nicoll, J.A.R. Invited Review—Understanding cause and effect in Alzheimer's pathophysiology: Implications for clinical trials. *Neuropathol. Appl. Neurobiol.* **2020**, *46*, 623–640. [CrossRef]
- 2. Sheppard, O.; Coleman, M. Alzheimer's Disease: Etiology, Neuropathology and Pathogenesis. In *Alzheimer's Disease: Drug Discovery*; Huang, X., Ed.; Exon Publications: Brisbane, Australia, 2020.
- Tanzi, R.E.; St George-Hyslop, P.H.; Gusella, J.F. Molecular genetic approaches to Alzheimer's disease. *Trends Neurosci.* 1989, 12, 152–158. [CrossRef]
- 4. Reitz, C.; Mayeux, R. Alzheimer disease: Epidemiology, diagnostic criteria, risk factors and biomarkers. *Biochem. Pharmacol.* 2014, *88*, 640–651. [CrossRef]
- 5. Reitz, C.; Brayne, C.; Mayeux, R. Epidemiology of Alzheimer disease. Nat. Rev. Neurol. 2011, 7, 137–152. [CrossRef]
- Emilien, G.; Maloteaux, J.M.; Beyreuther, K.; Masters, C.L. Alzheimer disease: Mouse models pave the way for therapeutic opportunities. *Arch. Neurol.* 2000, 57, 176–181. [CrossRef]
- Krasemann, S.; Madore, C.; Cialic, R.; Baufeld, C.; Calcagno, N.; El Fatimy, R.; Beckers, L.; O'Loughlin, E.; Xu, Y.; Fanek, Z.; et al. The TREM2-APOE Pathway Drives the Transcriptional Phenotype of Dysfunctional Microglia in Neurodegenerative Diseases. *Immunity* 2017, 47, 566–581.e9. [CrossRef]
- 8. Sandbrink, R.; Hartmann, T.; Masters, C.L.; Beyreuther, K. Genes contributing to Alzheimer's disease. *Mol. Psychiatry* **1996**, *1*, 27–40.
- 9. Qin, C.; Lu, Y.; Wang, K.; Bai, L.; Shi, G.; Huang, Y.; Li, Y. Transplantation of bone marrow mesenchymal stem cells improves cognitive deficits and alleviates neuropathology in animal models of Alzheimer's disease: A meta-analytic review on potential mechanisms. *Transl. Neurodegener.* 2020, *9*, 20. [CrossRef]
- 10. Perdigao, C.; Barata, M.A.; Araujo, M.N.; Mirfakhar, F.S.; Castanheira, J.; Guimas Almeida, C. Intracellular Trafficking Mechanisms of Synaptic Dysfunction in Alzheimer's Disease. *Front. Cell. Neurosci.* **2020**, *14*, 72. [CrossRef]
- Yu, L.; Chibnik, L.B.; Srivastava, G.P.; Pochet, N.; Yang, J.; Xu, J.; Kozubek, J.; Obholzer, N.; Leurgans, S.E.; Schneider, J.A.; et al. Association of Brain DNA methylation in SORL1, ABCA7, HLA-DRB5, SLC24A4, and BIN1 with pathological diagnosis of Alzheimer disease. *JAMA Neurol.* 2015, 72, 15–24. [CrossRef]
- Li, G.; Larson, E.B.; Sonnen, J.A.; Shofer, J.B.; Petrie, E.C.; Schantz, A.; Peskind, E.R.; Raskind, M.A.; Breitner, J.C.; Montine, T.J. Statin therapy is associated with reduced neuropathologic changes of Alzheimer disease. *Neurology* 2007, 69, 878–885. [CrossRef] [PubMed]
- 13. Storga, D.; Vrecko, K.; Birkmayer, J.G.; Reibnegger, G. Monoaminergic neurotransmitters, their precursors and metabolites in brains of Alzheimer patients. *Neurosci. Lett.* **1996**, *203*, 29–32. [CrossRef]
- 14. Yew, D.T.; Li, W.P.; Webb, S.E.; Lai, H.W.; Zhang, L. Neurotransmitters, peptides, and neural cell adhesion molecules in the cortices of normal elderly humans and Alzheimer patients: A comparison. *Exp. Gerontol.* **1999**, *34*, 117–133. [CrossRef]
- 15. Ulland, T.K.; Colonna, M. TREM2—A key player in microglial biology and Alzheimer disease. *Nat. Rev. Neurol.* **2018**, *14*, 667–675. [CrossRef]

- 16. Jonsson, T.; Stefansson, H.; Steinberg, S.; Jonsdottir, I.; Jonsson, P.V.; Snaedal, J.; Bjornsson, S.; Huttenlocher, J.; Levey, A.I.; Lah, J.J.; et al. Variant of TREM2 associated with the risk of Alzheimer's disease. *N. Engl. J. Med.* **2013**, *368*, 107–116. [CrossRef] [PubMed]
- Smith, M.A.; Zhu, X.; Tabaton, M.; Liu, G.; McKeel, D.W., Jr.; Cohen, M.L.; Wang, X.; Siedlak, S.L.; Dwyer, B.E.; Hayashi, T.; et al. Increased iron and free radical generation in preclinical Alzheimer disease and mild cognitive impairment. *J. Alzheimers Dis.* 2010, 19, 363–372. [CrossRef] [PubMed]
- Ghai, R.; Nagarajan, K.; Arora, M.; Grover, P.; Ali, N.; Kapoor, G. Current Strategies and Novel Drug Approaches for Alzheimer Disease. CNS Neurol. Disord. Drug Targets 2020, 19, 676–690. [CrossRef]
- 19. Alexander, G.C.; Emerson, S.; Kesselheim, A.S. Evaluation of Aducanumab for Alzheimer Disease: Scientific Evidence and Regulatory Review Involving Efficacy, Safety, and Futility. *JAMA* 2021, 325, 1717–1718. [CrossRef]
- 20. Long, J.M.; Holtzman, D.M. Alzheimer Disease: An Update on Pathobiology and Treatment Strategies. *Cell* **2019**, *179*, 312–339. [CrossRef]
- 21. Yiannopoulou, K.G.; Papageorgiou, S.G. Current and Future Treatments in Alzheimer Disease: An Update. *J. Cent. Nerv. Syst. Dis.* **2020**, *12*. [CrossRef]
- Shin, J.Y.; Park, H.J.; Kim, H.N.; Oh, S.H.; Bae, J.S.; Ha, H.J.; Lee, P.H. Mesenchymal stem cells enhance autophagy and increase beta-amyloid clearance in Alzheimer disease models. *Autophagy* 2014, 10, 32–44. [CrossRef]
- Nakano, M.; Kubota, K.; Kobayashi, E.; Chikenji, T.S.; Saito, Y.; Konari, N.; Fujimiya, M. Bone marrow-derived mesenchymal stem cells improve cognitive impairment in an Alzheimer's disease model by increasing the expression of microRNA-146a in hippocampus. *Sci. Rep.* 2020, *10*, 10772. [CrossRef] [PubMed]
- Wang, X.; Ma, S.; Yang, B.; Huang, T.; Meng, N.; Xu, L.; Xing, Q.; Zhang, Y.; Zhang, K.; Li, Q.; et al. Resveratrol promotes hUC-MSCs engraftment and neural repair in a mouse model of Alzheimer's disease. *Behav. Brain Res.* 2018, 339, 297–304. [CrossRef] [PubMed]
- 25. Fujiwara, N.; Shimizu, J.; Takai, K.; Arimitsu, N.; Ueda, Y.; Wakisaka, S.; Suzuki, T.; Suzuki, N. Cellular and molecular mechanisms of the restoration of human APP transgenic mouse cognitive dysfunction after transplant of human iPS cell-derived neural cells. *Exp. Neurol.* **2015**, *271*, 423–431. [CrossRef] [PubMed]
- Zhang, W.; Gu, G.J.; Zhang, Q.; Liu, J.H.; Zhang, B.; Guo, Y.; Wang, M.Y.; Gong, Q.Y.; Xu, J.R. NSCs promote hippocampal neurogenesis, metabolic changes and synaptogenesis in APP/PS1 transgenic mice. *Hippocampus* 2017, 27, 1250–1263. [CrossRef]
- Liu, Z.; Wang, C.; Wang, X.; Xu, S. Therapeutic Effects of Transplantation of As-MiR-937-Expressing Mesenchymal Stem Cells in Murine Model of Alzheimer's Disease. *Cell. Physiol. Biochem.* 2015, 37, 321–330. [CrossRef]
- Nasiri, E.; Alizadeh, A.; Roushandeh, A.M.; Gazor, R.; Hashemi-Firouzi, N.; Golipoor, Z. Melatonin-pretreated adipose-derived mesenchymal stem cells efficiently improved learning, memory, and cognition in an animal model of Alzheimer's disease. *Metab. Brain Dis.* 2019, 34, 1131–1143. [CrossRef]
- 29. Van den Hurk, M.; Erwin, J.A.; Yeo, G.W.; Gage, F.H.; Bardy, C. Patch-Seq Protocol to Analyze the Electrophysiology, Morphology and Transcriptome of Whole Single Neurons Derived From Human Pluripotent Stem Cells. *Front. Mol. Neurosci.* 2018, *11*, 261. [CrossRef]
- 30. Bae, J.S.; Jin, H.K.; Lee, J.K.; Richardson, J.C.; Carter, J.E. Bone marrow-derived mesenchymal stem cells contribute to the reduction of amyloid-beta deposits and the improvement of synaptic transmission in a mouse model of pre-dementia Alzheimer's disease. *Curr. Alzheimer Res.* **2013**, *10*, 524–531. [CrossRef]
- 31. Lampron, A.; Pimentel-Coelho, P.M.; Rivest, S. Migration of bone marrow-derived cells into the central nervous system in models of neurodegeneration. *J. Comp. Neurol.* 2013, 521, 3863–3876. [CrossRef]
- Hwang, J.H.; Shim, S.S.; Seok, O.S.; Lee, H.Y.; Woo, S.K.; Kim, B.H.; Song, H.R.; Lee, J.K.; Park, Y.K. Comparison of cytokine expression in mesenchymal stem cells from human placenta, cord blood, and bone marrow. *J. Korean Med. Sci.* 2009, 24, 547–554. [CrossRef]
- 33. Drachman, D.A. Do we have brain to spare? *Neurology* 2005, 64, 2004–2005. [CrossRef]
- 34. Jaldeep, L.; Lipi, B.; Prakash, P. Potential role of NGF, BDNF and their receptors in Oligodendrocytes differentiation from neural stem cell—An in-vitro study. *Cell Biol. Int.* 2020, 45, 432–446. [CrossRef]
- 35. Klein, R.; Mahlberg, N.; Ohren, M.; Ladwig, A.; Neumaier, B.; Graf, R.; Hoehn, M.; Albrechtsen, M.; Rees, S.; Fink, G.R.; et al. The Neural Cell Adhesion Molecule-Derived (NCAM)-Peptide FG Loop (FGL) Mobilizes Endogenous Neural Stem Cells and Promotes Endogenous Regenerative Capacity after Stroke. *J. Neuroimmune Pharmacol.* **2016**, *11*, 708–720. [CrossRef]
- Lian, H.; Litvinchuk, A.; Chiang, A.C.; Aithmitti, N.; Jankowsky, J.L.; Zheng, H. Astrocyte-Microglia Cross Talk through Complement Activation Modulates Amyloid Pathology in Mouse Models of Alzheimer's Disease. J. Neurosci. 2016, 36, 577–589. [CrossRef] [PubMed]
- Couturier, J.; Stancu, I.C.; Schakman, O.; Pierrot, N.; Huaux, F.; Kienlen-Campard, P.; Dewachter, I.; Octave, J.N. Activation of phagocytic activity in astrocytes by reduced expression of the inflammasome component ASC and its implication in a mouse model of Alzheimer disease. *J. Neuroinflamm.* 2016, 13, 20. [CrossRef] [PubMed]
- 38. Schafer, S.; Calas, A.G.; Vergouts, M.; Hermans, E. Immunomodulatory influence of bone marrow-derived mesenchymal stem cells on neuroinflammation in astrocyte cultures. *J. Neuroimmunol.* **2012**, *249*, 40–48. [CrossRef]
- Konttinen, H.; Gureviciene, I.; Oksanen, M.; Grubman, A.; Loppi, S.; Huuskonen, M.T.; Korhonen, P.; Lampinen, R.; Keuters, M.; Belaya, I.; et al. PPARbeta/delta-agonist GW0742 ameliorates dysfunction in fatty acid oxidation in PSEN1DeltaE9 astrocytes. *Glia* 2019, 67, 146–159. [CrossRef] [PubMed]

- 40. Yin, P.; Wang, X.; Wang, S.; Wei, Y.; Feng, J.; Zhu, M. Maresin 1 Improves Cognitive Decline and Ameliorates Inflammation in a Mouse Model of Alzheimer's Disease. *Front. Cell. Neurosci.* **2019**, *13*, 466. [CrossRef]
- Sierksma, A.; Lu, A.; Mancuso, R.; Fattorelli, N.; Thrupp, N.; Salta, E.; Zoco, J.; Blum, D.; Buee, L.; De Strooper, B.; et al. Novel Alzheimer risk genes determine the microglia response to amyloid-beta but not to TAU pathology. *EMBO Mol. Med.* 2020, 12, e10606. [CrossRef]
- 42. Zhang, B.; Wei, Y.Z.; Wang, G.Q.; Li, D.D.; Shi, J.S.; Zhang, F. Targeting MAPK Pathways by Naringenin Modulates Microglia M1/M2 Polarization in Lipopolysaccharide-Stimulated Cultures. *Front. Cell. Neurosci.* **2018**, *12*, 531. [CrossRef]
- Zhao, R.; Ying, M.; Gu, S.; Yin, W.; Li, Y.; Yuan, H.; Fang, S.; Li, M. Cysteinyl Leukotriene Receptor 2 is Involved in Inflammation and Neuronal Damage by Mediating Microglia M1/M2 Polarization through NF-kappaB Pathway. *Neuroscience* 2019, 422, 99–118. [CrossRef]
- 44. Kan, I.; Barhum, Y.; Melamed, E.; Offen, D. Mesenchymal stem cells stimulate endogenous neurogenesis in the subventricular zone of adult mice. *Stem Cell Rev. Rep.* 2011, 7, 404–412. [CrossRef] [PubMed]
- Bavamian, S.; Mellios, N.; Lalonde, J.; Fass, D.M.; Wang, J.; Sheridan, S.D.; Madison, J.M.; Zhou, F.; Rueckert, E.H.; Barker, D.; et al. Dysregulation of miR-34a links neuronal development to genetic risk factors for bipolar disorder. *Mol. Psychiatry* 2015, 20, 573–584. [CrossRef] [PubMed]
- 46. Liu, L.; Cao, J.X.; Sun, B.; Li, H.L.; Xia, Y.; Wu, Z.; Tang, C.L.; Hu, J. Mesenchymal stem cells inhibition of chronic ethanol-induced oxidative damage via upregulation of phosphatidylinositol-3-kinase/Akt and modulation of extracellular signal-regulated kinase 1/2 activation in PC12 cells and neurons. *Neuroscience* 2010, 167, 1115–1124. [CrossRef]
- 47. Singh, P.; Fukuda, S.; Liu, L.; Chitteti, B.R.; Pelus, L.M. Survivin Is Required for Mouse and Human Bone Marrow Mesenchymal Stromal Cell Function. *Stem Cells* **2018**, *36*, 123–129. [CrossRef]
- Xue, F.; Shi, C.; Chen, Q.; Hang, W.; Xia, L.; Wu, Y.; Tao, S.Z.; Zhou, J.; Shi, A.; Chen, J. Melatonin Mediates Protective Effects against Kainic Acid-Induced Neuronal Death through Safeguarding ER Stress and Mitochondrial Disturbance. *Front. Mol. Neurosci.* 2017, 10, 49. [CrossRef]
- Polis, B.; Samson, A.O. Neurogenesis versus neurodegeneration: The broken balance in Alzheimer's disease. *Neural Regen. Res.* 2021, 16, 496–497. [CrossRef]
- 50. Yu, S.; Hei, Y.; Liu, W. Upregulation of seladin-1 and nestin expression in bone marrow mesenchymal stem cell transplantation via the ERK1/2 and PI3K/Akt signaling pathways in an Alzheimer's disease model. *Oncol. Lett.* **2018**, *15*, 7443–7449. [CrossRef]
- 51. Gomez Del Pulgar, T.; De Ceballos, M.L.; Guzman, M.; Velasco, G. Cannabinoids protect astrocytes from ceramide-induced apoptosis through the phosphatidylinositol 3-kinase/protein kinase B pathway. J. Biol. Chem. 2002, 277, 36527–36533. [CrossRef]
- 52. Peltier, J.; O'Neill, A.; Schaffer, D.V. PI3K/Akt and CREB regulate adult neural hippocampal progenitor proliferation and differentiation. *Dev. Neurobiol.* 2007, 67, 1348–1361. [CrossRef]
- 53. Erlich, S.; Mizrachy, L.; Segev, O.; Lindenboim, L.; Zmira, O.; Adi-Harel, S.; Hirsch, J.A.; Stein, R.; Pinkas-Kramarski, R. Differential interactions between Beclin 1 and Bcl-2 family members. *Autophagy* **2007**, *3*, 561–568. [CrossRef]
- Pickford, F.; Masliah, E.; Britschgi, M.; Lucin, K.; Narasimhan, R.; Jaeger, P.A.; Small, S.; Spencer, B.; Rockenstein, E.; Levine, B.; et al. The autophagy-related protein beclin 1 shows reduced expression in early Alzheimer disease and regulates amyloid beta accumulation in mice. J. Clin. Investig. 2008, 118, 2190–2199. [CrossRef]
- 55. Zhang, J.; Ma, K.; Qi, T.; Wei, X.; Zhang, Q.; Li, G.; Chiu, J.F. P62 regulates resveratrol-mediated Fas/Cav-1 complex formation and transition from autophagy to apoptosis. *Oncotarget* **2015**, *6*, 789–801. [CrossRef]
- 56. Lee, J.A.; Gao, F.B. Regulation of Abeta pathology by beclin 1: A protective role for autophagy? *J. Clin. Investig.* **2008**, *118*, 2015–2018. [CrossRef]
- Rocchi, A.; Yamamoto, S.; Ting, T.; Fan, Y.; Sadleir, K.; Wang, Y.; Zhang, W.; Huang, S.; Levine, B.; Vassar, R.; et al. A Becn1 mutation mediates hyperactive autophagic sequestration of amyloid oligomers and improved cognition in Alzheimer's disease. *PLoS Genet.* 2017, 13, e1006962. [CrossRef]
- Wan, Y.; Liang, Y.; Liang, F.; Shen, N.; Shinozuka, K.; Yu, J.T.; Ran, C.; Quan, Q.; Tanzi, R.E.; Zhang, C. A Curcumin Analog Reduces Levels of the Alzheimer's Disease-Associated Amyloid-beta Protein by Modulating AbetaPP Processing and Autophagy. J. Alzheimer's Dis. 2019, 72, 761–771. [CrossRef]
- 59. Esler, W.P.; Wolfe, M.S. A portrait of Alzheimer secretases—New features and familiar faces. *Science* 2001, 293, 1449–1454. [CrossRef] [PubMed]
- 60. Zhou, R.; Yang, G.; Guo, X.; Zhou, Q.; Lei, J.; Shi, Y. Recognition of the amyloid precursor protein by human gamma-secretase. *Science* **2019**, *363*, eaaw0930. [CrossRef] [PubMed]
- Wang, B.J.; Wu, P.Y.; Chen, Y.W.; Chang, Y.T.; Bhore, N.; Wu, P.F.; Liao, Y.F. Quantitative Measurement of gamma-Secretasemediated Amyloid Precursor Protein and Notch Cleavage in Cell-based Luciferase Reporter Assay Platforms. *J. Vis. Exp.* 2018, 56795. [CrossRef]
- 62. Morroni, F.; Sita, G.; Tarozzi, A.; Rimondini, R.; Hrelia, P. Early effects of Abeta1-42 oligomers injection in mice: Involvement of PI3K/Akt/GSK3 and MAPK/ERK1/2 pathways. *Behav. Brain Res.* **2016**, *314*, 106–115. [CrossRef]
- 63. Yamamoto, N.; Tanida, M.; Kasahara, R.; Sobue, K.; Suzuki, K. Leptin inhibits amyloid beta-protein fibrillogenesis by decreasing GM1 gangliosides on the neuronal cell surface through PI3K/Akt/mTOR pathway. J. Neurochem. 2014, 131, 323–332. [CrossRef]

- Esmaeilzade, B.; Artimani, T.; Amiri, I.; Najafi, R.; Shahidi, S.; Sabec, M.; Farzadinia, P.; Zare, M.; Zahiri, M.; Soleimani Asl, S. Dimethyloxalylglycine preconditioning enhances protective effects of bone marrow-derived mesenchymal stem cells in Abetainduced Alzheimer disease. *Physiol. Behav.* 2019, 199, 265–272. [CrossRef] [PubMed]
- 65. Verret, L.; Jankowsky, J.L.; Xu, G.M.; Borchelt, D.R.; Rampon, C. Alzheimer's-type amyloidosis in transgenic mice impairs survival of newborn neurons derived from adult hippocampal neurogenesis. *J. Neurosci.* 2007, 27, 6771–6780. [CrossRef]
- Verret, L.; Trouche, S.; Zerwas, M.; Rampon, C. Hippocampal neurogenesis during normal and pathological aging. *Psychoneuroen*docrinology 2007, 32 (Suppl. 1), S26–S30. [CrossRef]
- 67. Gustavsson, T.; Syvanen, S.; O'Callaghan, P.; Sehlin, D. SPECT imaging of distribution and retention of a brain-penetrating bispecific amyloid-beta antibody in a mouse model of Alzheimer's disease. *Transl. Neurodegener.* **2020**, *9*, 37. [CrossRef]
- Honjo, Y.; Horibe, T.; Torisawa, A.; Ito, H.; Nakanishi, A.; Mori, H.; Komiya, T.; Takahashi, R.; Kawakami, K. Protein disulfide isomerase P5-immunopositive inclusions in patients with Alzheimer's disease. J. Alzheimer's Dis. 2014, 38, 601–609. [CrossRef]
- 69. Kuang, H.; Tan, C.Y.; Tian, H.Z.; Liu, L.H.; Yang, M.W.; Hong, F.F.; Yang, S.L. Exploring the bi-directional relationship between autophagy and Alzheimer's disease. *CNS Neurosci. Ther.* **2020**, *26*, 155–166. [CrossRef] [PubMed]
- Pierzynowska, K.; Podlacha, M.; Gaffke, L.; Majkutewicz, I.; Mantej, J.; Wegrzyn, A.; Osiadly, M.; Myslinska, D.; Wegrzyn, G. Autophagy-dependent mechanism of genistein-mediated elimination of behavioral and biochemical defects in the rat model of sporadic Alzheimer's disease. *Neuropharmacology* 2019, 148, 332–346. [CrossRef] [PubMed]
- 71. Wang, J.Z.; Wang, Z.H.; Tian, Q. Tau hyperphosphorylation induces apoptotic escape and triggers neurodegeneration in Alzheimer's disease. *Neurosci. Bull.* **2014**, *30*, 359–366. [CrossRef] [PubMed]
- Gourmaud, S.; Shou, H.; Irwin, D.J.; Sansalone, K.; Jacobs, L.M.; Lucas, T.H.; Marsh, E.D.; Davis, K.A.; Jensen, F.E.; Talos, D.M. Alzheimer-Like amyloid and tau alterations associated with cognitive deficit in temporal lobe epilepsy. *Brain* 2020, 143, 191–209. [CrossRef] [PubMed]
- 73. Zhu, X.; Raina, A.K.; Rottkamp, C.A.; Aliev, G.; Perry, G.; Boux, H.; Smith, M.A. Activation and redistribution of c-jun N-terminal kinase/stress activated protein kinase in degenerating neurons in Alzheimer's disease. J. Neurochem. 2001, 76, 435–441. [CrossRef]
- 74. Orejana, L.; Barros-Minones, L.; Aguirre, N.; Puerta, E. Implication of JNK pathway on tau pathology and cognitive decline in a senescence-accelerated mouse model. *Exp. Gerontol.* **2013**, *48*, 565–571. [CrossRef]
- 75. Wu, C.; Gong, W.G.; Wang, Y.J.; Sun, J.J.; Zhou, H.; Zhang, Z.J.; Ren, Q.G. Escitalopram alleviates stress-induced Alzheimer's disease-like tau pathologies and cognitive deficits by reducing hypothalamic-pituitary-adrenal axis reactivity and insulin/GSK-3beta signal pathway activity. *Neurobiol. Aging* **2018**, *67*, 137–147. [CrossRef]
- 76. Ye, T.; Li, X.; Zhou, P.; Ye, S.; Gao, H.; Hua, R.; Ma, J.; Wang, Y.; Cai, B. Chrysophanol improves memory ability of d-galactose and Abeta25-35 treated rat correlating with inhibiting tau hyperphosphorylation and the CaM-CaMKIV signal pathway in hippocampus. 3 Biotech 2020, 10, 111. [CrossRef]
- 77. Lee, I.S.; Jung, K.; Kim, I.S.; Lee, H.; Kim, M.; Yun, S.; Hwang, K.; Shin, J.E.; Park, K.I. Human neural stem cells alleviate Alzheimer-like pathology in a mouse model. *Mol. Neurodegener.* **2015**, *10*, 38. [CrossRef]
- Lee, J.K.; Jin, H.K.; Endo, S.; Schuchman, E.H.; Carter, J.E.; Bae, J.S. Intracerebral transplantation of bone marrow-derived mesenchymal stem cells reduces amyloid-beta deposition and rescues memory deficits in Alzheimer's disease mice by modulation of immune responses. *Stem Cells* 2010, *28*, 329–343. [CrossRef]
- Uddin, M.S.; Stachowiak, A.; Mamun, A.A.; Tzvetkov, N.T.; Takeda, S.; Atanasov, A.G.; Bergantin, L.B.; Abdel-Daim, M.M.; Stankiewicz, A.M. Autophagy and Alzheimer's Disease: From Molecular Mechanisms to Therapeutic Implications. *Front. Aging Neurosci.* 2018, 10, 4. [CrossRef]
- Menzies, F.M.; Fleming, A.; Caricasole, A.; Bento, C.F.; Andrews, S.P.; Ashkenazi, A.; Fullgrabe, J.; Jackson, A.; Jimenez Sanchez, M.; Karabiyik, C.; et al. Autophagy and Neurodegeneration: Pathogenic Mechanisms and Therapeutic Opportunities. *Neuron* 2017, 93, 1015–1034. [CrossRef]
- 81. Demirci, S.; Aynali, A.; Demirci, K.; Demirci, S.; Aridogan, B.C. The Serum Levels of Resistin and Its Relationship with Other Proinflammatory Cytokines in Patients with Alzheimer's Disease. *Clin. Psychopharmacol. Neurosci.* **2017**, *15*, 59–63. [CrossRef]
- 82. Ng, A.; Tam, W.W.; Zhang, M.W.; Ho, C.S.; Husain, S.F.; McIntyre, R.S.; Ho, R.C. IL-1beta, IL-6, TNF- alpha and CRP in Elderly Patients with Depression or Alzheimer's disease: Systematic Review and Meta-Analysis. *Sci. Rep.* **2018**, *8*, 12050. [CrossRef]
- Mosser, D.M.; Zhang, X. Interleukin-10: New perspectives on an old cytokine. *Immunol. Rev.* 2008, 226, 205–218. [CrossRef] [PubMed]
- 84. Guillot-Sestier, M.V.; Doty, K.R.; Gate, D.; Rodriguez, J., Jr.; Leung, B.P.; Rezai-Zadeh, K.; Town, T. Il10 deficiency rebalances innate immunity to mitigate Alzheimer-like pathology. *Neuron* 2015, *85*, 534–548. [CrossRef]
- 85. Lively, S.; Schlichter, L.C. Microglia Responses to Pro-inflammatory Stimuli (LPS, IFNgamma + TNFalpha) and Reprogramming by Resolving Cytokines (IL-4, IL-10). *Front. Cell Neurosci.* **2018**, *12*, 215. [CrossRef]
- 86. Jha, N.K.; Jha, S.K.; Kar, R.; Nand, P.; Swati, K.; Goswami, V.K. Nuclear factor-kappa beta as a therapeutic target for Alzheimer's disease. *J. Neurochem.* **2019**, 150, 113–137. [CrossRef] [PubMed]
- 87. Shi, Z.; Hong, Y.; Zhang, K.; Wang, J.; Zheng, L.; Zhang, Z.; Hu, Z.; Han, X.; Han, Y.; Chen, T.; et al. BAG-1M co-activates BACE1 transcription through NF-kappaB and accelerates Abeta production and memory deficit in Alzheimer's disease mouse model. *Biochim. Biophys. Acta Mol. Basis Dis.* **2017**, *1863*, 2398–2407. [CrossRef]

- Nilson, A.N.; English, K.C.; Gerson, J.E.; Barton Whittle, T.; Nicolas Crain, C.; Xue, J.; Sengupta, U.; Castillo-Carranza, D.L.; Zhang, W.; Gupta, P.; et al. Tau Oligomers Associate with Inflammation in the Brain and Retina of Tauopathy Mice and in Neurodegenerative Diseases. J. Alzheimer's Dis. 2017, 55, 1083–1099. [CrossRef]
- Nogueras-Ortiz, C.J.; Mahairaki, V.; Delgado-Peraza, F.; Das, D.; Avgerinos, K.; Eren, E.; Hentschel, M.; Goetzl, E.J.; Mattson, M.P.; Kapogiannis, D. Astrocyte- and Neuron-Derived Extracellular Vesicles from Alzheimer's Disease Patients Effect Complement-Mediated Neurotoxicity. *Cells* 2020, *9*, 1618. [CrossRef]
- Safar, M.M.; Arab, H.H.; Rizk, S.M.; El-Maraghy, S.A. Bone Marrow-Derived Endothelial Progenitor Cells Protect Against Scopolamine-Induced Alzheimer-Like Pathological Aberrations. *Mol. Neurobiol.* 2016, 53, 1403–1418. [CrossRef]
- 91. Piers, T.M.; Cosker, K.; Mallach, A.; Johnson, G.T.; Guerreiro, R.; Hardy, J.; Pocock, J.M. A locked immunometabolic switch underlies TREM2 R47H loss of function in human iPSC-derived microglia. *FASEB J.* **2020**, *34*, 2436–2450. [CrossRef]
- Qiao, P.; Ma, J.; Wang, Y.; Huang, Z.; Zou, Q.; Cai, Z.; Tang, Y. Curcumin Prevents Neuroinflammation by Inducing Microglia to Transform into the M2-phenotype via CaMKKbeta-dependent Activation of the AMP-Activated Protein Kinase Signal Pathway. *Curr. Alzheimer Res.* 2020, *17*, 735–752. [CrossRef]
- 93. Satoh, J.; Kino, Y.; Asahina, N.; Takitani, M.; Miyoshi, J.; Ishida, T.; Saito, Y. TMEM119 marks a subset of microglia in the human brain. *Neuropathology* **2016**, *36*, 39–49. [CrossRef]
- Zeng, H.; Liu, N.; Yang, Y.Y.; Xing, H.Y.; Liu, X.X.; Li, F.; La, G.Y.; Huang, M.J.; Zhou, M.W. Lentivirus-mediated downregulation of alpha-synuclein reduces neuroinflammation and promotes functional recovery in rats with spinal cord injury. *J. Neuroinflamm.* 2019, 16, 283. [CrossRef]
- Ji, J.; Xue, T.F.; Guo, X.D.; Yang, J.; Guo, R.B.; Wang, J.; Huang, J.Y.; Zhao, X.J.; Sun, X.L. Antagonizing peroxisome proliferatoractivated receptor gamma facilitates M1-to-M2 shift of microglia by enhancing autophagy via the LKB1-AMPK signaling pathway. *Aging Cell* 2018, 17, e12774. [CrossRef]
- 96. Zhang, Y.; He, M.L. Deferoxamine enhances alternative activation of microglia and inhibits amyloid beta deposits in APP/PS1 mice. *Brain Res.* 2017, 1677, 86–92. [CrossRef]
- 97. Wozniak, M.A.; Mee, A.P.; Itzhaki, R.F. Herpes simplex virus type 1 DNA is located within Alzheimer's disease amyloid plaques. *J. Pathol.* **2009**, *217*, 131–138. [CrossRef]
- Miklossy, J.; Khalili, K.; Gern, L.; Ericson, R.L.; Darekar, P.; Bolle, L.; Hurlimann, J.; Paster, B.J. Borrelia burgdorferi persists in the brain in chronic lyme neuroborreliosis and may be associated with Alzheimer disease. *J. Alzheimer's Dis.* 2004, *6*, 639–649; discussion 673–681. [CrossRef]
- 99. Pisa, D.; Alonso, R.; Rabano, A.; Rodal, I.; Carrasco, L. Different Brain Regions are Infected with Fungi in Alzheimer's Disease. *Sci. Rep.* 2015, *5*, 15015. [CrossRef]
- 100. Linard, M.; Letenneur, L.; Garrigue, I.; Doize, A.; Dartigues, J.F.; Helmer, C. Interaction between APOE4 and herpes simplex virus type 1 in Alzheimer's disease. *Alzheimer's Dement.* 2020, *16*, 200–208. [CrossRef]
- 101. Carter, C.J. Genetic, Transcriptome, Proteomic, and Epidemiological Evidence for Blood-Brain Barrier Disruption and Polymicrobial Brain Invasion as Determinant Factors in Alzheimer's Disease. J. Alzheimer's Rep. 2017, 1, 125–157. [CrossRef]
- 102. Wu, J.T.; Leung, K.; Bushman, M.; Kishore, N.; Niehus, R.; de Salazar, P.M.; Cowling, B.J.; Lipsitch, M.; Leung, G.M. Estimating clinical severity of COVID-19 from the transmission dynamics in Wuhan, China. *Nat. Med.* **2020**, *26*, 506–510. [CrossRef]
- 103. Caratozzolo, S.; Zucchelli, A.; Turla, M.; Cotelli, M.S.; Fascendini, S.; Zanni, M.; Bianchetti, A.; Psy, M.P.; Rozzini, R.; Boffelli, S.; et al. The impact of COVID-19 on health status of home-dwelling elderly patients with dementia in East Lombardy, Italy: Results from COVIDEM network. *Aging Clin. Exp. Res.* 2020, *32*, 2133–2140. [CrossRef] [PubMed]
- 104. Bergman, M.; Salman, H.; Beloosesky, Y.; Djaldetti, M.; Bessler, H. Are peripheral blood cells from patients with Alzheimer disease more sensitive to apoptotic stimuli? *Alzheimer Assoc. Disord.* 2002, 16, 156–160. [CrossRef] [PubMed]
- 105. Chan, G.; White, C.C.; Winn, P.A.; Cimpean, M.; Replogle, J.M.; Glick, L.R.; Cuerdon, N.E.; Ryan, K.J.; Johnson, K.A.; Schneider, J.A.; et al. CD33 modulates TREM2: Convergence of Alzheimer loci. *Nat. Neurosci.* 2015, *18*, 1556–1558. [CrossRef] [PubMed]
- 106. Monsonego, A.; Zota, V.; Karni, A.; Krieger, J.I.; Bar-Or, A.; Bitan, G.; Budson, A.E.; Sperling, R.; Selkoe, D.J.; Weiner, H.L. Increased T cell reactivity to amyloid beta protein in older humans and patients with Alzheimer disease. *J. Clin. Investig.* 2003, 112, 415–422. [CrossRef]
- 107. McQuade, A.; Kang, Y.J.; Hasselmann, J.; Jairaman, A.; Sotelo, A.; Coburn, M.; Shabestari, S.K.; Chadarevian, J.P.; Fote, G.; Tu, C.H.; et al. Gene expression and functional deficits underlie TREM2-knockout microglia responses in human models of Alzheimer's disease. *Nat. Commun.* 2020, *11*, 5370. [CrossRef]
- 108. Chen, H.; Lin, W.; Zhang, Y.; Lin, L.; Chen, J.; Zeng, Y.; Zheng, M.; Zhuang, Z.; Du, H.; Chen, R.; et al. IL-10 Promotes Neurite Outgrowth and Synapse Formation in Cultured Cortical Neurons after the Oxygen-Glucose Deprivation via JAK1/STAT3 Pathway. Sci. Rep. 2016, 6, 30459. [CrossRef]
- Naaldijk, Y.; Jager, C.; Fabian, C.; Leovsky, C.; Bluher, A.; Rudolph, L.; Hinze, A.; Stolzing, A. Effect of systemic transplantation of bone marrow-derived mesenchymal stem cells on neuropathology markers in APP/PS1 Alzheimer mice. *Neuropathol. Appl. Neurobiol.* 2017, 43, 299–314. [CrossRef]
- 110. Marsh, S.E.; Abud, E.M.; Lakatos, A.; Karimzadeh, A.; Yeung, S.T.; Davtyan, H.; Fote, G.M.; Lau, L.; Weinger, J.G.; Lane, T.E.; et al. The adaptive immune system restrains Alzheimer's disease pathogenesis by modulating microglial function. *Proc. Natl. Acad. Sci.* USA 2016, 113, E1316–E1325. [CrossRef]

- 111. Liu, W.; Rong, Y.; Wang, J.; Zhou, Z.; Ge, X.; Ji, C.; Jiang, D.; Gong, F.; Li, L.; Chen, J.; et al. Exosome-shuttled miR-216a-5p from hypoxic preconditioned mesenchymal stem cells repair traumatic spinal cord injury by shifting microglial M1/M2 polarization. *J. Neuroinflamm.* 2020, *17*, 47. [CrossRef]
- 112. Baufeld, C.; O'Loughlin, E.; Calcagno, N.; Madore, C.; Butovsky, O. Differential contribution of microglia and monocytes in neurodegenerative diseases. *J. Neural Transm. Vienna* **2018**, 125, 809–826. [CrossRef]
- 113. Zhong, Z.; Chen, A.; Fa, Z.; Ding, Z.; Xie, J.; Sun, Y.; Zhang, R.; Wang, Q. Adipose-Derived Stem Cells Modulate BV2 Microglial M1/M2 Polarization by Producing GDNF. Stem Cells Dev. 2020, 29, 714–727. [CrossRef]
- 114. Petukhova, E.O.; Mukhamedshina, Y.O.; Salafutdinov, I.; Garanina, E.E.; Kaligin, M.S.; Leushina, A.V.; Rizvanov, A.A.; Reis, H.J.; Palotas, A.; Zefirov, A.L.; et al. Effects of Transplanted Umbilical Cord Blood Mononuclear Cells Overexpressing GDNF on Spatial Memory and Hippocampal Synaptic Proteins in a Mouse Model of Alzheimer's Disease. J. Alzheimer's Dis. 2019, 69, 443–453. [CrossRef]
- 115. Ren, C.; Gu, X.; Li, H.; Lei, S.; Wang, Z.; Wang, J.; Yin, P.; Zhang, C.; Wang, F.; Liu, C. The role of DKK1 in Alzheimer's disease: A potential intervention point of brain damage prevention? *Pharmacol. Res.* **2019**, *144*, 331–335. [CrossRef] [PubMed]
- Yu, B.; Zhang, J.; Li, H.; Sun, X. Silencing of aquaporin1 activates the Wnt signaling pathway to improve cognitive function in a mouse model of Alzheimer's disease. *Gene* 2020, 755, 144904. [CrossRef]
- 117. Dobrowolski, R.; Vick, P.; Ploper, D.; Gumper, I.; Snitkin, H.; Sabatini, D.D.; De Robertis, E.M. Presenilin deficiency or lysosomal inhibition enhances Wnt signaling through relocalization of GSK3 to the late-endosomal compartment. *Cell Rep.* 2012, 2, 1316–1328. [CrossRef] [PubMed]
- 118. Arnes, M.; Casas Tinto, S. Aberrant Wnt signaling: A special focus in CNS diseases. J. Neurogenet. 2017, 31, 216–222. [CrossRef] [PubMed]
- 119. Tapia-Rojas, C.; Inestrosa, N.C. Wnt signaling loss accelerates the appearance of neuropathological hallmarks of Alzheimer's disease in J20-APP transgenic and wild-type mice. *J. Neurochem.* **2018**, *144*, 443–465. [CrossRef] [PubMed]
- Wang, Y.; Liang, G.; Liang, S.; Mund, R.; Shi, Y.; Wei, H. Dantrolene Ameliorates Impaired Neurogenesis and Synaptogenesis in Induced Pluripotent Stem Cell Lines Derived from Patients with Alzheimer's Disease. *Anesthesiology* 2020, 132, 1062–1079. [CrossRef]
- 121. Hayashi, Y.; Lin, H.T.; Lee, C.C.; Tsai, K.J. Effects of neural stem cell transplantation in Alzheimer's disease models. *J. Biomed. Sci.* **2020**, *27*, 29. [CrossRef]
- 122. Corbett, N.J.; Gabbott, P.L.; Klementiev, B.; Davies, H.A.; Colyer, F.M.; Novikova, T.; Stewart, M.G. Amyloid-beta induced CA1 pyramidal cell loss in young adult rats is alleviated by systemic treatment with FGL, a neural cell adhesion molecule-derived mimetic peptide. *PLoS ONE* 2013, *8*, e71479. [CrossRef] [PubMed]
- 123. Klein, R.; Blaschke, S.; Neumaier, B.; Endepols, H.; Graf, R.; Keuters, M.; Hucklenbroich, J.; Albrechtsen, M.; Rees, S.; Fink, G.R.; et al. The synthetic NCAM mimetic peptide FGL mobilizes neural stem cells in vitro and in vivo. *Stem Cell Rev. Rep.* 2014, 10, 539–547. [CrossRef]
- 124. Jiang, Y.; Gao, H.; Yuan, H.; Xu, H.; Tian, M.; Du, G.; Xie, W. Amelioration of postoperative cognitive dysfunction in mice by mesenchymal stem cell-conditioned medium treatments is associated with reduced inflammation, oxidative stress and increased BDNF expression in brain tissues. *Neurosci. Lett.* 2019, 709, 134372. [CrossRef] [PubMed]
- 125. Rosenblum, S.; Smith, T.N.; Wang, N.; Chua, J.Y.; Westbroek, E.; Wang, K.; Guzman, R. BDNF Pretreatment of Human Embryonic-Derived Neural Stem Cells Improves Cell Survival and Functional Recovery After Transplantation in Hypoxic-Ischemic Stroke. *Cell Transplant.* 2015, 24, 2449–2461. [CrossRef]
- 126. Chen, Y.; Pan, C.; Xuan, A.; Xu, L.; Bao, G.; Liu, F.; Fang, J.; Long, D. Treatment Efficacy of NGF Nanoparticles Combining Neural Stem Cell Transplantation on Alzheimer's Disease Model Rats. *Med. Sci. Monit.* **2015**, *21*, 3608–3615. [CrossRef]
- 127. Danielyan, L.; Beer-Hammer, S.; Stolzing, A.; Schafer, R.; Siegel, G.; Fabian, C.; Kahle, P.; Biedermann, T.; Lourhmati, A.; Buadze, M.; et al. Intranasal delivery of bone marrow-derived mesenchymal stem cells, macrophages, and microglia to the brain in mouse models of Alzheimer's and Parkinson's disease. *Cell Transplant.* 2014, 23 (Suppl. 1), S123–S139. [CrossRef] [PubMed]
- 128. Lee, M.; Ban, J.J.; Yang, S.; Im, W.; Kim, M. The exosome of adipose-derived stem cells reduces beta-amyloid pathology and apoptosis of neuronal cells derived from the transgenic mouse model of Alzheimer's disease. *Brain Res.* 2018, 1691, 87–93. [CrossRef]
- 129. Wang, X.; Yang, G. Bone marrow mesenchymal stem cells-derived exosomes reduce Abeta deposition and improve cognitive function recovery in mice with Alzheimer's disease by activating sphingosine kinase/sphingosine-1-phosphate signaling pathway. *Cell Biol. Int.* **2020**, *45*, 775–784. [CrossRef]
- 130. Losurdo, M.; Pedrazzoli, M.; D'Agostino, C.; Elia, C.A.; Massenzio, F.; Lonati, E.; Mauri, M.; Rizzi, L.; Molteni, L.; Bresciani, E.; et al. Intranasal delivery of mesenchymal stem cell-derived extracellular vesicles exerts immunomodulatory and neuroprotective effects in a 3xTg model of Alzheimer's disease. *Stem Cells Transl. Med.* **2020**, *9*, 1068–1084. [CrossRef]