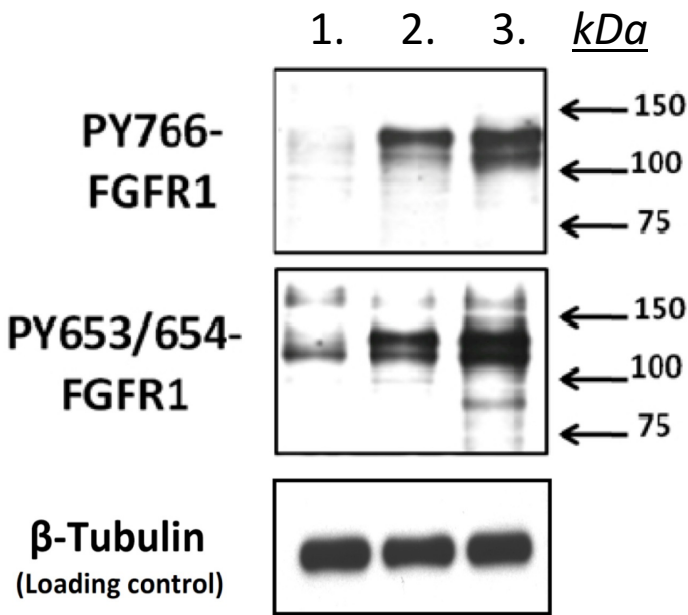


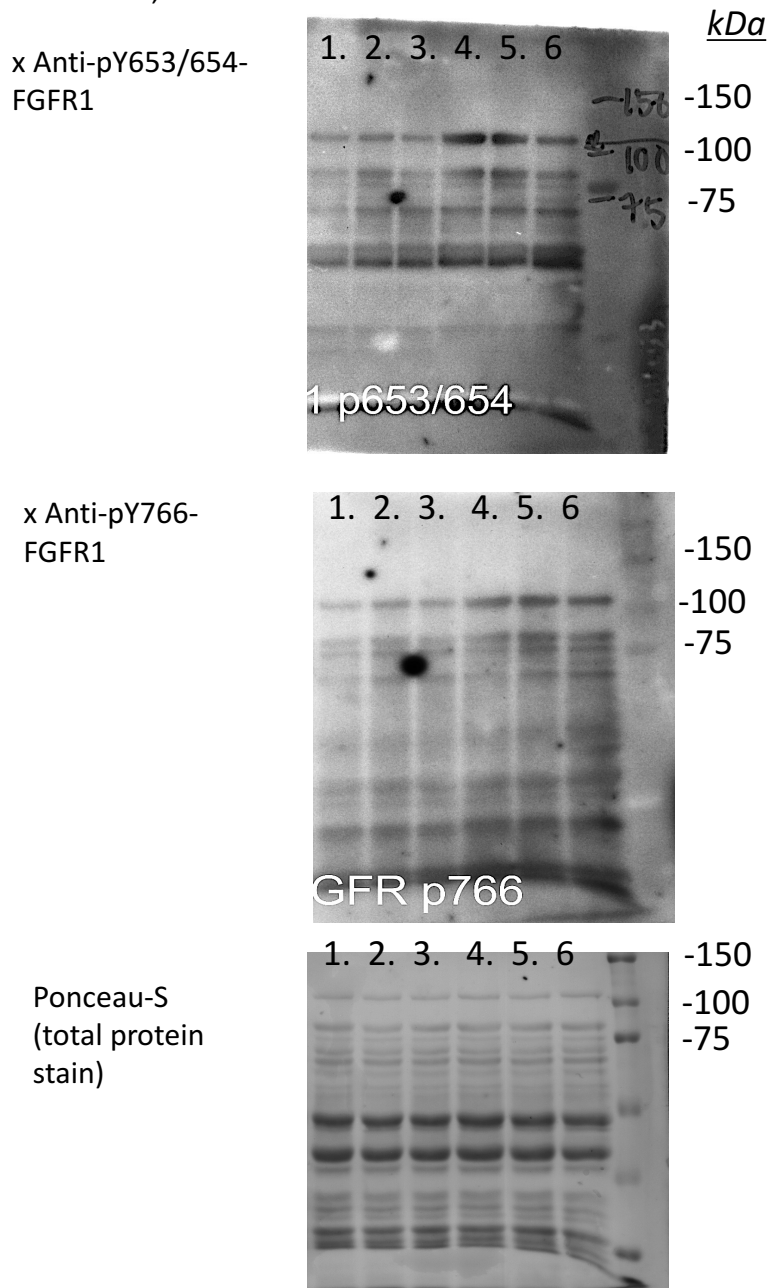
**Figure S1 Testing of anti-pY- FGFR1 antibodies**

Human endothelial kidney (HEK) 293 cells were transfected or not with a plasmid expressing mouse FGFR1 and stimulated or not with Lo-FGF2 (10 ng/ml, 15 minutes). Subsequently, cell lysates were analyzed by western blotting and probed with antibodies for pY766-FGFR1, pY653/654-FGFR1, or beta ( $\beta$ )-tubulin, as indicated. Untransfected cells presented negligible signal for pY766-FGFR1 , and detectable signal for pY653/654-FGFR1. In both cases, the immunoreactive signal increased substantially in transfected cells, especially after stimulation with Lo-FGF2. Intensity of signal for  $\beta$ -tubulin (loading control) appeared similar between the different samples.



1. Untransfected HEK 293 cells
2. Cells transfected with cDNA for FGFR1
3. Cells transfected with a cDNA for FGFR1 + FGF2

**Figure S2. Lo-FGF2 promotes mito-FGFR1 phosphorylation in hearts.** Western blot staining of cardiac mitochondria (IFM) from saline-perfused (lanes 1,2,3, three preparations) and hearts stimulated with Lo-FGF2 for 15 minutes, lanes 4,5,6, three preparations) for pY-FGFR1. Ponceau S staining indicates similar protein loading between all lanes. A stronger signal for immunoreactive bands (70-110 kDa) can be observed in lanes 4,5 and 6, compared to lanes 1,2 and 3.



Methodology for whole heart perfusion with a Lo-FGF2 containing buffer for 15 minutes, is described in references #17, and #32 (PhD thesis of the first author). Mitochondrial samples (20 microg/lane) were analyzed in 7.5% polyacrylamide gels.

**Table S1.** The effect of calcium overload, Lo-FGF2, and PKC $\epsilon$  inhibitors on cardiac (SSM) mitochondrial Optical Density (OD) at 545 nM

	Control	Lo-FGF2	Lo-FGF2 + eV1-2	Lo-FGF2 + scrambled peptide
<b>Calcium Retention Capacity</b> (nM Ca <sup>2+</sup> /mg protein)	500 $\pm$ 72	936 $\pm$ 5.8*	375 $\pm$ 56	900 $\pm$ 13.5*
<b>Rate of swelling</b> (OD <sub>545</sub> $\times$ 10 <sup>3</sup> /min)	73.1 $\pm$ 5.6	75.5 $\pm$ 9.9	61.8 $\pm$ 5.4	75.2 $\pm$ 6.6
<b>Amplitude of swelling</b> (% of maximal swelling)	30.0 $\pm$ 0.3	29.3 $\pm$ 2.5	29.6 $\pm$ 2.3	32.9 $\pm$ 3.3

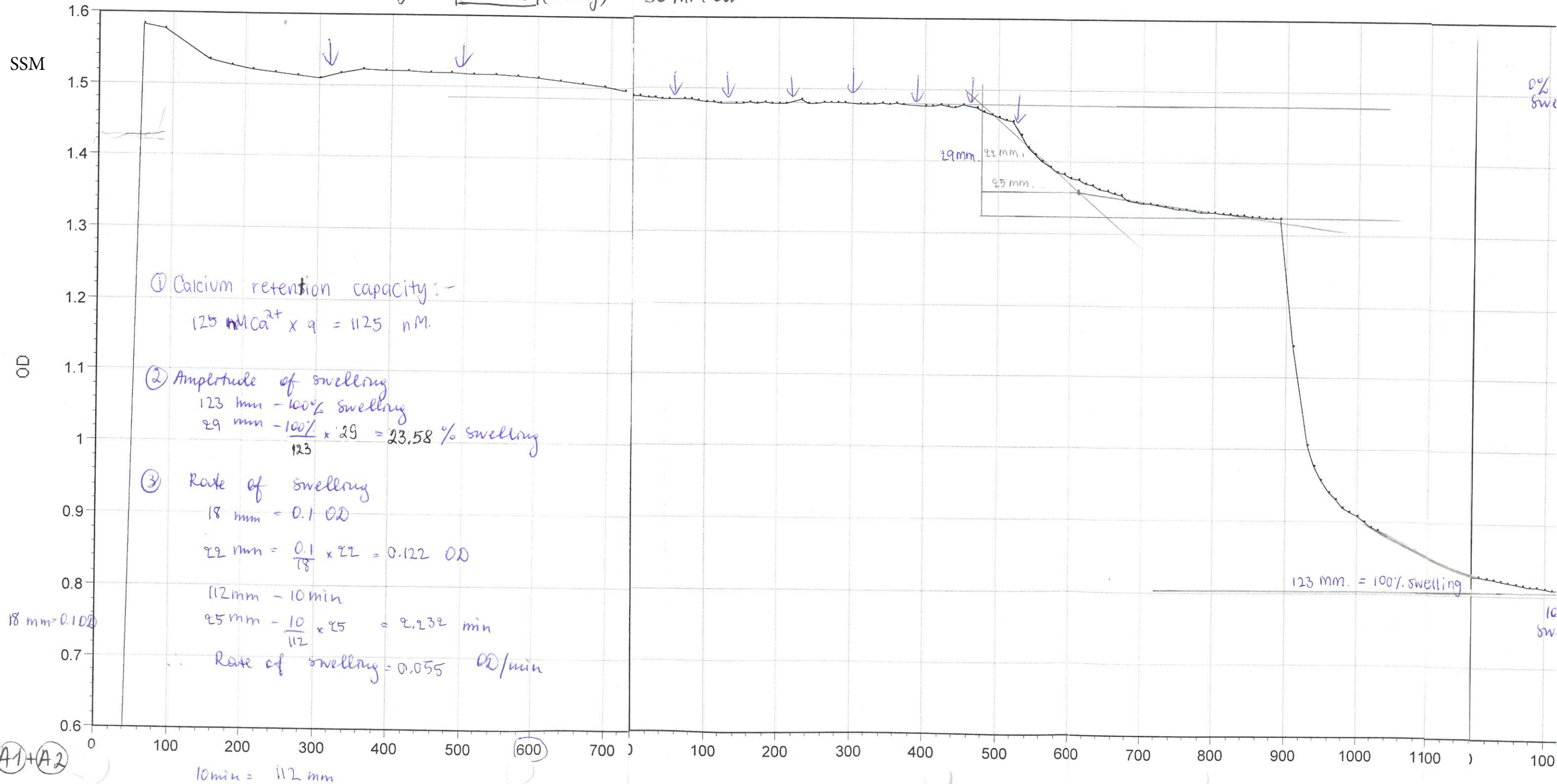
**Table S2.** The effect of Lo-FGF2 on cardiac mitochondria (SSM)  
Respiration

	Rate of Respiration		RCI (State 3/ State 4)	ADP/O ratio nmol ADP/ nmol O <sub>2</sub>	Rate of Oxidative Phosphorylation (nmol ATP/min/ mg protein)
	State 3	State 4			
<b>Control</b>	154.3 ± 6.7	19.9 ± 1.2	7.79 ± 0.26	2.51 ± 0.05	387.3 ± 19.5
<b>Lo-FGF2</b>	171.3 ± 10.3	20.0 ± 2.4	8.85 ± 0.87	2.56 ± 0.04	437.9 ± 25.8



December 7, 2007

ADP+oligo + FGF-2 (50ng) + 50 mM  $\text{Ca}^{2+}$



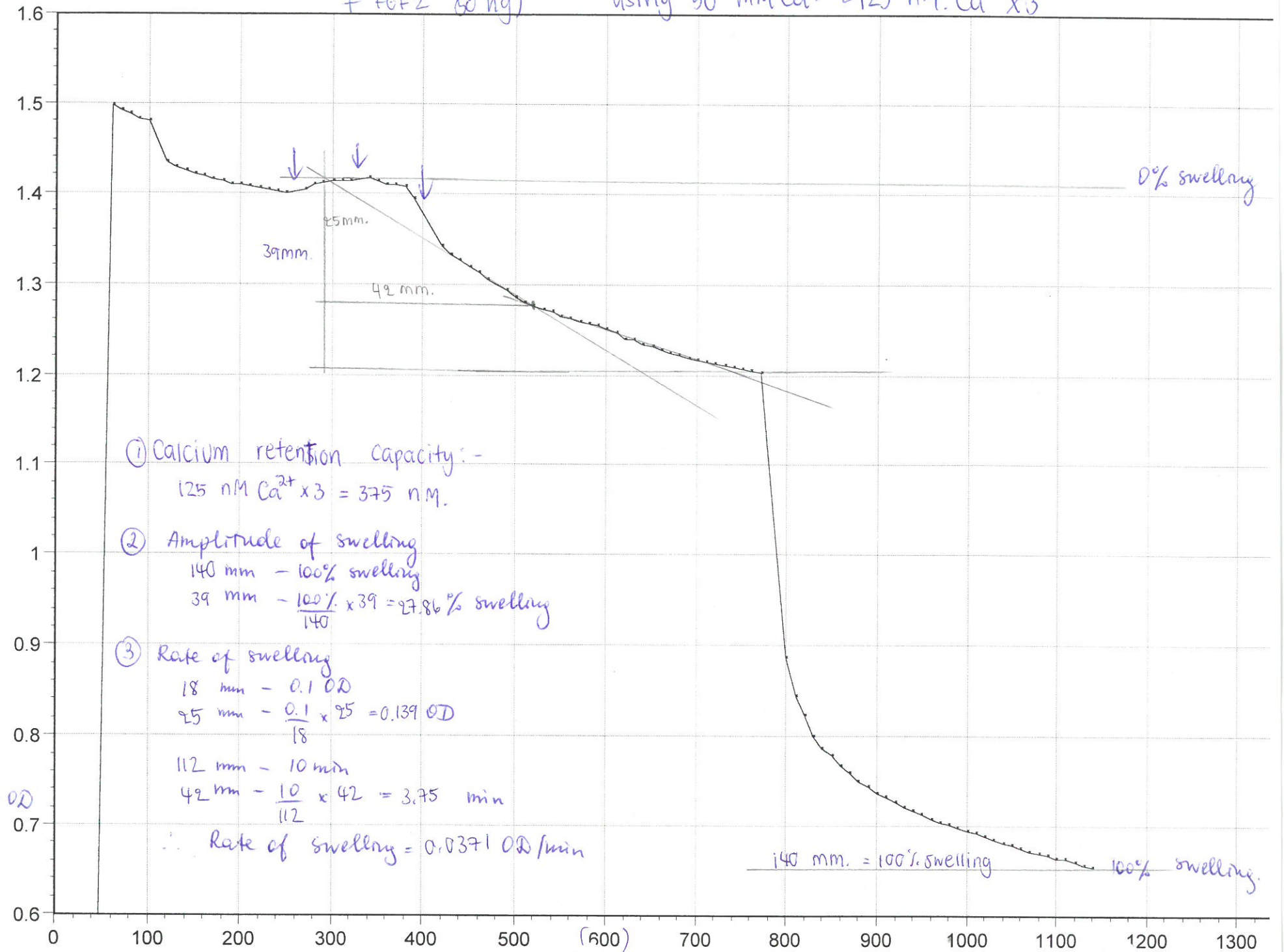
(A3)

ADP + Oligo + PKC $\epsilon$  inh (0.5  $\mu$ M)  
+ FGF2 (50 ng)using 50 mM Ca $^{2+}$   $\rightarrow$  125 nM Ca $^{2+}$  x3

Dec 7, 2007

SSM

OD



① Calcium retention capacity:-

$$125 \text{ nM Ca}^{2+} \times 3 = 375 \text{ nM.}$$

② Amplitude of swelling

$$140 \text{ mm} - 100\% \text{ swelling}$$

$$39 \text{ mm} - \frac{100\%}{140} \times 39 = 27.86\% \text{ swelling}$$

③ Rate of swelling

$$18 \text{ mm} - 0.1 \text{ OD}$$

$$25 \text{ mm} - \frac{0.1}{18} \times 25 = 0.139 \text{ OD}$$

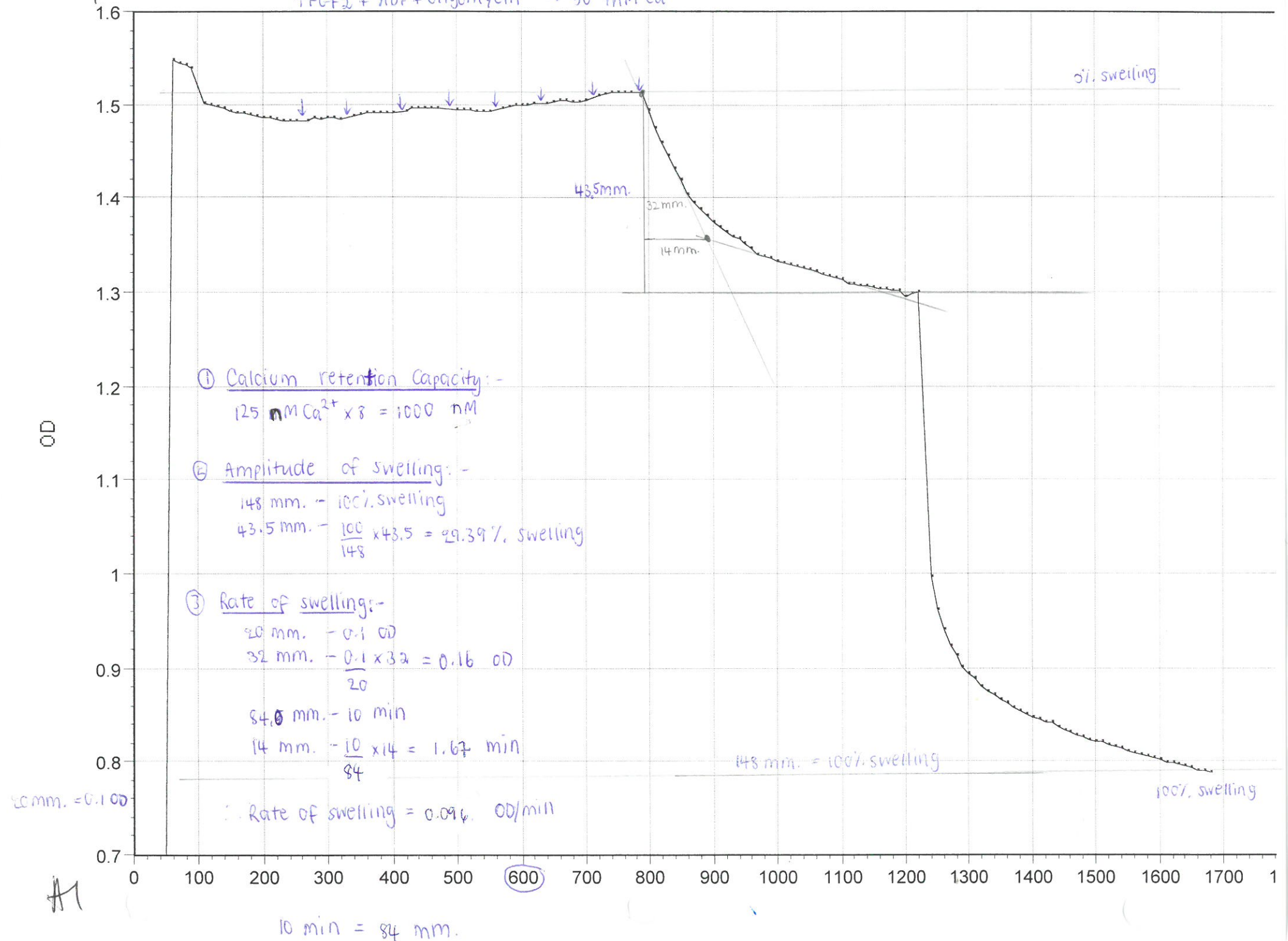
$$112 \text{ mm} - 10 \text{ min}$$

$$42 \text{ mm} - \frac{10}{112} \times 42 = 3.75 \text{ min}$$

$$\therefore \text{Rate of swelling} = 0.0371 \text{ OD/min}$$

Deck 2, 2007

$+ \text{FCF}_2 + \text{ADP} + \text{Oligomycin} \rightarrow 50 \text{ mM Ca}^{2+}$

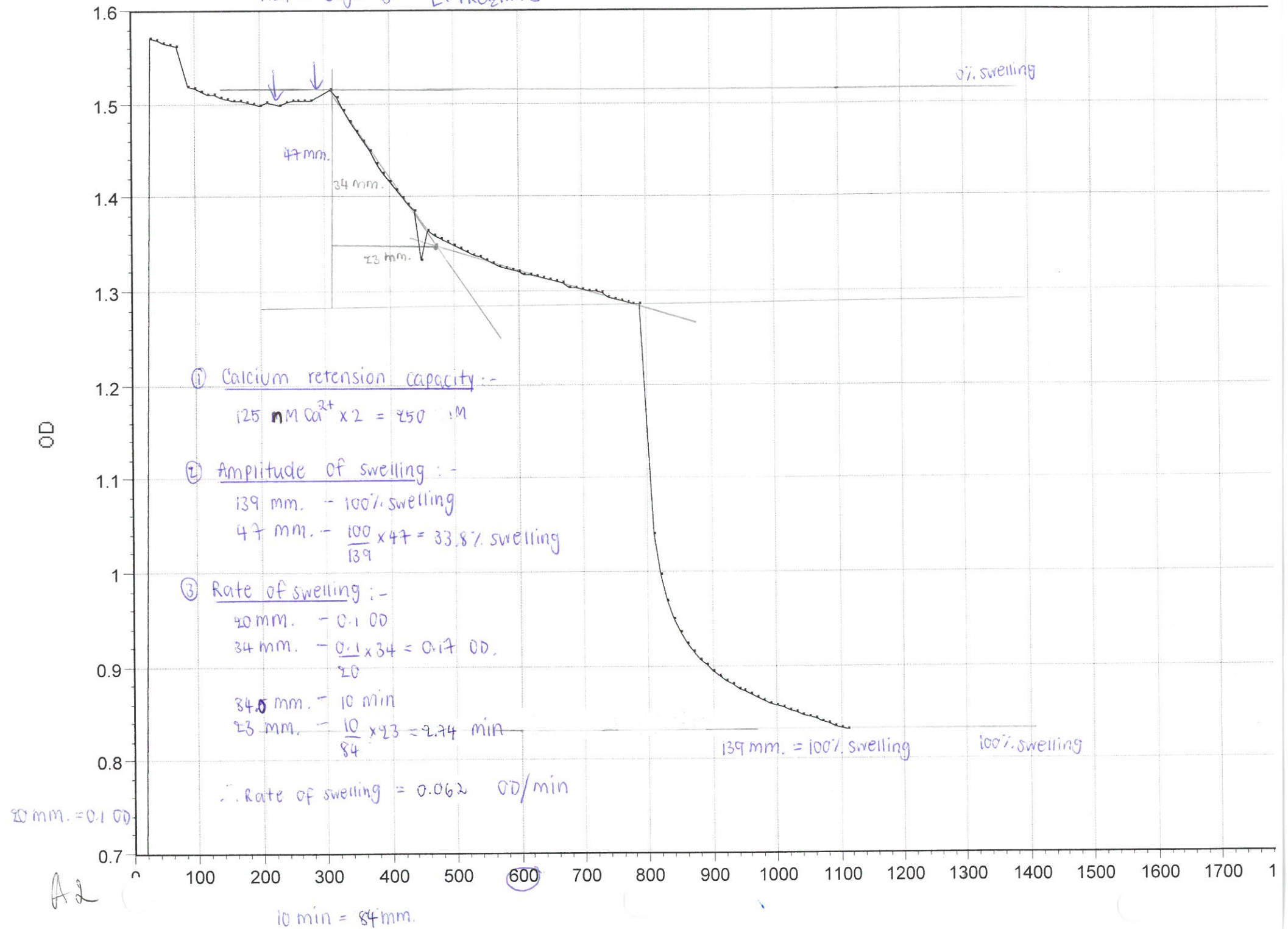




Dec 12, 2007

ADP + Oligomycin  $\left[ \begin{matrix} + \text{FGF2} \\ + \text{PKC}_{\text{inh.}} \end{matrix} \right] \rightarrow 50 \text{ mM Ca}^{2+}$

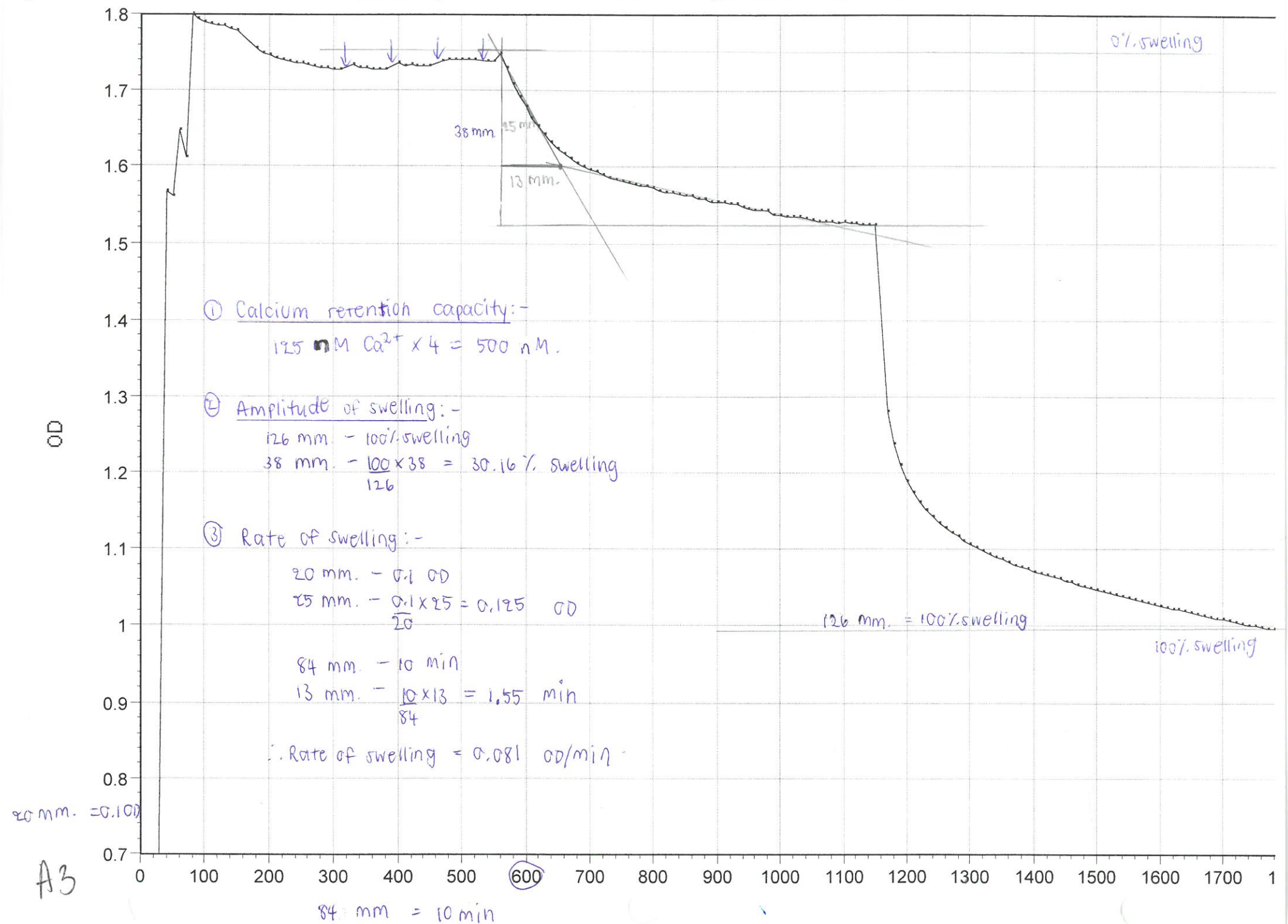
SSM



Dec 12, 2007

ADP + C<sub>12</sub> mycin. → 50 mM Ca<sup>2+</sup> NO FGF2 + NO PKC<sub>ε</sub> inh.

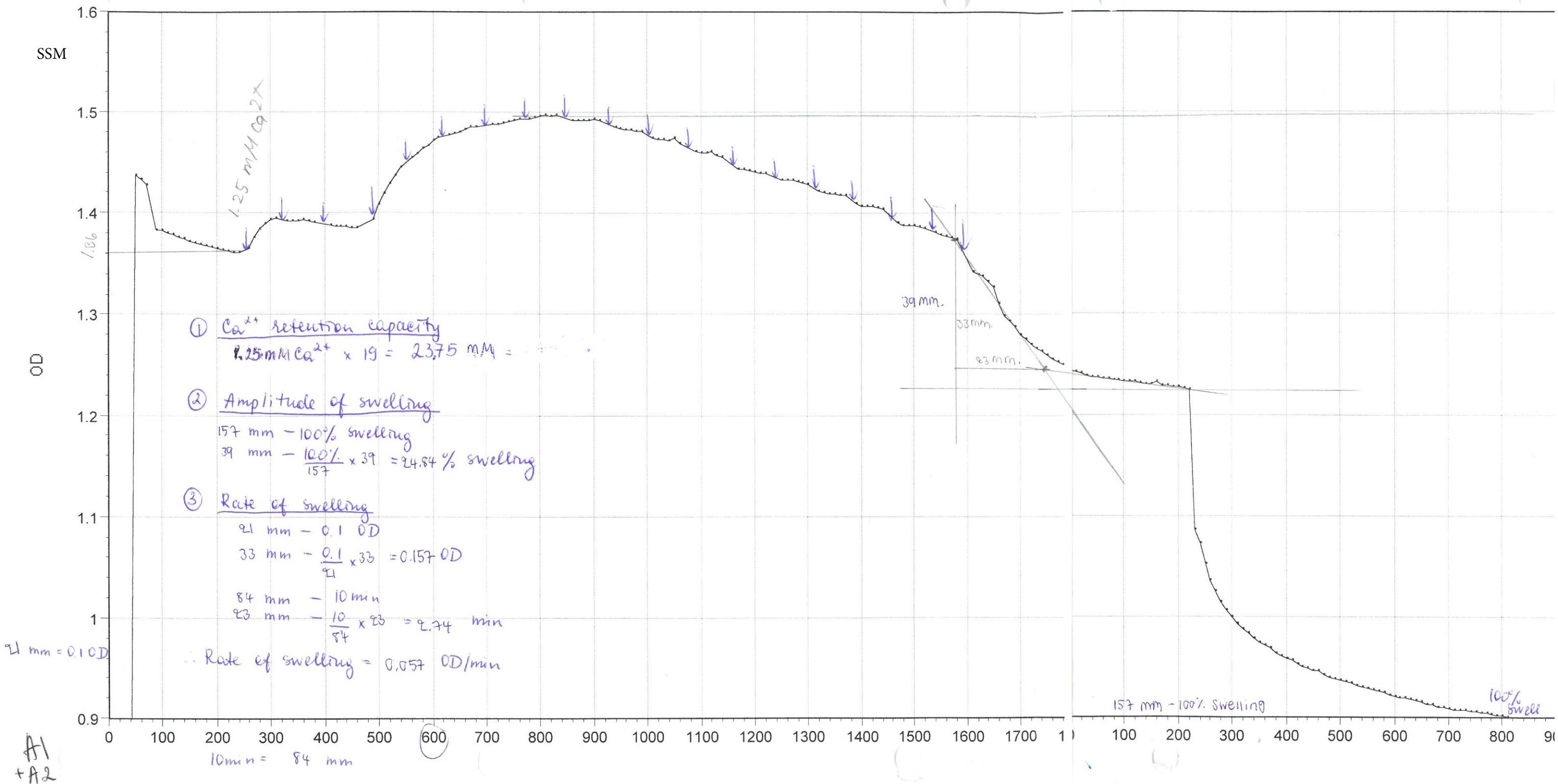
SSM



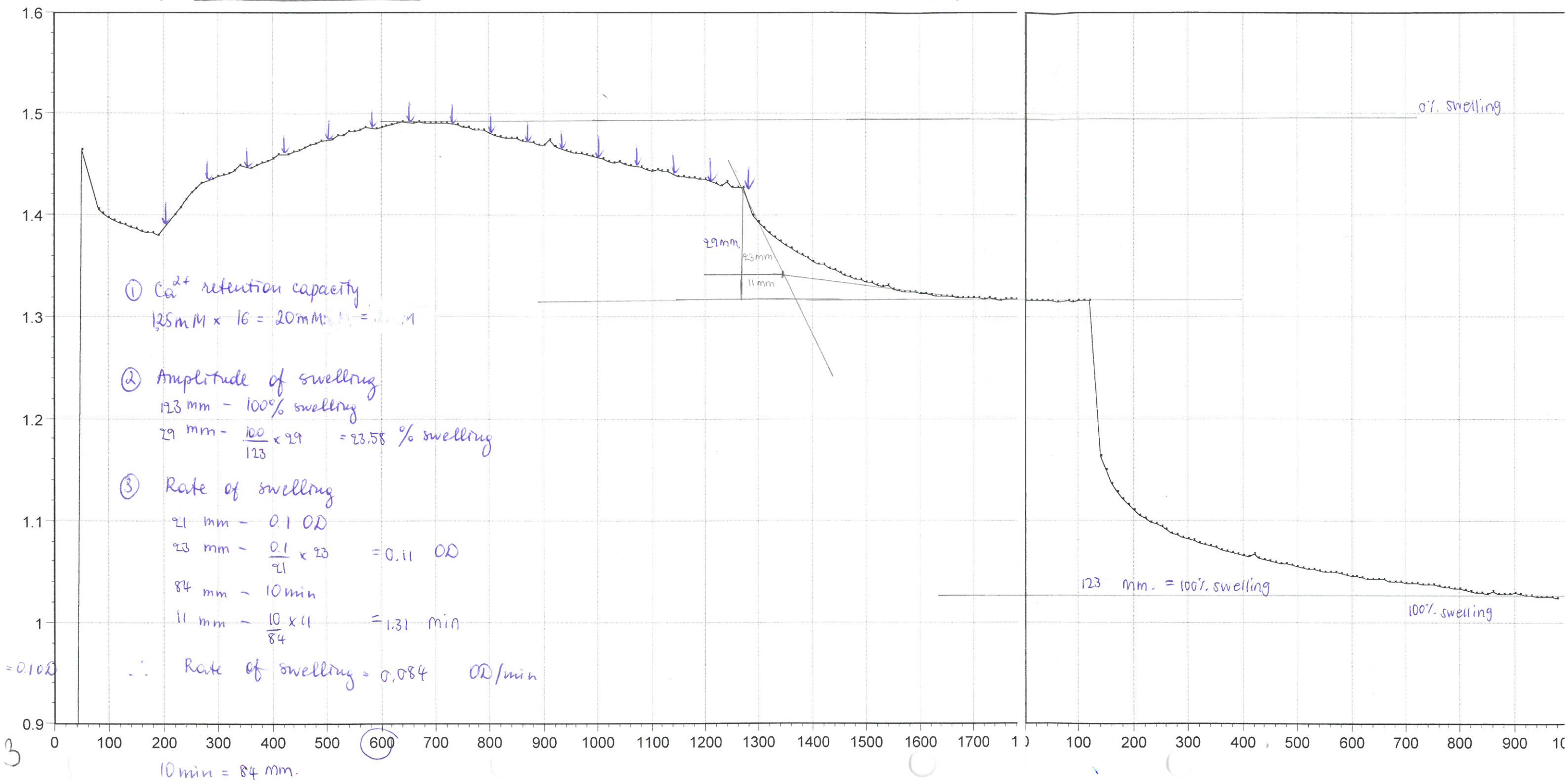
Dec 14, 2007

ADP + olig +  $\boxed{\text{FGF-2} + \text{CsA}}$   $\rightarrow 500 \text{ mM Ca}^{2+}$ 

Dec 14, 2007



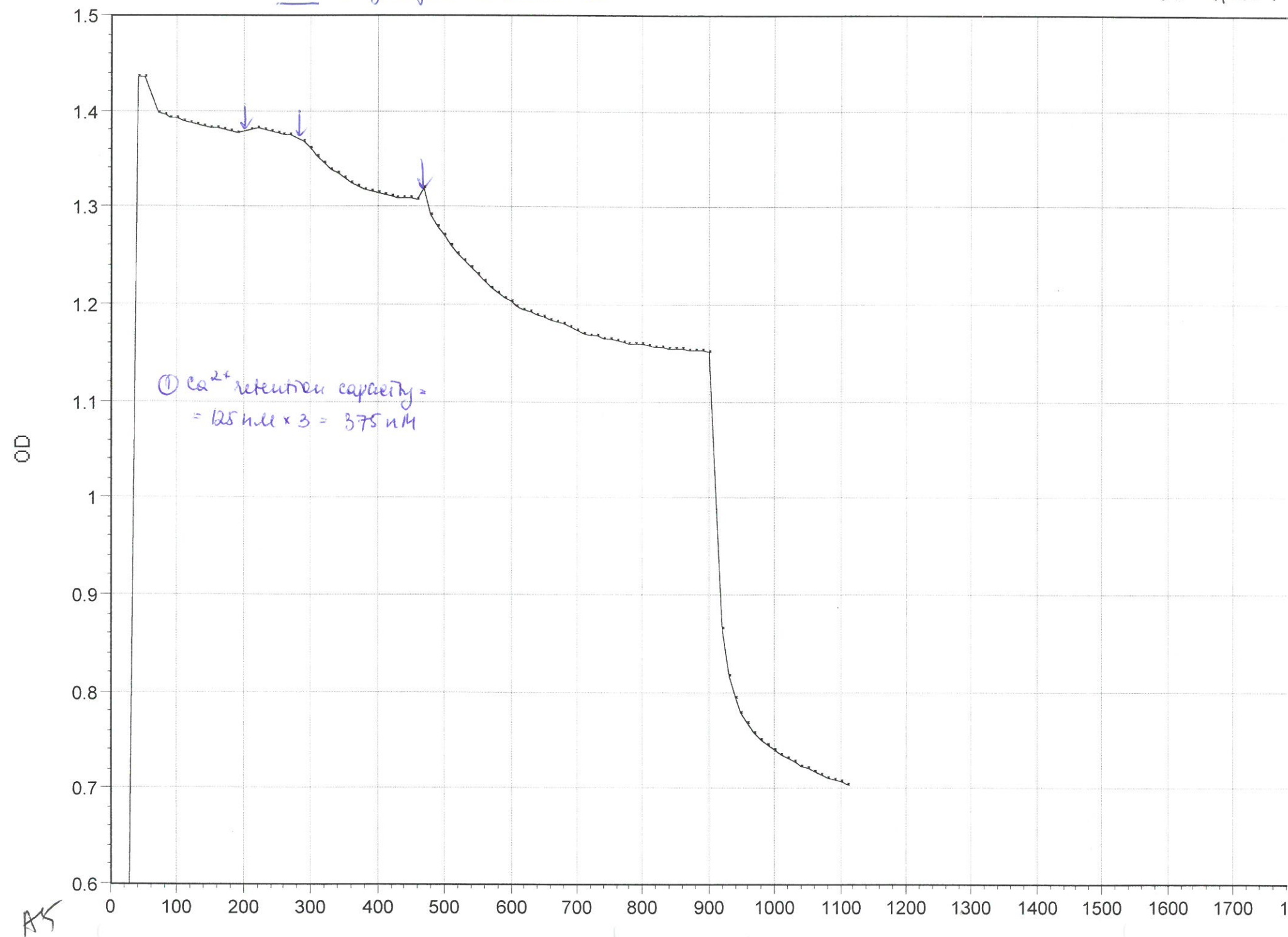




SSM

ATP + oligomycin  $\rightarrow$  50 mM  $\text{Ca}^{2+}$

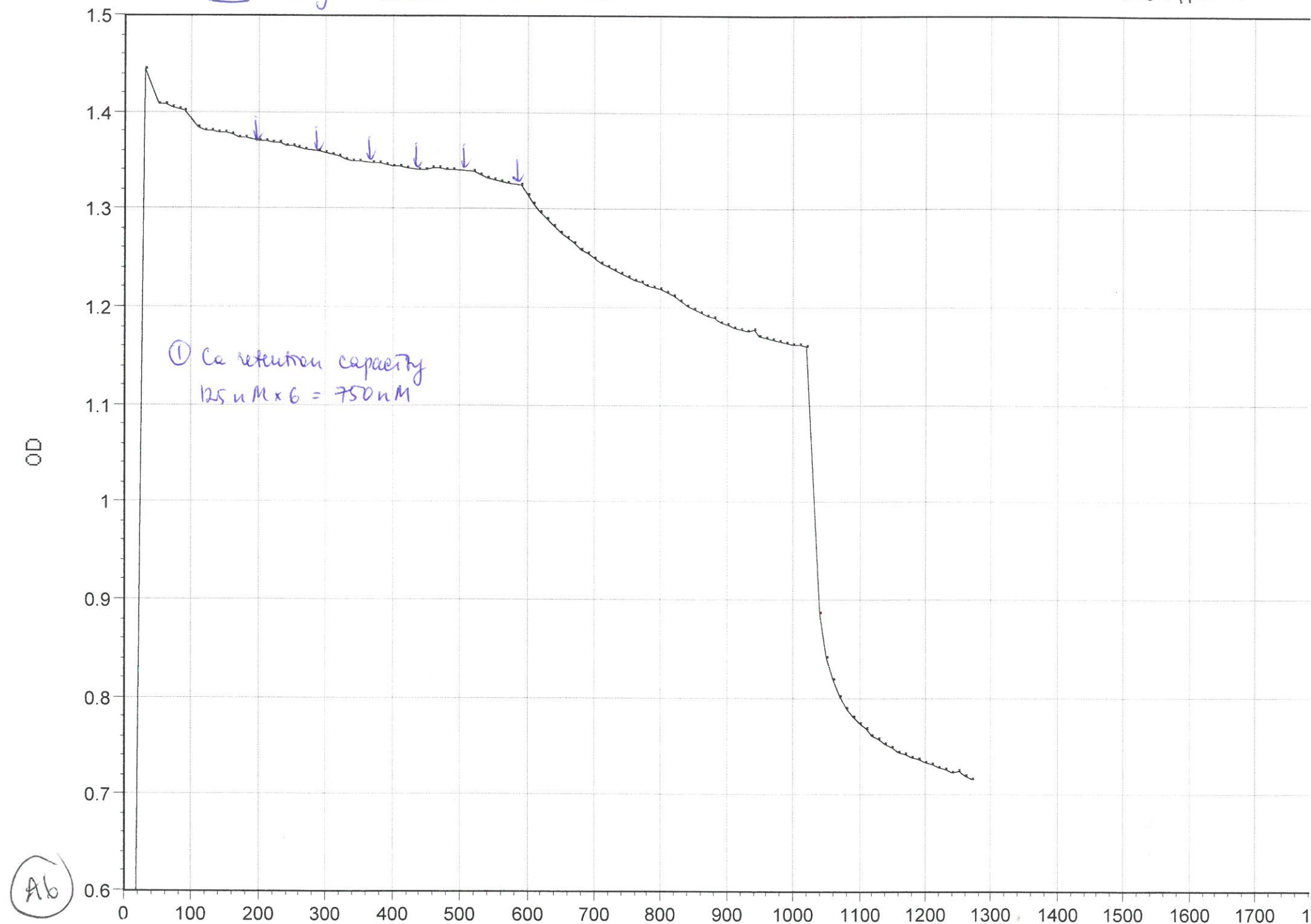
DEC 14, 2007





ATP + olig + FGF2  $\rightarrow$  50 mM  $\text{Ca}^{2+}$

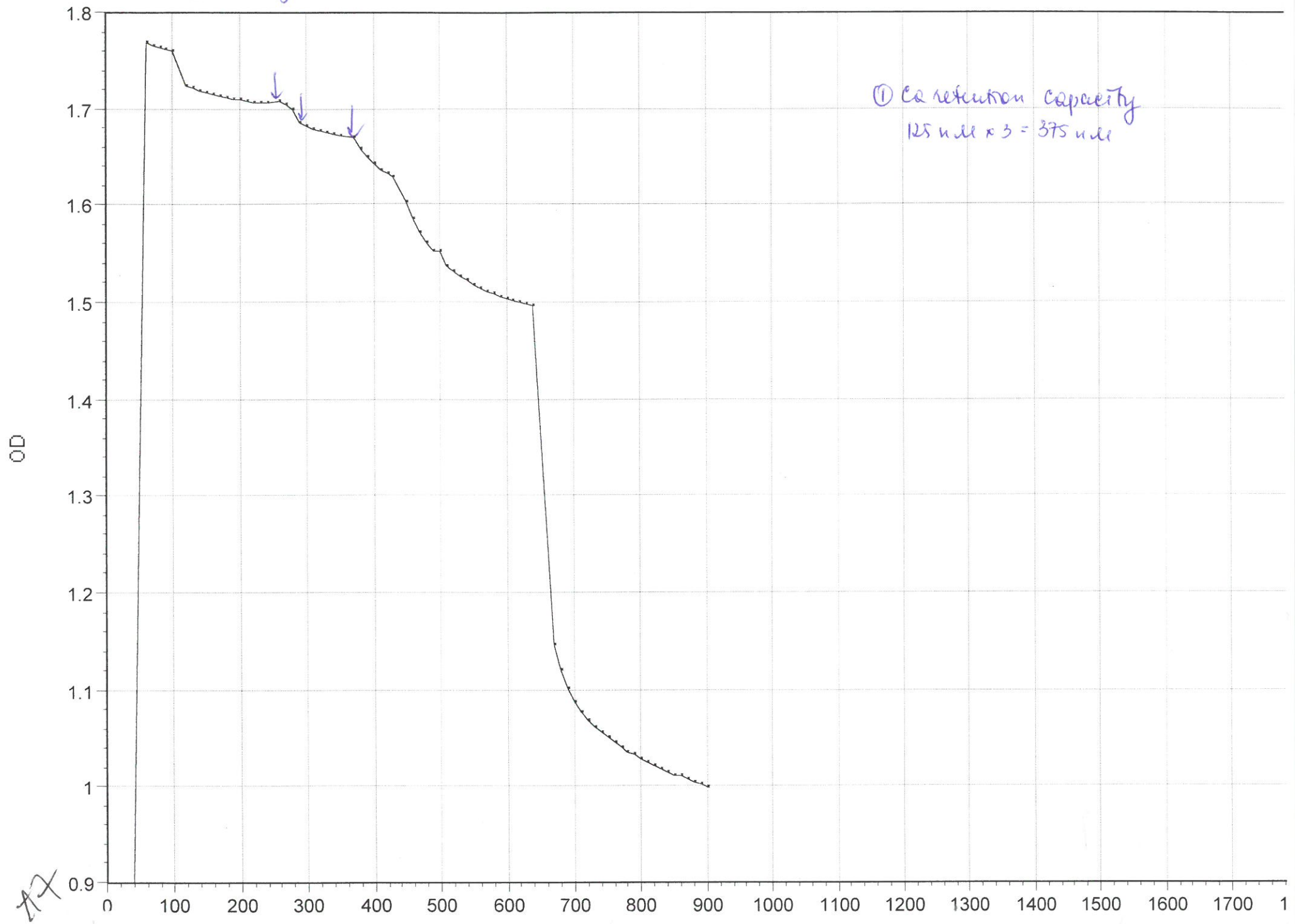
Dec 14, 2007



ATP + olig + FGF-2 + PKCinh.

Dec 14, 2007

SSM

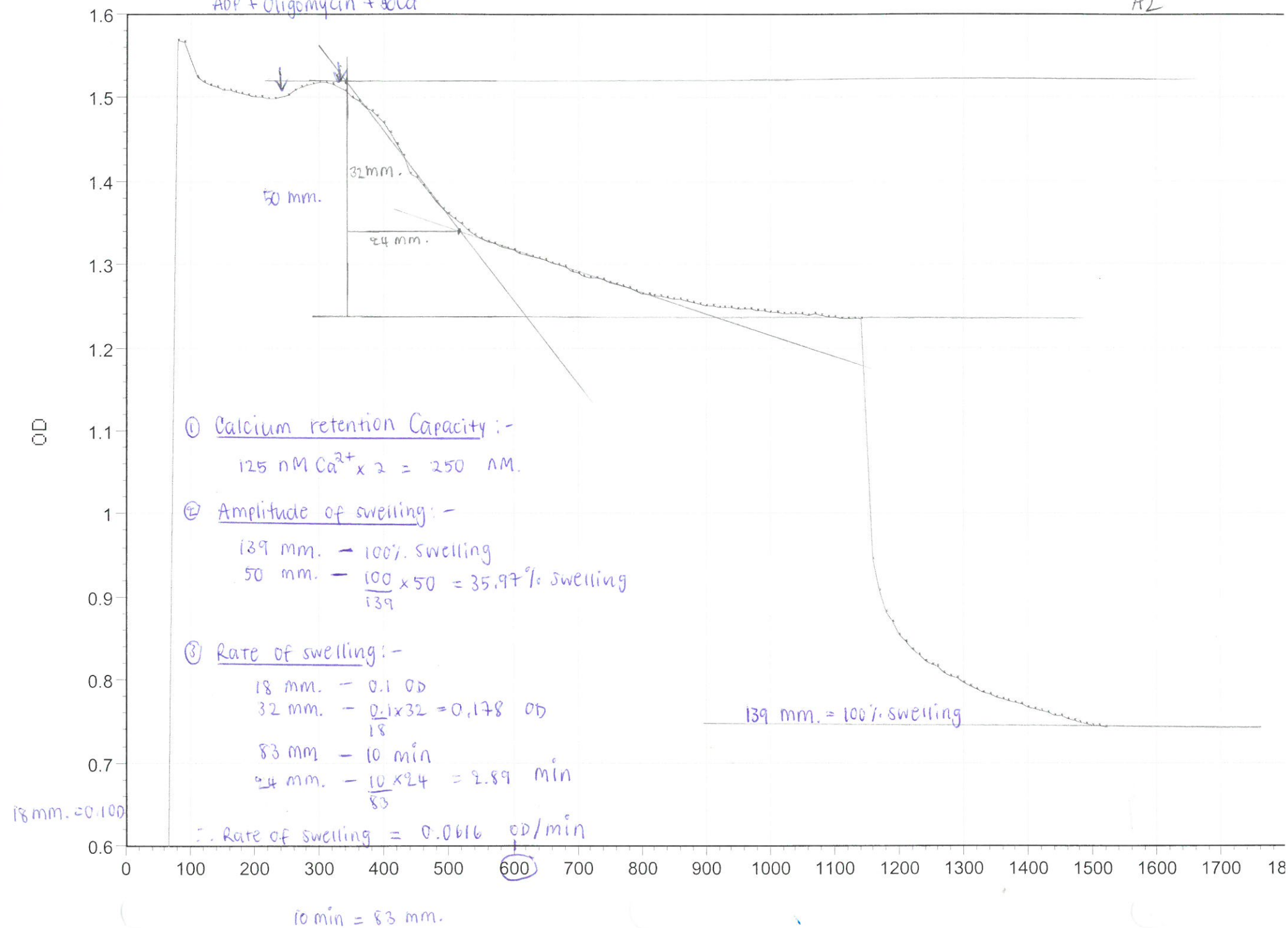


17

10 min = 83 mm.

SSM

• FGF2 + PKC $\epsilon$  inhibitor  
ADP + Oligomycin +  $\text{Ca}^{2+}$





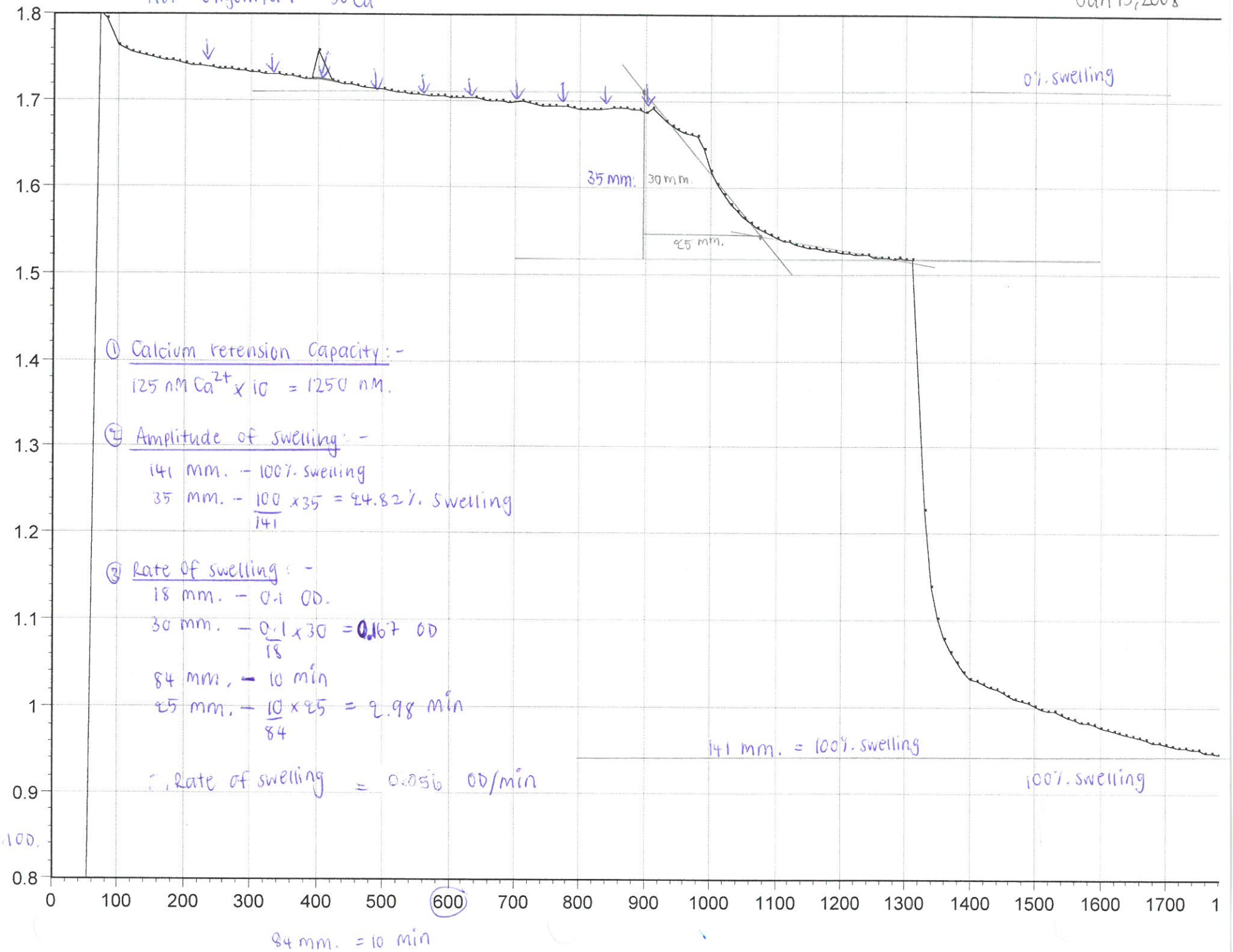
• FGF2 + PKC $\epsilon$  scramble  
ADP + Oligomycin + 50 Ca $^{2+}$

A3  
Jan 15, 2008

SSM

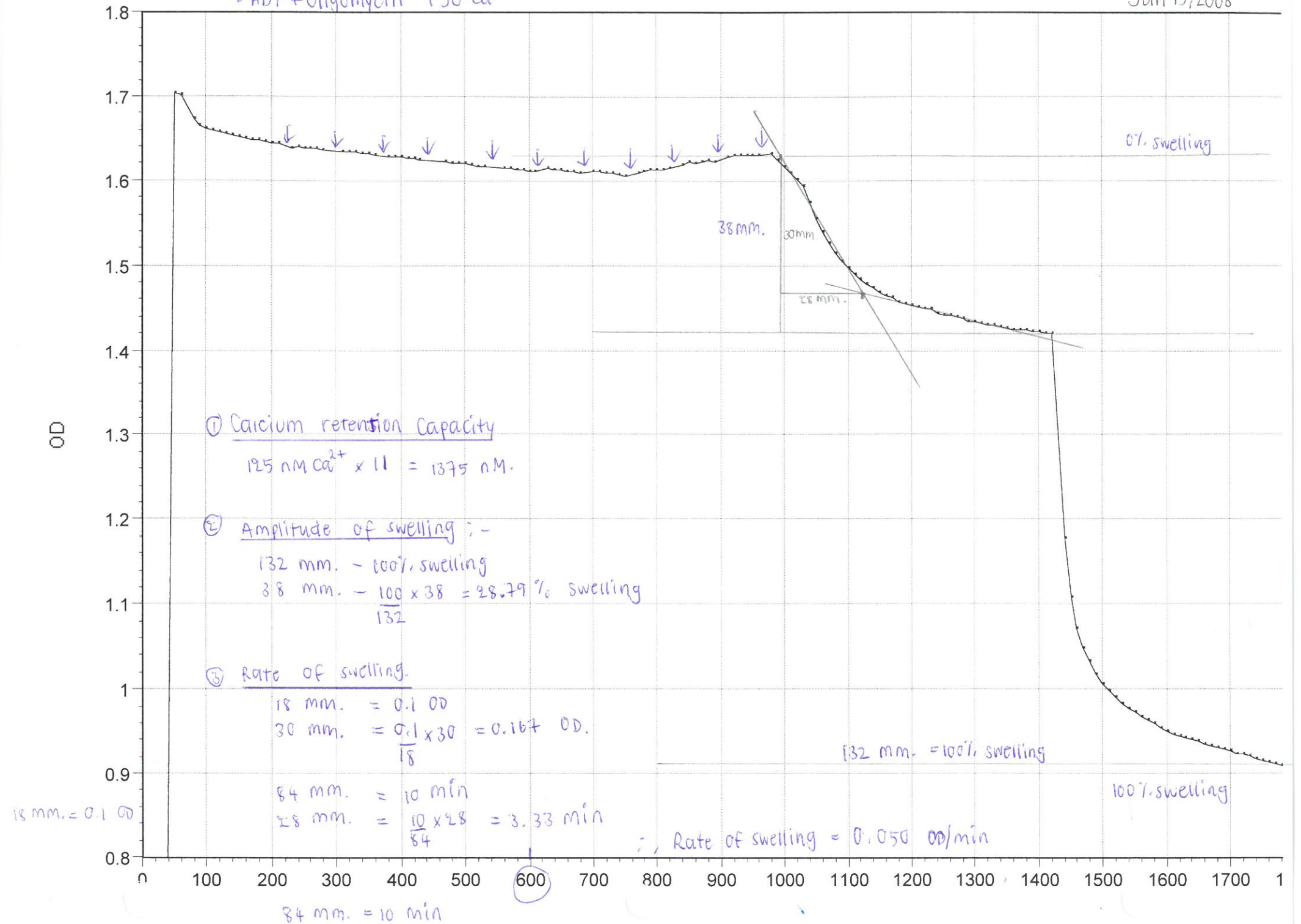
OD

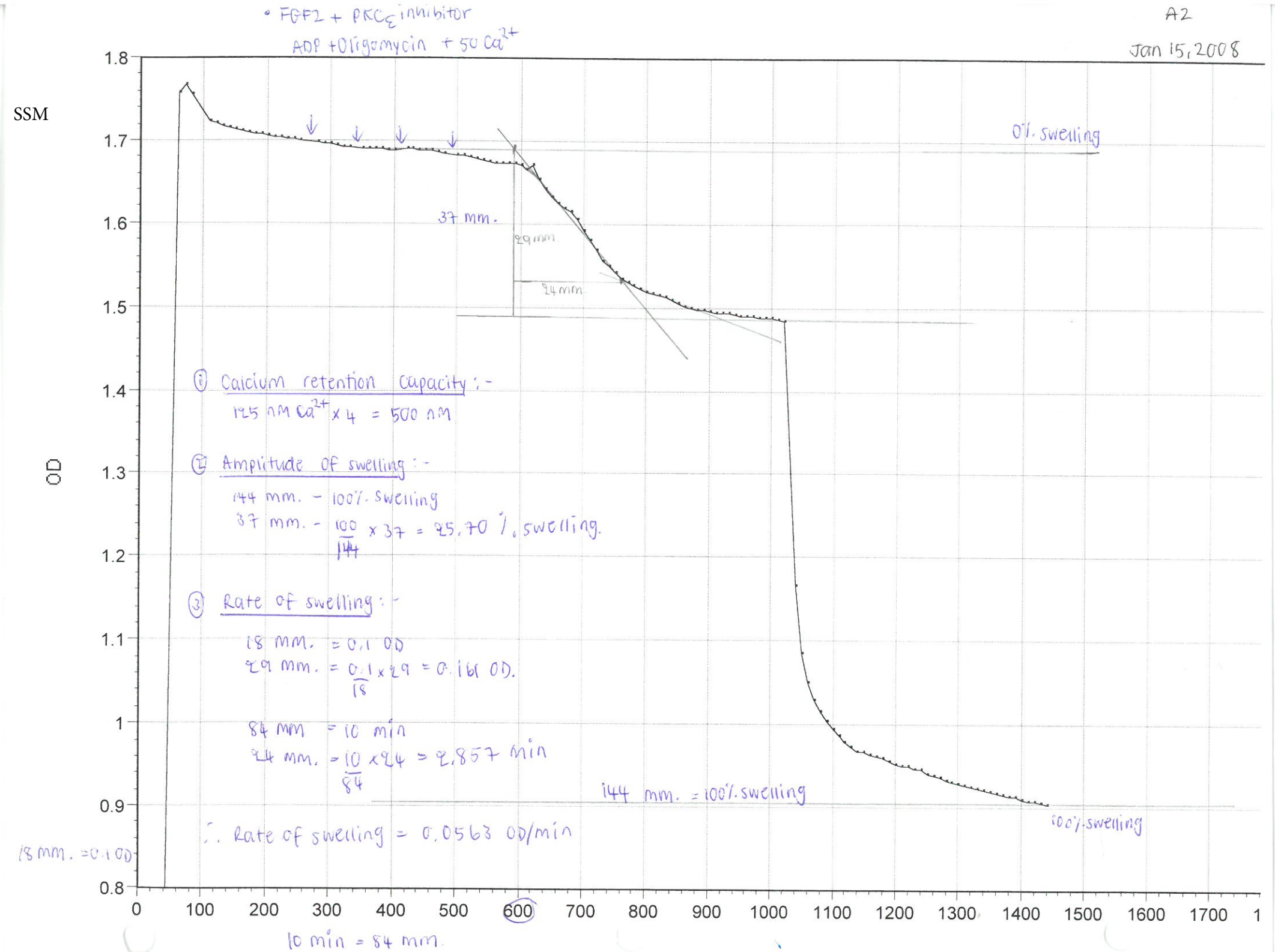
18 mm. = 0.100.



SSM

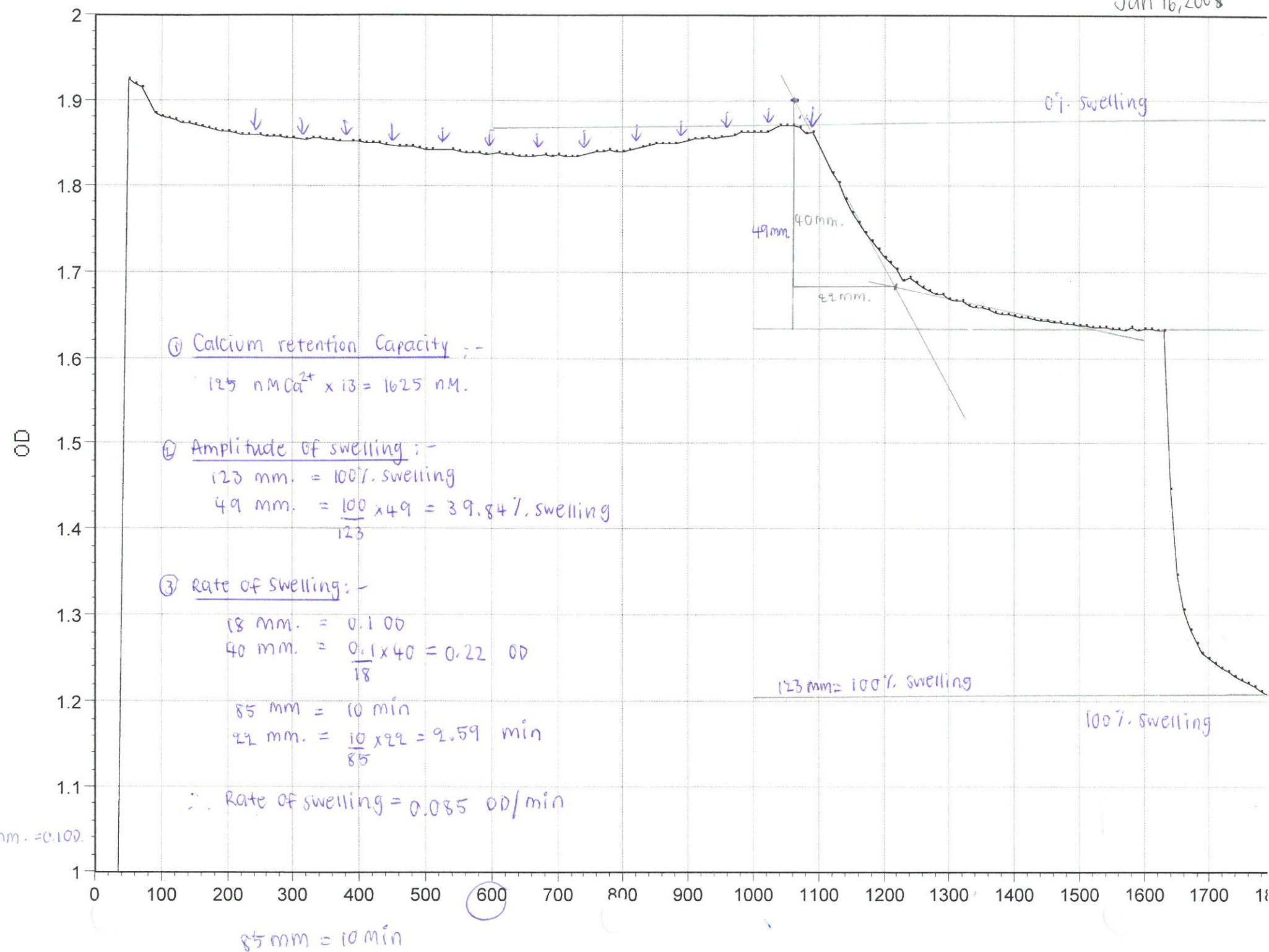
• FGF2  
• ADP + Oligomycin + 50  $\text{Ca}^{2+}$







SSM

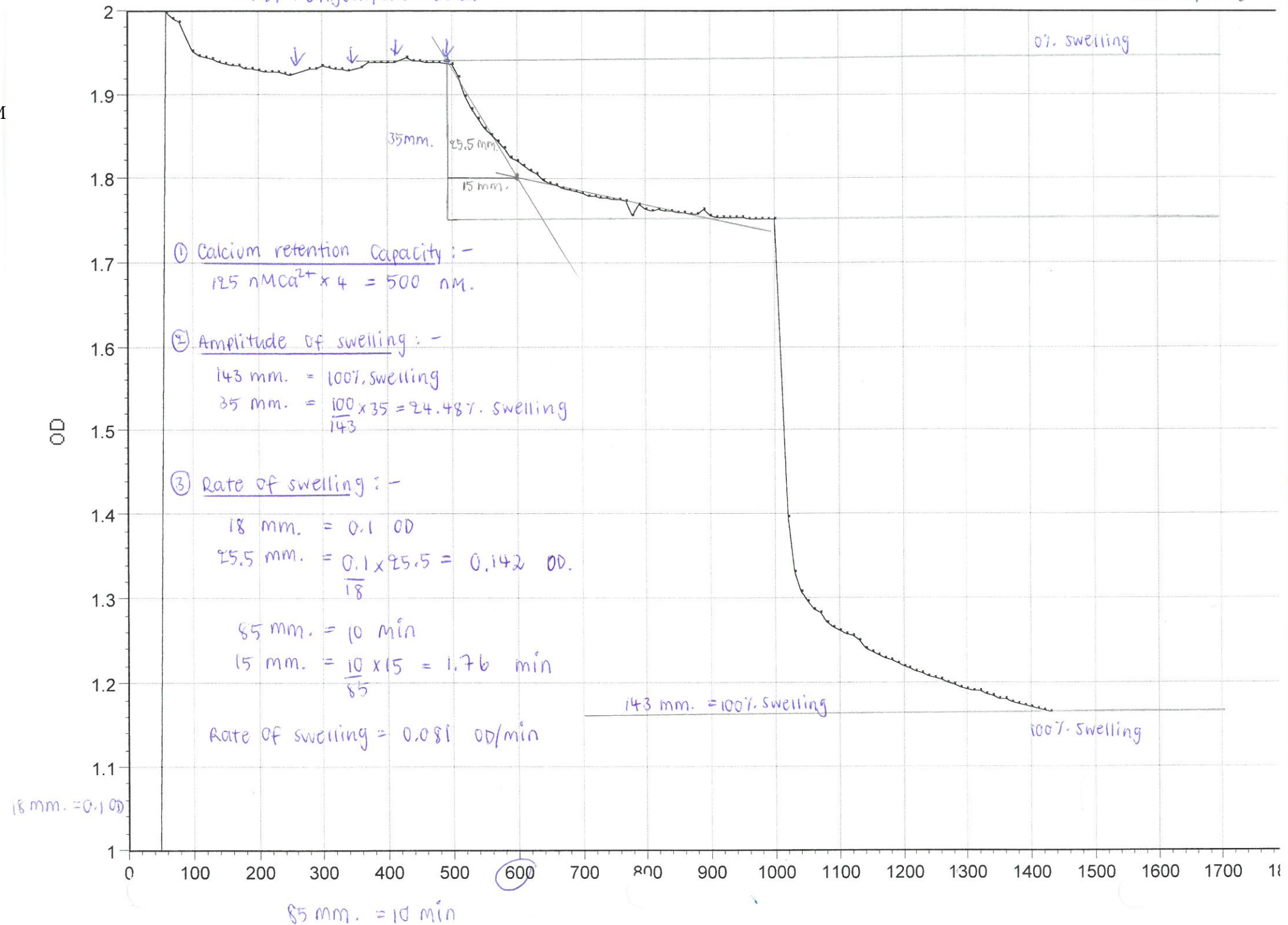




FGF2 + PKC<sub>E</sub> inhibitor  
 • ADP + Oligomycin + 50 Ca<sup>2+</sup>

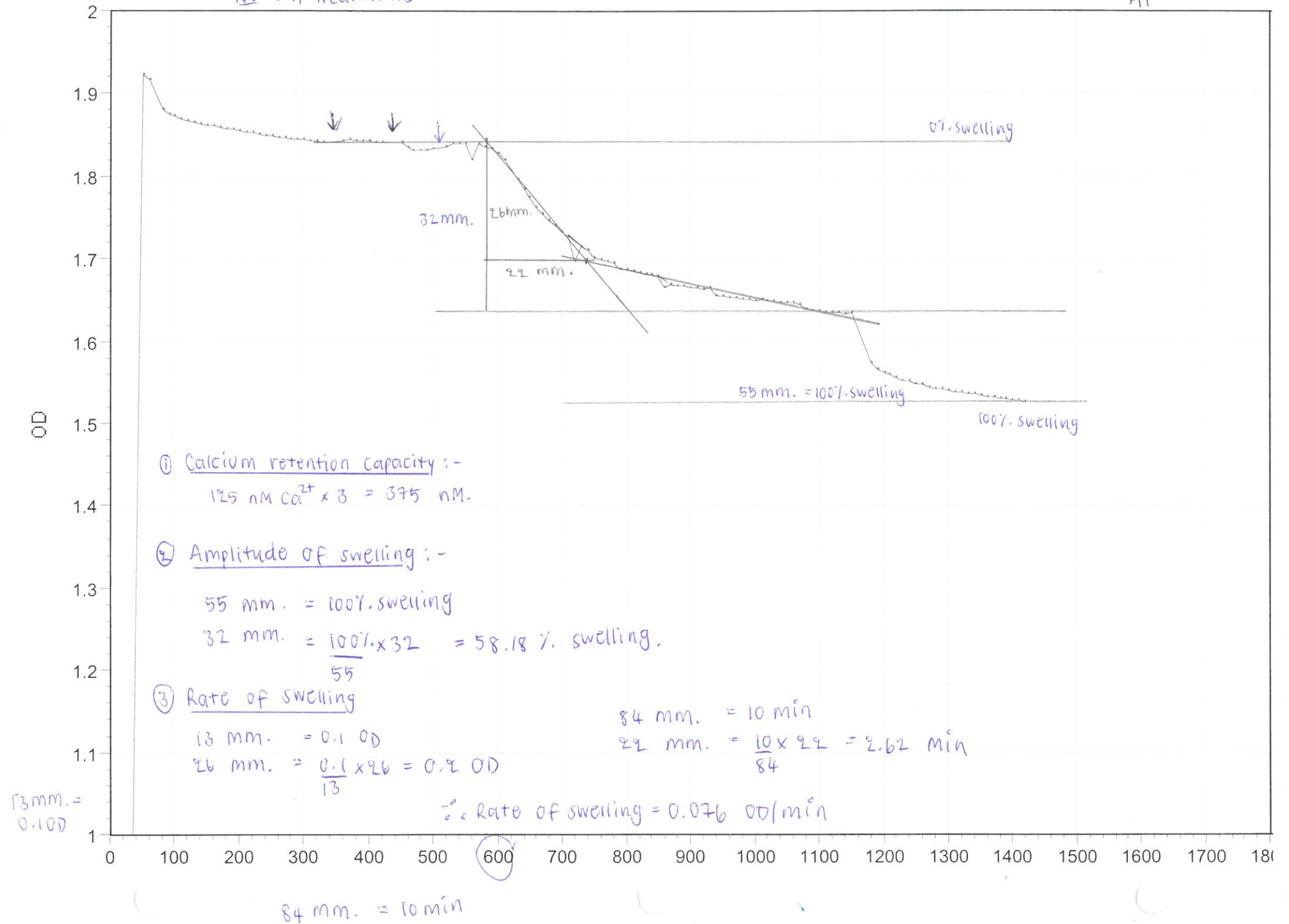
A2  
 Jan 16, 2008

SSM

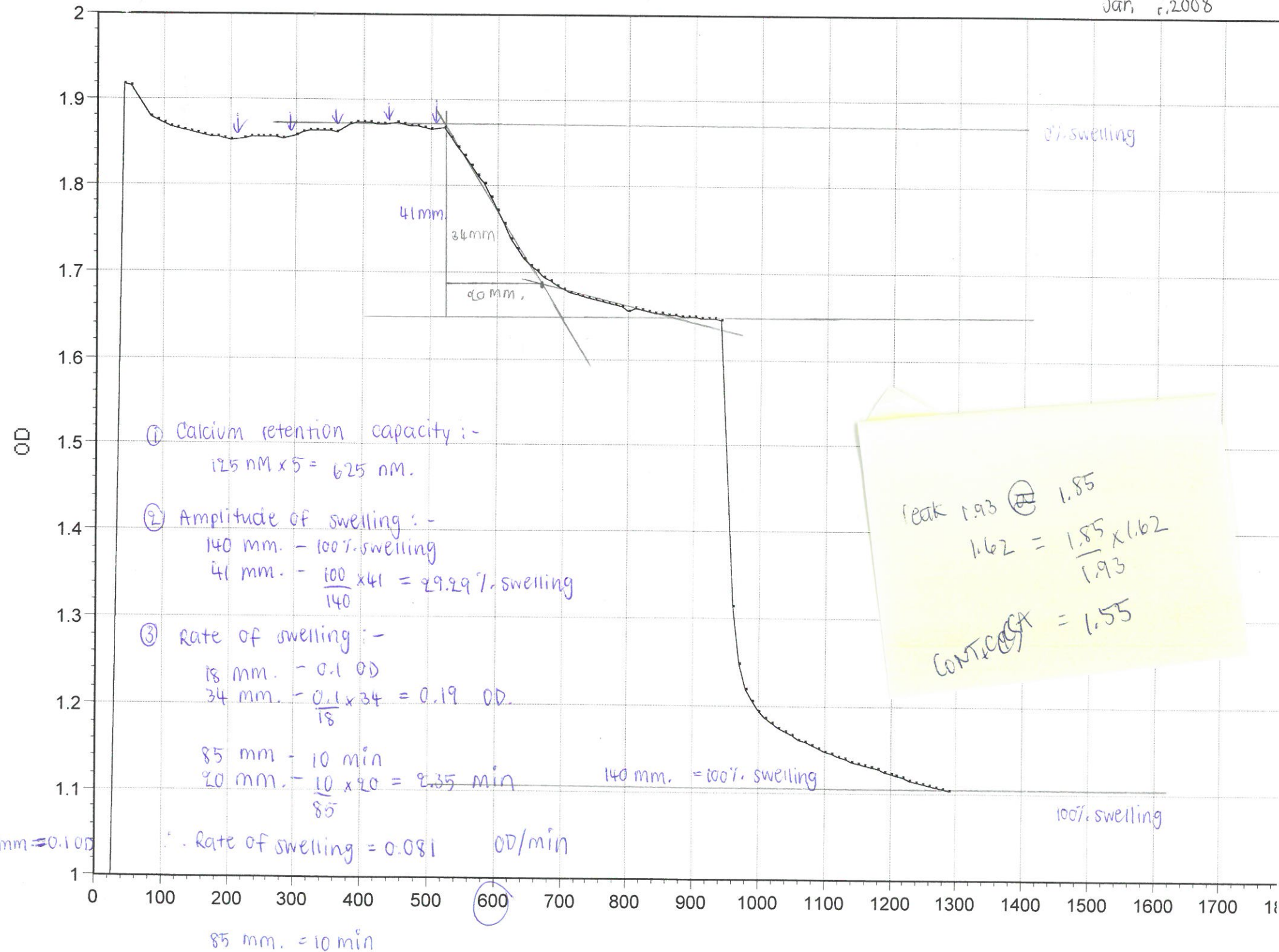


No any treatments

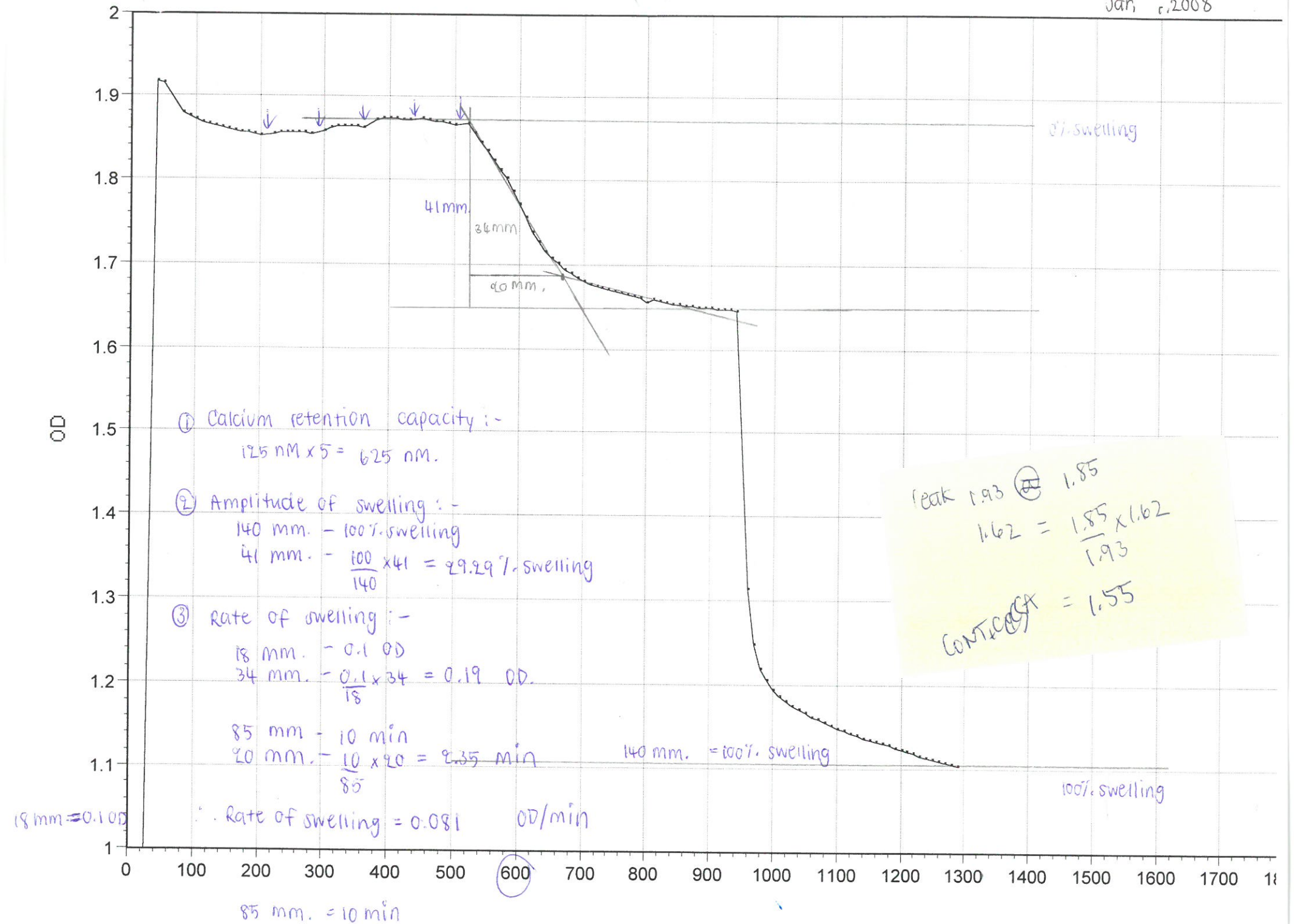
A1



Jan, 2008





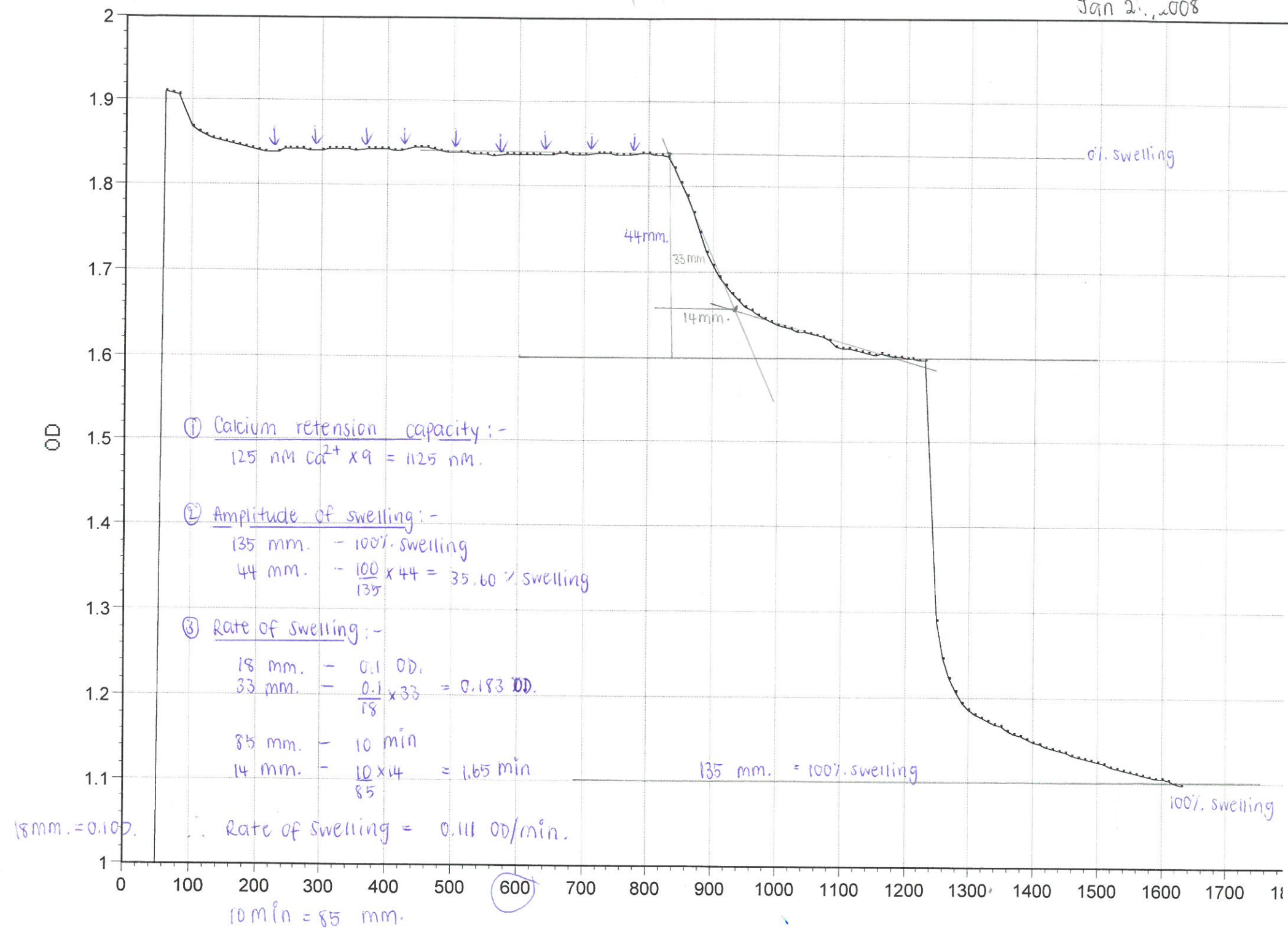


SSM

ADP + Oligomycin + F<sub>1</sub>F<sub>2</sub> (x9)

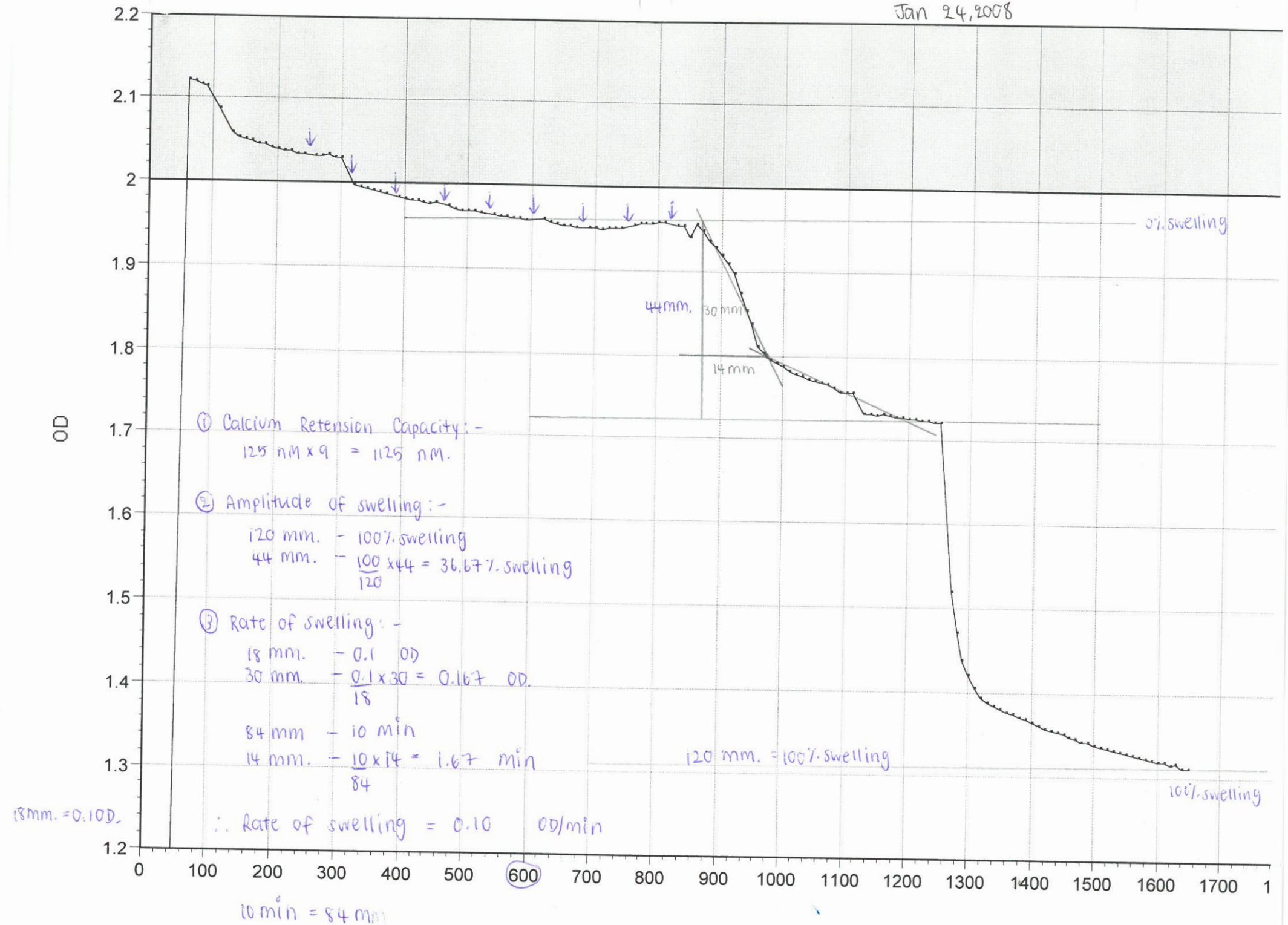
A2 (RHM)

Jan 21, 2008





Jan 24, 2008



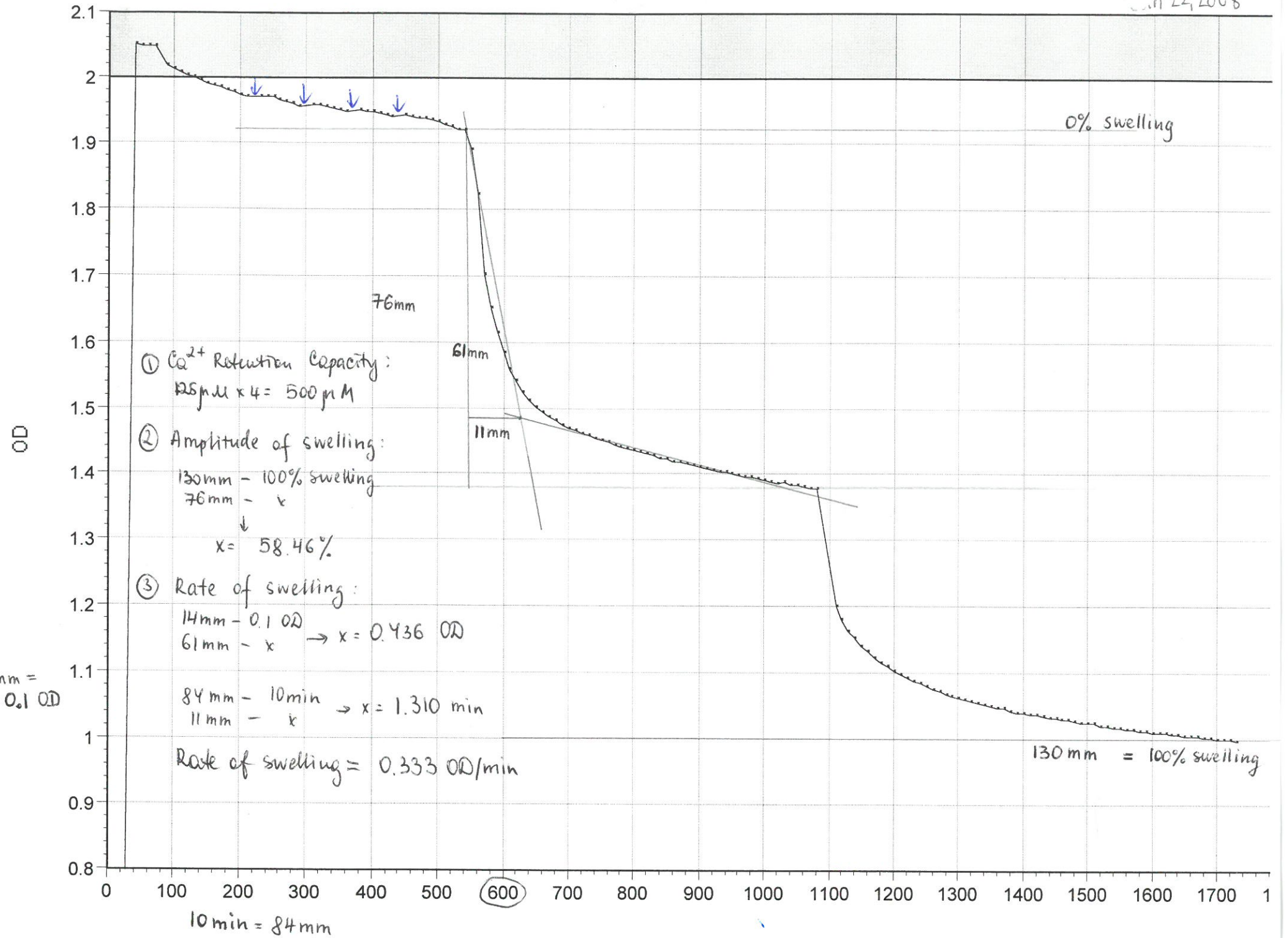
ADP + Oligo  $\rightarrow$  50 mM  $\text{Ca}^{2+}$  (x4)

liver

Xi (RLM)

Jan 22, 2008

LIVER



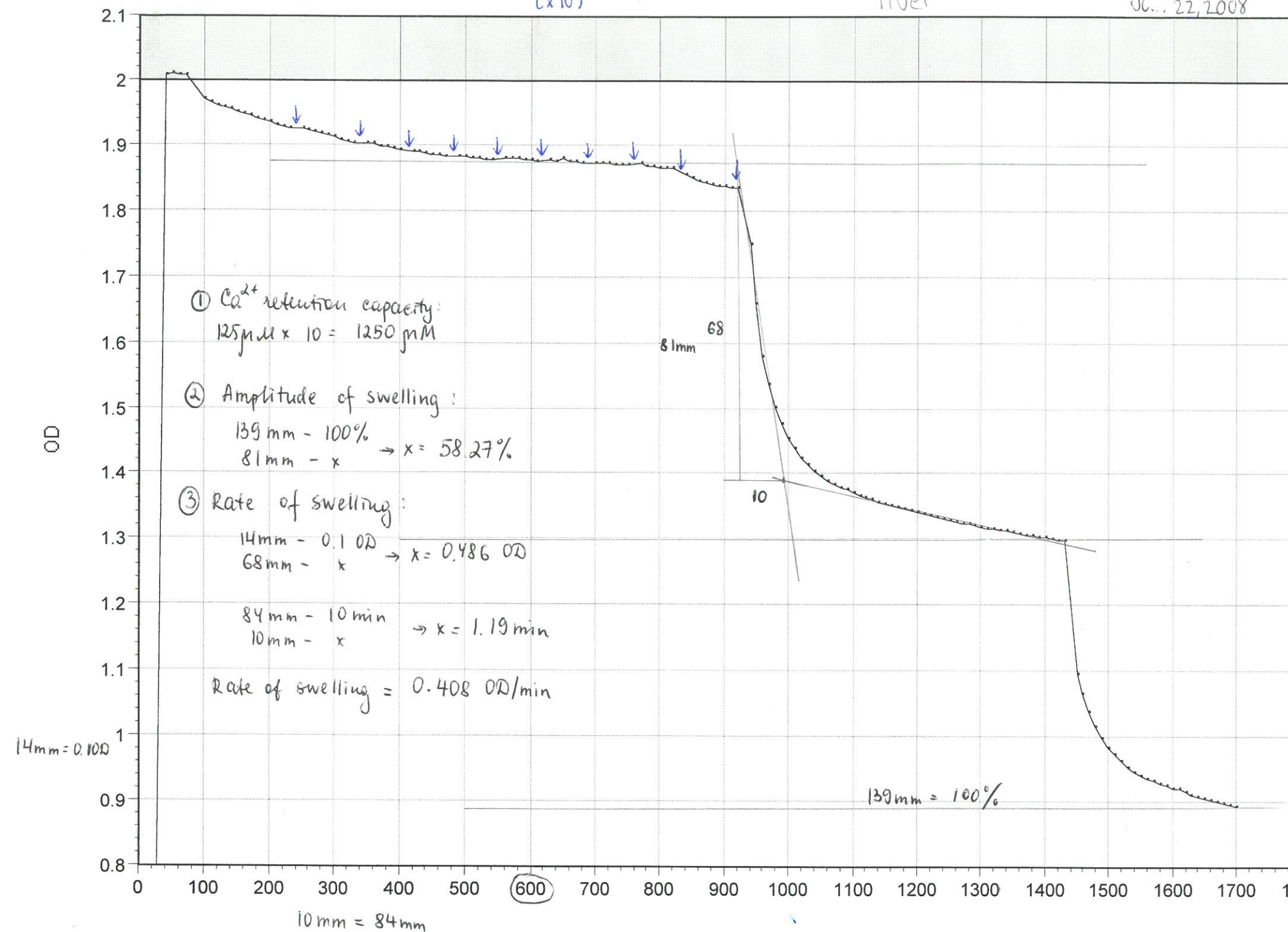


LIVER

ADP + Oligo + FGF2  $\rightarrow$  50 mM  $\text{Ca}^{2+}$   
(x 10)

liver

AZ (RLM)  
Jul 22, 2008



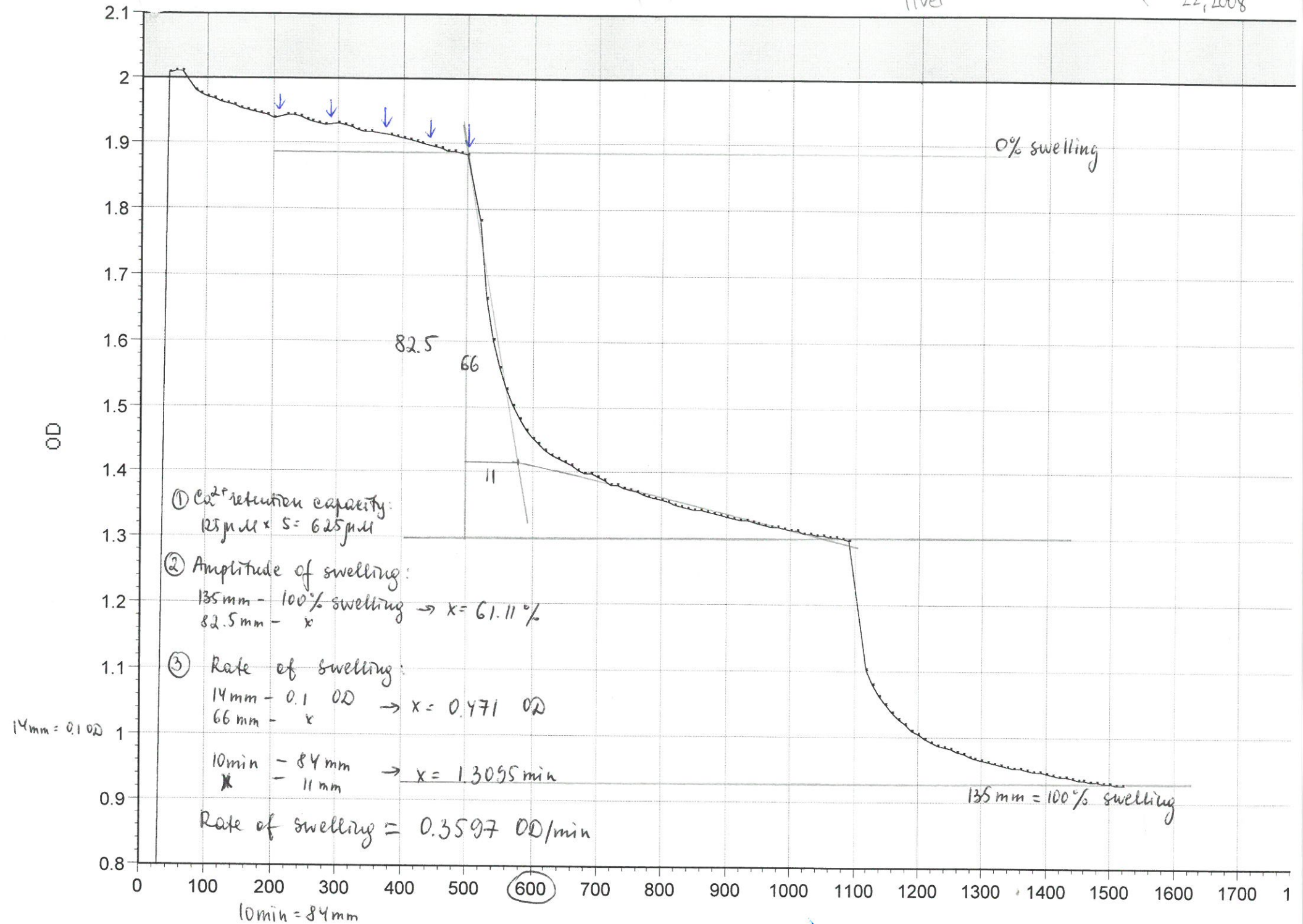


LIVER

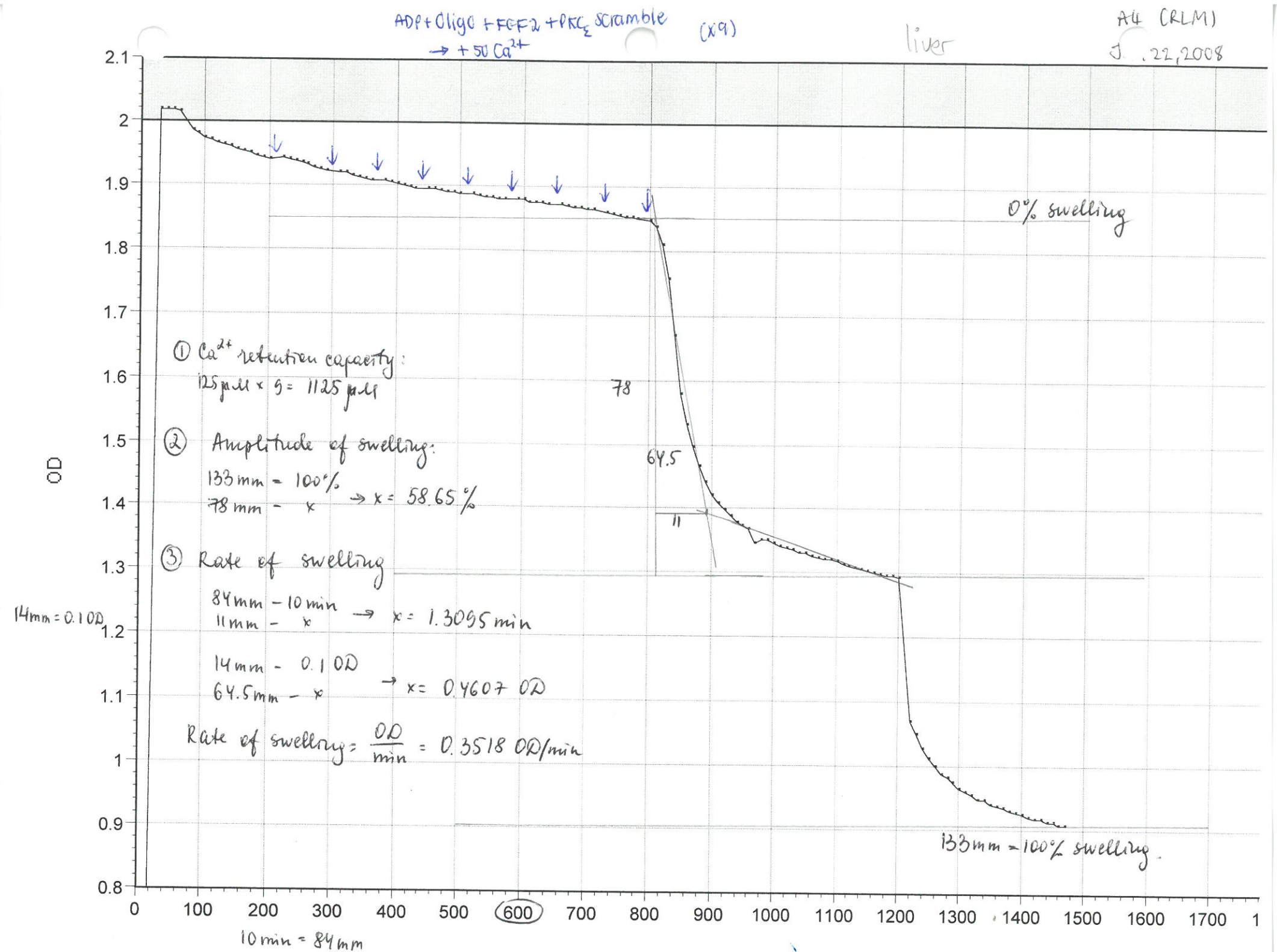
ADP + Oligo + FGF2 + PKC<sub>ε</sub> inhibitor → 50 mM Ca<sup>2+</sup> (x5)

liver

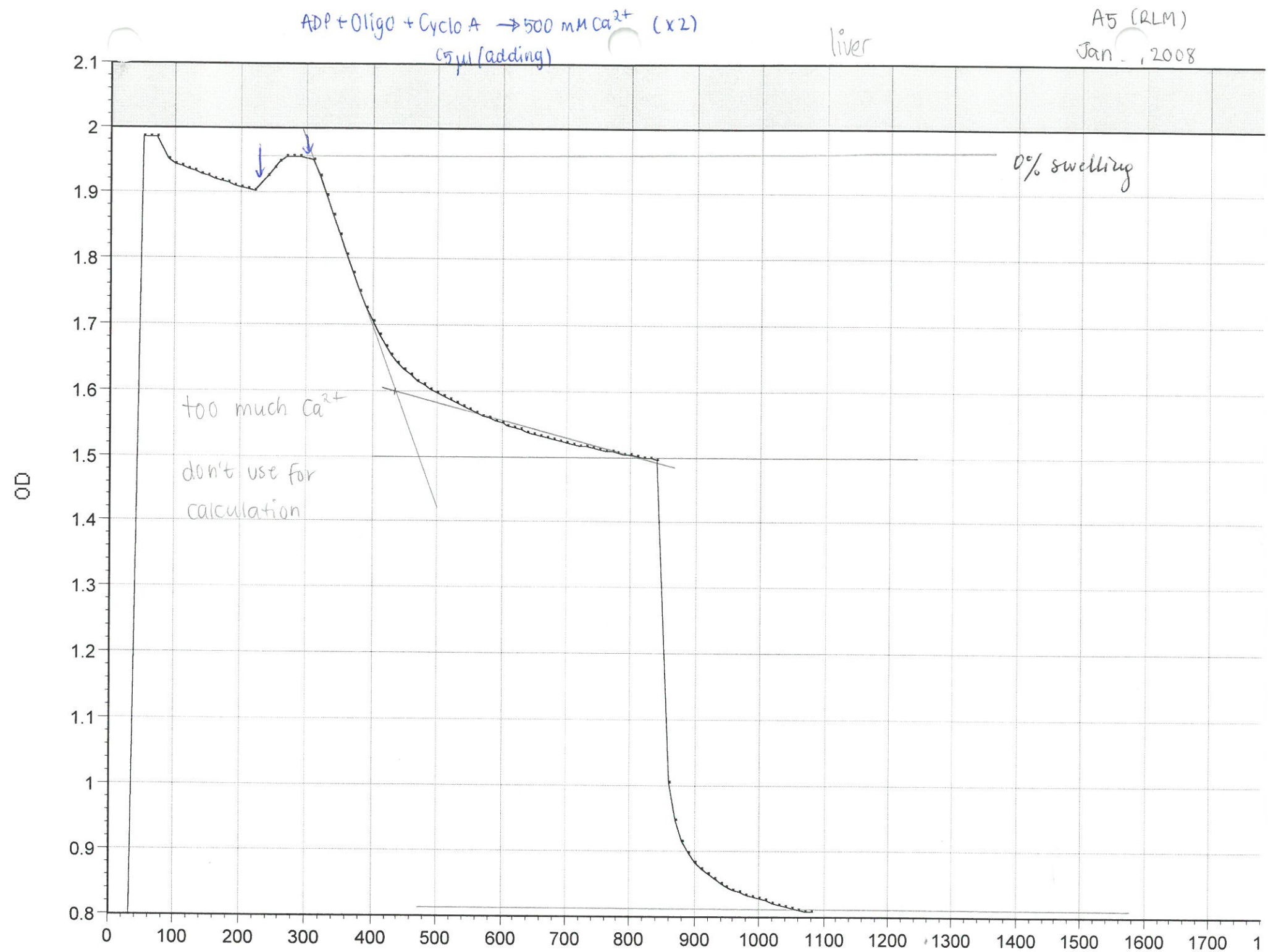
#3 (RLM)  
22, 2008



LIVER



LIVER





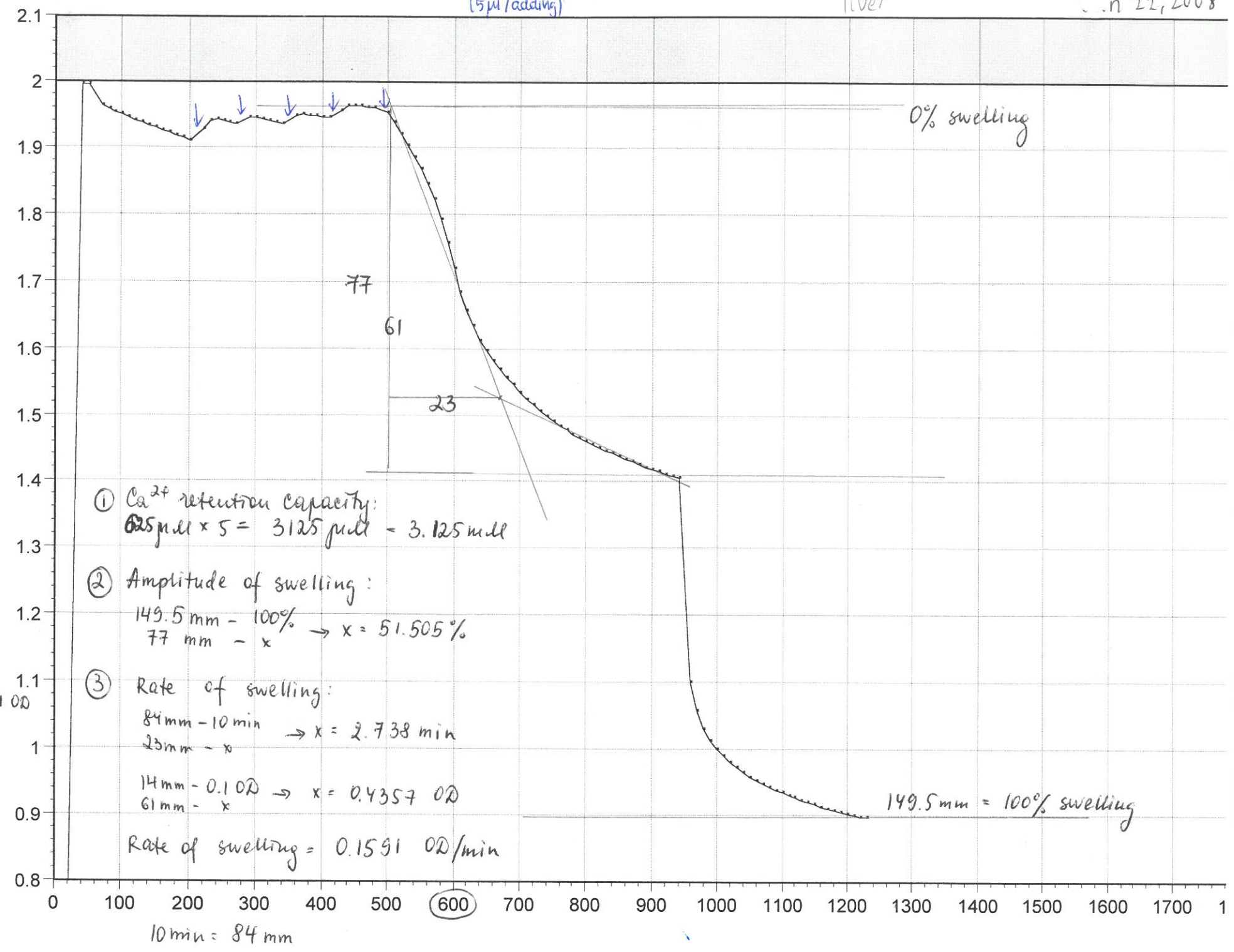
ADP + Oligo + Cyclo A  $\rightarrow$  250 mM  $\text{Ca}^{2+}$  (X<sup>-</sup>)  
(5  $\mu\text{l}$  / adding)

liver

Ab (RLM)  
n 22, 2008

LIVER

OD

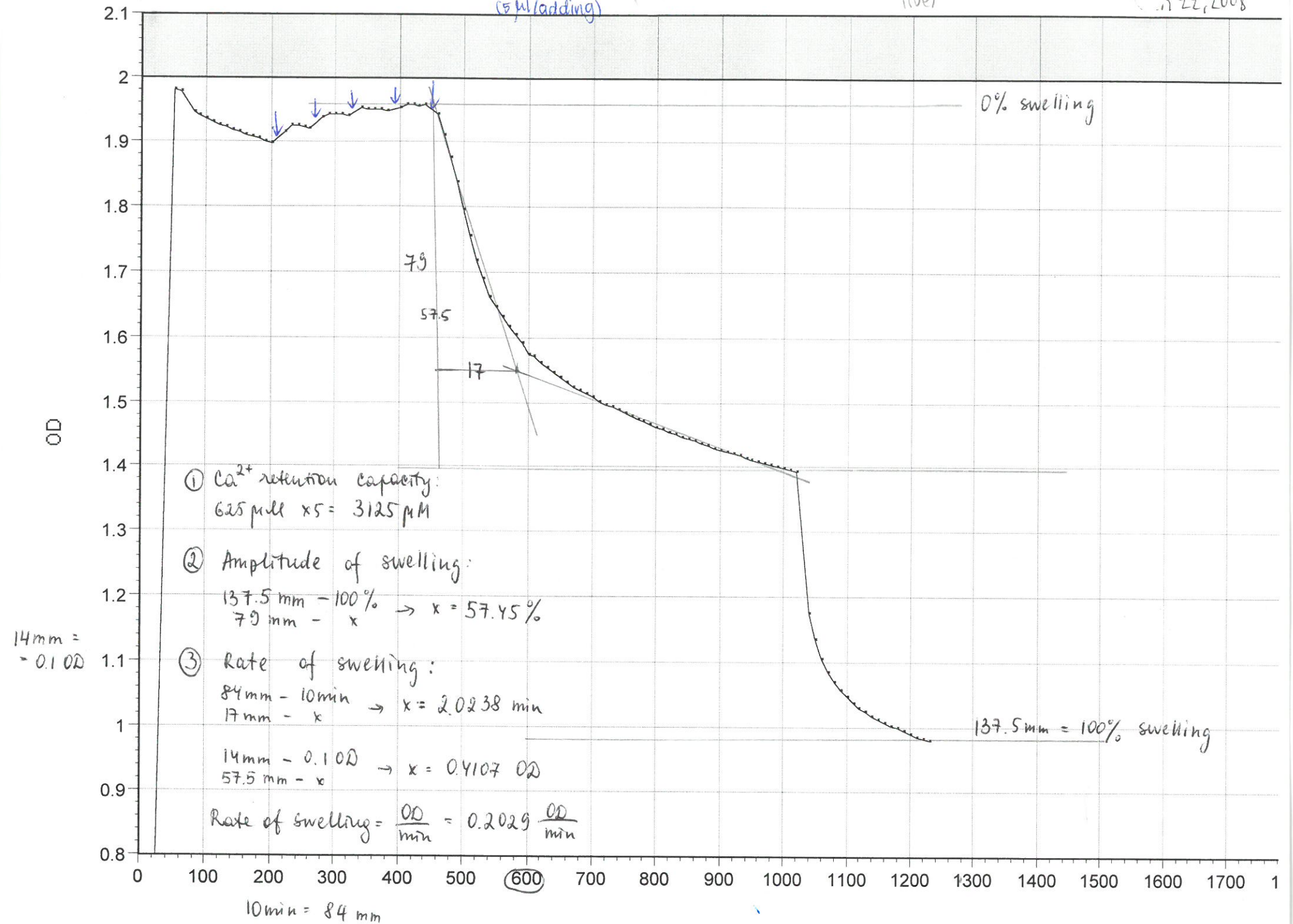


LIVER

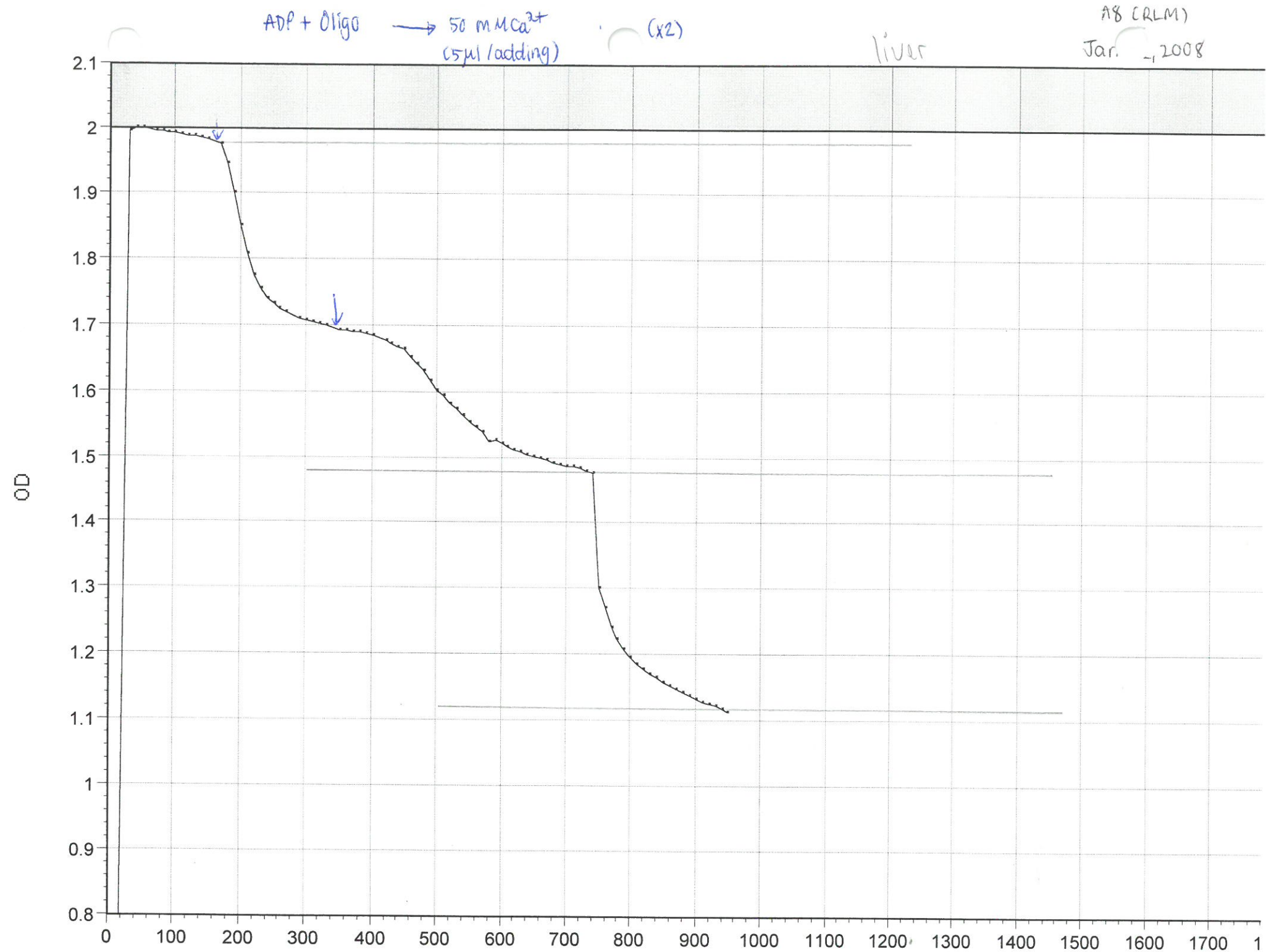
ADP + Oligo + Cyclo A + FGFA  $\rightarrow$  250 mM  $Ca^{2+}$  (X5)  
(5  $\mu$ l/adding)

liver

A7 (RLM)  
22, 2008



LIVER



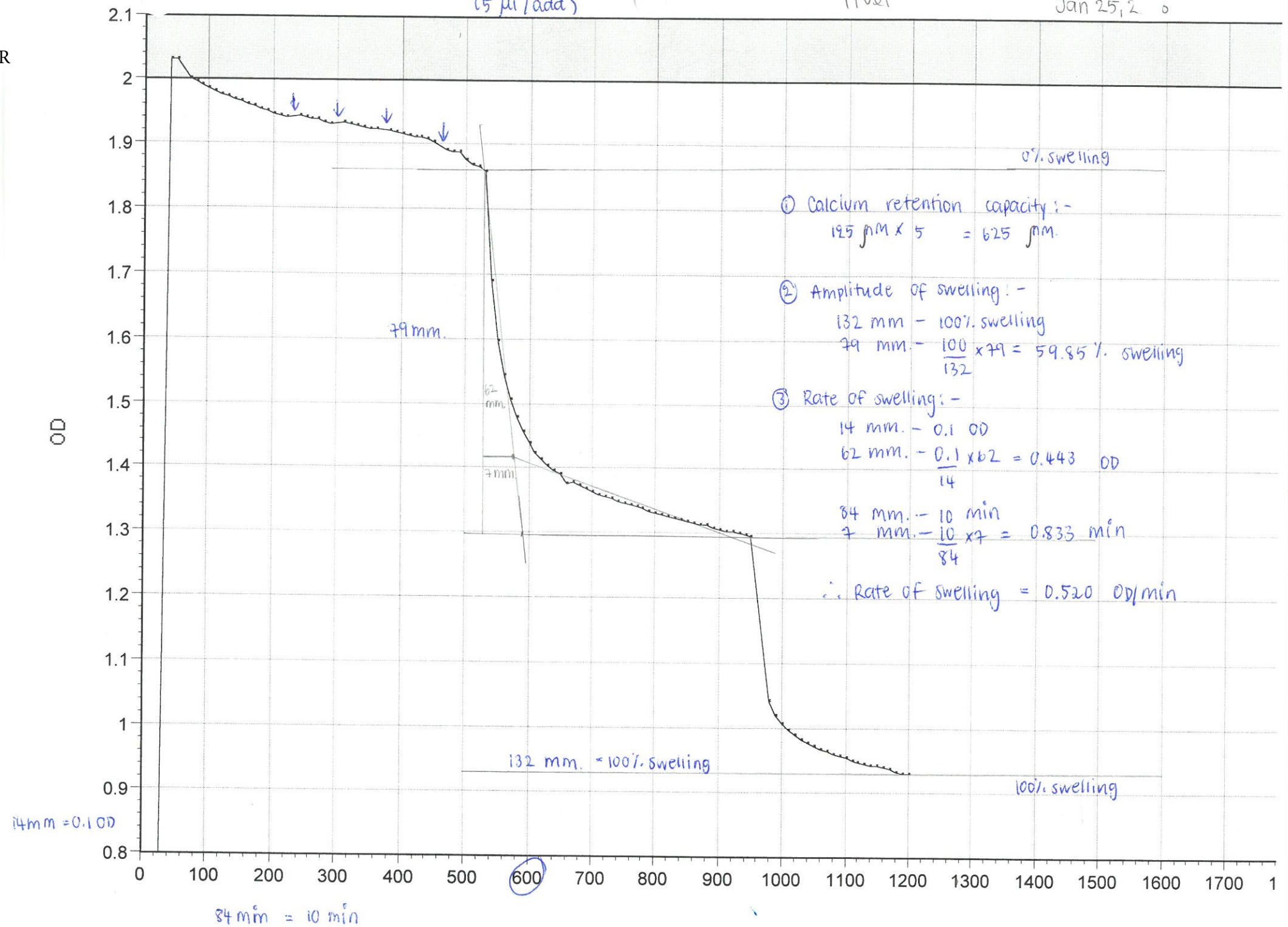


LIVER

ADP + Oligomycin  $\rightarrow$  50 mM  $\text{Ca}^{2+}$  (x)  
15  $\mu\text{l}$  / add)

liver

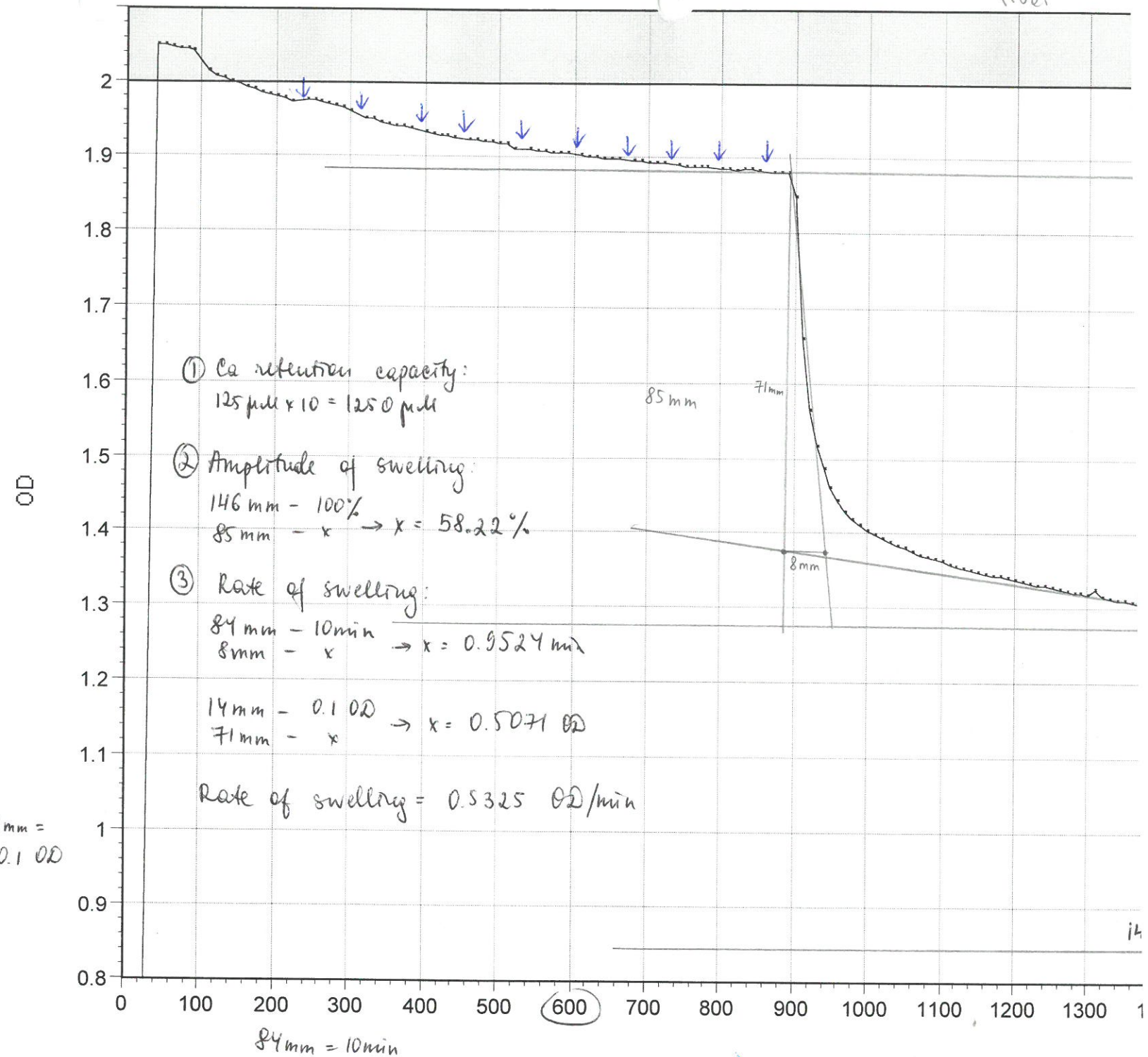
A1 (RLM)  
Jan 25, 20



LIVER

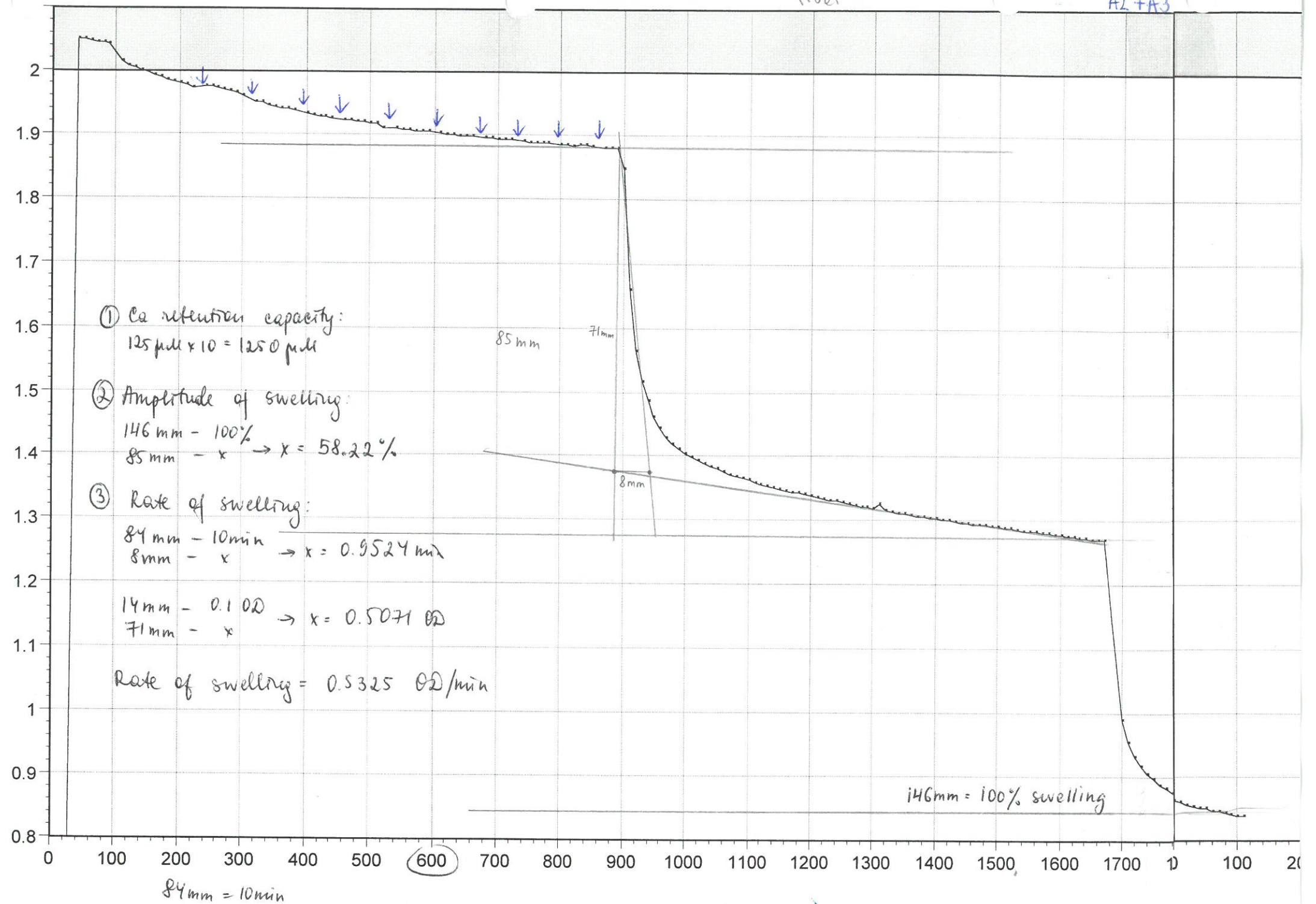
ADP + Oligomycin + FBF2 → 50 mM Ca<sup>2+</sup> (5 μl/adding)

liver





(RLM) Jan 25, 2008  
A2 + A3



LIVER

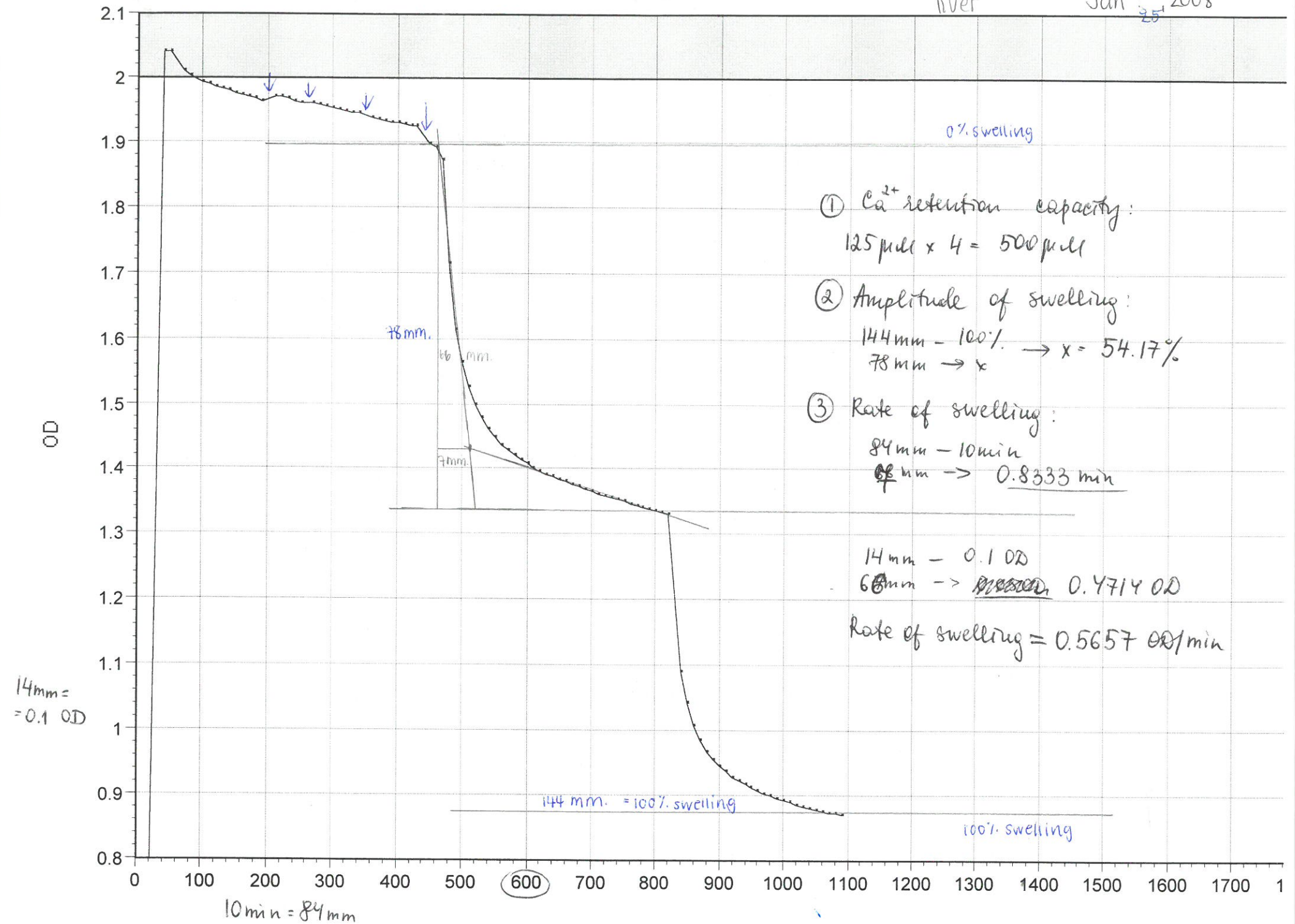
ADP + Oligo + PGE<sub>2</sub> + PKC<sub>2</sub> inh.

50mM Ca

liver

A4 (RLM)

Jan 25, 2008





LIVER

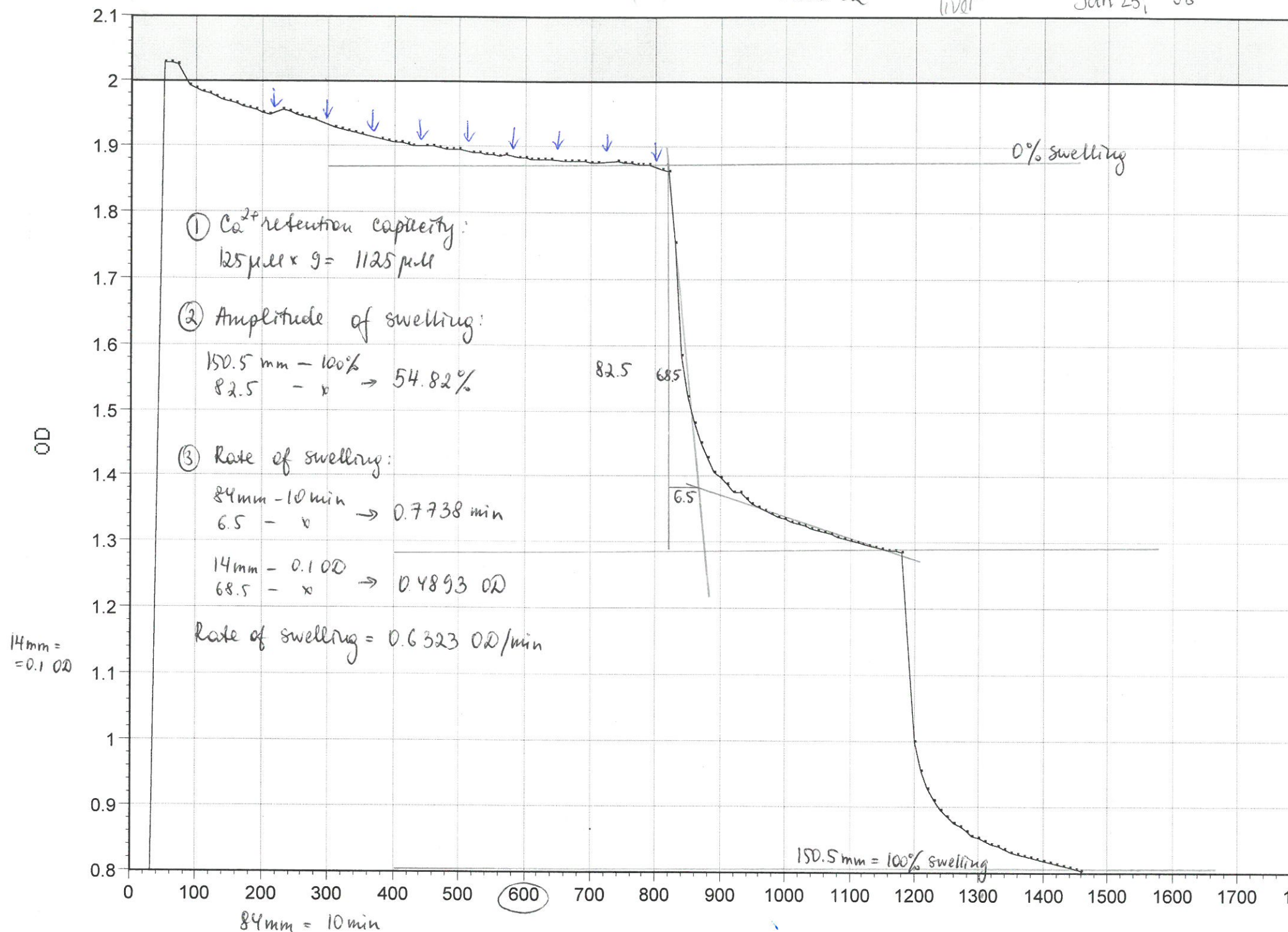
ADP + Oligo + FGF2 + PKC $\epsilon$  scramble (x9)

50ml Ca

liver

A5 (RLM)

Jan 25, 08



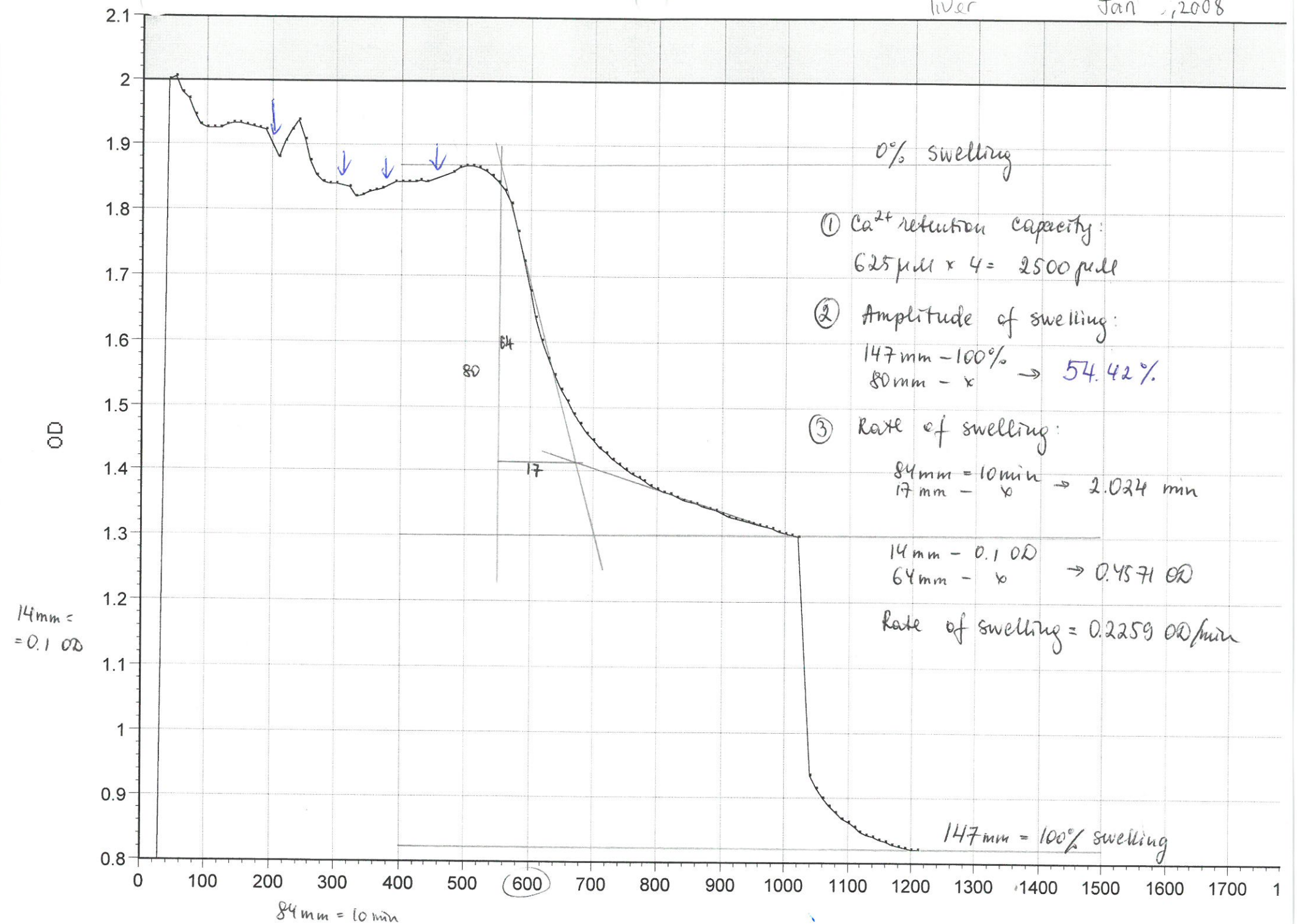


LIVER

ADP + Oligo + Cycle A  $\rightarrow$  250 mM  $\text{Ca}^{2+}$  (5  $\mu\text{M}$ /adding) (x4)

liver

Ab (KLM)  
Jan , 2008

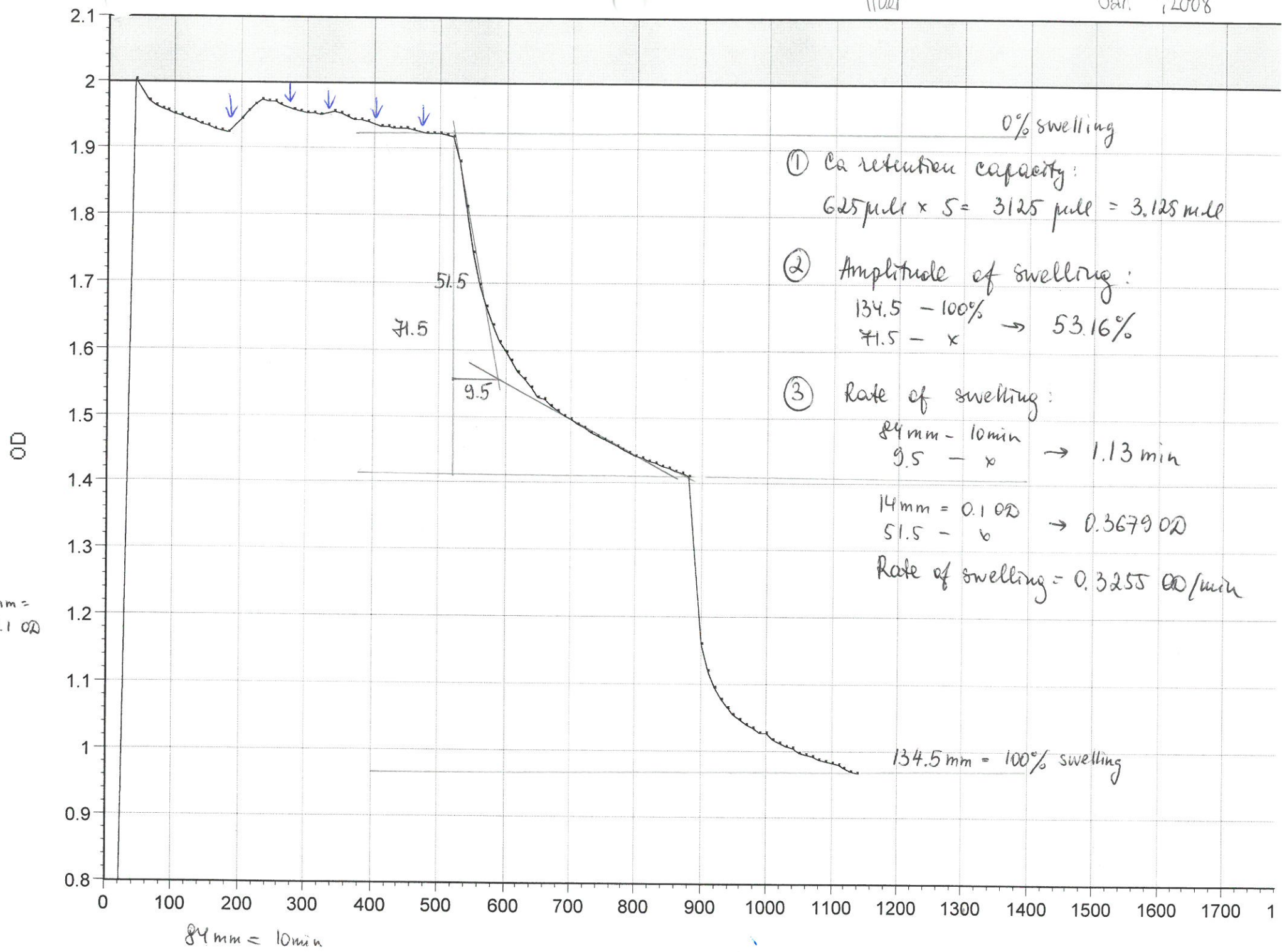


LIVER

ADP + Oligo + FGF2 + Cyclo A → 250 mM  $\text{Ca}^{2+}$  (5  $\mu\text{l}$ /adding)

liver

A7 (RLM)  
Jan. 2008



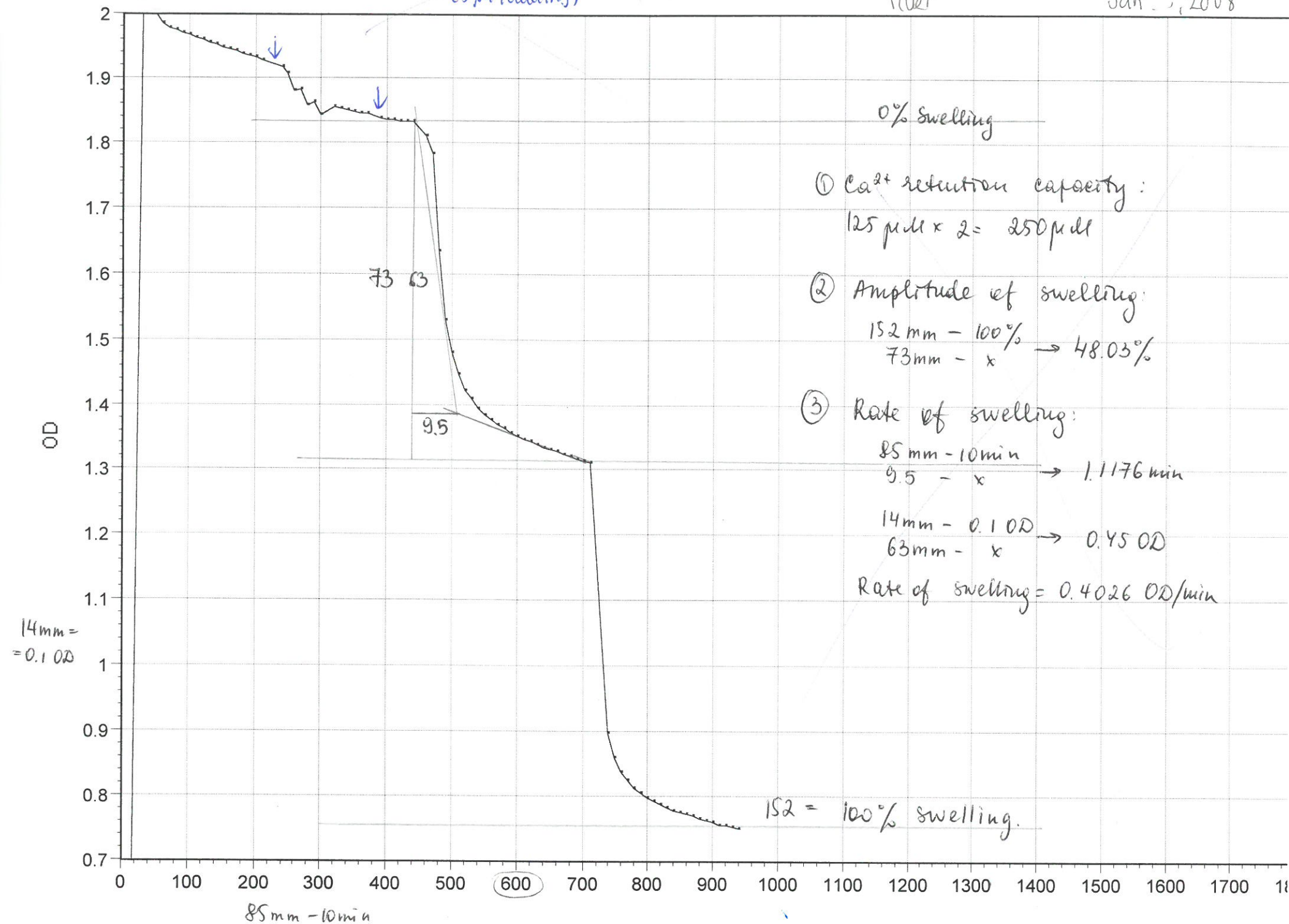


LIVER

ADP + Oligo → 50 mM Ca<sup>2+</sup>  
15 μl (adding)

liver

A8 (RLM)  
Jan 2008





A1

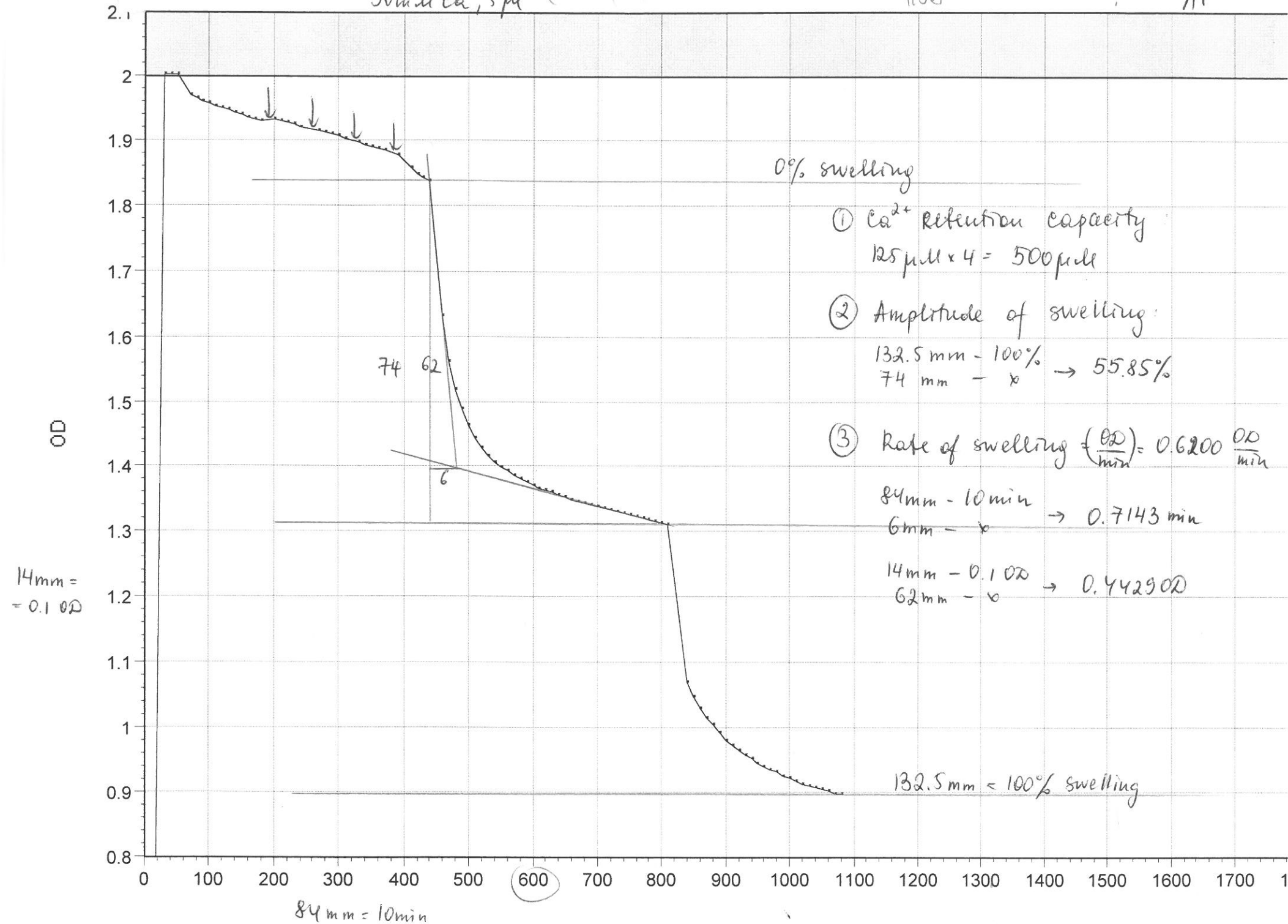
(ADP+olig) = Control  
50  $\mu$ M Ca, 5  $\mu$ M (X4)

liver

Feb 8 2008

A1

LIVER



A2

ADP+olig+ FGF2  $\rightarrow$  50  $\mu$ l cell  
5  $\mu$ l

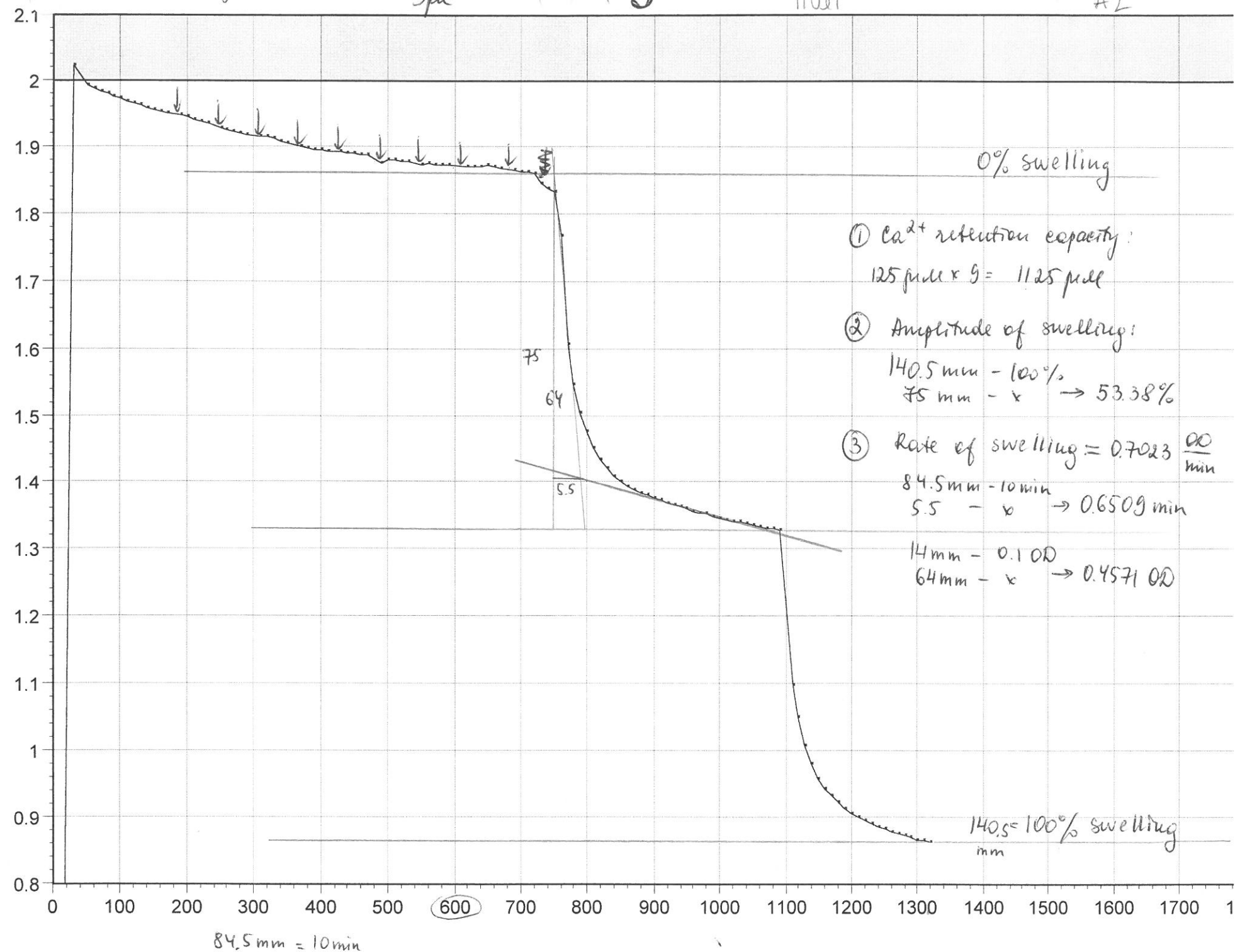
(x9)

liver

Feb 5, 2008  
A2

LIVER

OD

14 mm =  
= 0.1 OD

A3

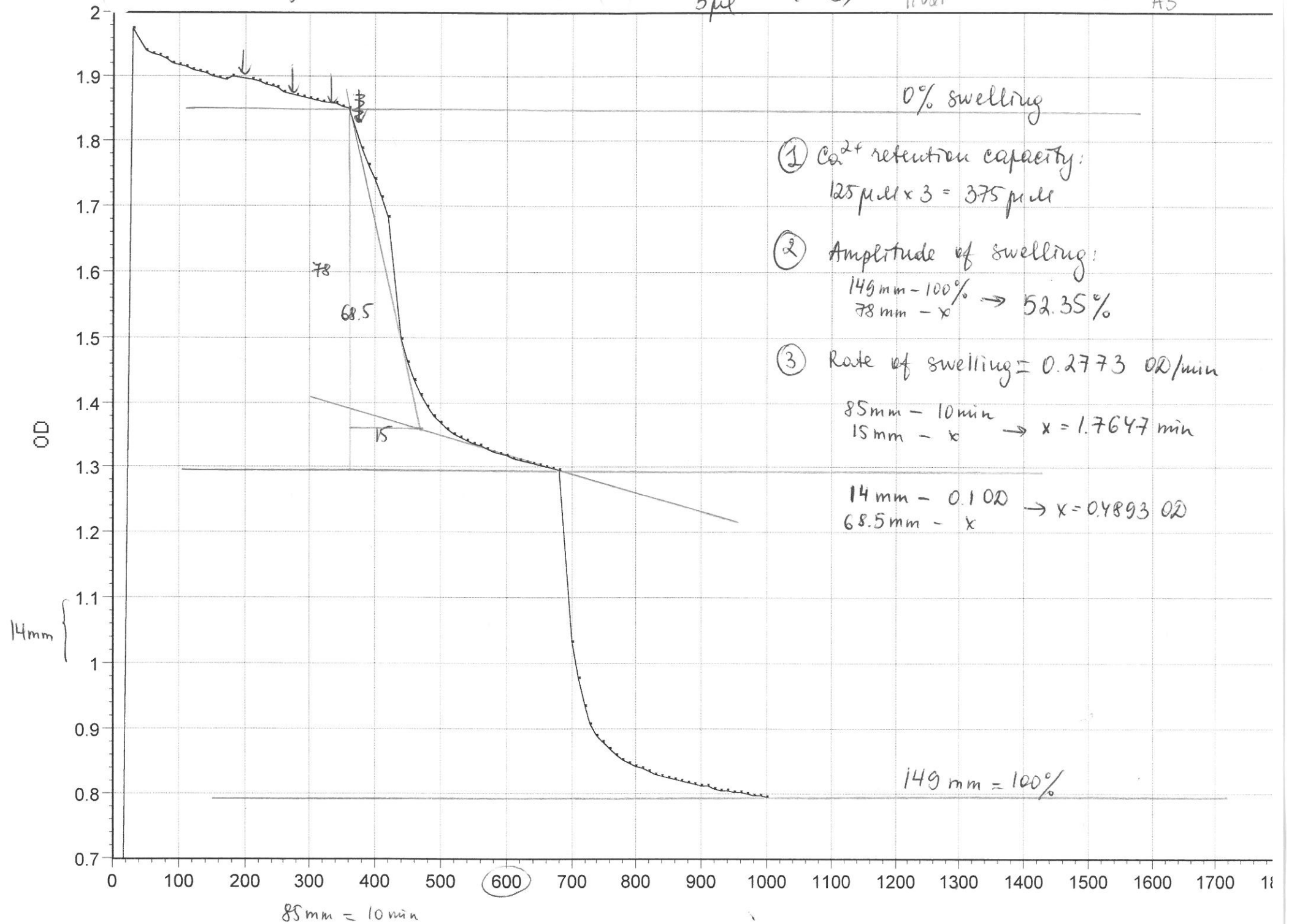
ADP + oligo + FGF + PKCE inhib →

50  $\mu$ mol Co (x3)  
5  $\mu$ l

liver

Feb 8, 2008  
A3

LIVER





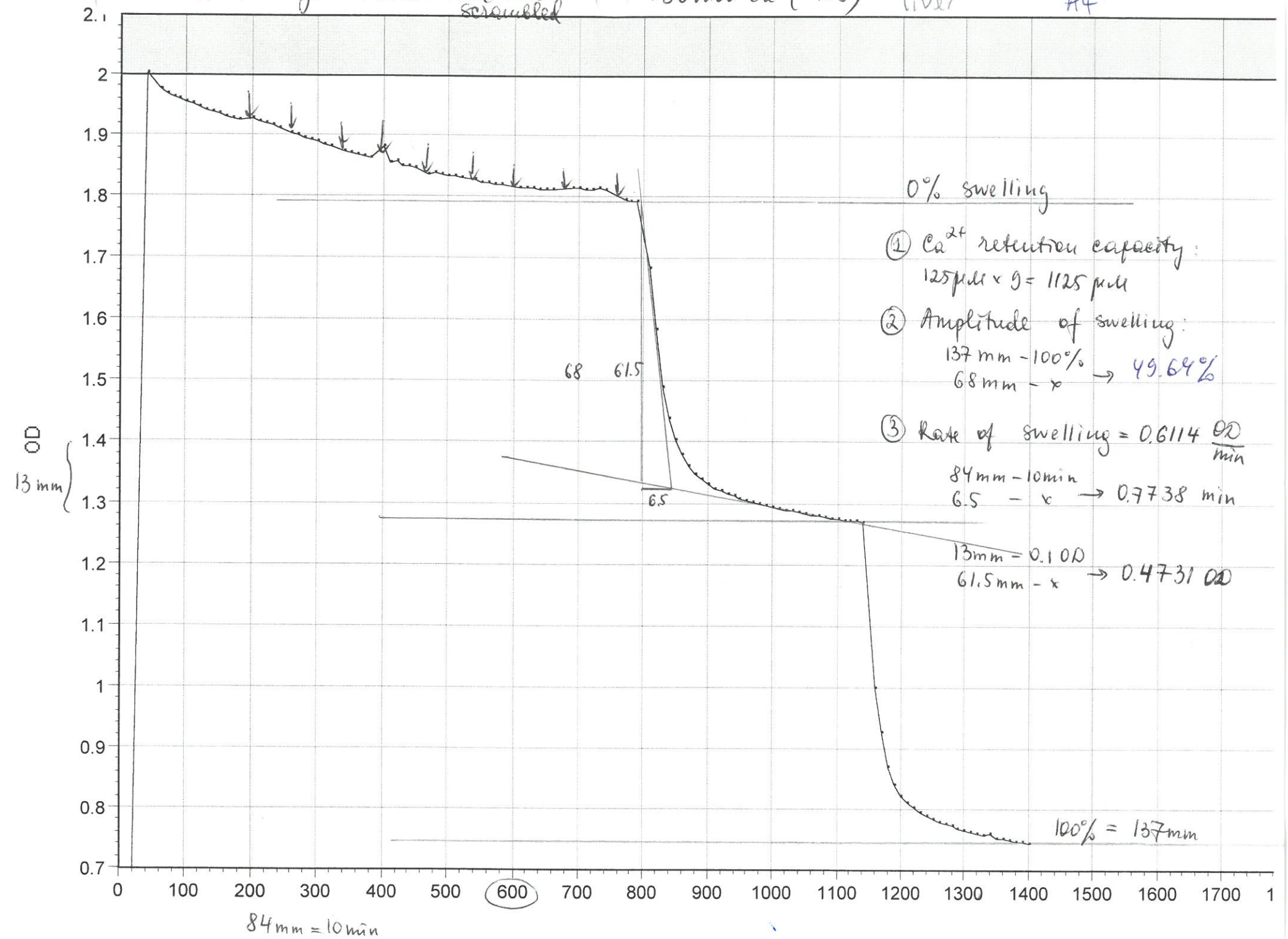
A4

ADP + oligo + FGF2 + PKC $\epsilon$  inh  
scrambled

→ 50 mM Ca (x9) liver

Feb 8, 2008  
A4

LIVER



A5 + A6

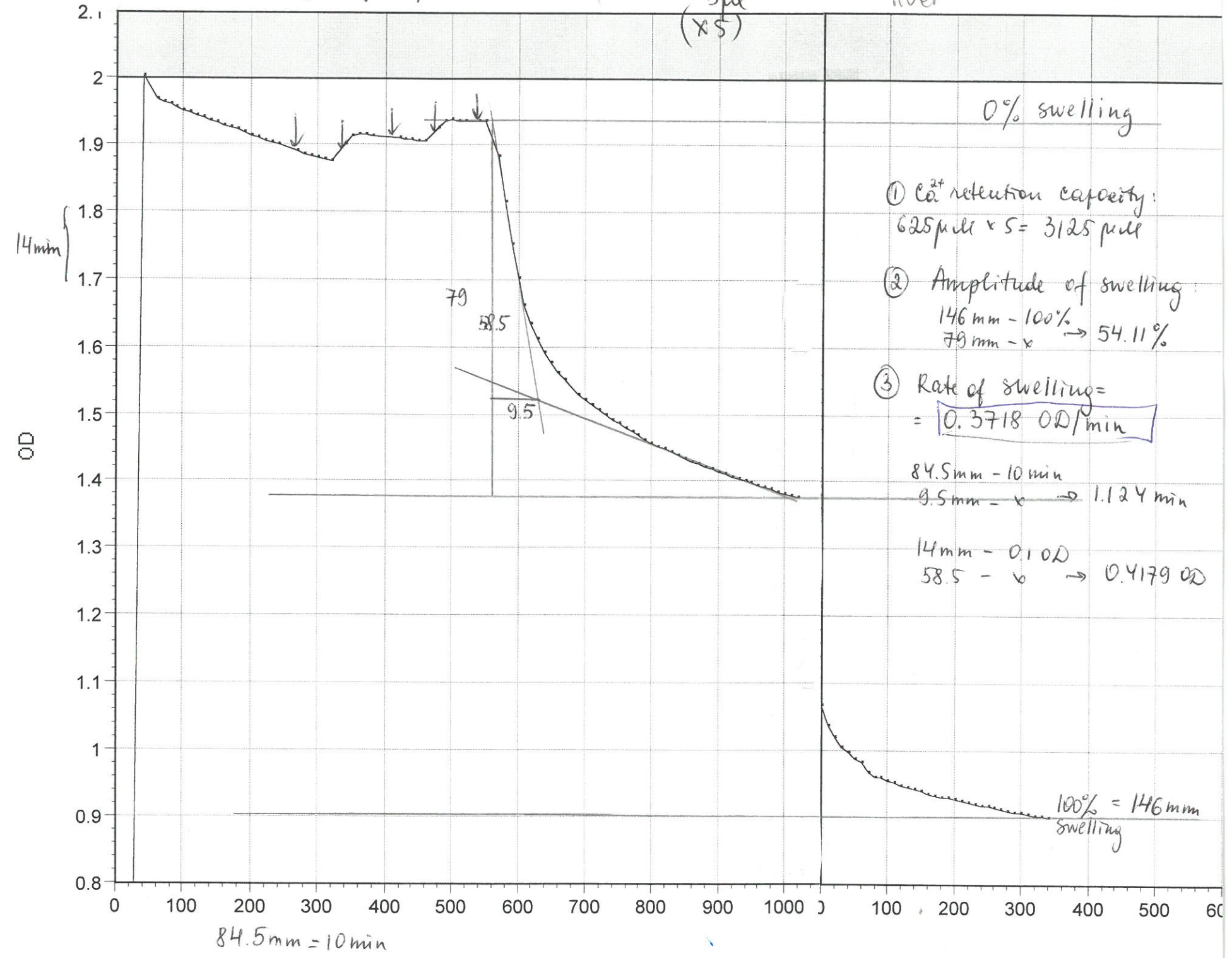
ADP + olig + Cyclosporin A →

→ 250  $\mu$ l  $Ca^{2+}$   
5  $\mu$ l  
(x5)

liver

Feb 8 '08

LIVER



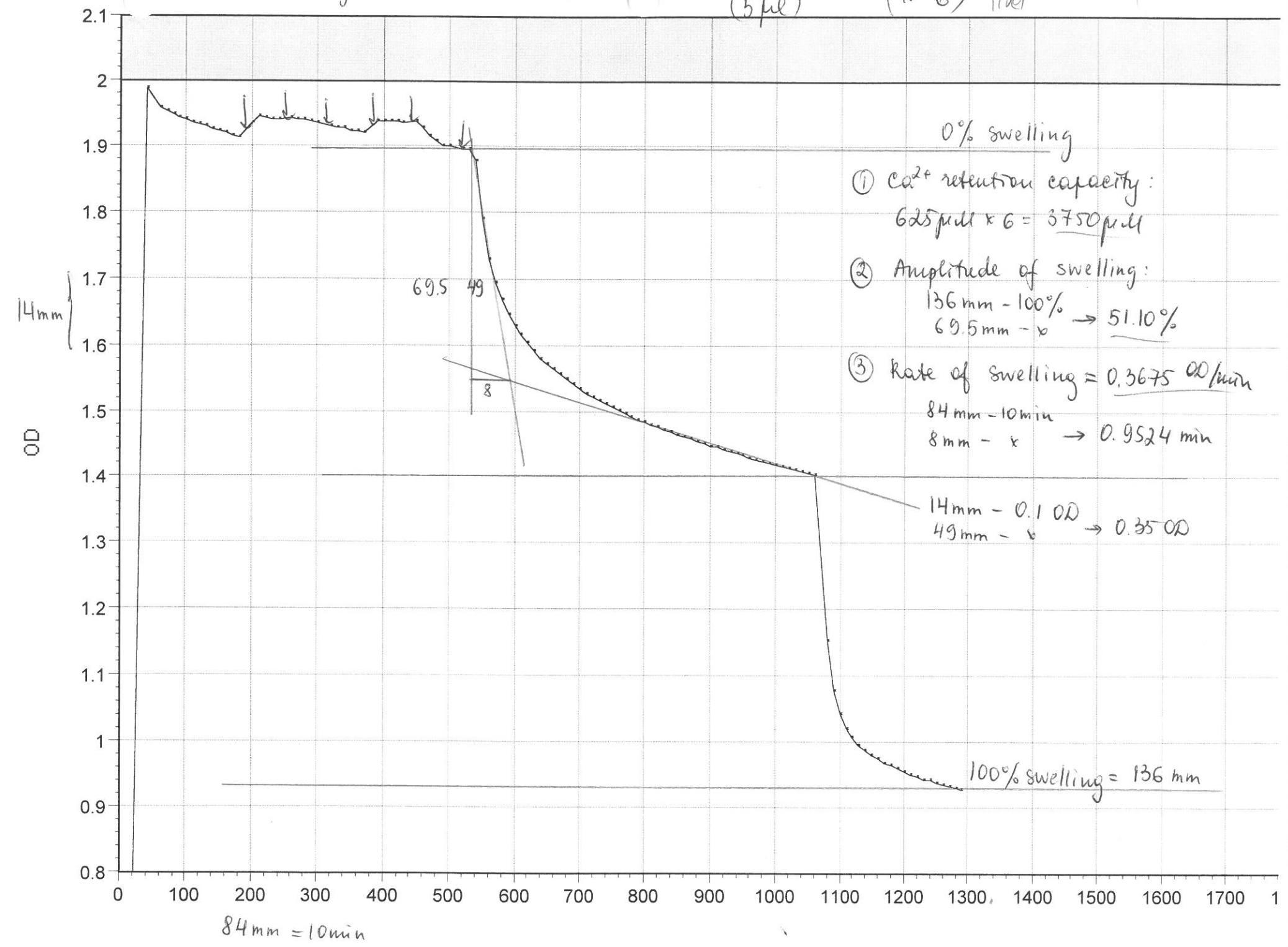
A7

ADP + olig + CsA + FGF2 →

→ 250  $\mu$ l Co (x 6) liver  
(5  $\mu$ l)

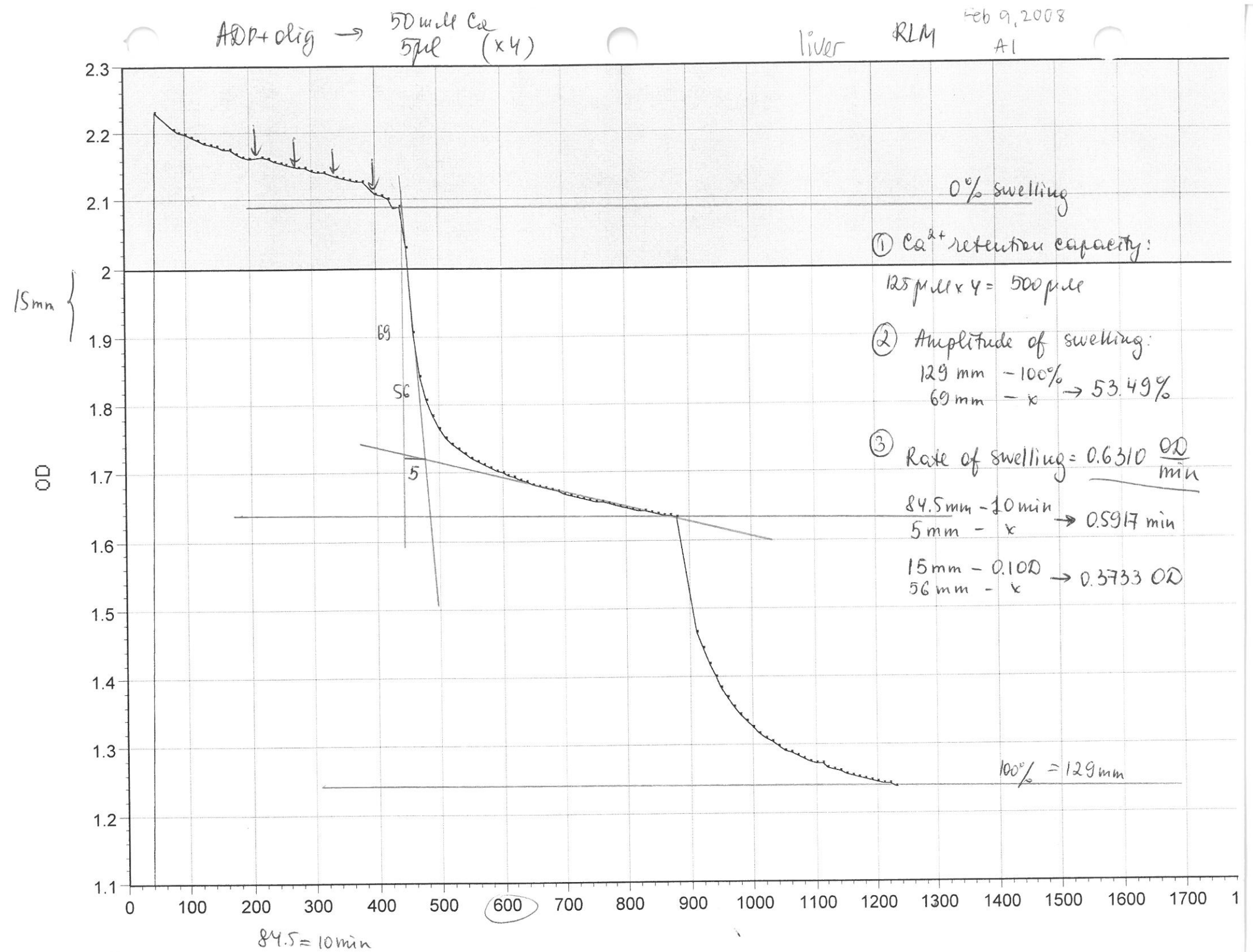
FEB 8, 2008

LIVER

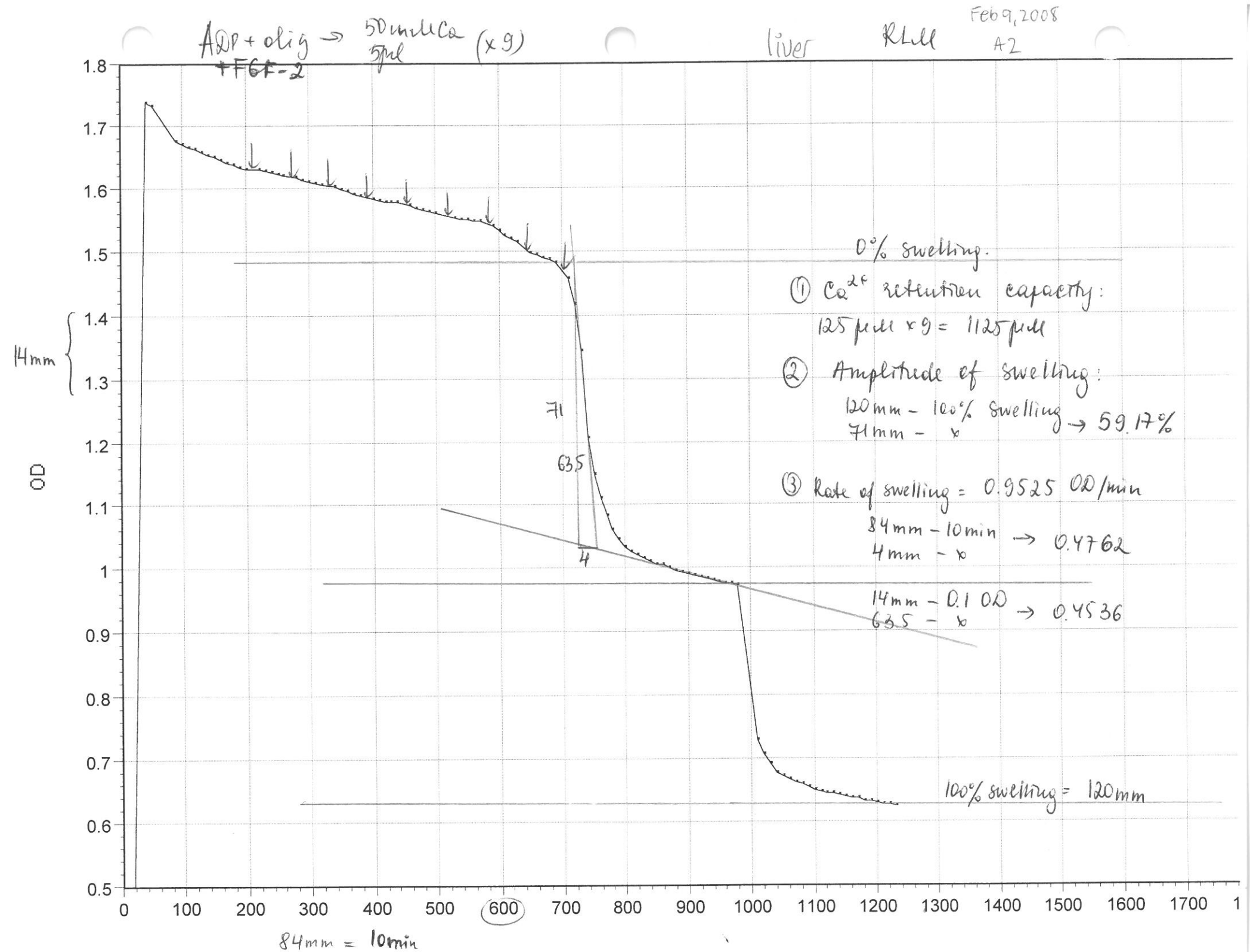




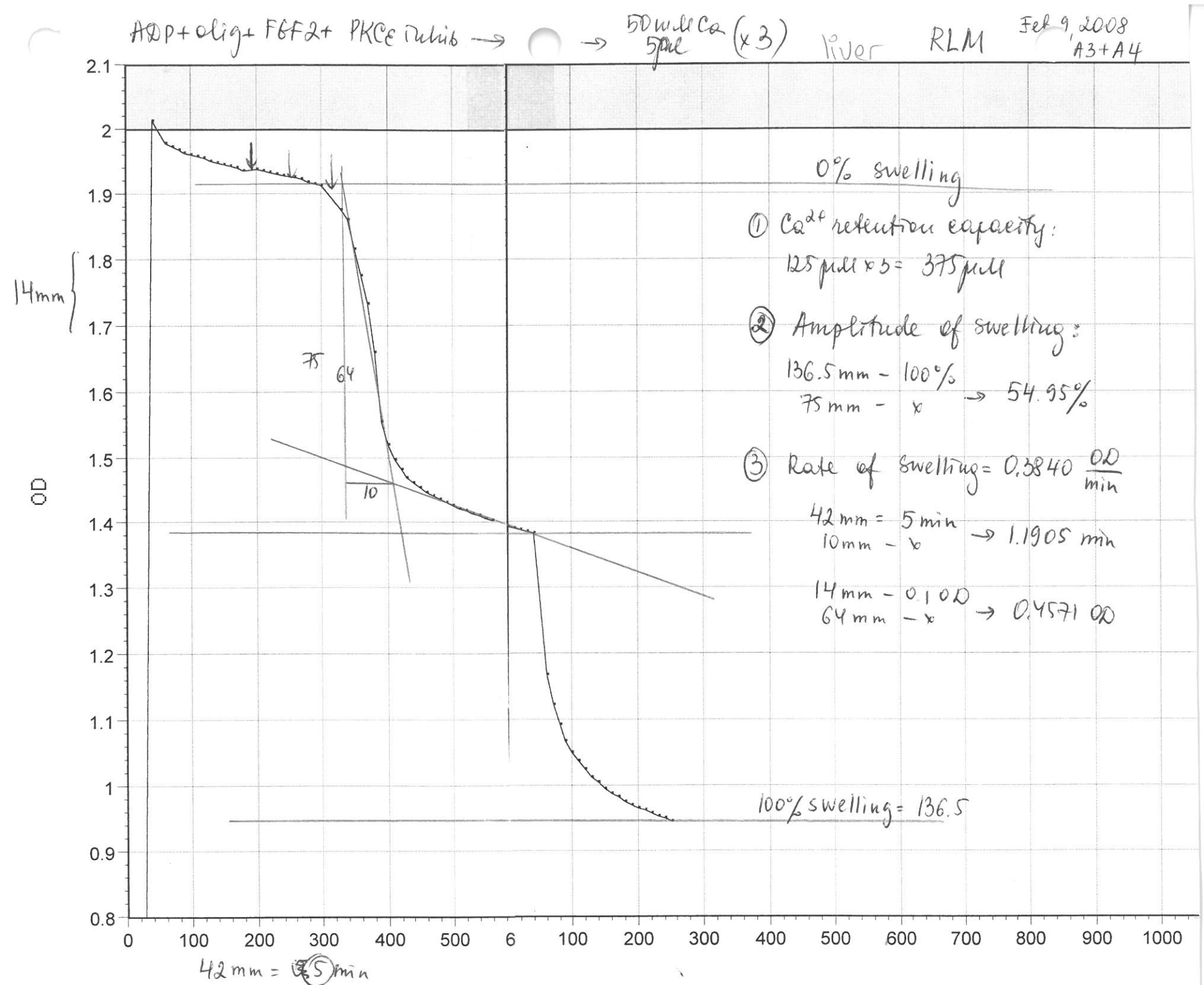
LIVER



LIVER



LIVER





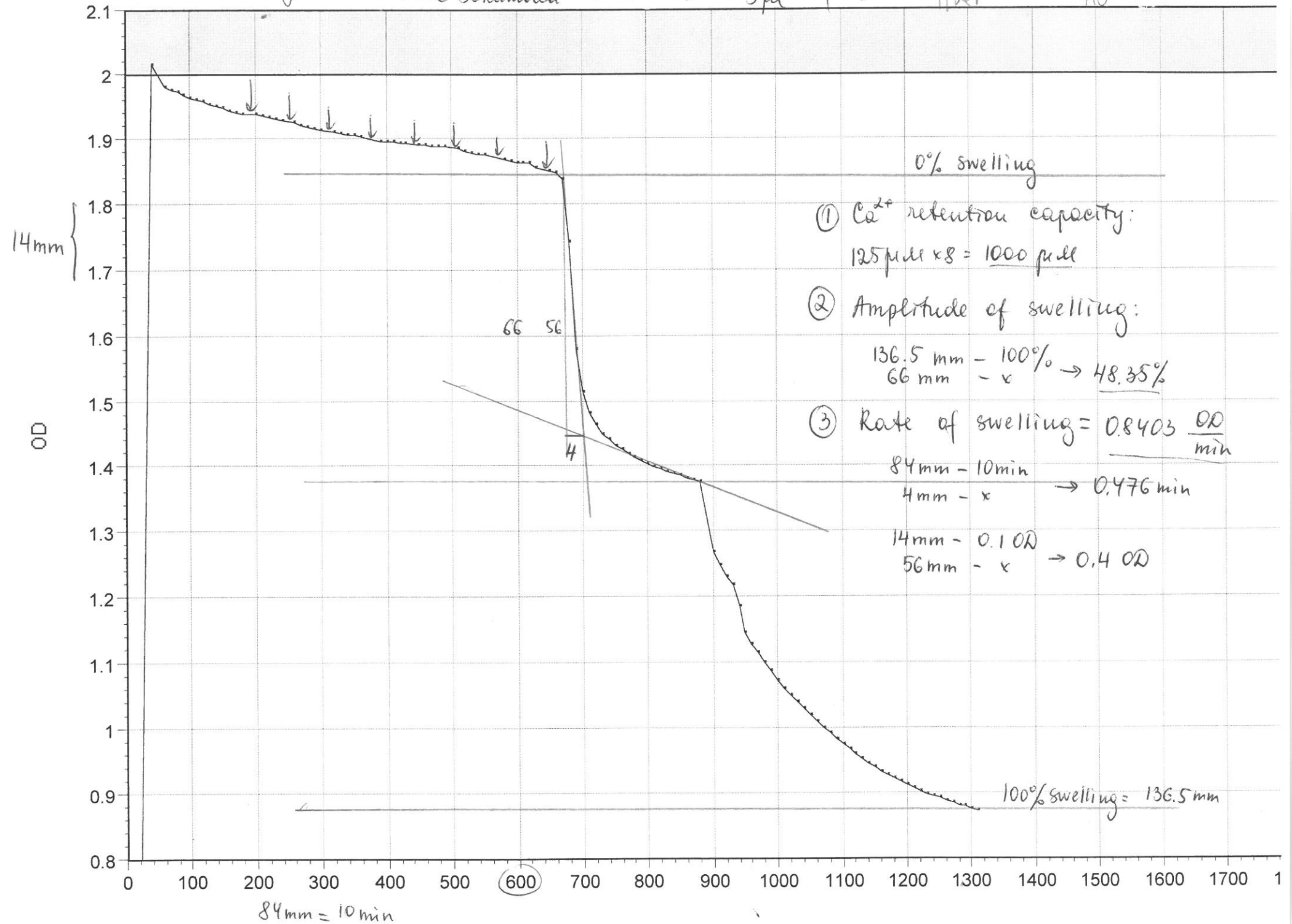
LIVER

ADD + olig + FGF2 + PKCE scrambled →

50  $\mu$ l  $Ca^{2+}$  (x8)  
5  $\mu$ l

liver

Feb 9, 2008  
A5



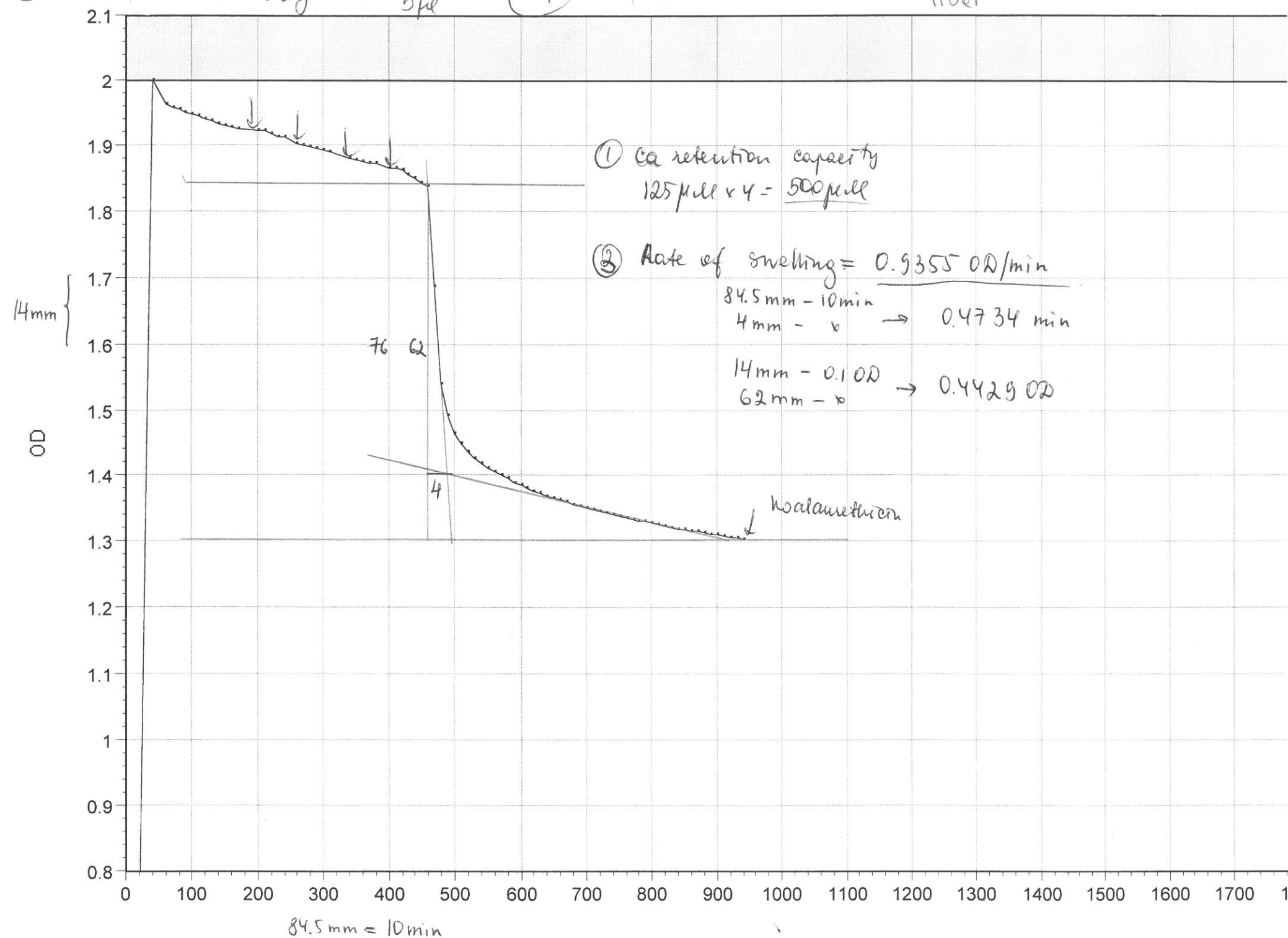
LIVER

(A6)

ADD+ olig → 50 ml C<sub>2</sub> 5 $\mu$ l (x4)

liver

Feb 9 2008



ADP + olig → 50  $\mu$ M Ca (x4)  
5  $\mu$ M

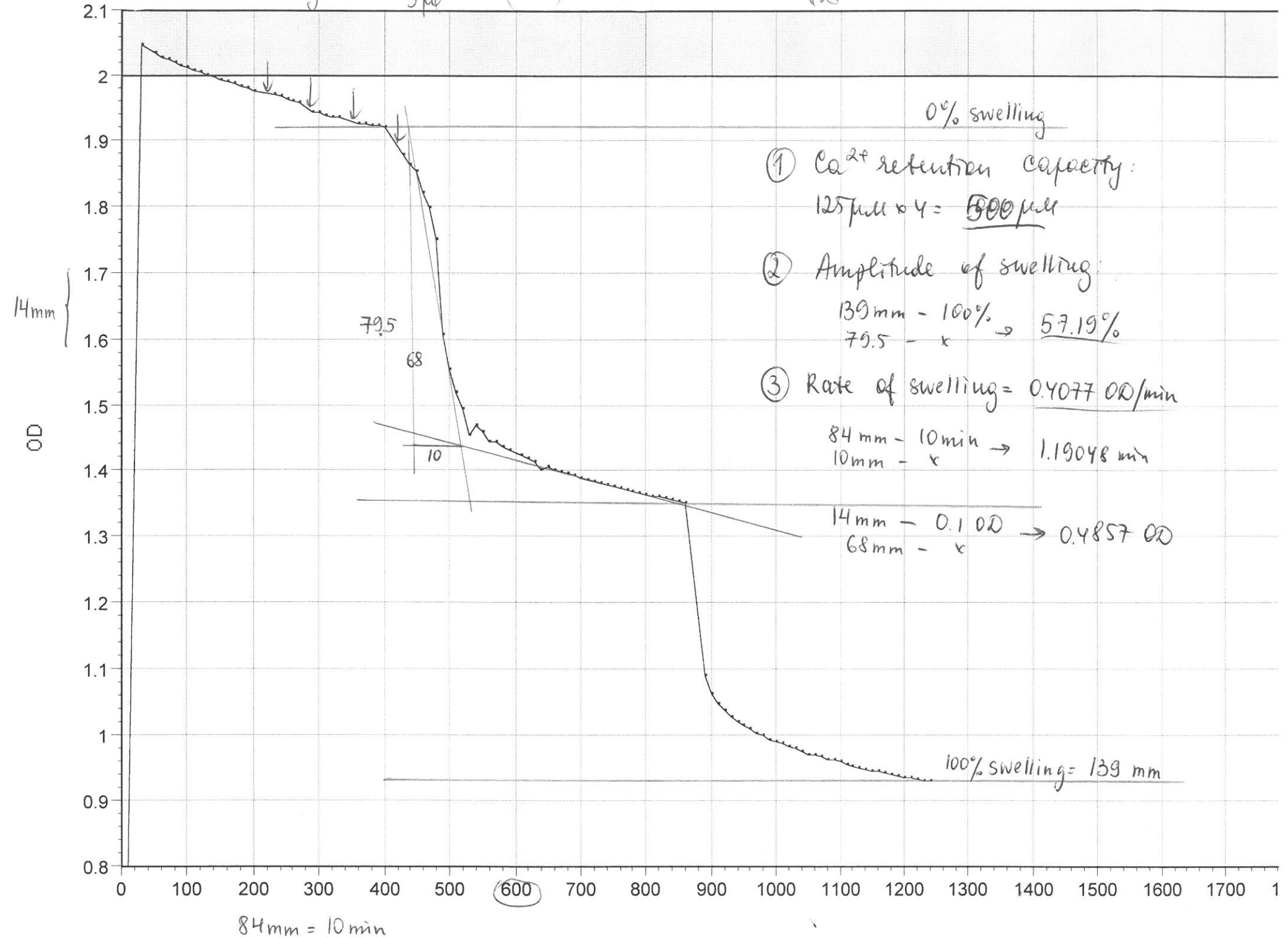
liver

RLM

A7

Feb 9, 2008

LLIVER





Original blots for Fig.3

Fig.3a

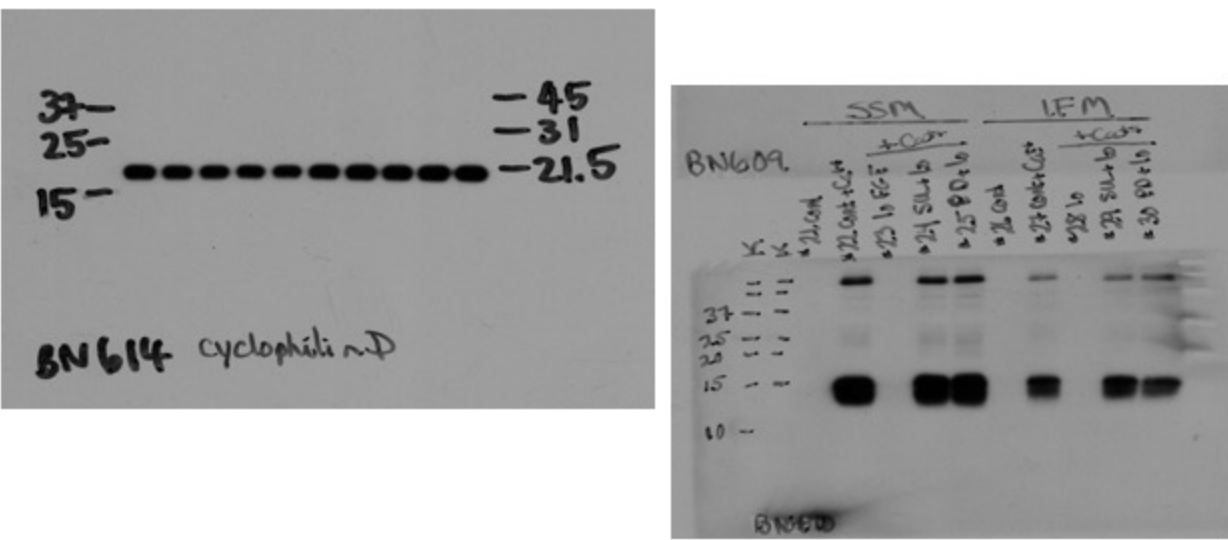
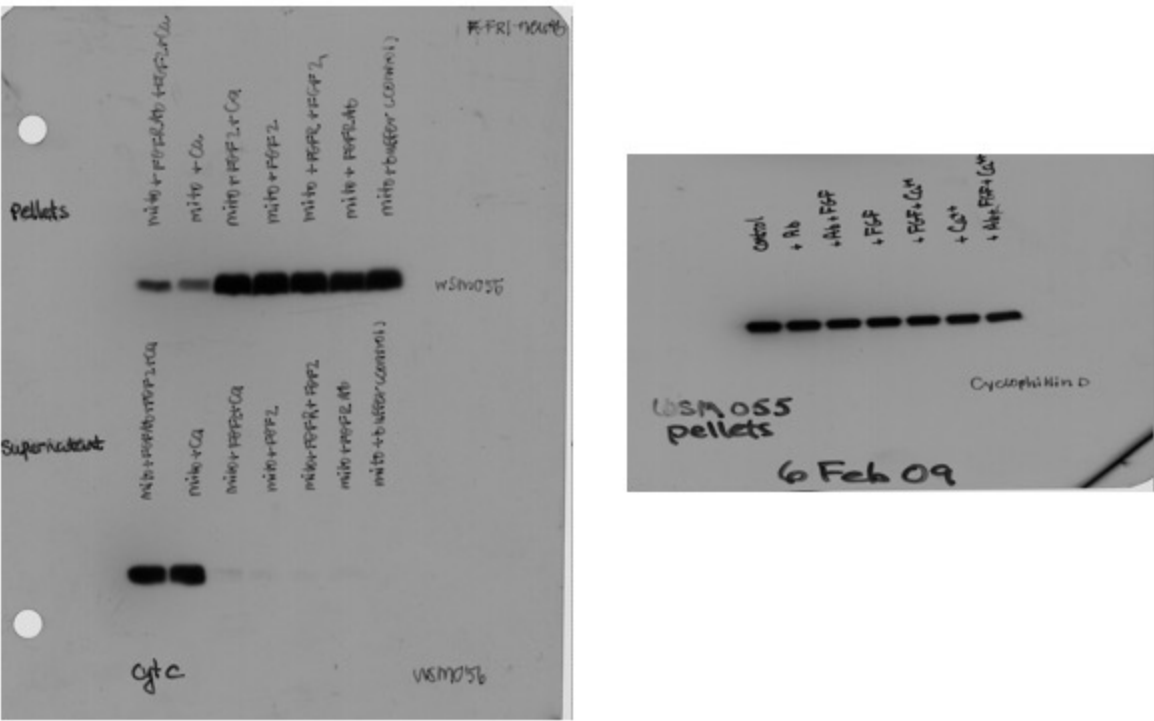
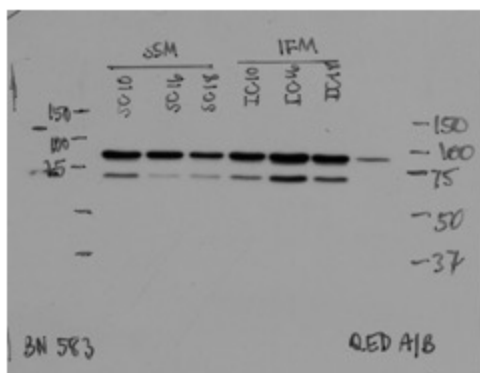


Fig.3b

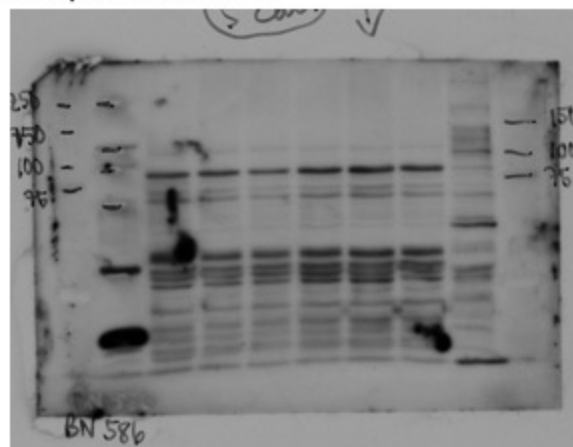


## Original blots for Fig.2a

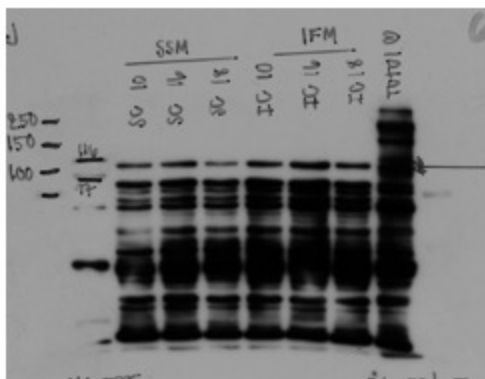
x anti-FGFR1, QED A/B.



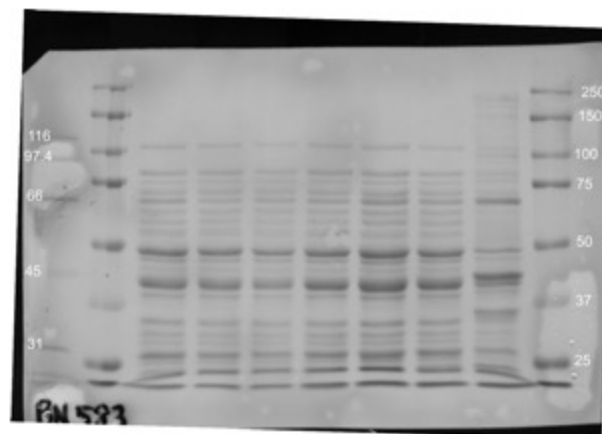
x anti-pY766-FGFR1



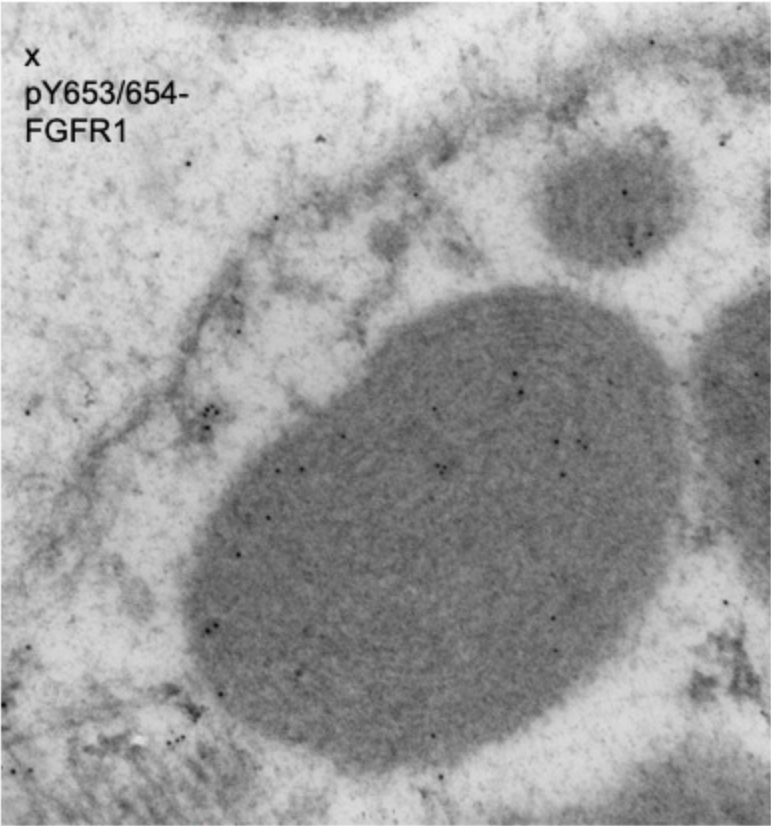
x anti-pY653/654-FGFR1.



Ponceau S stain

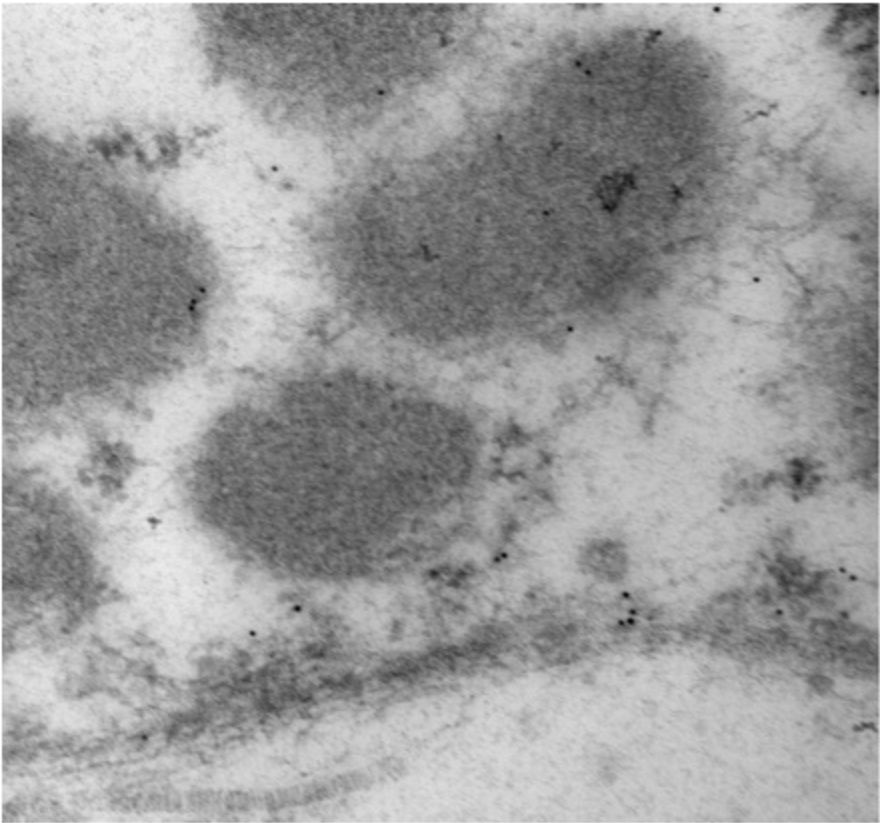


Original  
images  
for  
Fig.2b



*Mislabeled-* heart-pY653-654-FGFR-sigma-2.tif  
HEART SECTION  
Print Mag: 116000x @ 7.0 in  
11:57 10/31/08  
ito

100 nm  
HV=80.0kV  
Direct Mag: 100000x  
Facility of Medicine EM Unit



100 nm  
HV=80.0kV  
Direct Mag: 120000x  
Facility of Medicine EM Unit



Original blots for Fig.4

Fig.4b

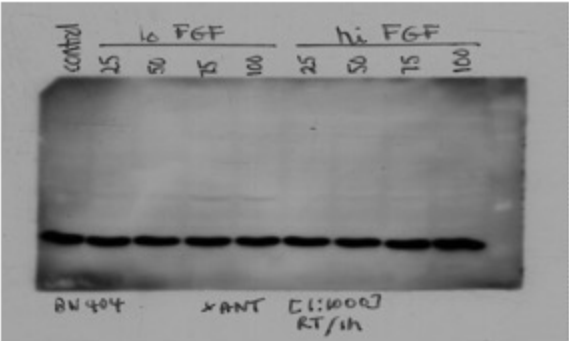


Fig.4a

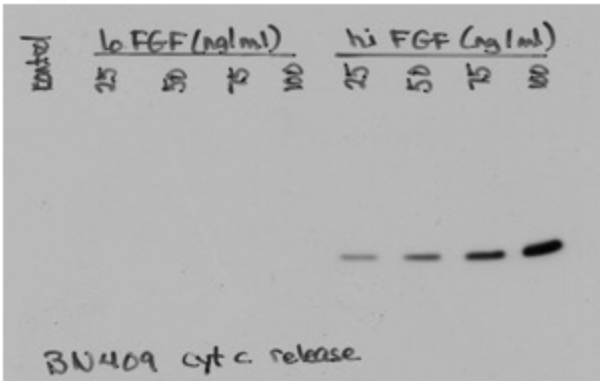


Fig.4c

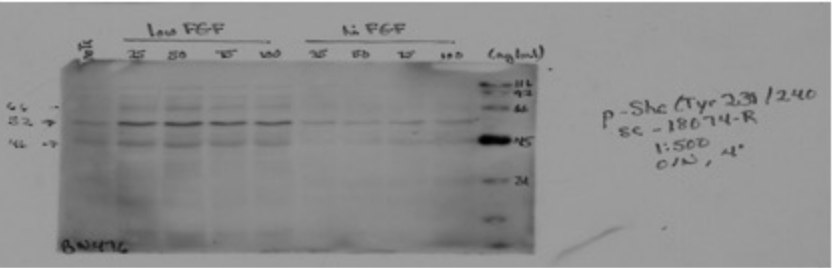


Fig.4d

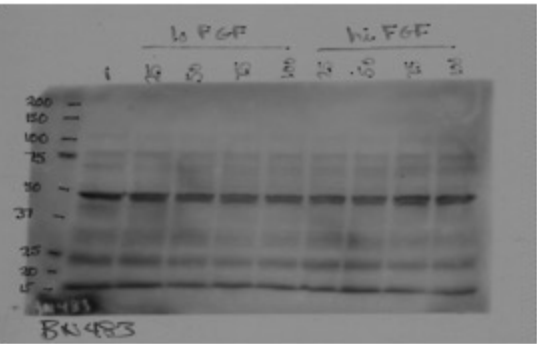
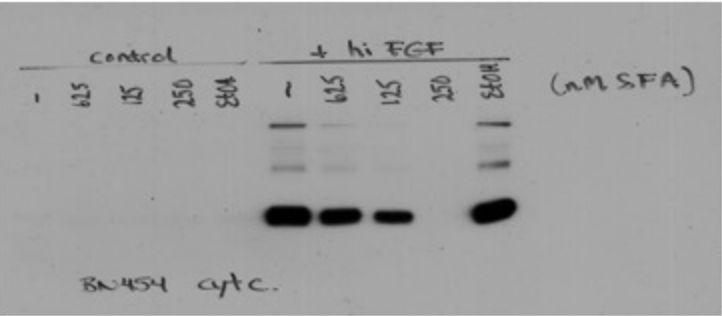


Fig.4e



Original blots for Fig.5

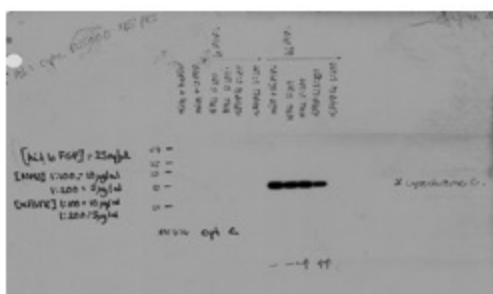


Fig.5a

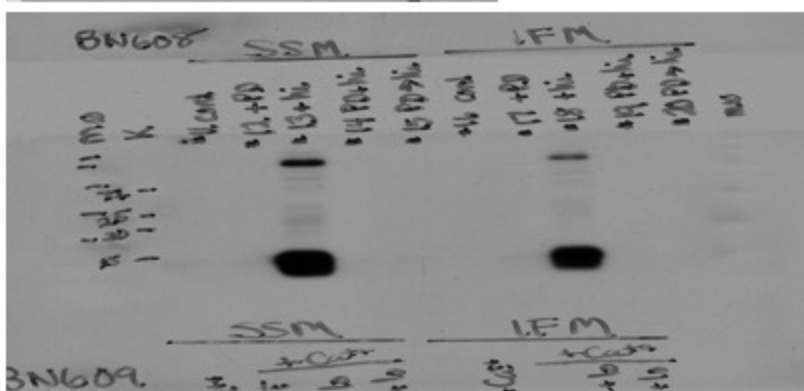


Fig.5b

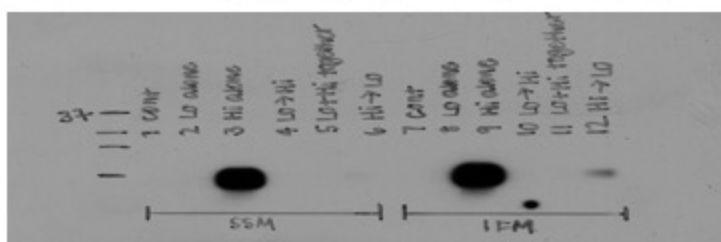
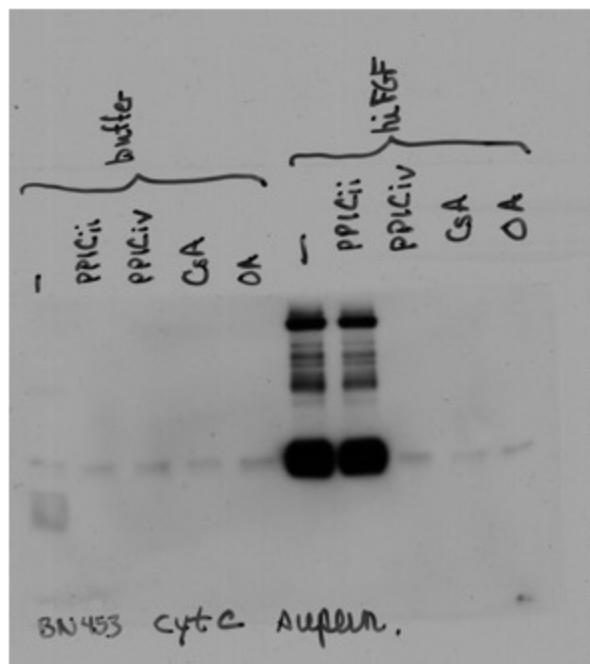
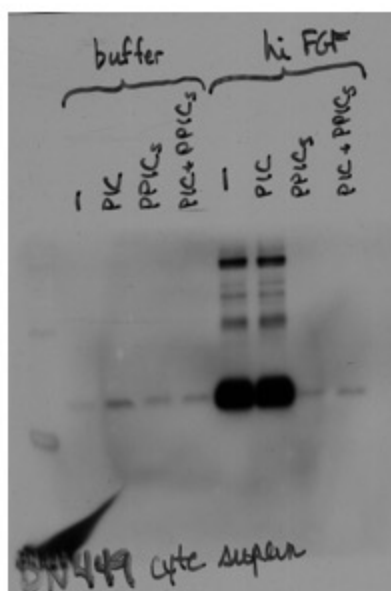
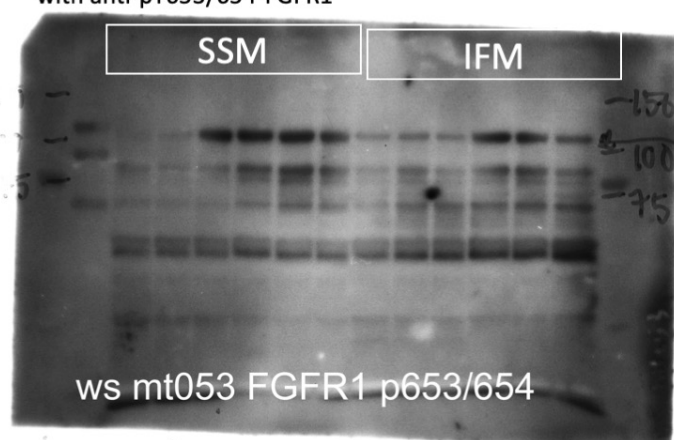


Fig.5c

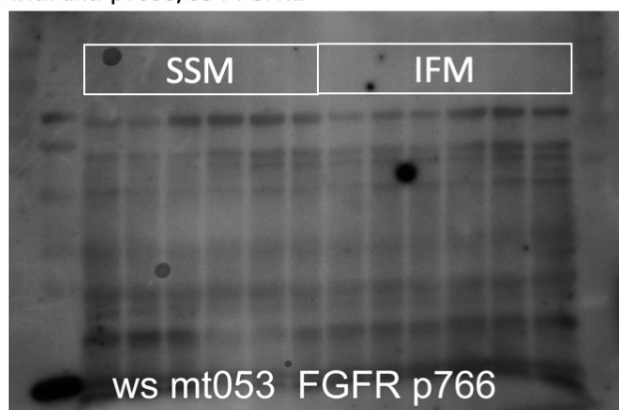
Fig.5e



Repeat experiment: SSM and IFM preparations probed  
with anti-pY653/654-FGFR1

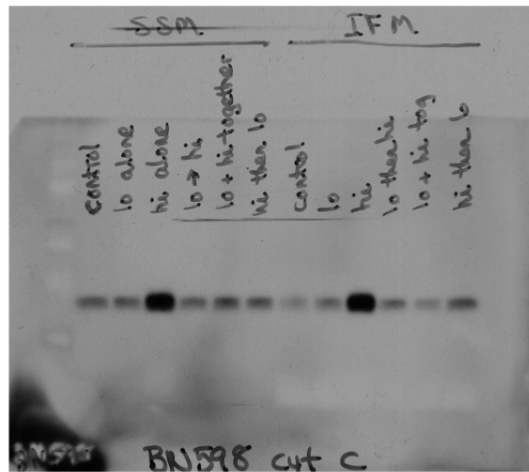
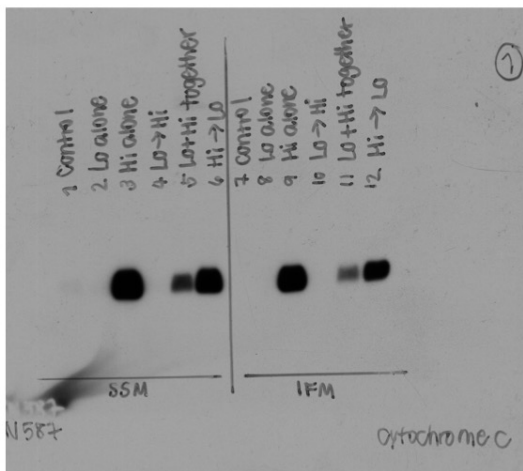


Repeat experiment: SSM and IFM preparations probed  
with anti-pY653/654-FGFR1

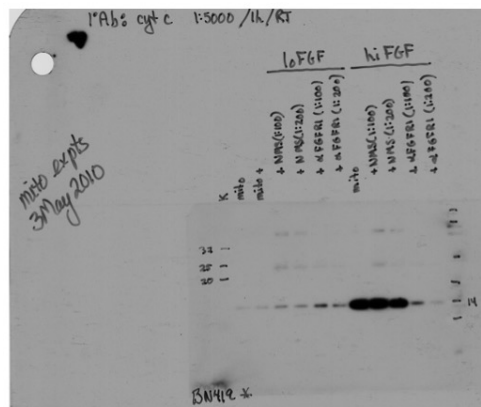
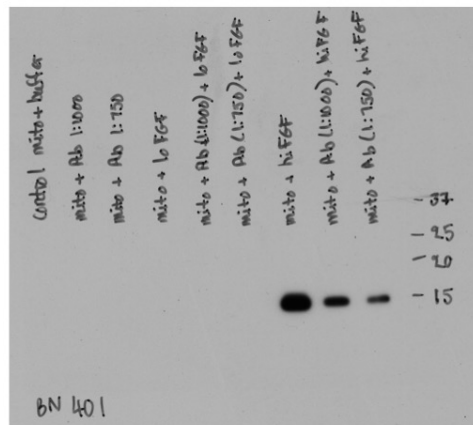
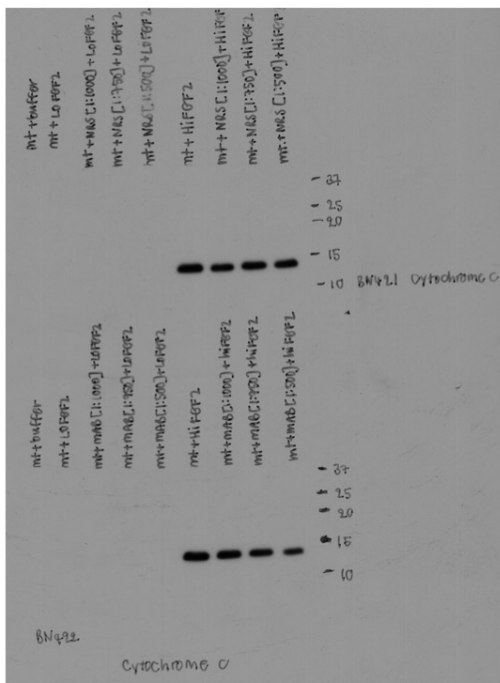




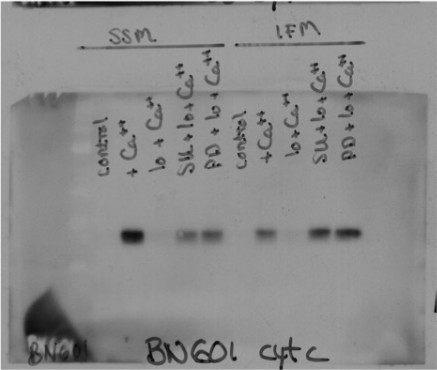
Repeat experiments: Western blots showing supernatant cytochrome c after exposure to Hi-FGF2, and Lo-FGF2



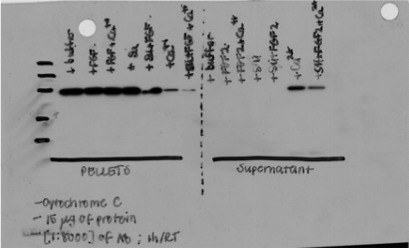
Repeat experiments: Western blots showing supernatant cytochrome c after exposure to Hi-FGF2 in the presence of non-immune antibodies (upper gel) or neutralizing antibodies (lower gel)



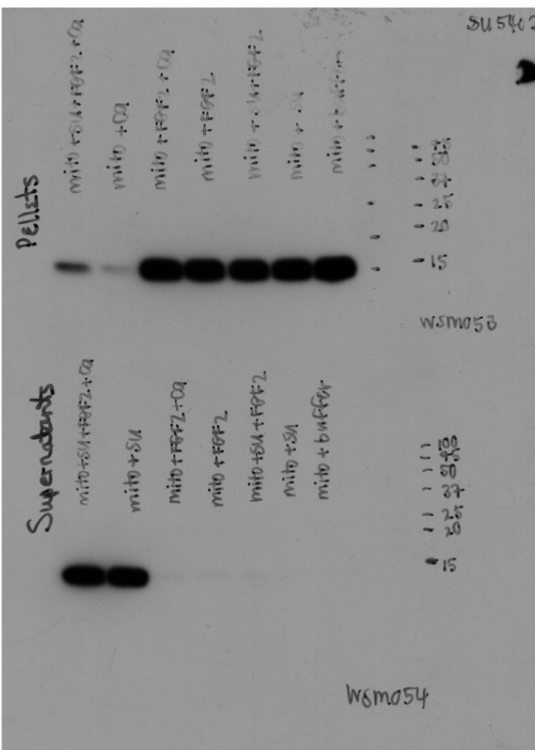
Repeat experiment: SSM and IFM treated with Lo-FGF2 in the presence and absence of FGFR1 inhibitors (SU-, PD-) and subjected to calcium overload: Cyt-C at supernatant



Repeat experiment: SSM treated with Lo-FGF2 in the presence and absence of FGFR1 inhibitor SU-, and subjected to calcium overload: Cyt-C at supernatant and at particulate fraction



Repeat experiment: SSM treated with Lo-FGF2 in the presence and absence of FGFR1 inhibitor SU-, and subjected to calcium overload: Cyt-C at supernatant and at particulate fraction



Repeat experiments: Western blots of SSM and IFM treated with Hi-FGF2 in the presence and absence of FGFR1 inhibitor PD-, SU-: Cyt-C at supernatant

