# **Supplementary Information**

## Title

Depletion of embryonic macrophages leads to a reduction in angiogenesis in the ex ovo chick chorioallantoic membrane assay

#### Authors

Hanna Tay<sup>1,2</sup>, Charis Du Cheyne<sup>1</sup>, Kristel Demeyere<sup>3</sup>, Jurgen De Craene<sup>1</sup>, Lobke De Bels<sup>1</sup>, Evelyne Meyer<sup>3</sup>, Andries Zijlstra<sup>4</sup>, Ward De Spiegelaere<sup>1,2</sup>

### Author affiliations

<sup>1</sup> Department of Morphology, Ghent University, Merelbeke, East-Flanders, Belgium

<sup>2</sup> Cancer Research Institute Ghent (CRIG), Ghent University, Merelbeke, East-Flanders, Belgium

<sup>3</sup> Department of Pharmacology, Toxicology and Biochemistry, Ghent University, Merelbeke, East-Flanders, Belgium

<sup>4</sup> Department of Pathology, Microbiology and Immunology, Vanderbilt University Medical Center, Nashville, Tennessee, USA

#### The supplementary information includes 7 figures and 3 tables



Supplementary Figure 1 The depletive effect on chicken macrophages after injection of 100 or 50  $\mu$ l of clodronate liposomes (right image and density plot) versus 100 or 50  $\mu$ l of PBS liposomes (left image and density plot) is comparable to that after the injection of 25  $\mu$ l of liposomes. The immunohistochemistry images are an example of the decrease in KUL01-positive stained macrophages after injection of 100  $\mu$ l clodronate liposomes, while the density dot-plots show the reduction of PE-KUL01 stained macrophages in the CD45-APC stained leukocyte population after injection of 50  $\mu$ l of clodronate liposomes.



**Supplementary Figure 2** Random placement of a grid on immunohistochemistry images allowed for the quantification of macrophages. The amount of positive cells and number of crossing points of the grid with embryonic tissue was determined. Then, the ratio of positive cells per crossing point was calculated.



**Supplementary Figure 3** Staining with an isotype control (right) shows that the background signal is negligible and that the observed PE-signal was primarily due to the specific binding of the PE-labelled KUL01 antibody to chicken macrophages (left).



**Supplementary Figure 4** Cell viability after sample preparation for flow cytometry varied between 85-90% (biological triplicates). This % viability is calculated as 100 - % PI<sup>+</sup>. PI = propidium iodide.

Experiment	Treatment	Embryos injected	Embryos survived	Survival (%)
	PBS liposomes	8	7	87.50
I	Clodronate liposomes	9	4	44.44
	PBS liposomes	8	6	75.00
2	Clodronate liposomes	9	4	44.44
2	PBS liposomes	8	7	87.50
3	Clodronate liposomes	9	5	55.56
4	PBS liposomes	8	7	87.50
4	Clodronate liposomes	9	5	55.56
5	PBS liposomes	8	6	75.00
	Clodronate liposomes	10	8	80.00
6	PBS	7	6	85.71
	Free clodronate	8	7	87.50
7	PBS	7	7	100
	Free clodronate	8	7	87.50
0	PBS	5	4	80.00
8	Free clodronate	6	4	66.67

Supplementary Table 1 Overview of the embryo survival rate in each CAM angiogenesis experiment.



**Supplementary Figure 5** Collagen/mesh onplants were recuperated for immunohistochemistry in the same fashion as the embryonic samples. During cutting of the collagen/mesh onplants, the samples easily ripped at the site where the nylon mesh was located. However, the rip and thus the area where the mesh had been, could easily be detected. In control embryos, a large amount of KUL01 positive macrophages could be detected in the CAM surrounding the area where the mesh was previously located (left). In clodronate liposome-injected embryos, the number of positive cells was considerably lower (right), confirming that depletion of macrophages was also detected at the level of the collagen/mesh onplants.



**Supplementary Figure 6** Injection of fluorescent (PE) Dil-liposomes and subsequent analysis with flow cytometry confirmed that these PE-positive liposomes are mainly ingested by chicken macrophages (detectable with KUL01-FITC conjugated antibody). The two density- dot plots (unstained - (left panel) and stained with (middle panel) KUL01-FITC) show that the PE-positive subpopulation, which represents the cells that have ingested Dil-liposomes, shifts towards the right after staining with KUL01-FITC, indicating that the majority of liposomes is ingested by macrophages. The PE-negative KUL01-FITC-positive subpopulation depicts the macrophages that have not ingested Dil-liposomes (7-8% of the leukocyte population). The histogram (right panel) shows the FITC signal after gating of all PE (Dil-liposomes)-positive events either unstained (black) or stained with (green) KUL01-FITC conjugated macrophage marker. The peak shifts to the right when the Dil-liposome ingesting population is stained with KUL01-FITC indicating that 91.35 % of these cells are KUL01-FITC positive.

Experiment	Treatment	Embryos injected	Embryos survived	Survival (%)
	PBS liposomes	8	7	87.50
	Clodronate liposomes	9	4	44.44
2	PBS liposomes	8	6	75.00
2	Clodronate liposomes	9	4	44.44
2	PBS liposomes	8	7	87.50
3	Clodronate liposomes	9	5	55.56
4	PBS liposomes	8	7	87.50
4	Clodronate liposomes	9	5	55.56
5	PBS liposomes	8	6	75.00
	Clodronate liposomes	10	8	80.00
6	PBS	7	6	85.71
	Free clodronate	8	7	87.50
7	PBS	7	7	100
	Free clodronate	8	7	87.50
	PBS	5	4	80.00
8	Free clodronate	6	4	66.67

Supplementary Table 2 Overview of the embryo survival rate in each CAM angiogenesis experiment.

**Supplementary Table 3** Overview of the angiogenic response (percentage) in all collagen plugs used in the CAM angiogenesis assay

Experiment	Embryo	Treatment	Onplant	Percentage
			Control	0.00
		PBS	Control	0.00
	1	liposomes	HT1080	14,58
			HT1080	29,14
		PBS liposomes	Control	4,69
	2		HT1080	47,00
			HT1080	23,61
		Clodronate	Control	0,00
	3	linosomes	Control	0,00
			HT1080	0,00
		PBS	Control	10,26
	4		Control	10,39
		iiposomes	HT1080	55,50
			Control	3,30
		Clodronate	Control	3 57
	5	linosomes	HT1080	1.85
			HT1080	0.00
1			Control	0,00
-			Control	0,00
	6	Clodronate	HT1080	4,55
		iiposomes	HT1080	11,90
			HT1080	5,00
			Control	0,00
	7	PBS	Control	0,00
	,	liposomes	HT1080	0,91
			HT1080	0,00
			Control	2,44
	8	Linesomos	LUT1080	0,00
		liposonies	HT1080	2,78
			Control	0.00
		PBS	Control	1.67
	9	liposomes	HT1080	2,60
			HT1080	2,48
	10	PBS	Control	1,59
		liposomes	HT1080	8,33
	11	PBS	Control	50,67
		liposomes	Control	64,81
		nposonies	HT1080	87,50
	12	Cladramata	Control	0,00
		linosomos		0,00
		liposonies	HT1080	0,00
			Control	5.00
		PBS liposomes Clodronate liposomes	Control	1.79
	13		HT1080	10.29
			HT1080	10,94
			Control	2,22
	14		Control	0,00
			HT1080	11,57
		PBS liposomes	Control	8,40
	15		Control	2,27
2	15		HT1080	13,41
Z			HT1080	14,53
	16	Clodronate liposomes	Control	0,00
			LUT1080	1,18
			HT1080	16,71
	17		Control	10,10
		PBS liposomes	Control	5,77
			HT1080	60,71
			HT1080	42,86
	18	Clodronate liposomes	Control	0,00
			Control	48,61
			HT1080	3,13
			HT1080	0,00
	10	PBS	Control	1,41
	13	liposomes	HT1080	0.00
	20		Control	0,00

-				
			Control	0,00
		Clodronate	HT1080	0.00
		liposomes	1111000	0,00
	L		H11080	0,00
			Control	55,56
		DBC	Control	7 95
	21	PD3	CONTION	7,95
		liposomes	HT1080	8,65
			HT1080	1 18
			0	1,10
			Control	0,83
		PBS	Control	7 14
	22	liposomes	Control	7,14
			HT1080	0,57
			HT1080	13 33
			0.1.1	20,00
			Control	7,14
		Clodronate	Control	3.33
	23	linesee	UT1000	C0 75
		liposomes	H11080	68,75
			HT1080	0.00
			Control	14.00
	24		Control	14,00
		PBS	Control	4,17
		linocomec	UT1000	12 10
		ilposomes	H11080	15,10
			HT1080	1,35
		Cladranata	Control	0.00
	25	Clouronate	CONTION	0,00
	20	liposomes	Control	0,00
			Control	16 70
			CONTION	10,78
	26	PBS	Control	10,71
	26	linosomes	HT1080	34 20
		1103011103	1111000	J4,23
			HT1080	63,64
1	[	Clodronate	Control	8 93
	27	ciouronate	0010101	0,00
_		liposomes	HT1080	8,00
2			Control	8 3 3
3	1		0	3,33
	28	PBS	Control	4,44
	20	liposomes	HT1080	28.67
		nposonies	1111000	20,07
			HT1080	10,77
			Control	0.00
			0 1 1	0,00
	20	Clodronate	Control	0,00
	25	liposomes	HT1080	5.13
			1171000	0.22
			H11080	8,33
			Control	2,27
	20	PBS	Control	1 20
	30	linosomes	Control	1,30
		nposonies	HT1080	5,36
		Clodronate	Control	0.00
	31	CONTION	0,00	
	_	liposomes	HT1080	0,00
			Control	0.00
			0 1 1	0,00
	32	PBS liposomes	Control	0,00
			HT1080	6.92
			UT1090	0,00
			H11000	0,00
			Control	3,57
	33	PBS liposomes	Control	21.68
			CONTION	21,00
			HT1080	23,81
			HT1080	44.32
	1			
		PBS liposomes	Control	4,55
1	Ι.		Control	6,49
	34		HT1090	10 70
			1111000	15,70
			HT1080	0,00
		Clodronate	Control	0.00
			C+- !	0.00
	35		Control	9,26
	35	liposomes	HT1080	0,00
			HT1020	10 71
			H11000	10,71
		PBS	Control	9,38
	36		Control	14.06
		liposomes	CONTION	14,00
			HT1080	13,75
	37		Control	1.00
		Clarker	Control	0.00
		Clodronate liposomes	CONTROL	0,00
- 1			HT1080	0,00
<u>л</u>			HT1080	2.86
-+	H			2,00
	1		Control	2,60
		PBS	Control	1.92
	38	liposomes	UT1000	6 70
			000110	0,/ð
			HT1080	6,92
		T I	Control	0.00
			0	0,00
	20	Clodronate	Control	0,00
	39 lip	liposomes	HT1080	1.12
			UT1000	0.00
		┥───┤	000110	0,00
			Control	0,00
		DDC	Control	0.00
	40	FD3	Control	0,00
	1	liposomes	HT1080	0,00
		1 1	HT1080	0.00
				0,00
	Δ1	PBS	Control	13,59

1				
			HT1080	47,25
			HT1080	35,37
		Clodronate	Control	7,94
	42	linosomes	HT1080	5 56
		liposonies	Control	1 65
		555	Control	1,05
	43	PBS	Control	12,34
	-	liposomes	HT1080	23,08
			HT1080	9,52
			Control	12,00
		PBS	Control	4,76
	44	linosomes	HT1080	19.15
			HT1080	25.76
			Control	23,70
		Clodronate	Control	0,00
	45		Control	1,01
		liposomes	HT1080	5,30
			HT1080	11,11
			Control	6,94
		PBS	Control	2,20
	46	liposomes	HT1080	33.68
			HT1080	0.00
			Control	0,00
	1		Control	0,00
	47	Cloaronate	Control	7,19
	1	liposomes	HT1080	16,67
	L	I	HT1080	23,81
			Control	12,99
		PBS	Control	3,79
	48	liposomes	HT1080	8.97
			HT1080	69.44
			Control	10.00
			Control	10,00
	49	Clodronate liposomes	Control	3,33
			HT1080	2,02
			HT1080	14,88
			Control	1,79
	50	Clodronate liposomes	Control	3,33
			HT1080	0,56
			HT1080	18.18
		PBS liposomes	Control	1.67
			Control	2,07
_	51			22,75
5			H11080	17,78
			HT1080	47,44
		Clodronate liposomes	Control	0,00
	52		Control	6,25
	52		HT1080	12,22
			HT1080	8,33
I		PBS	Control	8,33
I	53	liposomes	HT1080	30.61
	<u> </u>	nposonies	Control	3 52
		Clodronate	Control	0.00
	54		LUT1000	0,00
	1	liposomes	H11080	3,33
			HT1080	1,39
			Control	1,67
		PBS	Control	1,67
	55	liposomes	HT1080	38,89
	1		HT1080	10,00
		Clodronate	Control	12.12
			Control	1 79
	56		HT1020	25.10
		iiposoilles	UT1000	4.00
			080110	4,90
			Control	9,40
	57	PBS	Control	12,86
	57	PBS liposomes	Control HT1080	12,86 3,90
	57	PBS liposomes	Control HT1080 HT1080	12,86 3,90 8,33
	57	PBS liposomes	Control HT1080 HT1080 Control	12,86 3,90 8,33 20,00
	57	PBS liposomes Clodronate	Control HT1080 HT1080 Control Control	12,86 3,90 8,33 20,00 4,76
	57	PBS liposomes Clodronate	Control HT1080 HT1080 Control Control HT1080	12,86 3,90 8,33 20,00 4,76 0,83
	57	PBS liposomes Clodronate liposomes	Control HT1080 Control Control HT1080 HT1080	12,86 3,90 8,33 20,00 4,76 0,83 27,08



**Supplementary Figure 7** Injection with free clodronate did not have a significant effect on angiogenesis compared to injection with PBS. Statistical analysis using the Wilcoxon signed-rank test revealed that no significant difference could be observed in the amount of ingrowing capillaries in HT1080 plugs (P = 0.97) as well as in plain collagen plugs (P = 0.16). Data are presented as boxplots showing the median, first and third quartiles.