



Article Effect of Organic and Inorganic Fertilizer on the Growth and Yield Components of Traditional and Improved Rice (Oryza sativa L.) Genotypes in Malaysia

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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Abstract: Rice is the most important staple cereal human nutrition and consumed by 75% of the global population. Rice plants need a supply of essential nutrients for their optimal growth. Rice production has increased tremendously in Malaysia insensitive irrigation and the use of inorganic fertilizers and pesticides. However, the effect of using inorganic fertilizers resulted in contamination of ground water and decreased the productivity of soil, which in turn affected the rice production in the long term. The use of organic manure may help to regain the soil health, but that is insufficient for providing the essential nutrients to achieve optimal growth. Therefore, the use of organic manure combined with inorganic fertilizers is applied to obtain optimum yields. This study aims to test the effect of organic and inorganic fertilizers on the growth and yield components of 65 rice genotypes. The pot experiment was conducted at the net house on field 10, University Putra Malaysia, UPM, Malaysia, during the period of February to June 2019 and August to December 2019 in a randomized complete block design (RCBD) with three replications. There were three treatment combinations viz. T_1 : 5 t ha⁻¹ chicken manure (CM), T_2 : 2.5 t ha⁻¹ CM + 50% CFRR, T_3 : 100% (150 N: 60 P₂O₅: $60 \text{ K}_2\text{O} \text{ kg ha}^{-1}$) and chemical fertilizer recommended rate (CFRR). Grain and straw samples were collected for chemical analysis, and physical parameters were measured at the harvest stage. Results showed that most of the growth and yield components were significantly influenced due to the application of organic manure with chemical fertilizer. The application of chemical fertilizer alone or in combination with organic manure resulted in a significant increase in growth, yield component traits, and nutrient content (N, P, and K) of all rice genotypes. Treatment of 2.5 t ha⁻¹ CM + 50% CFRR as well as 100% CFRR showed a better performance than the other treatments. It was observed that the yield of rice genotypes can be increased substantially with the judicious application of organic manure with chemical fertilizer. The benefits of the mixed fertilization (organic + inorganic) were not only the crop yields but also the promotion of soil health, the reduction of chemical fertilizer input, etc.

Keywords: organic manure; crop productivity; rice; soil health; nutrient content

1. Introduction

Rice (*Oryza sativa* L.) is a widely farmed food crop that provides sustenance for more than half of the world's population. "Rice is life" is the most appropriate slogan for the world, as this grain is critical to our national food security and provides a source of income

for millions of rural people. Despite the enormous area under rice production, production is low due to a number of interrelated problems. One of the main causes of low production is an imbalance in fertilizer use, and the continued use of inorganic fertilizers has resulted in a decline in soil fertility. The use of inorganic fertilizers in conjunction with organic resources resulted in the highest grain and straw yields [1]. Using organic manure and chemical fertilizers in tandem would be very promising, not just in terms of increasing output stability but also in terms of improving soil fertility [2].

Organic manuring is becoming an increasingly significant part of environmentally healthy, long-term farming. Plant nutrients are replenished in agricultural soils primarily through inorganic, organic, and biofertilizers [3]. Inorganic fertilizers are used indefinitely, causing a decline in soil chemical, physical, and biological qualities, as well as soil health [4]. Chemical fertilizer's negative effects, combined with rising prices, have sparked a surge in interest in organic fertilizers as a nutritional source [4,5]. For sustainable agricultural production, the use of organic resources as fertilizers has obtained plenty of attention [6,7]. Organic materials have a plenty of potential as a source of numerous nutrients and as a method for improving soil properties [8].

However, due to the comparatively low nutrient content of organic manures, they may not be enough to meet the plant's needs. The application of organic manure with chemical fertilizer increases microbial activity, nutrient usage efficiency, and the availability of native nutrients to plants, resulting in increased nutrient uptake [9]. To obtain optimal yields, it is vital to employ organic manures in conjunction with inorganic fertilizers to provide the soil with all of the plant nutrients in easily available form and to maintain good soil health [10]. With a combination of safe modern technologies and traditional organic agriculture, which is not in its orthodox form, it has the potential to be approved for increased yields.

The use of organic manure in conjunction with chemical fertilizers has the potential to improve soil fertility and crop output. Integrated plant nutrition systems, particularly using organic manure, could improve crop productivity in intensive cropping systems. Organic manure has lately been discovered to be an excellent source of plant nutrients in the soil. Farmyard manure (FYM) and inorganic N and P fertilizers were used together to improve chemical and physical qualities, which could lead to increased and sustainable rice production [11]. Organic manure can provide a good amount of plant nutrients, which can help increase rice yield. As a result, in order to achieve a sustainable crop yield without depleting soil fertility, it is necessary to fertilize and manure in a coordinated manner.

Most cultivated soils in the world have less than 1.5% organic matter, although a good agricultural soil should have at least 2% organic matter. The use of organic manure in conjunction with chemical fertilizers can significantly boost rice output and soil productivity [12]. In a rice–rice cropping pattern, the integrated use of chemical and organic manure is critical for long-term crop productivity and soil fertility [2]. Soil organic matter boosts crop output by improving the physicochemical characteristics of the soil. Organic waste, farmyard manure, compost, and chicken manure have recently received attention as the most effective techniques for boosting soil fertility and thus crop output. Higher crop production necessitates a well-balanced mix of organic and inorganic fertilizer sources.

Inorganic fertilizers were employed with little or no addition of organic manure to generate increased rice yields. Even while inorganic fertilizers resulted in increased agricultural yields, the overuse of them was linked to deteriorated soil characteristics and degraded soils, resulting in lower yields in the future [13]. Chemical fertilizers, growth regulators, and pesticides are completely reliant on chemical fertilizers, growth regulators, and pesticides in the Western world to boost crop yield. Chemical fertilizer use has been linked to a number of negative health and environmental consequences [14]. Taking these factors into account, a middle ground between organic and inorganic fertilizer use for rice cultivation is necessary.

Chicken manure has been considered to be a soil additive to reduce the use of mineral fertilizers because it provides required nutrient amounts, increases cation-exchange capac-

ity, and improves water-holding capacity. Chicken manure not only increases the yield of rice but can also substitute chemical fertilizers to some extent.

However, the use of organic manure alone might not meet the plant requirement due to presence of a relatively low content of nutrients. The application of organic manure with chemical fertilizer accelerates the microbial activity, increases nutrient use efficiency, and enhances the availability of the native nutrients to the plants, resulting in a higher nutrient uptake. Therefore, in order to make the soil well supplied with all the plant nutrients in the readily available form and to maintain good soil health, it is necessary to use organic manure in combination with inorganic fertilizers to obtain optimum yields.

Therefore, the present research work was undertaken to investigate the effect of the combined application of organic and inorganic fertilizers on the growth and yield of traditional and improved rice genotypes.

2. Materials and Methods

2.1. Plant Material

A total of 64 traditional and improved rice cultivars were evaluated in this study. The cultivars were obtained from different sources. The genotypes names and origin are presented in Table 1. First, the seeds were dried under sunlight for 8 h before soaking in a Petri dish and being placed in a dark incubator for 2 days. After that, the germinated seeds were sown in the seed tray.

Table 1. Name, origin, grain size, shape, and status of sample of 64 traditional and improved rice genotypes.

Code No	Name of Genotype	Source Country	Grain Size and Shape	Status of Sample
G1	Pukhi	Bangladesh	MS	Traditional cultivar
G2	Panbira	Bangladesh	SB	Traditional cultivar
G3	Dharial	Bangladesh	MB	Traditional cultivar
G4	Utri	Bangladesh	MS	Traditional cultivar
G5	Luanga	Bangladesh	MS	Traditional cultivar
G6	Kaisa panja	Bangladesh	MS	Traditional cultivar
G7	Vanđana	Bangladesh	MS	Traditional cultivar
G8	Dular	Bangladesh	MB	Traditional cultivar
G9	Sondhamoni	Bangladesh	MB	Traditional cultivar
G10	Hasikamli	Bangladesh	MS	Traditional cultivar
G11	Dumai	Bangladesh	MS	Traditional cultivar
G12	Parija	Bangladesh	MS	Traditional cultivar
G13	Kataktara	Bangladesh	MS	Traditional cultivar
G14	Balirdia	Bangladesh	SB	Traditional cultivar
G15	Binnatoa	Bangladesh	MS	Traditional cultivar
G16	Parangi	Bangladesh	MB	Traditional cultivar
G17	Chengri	Bangladesh	MS	Traditional cultivar
G18	Dhala saitta	Bangladesh	SB	Traditional cultivar
G19	Morich boti	Bangladesh	SB	Traditional cultivar
G20	Saitta	Bangladesh	MB	Traditional cultivar
G21	Lal Dular	Bangladesh	MS	Traditional cultivar
G22	Nayan moni	Bangladesh	MS	Traditional cultivar
G23	Kalabokra	Bangladesh	MB	Traditional cultivar
G24	HUA565	Philippines	MS	Improved cultivar
G25	Takanari	Philippines	MB	Improved cultivar
G26	Kachalath	India	MS	Traditional cultivar
G27	Wkhi1	Philippines	MB	Improved cultivar
G28	Hukurikul193	Philippines	MB	Improved cultivar
G29	ML6	Malaysia	LS	Breeding line
G30	ML9	Malaysia	LS	Breeding line
G31	Wanxiam-P10	Malaysia	MS	Traditional cultivar
G32	RENGAN WANG	Malaysia	LS	Traditional cultivar
G33	PETEH PERAK	Malaysia	LB	Traditional cultivar
G34	WANGI PUTEH	Malaysia	MB	Traditional cultivar
G35	KUNYIT	Malaysia	MS	Traditional cultivar
G36	GHAU	Malaysia	LS	Traditional cultivar
G37	LALAMG	Malaysia	MS	Traditional cultivar
G38	MGAWA	Malaysia	MS	Traditional cultivar
G39	SUNGKAI	Malaysia	MS	Traditional cultivar
G40	UGAN	Malaysia	LS	Traditional cultivar

Code No	Name of Genotype	Source Country	Grain Size and Shape	Status of Sample
G41	TADOM	Malaysia	MS	Traditional cultivar
G42	BANGKUL	Malaysia	MS	Traditional cultivar
G43	NMR151	Malaysia	LS	Improved cultivar
G44	NMR152	Malaysia	LS	Improved cultivar
G45	MR297	Malaysia	LS	Improved cultivar
G46	Putra 1	Malaysia	LS	Improved cultivar
G47	Putra 2	Malaysia	LS	Improved cultivar
G48	MR 303	Malaysia	LS	Improved cultivar
G49	MR 309	Malaysia	LS	Improved cultivar
G50	BR24	Bangladesh	MS	Improved cultivar
G51	BRRI dhan48	Bangladesh	MS	Improved cultivar
G52	BRRI dhan82	Bangladesh	MS	Improved cultivar
G53	BRRI dhan72	Bangladesh	MS	Improved cultivar
G54	BRRI dhan28	Bangladesh	MS	Improved cultivar
G55	BRRI dhan39	Bangladesh	MS	Improved cultivar
G56	BRRI dhan42	Bangladesh	MS	Improved cultivar
G57	BRRI dhan43	Bangladesh	MS	Improved cultivar
G58	BRRI dhan46	Bangladesh	SB	Improved cultivar
G59	BRRI dhan75	Bangladesh	MS	Improved cultivar
G60	BRRI dhan55	Bangladesh	MS	Improved cultivar
G61	BRRI dhan69	Bangladesh	MS	Improved cultivar
G62	B370	India	LS	Improved cultivar
G63	BINASAIL	Bangladesh	MB	Improved cultivar
G64	BINA dhan7	Bangladesh	MB	Improved cultivar

Table 1. Cont.

2.2. Site of Experimentation

The pot experiment was conducted in a net house at the field 10, University Putra Malaysia (UPM), Malaysia. The experiment was conducted in two seasons, the first season being from February 2019 to June 2019 and the second season from August 2019 to December 2019. Geographically, the place is located at about 3°02′ N latitude and 101°42′ E longitude with an elevation of 31 m from the sea level, and it is characterized by a humid tropical climate. Details of the weather information are presented in Table 2.

Table 2. Month-wise average of daily maximum temperature, minimum temperature, mean temperature, and rainfall at UPM during experimentation period from February to June (1st planting season) and from August to December (2nd planting season) 2019.

	1st Pla	anting								
	Tempera	ture (°C)		Rain Fall		Rain Fall				
Month	Max.	Min.	Ave.	— (IIIII)	Month	Max.	Min.	Ave.	— (IIIII)	
February	35.52	26.23	30.88	118.76	August	33.48	25.57	29.53	114.76	
March	35.34	26.18	30.76	119.45	September	33.24	25.34	29.29	120.39	
April	35.48	26.14	30.81	120.62	Öctober	32.72	25.11	28.92	232.73	
May	34.75	25.77	30.26	121.58	November	32.66	24.82	28.74	235.41	
June	34.60	25.63	30.12	121.74	December	31.54	24.59	28.07	242.93	
Average	35.14	25.99	30.57		Average	32.73	25.09	28.91		
Total				602.15	Total				946.22	

2.3. Experimental Design and Treatments

The experiment was conducted following a randomized complete block design with three replication on each treatment. Twenty-day-old seedlings of each test genotypes were transplanted, and two seedlings were used per hill in 45 cm diameter and 52 cm height plastic pot with 15 kg soil and 20 cm spacing between hills. There were three (3) treatment combinations with chicken manure (CM) and chemical fertilizer recommended rate (CFRR) for high goal (HYG) as follows—T₁: CM (5 t ha⁻¹), T₂: CM (2.5 t ha⁻¹) + 50% CFRR (NPK) and T₃: 100% CFRR (NPK).

2.4. Application of Fertilizer and Operational of Intercultural

Organic fertilizer was incorporated into the soil before crop establishment, while a compound fertilizer (NPK 2.16:1.89:0.79) was applied at the rate of 5 t ha⁻¹. Triple super phosphate and muriate of potash were applied during final pot preparation, and urea was applied in two split doses at 25 days after seeding (DAS) and at 55 DAS, to supply total recommended nutrient of 150 N: 60 P₂O₅: 60 k₂O kg ha⁻¹. Both organic fertilizer and chemical fertilizer were applied as prescribed by the treatments. Weeding and other management practices were performed as and when required. Irrigation was also conducted whenever required.

2.5. Soil Analyses

Initial soil samples were taken from the surface to a depth of 0–15 cm. The samples were air-dried and crushed to pass through a 2 mm (10 meshes) sieve after being free of weeds, plant roots, stubbles, and stones. After that, the samples were placed in clean plastic bags to be analyzed chemically and mechanically. Standard procedures were used to assess the physical and chemical qualities of the initial and postharvest soil samples in Table 3. The textural class was calculated by projecting the values for percent sand, percent silt, and percent clay to the Marshall's Triangular Coordinate following the USDA methodology, and the particle size analysis of the soil was performed by hydrometer method [15]. Organic matter was determined by Walkley and Black method [16], soil pH (1:2.5 soil-water) by glass electrode pH meter method [17], total N by semi-micro Kjeldahl method [18], available P by Olsen method [19], exchangeable K by flame photometer after extraction with 1N NH₄OA_c at pH 7.0 [20], available S by extracting soil samples with CaCl2, solution (0.15%), and by measuring turbidity by spectrophotometer [21] method and CEC by sodium saturation method [15].

Table 3. Physiochemical characteristics of the initial and postharvest soil sample at the pot experiment on over two planting seasons, 2019.

	_			After Cro	p Harvest		
Soil Characters	Initial	5 t/h	a CM	2.5 t/ha CN	A + 50% CF	1009	% CF
Characters		1st Planting	2nd Planting	1st Planting	2nd Planting	1st Planting	2nd Planting
pН	5.9	6.01	6.03	6.02	6.03	6.0	6.01
EC $(\mu S/cm)$	54	57	59	61	64	63	67
CEC	16.72	17.23	17.83	18.54	19.36	19.22	19.48
Organic carbon (%)	0.67	0.71	0.74	0.76	0.81	0.77	0.80
Organic matter (%)	1.32	1.57	1.66	1.54	1.59	1.49	1.55
Total N (%)	0.07	0.09	0.12	0.11	0.14	0.15	0.18
Exchangeable K (cmolkg ⁻¹)	0.28	0.31	0.33	0.33	0.37	0.36	0.39
Available P (mgkg ⁻¹)	17.54	17.86	18.32	18.59	19.83	19.45	19.78
Sand	31.63	31.95	32.24	32.56	32.84	31.69	32.27
Silt	34.18	34.36	35.22	35.51	35.79	35.62	36.45
Clay	27.49	27.64	28.27	28.33	28.65	27.74	28.46
Soil texture	Clay loam	Clay loam	Clay loam	Clay loam	Clay loam	Clay loam	Clay loam

Chemical properties of the chicken manure (dry basis).								
Properties	Percentage							
Ν	2.16%							
Р	1.89%							
K	0.79%							

2.6. Plant Tissue Analyses

After harvest, plant samples were collected from each treatment, and the samples were separated into the shoot (above ground plant parts excluding the grains), root, and grain, after which they were oven-dried at 70 °C for 72 h. Oven-dried samples were ground in the laboratory using a Wiley hammer mill with 1 mm mesh size. The samples were analyzed for total nitrogen (N), phosphorus (P), potassium (K), calcium (Ca), and magnesium (Mg). The nutrients were determined using acid wet digestion method [22]. For the digestion process, ground samples of 0.25 g were transferred to clean 100 mL digestion flask, and 5 mL of concentrated sulfuric acid (H_2SO_4) was added to each flask. The samples were allowed to stand for 2 h, after which 2 mL of 50% hydrogen peroxide (H₂O₂) was added. The flasks were heated for 45 min at 285 °C and then allowed to cool. This process was repeated twice to let the digestion be clear (colorless). The flasks were then removed from the digestion block, cooled to room temperature, and made up to 100 mL with distilled water filtered through filter paper (Whatman no. 1). The digested samples were stored in plastic vials before analysis for N, P, K, Ca, and Mg. Nitrogen and potassium were determined with auto analyzer (AA) (Lachat instrument, Milwaukee, WI, USA), while potassium, calcium, and magnesium were determined using automatic absorption spectrometer (ASS) (Perkin Elmer, 5100, Waltham, MA, USA).

2.7. Obtaining the Data

The data on morphological, physiological, and yield characteristics were collected in this study, which includes the quantitative characters that can be counted or measured using specific measuring tools such as plant height (PH, cm), total number of tiller per plant (NT, no.), total number of panicle per plant (NP, no.), panicle length (PL, cm), number of filled grains per panicle (NFG, no), number of unfilled grains per panicle (UNFG, no), 1000 grain weight (TGW, g), grain yield per plant (YP, g), straw yield per plant (SY, g), harvest index (HI,%), and nutrient content (N, P, and K) of grain and straw samples.

2.8. Statistical Analysis

All evaluated data were analyzed by pooled statistical analysis software (SAS) version 9.4 to test for significant differences using the analysis of variance (ANOVA) procedure and least significant differences (LSD) ($p \le 0.001, 0.05$) to compare among the significant characteristics mean using the Duncan's new multiple range test (DNMRT) [23]. Prior to running ANOVA, data were tested for normal distribution and homogeneity of variance. These were used to determine the level of variation of all observed traits, which was brought about by genotypes, seasons, treatments, genotypes by treatments, genotypes by seasons, and genotypes by treatments by seasons to determine the level of variations.

3. Results

3.1. Morphological Traits

3.1.1. Plant Height

Plant height at the time when the plant reaches maturity varied from genotype to genotype; there was a significant difference ($p \le 0.01$) among the rice genotype, treatment, genotype by treatment, and genotype by season (combination of genotype and season) as presented in Table 4. Results indicated that the application of T₂ (2.5 t ha⁻¹ CM + 50% CFRR) produced the tallest plant height recorded in kalabokra (G23) (171.39 cm) followed by Kataktara (G13), Kaisa panja (G6), Saitta (G20), Nayan moni (G22), Parija (G12), and Panbira (G2) (169.98, 169.68, 163.97, 163.87, 163.45, and 162.65 cm, respectively), which were significantly higher than other treatments except T₃ (100% CFRR), which had a similar plant height. The application of T₁ (5 t ha⁻¹ CM) produced a shorter plant height recorded in BRRI dhan69 (G61), BRRI dhan46 (G58), BRRI dhan75 (G59), BRRI dhan42 (G56), BINA dhan7 (G64), BRRI dhan48 (G51), and BRRI dhan55 (G60) (114.77, 115.94, 115.99, 117.56, 117.74, 117.82, and 117.91 cm, respectively) was presented in Table 5.

Variable	DF	PH	NT	NFG	NUFG	NP	PL	1000-GW	YP	SY	HI
Replication with seasons (R)	4	188.37 **	21.66 **	1476.35 **	1110.73 **	12.53 **	7.87 **	7.79 **	1.71 **	86.66 **	0.01 **
Seasons (S)	1	137.78 **	227.57 **	1884.81 **	1146.12 **	147.20 **	0.46ns	5.68 **	75.21 **	124.79 **	0.01 **
Treatments (T)	2	2865.25 **	249.86 **	13191.15 **	3290.56 **	120.86 **	28.72 **	67.11 **	119.63 *	701.82 **	0.13 **
S×T	2	50.15 **	2.09 *	27.93 ns	34.96 **	9.04 **	16.95 **	1.10 **	5.76 **	16.10 **	0.01 **
$S \times R \times T$ (Error a)	8	0.84 ns	14.25 **	22.98 ns	5.11 ns	11.28 **	4.04 **	1.09 **	1.17 **	9.76 **	0.00 ns
Genotypes (G)	63	3433.63 **	54.74 **	8967.37 **	1982.55 **	28.44 **	125.48 **	152.32 **	186.68 **	159.08 **	0.20 **
G×S	63	1.92 **	0.94 **	32.94 **	16.30 **	0.90 **	2.50 **	0.27 **	0.88 **	1.14 **	0.00 **
$G \times T$	126	10.62 **	2.77 **	39.62 **	19.56 **	1.97 **	2.54 **	3.59 **	0.71 **	8.51 **	0.00 **
$G \times S \times T$	126	2.09 **	0.88 **	13.60 ns	4.96 **	0.82 **	2.27 **	0.20 ns	0.39 ns	0.83 ns	0.0 **
Error b	756	0.92	0.59	11.56	3.06	0.58	0.74	0.16	0.38	0.96	0.00

Table 4. Pooled analysis of variance mean square of growth traits across two planting seasons.

** = significant at $p \le 0.01$, * = significant at $p \le 0.05$, ns = not significant, DF = degree of freedom, PH = Plant height, NT = Number of tillers per plant, NFG = Number of filled grains per panicle, NUFG = Number of unfilled grains per panicle, NP = Number of panicles per plant, PL = panicle length, 1000-GW = 1000 grain weight, YP = Yield per plant, SY = Straw yield and HI = Harvest index.

3.1.2. Number of Tillers per Plant

The number of tillers per plant showed a significant difference ($p \le 0.01$) among the rice genotype, treatment, genotype by treatment, and genotype by season (combination of genotype and season) as presented in Table 4. Results indicated that the application of T₂ (2.5 t ha⁻¹ CM + 50% CFRR) produced a significantly higher number of tillers plant⁻¹ recorded in Takanari (G25) (14.88) followed by Putra 1 (G46), MR297 (G45), Putra 2 (G47), BR24 (G50), BRRI dhan48 (G51), and BRRI dhan39 (G55) (14.67, 14.56, 14.55, 13.77, 13.74, and 13.73, respectively), which were significantly higher than other treatments except T₃ (100% CFRR), which had a similar number of tillers plant⁻¹. The application of T₁ (5 t ha⁻¹ CM) gave the lowest number of tillers plant⁻¹ recorded in RENGAN WANG (G32), KUNYIT (G35), Nayan moni (G22), Kataktara (G13), Panbira (G2), BANGKUL (G42), and Saitta (G20) (6.46, 6.56, 6.55, 6.72, 6.73, 7.37, and 7.46, respectively) was presented in Table 5.

3.2. Physiological Traits

Straw Yield per Plant and Harvest Index

Straw yield per plant and harvest index showed a significant difference ($p \le 0.01$) among the rice genotype, treatment, genotype by treatment, and genotype by season (combination of genotype and season) as presented in Table 4. Results indicated that the application of T₂ (2.5 t ha⁻¹ CM + 50% CFRR) produced a significantly higher straw yield recorded in BRRI dhan39 (G55) (28.73 g plant⁻¹) followed by BR24 (G50), BRRI dhan75 (G59), BRRI dhan72 (G53), and BRRI dhan48 (G51) (28.63, 28.43, 28.15, and 27.84 g plant⁻¹, respectively), and harvest index recorded in MR309 (G49) (0.73%) followed by Putra 1 (G46), BR24 (G50), MR297 (G45), and Putra 2 (G47) (0.73%, 0.72%, 0.70%, and 0.67%, respectively) were significantly higher than other treatments except T₃ (100% CFRR), which had a similar straw yield and harvest index. The application of T₁ (5 t ha⁻¹ CM) recorded a lower straw yield in Kalabokra (G23), Vandana (G7), Kataktara (G13), Luanga (G5), and Panbira (G2) (15.18, 15.75, 16.44, 16.77, and 17.25), and harvest index recorded in RENGAN WANG (G32), Wanxiam-P10 (G31), BRRI dhan43 (G57), B370 (G62), and WANGI PUTEH (G34) (0.30%, 0.32%, 0.32%, 0.34%, and 0.35%, respectively) were presented in Table 6.

Genotype (G)	Treatment (T)	t	PH (cm)			NT		
C1	1	137.14b	140.56b	145.81b	125.87b	9.33a	9.72b	7.56b	11.72a
GI	3	139.15ab	143.98a	145.6b	124.27b	10.74a G17 10.25a	12.30a G33	9.11b	12.48a
LSD(0.0	05)	2.96	2.67	2.99	2.38	1.88	1.92	1.11	1.45
	1	155.11b	145.65b	141.69b	119.92b	6.73a	10.36b	7.99b	11.56a
G2	2	162.65a G18	149.17a G34	144.25a G50	122.64a G2	7.42a G18	12.62a G34	9.26a G50	13.73a
LSD(0)	05)	2 55	2 24	2 45	120.190	1 16	1.37ab	0.79a 1 13	11.09a 1.95
LOD (0.	1	136.84c	141.65a	138.04c	117.82b	8.38b	9.63b	6.56a	11.56b
G3	2	144.13a G19	142.66a G35	144.66a G51	122.43a G3	10.26a G19	11.28a G35	7.36a G51	13.73a
	3	140.86b	143.12a	140.66b	120.61a	9.66ab	10.82ab	7.11a 1.07	11.89ab
L3D(0.0	1	143.69a	161.74a	139.14a	121.07b	9.37a	7.46b	8.27b	11.34a
G4	2	146.14a G20	163.98a G36	141.91a G52	125.11a G4	10.66a G20	9.27 G36	9.72a G52	12.63a
LODIA	3	144.34a	163.55a	130.61b	124.97a	10.37a	8.79ab	8.32b	11.88a
LSD(0.0	05) 1	2.98 156 1ab	3.1 142.0b	2.9 126 5b	2.05 110.77b	1.62 8.45b	1.62	1.1 9.45ab	1.83
G5	2	160.08a G21	150.05a G37	140.57a G53	123.5a G5	10.73a G21	12.44a G37	9.68a G53	12.59a
60	3	154.63b	146.86b	141.04a	121.63ab	9.66b	11.56a	8.37b	11.28a
LSD(0.0	05)	4.12	3.04	2.72	2.97	1.45	1.88	1.11	1.74
C	1	164.8b	159.69b	147.22b 153.49p CE4	118.05a 120.74a CC	7.47b	6.56b	8.57a	11.46a
Go	$\frac{2}{3}$	163.82b	161.79ab	151.4a	120.74a Go 110.01a	9.07h	8.52a	8.79a G54	12.38a 11.88a
LSD(0.0	05)	3.35	3.78	3.14	3.1	1.16	1.17	1.07	2.44
-	1	155.23b	164.45b	143.11b	118.39a	8.38a	8.33b	7.58a	11.38
G7	23	160.37a G23	171.39a G39	150.24a G55	121.79a G7 121.55	9.89a G23	9.52ab G39	8.55a G55 7.69a	13.73
LSD(0.0	05)	3.16	3.59	3.47	3.76	1.46	1.39	1.61	1.61
(1	141.71b	121.69b	140.05b	117.65b	9.73a	8.67b	8.67b	9.67b
G8	2	144.72a G24	126.78a G40	143.78a G56	122.45a G8	11.56a G24	10.46ab G40	9.62a G56	12.61a
ISD(0)	05) 3	142.04ab 2.77	124.61ab 3.47	140.42b 2 38	121.12a 2.12	10.74a 2.4	9.66a 1.46	8.93b 1.09	11.3/a 2.19
L3D(0.0	1	144.1b	128.91b	149.51b	119.56b	8.45b	12.62a	7.66a	9.66a
G9	2	149.9a G25	133.67a G41	153.93a G57	124.16a G9	10.69a G25	14.88a G41	8.72a G57	11.83a
LCD(0)	3	143.78b	131.15ab	150.76ab	121.09b	9.53ab	14.47b	8.33a	11.25a
LSD(0.0	05) 1	3.44 140.26b	2.74 136.12b	3.49 147.1b	2.82 115 94b	1.64 10 55a	1.95 11.27b	1.45 7 37a	1.88 8.67b
G10	2	146.85a G26	141.49a G42	150.59a G58	121.4a G10	11.21a G26	12.68a G42	8.62a G58	11.62a
//	3	144.33a	137.78b	146.12b	122.01a	10.2a	11.85b	8.23a	9.56b
LSD(0.0	05)	2.71	1.9	2.89	3.34	1.13	1.89	1.61	1.72
G11	2	133.97C 141.99a C27	140.00a 143.55a C43	131.00D 137.73a C59	115.990 121.33a G11	10.62a 11.83a C27	6.25a 899a G43	9.560 12.62a C59	11.720 13.67a
011	3	137.92b	142.36a	131.68b	118.65b	10.75a	8.72a	11.38a	12.48ab
LSD(0.0	05)	3.17	2.96	2.71	1.75	1.59	1.82	1.13	1.52
C12	1	155.72b 163.45b C28	137.12 143.41 C44	127.26c	117.91c	8.32b	10.62ab	10.51a 12.27a C60	8.62b
GIZ	2	163.430 G28	139.04 G44	130.17b	123.00a G12 120.8b	9.02a G28 8.78b	12.07a G44 11.77b	12.27a G60 11.89a	10.37a
LSD(0.0	05)	3.29	3.13	2.38	1.13	1.22	1.98	2.37	1.49
	1	165.87b	132.12b	128.46b	114.77b	6.72b	11.56b	12.71a	10.72b
G13	2	169.98a G29	134.78a G45	133.37a G61	120.79a G13	8.67a G29	12.71a G45	14.56a G61	12.66a 11.45b
LSD(0.0	05)	3.5	2.41	3.44	2.59	1.15	1.85	2.54	1.85
	1	141.7b	127.22b	126.94b	121.6b	9.86b	10.78b	13.62a	8.52b
G14	2	146.11a G30	133.89a G46	132.66a G62	126.13a G14	11.36a G30	12.61a G46	14.67a G62	9.69a
ISD(0)	05) 3	143.67ab 2.91	131.58a 3 29	131.18a 1.65	124.68ab 3.49	11.72a 1.89	10.926	13.85a 2.48	9.28ab
L0D(0.	1	143.19b	143.83b	125.98c	119.15b	8.56b	7.56b	12.61b	8.36b
G15	2	148.12a G31	150.16a G47	131.07a G63	123.7a G15	12.62a G31	8.82a G47	14.56a G63	11.82a
	3	144.11b	145.89b	129.2b	121.09b	11.35a	7.64b	12.72ab	10.72a
LSD(0.0	1	3.10 145.75b	∠./o 138.84b	1.09 122.77c	∠.01 117.74b	1.02 9.26a	1.12 6.46b	1.07 10.61a	1.52 9.61a
G16	2	153.95a G32	142.38a G48	130.15a G64	119.59b G16	11.56a G32	7.88a G48	11.59a G64	10.77a
	3	151.35a	140.18ab	125.84b	124.64a	11.21a	7.17ab	11.27a	10.28a
LSD(0.0	05)	3.15	3.18	2.91	2.8	2.71	1.12	1.94	1.87

Table 5. Plant height and number of tillers per plant of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

In a column, means with similar letter (s) are statistically identical, and those with dissimilar letter (s) differ significantly as per 0.05 level of probability, $T_1 = 5 \text{ t ha}^{-1} \text{ CM}$, $T_2 = 2.5 \text{ t ha}^{-1} + 50\%$ CFRR, $T_3 = 100\%$ CFRR, PH= plant height, and NT= number of tillers per hill.

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Genotype 7 (G)	Freatment (G)		SY (g)				HI (%	.)	
	1	22.82b	23.38a	19.53c	25.55a	0.57a	0.58b	0.42a	0.68a
G1	2	24.63a G17	24.85a G33	23.48a G49	27.38a G1	0.57a G17	0.56b G33	0.47a G49	0.66a
/	3	24.28a	24.17a	21.36b	25.38a	0.56a	0.65a	0.44a	0.69a
LSD(0.05))	1.31	1.28	1.83	2.74	0.03	0.02	0.07	0.03
~	1	17.25a	22.95a	21.73b	26.33a	0.51a	0.63a	0.38a	0.68ab
G2	2	18.73a G18	24.89a G34	23.67a G50	28.63a G2	0.54a G18	0.6a G34	0.39a G50	0.676
1 50/0.05	5	10.54a 1.87	23.70a 2.44	20.190	27.10a 3.11	0.04a	0.05a	0.39a	0.71a
LOD(0.00)	1	24.56ab	23.56b	19.2a	25.68b	0.52b	0.59b	0.42a	0.63b
G3	2	25.38a G19	27.45a G35	23.45b G51	27.84a G3	0.54ab G19	0.61b G35	0.44a G51	0.63b
	3	23.68b	26.63a	20.63	27.07ab	0.58a	0.68a	0.46a	0.69a
LSD(0.05))	1.69	1.81	1.81	1.66	0.06	0.03	0.06	0.03
<u></u>	1	23.67ab	17.48b	22.36b	27.34a	0.54b	0.51b	0.36b	0.62a
G4	2	24.26a G20	18.93a G36	24.61a G52	18.56a G4	0.51b G20	0.54ab G36	0.41a G52	0.63a
I SD(0.05)	1	22.550	16.000	22.000	27.22a 2.86	0.384	0.59a	0.39a	0.05a
LOD(0.00)	1	16.77b	22.62ab	19.63b	27.44a	0.47a	0.6a	0.48ab	0.64a
G5	2	18.72a G21	25.21a G37	24.58a G53	28.15a G5	0.46a G21	0.56a G37	0.43b G53	0.64a
	3	18.58ab	21.46b	20.47b	27.8a	0.47a	0.57a	0.53a	0.67a
LSD(0.05))	1.96	2.85	2.49	3.05	0.04	0.04	0.06	0.04
<i>C</i> (1	17.83ab	17.27b	18.53ab	25.46a	0.46a	0.51ab	0.44b	0.39a
G6	2	20.35a G22	20.84a G38	20.74a G54	27.39a G6	0.396 G22	0.56 G38	0.44b G54	0.42a 0.45a
I SD(0.05)	2 66	17.110	2 10	23.20a 3.37	0.47a	0.06a	0.464	0.45a
LOD(0.00)	1	15.75b	15.18b	18.93b	27.16ab	0.44a	0.48a	0.37ab	0.6ab
G7	2	18.33a G23	19.34a G39	23.45a G55	28.73a G7	0.49a G23	0.44a G39	0.35b G55	0.57b
	3	16.72b	16.78b	19.75b	26.38b	0.47a	0.49a	0.39a	0.63a
LSD(0.05))	1.79	2.11	1.99	2.09	0.05	0.09	0.02	0.04
	1	22.95b	26.56a	20.53b	26.56ab	0.52b	0.68a	0.45b	0.38b
G8	2	25.74a G24	27.43a G40	22.39a G56	27.44a G8	0.55b G24	0.68ab G40	0.45b G56	0.356
1 50/0.05	5	23.360	23.95a 2.24	19.030	23.710	0.02	0.040	0.01	0.46a
L3D(0.03	1	24.53ab	22.97b	18.37b	2.09 24.78a	0.56b	0.64a	0.43a	0.36b
G9	2	25.98a G25	25.26a G41	21.83a G57	27.29a G9	0.53b G25	0.59b G41	0.4a G57	0.4b
	3	23.37b	22.57b	18.97b	25.48a	0.62a	0.65a	0.44a	0.36a
LSD(0.05))	2.44	2.22	1.67	3.08	0.03	0.04	0.04	0.02
610	1	22.89b	23.66a	18.35a	25.97a	0.59a	0.56a	0.42b	0.38a
G10	2	25.57a G26	24.39a G42	20.54a G58	27.45a G10	0.54b G26	0.54a G42	0.42b G58	0.39a
I SD(0.05)	3 21	22.05a 2.11	2 33	20.00a 3.48	0.03	0.37a	0.07	0.05
E6E(0.00)	′ 1	25.64c	19.37b	22.78b	27.37ab	0.55b	0.41a	0.37b	0.6b
G11	2	21.52a G27	21.19a G43	25.26a G59	28.43a G11	0.63a G27	0.43a G43	0.38b G59	0.6ab
	3	18.86b	19.24b	21.83b	26.35b	0.64a	0.41a	0.43a	0.62a
LSD(0.05))	2.33	1.64	1.84	2.21	0.05	0.03	0.02	0.02
C10	1	17.44b	24.08a	21.67b	24.35a	0.49b	0.59a	0.41b	0.366
GIZ	2	21.55a G28	25.56a G44	24.87a G60	20.00a G12	0.45ab G28	0.550 G44	0.44D G60	0.44a 0.44a
LSD(0.05)	1.73	2.16	1.69	2.95	0.08	0.05	0.04	0.05
202 (0100)	′ 1	16.44b	18.24c	25.46a	24.83a	0.56b	0.51a	0.67a	0.37a
G13	2	18.78a G29	25.64a G45	27.63a G61	26.74a G13	0.56b G29	0.4b G45	0.66a G61	0.37a
	3	17.52ab	22.86b	25.96a	25.16a	0.6a	0.43b	0.71a	0.36a
LSD(0.05))	1.69	1.80	2.27	2.95	0.03	0.04	0.07	0.07
C14	1	23.65b	20.75b	25.17a	22.36ab	0.58b	0.48a	0.68b	0.39a
G14	2	25.65a G30	23.34a G46 24.27a	20.75a G62	20.55a G14 24.72h	0.560 G30	0.370 G46 0.48b	0.050 G62	0.35a 0.36a
LSD(0.05)	1.45	1.94	2.16	2.58	0.05	0.02	0.02	0.10
202 (0.00)	1	22.26b	22.67c	26.38a	22.87a	0.059ab	0.48a	0.65a	0.41a
G15	2	25.63a G31	26.61a G47	26.77a G63	23.56a G15	0.056b G31	0.34b G47	0.66a G63	0.42a
	3	23.45b	24.78b	5.85a	22.45a	0.062a	0.35b	0.7a	0.43a
LSD(0.05))	2.27	1.31	2.49	1.65	0.04	0.04	0.06	0.05
C16	1	23.56a	20.75C	25.73a	22.53ab	0.580 0.57b C22	0.41a	0.65a	0.41a
G10	∠ 3	24.70a G32	22.40a G48	20.05a G64	23.20a G16 22.67b	0.570 G32	0.350 G48	0.00a G64	0.44a 0 44a
LSD(0.05)	3.28	2.48	2.93	2.58	0.04	0.07	0.03	0.04

Table 6. Straw yield and harvest index of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

In a column, means having similar letter (s) are statistically identical and those having dissimilar letter (s) differ significantly as per 0.05 level of probability, $T_1 = 5 \text{ t ha}^{-1} \text{ CM}$, $T_2 = 2.5 \text{ t ha}^{-1} + 50\%$ CFRR, $T_3 = 100\%$ CFRR, SY= straw yield per plant, and HI = harvest index.

3.3. Yield and Yield Contributing Traits

3.3.1. Number of Panicles per Plant and Panicle Length

In order to increase the grain yield, the most important aspect on the growth of rice is the panicle number per plant. There was a highly significant difference ($p \le 0.01$) among the rice genotype, treatment, genotype by treatment, and genotype by season (combination of genotype and season) on number of panicles $plant^{-1}$ and panicle length as presented in Table 4. Results indicated that the application of T₂ (2.5 t ha⁻¹ CM + 50%) CFRR) produced significantly higher numbers of panicles plant⁻¹ recorded in HUA565 (G24) (12.18) followed by MR297 (G45), Putra 1 (G46), Putra 2 (G47), and BRRI dhan39 (G55) (11.37, 11.26, 11.16, and 110.61, respectively) and longest panicle length recorded in MR297 (G45) (27.49 cm) followed by Putra 1 (G46), BRRI dhan28 (G54), BRRI dhan48 (G51), and Putra 2 (G47) (26.98, 26.89, 26.86 and 26.55 cm, respectively), which were significantly higher than other treatments except T₃ (100% CFRR), which had a similar number of panicle plant⁻¹ and panicle length. The application of T_1 (5 t ha⁻¹ CM) recorded a lower number of panicles $plant^{-1}$ in WANGI PUTEH (G34), Nayan moni (G22), Panbira (G2), Kataktara (G13), and RANGAN WANG (G32) (4.93, 5.28, 5.27, 5.72, and 5.75, respectively) and the shortest panicle length recorded in Kaisa panja (G6), Vandana (G7), Kataktara (G13), Kalabokra (G23), and Saitta (G20) (16.46, 17.16, 17.19, 17.20, and 17.43 cm, respectively) were presented in Table 7.

Table 7. Number of panicles per hill and panicle length of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

Genotype (G)	Treatment (T)		NP PL (cm)													
	1	8.33a		8.27b		6.46b		9.37a		22.24b		22.66a		19.53a		25.47b
G1	2	8.59a	G17	10.42a	G33	8.21a	G49	10.52a	G1	23.16a	G17	23.12a	G33	20.26a	G49	25.96a
01	3	7.28a	017	9.38ab	000	6.83ab	01/	10.29a	01	22.39b	01/	22.47a	000	26.13a	01/	25.56b
LSD(0.05	5)	1.99		1.51		1.49		1.65		0.02		0.61		6.83		0.23
(1	5.28a		8.52a		4.93b		9.57a		17.46b		22.29a		20.23b		25.43a
G2	2	5.63a	G18	9.73a	G34	7.37a	G50	9.71a	G2	18.31a	G18	22.48a	G34	20.43a	G50	25.56a
	3	5.49a		9.45a		7.15a		9.28a		17.16b		21.91b		20.26b		25.27a
LSD(0.05	5)	1.16		1.99		1.17		2.85		0.30		0.27		0.16		0.24
	´ 1	9.08a		8.15a		6.48a		9.28a		21.4b		21.46b		18.36b		26.36a
G3	2	8.27a	G19	9.24a	G35	6.64a	G51	10.09a	G3	22.33a	G19	22.27a	G35	19.03a	G51	26.79a
	3	7.82a		8.68a		6.71a		9.78a		22.06a		22.37a		18.86a		26.45a
LSD(0.05	5)	1.89		2.21		1.18		1.66		0.35		0.29		0.30		0.27
	1	7.56a		6.46a		7.38ab		8.75a		22.36b		17.43b		20.2a		26.52a
G4	2	8.37a	G20	7.38a	G36	8.44a	G52	10.24a	G4	23.27a	G20	18.33a	G36	20.35a	G52	26.4ab
	3	8.72a		7.19a		7.68b		9.68a		22.4b		17.29a		20.27a		26.23b
LSD(0.05	5)	1.65		1.63		1.17			1.91	0.39		0.31		0.35		0.18
	1	7.62ab		8.92a		8.26a		8.64ab		18.06b		21.36c		19.4a		25.73b
G5	2	8.37a	G21	9.66a	G37	8.53a	G53	9.72a	G5	19.37a	G21	22.23a	G37	19.94a	G53	26.08ab
//	3	6.86b		9.45a		7.88a		7.89b		19.14a		22.01b		19.67a		26.26a
LSD(0.05	5)	1.50		1.51		1.19		2.01		0.41		0.14		0.63		0.36
	1	6.79b		5.28b		5.78b		8.85a		16.46c		18.1a		20.93a		26.39a
G6	2	8.57a	G22	6.74a	G38	8.18a	G54	9.29a	G6	17.38a	G22	18.36a	G38	21.12a	G54	26.86a
	3	7.83ab		7.46a		7.63a		8.46a		17.18b		18.3a		20.98a		26.47a
LSD(0.03) 1	1.19		1.17		1.18		1.51		0.22		0.38		0.42		0.31
	1	7.16a	C 222	6.78a	G2 0	6./8a	055	9.2/a	07	17.160	C22	17.2D	C2 0	19.2a	055	25.43a
G/	2	7.34a	G23	7.59a	G39	7.65a	G55	10.61a	G7	17.78a	G23	18.26a	G39	19.8/a	G55	25.88a 25.27a
	5	7.22a 1.51		0.12a 1.40		0.00a 1.62		9.70d 1.10		10.090		0.25		19.02a		23.27a
L3D(0.0.	<i>)</i> /1	8 282		10.47		7.56ab		6.03h		21.380		23.22		18 362		22.28h
C ⁸	2	0.20a 0.13a	C24	10.47 a 12 18 a	C_{40}	8 73a	C56	0.950	C ⁸	21.50C 22.54a	C24	20.2a 26.13a	C40	10.50a	C56	22.200
60	3	8.66a	624	11.10a	640	7 53h	650	9.17a	Go	22.04a 22.28h	624	26.15a 26.06a	640	19.00a	650	23.30a 23.33a
LSD(0.05	5)	1 19		2 13		1 16		1 92		0.28		0.82		0.38		0.28
E0D (0.00	1	7.26a		9.26a		7 12a		7 29h		22.38c		23.26a		20.35h		21 54c
G9	2	8.41a	G25	10.52a	G41	7.34a	G57	8.74a	G9	23.28a	G25	23.3a	G41	21.04a	G57	22.69a
0,	3	7.79a	020	8.69a	011	6.85a	007	7.68ab	0/	22.76b	020	22.96b	011	20.76b	007	22.25b
LSD(0.05	5)	1.92		2.32		1.49		1.19		0.24		0.27		0.27		0.29
(0.00	1	7.19b		7.27b		7.83a		8.56a		23.46ab		22.36b		19.29b		20.47c
G10	2	9.35a	G26	9.43a	G42	7.19a	G58	10.28a	G10	23.75a	G26	22.96a	G42	19.86a	G58	21.28a
	3	8.29ab		8.48ab		6.96a		9.75a		23.49b		22.63b		19.49a		20.88b
LSD(0.05	5)	1.15		1.64		1.81		1.91		0.19		0.28		0.22		0.27
,	1	7.57a		6.69a		8.23b		6.79ab		22.42c		19.06ab		18.75ab		25.38b
G11	2	8.62a	G27	7.33a	G43	10.17a	G59	7.45a	G11	23.36a	G27	19.43a	G43	19.67a	G59	25.89a
	3	8.31a		7.24a		8.64ab		5.83b		22.89b		18.96b		19.35b		25.68b
LSD(0.05	5)	1.99		1.16		1.79		1.65		0.29		0.41		0.62		0.28

Genotype (G)	Treatment (T)				NP								PL (cn	า)		
	1	7.02a		9.57a		8.75a		8.62ab		18.24c		22.23a		19.18a		22.28ab
G12	2	6.64a	G28	9.31a	G44	9.29a	G60	9.56a	G12	19.35a	G28	22.5a	G44	19.48a	G60	22.76a
	3	6.59a		8.64a		9.45a		8.48b		19.08b		22.36a		19.37a		21.75b
LSD(0.	05)	1.48		1.99		1.66		1.66		0.32		0.27		0.21		0.39
	1	5.72b		10.73a		10.28a		8.27ab		17.19b		21.1ab		26.96c		23.06a
G13	2	7.46a	G29	9.46b	G45	11.37a	G61	9.49a	G13	18.27a	G29	21.4a	G45	27.49a	G61	23.48a
	3	5.68b		7.73c		10.28a		7.61b		17.3b		20.96b		27.25b		22.97a
LSD(0.	05)	1.18		1.17		1.18		1.67		0.35		0.40		0.24		0.39
	1	8.11a		10.23a		10.68a		6.72b		22.33a		20.13b		26.35a		21.28a
G14	2	8.57a	G30	8.72ab	G46	11.26a	G62	7.48a	G14	22.46a	G30	21.1a	G46	26.89a	G62	21.67a
	3	8.52a		7.92b		9.87a		7.83ab		21.63b		20.36b		26.67a		21.13a
LSD(0.	05)	2.2		1.79		2.23		1.19		0.60		0.39		0.49		0.41
	1	7.25b		5.88b		10.28a		6.67b		23.38a		19.26b		25.48b		22.38a
G15	2	9.43a	G31	6.63a	G47	11.16a	G63	9.18a	G15	23.29a	G31	20.3ab	G47	26.55a	G63	22.56a
	3	8.67ab		7.47a		10.46a		7.74ab		19.76a		26.2a		26.32a		22.29a
LSD(0.	05)	1.97		1.18		1.79		1.91		0.56		0.91		0.25		0.28
	1	7.83a		5.75b		9.47a		7.33a		21.52b		20.33b		26.46a		23.07b
G16	2	9.35a	G32	6.49ab	G48	8.72a	G64	8.18a	G16	22.37a	G32	21.03a	G48	26.49a	G64	23.47a
	3	8.74a		6.82a		9.38a		8.43a		21.95a		20.4b		26.28a		23.11b
LSD(0.	05)	2.31		1.18		1.51		1.9		0.288		0.33		0.33		0.35

Table 7. Cont.

In each column, means with similar letter (s) are statistically identical, and those with dissimilar letter (s) differ significantly as per 0.05 level of probability, $T_1 = 5 \text{ t} \text{ ha}^{-1} \text{ CM}$, $T_2 = 2.5 \text{ t} \text{ ha}^{-1} + 50\%$ CFRR, $T_3 = 100\%$ CFRR, NP = number of panicles per hill, and PL = panicle length.

3.3.2. Number of Filled Grains and Number of Unfilled Grains per Panicle

The number of filled and unfilled grains had a significant difference ($p \le 0.01$) among the rice genotype, treatment, genotype by treatment, and genotype by season (combination of genotype and season) as presented in Table 4. Results indicated that the application of T_2 $(2.5 \text{ t ha}^{-1} \text{ CM} + 50\% \text{ CFRR})$ was significantly higher on number of filled grains panicle⁻¹ recorded in BRRI dhan48 (G51) (176.41) followed by HUA565 (G24), MR297 (G45), BR24 (G50), and Putra 2 (G47) (171.42, 171.26, 170.53, and 168.27, respectively), and a similar number of filled grains panicle⁻¹ were produced in T₃ (100% CFRR). The application of T_1 (5 t ha⁻¹ CM) recorded the lowest number of filled grains panicle⁻¹ in TADOM (G41), Vandana (G7), GHAU (G36), MGAWA (G38), and Kataktara (G13) (85.39, 85.46, 88.45, 89.46, and 89.56, respectively). The application of T_1 (5 t ha⁻¹ chicken manure) was significantly higher on number of unfilled grains panicle⁻¹ recorded in BINASAIL (G63) (67.21) followed by BINA dhan5 (G65), GHAU (G36), RENGAN WANG (G32), and BINA dhan7 (G64) (65.32, 63.33, 63.27, and 62.70, respectively). The application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded the lowest number of unfilled grains panicle⁻¹ in BRRI dhan48 (G51), Putra 1 (G46), Kachalath (G26), Kalabokra (G23), and Putra 2 (G47) (19.26, 20.71, 21.75, 21.87, and 24.22, respectively), and a similar number of unfilled grains panicle⁻¹ also recorded in T₃ (100% CFRR) were presented in Table 8.

3.3.3. 1000 Grain Weight and Yield per Plant

When it comes to rice production, stable and high-yielding genotypes are necessary, and 1000-grain weight of grains is a measurement of grain size. Grain size multiplied by grain number results in a measurement of total yield of grain. Here, 1000-grain weight is expressed as an effective function of grain yield. There was a significant difference ($p \le 0.01$) among the rice genotype, treatment, genotype by treatment, and genotype by season (combination of genotype and season) as presented in Table 4. Results indicated that the application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded 1000-grain weight in BRRI dhan48 (G51) (25.85 g) followed by BR24 (G50), Putra 2 (G47), BRRI dhan55 (G60), and BRRI dhan69 (G61) (25.77, 25.73, 25.62, and 25.57 g, respectively), which were significantly higher than the other treatments except T_3 (100% CFRR), which had a similar 1000-grain weight. Further, the application of T_1 (5 t ha⁻¹ CM) produced the lowest 1000-grain weight observed in KUNYIT (G35), Kataktara (G13), Parija (G12), Nayan moni (G22), and Panbira (G2) (10.05, 14.26, 14.67, 15.58, and 15.70 g). The application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) produced a yield plant⁻¹ recorded in MR297 (G45) (17.34 g), followed by BR24 (G50), Putra 2 (G47), Putra 1 (G46), and BRRI dhan72 (G53) (17.32, 16.85,

16.81, and 16.78 g plant⁻¹, respectively), which were significantly higher than the other treatments except T_3 (100% CFRR), which had a similar yield plant⁻¹. The application of T_1 (5 t ha⁻¹ CM) produced lower yield plant⁻¹ observed in RANGAN WANG (G32), SUNGKAI (G39), Vandana (G7), Kalabokra (G23), and BANGKUL (G42) (6.15, 6.29, 6.34, 6.71, and 6.88 g plant⁻¹, respectively) were presented in Table 9.

Table 8. Number of filled grains and unfilled grains per plant of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

Genotype (G)	Treatment (T)		NFG			NUFG							
G1	1 2 3	121.28c 136.56a G17 131.48b	133.41a 140.27a G33 133.64a	99.72b 108.37a G49 105.835b	161.27a 166.35a G1 165.19a	32.27a 29.62a G17 34.56a	37.66a 30.83b G33 35.68a	62.67a 52.19b G49 54.73b	33.57a 28.96b 31.62ab				
LSD(0.05	5) 1	4.15 103.26b	7.35 124.57b	6.69 106.28b	6.88 164.19b	7.59 40.65a	5.45 46.58a	5.39 55.17a	4.89 30.51a				
G2	2 3	114.18a G18 111.09a	139.36a G34 134.62a	113.22a G50 110.15ab	170.53a G2 168.48ab	35.58a G18 38.77a	40.84b G34 44.36ab	45.2b G50 48.34b	26.38a 30.08a				
LSD(0.0	5) 1 2	6.06 128.31c 141.05a C10	6.35 141.28b 150.22a C25	5.7 105.28a 112.21a (CE1	5.61 172.28b	6.73 45.23a	5.66 36.73a 20.65b - C25	5.85 56.28a	4.31 21.88a				
GS LSD(0.0	3 5)	134.63b 5.84	130.32a G35 145.56b 5.2	112.51a G51 105.57a 7.87	170.41a G3 170.52ab 5.16	40.64a G19 43.53a 6.55	36.44a 4.61	48.476 G51 55.06a 4.41	19.26a 22.64a 6.77				
G4	1 2	135.18b 147.34a G20	106.34b 117.28a G36	88.45b 102.63a G52	165.27a 167.22a G4	37.23a 28.79b G20	46.28a 40.86b G36	63.33a 55.72b G52	36.32a 32.18a				
LSD(0.05	5) 1	142.29a 5.11 107.25c	111.74b 5.11 145.26b	94.57ab 8.31 109.28b	165.19a 5.88 160.185	33.64ab 5.86 39.65a	45.57ab 5.57 27.52ab	61.14a 5.17 41.56a	35.76a 4.8 36.44a				
G5	2 3	107.25C 122.34a G21 115.3b	143.360 161.52a G37 157.38a	109.280 118.36a G53 113.69ab	167.26a G5 161.42a	35.78a G21 40.59a	24.26b G37 30.63a	41.50a 35.25a G53 40.05a	32.38a 37.84a				
LSD(0.03	5) 1 2	5.49 94.57b 116.28a G22	5.42 108.17b 119.42a G38	5.11 89.46b 103.28a G54	7.23 116.45b 137.35a G6	5.78 46.33a 42.5a G22	4.85 39.76a 34.18a G38	6.86 53.28a 45.44b G54	5.69 60.32a 50.71b				
LSD(0.05	3 5)	111.19a 7.38	111.27a 5.07	98.57a 6.06	131.25a 7.38	44.18a 5.46	37.67a 5.67	50.23ab 6.55	56.27ab 5.68				
G7	1 2 3	85.46b 98.38a G23 95.27a	96.28b 113.45a G39 109.63a	110.27b 122.16a G55 119.42a	152.19b 165.26a G7 163.09a	35.73a 27.62b G23 35.41a	25.36ab 21.87b G39 29.44a	38.22a 30.18b G55 36.64ab	33.26a 30.64a 36.5a				
LSD(0.05 G8	5) 1 2	5.69 128.38b 140.41a G24	6.37 159.38b 171.42a G40	5.69 105.56b 113.32a G56	5.16 115.28c 126.41a G8	7.14 40.5a 35.27b G24	4.36 34.08a 28.47a G40	6.15 47.32a 40.52b G56	5.88 49.55a 43.27b				
LSD(0.05	3 5) 1	135.56ab 6.59 122.24b	168.56a 5.2 152.26b	118.24a 6.79 85.30b	121.58b 4.56 110.20b	40.18a 4.66 24.25ab	31.55a 6.55 39.61a	45.71ab 5.46 35.072	46.09ab 6.08 41.252				
G9	2 3	133.29a G25 132.42a	161.54a G41 156.44ab	103.18a G57 99.46a	110.290 122.34a G9 113.56b	32.4b G25 38.29a	33.56b G41 35.73ab	30.44a G57 33.76a	42.76a 44.2a				
LSD(0.05	5) 1	7.62 137.57b	6.88 141.18b	5.99 94.28b	6.75 111.23b	4.77 26.33b	5.38 26.37a	6.64 50.3a	5.54 37.22ab				
G10 LSD(0.05	2 3 5)	146.63a G26 143.74ab 6.69	150.28a G42 145.63ab 6.38	110.42a G58 105.74a 6.72	122.31a G10 115.4b 5.49	30.45ab G26 35.68a 5.77	21.75a G42 25.47a 7.08	42.62b G58 47.55ab 6.43	36.53b 41.8a 5.21				
G11	1 2	133.28b 146.16a G27	107.11b 121.07a G43	113.67b 122.28a G59	154.29b 165.31a G11	48.54b 37.67a G27	45.32a 40.4a G43	41.25a 35.69a G59	41.17a 34.86a				
LSD(0.05	5) 1	141.64a 6.94 93.74b	6.03 138.26b	6.51 103.19b	161.48a 5.93 117.28b	52.48b 6.52 42.07a	44.28a 4.85 31.35a	41.52a 5.94 52.73a	37.24a 7.23 40.56ab				
G12	2 3	111.47a G28 104.28a 9.65	150.32a G44 145.07a 6.17	116.28a G60 112.43a 5.81	128.31a G12 125.69a 6 79	36.85a G28 41.37a 5.49	28.73a G44 33.25a 6.45	40.45b G60 45.11b 5.47	37.32b 44.05a				
G13	1 2	89.56b 102.49a G29	108.67b 120.1a G45	162.18b 171.26a G61	115.57b 126.42a G13	52.35a 48.72a G29	51.47a 40.37c G45	33.28a 30.61a G61	51.23a 44.64a				
LSD(0.05	5) 1	96.37ab 6.65 122.17b	115.58a 5.11 115.67b	167.52ab 6.63	121.37ab 6.83	53.21a 4.74 26 55 a	45.11b 5.29	35.67a 5.13 24.44ab	50.6a 6.79				
G14	$\frac{1}{2}$	132.17b 141.34a G30 135.27ab	115.67b 126.45a G46 122.73a	162.41a G62 161.53a	94.696 111.26a G14 107.56a	35.27a G30 40.62a	55.37a 45.28c G46 50.71b	24.44ab 20.71b G62 31.69a	53.2b 56.31ab				
LSD(0.05	5) 1	6.75 138.26c	5.28 111.18b	6.49 162.46b	8.91 105.28b	6.54 32.42a	3.99 55.24a	9.01 25.35ab	5.29 67.21a				
G15 LSD(0.04	2 3 5)	152.19a G31 143.53b 5.15	120.34a G47 115.41ab 4.93	168.27a G63 167.71a 5.19	113.28a G15 110.73ab 7.71	30.58a G31 34.66a 7.53	51.62a G47 55.23a 5.04	24.22b G63 29.84a 4.57	61.08b 63.44ab 5.81				
G16	1 2	145.55b 157.48a G32	102.72b 111.33a G48	160.58a 164.27a G64	115.26b 123.62a G16	44.38a 39.64b G32	63.27a 51.77b G48	34.16a 31.62a G64	62.7a 55.27b				
LSD(0.05	5)	5.41	107.28ab 5.99	5.86	6.59	42.49ab 4.57	56.83b 5.93	5.36	61.32a 4.98				

In each column, means with similar letter (s) are statistically identical, and those with dissimilar letter (s) differ significantly as per 0.05 level of probability, $T_1 = 5 \text{ t ha}^{-1} \text{ CM}$, $T_2 = 2.5 \text{ t ha}^{-1} + 50\%$ CFRR, $T_3 = 100\%$ CFRR, NFG = number of filled grains per plant, and NUFG = number of unfilled grains per plant.

Genotype [(G)	Treatment (T)		1000-GW	(g)	1000-GW (g) YP (g)								
G1	1 2 3	18.26b 20.43a G17 18.79b	20.18b 21.84a G33 20.48b	18.53b 20.77a G49	24.33a 25.51a G1 25.07a	11.43b 12.27a G17 12.04ab	12.33a 7.4 13.25a G33 9.9 12.78a 84	47b 91a G49	15.51a 16.46a 16.08a				
LSD(0.05) 1	0.58 15.7a	0.50 19.55a	0.93 17.88c	0.87 24.76b	0.99 7.83a	0.88 1. 12.89a 7.3	06 34b	0.95 16.19b				
G2	2 3	16.25a G18 15.89a	20.15a G34 19.74a	19.25a G50 18.76b	25.77a G2 25.42ab	8.89a G18 8.68a	13.11a G34 8.3 13.82a 7.4	33a G50 45b	17.32a 17.05ab				
G3) 1 2	0.41 19.77b 21.43a G19	0.36 20.56a 21.11a G35	0.67 10.05b 20.68a G51	0.81 24.64b 25.85a G3	1.29 11.37b 12.51a G19	1.64 0. 12.36a 7.4 13.16a G35 9.5	52 49c 58a G51	15.24a 16.29a				
LSD(0.05	3	20.65b 0.52	20.73a 0.28	20.87a 0.69	24.79ab 0.99 24.885	12.11ab 0.84	12.77a 8.5 0.48 1.	54b 02	15.89a 1.38				
G4	1 2 3	20.85a G20 20.16a	15.82b 17.22a G36 16.65ab	21.43a G52 20.45b	24.880 25.51a G4 25.2ab	11.24a 11.85a G20 11.47a	9.16a G36 9.0 9.07a 7.8	03a G52 89b	16.4a 15.58a				
LSD(0.05) 1 2	0.65 17.67b	0.67 21.62b 22.56a C27	0.48 18.52b	0.69 24.9b 25.48a CE	1.11 7.02a 7.00a C21	0.59 0. 12.59a 8.4	99 14b	1.74 15.77a				
G5 LSD(0.05	3	18.25a G21 18.05a 0.64	22.36a G37 22.2ab 0.76	19.06ab 0.57	25.46a G5 25.26ab 0.56	7.68a 1.23	12.64a 9. 1.71 0.	5a G53 5a 79	16.35a 2.25				
G6	1 2 3	16.72b 17.85a G22 17.33a	15.58c 17.22a G38 16.46b	19.65a 20.45a G54 20.27a	23.67b 25.45a G6 24.97a	7.39a 7.56a G22 7.41a	7.79b 7.3 9.32a G38 8.1 9.06a 7.5	39b 17a G54 76ab	9.16a 10.33a 9.84a				
LSD(0.05) 1	0.61 16.35b	0.64 19.26b	0.62 17.88c	0.73 24.55b	0.75 6.34b	0.63 0. 6.71a 6.2	71 29b	1.09 14.71a				
G7 LSD(0.05	3	17.51a G23 16.89b 0.48	20.83a G39 20.35ab 0.73	19.74a G55 18.92b 0.564	25.45a G7 25.25a 0.56	7.75a G23 6.89ab 0.96	8.55a G39 7.3 7.68a 6.9 1.52 0.	96a G55 99a 63	15.43a 14.89a 0.94				
G8	1 2 3	17.78c 19.56a G24 18.58b	24.35c 22.07a G40 21.67b	18.37b 19.56a G56 18.84ab	23.52b 24.67a G8 24.11a	10.89b 12.78a G24 12.52a	13.64b 8.1 16.37a G40 9.0 16.22a 80	12b)5a G56	8.67a 9.43a 9.07a				
LSD(0.05) 1	0.54 19.26ab	0.59 20.62c	0.62 17.66a	0.81 24.08b	1.46 12.3a	1.48 0. 13.23a 7.2	59 21b	1.32 8.09b				
G9 LSD(0.05	2 3	20.72b G25 19.88a 0.86	24.18a G41 21.66b 0.75	18.16a G57 17.9a 0.57	25.33a G9 24.95a 0.67	12.88a G25 12.28a 1.22	13.89a G41 7.9 13.54a 7.4 0.78 0.	94a G57 48ab 56	9.27a 8.73ab 0.89				
G10	1 2 2	19.43c 21.66a G26	20.56c 22.43a G42	18.99a 19.64a G58	23.85b 24.55a G10	12.25a 12.28a G26	12.03a 6.8 14.26a G42 7.6	88a 61a G58	8.97a 9.64a				
LSD(0.05	3) 1	0.77 18.24c	0.79 18.37b	0.73 22.96b	24.24ab 0.82 24.62b	11.95a 1.45 12.25a	13.68a 7. 1.26 1. 7.22b 7.7	9a 19 76a	9.02a 1.21 14.73a				
G11	2 3	20.63a G27 19.52b	20.51a G43 19.67a	24.37a G59 23.45ab	25.37a G11 25.12a	12.8a G27 12.56a	8.61ab G43 8.3 7.93a 8.0	37a G59)3a 82	15.56a 14.88a				
G12) 1 2	14.67c 16.43a G28	20.55b 24.62a G44	22.51b 24.36a G60	24.55b 25.62a G12	7.67a 7.98a G28	12.34a 8.1 12.89a G44 9.5	17b 76a G60	8.11b 10.18a				
LSD(0.05	3) 1	15.88b 0.75 14.26c	21.58b 0.91 20.32b	23.78b 0.53 24.23b	25.23a 0.35 23.86c	7.73a 1.38 8.34b	12.57a 9.5 1.11 0. 8.39b 15	58a 96 662a	9.27a 1.05 8.33a				
G13	23	16.68a G29 15.32b	22.44a G45 20.74b	25.3a G61 24.87b	25.57a G13 24.88b	9.16a G29 9.4a	9.15a G45 17 8.44b 17	.34a G61 .11a	9.45a 8.96a				
G14) 1 2	0.64 19.88a 19.72a G30	0.67 19.62b 23.53a G46	0.57 24.17b 25.43a G62	0.38 22.76b 23.94a G14	0.79 12.48a 12.85a G30	0.62 2. 7.97b 15 8.83a G46 16	64 .36a .81a G62	1.34 7.81a 8.44a				
LSD(0.05	3)	19.98a 0.82 18.64b	20.66b 0.86 20.86b	25.08a 0.64 24.54b	24.26a 0.99 22.17b	12.81a 1.27 11.92a	8.31ab 16 0.64 1. 7.25b 15	.16a 92	7.82a 1.71				
G15	2 3	19.56a G31 19.08ab	20.865 22.23a G47 21.38b	25.73a G63 25.18a	24.36a G15 23.95a	12.93a G31 12.17a	7.550 15 8.06a G47 16 7.83ab 16	.85a G63 .52a	9.14a 9.02a				
LSD(0.05) $1 \\ 2$	0.93 19.55b 20.45a C32	0.71 20.18b 21.36a - C48	0.74 24.18b 25.32a C64	0.83 23.65b 24.57a C16	1.22 12.23a 12.58a C32	0.54 2. 6.15b 14 7.76a C48 15	22 .8a .52a C64	1.45 8.48b 10.46a				
LSD(0.05	3)	20.13ab 0.63	20.57b 0.62	24.67ab 1.01	24.21ab 0.73	12.37a 1.72	7.11ab 15 1.04 0.	.26a 77	10.23a 0.77				

Table 9. 1000-grain weight and yield per plant of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

In each column, means with similar letter (s) are statistically identical, and those with dissimilar letter (s) differ significantly as per 0.05 level of probability, $T_1 = 5 \text{ t ha}^{-1} \text{ CM}$, $T_2 = 2.5 \text{ t ha}^{-1} + 50\%$ CFRR, $T_3 = 100\%$ CFRR, SP = number of spikelet per panicle, and PFG = percent filled grains.

3.4. Nutrient Content

3.4.1. Nutrient Content (% N) of Grain and Straw

There was a significant effect on N contents in rice grain and straw among the genotype and treatment. Results indicated that the application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded on N content in grain on genotype Pukhi (G1) (1.40%) followed by Utri (G4) (1.39%), Panbira (G2) (1.36%), Kaisa panja (G6) (1.34%), Dumai (G11) (1.32%), Nayan moni (G22) (1.31%), and Vandana (G7) (1.29%), respectively, were significantly higher than the other treatments. In addition, the application of T_1 (5 t ha⁻¹ CM) recorded lower N content on genotype BRRI dhan28 (G54) (0.89%), BRRI dhan55 (G60) (0.91%), BRRI dhan42 (G56) (0.92%), B370 (G62) (0.93%), and NMR 151 (G43) (0.94%), respectively, were presented in Table 10. On the other hand, the application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded on N content in straw on genotype Utri (G4) (1.20%) followed by Panbira (G2) (1.19%), Pukhi (G1) (1.18%), Kaisa panja (G6) (1.17%), Dumai (G11) (1.15%), Luanga (G5) (1.14%), and Nayan moni (G22) (1.13%), respectively, were significantly higher than the other treatments, while the application of T_1 (5 t ha⁻¹ CM) recorded lower N content on genotype GHUA (G36) (0.81%), BRRI dhan28 (G54) (0.82%), BRRI dhan55 (G60) (0.83%), and B370 (G62) (0.84%), respectively, in Table 10.

3.4.2. Nutrient Content (% P) of Grain and Straw

There was a significant effect on P contents in rice grain and straw among the genotype and treatment. Results indicated that the application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded on P content in grain on genotype BRRI dhan72 (G53) (0.40%) followed by Vandana (G7) (0.38%), Pukhi (G1) (0.37%), Dharial (G3) (0.36%), Kaisa panja (G6) (0.35%), Utri (G4) (0.34%), and Dular (G8) (0.33%), respectively, were significantly higher than the other treatments. In addition, the application of T_1 (5 t ha⁻¹ CM) recorded lower P content on genotype BINASAIL (G63) (0.19%), MGAWA (G38) (0.20%), BANGKUL (G42) (0.21%), B370 (G62) (0.22%), BRRI dhan28 (G54) (0.23%), respectively, were presented in Table 11. On the other hand, the application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded on P content in straw on genotype Dharial (G3) (0.25%) followed by BRRI dhan72 (G53) (0.24%), Pukhi (G1) (0.23%), Kaisa panja (G6) (0.22%), Dharial (G3) (0.21%), Luanga (G5) (0.20%), and Kaisa panja (G6) (0.19%), respectively, were significantly higher than the other treatments, while the application of T_1 (5 t ha⁻¹ CM) recorded lower P content on genotype Sonhamoni (G9) (0.10%), BRRI dhan28 (G54) (0.11%), BRRI dhan42 (G56) (0.11%), and B370 (G62) (0.12%), respectively, in Table 11.

3.4.3. Nutrient Content (% K) of Grain and Straw

There was a significant effect on K contents in rice grain and straw among the genotype and treatment. Results indicated that the application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded on K content in grain on genotype Putra 2 (G47) (0.19%) followed by MR297(G45) (0.18%), BRRI dhan39 (G55) (0.17%), BRRI dhan75 (G59) (0.16%), and Pukhi (G1) (0.15%), respectively, were significantly higher than the other treatments, while application of T_1 (5 t ha⁻¹ CM) recorded lower P content on genotype BANGKUL (G42) (0.07%), B370 (G62) (0.08%), BRRI dhan55 (G60) (0.09%), BINASAIL (G63) (0.10%), and BRRI dhan39 (G55) (0.11%), respectively, were presented in Table 12. On the other hand, the application of T_2 (2.5 t ha⁻¹ CM + 50% CFRR) recorded on K content in straw on genotype Pukhil (G1) (1.84%) followed by Utri (G4) (1.81%), Panbira (G2) (1.79%), Vandana (G7) (1.78%), Parija (G12) (1.77%), Dharial (G3) (1.76%), and Kaisa panja (G6) (1.74%), respectively, were significantly higher than the other treatments, while the application of T_1 (5 t ha⁻¹ CM) recorded lower K content on genotype B370 (G62) (1.25%), BINASAIL (G63) (1.27%), BRRI dhan55 (G60) (1.28%), and BRRI dhan28 (G54) (1.29%), respectively, in Table 12.

Genotype	Treatment				Grain				Straw									
(G) *	(T)				% N								% N					
	1	1.12c	C17	1.05c	C^{22}	0.97c	C 40	0.96c	C1	1.03b	C17	0.88c	C22	0.85b	C40	0.85c		
GI	3	1.40a 1.26b	GI/	1.27a 1 16b	Goo	1.20a 1.09b	G49	1.22a 1 11h	GI	1.10a 1.06b	GI/	0.93h	Goo	0.88b	G49	0.90a 0.90b		
LSD	(0.05)	0.034		0.028		0.022		0.02		0.035		0.031		0.034		0.031		
	1	1.10c		1.04c		1.04c		0.92c		1.02b		0.85c		0.85c		0.83b		
G2	2	1.36a	G18	1.23a	G34	1.23a	G50	1.15a	G2	1.19a	G18	1.06a	G34	1.01a	G50	0.92a		
ISD	3	1.230		1.14b		1.120		1.076		1.050		0.920		0.900		0.850		
LSD	(0.03)	1.04c		1.06c		0.004 0.99a		0.025 0.95c		0.055 0.95b		0.052 0.85c		0.020		0.020 0.86c		
G3	2	1.25a	G19	1.28a	G35	1.21a	G51	1.16a	G3	1.11a	G19	1.09a	G35	0.99a	G51	0.94a		
	3	1.13b		1.16b		1.10b		1.07b		0.97b		0.94b		0.88b		0.91b		
LSD	(0.05)	0.03		0.027		0.032		0.034		0.029		0.025		0.025		0.028		
C4	1	1.11C 1.20a	C^{20}	1.06C	C26	1.03C	CED	0.93C	C_{1}	0.99C	C20	0.83C	C26	0.810	CED	0.84C		
G4	3	1.21b	G20	1.20a 1.14b	G30	1.29a	G52	1.07b	G4	1.03b	G20	0.91b	G30	0.96a	G52	0.97a		
LSD	(0.05)	0.032		0.025		0.037		0.025		0.031		0.028		0.028		0.026		
	1	1.09c		1.07c		1.03c		0.97c		0.95b		0.84c		0.89b		0.86c		
G5	2	1.28a	G21	1.24a	G37	1.25a	G53	1.22a	G5	1.14a	G21	1.06a	G37	1.03a	G53	0.99a		
ISD	(0.05)	1.190		1.150		1.14b		1.100		0.980		0.976		0.910		0.900		
LSD	(0.03)	1.10c		1.034		1.054		0.023		0.029 0.97h		0.03 0.95h		0.034 0.88h		0.02 0.82b		
G6	2	1.34a	G22	1.31a	G38	1.22a	G54	1.15a	G6	1.17a	G22	1.13a	G38	1.01a	G54	0.93a		
	3	1.22b		1.20b		1.13b		1.06b		0.99b		0.98b		0.90b		0.85b		
LSD	(0.05)	0.038		0.027		0.025		0.027		0.031		0.041		0.028		0.031		
C7	1	1.10c	C^{22}	1.10c	C20	1.00c	CEE	1.02c	C7	0.95b	C^{22}	0.92b	C20	0.83c	CEE	0.94c		
G/	3	1.29a 1.18h	G23	1.29a 1.18h	G39	1.20a 1.11h	G55	1.20a 1.15h	G/	0.97b	G23	0.94h	G39	1.05a 0.89h	G55	1.05a 0.96h		
LSD	(0.05)	0.031		0.026		0.028		0.026		0.026		0.028		0.022		0.023		
	1	1.07c		1.04c		1.08c		0.92c		0.92c		0.92b		0.90b		0.85c		
G8	2	1.26a	G24	1.23a	G40	1.32a	G56	1.16a	G8	1.09a	G24	1.08a	G40	1.08a	G56	0.92a		
LCD	3	1.15b		1.12b		1.19b		1.05b		0.97b		0.96b		0.92b		0.896		
L5D	(0.05)	1.029		0.03 1.06c		1.054		0.03 0.93c		0.025 0.93c		0.02 0.89c		0.05		0.028		
G9	2	1.07c	G25	1.25a	G41	1.24a	G57	1.17a	G9	1.12a	G25	1.02a	G41	1.04b	G57	0.95a		
	3	1.19b		1.13b		1.12b		1.08b		1.01b		0.93b		0.94b		0.88b		
LSD	(0.05)	0.034		0.031		0.033		0.032		0.025		0.028		0.023		0.02		
C10	1	1.13c	C2 (1.04c	C 10	0.99c	050	0.96c	C10	1.01b	C2 (0.85c	C 10	0.82c	050	0.83c		
GIU	$\frac{2}{3}$	1.39a 1.25b	G26	1.20a 1.11b	G42	1.20a 1.11b	G58	1.19a 1.08b	G10	1.18a 1.03b	G26	1.01a 0.90b	G42	0.98a 0.94b	G58	0.95a 0.91h		
LSD	(0.05)	0.027		0.025		0.034		0.024		0.03		0.016		0.040		0.027		
	1	1.11c		1.02c		0.94c		1.00c		0.95c		0.83c		0.85b		0.91c		
G11	2	1.32a	G27	1.24a	G43	1.18a	G59	1.24a	G11	1.15a	G27	1.03a	G43	0.97a	G59	1.02a		
	3	1.21b		1.10b		1.07b		1.13b		0.99b		0.91b		0.86b		0.96b		
LSD	(0.05)	0.029		1.028		0.028		0.034		0.027 0.92h		0.025 0.91b		0.023 0.84c		0.022		
G12	2	1.00C	G28	1.05C	G44	1.17a	G60	1.17b	G12	01.10a	G28	1.06a	G44	0.96a	G60	0.95a		
012	3	1.13b	020	1.13b	011	1.06b	000	1.06b	012	0.95b	020	0.92b	011	0.87b	000	0.90b		
LSD	(0.05)	0.03		0.034		0.025		0.031		0.025		0.027		0.024		0.025		
610	1	1.10c	~~~	1.05c	.	0.96c	0.11	0.93c	610	0.94b	~	0.89c	.	0.86b	0.11	0.85c		
G13	2	1.36a	G29	1.29a	G45	1.21a	G61	1.17a 1.10b	G13	1.14a	G29	1.09a	G45	0.98a	G61	0.97a		
LSD	(0.05)	0.031		0.03		0.025		0.031		0.970		0.940		0.090		0.900		
LOD	1	1.07c		1.03c		0.94c		0.93c		0.90c		0.88c		0.85c		0.84c		
G14	2	1.28a	G30	1.26a	G46	1.20a	G62	1.14a	G14	1.09a	G30	1.11a	G46	0.97a	G62	0.98a		
	3	1.15b		1.17b		1.08b		1.05b		0.96b		0.92b		0.89b		0.91b		
LSD	(0.05)	0.023		0.024		0.029		0.021		0.025		0.02		0.028		0.018		
C1E	1	1.05C 1.25c	C21	1.04C	C 47	0.97C	C(2)	0.96C	C1E	0.88C	C21	0.86C	C_{47}	0.87C	C(2)	0.860		
G15	23	1.20a 1.13b	G31	1.25a 1 12b	G47	1.19a 1.08b	G63	1.10a 1.09b	GIS	0.93b	G31	0.91h	G47	0.95a 0.91h	G63	0.95a 0.89b		
LSD	(0.05)	0.034		0.03		0.031		0.031		0.028		0.027		0.023		0.028		
	1	1.09c		1.01c		0.99c		1.04c		0.92b		0.85c		0.89b		0.92c		
G16	2	1.32a	G32	1.21a	G48	1.23a	G64	1.22a	G16	1.12a	G32	1.01a	G48	1.02a	G64	1.02a		
ICD	3	1.20b		1.09b		1.10b		1.13b		0.95b		0.89b		0.92b		0.95b		
LSD	(0.05)	0.023		0.025		0.034		0.037		0.022		0.02		0.035		0.027		

Table 10. Grain and straw nutrient content (% N) of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

In each column, means with similar letter (s) are statistically identical, and those with dissimilar letter (s) differ significantly as per 0.05 level of probability, T1 = 5 t ha⁻¹ CM, T2 = 2.5 t ha⁻¹ + 50% CFRR, T3 = 100% CFRR, and % N = percentage of nitrogen content.

Genotype Tre	atment				Grain								Straw	7					
(G) ¹	(T)	% P								% P									
	1	0.25c		0.23c		0.21c		0.26b		0.12b		0.10c		0.13c		0.17b			
G1	2	0.37a	G17	0.31a	G33	0.33a	G49	0.33a	G1	0.23a	G17	0.20a	G33	0.22a	G49	0.21a			
I SD(0.05)	3	0.316		0.276		0.276		0.31a 0.024		0.20a 0.03		0.166		0.160		0.19ab 0.027			
L3D(0.03)	1	0.22c		0.22b		0.020 0.22c		0.26b		0.00 0.11b		0.020 0.13c		0.13b		0.16b			
G2	2	0.31a	G18	0.27a	G34	0.28a	G50	0.33a	G2	0.19a	G18	0.21a	G34	0.19a	G50	0.23a			
	3	0.27b		0.25a		0.25b		0.3a		0.17a		0.17b		0.15b		0.20a			
LSD(0.05)	1	0.028		0.023 0.21c		0.025 0.21h		0.026 0.27c		0.028 0.15c		0.025 0.11b		0.028 0.12c		0.027 0.17b			
G3	2	0.26c	G19	0.21c	G35	0.210 0.27a	G51	0.27C	G3	0.15c	G19	0.11D 0.19a	G35	0.12c	G51	0.17D 0.24a			
60	3	0.30b	017	0.26b	000	0.23b	001	0.31b	00	0.21b	017	0.17a	000	0.15b	001	0.19b			
LSD(0.05)	1	0.027		0.024		0.024		0.03		0.034		0.027		0.025		0.029			
C_{1}	1	0.23C 0.34a	C^{20}	0.22C 0.34a	C26	0.21b	CE2	0.23C	C_{1}	0.11C 0.21a	C20	0.12b 0.22a	C26	0.12b	C52	0.14c 0.22a			
64	3	0.28b	G20	0.29b	G30	0.24ab	G52	0.35a 0.27b	64	0.17b	G20	0.22a 0.20a	G30	0.17a 0.15a	G52	0.22a 0.17b			
LSD(0.05)		0.028		0.026		0.023		0.028		0.028		0.025		0.023		0.024			
~-	1	0.22c		0.25c	~~~	0.23b	~ ~ ~	0.27c	~-	0.11c	C2 1	0.14c	~~~	0.13b	~	0.16b			
G5	2	0.31a 0.28b	G21	0.35a 0.31h	G37	0.29a 0.26a	G53	0.40a 0.33b	G5	0.20a 0.16b	G21	0.23a 0.19b	G37	0.18a 0.15a	G53	0.24a 0.19b			
LSD(0.05)	5	0.025		0.027		0.204		0.031		0.029		0.027		0.029		0.190			
()	1	0.26c		0.24b		0.20c		0.23c		0.14b		0.13c		0.11b		0.15b			
G6	2	0.35a	G22	0.29a	G38	0.25a	G54	0.34a	G6	0.22a	G22	0.19a	G38	0.17a	G54	0.21a			
I SD(0.05)	3	0.31b		0.27a 0.025		0.236		0.276		0.19a 0.034		0.160		0.16a 0.016		0.176			
L3D(0.03)	1	0.051 0.26c		0.025 0.23b		0.02 0.25c		0.020 0.22c		0.13c		0.022 0.12b		0.13b		0.020 0.14b			
G7	2	0.38a	G23	0.27a	G39	0.33a	G55	0.30a	G7	0.25a	G23	0.18a	G39	0.21a	G55	0.19a			
	3	0.32b		0.25a		0.29b		0.26b		0.21b		0.14b		0.19a		0.16b			
LSD(0.05)	1	0.023 0.24c		0.024 0.22h		0.028 0.24b		0.03		0.031 0.11b		0.025		0.038 0.13b		0.025 0.11c			
G8	2	0.33a	G24	0.220 0.26a	G40	0.240 0.29a	G56	0.22c 0.29a	G8	0.11D 0.21a	G24	0.19a	G40	0.13D 0.18a	G56	0.11c			
	3	0.29b		0.24ab		0.26ab		0.25b		0.19a		0.14b		0.16a		0.14b			
LSD(0.05)	1	0.025		0.027		0.027		0.023		0.03		0.017		0.024		0.023			
C9	1	0.22c 0.29a	C^{25}	0.26C 0.32a	C_{41}	0.21C 0.28a	C57	0.24c 0.32a	C^{0}	0.10b 0.17a	C^{25}	0.150	C_{41}	0.12b 0.17a	C57	0.14b 0.20a			
97	3	0.26b	G25	0.29b	041	0.24b	657	0.27b	G)	0.17a 0.15a	625	0.17ab	041	0.14b	G57	0.16b			
LSD(0.05)		0.025		0.027		0.023		0.025		0.022		0.031		0.022		0.029			
C10	1	0.22c	COL	0.21c	C 10	0.21b	050	0.26c	C10	0.12c	COK	0.13c	C 10	0.12c	CEO	0.15b			
G10	2	0.31a 0.27b	G26	0.29a 0.26b	G42	0.27a 0.25a	G58	0.35a 0.31b	GIU	0.19a 0.16b	G26	0.19a 0.16b	G42	0.19a 0.15b	G58	0.21a 0.19a			
LSD(0.05)	0	0.031		0.016		0.022		0.03		0.028		0.023		0.022		0.034			
· · · · · · · · · · · · · · · · · · ·	1	0.23b		0.22b		0.26c		0.25c		0.12b		0.11c		0.16b		0.15b			
G11	2	0.29a	G27	0.27a	G43	0.34a	G59	0.37a	G11	0.18a	G27	0.19a	G43	0.22a	G59	0.24a			
LSD(0.05)	3	0.27a 0.025		0.204		0.031		0.028		0.16a		0.025		0.029		0.180			
202 (0100)	1	0.24c		0.24b		0.24c		0.22c		0.11c		0.14b		0.14c		0.12c			
G12	2	0.32a	G28	0.33a	G44	0.35a	G60	0.31a	G12	0.20a	G28	0.21a	G44	0.22a	G60	0.20a			
	3	0.28b		0.30a		0.30b		0.26b		0.16b		0.17b		0.18b		0.17b			
L3D(0.03)	1	0.028 0.23c		0.031 0.25c		0.028 0.22c		0.025 0.22c		0.032 0.13c		0.03 0.15c		0.031 0.13b		0.025 0.12b			
G13	2	0.31a	G29	0.36a	G45	0.33a	G61	0.28a	G13	0.19a	G29	0.24a	G45	0.19a	G61	0.18a			
	3	0.28b		0.31b		0.27b		0.25b		0.15b		0.19b		0.17a		0.16a			
LSD(0.05)	1	0.023		0.028 0.27b		0.028		0.018 0.22h		0.028		0.027 0.16b		0.027 0.12b		0.031 0.12h			
G14	2	0.21C 0.28a	G30	0.270 0.36a	G46	0.21c 0.31a	G62	0.220 0.26a	G14	0.2C 0.18a	G30	0.10D 0.23a	G46	0.120 0.18a	G62	0.120 0.17a			
	3	0.23b	200	0.30ab	0.10	0.26b	202	0.23b	011	0.15b	200	0.18b	0.10	0.16a	202	0.14b			
LSD(0.05)	1	0.022		0.029		0.029		0.027		0.024		0.03		0.028		0.023			
C15	1	0.23c 0.30a	C^{21}	0.21c 0.26a	C^{47}	0.250	C^{42}	0.19c 0.27a	$C1^{r}$	0.11b 0.18a	C^{21}	0.12b 0.17a	C^{47}	0.17b 0.21a	C62	0.11C 0.185			
GIJ	$\frac{1}{3}$	0.27b	631	0.24b	64/	0.29b	605	0.24b	G10	0.16a	G31	0.17a 0.15a	64/	0.18ab	605	0.14b			
LSD(0.05)		0.016		0.023		0.023		0.024		0.024		0.026		0.029		0.016			
C1(1	0.22b	C 22	0.23c	C 40	0.24c	CA	0.24b	01/	0.12b	C00	0.13b	C 40	0.16b	CCA	0.13b			
G16	∠ 3	0.27a 0.24ab	G32	0.31a 0.26h	G48	0.32a 0.28h	G64	0.30a 0.26h	G16	0.19a 0.14h	G32	0.20a 0.16b	G48	0.20a 0.18ab	G64	0.19a 0.14h			
LSD(0.05)	5	0.028		0.024		0.025		0.027		0.023		0.024		0.025		0.025			

Table 11. Grain and straw nutrient content (% P) of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

In each column, means with similar letter (s) are statistically identical, and those with dissimilar letter (s) differ significantly as per 0.05 level of probability, $T_1 = 5 \text{ t ha}^{-1} \text{ CM}$, $T_2 = 2.5 \text{ t ha}^{-1} + 50\%$ CFRR, $T_3 = 100\%$ CFRR, and % P = percentage of phosphorus content.

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Cenotype	Freatment				Grain								Straw	7				
(G)	(T)	% K																
	1	0.111		0.001-	70 K	0.001-		0.101		1 51 -		1 41 -	70 K	1 20 -		1.00 -		
G1	$\frac{1}{2}$	0.11b 0.17a	G17	0.09b 0.16a	G33	0.09b 0.14a	C49	0.10b 0.16a	G1	1.51c 1.84a	G17	1.41c 1.76a	C33	1.39C	G49	1.32c 1.54a		
01	3	0.15a	017	0.15a	000	0.13a	01/	0.13b	GI	1.70b	017	1.54b	000	1.49b	01/	1.40b		
LSD(0.0)5)	0.020		0.025		0.023		0.030		0.044		0.035		0.034		0.033		
	1	0.10c		0.09c		0.08b		0.11c		1.47c		1.40c		1.37c		1.30c		
G2	2	0.16a	G18	0.16a	G34	0.16a	G50	0.19a	G2	1.79a	G18	1.74a	G34	1.71a	G50	1.59a		
	3	0.130		0.130		0.14a		0.160		1.680		1.530		1.540		1.42b		
L3D(0.0	1	0.020 0.12h		0.020 0.08c		0.023 0.09c		0.028 0.09b		1.052		1.37c		1.34c		1.31c		
G3	2	0.120 0.16a	G19	0.15a	G35	0.16a	G51	0.16a	G3	1.76a	G19	1.78a	G35	1.72a	G51	1.57a		
	3	0.14a		0.11b		0.13b		0.15a		1.63b		1.56b		1.53b		1.43b		
LSD(0.0)5)	0.023		0.022		0.011		0.020		0.040		0.034		0.027		0.034		
<u>C1</u>	1	0.12b	C2 0	0.09c	<u> </u>	0.10c	050	0.11b	C 1	1.46c	C2 0	1.34c	COL	1.38c	050	1.28c		
G4	2	0.15a 0.14ab	G20	0.17a 0.14b	G36	0.16a 0.14b	G52	0.16a 0.15a	G4	1.81a 1.70b	G20	1.70a 1.47h	G36	1.69a 1.50b	G52	1.40a 1.40b		
LSD(0))5)	0.025		0.020		0.023		0.031		0.041		0.028		0.038		0.033		
202 (0.	1	0.10b		0.09c		0.11c		0.10c		1.38c		1.38c		1.35c		1.31c		
G5	2	0.15a	G21	0.14a	G37	0.17a	G53	0.19a	G5	1.70a	G21	1.72a	G37	1.65a	G53	1.60a		
	3	0.13b		0.11b		0.15b		0.16b		1.59b		1.54b		1.51b		1.42b		
LSD(0.0	J5) 1	0.027		0.021		0.022		0.025		0.032		0.033		0.026		0.035		
C6	2	0.09C	C^{22}	0.100	C38	0.090 0.14a	C54	0.120	C6	1.400 1 74a	C^{22}	1.57C 1.74a	C38	1.55C	C54	1.29C		
90	3	0.13u 0.12b	622	0.13b	G30	0.11a 0.12a	654	0.14b	Gu	1.62b	622	1.53b	G30	1.49b	654	1.37b		
LSD(0.0)5)	0.025		0.028		0.025		0.03		0.032		0.030		0.041		0.037		
	1	0.09c		0.10b		0.11b		0.11a		1.42c		1.35c		1.39c		1.28c		
G7	2	0.13a	G23	0.14a	G39	0.15a	G55	0.17a	G7	1.78a	G23	1.70a	G39	1.72a	G55	1.50a		
I SD(0)	3	0.110		0.12ab		0.14a 0.025		0.130		1.630		1.520		1.510		1.360		
L3D(0.0	1	0.021 0.11c		0.025 0.11b		0.025 0.12b		0.021 0.13b		1.36c		1.37b		1.36c		1.27c		
G8	2	0.17a	G24	0.17a	G40	0.16a	G56	0.18a	G8	1.66a	G24	1.61a	G40	1.64a	G56	1.44a		
	3	0.14b		0.15a		0.15a		0.15ab		1.52b		1.48b		1.47b		1.34b		
LSD(0.0)5)	0.023		0.023		0.020		0.027		0.041		0.022		0.034		0.031		
CO	1	0.086	COL	0.10b	C 41	0.096	CET	0.10b	CO	1.44C	COF	1.41c	C 41	1.37c	CEZ	1.25c		
G9	23	0.10a 0.14a	G25	0.15a 0.14a	G41	0.15a 0.12a	G37	0.17a 0.15a	G9	1.71a 159h	G25	1.72a 1.50b	G41	1.09a 1.48h	G3/	1.40a 1.34h		
LSD(0.0)5)	0.025		0.022		0.021		0.023		0.034		0.034		0.029		0.035		
,	´ 1	0.08c		0.08b		0.07b		0.12b		1.40c		1.34c		1.34c		1.27c		
G10	2	0.15a	G26	0.13a	G42	0.12a	G58	0.17a	G10	1.79a	G26	1.65a	G42	1.71a	G58	1.50a		
	3	0.12b		0.12ab		0.11a		0.14b		1.66b		1.44b		1.45b		1.35b		
L5D(0.0	1	0.017 0.10b		0.021 0.09c		0.020 0.11b		0.028 0.11c		1.042 1.43c		1.025 1.34c		1.030		1.034		
G11	2	0.100 0.14a	G27	0.15a	G43	0.110 0.15a	G59	0.19a	G11	1.78a	G27	1.62a	G43	1.55a	G59	1.56a		
011	3	0.11b	.	0.12b	010	0.15a	007	0.16b	011	1.63b	01	1.41b	010	1.46b	007	1.38b		
LSD(0.0)5)	0.020		0.016		0.025		0.024		0.027		0.036		0.038		0.035		
C10	1	0.11b	C2 0	0.09c	C14	0.11c	C(0	0.09c	C10	1.40c	C2 0	1.37c	CAA	1.36c	<i>C</i> (0	1.28c		
GIZ	2	0.16a 0.13b	G28	0.16a 0.14b	G44	0.16a 0.14b	G60	0.16a 0.12b	GIZ	1.77a 1.61h	G28	1.70a 1.49h	G44	1.55a 1.44b	G60	1.4/a 1.34h		
LSD(0.0)5)	0.130		0.021		0.022		0.020		0.030		0.037		0.035		0.035		
	1	0.09c		0.08c		0.12c		0.11b		1.36c		1.39c		1.32c		1.28c		
G13	2	0.15a	G29	0.15a	G45	0.18a	G61	0.16a	G13	1.72a	G29	1.68a	G45	1.56a	G61	1.51a		
	3	0.12b		0.12b		0.15b		0.13ab		1.55b		1.50b		1.45b		1.35b		
LSD(0.0	J5) 1	0.016 0.00b		0.023		0.023		0.027 0.08b		1.034		0.036		1.034		1.250		
C14	2	0.090	C30	0.000 0.13a	C46	0.060	C62	0.000 0.14a	C14	1.57C	C30	1.54C	C46	1.55C	C62	1.25C		
011	3	0.17a	000	0.12ab	010	0.14a	002	0.11b	017	1.51b	0.50	1.45b	010	1.47b	002	1.33b		
LSD(0.0)5)	0.024		0.020		0.024		0.025		0.033		0.029		0.028		0.037		
-	1	0.08b		0.11b		0.11c		0.10c		1.34c		1.35c		1.34c		1.27c		
G15	2	0.14a	G31	0.16a	G47	0.19a	G63	0.14a	G15	1.65a	G31	1.54a	G47	1.59a 1.42h	G63	1.43a 1.25b		
I SD(0))5)	0.15a		0.14a		0.150		0.120		0.035		0.031		1.420 0.031		0.030		
	1	0.10b		0.020 0.09c		0.12b		0.10b		1.43c		1.35c		1.35c		1.29c		
G16	2	0.15a	G32	0.15a	G48	0.18a	G64	0.15a	G16	1.79a	G32	1.66a	G48	1.55a	G64	1.52a		
	3	0.14a		0.11b		0.14b		0.13a		1.62b		1.54b		1.39b		1.40b		
LSD(0.0	15)	0.023		0.022		0.028		0.025		0.034		0.027		0.035		0.031		

Table 12. Grain and straw nutrient content (% K) of 64 rice genotypes as influenced by treatment and genotype (pooled over two seasons).

In each column, means with similar letter (s) are statistically identical, and those with dissimilar letter (s) differ significantly as per 0.05 level of probability, $T1 = 5 \text{ t ha}^{-1} \text{ CM}$, $T2 = 2.5 \text{ t ha}^{-1} + 50\%$ CFRR, T3 = 100% CFRR, and % k = percentage of potassium content.

4. Discussion

The results of this experiment revealed that there is a high correlation between plant height and rice plant productivity or growth rate. During the developing stages of rice plants, they grow and flourish to a specific height [24]. Phenotype refers to the process of measuring the basic and complicated traits of a rice species, which include plant height. Organic fertilizer has a positive impact on the growth and production of various crops [25,26]. Plant height is also a major agronomic characteristic that indirectly affects rice plant yields. Traditionally, rice genotypes are tall in stature, are susceptible to loading at maturity, respond poorly to nitrogen fertilizer, and, therefore, produce low yields. For high-yielding varieties, moderate plant heights are desirable. Approximately half of the recommended chemical fertilizer is saved, according to the findings of this study. It differs from the findings of Chandini et al. [13], who discovered that organic fertilizers could potentially replace 50% of needed nitrogen and phosphorus fertilizers by improving. They examined the efficacy of suggested nitrogen and phosphorus fertilizers and lowering chemical fertilizer costs while also preventing environmental contamination from widespread use. Chemical N and P application can be reduced by 50%, while rice yield is boosted with the addition of 5 t ha⁻¹ organic fertilizer [27]. However, when organic fertilizers were used in conjunction with a half dose of inorganic fertilizer on lettuce (*Lactuca sativa*), twenty-five percent (25%) more growth was achieved than when only chemical fertilizer was used, and at least fifty percent (50%) of chemical fertilizer was saved by using organic fertilizer [28]. The enhanced vegetative growth and additional nitrogen contribution that occurs in response to the recommended fertilizer dose could be the primary reason for the increase in plant height [29]. The availability of main nutrients was equated to the variance in plant height caused by nutrient sources. Chemical fertilizers provide nutrients that are easily soluble in soil solutions and hence available to plants almost immediately. Microbial action and increased soil physical condition contribute to nutrient availability from organic sources. Bargaz et al. [30] agreed with these conclusions. Variations in the availability of key nutrients were thought to be the cause of plant height variation caused by nutrition source. Setiawati et al. [31] reported similar results in rice crops.

Tillering is a crucial feature for grain production and, as a result, a significant factor in rice output. Siavoshi et al. [32] found that different fertilizer mixes increased the number of tillers in rice plants. According to them, the increased number of tillers per square meter could be related to increased nitrogen availability, which is important for cell division. Organic sources provide plants with a better balanced diet, particularly micronutrients, which have a good impact on the number of tillers in plants [33]. The number of productive tillers (tillers that carry panicles) is more important than the overall number of tillers in determining rice plant productivity. The considerable difference in the number of tiller and panicle plant⁻¹ seen in this study can be attributed to genetic differences in their ability to use fertilizers, partition photosynthesis, and accumulate dry matter. The number of panicles grew with increasing nitrogen rates [34,35], and the number of panicles plant⁻¹ increased with increasing NPK rates. Organic manure and chemical fertilizers produced the most prolific tillers, which could be attributed to the nutrient availability in the soil. The availability of nutrients from organic sources, on the other hand, is attributed to microbial action and improved soil physical conditions. The excessive application of inorganic fertilizers is not required to generate good tillers if organic manures are supplemented, which also helps to provide vital micronutrients to the plants [36,37]. In rice crops, Mirza et al. [38] found similar findings.

Rice genotypes differed considerably in panicle length and grain yield. These findings are thought to be attributable to the rice plant receiving extra nutrients as a result of the soil amendment. The application of organic manure and chemical fertilizers resulted in a considerable increase in panicle length [1]. Similar findings were reported by [39,40].

In comparison to fertilizers, manure had a stronger effect in increasing the quantity of grains panicle⁻¹. It is possible that this owes to the manure's higher nutrient availability. The application of organic materials as fertilizers provided growth-regulating substances

that helped better grain filling and improved the physical, chemical, and microbial properties of the soil in this study, and the organic manure and chemical fertilizer had a significant effect because the application of organic materials as fertilizers provides growth-regulating substances that helped better grain filling and improved the physical, chemical, and microbial properties of the soil [41]. The use of organic manures and chemical fertilizers resulted in a considerable increase in grains per panicle [12]. These findings are also supported by Iqbal et al. [42].

The combined application of organic manure and artificial fertilizer resulted in statistically significant change in the weight of 1000 seeds. The combined use of organic manure and artificial fertilizers enhanced the 1000-grain weight of rice [43]. The use of organic manure and artificial fertilizer enhanced the 1000-grain weight of rice [44]. Hoque et al. [45] also found that combining organic manure with chemical fertilizers improved grain weight by 1000 grains. Geng et al. [46] reported that the availability of nutrients throughout the reproductive stage resulted in improved grain filling and thus increased grain weight.

The addition of organic manure to chemical fertilizers enhanced grain output significantly in all genotypes. This was due to the effect of organic and chemical fertilizers on encouraging growth and, as a result, increasing yields. The various fertilizers aided tiller growth and helped spikelet formation, resulting in a higher yield. Wang et al. [34] supported these findings. The fact that it improves soil quality, soil health, and crop output could explain this. The observations of [10] backed up this theory. It was demonstrated that applying organic manure can boost photosynthetic efficiency and nutrient availability [9]. Ye et al. [47] suggested the use of organic manure and chemical fertilizers enhanced grain output considerably. Organic manure and chemical fertilizers boosted rice straw yields [48]. These assumptions are supported by [40,49]. Increasing cropping intensity, the use of modern varieties (high-yielding varieties and hybrids), cultivation of high-biomass-potential crops, nutrient leaching, and unbalanced fertilizer application, with no or little addition of organic manure, have resulted in nutrient mining from the soils. To stop nutrient mining, it is not justified to increase the use of only inorganic fertilizers, but the use of organic sources of plant nutrients viz. cow dung, chicken manure, compost, and green manure should be also considered. In this study, nutrient contents of grain and straw of all genotypes showed that the highest N, P, and K contents were recorded in T_2 (2.5 t ha⁻¹ CM+ 50% CFRR). These findings are partially similar to these of [50,51], who obtained higher contents of nutrient elements such as N, P, and K in rice by applying chicken manure with inorganic fertilizers. The use of a combination of organic manures and inorganic fertilizers clearly aided plant vegetative growth, resulting in higher straw yield.

5. Conclusions

From the above results, it may be concluded that organic fertilizers in the form of chicken manure have the potential to increase the growth parameters, yield components, and nutritional quality of rice. The use of chicken manure as an organic fertilizer for rice also had positive effects on growth, yield, and nutrient content in the crop. All of the treatments had a significant impact on rice genotypes growth and production. In the current study, it was discovered that 2.5 tons of chicken manure per hectare, combined with 50% of the prescribed chemical fertilizer, resulted in a higher grain yield than the other treatments. From a financial standpoint, producers can employ a combination of organic fertilizer and a lower amount of inorganic fertilizer to increase rice yields while also maintaining and improving soil health.

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