


Article

NLR Genes Related Transcript Sets in Potato Cultivars Bearing Genetic Material of Wild Mexican Solanum Species

Alex V. Kochetov ^{1,2,3,*}, Dmitry A. Afonnikov ^{1,2} , Nikolay Shmakov ¹, Gennady V. Vasiliev ¹, Olga Y. Antonova ⁴, Natalja V. Shatskaya ¹, Anastasiya Y. Glagoleva ¹, Salmaz M. Ibragimova ¹, Aleksander Khiutti ⁵, Olga S. Afanasenko ⁵ and Tatjana A. Gavrilenko ⁴

¹ Institute of Cytology & Genetics, 630090 Novosibirsk, Russia; ada@bionet.nsc.ru (D.A.A.); shmakov@bionet.nsc.ru (N.S.); genn@bionet.nsc.ru (G.V.V.); shatskaya@bionet.nsc.ru (N.V.S.); glagoleva@bionet.nsc.ru (A.Y.G.); isola@bionet.nsc.ru (S.M.I.)

² Faculty of Natural Sciences, Novosibirsk State University, 630090 Novosibirsk, Russia

³ Novosibirsk State Agrarian University, 630039 Novosibirsk, Russia

⁴ N.I. Vavilov All-Russian Institute of Plant Genetic Resources (VIR), 190000 Saint Petersburg, Russia; olgaant326@mail.ru (O.Y.A.); tatjana9972@yandex.ru (T.A.G.)

⁵ All Russian Institute of Plant Protection, 196608 Saint Petersburg, Russia; alexanderkhyutti@mail.ru (A.K.); olga.s.afan@gmail.com (O.S.A.)

* Correspondence: ak@bionet.nsc.ru



Citation: Kochetov, A.V.; Afonnikov, D.A.; Shmakov, N.; Vasiliev, G.V.; Antonova, O.Y.; Shatskaya, N.V.; Glagoleva, A.Y.; Ibragimova, S.M.; Khiutti, A.; Afanasenko, O.S.; Gavrilenko, T.A. NLR Genes Related Transcript Sets in Potato Cultivars Bearing Genetic Material of Wild Mexican Solanum Species. *Agronomy* **2021**, *11*, 2426. <https://doi.org/10.3390/agronomy11122426>

Academic Editor: Rafael A. Cañas

Received: 11 November 2021

Accepted: 25 November 2021

Published: 28 November 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: The long history of potato breeding includes the numerous introgressions of resistance genes from many wild species of South and Central America as well as from cultivated species into the breeding gene pool. Most R genes belong to the NLR family with nucleotide-binding site–leucine-rich repeat. The aim of this research concerns an evaluation of NLR genes expression in transcriptomes of three potato cultivars (Evraziya, Siverskij, Sudarynya), which combine genetic material from wild and cultivated potato species, and each bears intragenic markers of *RB/Rpi-blb1/Rpi-sto1* genes conferring broad-range resistance to late blight. The transcriptomes of the cultivars were compared before and 24 h after the *Phytophthora infestans* inoculation. The induction of *RB/Rpi-blb1/Rpi-sto1* transcript after 24 h of inoculation was detected in the resistant cultivars Siverskij and Sudarynya but not in susceptible cv. Evraziya. This demonstrates the importance of transcriptomic assay for understanding the results of marker-assisted selection and phenotyping. Interestingly, assembling the transcriptomes de novo and analysis with NLR-parser tool revealed significant fractions of novel NLR genes with no homology to the reference genome from 103 (cv. Siverskij) to 160 (*S. stoloniferum*, 30514/15). Comparison of novel NLRs demonstrated a relatively small intersection between the genotypes that coincided with their complex pedigrees with several interspecific hybridization events. These novel NLRs may facilitate the discovery of new efficient R genes.

Keywords: potato cultivars; NLR genes; *P. infestans*; novel genes; introgression

1. Introduction

Marker-assisted selection (MAS) is efficiently exploited in selecting desirable genotypes with resistance genes (R genes) and stacking several R genes to facilitate the breeding of new cultivars with durable resistance [1–3]. MAS can increase the efficiency of conventional breeding, and it is applicable for traits conferred by both major qualitative genes and quantitative trait loci (QTLs). Occasionally, MAS can be less efficient (or predictable) because of the insufficient linkage between a marker and a gene (QTL), the complexity of the genetic control of a resistance trait, or the instability of QTL effects. In addition, in many cases the genetic factors providing resistance against a specific pathogen remains unidentified.

Most R genes belong to NLR family encoding proteins with nucleotide-binding site–leucine-rich repeat (NBS-LRR, about 80% of cloned genes [4]). Some classes of R genes and their analogs (RGAs) may be predicted by bioinformatics tools [5–9].

Various approaches are used for identification of novel efficient R genes and their introgression. The resequencing of genotype collections revealed a number of NBS-LRR-candidate genes [10–12]. Selective sequencing of NLR genes segments was proposed as an efficient tool for their characterization (for example, RenSeq [13]). Comparison of syntenic genomic regions is a promising way to locate candidate R genes in the genomes of various crops [14,15]. Transcriptomic analysis is critically important for the reconstruction of molecular mechanisms of resistance to various pathogens (e.g., [16–18]). An integration of genomic and transcriptomic data may be useful for resolving complicated cases. For instance, ascochyta blight resistance genes are located close to nine QTLs in the chickpea genome region with about 30 NLR genes, and comparison of their transcription patterns was useful for selecting the candidates [19]. Further verification of candidate R genes may include co-segregation analysis, effectoromics approaches [20], dsRNA-mediated suppression of a candidate gene in a resistant plant [21], overexpression of a candidate gene in a susceptible plant [22,23], etc.

In general, the isolation of a major R gene from a pool of RGAs is hampered by their natural variability: the genomes of cultivated plants commonly contain clusters with many duplicated and recombined RGAs with highly similar structures [24,25]. It is known that clusters of the R genes in plant genomes are rapidly evolving with respect to copy number variation and SNP accumulation [11,24]. The natural variability of NLR gene clusters mainly results from co-evolution with pathogens and their effectors. However, it was also found that genetic incompatibility contributes to this process. The mechanism of NLR-mediated resistance often involves local induction of programmed cell death at a site of pathogen invasion. In turn, the genetic incompatibility frequently results from hybrid necrosis. Systemic analysis of F1 hybrids of *Arabidopsis thaliana* L. strains demonstrated that incompatibility hot spots were frequently associated with genome regions containing NLR genes [26]. It was reported that a semi-dominant NLR allele with a single amino acid substitution causes the necrotic death of the entire seedling of wheat [27], and a DM10 allele of the TIR-NLR gene in *A. thaliana* causes hybrid necrosis at the cotyledon stage [28]. Interspecific hybridization can also be prevented by NLR incompatibility and hybrid necrosis. It was found that the seedling lethality of the *Nicotiana tabacum* L. × *N. africana* Merxm. interspecific cross was associated with the CC-NBS-LRR *Nt6549g30* gene [29], and the CC-NBS-LRR gene is the causal gene underlying the Le4 locus for interspecific hybrid lethality between *Gossypium barbadense* L. and *G. hirsutum* L. [30].

NLR genes trigger autoimmunity and cell death; thus, their expression should be strictly controlled. In addition to transcriptional control, molecular mechanisms of resistance involve various non-coding RNAs. In tomatoes, SI-lncRNA15492 interacts with SI-miR482a and affects *Solanum lycopersicum* L. resistance to *Phytophthora infestans* (Mont.) de Bary. SI-miR482a targets SI-NBS-LRR genes that may be needed as a negative regulator of NBS-LRR gene expression. In turn, SI-lncRNA15492 inhibits SI-miR482a and maintains homeostasis during the interaction between tomato and *P. infestans* [31]. The silencing of lncRNA23468 in tomatoes led to increased accumulation of miR482b and decreased accumulation of NBS-LRRs, as well as reduced plant resistance to *P. infestans* [32]. Inactivation of miR482b and miR482c, with the aid of gene editing, results in enhanced tomato resistance to *P. infestans* [33]. There may be some other mechanisms supplementing regulation by transcriptional factors and non-coding RNAs. For example, the RNA-binding protein FPA mediates widespread premature cleavage and polyadenylation of NLR transcripts, abolishing their expression [34]. These examples demonstrate the complexity of molecular mechanisms where transcriptional control is only a part of integral multilayer system.

The number and content of NLR genes may vary in different commercial genotypes and differ from the reference genomes. For example, quantitative trait loci in Spanish barley landrace on the long arm of chromosome 7H, which provides resistance against powdery mildew, contains a cluster of NLR genes but these genes are absent in the reference barley genome [35]. A comparison of 64 *A. thaliana* genomes revealed groups of well-conserved NLR clades that were missing from the reference Columbia-0 genome [36].

Systemic comparison of 1301 *A. thaliana* natural accession showed the presence of variants of defense-related genes in nearly all populations investigated [37].

The long history of potato breeding includes numerous introgression of resistance genes from wild and cultivated species into the breeding genepool. The number of introgression events may vary and reach up to one hundred [38]. An investigation of NBS domains in the genomes of 91 potato cultivars revealed 587 segments mapped on the reference genome of the diploid (doubled monoploid) *S. tuberosum* Group Phureja clone (clonal cultivar) DM 1–3 516 R44 [39]. As expected, a number of R genes detected in modern cultivars was absent in the reference genome. However, an evaluation of potato genotypes with genome sequencing approaches will reveal both functional and non-functional NLR genes because of their rapid evolution and high recombination rate in the gene clusters [38].

The aim of this research concerns an evaluation of NLR genes expression in the transcriptomes of three potato cultivars (Evraziya, Siverskij, Sudarynya), which combine genetic material from wild and cultivated potato species: *Solanum stoloniferum* Schltdl., *S. demissum* Lindl., *S. acaule* Bitter, and *S. tuberosum* L. ssp. *andigena* Hawkes [40] and may bear different subsets of resistance genes. The selected cultivars possessed several allele-specific markers of well-known broad-spectrum late blight resistance genes (*RB/Rpi-blb1/Rpi-sto1*). According to our knowledge, the *RB/Rpi-blb1/Rpi-sto1* homologue in potato cultivars developed by conventional breeding through interspecific crosses was first described in our work [40,41]. The *RB/Rpi-blb1* gene was first identified in wild Mexican potato species *S. bulbocastanum* [42–44]. The functional homolog of the *RB/Rpi-blb1* gene was detected in wild Mexican potato species *S. stoloniferum-Rpi-sto1* [45], as well as in accessions of *S. papita*, *S. cardiophyllum*, *S. brachistotrichum* [46,47]. With the exception of transgenic varieties of potato obtained by cisgenesis, the *RB/Rpi-blb1/Rpi-sto1* is extremely rare in the genomes of potato cultivars [48]. To reveal NLR genes' variability, we compared the de novo-assembled transcriptomes of cultivars Evraziya, Siverskij, Sudarynya before and 24 h after inoculation with *P. infestans*.

2. Materials and Methods

2.1. Plant Materials

Three potato cultivars chosen for this study, Evraziya, Siverskij, Sudarynya, were bred in Leningrad Research Institute for Agriculture “Belogorka” (LenNIISKh ‘Belogorka’) located in the Northwestern zone of the Russian Federation where prolonged periods of humid and cool summer weather are favorable for late blight. Cultivar Sudarynya showed late blight resistance under field conditions of 2010–2017, cv. Evraziya demonstrated moderate level of resistance or susceptibility depending on the season, whereas cv. Siverskij was released quite recently [40]. These cvs. originated from the hybrid clone 8889/3, which included genetic material from *S. stoloniferum*, *S. demissum*, *S. tuberosum* ssp. *andigena* (the detailed genetic pedigrees of these cultivars were published earlier and are available in [40] (Supplement 2)). These three cultivars were selected for the present study based on the results of marker-assisted selection—five allele-specific markers of broad-spectrum late blight resistance genes *RB/Rpi-blb1/Rpi-sto1* were detected in all three cultivars (for details, see [40,41]). The sequences of the *Rpi-sto1* and *BLB1* F/R-amplicons were identical in analyzed genotypes and showed 99 and 100% similarity to the corresponding fragments of the *Rpi-blb1* and *Rpi-sto1* genes from the GenBank database, respectively [41].

Samples of cvs. Evraziya, Siverskij, and Sudarynya were provided by the cultivar author Dr. Z. Evdokimova from LenNIISKh ‘Belogorka’; cv. Bintje, and clone 30514/15 of wild species *Solanum stoloniferum* Schltdl. were obtained from VIR collection. Cultivar Bintje and *S. stoloniferum*, clone 30514/15, were used as susceptible and resistant controls, respectively.

2.2. Resistance Assessment

P. infestans isolate VZR18 obtained in 2018 from leaves of cv. Charodey cultivated in the experimental field of All Russian Institute of Plant Protection (VIZR) contained 8 virulence genes (1.2.3.4.6.7.10.11.) and was used for inoculation of potato cultivars in this research. Isolate was propagated and stored in the dark at 15 °C on rye agar.

Potato plants were grown in 0.5 l plastic pots filled with soil «Terra vita» at 22 °C under artificial light (16 h of light/8 h of dark) and air humidity of about 70% during 30 days.

Inoculation of 22–24 plants of each cultivar in double replications with *P. infestans* was conducted according to Khiutti et al. [49]. Whole plants were inoculated by suspension with a concentration of approximately 50,000 sporangia/mL. Before inoculation, the suspension was incubated at 12 °C for 3 h to release zoospores. Plants were inoculated on both the underside and the topside of the leaves using a hand sprayer. Inoculated plants were covered with plastic boxes for two days to create a humid chamber (>95% relative humidity) at 23 °C during the day and 15 °C at night with a 14 h photoperiod.

Each cultivar was represented by 22–24 plants in double replication—six plants (3 at 0 hpi and 3 at 24 hpi) per cultivar/per replication were used for RNA extraction, and the remaining plants were cultivated further for the infection assay.

Plant resistance was evaluated on the 7th, 10th, and 13th days after inoculation. Each plant was evaluated on a 9-point scale, in which a score of 9 corresponded to resistance (absence of disease symptoms) and a score of 1 corresponded to susceptibility (90–100% area of leaf was covered with blight-infected lesions) [50]. Plants with a score between 9 and 6 were considered resistant. A score of 5 corresponds to moderate resistance; scores of 4 or below indicate susceptibility. In all experiments, control uninfected plants of all potato genotypes had no symptoms of disease.

2.3. RNA-seq

Three biological replicates were used for each cultivar analyzed, and the leaf samples were taken before and 24 h after inoculation with *P. infestans*. Total RNA was extracted with an RNeasy Plant Mini Kit (Qiagen, Hilden, Germany).

The quality of the RNA samples was evaluated using a Bioanalyzer 2100 (Agilent, Santa Clara, CA, USA). All samples had RIN 7.8 or higher. RNAseq library preparations were carried out with 1.5 µg of total RNA fraction using TruSeq® Stranded mRNA Library Prep Kit (Illumina, San Diego, CA, USA) according to the manufacturer's instructions for barcoded libraries with small modifications (4 min RNA fragmentation time and 12 PCR cycles were used). Final libraries quantification was performed with a Bioanalyzer 2100 and a DNA High Sensitivity Kit (Agilent). After normalization, barcoded libraries were pooled and sequenced on a NextSeq 550 sequencer 2 × 150 bp using NextSeq® 500/550 High Output v2.5 Kit 300 cycles (Illumina).

Raw sequencing data were deposited in NCBI Sequence Read Archive (PRJNA765923).

2.4. RT-qPCR

To prepare single-stranded cDNA by reverse transcription, 1 µg aliquots of total RNA samples were used with RevertAid™ kit (Thermo Fisher Scientific Inc., Waltham, MA, USA). Primers for RT-qPCR were designed with the IDT PrimerQuest software (<http://eu.idtdna.com/PrimerQuest/Home/>; accessed on 15 August 2021). β -tubulin gene was used as a reference (GenBank: Z33382; forward primer 5'-AGCTTCTGGTGGACGTTATG-3', reverse primer 5'-ACCAAGTTATCAGGACGGAAGA-3'). In addition, transcripts of several differentially expressed NLRs were selected from the transcriptomic data for RT-qPCR verification (R228311: forward primer 5'-TGGGCAGTTGGAAATAGATAG-3', reverse primer 5'-CCATTCACCTACCGACAGAAG-3', OG0043132: forward primer 5'-TTTGTCCGTCGTGGTAATG-3', reverse primer 5'-AGGGAAGCATTGGTTTGG-3', OG0042956: forward primer 5'-TGAGGAGGACGCGAATAA-3', reverse primer 5'-GTTCA-TCTCAGCCAAGGTC-3'). The RT-qPCR was conducted using a HS-qPCR Lo-ROX SYBR

kit (Biolabmix, Novosibirsk, Russia). For each reaction, three technical replicates were run (Figure S1).

2.5. Bioinformatic Analysis

2.5.1. Read Quality Assessment, Filtering and Mapping

Sequencing quality and length distributions were assessed using FastQC software version 0.11.5 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>; accessed on 21 October 2020). Program fastp v. 0.20.1 with default settings [51] was used to remove adapters and filter reads by quality. Read mapping was performed by Dart v. 1.3.6 [52] to the *Solanum tuberosum* SolTub_3.0 reference genome assembly [39] through the Ensembl Plants database v. 49 [53]. Alignments with up to 8 mismatches were accepted given high inter- and intra-specific genetic variability of *Solanum* genus [54].

FeatureCount [55] was used to estimate the libraries quality. These values were further used to perform multidimensional scaling analysis using plotMDS function in EdgeR v. 3.26.8 [56].

2.5.2. Transcriptome Assembly De Novo

We implemented the combined transcriptome assembly strategy suitable for accurate transcriptome reconstruction [57,58]. Transcripts were assembled independently by de novo- and genome-guided Trinity v. 2.11.0 modes [59] and with the aid of Trans-ABYSS v. 2.0.1 [60] for each genotype from their specific paired read libraries. Read alignments onto potato genome obtained with the aid of Dart were further used for Trinity genome-guided transcriptome assembly.

We used Trans-ABYSS assembler with four different k-mer length values (67, 87, 107, and 127) for each of the paired-end read libraries independently yielding 24 de novo transcriptome assemblies per genotype. These transcriptome sequences were merged for each genotype with the transabyss-merge tool with parameter k-range set varying from 67 to 127.

Finally, Trinity de novo- and genome-guided assemblies and Trans-ABYSS assembly for each of the five potato genotypes were concatenated into a single file. To remove redundancy for genotype-specific transcriptome we used tool uclust from the usearch package v. 10.0.240 [61] with identity threshold equal to 100% to remove the duplicated transcripts.

2.5.3. Transcriptome Analysis

TransDecoder v. 5.5.0 (<https://github.com/TransDecoder/TransDecoder/wiki>; accessed on 1 October 2020) was used to predict open reading frames. The completeness of amino acid sequence sets was checked with the aid of BUSCO v. 3.1.0 [62] and ‘embryophyta_odb10’ sequence dataset.

Nucleotide sequences from non-redundant transcriptome assemblies were aligned to the *S. tuberosum* reference genome SolTub_3.0 by Gmap v.20 March 2018 [63] at 90% identity and 90% transcript coverage thresholds (only the best alignment for each transcript was taken). The transcript matches a gene if it overlaps with at least half of gene exons.

To find orthologous amino acid sequences for protein-coding transcripts, *S. tuberosum* reference genome [39] and *S. lycopersicum* reference genome [64] were taken from Ensembl Plants database v. 49 [53], and OrthoFinder v. 2.5.1 [65] was used as outgroup. The phylogenetic tree of the genotypes investigated was reconstructed with concatenated multiple sequence alignments of amino acid sequences from single-copy orthologous groups. The alignment was made with Mafft 7.480 (L-INS-i, max. 2 iterations) [66], the tree was reconstructed using IQ-TREE v. 1.6.12 (parameters -st AA -m TESTNEW -bb 1000 -alrt 1000; JTT+F+I, substitution model was chosen automatically) [67].

Quantification of transcripts abundances was used with the aid of kallisto v. 0.46.1 [68]. Transcripts with low abundancies determined by filterByExpr function of EdgeR package v. 3.26.8 [56] were excluded from analysis. The expression threshold value for each

mRNA was determined using its distribution in all transcripts and all libraries [69]. Differentially expressed genes (DEGs) were identified by EdgeR using generalized linear model (glmQLFit function) and F-test of quasi-likelihood (glmQLFTest function) at the 'FDR < 0.01' threshold.

To identify NBS-LRR domains in the amino acid sequences, the NLR-parser tool was used [70].

2.5.4. Compilation of the Marker Genes Associated with Wide-Range Resistance to *P. infestans*

Several genes conferring late blight resistance (broad-spectrum and race-specific) were selected as known markers for transcriptome-*Rpi-blb1/Rpi-sto1, R3a, R3b, R8* (Table S1). A search for homologs of transcripts sequences with no similarity to reference genome was made with NCBI amino acid sequences database and ublast tool from the usearch package v. 10.0.240 [61]. Best-hit homologs were identified at 95% (or higher) sequence identity and coverage at the protein level.

2.5.5. Phylogenetic Analysis of the Novel NBS-LRR Sequences

A number of transcripts encoding amino acid sequences with NBS-LRR domain signatures did not show significant similarity to the potato reference genome. To characterize these putative NBS-LRR proteins, we performed phylogenetic tree reconstruction for novel and 438 known NBS-LRR potato amino acid sequences taken from Jupe et al. [13]. Five protein sequences from Table S1 were added to the dataset as well. Hmmscan from HMMER package v.3.1b2 [71] was used to identify Pfam domain signatures [72] in these sequences and extract fragments of the NB-ARC domains (Pfam id PF00931.23). Mean length of the NB-ARC domains is 209 residues; therefore, sequences with NB-ARC domain length less than 160 amino acids were removed. NB-ARC domain sequences were aligned with Mafft v. 7.480 (L-INS-i, iterative refinement method, max. 1000 iterations) [66]. Phylogenetic tree was reconstructed by maximum likelihood method using IQ-TREE v. 1.6.12 [67] (parameters -st AA -m TESTNEW -bb 1000 -alrt 1000; JTT+F+R7 substitution model was chosen automatically). SeqKit v0.16.1 [73] and Hmmpy (<https://github.com/EnzoAndree/Hmmpy>; accessed on 7 April 2021) tools were used for sequence data processing at various stages of this analysis.

3. Results

3.1. Assessment of Resistance to *P. infestans*

The results of evaluation of potato cultivars resistance to *P. infestans* isolate VZR18 are presented in Table 1. On the 7th day after inoculation (DAI), plants of the susceptible cultivar Bintje in two independent replications were severely affected, and on the 13th day, the almost died (score 1), indicating sufficient infection load and favorable conditions for disease development, allowing a reliable assessment of resistance. Cultivars Sudarynya and Siverskij showed resistance to isolate VZR18. *S. stoloniferum* clone 30514/15 and had no symptoms of the disease. On the 13th day, the cultivar Evraziya was affected at the level of susceptible cv. Bintje (Table 1).

3.2. Libraries Comparison and Quality Evaluation

The libraries comprise approximately 2.02 billion paired reads (298 billion bases; statistical metrics presented in more details in Table S2). After filtering by quality, 1.81 billion short reads (90% of raw libraries) comprising 269 billion bases remained. The number of bases with q > 30 in filtered reads varies from 93.4% (Bintje, 0 h, library 1) to 94.9% (Sudarynya, 24 hpi, library 3). The average number of paired reads per library is approximately 60.4 million, the average lengths of reads is 148 bases, and the average number of bases with Q > 30 is 94.

Table 1. Potato cultivars resistance to *P. infestans* isolate VZR18.

Potato Cultivar	Disease Assessment (9-Point Scale) *					
	Replicate 1			Replicate 2		
	Days after Inoculation					
	7	10	13	7	10	13
Sudarynya	8.2 ± 0.23	8.0 ± 0.23	8.0 ± 0.22	9.0 ± 0.00	6.5 ± 0.09	7.0 ± 0.08
Siverskij	6.9 ± 0.15	6.7 ± 0.13	6.2 ± 0.13	8.0 ± 0.00	7.0 ± 0.20	4.7 ± 0.09
Evraziya	4.8 ± 0.27	4.2 ± 0.19	1.8 ± 0.11	3.4 ± 0.17	2.1 ± 0.17	1.1 ± 0.05
Bintje (susceptible control)	4.5 ± 0.15	2.5 ± 0.14	1.0 ± 0.00	1.7 ± 0.14	1.0 ± 0.00	1.0 ± 0.00
<i>S. stoloniferum</i> , clone 30514/15 (resistant control)	9.0 ± 0.00	9.0 ± 0.00	9.0 ± 0.00	-	-	-

* score of 9 to 6, resistant; score 4 and below, susceptible.

The multidimensional scaling analysis demonstrates that libraries of the same genotype and conditions are clustered on the MDS plot (Figure 1). The transcriptomes corresponding to control (0 hpi) and *P. infestans* infection (24 hpi) were clustered separately. As expected, the libraries from *S. stoloniferum* clone 30514/15 differ from cultivated potato genotypes. The changes in transcriptomes of cultivated potato genotypes revealed two clusters: smaller changes in response to *P. infestans* were characteristic for cv. Evraziya and cv. Bintje, and more pronounced changes were detected for cv. Siverskij and cv. Sudarynya. This difference could be explained by a genotype-specific response to pathogen invasion.

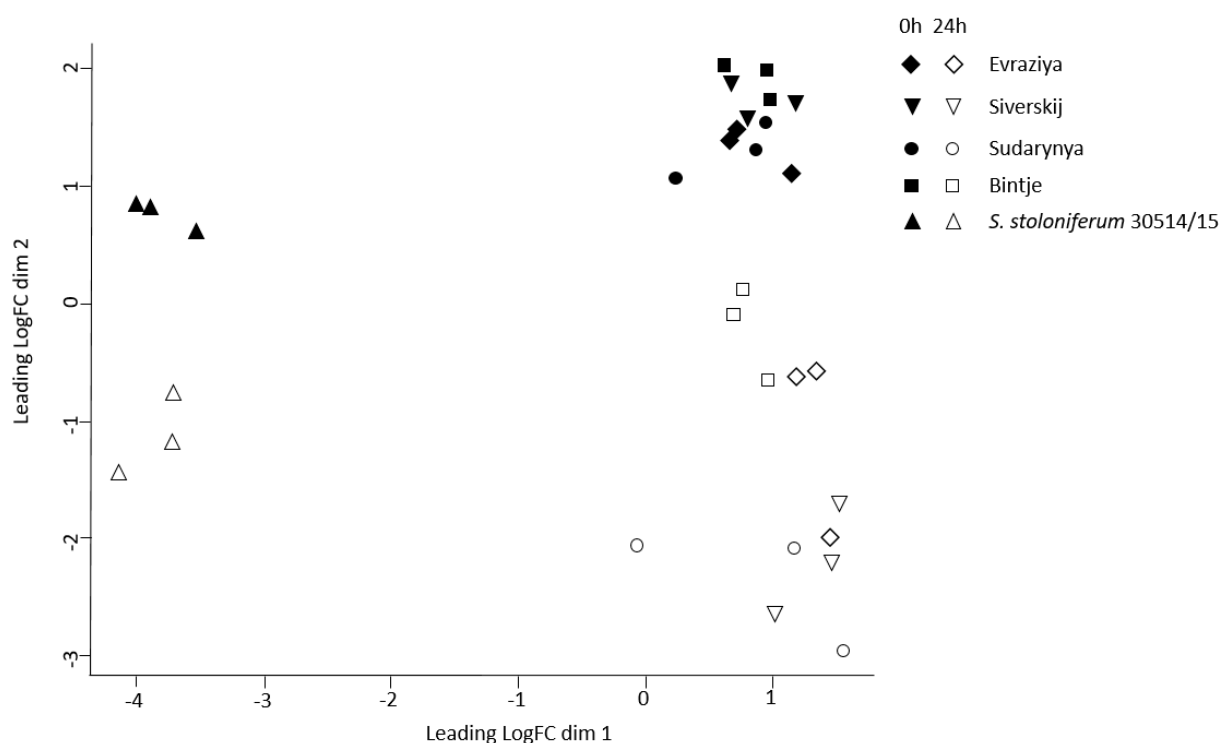


Figure 1. Comparison of libraries based on the contigs expression recorded at 24 h after inoculation (empty symbols) with the control (filled symbols) using multidimensional scaling plot for component 1 (X axis) and 2 (Y axis). Libraries from five genotypes are marked by distinct symbol shapes as shown on the legend in the top right part of figure.

3.3. De novo Transcriptome Assembly and Reconstruction of the Orthologous Groups

Transcriptomes were assembled de novo (Table S3). The assembly parameter N50 varies from 2337 to 2444. Transcript lengths median values vary from 1691 to 1742 between

the five reconstructed transcriptomes. After redundancy filtering, 32% of transcripts were removed on average.

De novo-reconstructed non-redundant transcriptomes contain between 62,179 (cv. Sudarynya) and 110,556 (*S. stoloniferum*, clone 30514/15) transcripts. The BUSCO analysis demonstrated the high completeness of the transcriptomes assemblies, with the verified BUSCO sequence fraction varying from 93.2 to 97.3% (Table S4).

A search of orthologous groups in the reconstructed transcriptomes and in the reference transcriptomes of *S. tuberosum* and *S. lycopersicum* identified 53,775 orthogroups. A total of 172 orthogroups contain exactly one sequence in each of the seven transcriptomes (single-copy orthogroups). Based on the alignments of protein sequences in these orthogroups, the phylogenetic tree was reconstructed with the aid of maximum likelihood method (Figure 2).

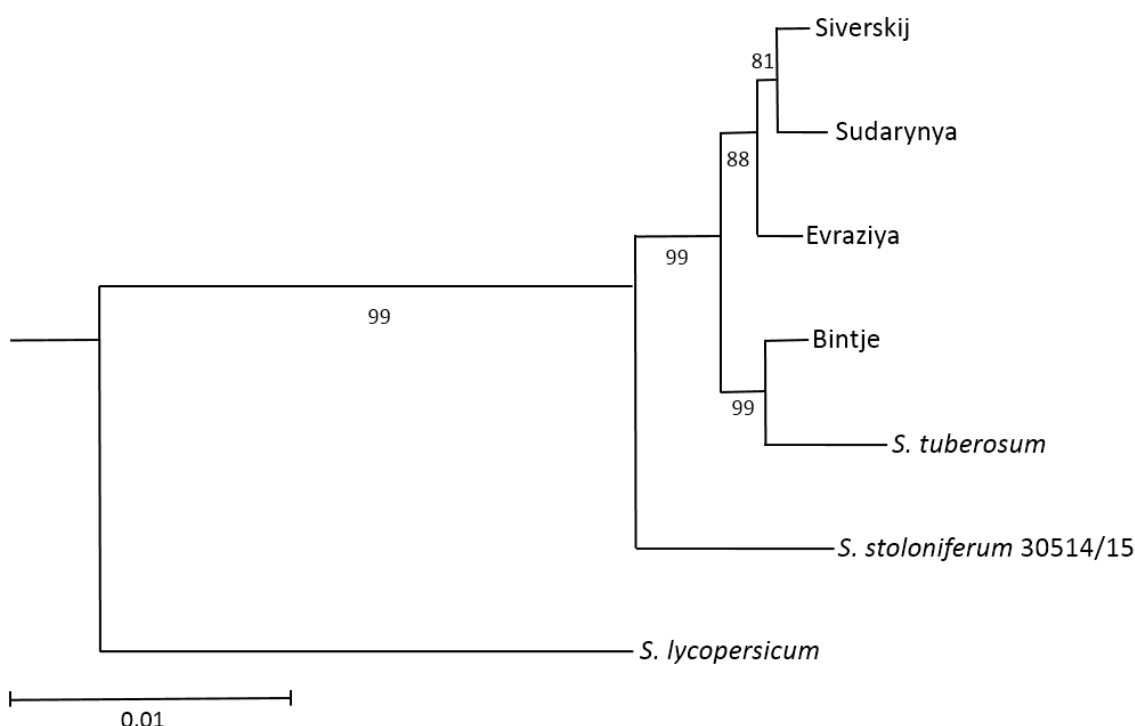


Figure 2. Phylogenetic tree reconstructed by IQ-TREE maximum likelihood method for concatenated sequence alignment of 172 single-copy orthogroups. Tree is rooted at *S. lycopersicum* node (outgroup). Distance scale shown below the tree.

As expected, three cultivars of common origin, Evraziya, Siverskij, and Sudarynya, form a well-separated subcluster. It is known that the pedigree of cv. Bintje (bred in the Netherlands in 1904) does not include crosses with wild potato species. *S. stoloniferum* 30514/15 represent a sister group for the cultivated potato genotypes and the reference genome.

3.4. Identification and Analysis of the NBS-LRR Proteins

The analysis of amino acid sequences encoded by the de novo-reconstructed transcripts with the aid of NLR-parser tool [70] predicted the proteins containing complete or partial NBS-LRR domains (Table 2).

The number of amino acid sequences with complete or partial NBS-LRR domain signatures is similar between four potato genotypes and varies from 960 (cv. Sudarynya) to 1188 (cv. Siverskij). The number of sequences with complete NBS-LRR domains is two to three times lower and varies from 334 (cv. Sudarynya) to 504 (*S. stoloniferum* 30514/15). Most of transcripts encoding complete NBS-LRR domains (>80% for each genotype) have

an expression level above 1 TPM. Detailed information on predicted NBS-LRR proteins is available in File S1.

Table 2. Proteins encoded by de novo-assembled potato transcriptomes and containing predicted NBS-LRR domains.

Potato Genotype	Number of Proteins with NBS-LRR Domains	Number of Proteins with Complete NBS-LRR Domains	Number of Proteins with Complete NBS-LRR Domains and TPM > 1
Siverskij	1188	409	358 (88%)
Evraziya	1331	449	376 (84%)
Sudarynya	960	334	310 (93%)
Bintje	1185	444	384 (86%)
<i>S. stoloniferum</i> , 30514/15	1333	504	417 (83%)

3.5. Novel Transcripts Encoding Proteins with NBS-LRR Domains

De novo-assembled transcripts encoding proteins with complete NBS-LRR domains (Table 2) were analyzed for similarity to the reference *S. tuberosum* genome. It was found that a fraction of predicted NLR proteins has no counterparts in the genome (Table 3). The origin of these novel transcripts may relate to the introgression of resistance genes from wild and cultivated potato species as well as the intense recombination within the NBS-LRR genes clusters [74,75].

Table 3. Novel NBS-LRR sequences in five potato transcriptomes.

Genotype	Number of Transcripts			Differential Expression (FDR < 0.05)	
	Total	CNL	TNL	Upregulated	Downregulated
Evraziya	104	66	38	3	1
Siverskij	103	67	36	7	5
Sudarynya	109	70	39	11	3
Bintje	133	94	39	7	9
<i>S. stoloniferum</i> 30514/15	160	138	22	41	16

Transcriptomes of cvs. Siverskij, Evraziya, Sudarynya have similar numbers of the novel NBS-LRR transcripts, including the CNL/TNL classes, and differ from the cv. Bintje or *S. stoloniferum* 30514/15 transcriptomes (Table 3). Comparison of transcriptomes taken before (0 hpi) and after *P. infestans* inoculation (24 hpi) revealed subsets of transcripts with either increased or decreased levels in response to the pathogen invasion (Table 3; data for novel transcripts are presented in Files S2 (description) and S3 (sequences)). As expected, *S. stoloniferum* 30514/15 was characterized with much higher number of *P. infestans*-induced novel NLR transcripts.

We also checked the transcriptomes of *S. tuberosum* genotypes for mRNAs potentially encoding well-known broad-spectrum resistance genes against *P. infestans* (*Rpi-blb1* (*Rpi-sto1*), *R3a*, *R3b*, *R8*, homology threshold >99%) (Table 4; cv. Bintje contains no transcripts from this list).

The results shown in Table 4 demonstrate that the transcriptomes of genotypes Siverskij, Sudarynya, and Euraziya encode proteins nearly identical to the *R3b* gene from *S. demissum* (identity > 99%). The transcriptomes of cv. Siverskij and cv. Sudarynya bear transcripts highly similar to *Rpi-blb1* from *S. bulbocastanum* (or *Rpi-sto1* from *S. stoloniferum*), identity >99% at amino acid level. Interestingly, cv. Euraziya transcriptome does not contain mRNA for *RB/Rpi-blb1/Rpi-sto1* either at 0 or at 24 hpi despite the genome of this cultivar containing corresponding intragenic markers [40,41]

Table 4. Expression of *P. infestans* resistance genes in the transcriptomes of potato genotypes.

Potato Cultivars	Transcript ¹	Homolog/Identity	Abundance at 0 hpi, TPM	Abundance at 24 hpi, TPM	Log ₂ (FC) 0 vs. 24 hpi	FDR
Siverskij	R450380	<i>R3b</i> /99.9%	7.42	5.82	−0.52	0.21
	R488392	<i>Rpi-blb1</i> /99.8%	2.04	7.43	2.14	0.004
Evraziya	R595896	<i>R3b</i> /99.9%	5.66	5.75	0.49	0.55
	R489301	<i>R3b</i> /99.8%	2.52	3.05	0.09	0.95
Sudarynya	R443838	<i>Rpi-blb1</i> /99.8%	0.90	5.43	4.21	0.007

¹ IDs for de novo predicted transcripts.

To characterize the novel NLR-encoding transcripts, the phylogenetic tree was reconstructed for their NB-ARC domains. The initial dataset contained 1052 sequences, including 608 novel NBS-LRR genes found in this research (description is available in File S3), 438 sequences that were taken from [13], and a few sequences for wide-range resistance genes against *P. infestans* (description is available in Table S1). After filtering (extraction of partial NB-ARC domains, removal of identical sequences), 812 non-redundant NB-ARC domains were used for tree reconstruction (File S4). The tree was rooted at CNL-R node. The part of tree with phylogenetic relationships in CNL6, 7, and 8 classes is shown in Figure 3. The full phylogenetic tree for NB-ARC domains from known and novel sequences and marker genes is presented in Newick, PDF, and SVG formats in File S4.

The distribution of NB-ARC domains of novel NLR proteins between the ten classes from [13] is non-uniform (Figure 4). The CNL-8 and TNL classes contain many novel sequences, while CNL-2 has only a few, and the CNL-R class does not contain novel sequences at all. The CNL-8 class includes the R3a protein from *S. tuberosum* and R3b-like protein from *S. stoloniferum* 30514/15. The CNL-6 class includes *Rpi-sto1* of *S. stoloniferum* and *Rpi-blb1* of *S. bulbocastanum* with identical NB-ARC domains. The R8 gene from *S. demissum* does not fall into these classes but into the large cluster sister to the CNL-1 class.

The novel NLR proteins of different potato cultivars under investigation (Table 3) were compared with the aid of the CD-hit program (proteins with identity >99% were combined).

One may see (Figure 5) that the intersections are rather small; that is, most of the novel NLR-like proteins are genotype specific. As expected, related genotypes (cv. Sudarynya, cv. Siversky, cv. Evraziya) have a larger number of common NLR proteins than cv. Bintje or *S. stoloniferum* 30514/15.

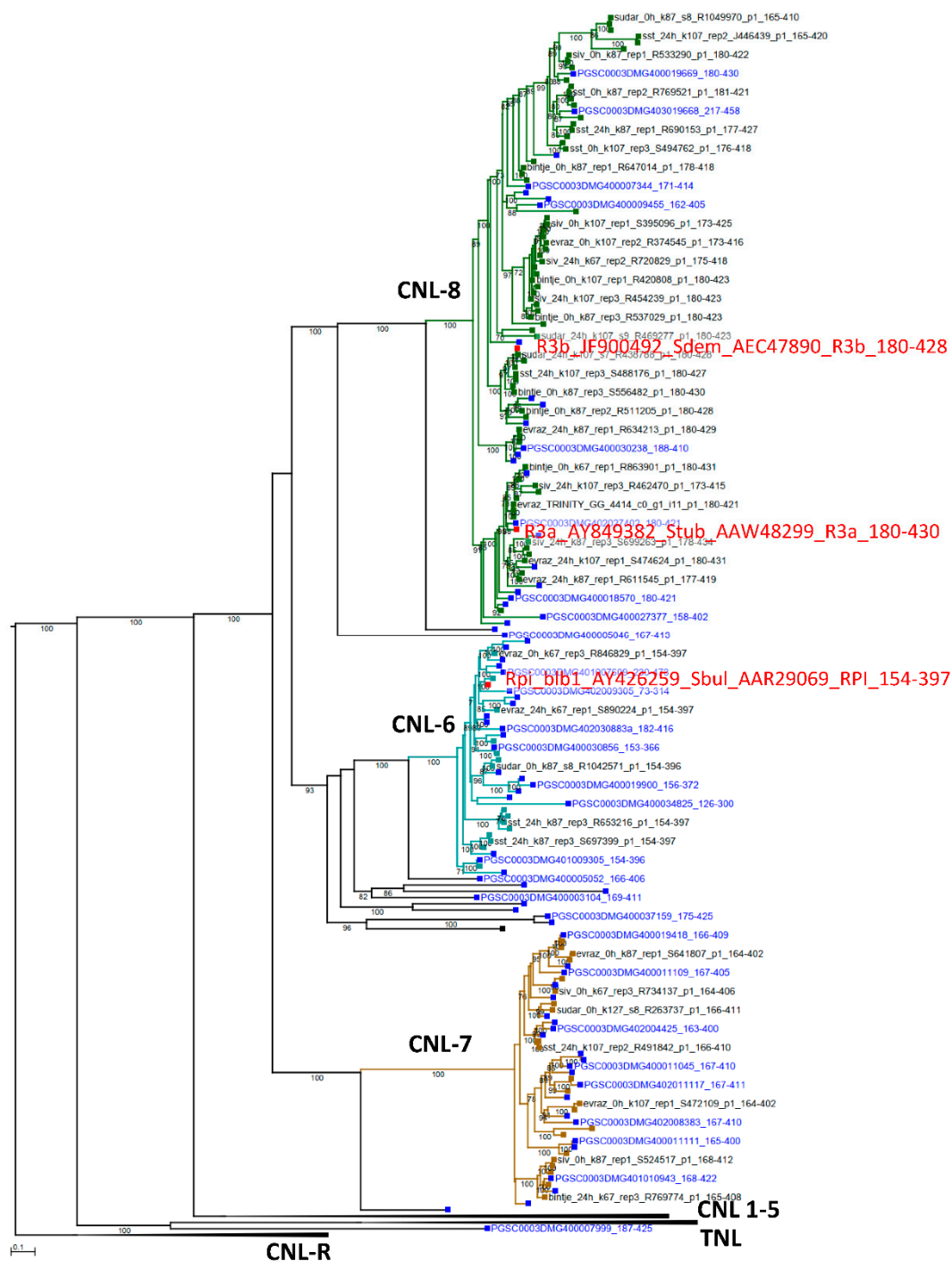


Figure 3. Fragment of the phylogenetic tree of NB-ARC domain containing proteins and their classes according to [13] (only clusters CNL-8, CNL-6, and CNL-7 are shown, classes TNL, CNL-R, and CNL 1–CNL 5 are collapsed; full phylogenetic tree is shown in the File S4). The IDs of known genes are highlighted in blue, wide-range resistance genes (*R3a*, *R3b*, *Rpi-blb1*) are shown in red, and black color highlights the proteins IDs with no homology to the reference genome (novel NLR genes, File S3). Bootstrap support values greater than 70 are shown. The scale of branches lengths is provided at the bottom of the figure.

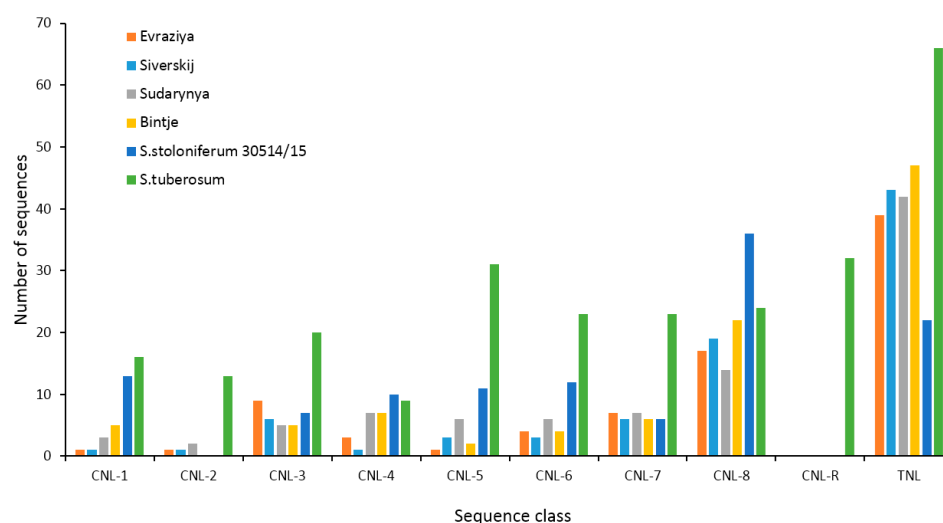


Figure 4. The distribution of the numbers of NBS-LRR novel sequences from five investigated genotypes and *S. tuberosum* between different NB-ARC classes according to [13]. X-axis: protein class; Y-axis: number of sequences. The bar colors for different genotypes are shown in the left top part of the plot.

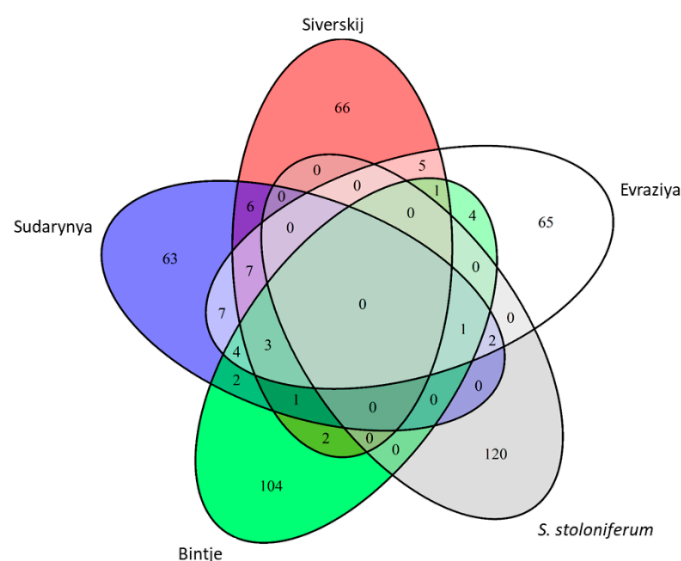


Figure 5. Intersection between the sets of new NLR proteins for cv. Siverskij (red), cv. Sudarynya (blue), cv. Evraziya (white), cv. Bintje (green), and *S. stoloniferum* clone 30514/15 (grey).

4. Discussion

NLR genes play a vital role in plant defense against various pathogens. Rapid co-evolution of pathogens and host plants results in a huge natural diversity of NBS-LRR receptors intensively screened for new valuable R genes for crop improvement. In addition, some incompatible NLR gene combinations might decrease plant fitness (e.g., [26,27,29]). Plant breeding has been conducted for many decades, and cultivars may bear rather different subsets of NLR genes. In addition, NLR loci frequently containing gene clusters are characterized with a high recombination rate; these evolution hot spots may facilitate the selection of new efficient resistance genes in response to newly arising strains of pathogens. Here, we investigated the transcriptomes of potato cultivars combining genetic material from wild and cultivated potato species for the presence of NLR-related transcripts.

The first 24 h period after inoculation is considered the most critical for the interaction between potato and *P. infestans* (e.g., [76]); thus, the samples were analyzed either before or 24 h after inoculation with *P. infestans*. The transcriptomes were assembled de novo to take

into account the pool of transcripts with no homology to reference genome, and NBS-LRR proteins were predicted computationally with the aid of an NLR-parser tool [70].

Three related cultivars, Evraziya, Siverskij, and Sudarynya, with the interspecific hybrid with wild Mexican species *S. stoloniferum* and *S. demissum* in their pedigrees, and two controls (susceptible cv. Bintje and resistant clone 30514/15 of the wild species *S. stoloniferum*) were taken for this analysis. Cultivars Sudarynya and Siverskij were evaluated as resistant to late blight in field trials in the 2016–2017 season (score 7–8) and to *P. infestans* isolate VZR18 with virulence formula 1.2.3.4.6.7.10.11 in laboratory assay (Table 1). Cv. Evraziya was evaluated as susceptible to isolate VZR18. The transcriptomes of four potato cultivars (Evrasiya, Siverskij, Sudarynya, Bintjee) with different response to late blight were compared for the presence of NLR genes-related mRNAs.

The comparison of cultivars demonstrated the difference between transcriptomes taken before and after *P. infestans* inoculation. Interestingly, the *S. tuberosum* transcriptomes at 24 hpi were divided into two clusters: those susceptible to late blight cvs. Evraziya and Bintje versus resistant cvs. Sudarynya and Siverskij, which may reflect their differential response to *P. infestans* (Figure 1). The reconstructed phylogenetic tree for orthologous groups of genes also revealed a separate cluster containing a close group from cv. Sudarynya and cv. Siverskij together with cv. Evraziya, whereas cv. Bintje was closer to the reference genome (Figure 2). This coincided with the fact that the pedigree of cv. Bintje (bred in the Netherlands in 1904) does not include crosses with wild potato species.

Analysis of de novo-assembled transcriptomes with NLR-parser [70] revealed a large number of transcripts encoding proteins with NBS-LRR domains (Table 2). Interestingly, a considerable part of these NLR-omes (from 103 transcripts in cv. Siverskij to 133 mRNAs in cv. Bintje, TPM > 1) had no counterparts in the reference *S. tuberosum* genome, and some of these novel transcripts differentially expressed in response to *P. infestans* (Table 3). A targeted analysis of *S. tuberosum* transcriptomes for mRNAs of known *P. infestans* resistance genes (*R3a*, *R3b*, *R8*, *Rpi-blb1/Rpi-sto1*) showed the presence of transcripts homologous (>99.5%) to *R3b* (cvs. Siverskij, Euraziya, Sudarynya) and *Rpi-blb1/Rpi-sto1* (cvs. Siverskij, Sudarynya). Most transcripts were characterized with moderate expression level (TPM > 1). However, the comparison of transcript abundances at 0 hpi and 24 hpi showed significant induction only for mRNA R488392 from cv. Siverskij and mRNA R443838 from cv. uy77i87Sudarynya (both similar to *Rpi-blb1*), whereas cv. Euraziya contained no *Rpi-blb1*-related mRNA either at 0 or at 24 hpi.

Actually, these results demonstrated the importance of transcriptomic evaluations for resolving the contradictory data between the marker-assisted selection and phenotyping. In some cases, the cultivar phenotypic resistance may differ from the DNA-marker-based prediction because of inappropriate expression pattern of the targeted R gene. For example, all the cultivars (Siverskij, Evraziya, Sudarynya) originating from hybrid clone 8889/3 (containing genetic material from *S. stoloniferum*) have the diagnostic intragenic markers of the *RB/Rpi-blb1/Rpi-sto1* gene in their genomes (for detail, see [40]). It may be expected that this NLR (if functional) will contribute to late blight resistance in all three genotypes. However, despite the presence of the DNA-markers, cv. Evraziya transcriptome does not contain *RB/Rpi-blb1/Rpi-sto1* mRNA, and this cultivar was susceptible to *P. infestans* in both field and laboratory evaluations. In our opinion, MAS results could be more precise if accompanied with transcriptomic assay or at least targeted RT-qPCR.

Despite the resistance of tested cultivars to the *P. infestans* isolate VZR18 and their field resistance to late blight in 2016–2017, it was later shown that in 2020 all these cultivars (Siverskij, Evraziya, Sudarynya) were severely affected by *P. infestans* in the field trials under conditions of high disease severity; they also appeared to be susceptible to *P. infestans* isolate MP1841 with virulence formula 1.2.3.4.5.6.7.8.9.10.11 (score 1–3, data not shown). Thus, the resistance of these cultivars turned out to be race-specific, which might be explained by insufficient expression level of *RB/Rpi-blb1/Rpi-sto1*. The last data indicate that for reaching the durable late blight resistance the stacks from several favorable genes and their simultaneous expression is important.

Indeed, the introgression of the *RB/Rpi-blb1/Rpi-sto1* gene per se does not necessarily provide a potato cultivar with high-level resistance against *P. infestans*. It has been previously reported that transgenic potato cultivars bearing the *RB/Rpi-blb1* gene from *S. stoloniferum* in its native form (8.5 kb long segment with promoter, introns, etc.) were resistant against *P. infestans*. However, the resistance level of transgenic lines varied greatly (susceptible, moderate susceptible, moderate resistant, resistant) and correlated with transgene copy numbers and expression rates [77]. Similar results were reported by another group [78].

Another outcome of this study concerns the considerable abundance of NLR transcript absent in the reference genome. Interestingly, the transcriptomes of the potato cultivars tested contained significant numbers of novel NLR transcripts. The reconstructed phylogenetic tree (File S4, Figure 3) showed the distribution of novel transcripts according to the classification of NBS-LRR genes based on their domain structures [13]. These potential novel NLR proteins are distributed non-uniformly but are present in almost all groups except CNL-R. In fact, their structure is close to well-known functional NLRs, and they may be involved in regulation of various biological processes. Indeed, the functionality of novel NLRs needs further experimental verification.

The origin of novel NLR transcripts may relate to introgression of resistance genes from other species as well as to intense recombination within the NBS-LRR genes clusters [74,75]. Different subsets of NLR alleles were introgressed into the genepool of modern cultivars from many wild species of Central and South America as well as from cultivated species. A comparison of proteins encoded by novel NLR transcripts demonstrated a relatively small intersection between the genotypes (Figure 5). This observation coincided well with their complex pedigrees, which contain hybridizations with introgressive lines originating from hybrid clones with *S. stoloniferum*, *S. demissum*, and *S. acaule*, *S. tuberosum* ssp. *tuberosum* [40]. The novel cultivar-specific NLR transcripts may also result from intense recombination inside the gene loci. It is likely that NLR gene loci represent some kind of playground for the rapid evolution of resistance genes to provide a reservoir for selection under the severe pressure of newly arising pathogen strains.

Investigation of genotype-specific NLR genes at the level of de novo-assembled transcriptome may considerably supplement genomic data because a cluster structure of NLR loci makes prediction at the genome level complicated. Taking novel NLRs into account may considerably extend the interpretation of major resistance genes mapping experiments.

One of the aims of this study was to compile a set of new candidate R genes (File S2) for further investigation. Association of a candidate R gene with the cultivar specific resistance against some pathogen provides opportunity for further study and inclusion in potato breeding programs or other biotechnological approaches for plant improvement (e.g., [3,79–85]). Further investigation of candidate R genes may be conducted with the aid of reverse genetics (suppression or increased expression of a targeted gene under testing combined with phenotype evaluation). This way may be of use to reveal novel R genes in selected resistant clones from the genetic collections. In this work, the late blight resistance of analyzed potato cultivars was race specific. However, *S. stoloniferum* clone 30514/15 is highly resistant to late blight, and its transcriptome contained 41 novel mRNA encoding NLR proteins. Further detailed investigation of these mRNAs could reveal new NLRs that are potentially useful for the further improvement of *S. tuberosum* commercial cultivars. *P. infestans* is rapidly evolving [84] and may gain the resistance to fungicides [85], and some new strains are rather dangerous [86], which makes molecular breeding quite pertinent [87]. The transcriptomic approach has proved itself as a useful tool to reveal new potential R genes for further introgression.

5. Conclusions

Three potato cultivars (Evraziya, Siverskij, Sudarynya) combining genetic material from wild and cultivated potato species and bearing the same intragenic markers of the *RB/Rpi-blb1/Rpi-sto1* gene were investigated. It was found that:

1. The transcriptomic assay may resolve the contradictive results of marker-assisted-selection and phenotypic data. Here, both resistant and susceptible cultivars have the same intragenic markers of the *RB/Rpi-blb1/Rpi-sto1*; however, the induction of *RB/Rpi-blb1/Rpi-sto1* transcript was found only in cultivars resistant to *P. infestans* isolate VZR18.

2. The significant fractions of transcriptomes analyzed (cvs. Evraziya, Siverskij, Sudarynya, Bintjee, *S. stoloniferum* clone 30514/15) represent novel NLR genes with no homology to the reference genome. Taking these NLRs into account may extend the limitations of the reference genome and increase the efficiency of novel R genes mapping.

Further functional analysis of novel NLR genes may provide a valuable resource for breeding.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/agronomy11122426/s1>, Figure S1: RT-qPCR verification of transcriptomic data; Table S1: Marker NLR genes associated with wide-range resistance against *P. infestans*; Table S2: RNA-seq libraries statistics for raw data and after filtering by quality; Table S3: De novo-assembled transcriptomes metrics; Table S4: BUSCO scores for de novo transcriptomes; File S1: Data on NBS-LRR encoding transcripts predicted in 5 potato transcriptomes before and after *P. infestans* inoculation; File S2: Data on NLR-encoding transcripts without similarity to *S. tuberosum* reference genome; File S3: nucleotide and amino acid sequences for novel NLR-encoding transcripts (describes in File S2); File S4: Multiple sequence alignment for non-redundant NB-ARC domains from novel, known *S. tuberosum* NBS-LRR proteins and five marker genes (novel_ref_R5_nbspep_nbarc_L160_nr_aln.fa). Sequences with identical domains are listed in Identical_NB_ARC.xlsx. Full phylogenetic tree is represented in three formats, Newick (novel_soltub_R5_NB-ARC-tree.nph), PDF (novel_soltub_R5_NB-ARC-tree.pdf) and SVG (novel_soltub_R5_NB-ARC-tree.svg), see also NB-ARC tree readme.txt in this archive.

Author Contributions: Conceptualization, A.V.K., O.S.A. and T.A.G.; Methodology, G.V.V., N.V.S., O.Y.A. and S.M.I.; Software, D.A.A. and N.S.; Validation, A.Y.G.; Investigation, A.K., O.S.A., A.V.K. and T.A.G.; Resources, T.A.G.; Writing A.V.K., D.A.A., O.S.A. and T.A.G.; Project Administration, A.V.K. and T.A.G. All authors have read and agreed to the published version of the manuscript.

Funding: The research was supported by Kurchatov Genomics Center of IC&G (Agreement with Ministry of Science and Higher Education RF 075–15–2019-1662) in the frame of potato pangenome and pantranscriptome sequencing and bioinformatics analysis. NGS sequencing was conducted in the ICG Joint Center for Genome Studies (supported in the frame of ICG budget project 0259-2021-0012). The computational data analysis was conducted using the resources of the ICG “Bioinformatics” Joint Computational Center.

Data Availability Statement: Raw sequencing data were deposited in the NCBI Sequence Read Archive (PRJNA765923).

Acknowledgments: The authors are grateful to Z.Z. Evdokimova (Leningrad Research Institute for Agriculture “Belogorka, branch of the A.G. Lorch Russian Potato Research Center) who provided the potato cultivars for this research.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Caruana, B.; Rodoni, B.; Constable, F.; Slater, A.; Cogan, N. Genome Enhanced Marker Improvement for Potato Virus Y Disease Resistance in Potato. *Agronomy* **2021**, *11*, 832. [CrossRef]
2. Stefańczyk, E.; Plich, J.; Janiszewska, M.; Smyda-Dajmund, P.; Sobkowiak, S.; Śliwka, J. Marker-assisted pyramiding of potato late blight resistance genes *Rpi-rzc1* and *Rpi-phu1* on di- and tetraploid levels. *Mol. Breed.* **2020**, *40*, 1–12. [CrossRef]
3. Rakosy-Tican, E.; Thieme, R.; König, J.; Nachtigall, M.; Hammann, T.; Denes, T.-E.; Kruppa, K.; Molnár-Láng, M. Introgression of Two Broad-Spectrum Late Blight Resistance Genes, *Rpi-Blb1* and *Rpi-Blb3*, From *Solanum bulbocastanum* Dun Plus Race-Specific R Genes Into Potato Pre-breeding Lines. *Front. Plant Sci.* **2020**, *11*, 699. [CrossRef]

4. Shao, Z.-Q.; Wang, B.; Chen, J.-Q. Tracking ancestral lineages and recent expansions of NBS-LRR genes in angiosperms. *Plant Signal. Behav.* **2016**, *11*, e1197470. [[CrossRef](#)] [[PubMed](#)]
5. Lozano, R.; Ponce, O.P.; Ramírez, M.; Mostajo, N.; Orjeda, G. Genome-Wide Identification and Mapping of NBS-Encoding Resistance Genes in *Solanum tuberosum* Group Phureja. *PLoS ONE* **2012**, *7*, e34775. [[CrossRef](#)] [[PubMed](#)]
6. Sekhwal, M.K.; Li, P.; Lam, I.; Wang, X.; Cloutier, S.; You, F.M. Disease Resistance Gene Analogs (RGAs) in Plants. *Int. J. Mol. Sci.* **2015**, *16*, 19248–19290. [[CrossRef](#)] [[PubMed](#)]
7. Zhao, Y.; Weng, Q.; Song, J.; Ma, H.; Yuan, J.; Dong, Z.; Liu, Y. Bioinformatics Analysis of NBS-LRR Encoding Resistance Genes in *Setaria italica*. *Biochem. Genet.* **2016**, *54*, 232–248. [[CrossRef](#)]
8. Pal, T.; Jaiswal, V.; Chauhan, R.S. DRPPP: A machine learning based tool for prediction of disease resistance proteins in plants. *Comput. Biol. Med.* **2016**, *78*, 42–48. [[CrossRef](#)]
9. Toda, N.; Rustenholz, C.; Baud, A.; Le Paslier, M.-C.; Amselem, J.; Merdinoglu, D.; Faivre-Rampant, P. NLGenomeSweeper: A Tool for Genome-Wide NBS-LRR Resistance Gene Identification. *Genes* **2020**, *11*, 333. [[CrossRef](#)]
10. Liu, W.; Ghouri, F.; Yu, H.; Li, X.; Yu, S.; Shahid, M.Q.; Liu, X. Genome wide re-sequencing of newly developed Rice Lines from common wild rice (*Oryza rufipogon* Griff.) for the identification of NBS-LRR genes. *PLoS ONE* **2017**, *12*, e0180662. [[CrossRef](#)]
11. Zhou, P.; Silverstein, K.A.T.; Ramaraj, T.; Guhlin, J.; Denny, R.; Liu, J.; Farmer, A.D.; Steele, K.P.; Stupar, R.M.; Miller, J.R.; et al. Exploring structural variation and gene family architecture with De Novo assemblies of 15 Medicago genomes. *BMC Genom.* **2017**, *18*, 261. [[CrossRef](#)] [[PubMed](#)]
12. Li, Q.; Jiang, X.-M.; Shao, Z.-Q. Genome-Wide Analysis of NLR Disease Resistance Genes in an Updated Reference Genome of Barley. *Front. Genet.* **2021**, *12*. [[CrossRef](#)] [[PubMed](#)]
13. Jupe, F.; Witek, K.; Verweij, W.; Śliwka, J.; Pritchard, L.; Etherington, G.J.; Maclean, D.; Cock, P.J.; Leggett, R.M.; Bryan, G.J.; et al. Resistance gene enrichment sequencing (RenSeq) enables reannotation of the NB-LRR gene family from sequenced plant genomes and rapid mapping of resistance loci in segregating populations. *Plant J.* **2013**, *76*, 530–544. [[CrossRef](#)] [[PubMed](#)]
14. Quirin, E.A.; Mann, H.; Meyer, R.S.; Traini, A.; Chiusano, M.L.; Litt, A.; Bradeen, J. Evolutionary Meta-Analysis of Solanaceous Resistance Gene and Solanum Resistance Gene Analog Sequences and a Practical Framework for Cross-Species Comparisons. *Mol. Plant-Microbe Interact.* **2012**, *25*, 603–612. [[CrossRef](#)]
15. Morata, J.; Puigdomènech, P. Variability among Cucurbitaceae species (melon, cucumber and watermelon) in a genomic region containing a cluster of NBS-LRR genes. *BMC Genom.* **2017**, *18*, 138. [[CrossRef](#)]
16. Cao, W.; Gan, L.; Shang, K.; Wang, C.; Song, Y.; Liu, H.; Zhou, S.; Zhu, C. Global transcriptome analyses reveal the molecular signatures in the early response of potato (*Solanum tuberosum* L.) to *Phytophthora infestans*, *Ralstonia solanacearum*, and Potato virus Y infection. *Planta* **2020**, *252*, 1–13. [[CrossRef](#)]
17. Kochetov, A.V.; Glagoleva, A.Y.; Strygina, K.V.; Khlestkina, E.K.; Gerasimova, S.; Ibragimova, S.M.; Shatskaya, N.V.; Vasilyev, G.V.; Afonnikov, D.A.; Shmakov, N.A.; et al. Differential expression of NBS-LRR-encoding genes in the root transcriptomes of two *Solanum phureja* genotypes with contrasting resistance to *Globodera rostochiensis*. *BMC Plant Biol.* **2017**, *17*, 251. [[CrossRef](#)]
18. Kochetov, A.V.; Egorova, A.A.; Glagoleva, A.Y.; Strygina, K.V.; Khlestkina, E.K.; Gerasimova, S.V.; Shatskaya, N.V.; Vasilyev, G.V.; Afonnikov, D.A.; Shmakov, N.A.; et al. The mechanism of potato resistance to *Globodera rostochiensis*: Comparison of root transcriptomes of resistant and susceptible *Solanum phureja* genotypes. *BMC Plant Biol.* **2020**, *20*, 350. [[CrossRef](#)]
19. Sagi, M.S.; Deokar, A.A.; Tar'An, B. Genetic Analysis of NBS-LRR Gene Family in Chickpea and Their Expression Profiles in Response to *Ascochyta* Blight Infection. *Front. Plant Sci.* **2017**, *8*, 838. [[CrossRef](#)]
20. Vleeshouwers, V.G.A.A.; Oliver, R.P. Effectors as Tools in Disease Resistance Breeding Against Biotrophic, Hemibiotrophic, and Necrotrophic Plant Pathogens. *Mol. Plant-Microbe Interact.* **2015**, *2015*, 17–27. [[CrossRef](#)]
21. Brendolise, C.; Montefiori, M.; Dinis, R.; Peeters, N.; Storey, R.D.; Rikkerink, E.H. A novel hairpin library-based approach to identify NBS-LRR genes required for effector-triggered hypersensitive response in *Nicotiana benthamiana*. *Plant Methods* **2017**, *13*, 32. [[CrossRef](#)]
22. Tran, P.-T.; Choi, H.; Kim, S.-B.; Lee, H.-A.; Choi, D.; Kim, K.-H. A simple method for screening of plant NBS-LRR genes that confer a hypersensitive response to plant viruses and its application for screening candidate pepper genes against Pepper mottle virus. *J. Virol. Methods* **2014**, *201*, 57–64. [[CrossRef](#)]
23. Zhang, C.; Chen, H.; Cai, T.; Deng, Y.; Zhuang, R.; Zhang, N.; Zeng, Y.; Zheng, Y.; Tang, R.; Pan, R.; et al. Overexpression of a novel peanut NBS-LRR gene AhRRS5 enhances disease resistance to *Ralstonia solanacearum* in tobacco. *Plant Biotechnol. J.* **2017**, *15*, 39–55. [[CrossRef](#)]
24. Wei, C.; Chen, J.; Kuang, H. Dramatic Number Variation of R Genes in Solanaceae Species Accounted for by a Few R Gene Subfamilies. *PLoS ONE* **2016**, *11*, e0148708. [[CrossRef](#)]
25. Qian, L.-H.; Zhou, G.-C.; Sun, X.-Q.; Lei, Z.; Zhang, Y.-M.; Xue, J.-Y.; Hang, Y.-Y. Distinct Patterns of Gene Gain and Loss: Diverse Evolutionary Modes of NBS-Encoding Genes in Three Solanaceae Crop Species. *G3 Genes Genomes Genet.* **2017**, *7*, 1577–1585. [[CrossRef](#)]
26. Chae, E.; Bomblies, K.; Kim, S.-T.; Karelina, D.; Zaidem, M.; Ossowski, S.; Martín-Pizarro, C.; Laitinen, R.A.; Rowan, B.A.; Tenenboim, H.; et al. Species-wide Genetic Incompatibility Analysis Identifies Immune Genes as Hot Spots of Deleterious Epistasis. *Cell* **2014**, *159*, 1341–1351. [[CrossRef](#)]
27. Jia, H.; Xue, S.; Lei, L.; Fan, M.; Peng, S.; Li, T.; Nagarajan, R.; Carver, B.; Ma, Z.; Deng, J.; et al. A semi-dominant NLR allele causes whole-seedling necrosis in wheat. *Plant Physiol.* **2021**, *186*, 483–496. [[CrossRef](#)]

28. Barragan, A.C.; Collenberg, M.; Wang, J.; Lee, R.R.Q.; Cher, W.Y.; Rabanal, F.A.; Ashkenazy, H.; Weigel, D.; Chae, E. A Truncated Singleton NLR Causes Hybrid Necrosis in *Arabidopsis thaliana*. *Mol. Biol. Evol.* **2021**, *38*, 557–574. [\[CrossRef\]](#)
29. Ma, J.; Hancock, W.G.; Nifong, J.M.; Kernodle, S.P.; Lewis, R.S. Identification and editing of a hybrid lethality gene expands the range of interspecific hybridization potential in *Nicotiana*. *Theor. Appl. Genet.* **2020**, *133*, 2915–2925. [\[CrossRef\]](#)
30. Deng, J.; Fang, L.; Zhu, X.; Zhou, B.; Zhang, T. A CC-NBS-LRR gene induces hybrid lethality in cotton. *J. Exp. Bot.* **2019**, *70*, 5145–5156. [\[CrossRef\]](#)
31. Jiang, N.; Cui, J.; Hou, X.; Yang, G.; Xiao, Y.; Han, L.; Meng, J.; Luan, Y. Sl-lncRNA15492 interacts with Sl-miR482a and affects *Solanum lycopersicum* immunity against *Phytophthora infestans*. *Plant J.* **2020**, *103*, 1561–1574. [\[CrossRef\]](#)
32. Jiang, N.; Cui, J.; Shi, Y.; Yang, G.; Zhou, X.; Hou, X.; Meng, J.; Luan, Y. Tomato lncRNA23468 functions as a competing endogenous RNA to modulate NBS-LRR genes by decoying miR482b in the tomato-*Phytophthora infestans* interaction. *Hortic. Res.* **2019**, *6*, 1–11. [\[CrossRef\]](#)
33. Hong, Y.-H.; Meng, J.; Zhang, M.; Luan, Y.-S. Identification of tomato circular RNAs responsive to *Phytophthora infestans*. *Gene* **2020**, *746*, 144652. [\[CrossRef\]](#)
34. Parker, M.T.; Knop, K.; Zacharaki, V.; Sherwood, A.V.; Tome, D.; Yu, X.; Martin, P.G.; Beynon, J.; Michaels, S.; Barton, G.J.; et al. Widespread premature transcription termination of *Arabidopsis thaliana* NLR genes by the spen protein FPA. *eLife* **2021**, *10*, 65537. [\[CrossRef\]](#)
35. Cantalapiedra, C.; Contreras-Moreira, B.; Silvar, C.; Perovic, D.; Ordon, F.; Gracia, M.P.; Igartua, E.; Casas, A.M. A Cluster of Nucleotide-Binding Site–Leucine-Rich Repeat Genes Resides in a Barley Powdery Mildew Resistance Quantitative Trait Loci on 7HL. *Plant Genome* **2016**, *9*. [\[CrossRef\]](#)
36. Lee, R.R.; Chae, E. Variation Patterns of NLR Clusters in *Arabidopsis thaliana* Genomes. *Plant Commun.* **2020**, *1*, 100089. [\[CrossRef\]](#)
37. Göktay, M.; Fulgione, A.; Hancock, A.M. A New Catalog of Structural Variants in 1301 *A. thaliana* Lines from Africa, Eurasia, and North America Reveals a Signature of Balancing Selection at Defense Response Genes. *Mol. Biol. Evol.* **2021**, *38*, 1498–1511. [\[CrossRef\]](#)
38. Prakash, C.; Trognitz, F.C.; Venhuizen, P.; Von Haeseler, A.; Trognitz, B. A compendium of genome-wide sequence reads from NBS (nucleotide binding site) domains of resistance genes in the common potato. *Sci. Rep.* **2020**, *10*, 1–21. [\[CrossRef\]](#)
39. Potato Genome Sequencing Consortium. Genome sequence and analysis of the tuber crop potato. *Nature* **2011**, *475*, 189–195. [\[CrossRef\]](#)
40. Gavrilenko, T.A.; Klimenko, N.S.; Antonova, O.Y.; Lebedeva, V.A.; Evdokimova, Z.Z.; Gadjiyev, N.M.; Apalikova, O.V.; Alpatyeva, N.V.; Kostina, L.I.; Zoteyeva, N.M.; et al. Molecular screening of potato varieties bred in the northwestern zone of the Russian Federation. *Vavilovskii Zhurnal Genet. I Sel. Vavilov J. Genet. Breed.* **2018**, *22*, 35–45. [\[CrossRef\]](#)
41. Antonova, O.Y.; Klimenko, N.S.; Evdokimova, Z.Z.; Kostina, L.I.; Gavrilenko, T.A. Finding RB/Rpi-blb1/Rpi-sto1-like sequences in conventionally bred potato varieties. *Vavilov J. Genet. Breed.* **2018**, *22*, 693–702. [\[CrossRef\]](#)
42. Naess, S.K.; Bradeen, J.M.; Wielgus, S.M.; Haberlach, G.T.; McGrath, J.M.; Helgeson, J.P. Resistance to late blight in *Solanum bulbocastanum* is mapped to chromosome 8. *Theor. Appl. Genet.* **2000**, *101*, 697–704. [\[CrossRef\]](#)
43. Song, J.; Bradeen, J.M.; Naess, S.K.; Raasch, J.A.; Wielgus, S.M.; Haberlach, G.T.; Liu, J.; Kuang, H.; Austin-Phillips, S.; Buell, C.R.; et al. Gene RB cloned from *Solanum bulbocastanum* confers broad spectrum resistance to potato late blight. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 9128–9133. [\[CrossRef\]](#)
44. Van Der Vossen, E.; Sikkema, A.; Hekkert, B.T.L.; Gros, J.; Stevens, P.; Muskens, M.; Wouters, D.; Pereira, A.; Stiekema, W.; Allefs, S. An ancient R gene from the wild potato species *Solanum bulbocastanum* confers broad-spectrum resistance to *Phytophthora infestans* cultivated potato and tomato. *Plant J.* **2003**, *36*, 867–882. [\[CrossRef\]](#)
45. Vleeshouwers, V.G.A.A.; Rietman, H.; Krenek, P.; Champouret, N.; Young, C.; Oh, S.-K.; Wang, M.; Bouwmeester, K.; Vosman, B.; Visser, R.G.F.; et al. Effector Genomics Accelerates Discovery and Functional Profiling of Potato Disease Resistance and *Phytophthora infestans* Avirulence Genes. *PLoS ONE* **2008**, *3*, e2875. [\[CrossRef\]](#)
46. Wang, M.; Allefs, S.; Berg, R.G.V.D.; Vleeshouwers, V.G.A.A.; van der Vossen, E.A.G.; Vosman, B. Allele mining in *Solanum*: Conserved homologues of Rpi-blb1 are identified in *Solanum stoloniferum*. *Theor. Appl. Genet.* **2008**, *116*, 933–943. [\[CrossRef\]](#)
47. Lokossou, A.A.; Rietman, H.; Wang, M.; Krenek, P.; van der Schoot, H.; Henken, B.; Hoekstra, R.; Vleeshouwers, V.G.A.A.; van der Vossen, E.A.G.; Visser, R.G.F.; et al. Diversity, Distribution, and Evolution of *Solanum bulbocastanum* Late Blight Resistance Genes. *Mol. Plant-Microbe Interact.* **2010**, *23*, 1206–1216. [\[CrossRef\]](#) [\[PubMed\]](#)
48. Haverkort, A.J.; Boonekamp, P.M.; Hutten, R.; Jacobsen, E.; Lotz, L.A.P.; Kessel, G.J.T.; Vossen, J.H.; Visser, R.G.F. Durable Late Blight Resistance in Potato Through Dynamic Varieties Obtained by Cisgenesis: Scientific and Societal Advances in the DuRPh Project. *Potato Res.* **2016**, *59*, 35–66. [\[CrossRef\]](#)
49. Khiutti, A.; Spooner, D.M.; Jansky, S.H.; Halterman, D.A. Testing Taxonomic Predictivity of Foliar and Tuber Resistance to *Phytophthora infestans* in Wild Relatives of Potato. *Phytopathology* **2015**, *105*, 1198–1205. [\[CrossRef\]](#)
50. Cruickshank, G.; Stewart, H.E.; Wastie, R.L. An illustrated assessment key for foliage blight of potatoes. *Potato Res.* **1982**, *25*, 213–214. [\[CrossRef\]](#)
51. Chen, S.; Zhou, Y.; Chen, Y.; Gu, J. fastp: An ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* **2018**, *34*, i884–i890. [\[CrossRef\]](#)
52. Lin, H.-N.; Hsu, W.-L. DART: A fast and accurate RNA-seq mapper with a partitioning strategy. *Bioinformatics* **2017**, *34*, 190–197. [\[CrossRef\]](#)

53. Howe, K.L.; Contreras-Moreira, B.; De Silva, N.; Maslen, G.; Akanni, W.; Allen, J.; Alvarez-Jarreta, J.; Barba, M.; Bolser, D.M.; Cambell, L.; et al. Ensembl Genomes 2020—enabling non-vertebrate genomic research. *Nucleic Acids Res.* **2020**, *48*, D689–D695. [\[CrossRef\]](#)
54. Hardigan, M.A.; Laimbeer, F.P.E.; Newton, L.; Crisovan, E.; Hamilton, J.; Vaillancourt, B.; Wiegert-Rininger, K.; Wood, J.; Douches, D.S.; Farré, E.M.; et al. Genome diversity of tuber-bearing *Solanum* uncovers complex evolutionary history and targets of domestication in the cultivated potato. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E9999–E10008. [\[CrossRef\]](#)
55. Liao, Y.; Smyth, G.K.; Shi, W. featureCounts: An efficient general purpose program for assigning sequence reads to genomic features. *Bioinformatics* **2014**, *30*, 923–930. [\[CrossRef\]](#)
56. Robinson, M.D.; McCarthy, D.J.; Smyth, G.K. edgeR: A Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* **2010**, *26*, 139–140. [\[CrossRef\]](#)
57. Roulin, A.C.; Wu, M.; Pichon, S.; Arbore, R.; Kühn-Bühlmann, S.; Kölliker, M.; Walser, J.-C. De Novo Transcriptome Hybrid Assembly and Validation in the European Earwig (Dermaptera, Forficula auricularia). *PLoS ONE* **2014**, *9*, e94098. [\[CrossRef\]](#)
58. Shmakov, N.A. Improving the quality of barley transcriptome de novo assembling by using a hybrid approach for lines with varying spike and stem coloration. *Vavilov J. Genet. Breed.* **2021**, *25*, 30–38. [\[CrossRef\]](#)
59. Grabherr, M.G.; Haas, B.J.; Yassour, M.; Levin, J.Z.; Thompson, D.A.; Amit, I.; Adiconis, X.; Fan, L.; Raychowdhury, R.; Zeng, Q.; et al. Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nat. Biotechnol.* **2011**, *29*, 644–652. [\[CrossRef\]](#)
60. Robertson, G.; Schein, J.; Chiu, R.; Corbett, R.; Field, M.; Jackman, S.D.; Mungall, K.; Lee, S.; Okada, H.M.; Qian, J.Q.; et al. De novo assembly and analysis of RNA-seq data. *Nat. Methods* **2010**, *7*, 909–912. [\[CrossRef\]](#)
61. Edgar, R.C. Search and clustering orders of magnitude faster than BLAST. *Bioinformatics* **2010**, *26*, 2460–2461. [\[CrossRef\]](#) [\[PubMed\]](#)
62. Simão, F.A.; Waterhouse, R.M.; Ioannidis, P.; Kriventseva, E.V.; Zdobnov, E.M. BUSCO: Assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* **2015**, *31*, 3210–3212. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Wu, T.D.; Watanabe, C.K. GMAP: A genomic mapping and alignment program for mRNA and EST sequences. *Bioinformatics* **2005**, *21*, 1859–1875. [\[CrossRef\]](#)
64. Sato, S.; Tabata, S.; Hirakawa, H.; Asamizu, E.; Shirasawa, K.; Isobe, S.; Kaneko, T.; Nakamura, Y.; Shibata, D.; Aoki, K.; et al. The tomato genome sequence provides insights into fleshy fruit evolution. *Nature* **2012**, *485*, 635–641. [\[CrossRef\]](#)
65. Emms, D.M.; Kelly, S. OrthoFinder: Phylogenetic orthology inference for comparative genomics. *Genome Biol.* **2019**, *20*, 1–14. [\[CrossRef\]](#)
66. Katoh, K.; Rozewicki, J.; Yamada, K.D. MAFFT online service: Multiple sequence alignment, interactive sequence choice and visualization. *Brief. Bioinform.* **2019**, *20*, 1160–1166. [\[CrossRef\]](#)
67. Nguyen, L.-T.; Schmidt, H.A.; Von Haeseler, A.; Minh, B.Q. IQ-TREE: A Fast and Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. *Mol. Biol. Evol.* **2015**, *32*, 268–274. [\[CrossRef\]](#)
68. Bray, N.L.; Pimentel, H.; Melsted, P.; Pachter, L. Near-optimal probabilistic RNA-seq quantification. *Nat. Biotechnol.* **2016**, *34*, 525–527. [\[CrossRef\]](#)
69. Chen, Y.; Lun, A.T.L.; Smyth, G.K. From reads to genes to pathways: Differential expression analysis of RNA-Seq experiments using Rsubread and the edgeR quasi-likelihood pipeline. *F1000Research* **2016**, *5*, 1438. [\[CrossRef\]](#)
70. Steuernagel, B.; Jupe, F.; Witek, K.; Jones, J.D.; Wulff, B.B. NLR-parser: Rapid annotation of plant NLR complements. *Bioinformatics* **2015**, *31*, 1665–1667. [\[CrossRef\]](#)
71. Eddy, S.R. Accelerated Profile HMM Searches. *PLoS Comput. Biol.* **2011**, *7*, e1002195. [\[CrossRef\]](#)
72. Finn, R.D.; Coghill, P.; Eberhardt, R.Y.; Eddy, S.R.; Mistry, J.; Mitchell, A.L.; Potter, S.C.; Punta, M.; Qureshi, M.; Sangrador-Vegas, A.; et al. The Pfam protein families database: Towards a more sustainable future. *Nucleic Acids Res.* **2016**, *44*, D279–D285. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Shen, W.; Le, S.; Li, Y.; Hu, F. SeqKit: A Cross-Platform and Ultrafast Toolkit for FASTA/Q File Manipulation. *PLoS ONE* **2016**, *11*, e0163962. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Pronozin, A.Y.; Bragina, M.K.; Salina, E.A. Crop pangenomes. *Vavilov J. Genet. Breed.* **2021**, *25*, 57–63. [\[CrossRef\]](#)
75. Golicz, A.; Batley, J.; Edwards, D. Towards plant pangenomics. *Plant Biotechnol. J.* **2015**, *14*, 1099–1105. [\[CrossRef\]](#) [\[PubMed\]](#)
76. He, M.; Zhou, Y.; Ye, G.; Zheng, J.; Meng, Y.; Wang, J.; Shan, W. Serial Transcriptome Analysis Reveals Genes Associated with Late Blight Resistance in Potato Cultivar Qingshu. *Agronomy* **2021**, *11*, 1919. [\[CrossRef\]](#)
77. Bradeen, J.M.; Iorizzo, M.; Molloy, D.S.; Raasch, J.; Kramer, L.C.; Millett, B.P.; Austin-Phillips, S.; Jiang, J.; Carputo, D. Higher Copy Numbers of the PotatoRBTransgene Correspond to Enhanced Transcript and Late Blight Resistance Levels. *Mol. Plant-Microbe Interact.* **2009**, *22*, 437–446. [\[CrossRef\]](#) [\[PubMed\]](#)
78. Kramer, L.C.; Choudoir, M.J.; Wielgus, S.M.; Bhaskar, P.B.; Jiang, J. Correlation Between Transcript Abundance of the RB Gene and the Level of the RB-Mediated Late Blight Resistance in Potato. *Mol. Plant-Microbe Interact.* **2009**, *22*, 447–455. [\[CrossRef\]](#) [\[PubMed\]](#)
79. Kochetov, A.V.; Titov, S.; Kolodyazhnaya, Y.S.; Komarova, M.L.; Koval, V.S.; Makarova, N.N.; Il'Yinskyi, Y.Y.; Trifonova, E.A.; Shumny, V. Tobacco Transformants Bearing Antisense Suppressor of Proline Dehydrogenase Gene, Are Characterized by Higher Proline Content and Cytoplasm Osmotic Pressure. *Russ. J. Genet.* **2004**, *40*, 216–218. [\[CrossRef\]](#)

80. Trifonova, E.A.; Sapotsky, M.V.; Komarova, M.L.; Scherban, A.B.; Shumny, V.K.; Polyakova, A.M.; Lapshina, L.A.; Kochetov, A.V.; Malinovskiy, V.I. Protection of transgenic tobacco plants expressing bovine pancreatic ribonuclease against tobacco mosaic virus. *Plant Cell Rep.* **2007**, *26*, 1121–1126. [[CrossRef](#)]
81. Sugawara, T.; Trifonova, E.A.; Kochetov, A.V.; Kanayama, Y. Expression of an extracellular ribonuclease gene increases resistance to Cucumber mosaic virus in tobacco. *BMC Plant Biol.* **2016**, *16*, 246. [[CrossRef](#)]
82. Li, X.; Zhang, Y.; Yin, L.; Lu, J. Overexpression of pathogen-induced grapevine TIR-NB-LRR gene VaRGA1 enhances disease resistance and drought and salt tolerance in *Nicotiana benthamiana*. *Protoplasma* **2017**, *254*, 957–969. [[CrossRef](#)]
83. Ghislain, M.; Byarugaba, A.A.; Magembe, E.; Njoroge, A.; Rivera, C.; Román, M.L.; Tovar, J.C.; Gamboa, S.; Forbes, G.A.; Kreuze, J.; et al. Stacking three late blight resistance genes from wild species directly into African highland potato varieties confers complete field resistance to local blight races. *Plant Biotechnol. J.* **2018**, *17*, 1119–1129. [[CrossRef](#)] [[PubMed](#)]
84. Morales, J.G.; Gaviria, A.E.; Gilchrist, E. Allelic Variation and Selection in Effector Genes of *Phytophthora infestans* (Mont.) de Bary. *Pathogens* **2020**, *9*, 551. [[CrossRef](#)] [[PubMed](#)]
85. Wang, W.; Liu, X.; Han, T.; Li, K.; Qu, Y.; Gao, Z. Differential Potential of *Phytophthora capsici* Resistance Mechanisms to the Fungicide Metalaxyl in Peppers. *Microorganisms* **2020**, *8*, 278. [[CrossRef](#)]
86. Zheng, J.; Duan, S.; Armstrong, M.R.; Duan, Y.; Xu, J.; Chen, X.; Hein, I.; Jin, L.; Li, G. New Findings on the Resistance Mechanism of an Elite Diploid Wild Potato Species JAM1-4 in Response to a Super Race Strain of *Phytophthora infestans*. *Phytopathology* **2020**, *110*, 1375–1387. [[CrossRef](#)]
87. Frolova, T.S.; Cherenko, V.A.; Sinitsyna, O.I.; Kochetov, A.V. Genetic aspects of potato resistance to phytophthorosis. *Vavilov J. Genet. Breed.* **2021**, *25*, 164–170. [[CrossRef](#)]