



Review

Achievements and Challenges towards a Sustainable Conservation and Use of ‘Galega vulgar’ *Olea europaea* Variety

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Abstract: The cultivated *Olea europaea* L., or olive tree, is an ancient crop extremely relevant for the bioeconomy of Mediterranean countries, especially for Portugal. With orchard modernization, Portugal has the potential to become the third-largest producer of olive oil over the next decade. In this country, the main national variety is ‘Galega vulgar,’ characterized by an excellent olive oil quality, rusticity and tolerance to drought. Nevertheless, its production has suffered a reduction due to replacement by foreign varieties. The present narrative review offers an overall perspective of present gaps and challenges to the conservation and use of ‘Galega vulgar.’ Existing information about traditional and innovative olive production systems, and the importance of life cycle thinking approaches for a sustainable olive chain were synthesized, focusing particularly in the last 10 years. Furthermore, the olive molecular characterization advances and their breeding importance were also evaluated considering their application to this Portuguese variety. To ensure an efficient and sustainable exploitation of the ‘Galega vulgar,’ we propose that efforts should concentrate on the characterization of the existing variability and development of genotype to phenotype prediction tools, integrating detailed molecular marker genotypic and environmental characterization, to support better informed conservation and breeding decisions in a multi-environment context.

Keywords: bioeconomy; genetic resources; genotype-by-environment-interaction; LCA; molecular markers; *Olea europaea*

1. Introduction

Olive (*Olea europaea* L., subsp. *europaea*, var. *europaea*) is a multifunctional long-living Mediterranean tree species with an extraordinarily ancient history and tradition. This crop holds an important role both for table olive and oil production and for shaping and protecting the regional landscape. Moreover, it assumes an enormous relevance on human nutrition and rural lifestyle [1]. The olive tree genetic breeding started when human populations domesticated wild olives, selecting the best phenotypes. This selection continued across time, throughout the Mediterranean Basin, leading to the present complex genetic relationships existing among varieties [2,3].

Currently, *O. europaea* is the most cultivated temperate fruit crop worldwide and is characterized by a vast heritage of clonally propagated traditional varieties [4]. Important socioeconomic changes in several Mediterranean countries led to significant technological improvement in olive cultivation, which increased the risk of genetic erosion on the traditional olive germplasm. This genetic erosion phenomenon is also occurring in Portugal, especially affecting the main national variety ‘Galega vulgar,’

which has been replaced despite its excellent olive oil qualities. Since high-yielding and low-vigor modern varieties, more suitable for mechanically harvested plantations, are preferred to the traditional varieties, the characterization and conservation of existing traditional varieties diversity is essential to ensure their sustainable use in the future [5].

In this narrative review we searched international data bases for English-language articles, and national state technical reports and University dissertations (in Portuguese). Keywords used covered the following topics: Economic and social impact of *Olea europaea*; Olive Production Systems, LCA, Genetic Diversity, Domestication and origin, Olive germplasm conservation, Conventional and precision breeding techniques in olive, Marker assisted selection, GxE, Genotype-to-phenotype (G2P) models. Search was focused mainly on the last 10 years (2010–2020), with additional older studies scanned from the obtained reference list, giving a special emphasis to ‘Galega vulgar’ variety.

Thus, the present narrative review provides an overview of the millenary culture of *O. europaea*, the changes that olive production systems have been confronted with over time and how life cycle thinking approaches might tackle these modifications and contribute to a more sustainable olive chain. Furthermore, the olive molecular characterization efforts that have taken place in different olive varieties and their importance for breeding programs are also reviewed. A special emphasis is given to the ‘Galega’ variety, highlighting the existing gaps and future challenges concerning this variety conservation and use.

2. Olive Tree: A Millenary Crop with a Massive Economic Impact Worldwide

The olive tree crop, together with the vine crop, represent the oldest perennial crops for human food [3]. Across time, olive trees assumed a special, even sacred, symbolic meaning in Mediterranean communities. Beyond the immense importance that olives and olive oil hold in gastronomy, considered today as cornerstones of the healthy Mediterranean diet, the olive tree is used during many spiritual celebrations [6]. More recently, olive has been recognized as a functional food due to its various bioactive constituents, particularly the phenolic compounds (such as oleuropein, the major phenolic constituent of *O. europaea*), and their pharmacological activities [7]. The olive high content of monounsaturated fatty acids (mostly oleic acid, representing 70% to 80% of the total olive oil) plays an important role in preventing cardiovascular diseases when consumed. It is a stimulant of the biliary tract (improving digestion and intestinal function) and encourages the absorption of fat-soluble vitamins (A, D, K and E), being rich in vitamin E [8]. Because of this dietetic value, health benefits and economic relevance, there is a high market demand and a regular increase in olive trees cultivated areas [9].

Mediterranean countries are the main consumers of olive oil, representing around 63% of world intake, which correspond to 1836.7 thousand tons [10]. However, olive oil consumption has been increasing in countries outside this region. This group is led by the United States, with about 11% of world consumption—Figure 1a. Regarding the total intake in the European Union (EU), Spain is the main consumer followed by Italy that together consume almost a third of the worldwide total (Figure 1b).

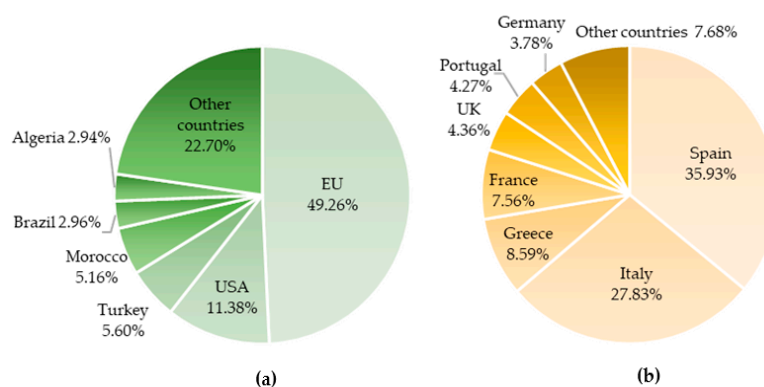


Figure 1. Consumption of olive oil. Provisory data from the campaign year 2018/2019: (a) World consumption; (b) European consumption (data from International Olive Council (IOC) [10]).

Considering the production of olive oil, the main producer is Spain (about 56% of world total and 79.08% of EU production, corresponding to 1789.9 thousand tons), followed by Greece and Italy—Figure 2a,b.

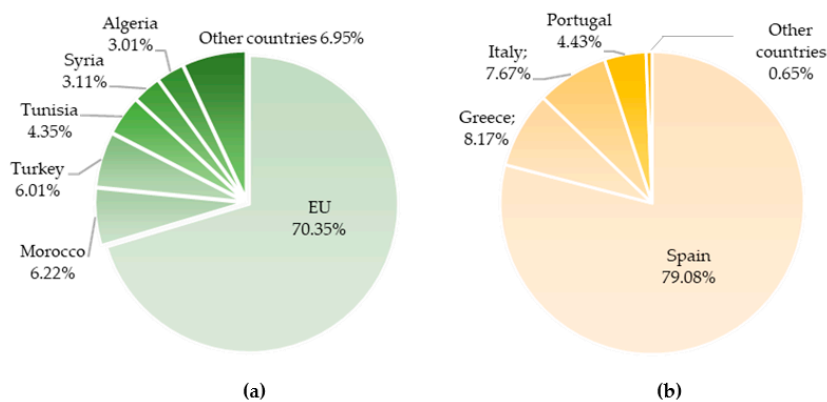


Figure 2. Production of olive oil. Provisory data from the campaign year 2018/2019: (a): World production; (b): European production (data from IOC [10]).

In the case of Portugal, in 2017/2018 and 2018/2019, this country ranked seventh in the world production of olive oil ($\approx 4\%$), and the fourth largest olive producer in Europe, behind Spain, Italy and Greece [10]. However, with the modernization of Portuguese olive groves, this country has the potential to become the third-largest producer of olive oil in the next 10 years. In fact, the olive oil production in Portugal has increased over the last decade, generating in the last three years a turnover 2.5 times higher (€620 million) than the one recorded between 2010 and 2012 [11].

In first decades of the last century, there was an expansion of the olive orchards and the production of olives and olive oil in Portugal. However, during the 1960s, olive cultivation suffered a drastic decrease due to changes in policies in the olive sector, the increase of production costs and the market introduction of other plant oils [12]. It was only in 1986, when Portugal became an EU member, that the modernization in the olive sector started, with the approval of the National Plan for the olive crop. With funds from EU, Portugal was able to renew the production models and started a technological revolution in its orchards, leading to an increased productivity and better quality of Portuguese olive oil [11]. In fact, the Portuguese olives and olive oil production already exceeds the production obtained in 1950/1960 (around 120 thousand tons of olive oil) [11], recording that in 2017, the total olive oil, olives for oil and table olives production was 134.8, 858.4 and 134.8 thousand tons, respectively (Figure 3) [10,13].

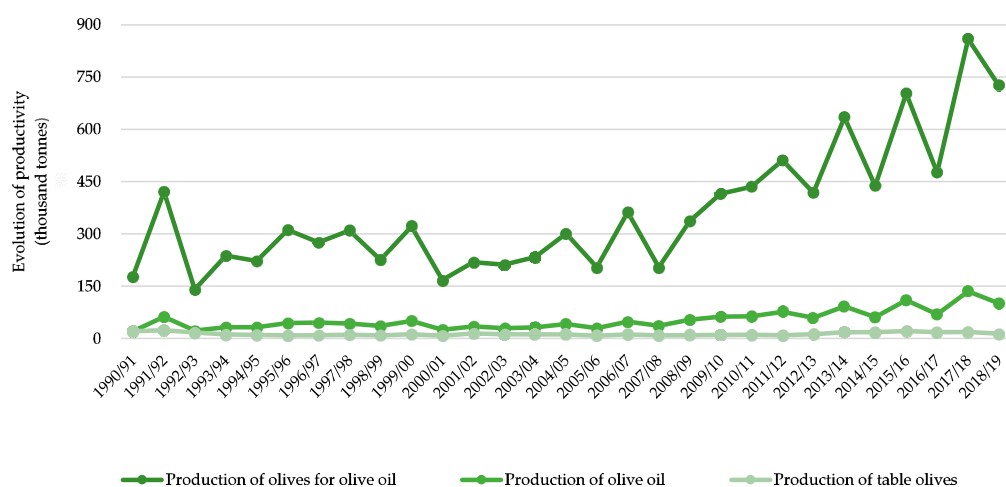


Figure 3. Evolution of production of olive oil and olives in the Portuguese olive sector since the 1990/1991 campaign year (data from IOC [10] and Instituto Nacional de Estatística (INE) [13]).

In Portugal, there are 361.177 thousand hectares (ha) of olive groves, with around 97.6% of them used for olive oil production. In this way, table olives production occupies only 8773 ha [13]. The main national producing region both for table olives and for olive oil is the “Alentejo” region. This southern territory holds a production of 6.9 thousand tons for table olives and around 551.4 thousand tons for olives for oil (Figure 4a). Concerning the surface occupied in Portugal for olive orchards, “Alentejo” is still the principal region for olive oil (184.936 thousand ha) and for table olives (3258 ha) (Figure 4b) [13].

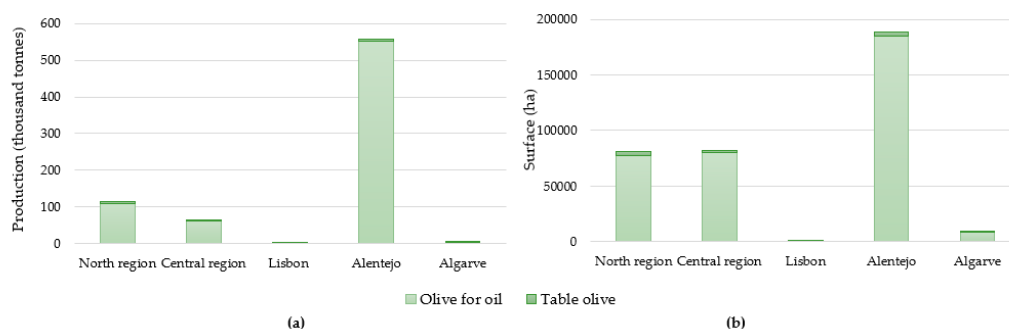


Figure 4. Production (thousand tons) and surface (ha) of olives for oil and table olives by NUTS (nomenclature of territorial units for statistics). Provisory data from the campaign year 2018/2019: (a) Portuguese production of olives for oil and table olives; (b) surface of olives for olive oil and table olives occupied in Portugal (data from INE [13]).

In Portugal, as in the other traditional olive-growing countries, there is a high number of olive varieties. It is estimated that worldwide this number may exceed 3000 with the particularity that only a small number is grown in more than one region [14].

2.1. ‘Galega Vulgar’, the Most Important Portuguese Olive Variety

In Portugal, the production of olive oil and table olive is mostly concentrated in 22 different varieties: ‘Galega vulgar’ or ‘Galega’, ‘Carrasquenha’, ‘Redondil’, ‘Azeitoneira’ or ‘Azeitoura’, ‘Branquita’ or ‘Blanqueta’, ‘Conserva de Elvas’, ‘Negrinha’, ‘Madural’, ‘Cobrançosa’, ‘Verdeal Transmontana’ or ‘Verdeal de Serpa’, ‘Cordovil de Castelo Branco’, ‘Bical de Castelo Branco’, ‘Maçanilha’ or ‘Maçanilha Fina’, ‘Hojiblanca’ and ‘Gordal’ [15].

The main national variety is ‘Galega vulgar’, also called ‘Galega’ (accounting for about 60% of all olive trees in the country) [16] and is associated with five of the six national PDOs (Protected Designations of Origin) regions [17]. The PDO oils are produced in defined geographical areas, with a characteristic soil and climate combination, and are exclusively made from certain olive varieties. These factors, together with the management of the orchard, harvesting and transport to the mill and the oil-makings conditions, give rise to olive oils with unique chemical and sensory characteristics that allow to distinguish them from other oils [18].

A high rusticity and alternate bearing characterize the ‘Galega’ variety (Figure 5). With a low weight fruit (<2 g), an endocarp with a low to medium weight (<0.30 g to 0.30–0.45 g) and a low flesh to stone ratio (5.6 on average) [15,19,20], it stands out for its excellent quality olive oil, and resistance to drought, despite median productivity, partly due to the sensitivity to pests and diseases [19]. ‘Galega’ has a low yield in olive oil (below 18%); this oil is characterized by a low percentage of linoleic acid and a high stability [19,21]. This olive variety shows a high sensitivity to the anthracnose fungus (*Colletotrichum* spp.), oleander knot (*Pseudomonas savastanoi* pv. *savastanoi*), olive fruit fly (*Bactrocera oleae*) and the black scale (*Saissetia oleae*). It also exhibits some susceptibility to olive peacock spot (*Spilocaea oleagina*) [19].



Figure 5. ‘Galega’ olive tree, the main Portuguese variety: (a) ‘Galega’ in a traditional olive orchard in Portugal; (b) Abaxial and adaxial surfaces of ‘Galega’ leaves; (c) Detail of the ‘Galega’ inflorescence before the pollination; (d) Flowers of ‘Galega’; (e) Drupes in ‘Galega’ tree; (f) Endocarp inside the drupe and the mesocarp; (g) Endocarp of ‘Galega.’ Photographs by Hélia Sales.

The ‘Galega’ variety presents a high potential for improvement/breeding due to its intra-variety diversity [22]. Significant progress in ‘Galega’ research has been attained in last years by several national teams, particularly in the areas of ecophysiology, in vitro culture, genetic resources characterization, improvement in production systems and cultural techniques, phytosanitary protection and technology, quality and market organization (Table 1).

Table 1. Research developed considering the ‘Galega’ variety in the last years.

Area	Applications	References
Ecophysiology	Water use efficiency; salinity stress	[23,24]
	Rooting	[25]
	Effect of chemical and natural compounds application; influence UV-B radiation	[26–28]
	Phenology studies	[29]
	Different types of pollination; profilin polymorphism	[26,30,31]
In vitro culture	Pollen viability and germination	[32]
	Gene characterization and expression	[33,34]
	Adventitious root formation	[35,36]
	Micropropagation	[37,38]
Genetic resources	Olive varieties identification and genetic relationships	[39–47]
	Genetic variability; genotyping and genetic mapping	[22,44,48–53]
Improvement in production systems and cultural techniques	Mechanical pruning and harvesting; effect of rejuvenation pruning	[54,55]
Phytosanitary protection	Richness and diversity of fungal communities	[56–58]
	Susceptibility to fungal diseases; response to pathogens attack	[59–62]
Technology, quality and markets	Analytical characterization of olive oil compounds; certification and olive oil traceability	[63,64]
	Effect of blending and storage time of olive oil	[65]
	Effect of fertilization on olive oil polyphenol, sterol and wax content	[66]
	Extraction of monovarietal olive oils with natural compounds	[67]
	Physicochemical, nutritional and microbiological characterization; health effects; agro-environmental factors on the olive oil mineral content	[68–70]

The ‘Galega’ productivity currently ranges between 3 and 5 tones/ha (with up to 200 trees/ha in non-irrigated systems) [71]. Comparatively, foreigner varieties productivity is around 8 to 13 tones/ha in super intensive systems [72]. These data point out the importance and the need to improve the productivity of the ‘Galega’ olive trees, in order to hamper its genetic erosion, by increasing its attractiveness to farmers. Nevertheless, to make ‘Galega’ more competitive regarding foreign varieties, it is very important to maintain its high quality while improving its productivity and adaptation to modern production systems.

2.2. Recent Changes on Olive Production Systems and the Importance of a Life Cycle Assessment

Olive trees naturally grow in relatively dry, rustic environments with a Mediterranean climate, characterized by warm, dry summers and rainy, cool winters [73]; these factors shape traditional olive plantations [5].

Traditional olive varieties are the result of empirical selection processes performed by olive growers in a given combination of soil and climatic conditions, with subsequent preservation by vegetative multiplication [14]. In this way, traditional varieties hold a high genetic variability, mainly because of olive allogamy and predominantly self-incompatible nature, tree longevity and long tradition of cultivation [74]. Consequently, different Mediterranean regions are characterized by different local genetic variability, resulting from a mixture of environments and growing systems [75,76]. However, in recent years, the olive germplasm diversity has been modified drastically because of the changes on the determinant characteristics in the choice of a variety. The improvement of the socioeconomic conditions and the exponential progress of technology worldwide have been the principal boosters in a progressive transformation in olive growing techniques [5,77]. The adaptation to the environment, which was the main selection criterion in the past, has been presently replaced by productivity, higher yield in oil and regularity in production, at reduced costs [14]. Consequently, the traditional, manually harvested non-irrigated olive orchards are being replaced by new high-density irrigated and mechanically harvested modern olive plantations [5].

Countries such as Spain, where modern olive groves have been established intensively, have its olive-growing areas predominantly dominated by only a few varieties, with ‘Picual’, ‘Hojiblanca’ and ‘Arbequina’ as the most common [78]. Indeed, ‘Arbequina’ is not only a variety of choice in Spain, but all over the world, and the globalization of some olive varieties may give rise to the tendency to reduce the genetic variability in the modern olive groves even more [76]. In Portugal, this is also occurring, with the main traditional variety in the country (‘Galega’) being replaced, despite its superior-quality olive oil, probably leading to the loss of alleles or of combinations of alleles, in a clear case of genetic erosion [79,80]. Therefore, it is crucial to characterize, conserve and explore the still-existing traditional olive genetic resources in order to prevent the risk of further genetic erosion, towards an efficient use in breeding programs [76].

The replacement of traditional olive varieties by a few foreign ones not only entails negative impacts related to the loss of genetic variability but could also enhance the negative impacts on the environment, related with the different practices and techniques typically associated with these replacement foreigner varieties. Examples are the depletion of natural resources, land degradation, air emissions, waste generation, soil erosion and water scarcity [81,82]. In order to counteract this negative tendency and ensure the olive sector sustainability, life cycle thinking approaches and assessment methods have been applied, allowing a deeper understanding of the role of the impacts from a life cycle perspective. Additionally, these methodologies are fundamental tools to support decision-making in complex value chains, such as the ones related to bioeconomy [81].

A summary of the existing olive sector Life Cycle Assessment (LCA) studies may be found on Table 2, taking into consideration diverse scenarios at different stages of the olive chain.

Table 2. Life Cycle Assessment (LCA) studies performed in the olive sector in the last 10 years.

Aim of the Application of LCA Methodology	FU ¹	Country	References
olive oil production	1 ton of olives	Italy	[83]
	1 ha	Iran	[84]
	1 L EVOO ²	Italy	[85]
	1 L EVOO	Greece	[86]
	1 kg of olive oil	Jordan	[87]
	5 L EVOO	Italy	[88]
distribution processes of olive oil supply chain and packaging	1 L EVOO	Italy	[89]
	1 L of bottling capacity	Italy	[90]
	0.5 L bottle of VOO ³	Spain	[91]
olive oil industry waste treatments	1 L olive mill waste	Greece	[92]
	1 mg of olive solid waste	Australia	[93]
	1 ton of torrefied olive husk	Cyprus	[94]
	100 kg olive pomace	Spain	[95]
	1MJ of energy production	Spain	[96]
field agricultural practices	1 ha	Italy	[97,98]
	1 ton of olives	Spain	[99]
olive-harvesting practices	1 ha; 1 kg	Italy	[100]

¹ FU—Functional Unit ² EVOO—Extra virgin olive oil. ³ VOO—virgin olive oil.

From these different studies, the agricultural phase was the issue reported as being the most impactful in the environment (i.e., the principal hotspot in the olive oil cycle), mostly due to the utilization of fertilizers, irrigation and phytosanitary treatments [82]. Moreover, distribution and waste management were also considered crucial issues. However, due to the extremely variation of the distribution phase scenarios, they are often excluded from studies. The impacts of waste treatment are also highly variable between studies. Consequently, they might be considered as “avoided” impacts due to energy or nutrient recovery [82,83].

Although the LCA methodology has been applied to calculate and evaluate the potential environmental impacts of the recent changes on olive production systems from the extraction of the raw materials to the oil mill gate, an integrative methodology that takes into consideration environmental, economic and social constraints is still missing. Particularly in Portugal, LCA studies in the olive sector are almost inexistent hampering a deeper characterization and evaluation of the Portuguese olive sector and its principal impacts. Since the olive sector is one of the most valuable bioeconomic value chains in Portugal, the enrichment of Portuguese inventories and impact assessment methods in olive chain is of utmost importance. These characterizations, which will allow future comparisons and inclusions in international studies, together with a deeper characterization of the diversity of Portuguese olive genetic resources, will allow a better knowledge, management, improvement and conservation of the Portuguese olive heritage.

3. *Olea europaea* Diversity Evolution

3.1. Domestication and Ancestral Origin of Olive Varieties

The geographic origin of olive tree, its domestication and history of its early use are still under debate [101]. According to archaeological and genetic studies, the cultivated olive tree might have developed by domestication of its wild ancestor “oleaster” (*Olea europaea* subsp. *europaea* var. *sylvestris*), approximately 6000 years ago in the Near East [2,102]. Hence, with the discovery of clonal propagation techniques that allowed the vegetative propagation of the best specimens for human consumption, the olive cultivation was boosted across the Mediterranean Basin [101]. Under this scenario of

domestication, probably the wild olives behaved as pollen donors to the primary domesticated varieties, diminishing the deleterious effects that might occur in the inbreeding events and giving rise to locally adapted varieties [103].

However, recent evidences clearly indicate the existence of multiple centers of diversity across the Mediterranean area for cultivated olive, and it remains unclear whether the centers of diversity result from one or two local domestication events (Figure 6) [102]. Nuclear microsatellites characterization disclosed the existence of three main cultivated genepools (Q1, Q2 and Q3, Figure 6b), mostly corresponding to three geographic areas: the west (WW), center and east (WE) Mediterranean Basin (Figure 6a) [102,104,105].

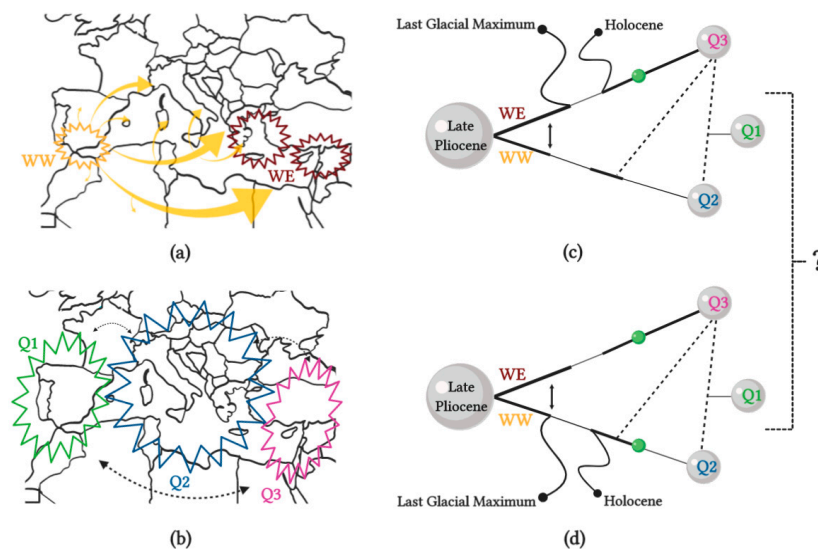


Figure 6. Single or multiple independent domestications for cultivated olive trees, the unanswered question: (a,b) distribution olive genepools in the Mediterranean Basin, accordingly to Besnard et al. [104] and Diez et al. [105], respectively; (c,d) simplified scenarios of domestication; (c) one single primary domestication event; (d) two independent primary domestication events. Green spheres indicate the primary domestication events, horizontal arrows indicate the possible occurrence of gene flow and the dashed horizontal connections indicate the admixture events (schematic representation made in ©BioRender—biorender.com).

Two simplified scenarios of domestication may be considered (Figure 6c,d), during the Late Pliocene with both western and eastern oleaster genepools diverging, with possible gene flow [2,106]. Later, during the Last Glacial Maximum and the Holocene, a population reduction occurred, followed by successive expansion. On the first scenario (Figure 6c), a primary domestication event (green sphere in the figure) is speculated to have taken place in the Eastern Mediterranean Basin, leading to Q3, while Q2 and Q1 were derived from admixture events between Q3 and WW (for Q2) or Q3 and Q2 (for Q1). On the second scenario (Figure 6d), two independent primary domestication events are thought to have occurred, one for Q3 in the Eastern Mediterranean Basin, and one for Q2 in the Central Mediterranean Basin [102].

Regardless of the primary origin of the olive varieties, since the genotypes were clonally propagated, they could be disseminated via human migration [103]. Although the migration history of olive trees is complex, Phoenicians seemed to have started disseminating the olive cultivation in the Iberian Peninsula, but it was the Roman Empire that was principally responsible for extending and encouraging this crop from east to west through both the northern and southern coasts of the Mediterranean Basin [101,107].

The domestication of olive trees promoted larger fruits and seeds, with more pointed ends in cultivated forms, in comparison with wild forms [108]. Nevertheless, some olive varieties such as ‘Galega’ still show considerable morphological diversity with round to acute endocarps of medium to

small weights. This could be a sign of an ongoing domestication process or the effect of a long-term clonal propagation process, reducing the number of generations from the wild ancestor or facilitating the existence of overlapping generations [103]. Corroborating this hypothesis, the ‘Galega’ reference sample at the World Olive Germplasm Bank (WOGB) appears as a blend of West and Central Mediterranean germplasm [109] and cultivated-wild admixture [105], and thus, it is possible that ‘Galega’ still holds undomesticated and interesting novel combinations of traits, which might be beneficial for the selection of interesting phenotypes in breeding programs.

As far as the origin of ‘Galega’ is concerned, little is known. Gemas et al. [22] hypothesized that the morphologically diverse ‘Galega’ had a polyclonal origin, supported by the occurrence of somatic mutations and possible sexual reproduction. Hence, the ‘Galega’ variety might have been originated by a naturalization process or as result of hybridization with wild olive trees. However, the study that led to these conclusions was performed using a small non-representative set of the whole ‘Galega’ diversity in terms of tree ages and geographical locations, and thus, a more comprehensive characterization is still needed to unravel the ancestral origin of the ‘Galega’ variety.

Knowledge on the genetic diversity, associated to the morphological differences, detected among and especially within olive varieties is essential to optimize the exploitation of the existing germplasm resources in a long-term successful breeding program.

3.2. The Use of Molecular Markers on Olive Diversity Characterization

Because of the early domestication and following dissemination throughout the Mediterranean area, the olive crop holds a vast number of varieties [110]. Most of these olive varieties are cultivated at the regional/national level, with few varieties dispersed globally [111], and the presence of several synonyms and homonyms makes their classification and description a difficult task [110]. This great diversity brings with it the need to develop tools to easily and efficiently distinguish the different varieties and characterize their genetic variability [112].

Morphological and agronomic characteristics are often the first choice for germplasm description and classification and their use represent an initial step in plant breeding [113]. In 1985, the Unité pour la Protection des Obtentions Végétales (UPOV) developed the olive descriptor list, with the aim of standardizing germplasm data collection and, currently, together with IPGRI (International Plant Genetic Resources Institute), hold the official identification and description of varieties and rootstocks, based on morphological characteristics. However, this methodology is complex and time-consuming due to the long juvenile phase of the trees and the potential environmental influence on morphology [114]. These drawbacks led to the incorporation of new methodologies into fruit certification schemes and characterization of plant varieties namely at DNA level, with molecular markers, which can accelerate and optimize the identification process, by allowing the identification of each genotype at any stage of development and independently of environmental factors [115].

Molecular marker techniques have also become very powerful for the analysis of olive genetic diversity. Additionally, if certain molecular markers are associated with interesting characteristics, these markers may become valuable tools for accelerating selection in breeding programs [116].

In olive, the lack of sufficient genetic information hampered the use of species-specific markers until the 2000s [117]; however, many studies have been performed with non-species-specific molecular markers. In Table 3, some applications of these initial non-species-specific molecular markers in olive varieties are shown.

Table 3. Applications of some non-species-specific molecular markers in olive varieties.

Method	Applications	References
Isoenzymes	Olive varieties identification; genetic diversity; genetic relationships	[118]
RFLP (Restriction Fragment Length Polymorphism)	Genetic distance/genetic relationships Genetic linkage map	[119] [120]
RAPD (Randomly Amplified Polymorphic DNA)	Olive varieties identification; genetic diversity; genetic relationships Genetic distance estimation between wild and cultivated olive genotypes Genetic linkage map	[22,41,45,49,51,53,118,121–123] [119,124] [120,125,126]
ISSRs (InterSimple Sequence Repeats)	Olive varieties identification; genetic diversity; genetic relationships	[22,41,42,53,122,123,127,128]
AFLPs (Amplified Fragment Length Polymorphism)	Olive varieties identification; genetic diversity; genetic relationships Genetic linkage map	[129–131] [120,126,132,133]

However, considering the drawbacks of these kind of molecular markers, such as low reproducibility (RAPDs) or being time consuming or expensive (RFLPs, AFLPs), the use of species-specific markers as SSRs (Simple Sequence Repeats, also called microsatellites) and SNPs (Single-Nucleotide Polymorphisms) have been increasing over the last few years in olive analysis [76,134].

SSRs molecular markers, consisting of tandem repeats of mono-, bi-, tri- or tetra-nucleotides in the eukaryotic genome, present high polymorphism based on the number of the nucleotide motif repeats [135]. SSRs are also characterized by a codominant inheritance, good reproducibility between laboratories and easy detection by PCR. In addition, SSRs can be transferred to related species since the sequences flanking the microsatellites are highly conserved [136,137]. Due to these characteristics, SSRs have been the marker of choice for the molecular characterization, genetic diversity and relationship studies in olive trees [46,105,137–139] until the expansion of SNP markers.

SNP markers are individual nucleotide base differences between two DNA sequences, while the polymorphism of microsatellite markers is due to length variations [117,140]. SNPs can be classified according to nucleotide substitution as either transitions or transversions. As a nucleotide base is the smallest unit of inheritance, SNPs provide the ultimate form of molecular marker [140]. Compared with other genetic markers, SNPs are more abundant in the genome, are stably inherited, and can occur in coding regions, occasionally leading to amino acid changes in the polypeptides encoded. They are, in most cases, bi-allelic, codominant, and amenable to high-throughput genotyping and automation [141]. SNP technology is being used to characterize germplasm towards its application in breeding programs. Broad adoption of this technology would be useful also to the plant protection regulatory systems, especially for variety identification and protection purposes. While SSRs are now generally accepted in the courts, some inherent limitations would be overcome using SNP. For instance, high-density oligonucleotide arrays on DNA chips have been developed to analyze genotypes with SNPs. These chips use nucleic acids immobilized on solid-state surfaces, to be hybridized with the sample, without requiring the time-consuming and labor-demanding assays that are needed for most molecular markers (often SSR), essential for size separation of multiple DNA fragments [140]. However, SSRs are still the most used molecular markers for varieties characterization and for diversity studies, since they are cheaper and less technically demanding compared to SNPs.

Table 4 elucidates the growing importance of SSRs and SNPs in olive, by representing the different analysis performed with these markers.

Table 4. Applications of SSRs and SNPs in olive varieties.

Method	Applications	References
SSRs	Identification/characterization of markers	[39,40,136,142,143]
	Olive varieties identification; genetic diversity; genetic relationships	[4,40,42,46–48,50,75,105,112,123,138,144–148]
	Genetic linkage map	[120,125,126,132,133,149]
	Paternity analysis	[150,151]
SNPs	Olive varieties identification; genetic diversity; genetic relationships	[52,141,152,153]
	Genetic linkage map	[154–158]

3.3. The Importance of *Olea* Germplasm Conservation

The recent renewal of old olive orchards in the main olive-producing countries, due to socioeconomic changes, lead to intensive modes of production, with the use of high-yielding and low-vigor varieties. This trend may potentially lead to local olive germplasm erosion, due to the progressive abandonment of several autochthonous and ‘local’ varieties [110,159]. In a climate changing world, with increased frequencies of severe droughts and flooding and the appearance of new diseases and pests, the presence of few varieties in the olive orchards could represent a problem, since monovarietal systems might be less resilient than the more traditional systems [159]. Taking all this into account, the study of traditional varieties and the preservation of its genetic diversity in in situ or ex situ collections are key points, towards the efficient exploitation of these genetic resources, namely by providing breeding material that could improve and diversify modern orchards [110,147,159].

International policies have been taken and projects developed to tackle the continuous loss of olive genetic diversity, due to social, economic and climate changes. For instance, the International Olive Oil Council has been the promoter in 1995 of the European project RESGEN (‘Conservation, characterization, collection and utilization of the olive genetic resources’), which aimed at the collection, characterization and conservation of olive genetic resources as well as at the introduction of germplasm from different countries in national in field ex-situ collections [110]. Moreover, in 2001, the International Treaty on Plant Genetic Resources for Food and Agriculture [160] established a plan for an equitable use of these resources and pictured a global information system to recover and share plant genetic resources [161]. Consequently, prospecting surveys and exchanges of olive varieties between many countries have contributed to the ex situ conservation of this germplasm in several institutions [162]. Currently, important collections exist in all the main olive-growing countries, being the two largest existing ex situ olive germplasm collections, the worldwide olive germplasm Bank of Córdoba, in Spain (WOGBC) and the worldwide olive germplasm Bank of Marrakech, in Morocco (WOGBM) [110,159,162]. In Portugal, a reference collection of olive varieties was set by the Instituto Nacional de Investigação Agrária e Veterinária, I.P (INIAV) in 2012, aiming to conserve and study, under the same edaphoclimatic conditions, the Portuguese olive varieties [16].

In what concerns the ‘Galega’ variety, due to its current replacement in the Portuguese groves, it is essential to prospect and characterize the still available diversity and invest in its conservation, either in situ, through the preservation of ancient isolated individuals or centenary old groves, or ex situ, through the installation of the prospected diversity in comparative field trials.

4. Olive Breeding Worldwide

Across time, olive breeding evolved from an empirical selection performed by olive growers to a more scientific-based selection, using new genomic tools to improve and accelerate breeding programs. This evolution is closely related to the characteristics used to choose a particular variety, which have been changing over time. In fact, presently, breeding objectives include improving yield

while including greater quality and value-added traits, whereas adaptation to the environment was the main selection criteria in the past [14,163].

Compared to other fruit crops, olive breeding is in its infancy, since until recently the agronomic evaluation of olive varieties was incomplete and the efforts to explore and characterize true-to-type olive varieties was very fragmented [110,162]. Although some progress has been achieved, it is fundamental to plan more efficient and faster breeding programs, through the use of new approaches [164]. In fact, the majority of the studies performed in wild and cultivated germplasm have focused on describing the existent variability and not in the direct implication in breeding programs. Thus, there is still a gap of knowledge between the available olive germplasm and its effective use for breeding purposes [165].

4.1. From Conventional to Precision Olive Breeding

The origin of olive growing is connected with the discovery of vegetative propagation using cuttings in the Middle East [164], and since then, the selection and propagation were performed by empirical knowledge by the olive growers. Across time, this empirical selection progressively evolved to a more scientific based selection, mainly performed on the direct observation, clonal selection and cross-breeding of individuals with the most desirable phenotypic characteristics by breeders [110]. In fact, most of the traditional olive varieties were selected from singular seedlings with remarkable traits and then clonal propagated, resulting in all current varieties from selected clones [162]. Presently, with the emergence of molecular techniques and the arising of genomic tools, olive breeding is being improved.

Classic methodologies have been applied worldwide to obtain high yield, early bearing varieties, adapted to the new mechanized orchards [5,166], resistant to plagues and diseases [167,168] or to enlarge the basis of genetic variability [168–170]. Additionally, in Portugal olive breeding was mostly based on the classic hybridization and selection in the progeny. In 2002 an olive breeding program with ‘Galega’ and ‘Cobrançosa’ varieties was initiated, aiming at more productive varieties, with good quality olive oil and higher levels of resistance to diseases and pests, and with better adaptation to mechanized harvesting [16].

Most of the conventional methods are time consuming and failed to meet several requirements of olive production and the demands of the olive industry, namely in enhancing yield and quality of its products, and accelerate development of new varieties and rootstocks [171]. In order to overcome these limitations, several biotechnological advances are currently used in *Olea* breeding [172]. In Table 5, several precision breeding approaches and breeding innovations in olive are summarized.

Table 5. Precision breeding approaches and breeding innovations in *Olea* sp.

Area	References
In vitro techniques for supporting unconventional methods of genetic improvement	[33–35,37,38,173–175]
Plant regeneration from in vitro cultured tissues	[176,177]
Genetic transformation and plant recovery	[178,179]
Transcriptome analysis	[180–191]
Plastome analysis	[2,43,192,193]

In vitro techniques have been developed and successfully used to propagate olive varieties and, in several cases, the resulting in vitro plants have been transplanted in the field [171,175]. The micropropagated materials have been used for screening resistance to biotic and abiotic stress, pathogen virulence and host interaction with parasites focusing on olive genetic improvement [172]. The use of axillary bud stimulation, organogenesis and somatic embryogenesis techniques are also being used to produce plants on a large scale. These techniques allow the production of pathogen-free genotypes, embryo germination, germplasm preservation, plant regeneration from cell tissues and

synthetic seed production [171]. Olive genetic transformation has been used to correct some defects of several varieties, reducing the time and cost of genetic improvement. Currently, this ‘plant gene therapy’ is used with a diversity of objectives, such as improvement of rooting ability, modification of canopy architecture and improvement of the tolerance to biotic and abiotic stresses [172].

Many efforts have also been made in the identification and annotation of genes, through expression analysis studies, based for instance on the identification of ESTs (expressed sequence tags). These studies concerned mainly allergens in olive pollen, characteristics of fruits, olive oil and disease resistance [171,180]. Furthermore, important progress has been accomplished in olive transcriptomic research, with several olive transcriptomes generated from different organs and adaptive responses [190], namely to address flower and fruit development [180,185,190,191], fruit abscission [182], response to abiotic and biotic stresses [181,186,187], plant architecture [188], miRNA [183] and comparative transcriptomics [189].

The entire olive plastome [43,192] provided new information on the olive nucleotide sequence and has been used in different studies such as the assessment of the genetic relationship of olive varieties and the phylogeny of olive [2,102].

Although the described advances in precision breeding techniques, the knowledge about the inheritance of most genes controlling complex agronomical performance and quality traits is still scarce, which consequentially have restricted breeding strategies to clonal or varietal selection [194]. Therefore, understanding the basis of quantitative agronomic traits might help plant breeders to improve important characteristics that are controlled by multiple genes and start a precision olive genetic improvement [171,195].

4.2. Marker Assisted Selection in Olive Breeding

Marker-assisted selection (MAS) for traits controlled by major genes or quantitative trait loci (QTLs) is the primary use of genomics in breeding [164]. However, despite the development and the successful application of different molecular marker systems in olive diversity analysis, their direct application for dissecting agronomic traits or develop trait-associated markers for MAS in breeding has been limited.

The first olive linkage map was developed by De la Rosa et al. [120] using dominant markers (RAPDs and AFLPs) and a limited number of codominant markers (RFLPs and SSRs) in 2003. This pioneer work was followed by others aiming at more saturated, with more markers and longer linkage groups maps [44,125,126,133,149].

All the olive genetic maps available until 2015 were based on non-sequence-based markers (for instance AFLPs) or on low-coverage markers (for example SSRs). However, with the rising of sequencing technologies new high-throughput markers became available, such as SNPs. In fact, genotyping-by-sequencing (GBS), an SNP identification method based on next-generation sequencing (NGS) technologies, showed to be very effective for identify high number of SNP markers and construct high-density genetic linkage maps in olive [156]. Based on this new technology, a high-density genetic linkage map for the olive genome was developed by İpek et al. [157], using 5643 markers (21 SSRs, 203 AFLPs, and 5736 SNPs), representing the most saturated genetic linkage map in olive up to date. Only after this study, it was possible to start locating QTLs for MAS breeding (excepting for some preliminary data from Ben Sadok et al. [149], Atienza et al. [196], Ates [197] and González-Plaza et al. [188]). Additionally, in 2016, Marchese et al. [158] using GBS, constructed a genetic map with 1597 SNP markers, with this linkage map being a useful resource for the study of tree habit and vigor traits segregating in the respective progeny.

Understanding the genetic architecture of important agronomic traits is essential for an effective use of the existent genetic variation in this plant breeding [198,199]. With the development of high-throughput genotyping a transition between the traditional QTL mapping to association or LD (Linkage Disequilibrium) mapping occurred, which assess the correlation between the phenotype and genotype in populations of unrelated individuals instead of familiar related [199].

Genome-wide association studies (GWAS), can take advantage of many generations of historical recombination in a diverse population and avoid the time-consuming generations of controlled crosses [199,200], providing a higher mapping resolution [201].

Despite the high potential of GWAS, this approach has been almost unused in olive. Kaya et al. [195] published the first olive GWAS study, where some significant markers-trait associations were detected for fruit and stone weight. More recently, GWAS was performed in a diverse panel of olive accessions to identify markers associated with five agronomic traits [201] and using 97 olive varieties, the association of the 5'UTR intron of the *FAD2-2* gene with oleic and linoleic acid content [202].

Concerning 'Galega', a detailed characterization of the existing genetic variability, with an effective environmental interaction control, using molecular and agro-morphological information is still missing, hampering the application of these genomic approaches. Consequently, in order to start the application of these tools towards a 'Galega' breeding program, is important to establish a representative collection of the genetic variability of this variety, under the same environment, to allow phenotypic data screening, which will integrate a future GWAS.

4.3. Genotype to Phenotype (G2P) Prediction Models: The Growing Importance of Integrating Environmental Sensitivities and Crop Growth Models Information

As already highlighted, most of the times, the target traits in olive breeding, such as yield, oil quality, and stress resistance, are complex traits, characterized by the contribution of several genes with relatively small effects, and influenced by genotype-by-environment interactions ($G \times E$) [203]. These complex traits very often difficult in the breeding process [204]. Nevertheless, the basis for the success of a plant breeding program lies on plant genotypes with higher performance (phenotype) in yield and/or quality in different environmental conditions. However, to guarantee this high performance, an understanding of the factors that contribute to the desirable phenotype is needed [205].

Traditional empirical methodologies have been used to identify superior individuals, which involve measuring phenotypic performance in broad segregating populations in different environmental trials, and subsequent application of statistical methods based on quantitative genetics. However, this methodology holds some drawbacks, as interactions with management are not integrated and the $G \times E$ might puzzle the selection [206]. Therefore, suitable genotype-to-phenotype (G2P) models that are able to detect genetic and environmental factors, which influence phenotypic variation, and predict the phenotype from genetic and environmental inputs, are needed [207].

The existing models for phenotype prediction hold a blend of statistical, genetic and physiological elements, as is the case of the linear mixed models (LMMs) and crop growth models (CGMs) [203,208]. LMMs can be described with a fixed part, which corresponds to the mean and a random part, composed of variances and covariances [208]. Smith et al. [209] performed a review of the use of mixed models to analyze complex data sets in plant breeding. CGMs integrate genetic and environmental variables in a natural way over time [203] and they have been widely used in plant breeding, namely in the characterization of environments, prediction of consequences of trait variation on yield within a genotype \times environment \times management context, assessment of hybrid performance and evaluation of breeding strategies [210].

The G2P models are of high relevance, since they can address several design issues, namely in the decision of the optimal environments and genotypes to use for phenotyping, the choice of the most important traits in accordance with the different environmental types and the selection of which phenotyping schedule to use [203]. On the other hand, the use of olive CGM [211] will indicate what is the optimum time to measure the different agronomic traits.

Since 'Galega' is present in almost all the Portuguese territory, it is important to phenotype its genetic variability in different environments, possibly with different managements practices, and with a good climate and soil characterizations, for the development of an optimal G2P prediction model. With this model integrating the phenotypic and genotypic diversity, using the soil and climate characteristics

as covariates, it will be possible to predict the production and quality of an olive clone in a certain environment or production system.

5. Future Prospects for a Sustainable Conservation and Use of the ‘Galega’ Variety

The traditional varieties of olives due to the many centuries of selection and propagation are not genetically homogenous entities, being composed of different genotypes with different yield and quality potential. Thus, the full exploitation of their potential has to go through the selection of the best intra-varietal genotypes that should provide the highest yields and the best quality, but considering also that from the sustainability point of view, it is important to maintain a certain level of diversity.

From the above, a more efficient and effective selection will be possible through the complementary morpho-agronomic and molecular characterization of the existent diversity, with a clear understanding of the environmental influence on the trait's expression. This selection will allow the identification of the most interesting clones as the basis for future breeding programs. The development of high-throughput DNA sequencing technologies made available large molecular resources and will support the discovery of controlling genes and regulatory sequences also in olives. In addition, with the assembled draft genome of *O. europaea* [212], breeding programs will be enhanced, as this genomic tool will facilitate the study of the genotype and its relationship with the phenotype.

However, before the accomplishment of the above, it is essential to start by characterizing the current variability in olive varieties.

In the case of Portugal, since ‘Galega’ is the main variety and is being replaced by other foreign varieties, it is essential to characterize the available diversity and create the indispensable conditions to overcome the agronomic limitations still found in this variety. Therefore, to tackle these limitations we propose that efforts should be made to molecularly characterize the existent genetic diversity at the traditional Portuguese olive orchards, allowing as well to unravel the ‘Galega’ ancestral origin. Furthermore, it will be necessary to create conditions to install, under the same environment, the genetic diversity found, to properly apply association mapping approaches. Thus, the identification of the genetic control of the characteristics to improve within ‘Galega’ will be possible with higher resolution and by developing these molecular tools and associated phenotype predictive models (G2P), a more accurate selection and exploitation of this variety could be accomplished. Hence, the valorization of the Portuguese bioeconomic olive sector will be attained by the introduction of adapted improved ‘Galega’ clones, reinforcing the Portuguese international market niche associated with olive oils and olives of excellent quality.

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References

1. Rugini, E.; Baldoni, L.; Muleo, R.; Sebastiani, L. Preface. In *The Olive Tree Genome*; Rugini, E., Baldoni, L., Muleo, R., Sebastiani, L., Eds.; Springer: Cham, Switzerland, 2016; pp. v–vi. [\[CrossRef\]](#)
2. Besnard, G.; Khadari, B.; Navascués, M.; Fernández-Mazuecos, M.; El Bakkali, A.; Arrigo, N.; Baali-Cherif, D.; Brunini-Bronzini de Caraffa, V.; Santoni, S.; Vargas, P.; et al. The complex history of the olive tree: From Late Quaternary diversification of Mediterranean lineages to primary domestication in the northern Levant. *Proc. R. Soc. B* **2013**, *280*, 20122833. [\[CrossRef\]](#)

3. Böhm, J. Domesticação da oliveira na antiguidade na bacia mediterrânica. In *O Grande Livro da Oliveira e do Azeite—Portugal Oleícola*; Böhm, J., Ed.; Dinalivro Editora: Lisboa, Portugal, 2013; pp. 52–65. ISBN 978-972-576-620-0.
4. Trujillo, I.; Ojeda, M.A.; Urdirroz, N.M.; Potter, D.; Barranco, D.; Rallo, L.; Diez, C.M. Identification of the Worldwide Olive Germplasm Bank of Córdoba (Spain) using SSR and morphological markers. *Tree Genet. Genomes* **2014**, *10*, 141–155. [CrossRef]
5. Rallo, L.; Barranco, D.; Castro-García, S.; Connor, D.J.; Gómez del Campo, M.; Rallo, P. High-density olive plantations. *Hortic. Rev.* **2013**, *41*, 303–383. [CrossRef]
6. FAO; Plan Bleu. *State of Mediterranean Forests 2018*; Food and Agriculture Organization of the United Nations: Rome, Italy; Plan Bleu: Marseille, France, 2018. Available online: https://ec.europa.eu/knowledge4policy/publication/state-mediterranean-forests-2018_en (accessed on 20 June 2020).
7. Hashmi, M.A.; Khan, A.; Hanif, M.; Farooq, U.; Perveen, S. Traditional uses, phytochemistry, and pharmacology of *Olea europaea* (olive). *Evid. Based Complement. Altern. Med.* **2015**, *2015*, 541591. [CrossRef]
8. Saldanha, M.H. *Benefícios do Azeite na Saúde Humana*. Coleção Estudos e Análises 7; Direção-Geral de Desenvolvimento Rural: Lisboa, Portugal, 1999. Available online: <https://www.dgadr.gov.pt/component/jdownloads/send/3-agricultura-e-desenvolvimento-rural/12-beneficios-do-azeite-na-saude-humana> (accessed on 20 June 2020).
9. Khadari, B.; Breton, C.; Moutier, N.; Roger, J.; Besnard, G.; Bervillé, A.; Dosba, F. The use of molecular markers for germplasm management in a French olive collection. *Theor. Appl. Genet.* **2003**, *106*, 521–529. [CrossRef]
10. International Olive Council (IOC). Economic Affairs & Promotion Unit—Figures. Available online: <https://www.internationaloliveoil.org/what-we-do/economic-affairs-promotion-unit/#figures> (accessed on 20 June 2020).
11. CONSULAI; Juan-Vilar, Consultores Estratégicos. Alentejo: A Liderar a Olivicultura Moderna Internacional. Olivum. 2019. Available online: https://13b249f8-94d9-4c10-832b9c2323cab575.filesusr.com/ugd/a303d9_5993f29b65054e46a54accff8c90cf7f.pdf (accessed on 20 June 2020).
12. Reis, P. *O Olival em Portugal. Dinâmicas, Tecnologias e Relação com o Desenvolvimento Rural*; Animar—Associação Portuguesa para o Desenvolvimento Local: Lisboa, Portugal, 2014; ISBN 978-989-8748-06-5.
13. Instituto Nacional de Estatística (INE). Estatísticas Agrícolas. Available online: https://www.ine.pt/xportal/xmain?xpid=INE&xpgid=ine_publicacoes&PUBLICACOESpagenumber=1&PUBLICACOEScolecao=107660&PUBLICACOESTipo=ea&selTab=tab0 (accessed on 20 June 2020).
14. Cordeiro, A.M.; Inês, C.; Morais, N. Aspetos gerais da cultura da oliveira: Principais cultivares de oliveira existentes em Portugal. In *Boas Práticas no Olival e no Lagar*; INIAV, I.P., Ed.; SIG—Sociedade Industrial Gráfica: Lisboa, Portugal, 2014; pp. 37–54. ISBN 978-972-579-041-0.
15. Leitão, F.; Pontes, M.F.; Calado, M.L.; Almeida, F.J. *Descrição de 22 Variedades de Oliveira Cultivadas em Portugal*; Ministério da Agricultura, Pescas e Alimentação, Direcção-Geral do Planeamento e Agricultura: Lisboa, Portugal, 1986. Available online: <https://www.dgadr.gov.pt/component/jdownloads/send/10-diversos/26-descricao-de-22-variedades-de-oliveira-cultivadas-em-portugal> (accessed on 20 June 2020).
16. Arias-Calderón, R.; Carvalho, M.T.; Cordeiro, A. *A Importância dos Progenitores no Programa de Melhoramento da Oliveira por Hibridação*; Instituto Nacional de Investigação Agrária e Veterinária, I.P. (INIAV): Lisboa, Portugal, 2017. Available online: http://www.iniaiv.pt/fotos/editor2/a_importancia_dos_progenitores_oliveiras.pdf (accessed on 15 July 2020).
17. Gouveia, J.; Saldanha, J.; Martins, A.; Modesto, M.; Sobral, V. *O Azeite em Portugal*; Edições Inapa: Sintra, Portugal, 2002; ISBN 978-972-797-024-7.
18. Costa, R. Caracterização do Perfil Fenológico e da Ultraestrutura do Pólen de Cultivares de *Olea europaea* L. como Contributo para uma Classificação Varietal. Master's Thesis, University of Coimbra, Coimbra, Portugal, 2011. Available online: <http://hdl.handle.net/10316/20275> (accessed on 21 June 2020).
19. Cordeiro, A.M.; Calado, M.L.; Morais, N.; Miranda, A.; Carvalho, M.T. *Variedades de Oliveira: 'Galega Vulgar'*; Direcção Regional de Agricultura e Pescas do Centro: Castelo Branco, Portugal, 2010. Available online: https://www.drapc.gov.pt/base/documentos/018_variedades_oliveira_galega_vulgar_nov10.pdf (accessed on 20 June 2020).
20. Del Río, C.; Caballero, J.M. Variability and classification of olive cultivars by fruit weight, flesh/stone ratio and oil percentage. *Acta Hortic.* **2008**, *791*, 39–44. [CrossRef]

21. Albuquerque, T.G.; Costa, H.S.; Oliveira, M.B.P.P. An overview of Portuguese olive oils and table olives with protected designation of origin. *Eur. J. Lipid Sci. Tech.* **2019**, *121*, 1800129. [\[CrossRef\]](#)
22. Gemas, V.J.; Almadanim, M.C.; Tenreiro, R.; Martins, A.; Fevereiro, P. Genetic diversity in the olive tree (*Olea europaea* L. subsp. *europaea*) cultivated in Portugal revealed by RAPD and ISSR markers. *Genet. Resour. Crop Evol.* **2004**, *51*, 501–511. [\[CrossRef\]](#)
23. Serrano, M.C.F.; Amaral, L. Irrigation effects on some fructification parameters of olive cvs. *Olea europaea* L. ‘Galega vulgar’, ‘Cordovil de Serpa’ and ‘Verdeal Alentejana’ in Alentejo. *Acta Hortic.* **2008**, *791*, 325–331. [\[CrossRef\]](#)
24. Coelho, R.; Sousa, A.; Rato, A.; Vaz, M. Response to salinity in young olive trees of three Iberian varieties. In Proceedings of the 12th Portuguese-Spanish Symposium on Plant Water Relations, Évora, Portugal, 30 September–3 October 2014; pp. 99–104.
25. Peixe, A.; Antunes, A.; Hegewald, H.; Costa, C.; Pinto, A.P. Preliminary results on the evaluation of auxin endogenous levels and oxidative enzymes activity, among the rooting of two olive (*Olea europaea* L.) cultivars ‘Galega vulgar’ and ‘Cobrançosa’. In *Atas Portuguesas de Horticultura 14, Proceedings of the 5th Simpósio Nacional de Olivicultura, Santarém, Portugal, 24–26 September 2009*; Associação Portuguesa de Horticultura: Lisboa, Portugal, 2011; pp. 13–22.
26. Lopes, J.P.P.S. Polinização em Oliveira cvs. Galega e Cobrançosa: Avaliação do Vingamento em Ensaios de Polinização Controlada e do Efeito da Aplicação de um Bioestimulante. Master’s Thesis, Instituto Superior de Agronomia, University of Lisboa, Lisboa, Portugal, 2011. Available online: <http://hdl.handle.net/10400.5/4122> (accessed on 29 June 2020).
27. Azevedo, A.; Bernardes, P. Influência das condições ecológicas no crescimento do olival em regime superintensivo. Reposta da cultivar Galega vulgar à fertilização azotada. In Proceedings of the 1st Congresso Nacional das Escolas Superiores Agrárias, Bragança, Portugal, 2–3 December 2015.
28. Dias, M.C.; Pinto, D.C.G.A.; Freitas, H.; Santos, C.; Silva, A.M.S. The antioxidant system in *Olea europaea* to enhanced UV-B radiation also depends on flavonoids and secoiridoids. *Phytochemistry* **2020**, *170*, 112199. [\[CrossRef\]](#)
29. Maia, F.B. Contribuição para o Estudo da Fenologia de Quatro Variedades de Oliveira (*Olea europaea* L.): ‘Arbequina’, ‘Cobrançosa’, ‘Galega Vulgar’ e ‘Picual’. Master’s Thesis, Instituto Superior de Agronomia, University of Lisboa, Lisboa, Portugal, 2010. Available online: <http://hdl.handle.net/10400.5/15199> (accessed on 29 June 2020).
30. Cordeiro, A.M.; Martins, P.C.S.; Ramos, A.; Sequeira, P. Characterization of olive cultivars fruit set in self-pollination. In *Atas Portuguesas de Horticultura 14, Proceedings of the 5th Simpósio Nacional de Olivicultura, Santarém, Portugal, 24–26 September 2009*; Associação Portuguesa de Horticultura: Lisboa, Portugal, 2011; pp. 23–30.
31. Jimenez-Lopez, J.C.; Morales, S.; Castro, A.J.; Volkmann, D.; Rodríguez-García, M.I.; Alche, J.D. Characterization of profilin polymorphism in pollen with a focus on multifunctionality. *PLoS ONE* **2012**, *7*, e30878. [\[CrossRef\]](#)
32. Ribeiro, H.; Calado, L.; Cunha, M.; Abreu, I. Evaluation of pollen viability and germination from 20 varieties of *Olea europaea* L. grown in Portugal. In *Atas Portuguesas de Horticultura 14, Proceedings of the 5th Simpósio Nacional de Olivicultura, Santarém, Portugal, 24–26 September 2009*; Associação Portuguesa de Horticultura: Lisboa, Portugal, 2011; pp. 39–46.
33. Velada, I.; Grzebelus, D.; Lousa, D.; Soares, C.M.; Macedo, E.S.; Peixe, A.; Arnholdt-Schmitt, B.; Cardoso, H.G. AOX1-subfamily gene members in *Olea europaea* cv. “Galega Vulgar”—gene characterization and expression of transcripts during IBA-induced in vitro adventitious rooting. *Int. J. Mol. Sci.* **2018**, *19*, 597. [\[CrossRef\]](#)
34. Velada, I.; Cardoso, H.; Porfirio, S.; Peixe, A. Expression profile of PIN-formed auxin efflux carrier genes during IBA-induced in vitro adventitious rooting in *Olea europaea* L. *Plants* **2020**, *9*, 185. [\[CrossRef\]](#)
35. Macedo, E.; Vieira, C.; Carrizo, D.; Porfirio, S.; Hegewald, H.; Arnholdt-Schmitt, B.; Calado, M.L.; Peixe, A. Adventitious root formation in olive (*Olea europaea* L.) microshoots: Anatomical evaluation and associated biochemical changes in peroxidase and polyphenol oxidase activities. *J. Hortic. Sci. Biotech.* **2013**, *88*, 53–59. [\[CrossRef\]](#)
36. Porfirio, S.; Calado, M.L.; Noceda, C.; Cabrita, M.J.; Silva, M.G.; Azadi, P.; Peixe, A. Tracking biochemical changes during adventitious root formation in olive (*Olea europaea*). *Sci. Hortic.* **2016**, *204*, 41–53. [\[CrossRef\]](#)

37. Ribeiro, A.V.A.P. Simplificação do Processo de Multiplicação in vitro da Oliveira "*Olea europaea* L.". Master's Thesis, University of Évora, Évora, Portugal, 2016. Available online: <http://hdl.handle.net/10174/20702> (accessed on 29 June 2020).
38. Pires, R.N. Ensaios com Vista à Indução de Embriogénese Somática em Oliveira (*Olea europaea* L.). Master's Thesis, University of Évora, Évora, Portugal, 2018. Available online: <http://hdl.handle.net/10174/23981> (accessed on 29 June 2020).
39. Rallo, P.; Dorado, G.; Martín, A. Development of simple sequence repeats in olive tree (*Olea europaea* L.). *Theor. Appl. Genet.* **2000**, *101*, 984–989. [\[CrossRef\]](#)
40. Díaz, A.; De la Rosa, R.; Martín, A.; Rallo, P. Development, characterization and inheritance of new microsatellites in olive (*Olea europaea* L.) and evaluation of their usefulness in cultivar identification and genetic relationship studies. *Tree Genet. Genomes* **2006**, *2*, 165–175. [\[CrossRef\]](#)
41. Martins-Lopes, P.; Lima-Brito, J.; Gomes, S.; Meirinhos, J.; Santos, L.; Guedes-Pinto, H. RAPD and ISSR molecular markers in *Olea europaea* L.: Genetic variability and molecular cultivar identification. *Genet. Resour. Crop Evol.* **2007**, *54*, 117–128. [\[CrossRef\]](#)
42. Gomes, S.; Martins-Lopes, P.; Lopes, J.; Guedes-Pinto, H. Assessing genetic diversity in *Olea europaea* L. using ISSR and SSR markers. *Plant Mol. Biol. Rep.* **2009**, *27*, 365–373. [\[CrossRef\]](#)
43. Mariotti, R.; Cultrera, N.G.M.; Díez, C.M.; Baldoni, L.; Rubini, A. Identification of new polymorphic regions and differentiation of cultivated olives (*Olea europaea* L.) through plastome sequence comparison. *BMC Plant Biol.* **2010**, *10*, 211. [\[CrossRef\]](#)
44. Domínguez-García, M.C.; Belaj, A.; De la Rosa, R.; Šatović, Z.; Heller-Uszynska, K.; Kilian, A.; Martín, A.; Atienza, S.G. Development of DArT markers in olive (*Olea europaea* L.) and usefulness in variability studies and genome mapping. *Sci. Hortic.* **2012**, *136*, 50–60. [\[CrossRef\]](#)
45. Parra-Lobato, M.C.; Delgado-Martinez, F.J.; Gomez-Jimenez, M.C. Morphological traits and RAPD markers for characterization and identification of minor Spanish olive cultivars from the Extremadura region. *Genet. Mol. Res.* **2012**, *11*, 2401–2411. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Lazović, B.; Adakalić, M.; Pucci, C.; Perović, T.; Bandelj, D.; Belaj, A.; Mariotti, R.; Baldoni, L. Characterizing ancient and local olive germplasm from Montenegro. *Sci. Hortic.* **2016**, *209*, 117–123. [\[CrossRef\]](#)
47. Veloso, M.; Simões-Costa, M.C.; Carneiro, L.C.; Guimarães, J.B.; Mateus, C.; Feveiro, P.; Pinto-Ricardo, C. Olive tree (*Olea europaea* L.) diversity in traditional small farms of Ficalho, Portugal. *Diversity* **2018**, *10*, 5. [\[CrossRef\]](#)
48. Lopes, M.S.; Mendonça, D.; Sefc, K.M.; Gil, F.S.; Da Câmara Machado, A. Genetic evidence of intra-cultivar variability within Iberian olive cultivars. *HortScience* **2004**, *39*, 1562–1565. [\[CrossRef\]](#)
49. Cordeiro, A.I.; Sanchez-Sevilla, J.F.; Alvarez-Tinaut, M.C.; Gomez-Jimenez, M.C. Genetic diversity assessment in Portugal accessions of *Olea europaea* by RAPD markers. *Biol. Plant.* **2008**, *52*, 642–647. [\[CrossRef\]](#)
50. Feveiro, P.; Leitão, F.; Potes, F.; Gemas, V.; Alves, M.; Favoretto, P. The Portuguese olive (*Olea europaea* subsp. *europaea*) germplasm. *Acta Hortic.* **2011**, *924*, 291–298. [\[CrossRef\]](#)
51. Figueiredo, E.; Canhoto, J.; Ribeiro, M.M. Fingerprinting and genetic diversity of *Olea europaea* L. ssp. *europaea* accessions from the cultivar Galega using RAPD markers. *Sci. Hortic.* **2013**, *156*, 24–28. [\[CrossRef\]](#)
52. Cultrera, N.G.M.; Sarri, V.; Lucentini, L.; Ceccarelli, M.; Alagna, F.; Mariotti, R.; Mousavi, S.; Ruiz, C.G.; Baldoni, L. High levels of variation within gene sequences of *Olea europaea* L. *Front. Plant Sci.* **2019**, *9*, 1932. [\[CrossRef\]](#)
53. Pereira, M.R. Diversidade Genómica e Genética Intra-Varietal em Oliveira 'Galega Vulgar'. Master's Thesis, Instituto Superior de Agronomia, University of Lisboa, Lisboa, Portugal, 2019. Available online: <http://hdl.handle.net/10400.5/18359> (accessed on 29 June 2020).
54. Dias, A.B.; Pimentel, R.S.; Pinheiro, A.; Peça, J.O. Resultados da avaliação da poda com máquina de discos num olival intensivo da variedade "Galega vulgar". In *Actas de Horticultura 73, Proceedings of the 1st Congreso Ibérico de Olivicultura, 5th Jornadas Nacionales del Grupo de Olivicultura de la Sociedad Española de Ciencias Hortícolas (SECH) and 7th Simpósio Nacional de Olivicultura da Associação Portuguesa de Horticultura (APH), Badajoz-Elvas, Spain, Portugal, 13–15 April 2016*; Sociedad Española de Ciencias Hortícolas: Córdoba, Spain, 2018; pp. 194–201.
55. Goldental-Cohen, S.; Biton, I.; Many, Y.; Ben-Sason, S.; Zemach, H.; Avidan, B.; Ben-Ari, G. Green olive browning differ between cultivars. *Front. Plant Sci.* **2019**, *10*, 1260. [\[CrossRef\]](#)

56. Landum, M.C.; Félix, M.R.; Alho, J.; Garcia, R.; Cabrita, M.J.; Rei, F.; Varanda, C.M.R. Antagonistic activity of fungi of *Olea europaea* L. against *Colletotrichum acutatum*. *Microbiol. Res.* **2016**, *183*, 100–108. [\[CrossRef\]](#) [\[PubMed\]](#)
57. Materatski, P.; Varanda, C.; Carvalho, T.; Dias, A.B.; Campos, M.D.; Rei, F.; Félix, M.R. Spatial and temporal variation of fungal endophytic richness and diversity associated to the phyllosphere of olive cultivars. *Fungal Biol.* **2019**, *123*, 66–76. [\[CrossRef\]](#) [\[PubMed\]](#)
58. Varanda, C.M.R.; Materatski, P.; Landum, M.; Campos, M.D.; Félix, M.R. Fungal communities associated with peacock and cercospora leaf spots in olive. *Plants* **2019**, *8*, 169. [\[CrossRef\]](#) [\[PubMed\]](#)
59. Campos, M.D.; Zellama, M.S.; Varanda, C.; Materatski, P.; Peixe, A.; Chaouachi, M.; Félix, M.R. Establishment of a sensitive qPCR methodology for detection of the olive-infecting viruses in Portuguese and Tunisian orchards. *Front. Plant Sci.* **2019**, *10*, 694. [\[CrossRef\]](#)
60. Gouvínhas, I.; Martins-Lopes, P.; Carvalho, T.; Barros, A.; Gomes, S. Impact of *Colletotrichum acutatum* pathogen on olive phenylpropanoid metabolism. *Agriculture* **2019**, *9*, 173. [\[CrossRef\]](#)
61. Materatski, P.; Varanda, C.; Carvalho, T.; Dias, A.B.; Campos, M.D.; Gomes, L.; Nobre, T.; Rei, F.; Félix, M.R. Effect of long-term fungicide applications on virulence and diversity of *Colletotrichum* spp. associated to olive anthracnose. *Plants* **2019**, *8*, 311. [\[CrossRef\]](#)
62. Azevedo-Nogueira, F.; Gomes, S.; Carvalho, T.; Martins-Lopes, P. Development of high-throughput real-time PCR assays for the *Colletotrichum acutatum* detection on infected olive fruits and olive oils. *Food Chem.* **2020**, *317*, 126417. [\[CrossRef\]](#)
63. Alves, E.; Melo, T.; Barros, M.P.; Domingues, M.R.M.; Domingues, P. Lipidomic profiling of the olive (*Olea europaea* L.) fruit towards its valorisation as a functional food: In-depth identification of triacylglycerols and polar lipids in Portuguese olives. *Molecules* **2019**, *24*, 2555. [\[CrossRef\]](#)
64. Ferro, M.D.; Santos, S.A.O.; Silvestre, A.J.D.; Duarte, M.F. Chromatographic separation of phenolic compounds from extra virgin olive oil: Development and validation of a new method based on a biphenyl HPLC column. *Int. J. Mol. Sci.* **2019**, *20*, 201. [\[CrossRef\]](#)
65. Peres, M.F.; Henriques, L.R.; Simões-Lopes, P.; Pinheiro-Alves, M.C. Azeites da ‘Galega Vulgar’—efeito do loteamento e do armazenamento. In *Atas Portuguesas de Horticultura 13, Proceedings of the 3rd Simpósio Nacional de Olivicultura, Castelo Branco, Portugal, 29–31 October 2003*; Associação Portuguesa de Horticultura: Lisboa, Portugal, 2009; pp. 186–191.
66. Marcelo, M.E.; Lopes, J.I.; Soares, F.M.; Centeno, M.S.L.; Cordeiro, A.M.; Vasconcelos, P.M. Effect of fertilization on polyphenols, sterols and waxes contents of the olive oil from six cultivars in Portugal. In *Atas Portuguesas de Horticultura 14, Proceedings of the 5th Simpósio Nacional de Olivicultura, Santarém, Portugal, 24–26 September 2009*; Associação Portuguesa de Horticultura: Lisboa, Portugal, 2011; pp. 141–150.
67. Pita, D.; Vitorino, M.C.; Gouveia, C.; Peres, F. Extraction of monovarietal olive oils with natural microtalc. In *Atas Portuguesas de Horticultura 14, Proceedings of the 5th Simpósio Nacional de Olivicultura, Santarém, Portugal, 24–26 September 2009*; Associação Portuguesa de Horticultura: Lisboa, Portugal, 2011; pp. 151–157.
68. Quintero-Flórez, A.; Nieva, L.S.; Sánchez-Ortíz, A.; Beltrán, G.; Perona, J.S. The fatty acid composition of virgin olive oil from different cultivars is determinant for foam cell formation by macrophages. *J. Agric. Food Chem.* **2015**, *63*, 6731–6738. [\[CrossRef\]](#)
69. Gouvínhas, I.; Domínguez-Perles, R.; Machado, N.; Carvalho, T.; Matos, C.; Barros, A.I.R.N.A. Effect of agro-environmental factors on the mineral content of olive oils: Categorization of the three major Portuguese cultivars. *J. Am. Oil Chem. Soc.* **2016**, *93*, 813–822. [\[CrossRef\]](#)
70. Pires-Cabral, P.; Barros, T.; Nunes, P.; Quintas, C. Physicochemical, nutritional and microbiological characteristics of traditional table olives from Southern Portugal. *Emir. J. Food Agric.* **2018**, *30*, 611–620. [\[CrossRef\]](#)
71. Saramago, I.S.L. Olival em Modo de Produção Biológico. Master’s Thesis, Escola Superior Agrária, Instituto Politécnico de Beja, Beja, Portugal, 2009. Available online: <http://hdl.handle.net/10400.26/3979> (accessed on 15 September 2020).
72. Tous, J.; Romero, A.; Hermoso, J.F.; Mallén, N. Sistemas de producción del olivo en seto. Experiencias en Cataluña. *Agricultura* **2007**, *896*, 360–367.
73. Guerrero Maldonado, N.; López, M.J.; Caudullo, G.; de Rigo, D. *Olea europaea* in Europe: Distribution, habitat, usage and threats. In *European Atlas of Forest Tree Species*; San-Miguel-Ayán, J., de Rigo, D., Caudullo, G., Houston Durrant, T., Mauri, A., Eds.; Publ. Off. EU: Luxembourg, 2016; p. e01534b+.

74. Beghè, D.; Ferrarini, A.; Ganino, T.; Fabbri, A. Molecular characterization and identification of a group of local *Olea europaea* L. varieties. *Tree Genet. Genomes* **2011**, *7*, 1185–1198. [[CrossRef](#)]
75. Marra, F.P.; Caruso, T.; Costa, F.; Di Vaio, C.; Mafra, R.; Marchese, A. Genetic relationships, structure and parentage simulation among the olive tree (*Olea europaea* L. subsp. *europaea*) cultivated in Southern Italy revealed by SSR markers. *Tree Genet. Genomes* **2013**, *9*, 961–973. [[CrossRef](#)]
76. Belaj, A.; Veral, M.G.; Sikaoui, H.; Moukhli, A.; Khadari, B.; Mariotti, R.; Baldoni, L. Olive genetic resources. In *The Olive Tree Genome*; Rugini, E., Baldoni, L., Muleo, R., Sebastiani, L., Eds.; Springer: Cham, Switzerland, 2016; pp. 27–54. [[CrossRef](#)]
77. Lavee, S. Evaluation of the need and present potential of olive breeding indicating the nature of the available genetic resources involved. *Sci. Hortic.* **2013**, *161*, 333–339. [[CrossRef](#)]
78. Belaj, A.; Muñoz-Diez, C.; Baldoni, L.; Šatović, Z.; Barranco, D. Genetic diversity and relationships of wild and cultivated olives at regional level in Spain. *Sci. Hortic.* **2010**, *124*, 323–330. [[CrossRef](#)]
79. Maxted, N.; Guarino, L. Genetic erosion and genetic pollution of crop wild relatives. In Proceedings of the PGR Forum Workshop 5, Genetic Erosion and Pollution Assessment Methodologies, Terceira Island, Autonomous Region of the Azores, Portugal, 8–11 September 2004; pp. 35–45.
80. Brown, A.H.D.; Hodgkin, T. Indicators of Genetic Diversity, Genetic Erosion, and Genetic Vulnerability for Plant Genetic Resources. In *Genetic Diversity and Erosion in Plants. Sustainable Development and Biodiversity*; Ahuja, M., Jain, S., Eds.; Springer: Cham, Switzerland, 2015; Volume 7, pp. 25–53. [[CrossRef](#)]
81. Salomone, R.; Cappelletti, G.M.; Malandrino, O.; Mistretta, M.; Neri, E.; Nicoletti, G.; Notarnicola, B.; Pattara, C.; Russo, C.; Saija, G. Life cycle assessment in the olive oil sector. In *Life Cycle Assessment in the Agri-Food Sector*; Notarnicola, B., Salomone, R., Petti, L., Renzulli, P., Roma, R., Cerutti, A., Eds.; Springer: Cham, Switzerland, 2015; pp. 57–121. [[CrossRef](#)]
82. Espadas-Aldana, G.; Vialle, C.; Belaud, J.P.; Vaca-Garcia, C.; Sablayrolles, C. Analysis and trends for Life Cycle Assessment of olive oil production. *Sustain. Prod. Consum.* **2019**, *19*, 216–230. [[CrossRef](#)]
83. Salomone, R.; Ioppolo, G. Environmental impacts of olive oil production: A life cycle assessment case study in the province of Messina (Sicily). *J. Clean. Prod.* **2012**, *28*, 88–100. [[CrossRef](#)]
84. Rajaeifar, M.A.; Akram, A.; Ghobadian, B.; Rafiee, S.; Heidari, M.D. Energy economic life cycle assessment (LCA) and greenhouse gas emissions analysis of olive oil production in Iran. *Energy* **2014**, *66*, 139–149. [[CrossRef](#)]
85. Proietti, S.; Sdringola, P.; Regni, L.; Evangelisti, N.; Brunori, A.; Ilarioni, L.; Nasini, L.; Proietti, P. Extra virgin olive oil as carbon negative product: Experimental analysis and validation of results. *J. Clean. Prod.* **2017**, *166*, 550–562. [[CrossRef](#)]
86. Tsarouhas, P.; Achillas, C.; Aidonis, D.; Folinas, D.; Maslis, V. Life cycle assessment of olive oil production in Greece. *J. Clean. Prod.* **2015**, *93*, 75–83. [[CrossRef](#)]
87. El Hanandeh, A.; Gharaibeh, M.A. Environmental efficiency of olive oil production by small and micro-scale farmers in northern Jordan: Life cycle assessment. *Agric. Syst.* **2016**, *148*, 169–177. [[CrossRef](#)]
88. Pattara, C.; Salomone, R.; Cichelli, A. Carbon footprint of extra virgin olive oil: A comparative and driver analysis of different production processes in Centre Italy. *J. Clean. Prod.* **2016**, *127*, 533–547. [[CrossRef](#)]
89. Accorsi, R.; Cascini, A.; Ferrari, E.; Manzini, R.; Pareschi, A.; Versari, L. Life cycle assessment of an extra-virgin olive oil supply chain. In Proceedings of the 18th Summer School “Francesco Turco”—Industrial Mechanical Plants, Senigallia, Italy, 11–13 September 2013; pp. 172–178.
90. Guiso, A.; Parenti, A.; Masella, P.; Guerrini, L.; Baldi, F.; Spugnoli, P. Environmental impact assessment of three packages for high-quality extra-virgin olive oil. *Agric. Eng.* **2016**, *47*, 191–196. [[CrossRef](#)]
91. Navarro, A.; Puig, R.; Martí, E.; Bala, A.; Fullana-i Palmer, P. Tackling the relevance of packaging in life cycle assessment of virgin olive oil and the environmental consequences of regulation. *Environ. Manag.* **2018**, *62*, 277–294. [[CrossRef](#)]
92. Chatzisyseon, E.; Foteinis, S.; Mantzavinos, D.; Tsoutsos, T. Life cycle assessment of advanced oxidation processes for olive mill wastewater treatment. *J. Clean. Prod.* **2013**, *54*, 229–234. [[CrossRef](#)]
93. El Hanandeh, A. Energy recovery alternatives for the sustainable management of olive oil industry waste in Australia: Life cycle assessment. *J. Clean. Prod.* **2015**, *91*, 78–88. [[CrossRef](#)]
94. Christoforou, E.A.; Fokaides, P.A. Life cycle assessment (LCA) of olive husk torrefaction. *Renew. Energy* **2016**, *90*, 257–266. [[CrossRef](#)]

95. Parascanu, M.M.; Gamero, M.P.; Sánchez, P.; Soreanu, G.; Valverde, J.L.; Sanchez-Silva, L. Life cycle assessment of olive pomace valorisation through pyrolysis. *Renew. Energy* **2018**, *122*, 589–601. [\[CrossRef\]](#)
96. Parascanu, M.M.; Sánchez, P.; Soreanu, G.; Valverde, J.L.; Sanchez-Silva, L. Environmental assessment of olive pomace valorization through two different thermochemical processes for energy production. *J. Clean. Prod.* **2018**, *186*, 771–781. [\[CrossRef\]](#)
97. Mohamad, R.S.; Verrastro, V.; Cardone, G.; Bteich, M.R.; Favia, M.; Moretti, M.; Roma, R. Optimization of organic and conventional olive agricultural practices from a life cycle assessment and life cycle costing perspectives. *J. Clean. Prod.* **2014**, *70*, 78–89. [\[CrossRef\]](#)
98. De Luca, A.I.; Falcone, G.; Stillitano, T.; Iofrida, N.; Strano, A.; Gulisano, G. Evaluation of sustainable innovations in olive growing systems: A life cycle sustainability assessment case study in southern Italy. *J. Clean. Prod.* **2018**, *171*, 1187–1202. [\[CrossRef\]](#)
99. Romero-Gámez, M.; Castro-Rodríguez, J.; Suárez-Rey, E.M. Optimization of olive growing practices in Spain from a life cycle assessment perspective. *J. Clean. Prod.* **2017**, *149*, 25–37. [\[CrossRef\]](#)
100. Bernardi, B.; Falcone, G.; Stillitano, T.; Benalia, S.; Strano, A.; Bacenetti, J.; De Luca, A.I. Harvesting system sustainability in Mediterranean olive cultivation. *Sci. Total Environ.* **2018**, *625*, 1446–1458. [\[CrossRef\]](#)
101. Kaniewski, D.; Van Campo, E.; Boiy, T.; Terral, J.F.; Khadari, B.; Besnard, G. Primary domestication and early uses of the emblematic olive tree: Palaeobotanical, historical and molecular evidences from the Middle East. *Biol. Rev.* **2012**, *87*, 885–899. [\[CrossRef\]](#)
102. Besnard, G.; Terral, J.; Cornille, A. On the origins and domestication of the olive: A review and perspectives. *Ann. Bot.* **2018**, *121*, 385–403. [\[CrossRef\]](#)
103. Díez, C.M.; Moral, J.; Barranco, D.; Rallo, L. Genetic diversity and conservation of olive genetic resources. In *Genetic Diversity and Erosion in Plants. Sustainable Development and Biodiversity*; Ahuja, M., Jain, S., Eds.; Springer: Cham, Switzerland, 2015; Volume 8, pp. 337–356. [\[CrossRef\]](#)
104. Besnard, G.; El Bakkali, A.; Haouane, H.; Baali-Cherif, D.; Moukhli, A.; Khadari, B. Population genetics of Mediterranean and Saharan olives: Geographic patterns of differentiation and evidence for early-generations of admixture. *Ann. Bot.* **2013**, *112*, 1293–1302. [\[CrossRef\]](#)
105. Díez, C.M.; Trujillo, I.; Martínez-Urdiroz, N.; Barranco, D.; Rallo, L.; Marfil, P.; Gaut, B.S. Olive domestication and diversification in the Mediterranean Basin. *New Phytol.* **2015**, *206*, 436–447. [\[CrossRef\]](#)
106. Besnard, G.; Rubio de Casas, R.; Christin, P.A.; Vargas, P. Phylogenetics of *Olea* (Oleaceae) based on plastid and nuclear ribosomal DNA sequences: Tertiary climatic shifts and lineage differentiation times. *Ann. Bot.* **2009**, *104*, 143–160. [\[CrossRef\]](#)
107. Ladisa, G.; Calabrese, E.; Perrino, E.V. The origin and distribution of olive tree and olive crop. In *Study on Biodiversity in Century-Old Olive Groves*; Calabrese, G., Tartaglini, N., Ladisa, G., Eds.; CIHEAM—Mediterranean Agronomic Institute of Bari: Bari, Italy, 2012; pp. 1–3.
108. Fuller, D.Q. Long and attenuated: Comparative trends in the domestication of tree fruits. *Veg. Hist. Archaeobotany* **2018**, *27*, 165–176. [\[CrossRef\]](#) [\[PubMed\]](#)
109. Belaj, A.; Dominguez-García, M.C.; Atienza, S.G.; Urdiroz, N.M.; De la Rosa, R.; Šatović, Z.; Martín, A.; Kilian, A.; Trujillo, I.; Valpuesta, V.; et al. Developing a core collection of olive (*Olea europaea* L.) based on molecular markers (DARs, SSRs, SNPs) and agronomic traits. *Tree Genet. Genomes* **2012**, *8*, 365–378. [\[CrossRef\]](#)
110. Fabbri, A.; Lambardi, M.; Ozden-Tokatli, Y. Olive Breeding. In *Breeding Plantation Tree Crops: Tropical Species*; Jain, S.M., Priyadarshan, P.M., Eds.; Springer: New York, NY, USA, 2009; pp. 423–465. ISBN 978-038-771-199-7.
111. Doveri, S.; Baldoni, L. Olive. In *Genome Mapping and Molecular Breeding in Plants—Fruits and Nuts*; Kole, C., Ed.; Springer: Berlin/Heidelberg, Germany, 2007; Volume 4, pp. 253–256. ISBN 978-354-034-531-2.
112. Alves, M.L.V. Caracterização e Estrutura Genéticas da Cultivar de Oliveira ‘Cobrançosa’ e Sua Relação com o Zambujeiro. Master’s Thesis, University of Lisboa, Lisboa, Portugal, 2007. Available online: <http://hdl.handle.net/10451/1358> (accessed on 30 June 2020).
113. Belaj, A.; León, L.; Šatović, Z.; De la Rosa, R. Variability of wild olives (*Olea europaea* subsp. *europaea* var. *sylvestris*) analyzed by agro-morphological traits and SSR markers. *Sci. Hortic.* **2011**, *129*, 561–569. [\[CrossRef\]](#)
114. Bouhadida, M.; Casas, A.M.; Moreno, M.A.; Gogorcena, Y. Molecular characterization of Miraflores peach variety and relatives using SSRs. *Sci. Hortic.* **2007**, *111*, 140–145. [\[CrossRef\]](#)
115. Wünsch, A.; Hormaza, J.I. Cultivar identification and genetic fingerprinting of temperate fruit tree species using DNA markers. *Euphytica* **2002**, *125*, 59–67. [\[CrossRef\]](#)

116. Hammer, K.; Arrowsmith, N.; Gladis, T. Agrobiodiversity with emphasis on plant genetic resources. *Naturwissenschaften* **2003**, *90*, 241–250. [\[CrossRef\]](#)
117. Pasqualone, A.; Montemurro, C.; Di Rienzo, V.; Summo, C.; Paradiso, V.M.; Caponio, F. Evolution and perspectives of cultivar identification and traceability from tree to oil and table olives by means of DNA markers. *J. Sci. Food Agric.* **2016**, *96*, 3642–3657. [\[CrossRef\]](#)
118. Lazović, B.; Bosković, R.; James, C.; Tobutt, K.R.; Gasic, K. Genetic diversity of olives grown along the coast of Montenegro. *Acta Hortic.* **2002**, *586*, 167–170. [\[CrossRef\]](#)
119. Besnard, G.; Baradat, P.; Chevalier, D.; Tagmount, A.; Bervillé, A. Genetic differentiation in the olive complex (*Olea europaea* L.) revealed by RAPDs and RFLPs in the rRNA genes. *Genet. Resour. Crop Evol.* **2001**, *48*, 165–182. [\[CrossRef\]](#)
120. De la Rosa, R.; Angiolillo, A.; Rallo, L.; Guerrero, C.; Pellegrini, M.; Besnard, G.; Bervillé, A.; Martin, A.; Baldoni, L. A first linkage map of olive (*Olea europaea* L.) cultivars using RAPD, AFLP, RFLP and SSR markers. *Theor. Appl. Genet.* **2003**, *106*, 1273–1282. [\[CrossRef\]](#)
121. Belaj, A.; Šatović, Z.; Trujillo, I.; Rallo, L. Genetic relationships of Spanish olive cultivars using RAPD markers. *HortScience* **2004**, *39*, 948–951. [\[CrossRef\]](#)
122. Brake, M.; Migdadi, H.; Al-Gharaibeh, M.; Ayoub, S.; Haddad, N.; El Oqlah, A. Characterization of Jordanian olive cultivars (*Olea europaea* L.) using RAPD and ISSR molecular markers. *Sci. Hortic.* **2014**, *176*, 282–289. [\[CrossRef\]](#)
123. Linos, A.; Nikoloudakis, N.; Katsiotis, A.; Hagidimitriou, M. Genetic structure of the Greek olive germplasm revealed by RAPD, ISSR and SSR markers. *Sci. Hortic.* **2014**, *175*, 33–43. [\[CrossRef\]](#)
124. Besnard, G.; Bervillé, A. Multiple origins for Mediterranean olive (*Olea europaea* L. ssp. *europaea*) based upon mitochondrial DNA polymorphisms. *Comptes Rendus Acad. Sci.* **2000**, *323*, 173–181. [\[CrossRef\]](#)
125. Wu, S.B.; Collins, G.; Sedgley, M. A molecular linkage map of olive (*Olea europaea* L.) based on RAPD, microsatellite, and SCAR markers. *Genome* **2004**, *47*, 26–35. [\[CrossRef\]](#)
126. Charafi, J.; Zine El Aabidine, A.; Grout, C.; Rahioui, B.; El Meziane, A.; Moukhli, A.; Boulouha, B.; El Modafar, C.; Khadari, B. A genetic linkage map of *Olea europaea* L. using a pseudo-test cross- mapping strategy based on SSR, AFLP, ISSR, RAPD and SCAR markers. *Acta Hortic.* **2009**, *814*, 609–614. [\[CrossRef\]](#)
127. Terzopoulos, P.J.; Kolano, B.; Bebeli, P.J.; Kaltsikes, P.J.; Metzidakis, I. Identification of *Olea europaea* L. cultivars using inter-simple sequence repeat markers. *Sci. Hortic.* **2005**, *105*, 45–51. [\[CrossRef\]](#)
128. Essadki, M.; Ouazzani, N.; Lumaret, R.; Moumni, M. ISSR variation in olive-tree cultivars from Morocco and other western countries of the Mediterranean basin. *Genet. Resour. Crop Evol.* **2006**, *53*, 475–482. [\[CrossRef\]](#)
129. Owen, C.A.; Bitá, E.C.; Banilas, G.; Hajjar, S.E.; Sellianakis, V.; Aksoy, U.; Hepaksoy, S.; Chamoun, R.; Talhook, S.N.; Metzidakis, I.; et al. AFLP reveals structural details of genetic diversity within cultivated olive germplasm from the Eastern Mediterranean. *Theor. Appl. Genet.* **2005**, *110*, 1169–1176. [\[CrossRef\]](#)
130. Ercisli, S.; Barut, E.; Ipek, A. Molecular characterization of olive cultivars using amplified fragment length polymorphism markers. *Genet. Mol. Res.* **2009**, *8*, 414–419. [\[CrossRef\]](#)
131. Ipek, M.; Seker, M.; Ipek, A.; Gul, M.K. Identification of molecular markers associated with fruit traits in olive and assessment of olive core collection with AFLP markers and fruit traits. *Genet. Mol. Res.* **2015**, *14*, 2762–2774. [\[CrossRef\]](#) [\[PubMed\]](#)
132. El Aabidine, A.Z.; Charafi, J.; Grout, C.; Doligez, A.; Santoni, S.; Moukhli, A.; Jay-Allemand, C.; El Modafar, C.; Khadari, B. Construction of a genetic linkage map for the olive based on AFLP and SSR markers. *Crop Sci.* **2010**, *50*, 2291–2302. [\[CrossRef\]](#)
133. Khadari, B.; El Aabidine, A.Z.; Grout, C.; Ben Sadok, I.; Doligez, A.; Moutier, N.; Santoni, S.; Costes, E. A genetic linkage map of olive based on amplified fragment length polymorphism, intersimple sequence repeat and simple sequence repeat markers. *J. Am. Soc. Hortic. Sci.* **2010**, *135*, 548–555. [\[CrossRef\]](#)
134. Ben-Ayed, R.; Ennouri, K.; Ben-Hassen, H.; Triki, M.A.; Rebaï, A. Comparison between DNA-based, pomological and chemical markers accomplished by bioinformatic tools to distinguish within Tunisian olive cultivars. *J. Fundam. Appl. Sci.* **2015**, *7*, 408–421. [\[CrossRef\]](#)
135. Morgante, M.; Olivieri, A.M. PCR-amplified microsatellites as markers in plant genetics. *Plant J.* **1993**, *3*, 175–182. [\[CrossRef\]](#)
136. De la Rosa, R.; James, C.M.; Tobutt, K.R. Isolation and characterization of polymorphic microsatellites in olive (*Olea europaea* L.) and their transferability to other genera in the *Oleaceae*. *Mol. Ecol. Notes* **2002**, *2*, 265–267. [\[CrossRef\]](#)

137. Rallo, P.; Tenzer, I.; Gessler, C.; Baldoni, L.; Dorado, G.; Martín, A. Transferability of olive microsatellite loci across the genus *Olea*. *Theor. Appl. Genet.* **2003**, *107*, 940–946. [\[CrossRef\]](#)
138. Belaj, A.; Cipriani, G.; Testolin, R.; Rallo, L.; Trujillo, I. Characterization and identification of the main Spanish and Italian olive cultivars by simple-sequence-repeat markers. *HortScience* **2004**, *39*, 1557–1561. [\[CrossRef\]](#)
139. Salimonti, A.; Simeone, V.; Cesari, G.; Lamaj, F.; Cattivelli, L.; Perri, E.; Desiderio, F.; Fanizzi, F.P.; Del Coco, L.; Zelasco, S. A first molecular investigation of monumental olive trees in Apulia region. *Sci. Hortic.* **2013**, *162*, 204–212. [\[CrossRef\]](#)
140. Xu, Y. *Molecular Plant Breeding*; CAB International: Oxfordshire, UK, 2010; pp. 195–247. ISBN 978-184-593-392-0.
141. Reale, S.; Doveri, S.; Díaz, A.; Angiolillo, A.; Lucentini, L.; Pilla, F.; Martín, A.; Donini, P.; Lee, D. SNP-based markers for discriminating olive (*Olea europaea* L.) cultivars. *Genome* **2006**, *49*, 1193–1205. [\[CrossRef\]](#)
142. Carriero, F.; Fontanazza, G.; Cellini, F.; Giorio, G. Identification of simple sequence repeats (SSRs) in olive (*Olea europaea* L.). *Theor. Appl. Genet.* **2002**, *104*, 301–307. [\[CrossRef\]](#) [\[PubMed\]](#)
143. Baldoni, L.; Cultrera, N.G.; Mariotti, R.; Riccioloni, C.; Arcioni, S.; Vendramin, G.G.; Buonamici, A.; Porceddu, A.; Sarri, V.; Ojeda, M.A.; et al. A consensus list of microsatellite markers for olive genotyping. *Mol. Breed.* **2009**, *24*, 213–231. [\[CrossRef\]](#)
144. Belaj, A.; Muñoz-Díez, C.; Baldoni, L.; Porceddu, A.; Barranco, D.; Šatović, Z. Genetic diversity and population structure of wild olives from the North-western Mediterranean by SSR markers. *Ann. Bot.* **2007**, *100*, 449–458. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Ipek, A.; Barut, E.; Gulen, H.; Ipek, M. Assessment of inter-and intra-cultivar variations in olive using SSR markers. *Sci. Agric.* **2012**, *69*, 327–335. [\[CrossRef\]](#)
146. Caruso, T.; Marra, F.P.; Costa, F.; Campisi, G.; Macaluso, L.; Marchese, A. Genetic diversity and clonal variation within the main Sicilian olive cultivars based on morphological traits and microsatellite markers. *Sci. Hortic.* **2014**, *180*, 130–138. [\[CrossRef\]](#)
147. Abdessemed, S.; Muzzalupo, I.; Benbouza, H. Assessment of genetic diversity among Algerian olive (*Olea europaea* L.) cultivars using SSR marker. *Sci. Hortic.* **2015**, *192*, 10–20. [\[CrossRef\]](#)
148. Lazović, B.; Klepo, T.; Adakalić, M.; Šatović, Z.; Arbeiter, A.B.; Hladnik, M.; Strikić, F.; Liber, Z.; Bandelj, D. Intra-varietal variability and genetic relationships among the homonymic East Adriatic olive (*Olea europaea* L.) varieties. *Sci. Hortic.* **2018**, *236*, 175–185. [\[CrossRef\]](#)
149. Ben Sadok, I.; Celton, J.M.; Essalouh, L.; El Aabidine, A.Z.; Garcia, G.; Martinez, S.; Grati-Kamoun, N.; Rebai, A.; Costes, E.; Khadari, B. QTL mapping of flowering and fruiting traits in olive. *PLoS ONE* **2013**, *8*, e62831. [\[CrossRef\]](#)
150. Mookerjee, S.; Guerin, J.; Collins, G.; Ford, C.; Sedgley, M. Paternity analysis using microsatellite markers to identify pollen donors in an olive grove. *Theor. Appl. Genet.* **2005**, *111*, 1174–1182. [\[CrossRef\]](#)
151. De la Rosa, R.; Belaj, A.; Muñoz-Mérida, A.; Trelles, O.; Ortiz-Martín, I.; González-Plaza, J.J.; Valpuesta, V.; Beuzón, C.R. Development of EST-derived SSR markers with long-core repeat in olive and their use for paternity testing. *J. Am. Soc. Hortic. Sci.* **2013**, *138*, 290–296. [\[CrossRef\]](#)
152. Hakim, I.R.; Grati-Kammoun, N.; Makhoulfi, E.; Rebaï, A. Discovery and potential of SNP markers in characterization of Tunisian olive germplasm. *Diversity* **2010**, *2*, 17–27. [\[CrossRef\]](#)
153. Domínguez-García, M.C.; Laib, M.; De La Rosa, R.; Belaj, A. Characterisation and identification of olive cultivars from North-eastern Algeria using molecular markers. *J. Hortic. Sci. Biotechnol.* **2012**, *87*, 95–100. [\[CrossRef\]](#)
154. Biton, I.; Doron-Faigenboim, A.; Jamwal, M.; Mani, Y.; Eshed, R.; Rosen, A.; Sherman, A.; Ophir, R.; Lavee, S.; Avidan, B.; et al. Development of a large set of SNP markers for assessing phylogenetic relationships between the olive cultivars composing the Israeli olive germplasm collection. *Mol. Breed.* **2015**, *35*, 107–120. [\[CrossRef\]](#)
155. Zhu, S.; Niu, E.; Shi, A.; Mou, B. Genetic diversity analysis of olive germplasm (*Olea europaea* L.) with Genotyping-by-Sequencing Technology. *Front. Genet.* **2019**, *10*, 755. [\[CrossRef\]](#)
156. Baldoni, L.; Khadari, B.; De la Rosa, R. Genetic mapping and detection of Quantitative Trait Loci. In *The Olive Tree Genome*; Rugini, E., Baldoni, L., Muleo, R., Sebastiani, L., Eds.; Springer: Cham, Switzerland, 2016; pp. 65–74. [\[CrossRef\]](#)
157. İpek, A.; Yılmaz, K.; Sıkıcı, P.; Tangu, N.A.; Öz, A.T.; Bayraktar, M.; İpek, M.; Gülen, H. SNP discovery by GBS in olive and the construction of a high-density genetic linkage map. *Biochem. Genet.* **2016**, *54*, 313–325. [\[CrossRef\]](#)

158. Marchese, A.; Marra, F.P.; Caruso, T.; Mhelembe, K.; Costa, F.; Fretto, S.; Sargent, D.J. The first high-density sequence characterized SNP-based linkage map of olive (*Olea europaea* L. subsp. *europaea*) developed using genotyping by sequencing. *Aust. J. Crop Sci.* **2016**, *10*, 857. [CrossRef]
159. El Bakkali, A.; Essalouh, L.; Tollon, C.; Rivallan, R.; Mournet, P.; Moukhli, A.; Zaher, H.; Mekkaoui, A.; Hadidou, A.; Sikaoui, L.; et al. Characterization of worldwide olive germplasm banks of Marrakech (Morocco) and Córdoba (Spain): Towards management and use of olive germplasm in breeding programs. *PLoS ONE* **2019**, *14*, e0223716. [CrossRef]
160. FAO. *International Treaty on Plant Genetic Resources for Food and Agriculture*; Food and Agriculture Organization of the United Nations: Rome, Italy, 2001. Available online: <http://www.fao.org/plant-treaty/overview/en/> (accessed on 15 July 2020).
161. Sardaro, R.; Bozzo, F.; Petrontino, A.; Fucilli, V. Community preferences in support of a conservation programme for olive landraces in the Mediterranean area. *Acta Hort.* **2018**, *1199*, 183–188. [CrossRef]
162. Rallo, L.; Barranco, D.; Díez, C.M.; Rallo, P.; Suárez, M.P.; Trapero, C.; Pliego-Alfaro, F. Strategies for olive (*Olea europaea* L.) breeding: Cultivated genetic resources and crossbreeding. In *Advances in Plant Breeding Strategies: Fruits*; Al-Khayri, J., Jain, S.M., Johnson, D.V., Eds.; Springer: Cham, Switzerland, 2018; Volume 3, pp. 536–600.
163. Baenziger, P.S.; Russell, W.K.; Graef, G.L.; Campbell, B.T. Improving lives: 50 years of crop breeding, genetics, and cytology (C-1). *Crop Sci.* **2006**, *46*, 2230–2244. [CrossRef]
164. Rallo, L.; Caruso, T.; Díez, C.M.; Campisi, G. Olive growing in a time of change: From empiricism to genomics. In *The Olive Tree Genome*; Rugini, E., Baldoni, L., Muleo, R., Sebastiani, L., Eds.; Springer: Cham, Switzerland, 2016; pp. 55–64. [CrossRef]
165. Baldoni, L.; Belaj, A. Olive. In *Oil Crops. Handbook of Plant Breeding*; Vollmann, J., Rajcan, I., Eds.; Springer: New York, NY, USA, 2009; Volume 4, pp. 397–421. [CrossRef]
166. Rallo, L. Breeding oil and table olives for mechanical harvesting in Spain. *Hortech* **2014**, *24*, 295–300. [CrossRef]
167. Arias-Calderón, R.; Rodríguez-Jurado, D.; León, L.; Bejarano-Alcázar, J.; De la Rosa, R.; Belaj, A. Pre-breeding for resistance to Verticillium wilt in olive: Fishing in the wild relative gene pool. *Crop Prot.* **2015**, *75*, 25–33. [CrossRef]
168. Trapero, C.; Rallo, L.; López-Escudero, F.J.; Barranco, D.; Díez, C.M. Variability and selection of verticillium wilt resistant genotypes in cultivated olive and in the *Olea* genus. *Plant Pathol.* **2015**, *64*, 890–900. [CrossRef]
169. Klepo, T.; Toumi, A.; De la Rosa, R.; León, L.; Belaj, A. Agronomic evaluation of seedlings from crosses between the main Spanish olive cultivar ‘Picual’ and two wild olive trees. *J. Hort. Sci. Biotechnol.* **2014**, *89*, 508–512. [CrossRef]
170. Caceres, M.E.; Ceccarelli, M.; Pupilli, F.; Sarri, V.; Mencuccini, M. Obtainment of inter-subspecific hybrids in olive (*Olea europaea* L.). *Euphytica* **2015**, *201*, 307–319. [CrossRef]
171. Rugini, E.; Cristofori, V.; Silvestri, C. Genetic improvement of olive (*Olea europaea* L.) by conventional and in vitro biotechnology methods. *Biotechnol. Adv.* **2016**, *34*, 687–696. [CrossRef]
172. Rugini, E.; De Pace, C. Olive breeding with classical and modern approaches. In *The Olive Tree Genome*; Rugini, E., Baldoni, L., Muleo, R., Sebastiani, L., Eds.; Springer: Cham, Switzerland, 2016; pp. 163–193. [CrossRef]
173. Leva, A.R. Innovative protocol for “ex vitro rooting” on olive micropropagation. *Cent. Eur. J. Biol.* **2011**, *6*, 352–358. [CrossRef]
174. Benelli, C.; De Carlo, A.; Englmen, F. Recent advances in the cryopreservation of shoot-derived germplasm of economically important fruit trees of *Actinidia*, *Diospyros*, *Malus*, *Olea*, *Prunus*, *Pyrus* and *Vitis*. *Biotechnol. Adv.* **2013**, *31*, 175–185. [CrossRef]
175. Rugini, E.; Silvestri, C.; Ceccarelli, M.; Muleo, R.; Cristofori, V. Mutagenesis and biotechnology techniques as tools for selecting new stable diploid and tetraploid olive genotypes and their dwarfing agronomical characterization. *HortScience* **2016**, *51*, 1–6. [CrossRef]
176. Capelo, A.M.; Silva, S.; Brito, G.; Santos, C. Somatic embryogenesis induction in leaves and petioles of a mature wild olive. *Plant Cell Tissue Org.* **2010**, *103*, 237–242. [CrossRef]
177. Toufik, I.; Guenoun, F.; Belkoura, I. Embryogenesis expression from somatic explants of olive (*Olea europaea* L.) cv Picual. *Moroc. J. Biol.* **2014**, *11*, 17–25.

178. Torreblanca, R.; Cerezo, S.; Palomo-Ríos, E.; Mercado, J.A.; Pliego-Alfaro, F. Development of a high throughput system for genetic transformation of olive (*Olea europaea* L.) plants. *Plant Cell Tissue Org.* **2010**, *103*, 61–69. [\[CrossRef\]](#)
179. Titouh, K.; Khelifi, L.; Moussa, K.T.; Cerezo-Medina, S.; Mercado, J.A.; Pliego-Alfaro, F. Evaluation of the effect of phosphinothricin, as selection agent, on the growth of olive somatic embryos. *Acta Hort.* **2014**, *1057*, 533–542. [\[CrossRef\]](#)
180. Ozgenturk, N.O.; Oruç, F.; Sezerman, U.; Kuçukural, A.; Korkut, S.V.; Toksoz, F.; Un, C. Generation and analysis of expressed sequence tags from *Olea europaea* L. *Comp. Funct. Genom.* **2010**, *2010*, 757512. [\[CrossRef\]](#)
181. Bazakos, C.; Manioudaki, M.E.; Therios, I.; Voyiatzis, D.; Kafetzopoulos, D.; Awada, T.; Kalaitzis, P. Comparative transcriptome analysis of two olive cultivars in response to NaCl-stress. *PLoS ONE* **2012**, *7*, e42931. [\[CrossRef\]](#) [\[PubMed\]](#)
182. Parra, R.; Paredes, M.A.; Sanchez-Calle, I.M.; Gomez-Jimenez, M.C. Comparative transcriptional profiling analysis of olive ripe-fruit pericarp and abscission zone tissues shows expression differences and distinct patterns of transcriptional regulation. *BMC Genom.* **2013**, *14*, 866. [\[CrossRef\]](#)
183. Yanik, H.; Turktas, M.; Dundar, E.; Hernandez, P.; Dorado, G.; Unver, T. Genome-wide identification of alternate bearing-associated microRNAs (miRNAs) in olive (*Olea europaea* L.). *BMC Plant Biol.* **2013**, *13*, 10. [\[CrossRef\]](#)
184. Cabanás, C.G.; Schilirò, E.; Valverde-Corredor, A.; Mercado-Blanco, J. Systemic responses in a tolerant olive (*Olea europaea* L.) cultivar upon root colonization by the vascular pathogen *Verticillium dahliae*. *Front. Microbiol.* **2015**, *6*, 928. [\[CrossRef\]](#)
185. Carmona, R.; Zafra, A.; Seoane, P.; Castro, A.J.; Guerrero-Fernández, D.; Castillo-Castillo, T.; Medina-García, A.; Cánovas, F.M.; Aldana-Montes, J.F.; Navas-Delgado, I.; et al. ReprOlive: A database with linked data for the olive tree (*Olea europaea* L.) reproductive transcriptome. *Front. Plant Sci.* **2015**, *6*, 625. [\[CrossRef\]](#)
186. Leyva-Pérez, M.O.; Valverde-Corredor, A.; Valderrama, R.; Jiménez-Ruiz, J.; Muñoz-Merida, A.; Trelles, O.; Barroso, J.B.; Mercado-Blanco, J.; Luque, F. Early and delayed long-term transcriptional changes and short-term transient responses during cold acclimation in olive leaves. *DNA Res.* **2015**, *22*, 1–11. [\[CrossRef\]](#)
187. Giampetruzzi, A.; Morelli, M.; Saponari, M.; Loconsole, G.; Chiumenti, M.; Boscia, D.; Saldarelli, P. Transcriptome profiling of two olive cultivars in response to infection by the OQDS strain of *Xylella fastidiosa* ssp. *pauca*. *BMC Genom.* **2016**, *17*, 1. [\[CrossRef\]](#)
188. González-Plaza, J.J.; Ortiz-Martín, I.; Muñoz-Mérida, A.; García-López, C.; Sánchez-Sevilla, J.F.; Luque, F.; Trelles, O.; Bejarano, E.R.; De la Rosa, R.; Valpuesta, V.; et al. Transcriptomic analysis using olive varieties and breeding progenies identifies candidate genes involved in plant architecture. *Front. Plant Sci.* **2016**, *7*, 240. [\[CrossRef\]](#) [\[PubMed\]](#)
189. Sarah, G.; Homa, F.; Pointet, S.; Contreras, S.; Sabot, F.; Nabholz, B.; Santoni, S.; Sauné, L.; Ardisson, M.; Chantret, N.; et al. A large set of 26 new reference transcriptomes dedicated to comparative population genomics in crops and wild relatives. *Mol. Ecol. Resour.* **2017**, *17*, 565–580. [\[CrossRef\]](#) [\[PubMed\]](#)
190. Zafra, A.; Carmona, R.; Traverso, J.A.; Hancock, J.T.; Goldman, M.H.S.; Claros, M.G.; Hiscock, S.J.; Alche, J.D. Identification and functional annotation of genes differentially expressed in the reproductive tissues of the olive tree (*Olea europaea* L.) through the generation of subtractive libraries. *Front. Plant Sci.* **2017**, *8*, 1576. [\[CrossRef\]](#) [\[PubMed\]](#)
191. Guodong, R.; Jianguo, Z.; Xiaoxia, L.; Ying, L. Identification of putative genes for polyphenol biosynthesis in olive fruits and leaves using full-length transcriptome sequencing. *Food Chem.* **2019**, *300*, 125246. [\[CrossRef\]](#)
192. Besnard, G.; Hernández, P.; Khadari, B.; Dorado, G.; Savolainen, V. Genomic profiling of plastid DNA variation in the Mediterranean olive tree. *BMC Plant Biol.* **2011**, *11*, 80. [\[CrossRef\]](#)
193. Kaya, E.; Vatansever, R.; Filiz, E. Assessment of the genetic relationship of Turkish olives (*Olea europaea* subsp. *europaea*) cultivars based on cpDNA *trnL-F* regions. *Acta Bot. Croat.* **2018**, *77*, 88–92. [\[CrossRef\]](#)
194. Rugini, E.; De Pace, C.; Gutiérrez-Pesce, P.; Muleo, R. Olea. In *Wild Crop Relatives: Genomic and Breeding Resources*; Kole, C., Ed.; Springer: Berlin/Heidelberg, Germany, 2011; pp. 79–144. [\[CrossRef\]](#)
195. Kaya, H.B.; Cetin, O.; Kaya, H.S.; Sahin, M.; Sefer, F.; Tanyolac, B. Association mapping in Turkish olive cultivars revealed significant markers related to some important agronomic traits. *Biochem. Genet.* **2016**, *54*, 506–533. [\[CrossRef\]](#)

196. Atienza, S.G.; De La Rosa, R.; León, L.; Martín, A.; Belaj, A. Identification of QTL for agronomic traits of importance for olive breeding. *Mol. Breed.* **2014**, *34*, 725–737. [[CrossRef](#)]
197. Ates, D. Identification of QTLs controlling genes of ripening time, flesh detachment from stone and firmness in olive (*Olea europaea* L.). In Proceedings of the 24th Plant and Animal Genome Conference, San Diego, CA, USA, 9–13 January 2016; p. P1163.
198. Mauricio, R. Mapping quantitative trait loci in plants: Uses and caveats for evolutionary biology. *Nat. Rev. Genet.* **2001**, *2*, 370–381. [[CrossRef](#)]
199. Morrell, P.L.; Buckler, E.S.; Ross-Ibarra, J. Crop genomics: Advances and applications. *Nat. Rev. Genet.* **2012**, *13*, 85–96. [[CrossRef](#)]
200. Yu, J.; Buckler, E.S. Genetic association mapping and genome organization of maize. *Curr. Opin. Biotechnol.* **2006**, *17*, 155–160. [[CrossRef](#)] [[PubMed](#)]
201. Kaya, H.B.; Akdemir, D.; Lozano, R.; Cetin, O.; Kaya, H.S.; Sahin, M.; Smith, J.L.; Tanyolac, B.; Jannink, J.L. Genome wide association study of 5 agronomic traits in olive (*Olea europaea* L.). *Sci. Rep.* **2019**, *9*, 1–14. [[CrossRef](#)]
202. Salimonti, A.; Carbone, F.; Romano, E.; Pellegrino, M.; Benincasa, C.; Micali, S.; Tondelli, A.; Conforti, F.L.; Perri, E.; Ienco, A.; et al. Association study of the 5' UTR intron of the *FAD2-2* gene with oleic and linoleic acid content in *Olea europaea* L. *Front. Plant Sci.* **2020**, *11*, 66. [[CrossRef](#)] [[PubMed](#)]
203. Van Eeuwijk, F.A.; Bustos-Korts, D.; Millet, E.J.; Boer, M.P.; Kruijer, W.; Thompson, A.; Malosetti, M.; Iwata, H.; Quiroz, R.; Kuppe, C.; et al. Modelling strategies for assessing and increasing the effectiveness of new phenotyping techniques in plant breeding. *Plant Sci.* **2019**, *282*, 23–39. [[CrossRef](#)] [[PubMed](#)]
204. Podlich, D.W.; Winkler, C.R.; Cooper, M. Mapping as you go. An effective approach for marker assisted selection of complex traits. *J. Crop Sci.* **2004**, *44*, 1560–1571. [[CrossRef](#)]
205. Malosetti, M.; Ribaut, J.M.; Van Eeuwijk, F.A. The statistical analysis of multi-environment data: Modeling genotype-by-environment interaction and its genetic basis. *Front. Physiol.* **2013**, *4*, 44. [[CrossRef](#)]
206. Hammer, G.; Messina, C.; Van Oosterom, E.; Chapman, S.; Singh, V.; Borrell, A.; Jordan, D.; Cooper, M. Molecular breeding for complex adaptive traits: How integrating crop ecophysiology and modelling can enhance efficiency. In *Crop Systems Biology*; Yin, X., Struik, P., Eds.; Springer: Cham, Switzerland, 2016; pp. 147–162. [[CrossRef](#)]
207. Van Eeuwijk, F.A.; Bink, M.C.A.M.; Chenu, K.; Chapman, S.C. Detection and use of QTL for complex traits in multiple environments. *Curr. Opin. Plant Biol.* **2010**, *13*, 193–205. [[CrossRef](#)]
208. Bustos-Korts, D.; Malosetti, M.; Chapman, S.; Van Eeuwijk, F. Modelling of genotype by environment interaction and prediction of complex traits across multiple environments as a synthesis of crop growth modelling, genetics and statistics. In *Crop Systems Biology*; Yin, X., Struik, P., Eds.; Springer: Cham, Switzerland, 2016; pp. 55–82. [[CrossRef](#)]
209. Smith, A.B.; Cullis, B.R.; Thompson, R. The analysis of crop cultivar breeding and evaluation trials: An overview of current mixed model approaches. *J. Agric. Sci.* **2005**, *143*, 449–462. [[CrossRef](#)]
210. Technow, F.; Messina, C.D.; Totir, L.R.; Cooper, M. Integrating crop growth models with whole genome prediction through approximate bayesian computation. *PLoS ONE* **2015**, *10*, e0130855. [[CrossRef](#)]
211. López-Bernal, Á.; Morales, A.; García-Tejera, O.; Testi, L.; Orgaz, F.; De Melo-Abreu, J.P.; Villalobos, F.J. OliveCan: A process-based model of development, growth and yield of olive orchards. *Front. Plant Sci.* **2018**, *9*, 632. [[CrossRef](#)]
212. Cruz, F.; Julca, I.; Gómez-Garrido, J.; Loska, D.; Marcet-Houben, M.; Cano, E.; Galán, B.; Frias, L.; Ribeca, P.; Derdak, S.; et al. Genome sequence of the olive tree, *Olea europaea*. *GigaScience* **2016**, *5*, 29. [[CrossRef](#)] [[PubMed](#)]

